

1 **Table S1.** Crystallographic data, structure solution and refinement statistics.

	Derivative	Native 1	Native 2	GlcNAc-soaked
<i>Data</i>				
Synchrotron beamline	ALBA BL13-Xaloc	ALBA BL13-Xaloc	ESRF ID29	ESRF ID23-2
Data collection date	31 January 2014	31 January 2014	21 February 2014	31 October 2014
Detector	Dectris Pilatus 6M	Dectris Pilatus 6M	ADSC Quantum 315r	MAR Mosaic 225
Spacegroup	<i>C</i> 222 ₁	<i>P</i> 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>I</i> 2 ₃
Cell parameters a, b, c (Å)	92.1, 164.3, 96.4	93.88, 100.60, 95.47	55.87, 91.11, 128.92	163.41, 163.41, 163.41
Cell angles α , β , γ (°)	90.0, 90.0, 90.0	90.00, 116.28, 90.00	90.00, 90.00, 90.00	90.00, 90.00, 90.00
Wavelength (Å)	1.0024	1.0023	0.9763	0.8729
Average mosaicity (°)	0.14	0.48	0.27	0.41
Resolution (Å)	96.4-2.76 (3.02-2.76)	29.7-1.80 (1.83-1.80)	34.1-1.70 (1.74-1.70)	28.9-2.00 (2.05-2.00)
Observed reflections	19199 (4473)	144435 (7096)	68912 (3722)	48930 (3642)
Completeness	0.997 (0.988)	0.982 (0.978)	0.943 (0.696)	1.000 (1.000)
Multiplicity	11.7 (8.0)	2.8 (2.9)	3.5 (1.9)	7.9 (7.8)
Rmerge	0.104 (0.589)	0.078 (0.334)	0.116 (0.233)	0.101 (0.404)
I/sigma(I)	19.6 (3.2)	7.1 (2.5)	6.3 (2.1)	13.2 (4.8)
CC1/2	0.998 (0.867)	0.989 (0.857)	0.983 (0.857)	0.998 (0.714)
Wilson B (Å ²)	71.3	16.5	14.5	20.0
<i>Phasing</i>				
Number of heavy atoms	12 Hg	-	-	-
Correlation coefficient	0.462	-	-	-
Anomalous phasing power	0.753	-	-	-
Figure of merit	0.250	-	-	-
Solvent flattening (solvent fraction)	0.539	-	-	-
Hand score (original / inverted)	0.2471 / 0.1310	-	-	-
Correlation on E ² / contrast	2.5866	-	-	-
<i>Refinement</i>				
Resolution	82.1-2.76 (2.83-2.76)	29.7-1.80 (1.90-1.80)	34.1-1.70 (1.82-1.70)	28.0-2.00 (2.05-2.00)
Reflections used	18197 (1269)	140696 (20353)	65280 (9409)	46439 (3444)
Reflections used for Rfree	985 (69)	3644 (499)	3559 (540)	2409 (165)
R-factor	0.203 (0.327)	0.168 (0.232)	0.172 (0.257)	0.165 (0.269)
Rfree	0.248 (0.377)	0.201 (0.253)	0.216 (0.266)	0.205 (0.294)
No. of atoms (total / protein / Hg / water / chloride / glycerol / sulfate / GlcNAc)	4578 / 4529 / 5 / 44 / 0 / 0 / 0 / 0	10928 / 9213 / 0 / 1664 / 8 / 42 / 0 / 0	5285 / 4607 / 0 / 672 / 0 / 6 / 0 / 0	5036 / 4580 / 0 / 350 / 0 / 6 / 55 / 45
Average B (overall / protein / Hg / water / chloride / glycerol / sulfate / GlcNAc)	62.4 / 62.6 / 70.0 / 41.6 / - / - / - / -	25.7 / 23.7 / - / 36.8 / 33.3 / 37.0 / - / -	21.1 / 19.8 / - / 30.1 / - / 48.1 / - / -	29.9 / 29.1 / - / 36.7 / - / 40.1 / 44.4 / 30.2
Ramachandran (favored / allowed)	0.929 / 0.990	0.968 / 1.000	0.976 / 1.000	0.971 / 0.998
R.m.s.d. bonds (Å) / angles (°)	0.009 / 1.5	0.013 / 1.6	0.012 / 1.5	0.010 / 1.4
PDB code	5N83	5N8D	5NBH	5NC1

2 Values in parentheses are for the highest resolution bin

4 Table S2. Abbreviations, names, print concentrations and primary structures of the glycans used in
5 the microarray.

Abbreviation	Neoglycoconjugate/ glycoprotein	Print conc (mg/mL)	Structure
1 Fetuin	Fetuin	1	Bovine fetuin
2 Ov	Ovalbumin	1	Hen ovalbumin
3 4APHS	4AP-HSA	1	4AP-HSA, linker alone attached to HSA
4 a-C	α -Crystallin from bovine lens	1	α -Crystallin from bovine lens, A and B subunits
5 M3BSA	Man α 1,3(Man α 1,6)Man-BSA	1	Man α -(1,3)-(Man α -(1,6)-)Man-BSA
6 GlcNAcBSA	GlcNAc-BSA	1	GlcNAc-Sp14-NH2(Lys)-BSA
7 3SLacHSA	3'-Sialyllactose-APD-HSA	1	Neu5Ac α -(2,3)-Gal- β -(1,4)-Glc-APD-HSA
8 6SLacHSA	6'-Sialyllactose-APD-HSA	1	Neu5Ac α -(2,6)-Gal- β -(1,4)-Glc-APD-HSA
9 H2BSA	H Type II-APE-BSA	1	Fuc α -(1,2)-Gal- β -(1,4)-GlcNAc β -APE-BSA
10 GGGHSA	Gak1,3Gal β 1,4GlcNAc-HSA	1	Gal α -(1,3)-Gal- β -(1,4)-GlcNAc-HSA
11 Ga3GBSA	Gak1,3Gal-BSA	1	Gal α -(1,3)-Gal- β -Sp3-BSA
12 4APBSA	4AP-BSA	1	4AP-HSA, linker alone attached to BSA
13 LNPFIBSA	Lacto-N-fucopentaose I-BSA	1	Fuc α -(1,2)-Gal- β -(1,3)-GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-BSA
14 LebBSA	LDI-BSA/ Lewis b-BSA	1	Fuc α -(1,2)-Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-APD-BSA
15 LexBSA	Lewis x-BSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc-BSA
16 3LexHSA	Tri-Lex-APE-BSA	1	Gal- β -(1,2)-Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc-BSA
17 6SLeaBSA	6-Sulfo Lewis a-BSA	1	[SO4]6Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc-Sp3-BSA
18 LeyHSA	Lewis y-tetrasaccharide-APE-HSA	1	Fuc α -(1,2)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -O-APE-HSA
19 LNTHSA	Lacto-N-tetraose-APD-HSA	1	Gal β -(1,3)-GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-APD-HSA
20 GlobNTHSA	Globotriose-APD-HSA	1	GalNAc β -(1,3)-Gal- β -(1,4)-Gal- β -(1,4)-Glc-APD-HSA
21 GlobTHSA	Globotriose-APE-HSA	1	Gal α -(1,4)-Gal- β -(1,4)-Glc- β -APE-HSA
22 ASF	Asialofetuin	1	Bovine asialofetuin
23 PBS	PBS	-	Phosphate buffered saline
24 RB	RNAse B	1	Ribonuclease B
25 Xferrin	Transferrin	1	Bovine transferrin
26 LacNAcBSA	LacNAc-BSA	1	Gal β -(1,4)-GlcNAc-Sp3-BSA
27 3SLNBSA	3'SialylLacNAc-BSA	1	Neu5Ac α -(2,3)-Gal- β -(1,4)-GlcNAc-BSA
28 2FLBSA	2'Fucosyllactose-BSA	1	Fuc α -(1,2)-Gal- β -(1,4)-Glc-Sp3-BSA
29 3SFLBSA	3'Sialyl-3-fucosyllactose-BSA	1	Neu5Ac α -(2,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]Glc-Sp3-BSA
30 BGABSA	Blood Group A-BSA	1	GalNAc α -(1,3)-[Fuc α -(1,2)-]Gal- β -(1,4)-GlcNAc-Sp6-BSA
31 BGHBSA	Blood Group B-BSA	1	Gal α -(1,3)-[Fuc α -(1,2)-]Gal- β -(1,4)-GlcNAc-Sp6-HSA
32 Gb4GBSA	Galb1,4GalBSA	1	Gal- β -(1,4)-Gal-Sp3-BSA
33 Ga2GBSA	Galb1,2GalBSA	1	Gal- β -(1,2)-Gal-Sp3-BSA
34 LNPFIIIBSA	Lacto-N-fucopentaose II-BSA	1	Fuc α -(1,3)-Gal- β -(1,3)-GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-BSA
35 LNPFIIIIBSA	Lacto-N-fucopentaose III-BSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-BSA
36 LDNHIBSA	Lacto-N-difucosylhexaose I-BSA	1	Fuc α -(1,2)-Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-Sp3-BSA
37 DILexBSA	DI-Lex-APE-BSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -O-APE-BSA
38 DILexHSA	DI-Lewisx-APE-HSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -O-APE-HSA
39 3SLexBSA3	3'Sialyl Lewis x-BSA	1	Neu5Ac α -(2,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc-Sp3-BSA
40 SLexBSA14	3'Sialyl Lewis x-BSA	1	Neu5Ac α -(2,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc-Sp14-BSA
41 6SLuLexBSA	6-Sulfo Lewis x-BSA	1	(SO4)6Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc-Sp3-BSA
42 3SuLeaBSA	3-Sulfo Lewis a-BSA	1	(SO4)3Gal- β -1-3-[Fuc α -(1,4)-]GlcNAc-Sp3-BSA
43 3SLuLexBSA	3-Sulfo Lewis x-BSA	1	(SO4)3Gal- β -1-3-[Fuc α -(1,4)-]GlcNAc-Sp3-BSA
44 DFPLNHHS	Difucosyl-para-lacto-N-hexaose-APD-HSA, (Lea/Lex)	1	Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-BSA
45 LeaBSA	Lewis a-BSA	1	Fuc α -(1,2)-Gal- β -(1,3)-[Fuc α -(1,4)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-Sp3-BSA
46 3FLeyHSA	Tri-fucosyl-Ley-heptasaccharide-APE-HSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -O-APE-BSA
47 LNTHSA	Lacto-N-neotetraose-APD-HSA	1	Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -O-APE-HSA
48 SLNFVHSA	Sialyl-LNF V-APD-HSA	1	Fuc α -(1,2)-Gal- β -(1,4)-[Fuc α -(1,3)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-APD-HSA
49 MMLNhHSA	Monofucosyl monosialylacto-N-neohexaose-APD-HSA	1	Fuc α -(1,2)-Gal- β -(1,3)-[NeuAc α -(2,6)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-APD-HSA
50 SLNnHSA	Sialyl-LNF N-penta-APD-HSA	1	Neu5Ac α -(2,3)-Gal- β -(1,3)-[NeuAc α -(2,6)-]GlcNAc β -(1,3)-Gal- β -(1,4)-Glc-APD-HSA
51 GM1HSA	GM1-pentasaccharide-APD-HSA	1	Gal- β -(1,3)-GalNAc β -(1,4)-[Neu5Ac α -(2,3)-]Gal- β -(1,4)-Glc-APD-HSA
52 aGM1HSA	Asialo-GM1-tetrasaccharide-APD-HSA	1	Gal- β -(1,3)-GalNAc β -(1,4)-Gal- β -(1,4)-Glc-APD-HSA
53 Inv	Invertase	1	Yeast invertase, grade VII
54 Fibrin	Fibrinogen	0.5	Fibrinogen from human plasma
55 A1AT	alpha-1-antitrypsin	1	alpha-1-antitrypsin
56 Cerulo	Ceruloplasmin	1	Ceruloplasmin, human, type III

57 AGP	alpha-1-acid glycoprotein	1	alpha-1-acid glycoprotein, human
58 LacNAcBSA	LacNAc α -4AP-BSA	1	Gal- β -(1,4)-GlcNAc α -4AP-BSA
59 LacAcbBSA	LacNAc β -4AP-BSA	1	Gal- β -(1,4)-GlcNAc β -4AP-BSA
60 PBS	Phosphate buffered saline	-	Phosphate buffered saline
61 Ovomuc	Ovomucoid	0.5	Partially purified ovomucoid, chicken
62 RhaBSA	L-Rhamnose-Sp14-BSA	1	L-Rhamnose-Sp14-BSA
63 GalaPITCBSA	Gal-a-PITC-BSA	1	Gal-a-PITC-BSA
64 XManaBSA	Man α -ITC-BSA	1	Man α -ITC-BSA
65 XLacbBSA	Lac β -4AP-BSA	1	Lac β -4AP-BSA
66 XMannBSA	Man β -4AP-BSA	1	Man β -4AP-BSA
67 XGalbBSA	Gal β -ITC-BSA	1	Gal β -ITC-BSA
68 XylbBSA	Xyl β -4AP-BSA	1	Xyl β -4AP-BSA
69 XylabBSA	Xyl α -4AP-BSA	1	Xyl α -4AP-BSA
70 XGlcBSA	Glc β -4AP-BSA	1	Glc β -4AP-BSA
71 FucabBSA	Fuc α -4AP-BSA	1	Fuc α -4AP-BSA
72 FucbBSA	Fuc β -4AP-BSA	1	Fuc β -4AP-BSA
73 GlcblTCBSA	Glc β -ITC-BSA	1	Glc β -ITC-BSA
74 Galb4APBSA	Gal β -4AP-BSA	1	Gal β -4AP-BSA
75 Neu5Gc α -4APBSA	Neu5Gc α -4AP-BSA	1	Neu5Gc α -4AP-BSA
76 D-GlobTHSA	Globotriose-HSA	1	Gal- α -(1,4)-Gal- β -(1,4)-Glc-Sp3-BSA

Linker key
 Sp3 = 3 atom spacer
 Sp6 = 6 atom spacer
 Sp14 = 14 atom linker
 4AP = 4-aminophenyl
 APE = aminophenylethyl
 APD = acetylphenylenediamine

8 **Table S3.** List of gastrointestinal tract mucins and glycoproteins printed and printing conditions for
9 the gastrointestinal tract mucin microarray.

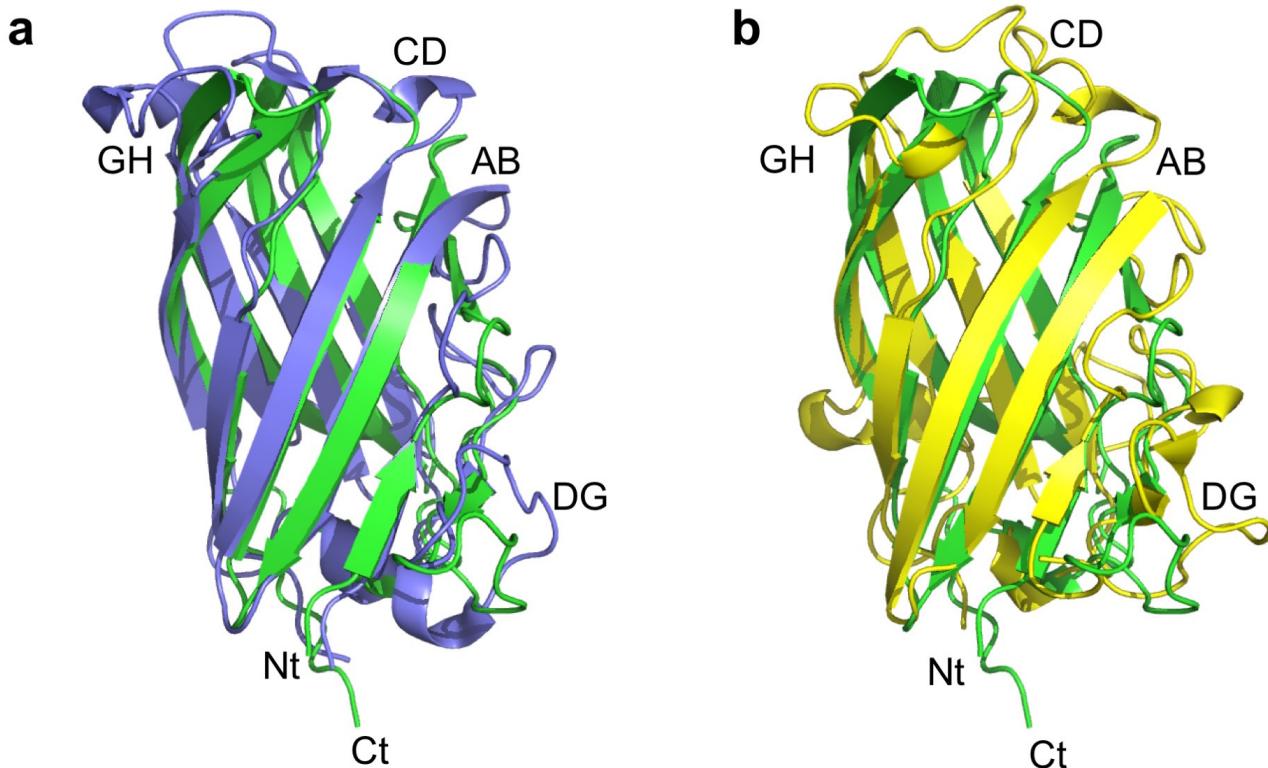
Number	Source	Print concentration (mg/mL)	Print buffer
1	Equine stomach	0.25	PBS 0.01% Tween 20
2	Ovine abomasum antrum	0.1	PBS 0.01% Tween 20
3	HT29MTXE12	0.5	PBS 0.025% Tween 20
4	Ovine descending colon	0.15	PBS 0.01% Tween 20
5	Ovine ileum	0.15	PBS 0.01% Tween 20
6	Ovine spiral colon	0.5	PBS
7	Chicken proximal small intestine	0.25	PBS 0.025% Tween 20
8	Ovine jejunum	0.5	PBS 0.01% Tween 20
9	Ovine duodenum	0.15	PBS 0.01% Tween 20
10	Porcine gastric mucin	0.33	PBS 0.01% Tween 20
11	Chicken large intestine	0.2	PBS 0.01% Tween 20
12	Equine duodenum	0.3	PBS 0.01% Tween 20
13	Deer jejunum	0.25	PBS 0.025% Tween 20
14	Deer spiral ascending colon	0.75	PBS 0.025% Tween 20
15	Bovine abomasum	0.25	PBS 0.01% Tween 20
16	Bovine duodenum	0.5	PBS 0.01% Tween 20
17	Equine small intestine	0.25	PBS
18	Equine left ventral colon	0.25	PBS 0.01% Tween 20
19	Bovine spiral colon	0.25	PBS 0.01% Tween 20
20	Deer duodenum	0.5	PBS 0.025% Tween 20
21	Equine right ventral colon	0.15	PBS 0.01% Tween 20
22	Equine dorsal colon	0.25	PBS 0.01% Tween 20
23	Deer abomasum	0.25	PBS 0.01% Tween 20
24	Chicken cecum	0.5	PBS 0.025% Tween 20
25	LS174T	0.5	PBS 0.01% Tween 20
26	Porcine descending colon	0.5	PBS 0.025% Tween 20
27	Porcine jejunum	0.25	PBS
28	Porcine spiral colon	0.6	PBS 0.025% Tween 20
29	Porcine stomach	0.5	PBS 0.025% Tween 20
30	Porcine ceca	0.5	PBS 0.025% Tween 20
31	Mouse large intestine	0.4	PBS 0.025% Tween 20
32	Mouse cecum	0.3	PBS 0.025% Tween 20
33	Mouse stomach	0.5	PBS 0.025% Tween 20
34	Mouse small intestine	0.25	PBS 0.025% Tween 20
35	Rat ileum	0.5	PBS 0.025% Tween 20
36	Rat duodenum and jejunum	0.5	PBS 0.025% Tween 20
37	Rat cecum	0.5	PBS 0.025% Tween 20
38	Rat stomach	0.5	PBS 0.025% Tween 20
39	PBS		
40	Asialofetuin	1	PBS 0.05% Tween 20
41	RNase B	1	PBS 0.05% Tween 20
42	Fetuin	1	PBS 0.05% Tween 20
43	Transferrin	1	PBS 0.05% Tween 20
44	Ovomucoid	0.5	PBS 0.05% Tween 20
45	Human a1-acid glycoprotein	1	PBS 0.05% Tween 20
46	PBS 0.025% Tween 20		

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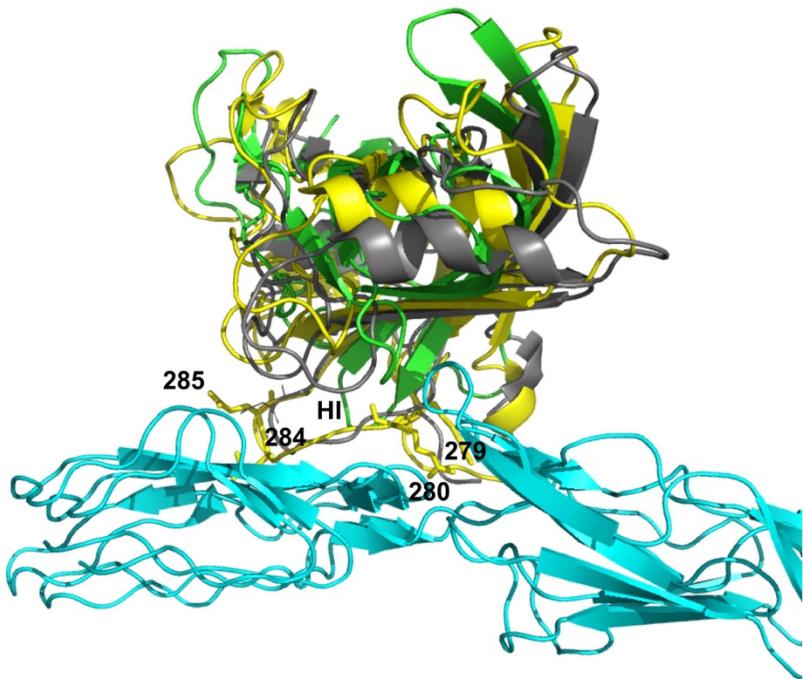
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12 **Table S4.** Inhibition percentage of binding in the presence of 100 mM GlcNAc. Ten percent
 13 inhibition or greater is highlighted by shaded cells.
 14

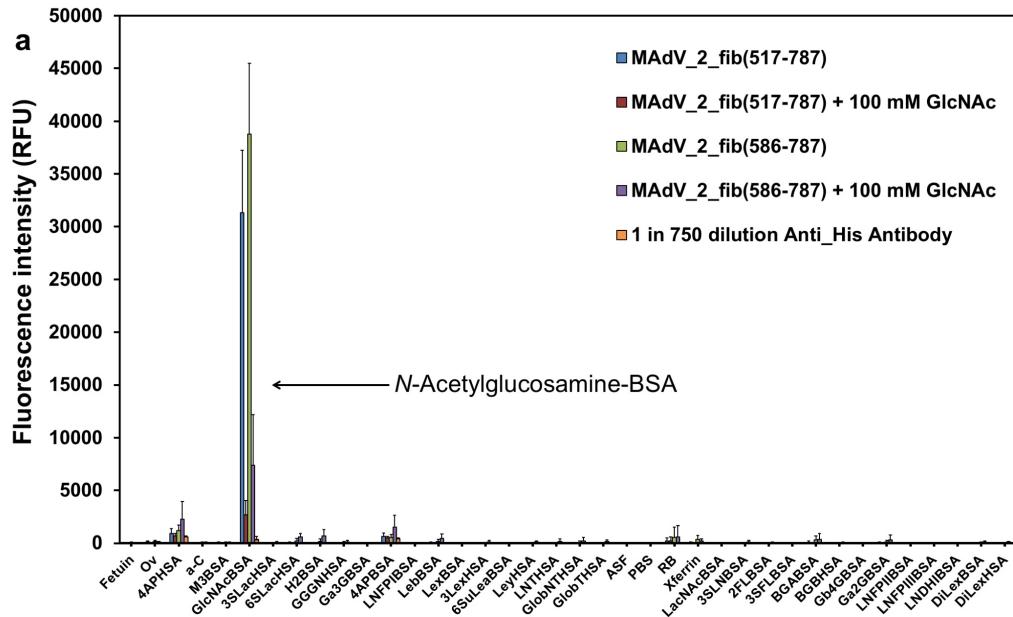
Binding probe	MAdV-2fib(517-787)	MAdV-2fib(586-787)	WGA
Equine stomach mucin	0.0	23	86
Ovine abomasum antrum mucin	1.8	19	93
HT29MTXE12 mucin	39.4	44	63
Ovine descending colon mucin	41.9	61	83
Ovine ileum mucin	19.2	34	55
Ovine spiral colon mucin	43.2	64	45
Chicken proximal small intestine mucin	83.9	85	0
Ovine jejunum mucin	10.8	48	66
Ovine duodenum mucin	60.0	75	22
Porcine gastric mucin	37.7	79	0
Chicken large intestine mucin	43.9	65	0
Equine duodenum mucin	43.1	75	67
Deer jejunum mucin	0.0	0	0
Deer spiral ascending colon mucin	0.0	0	55
Bovine abomasum mucin	9.4	40	90
Bovine duodenum mucin	40.4	27	64
Equine small intestine mucin	62.4	80	88
Equine left ventral colon mucin	52.1	50	90
Bovine spiral colon mucin	0.0	0	0
Deer duodenum mucin	0.0	29	0
Equine right ventral colon mucin	0.0	3	91
Equine dorsal colon mucin	0.0	7	91
Deer abomasum mucin	100.0	100	0
Chicken cecum mucin	66.6	99	0
LS174T mucin	71.4	83	94
Porcine descending colon mucin	76.8	82	54
Porcine jejunum mucin	0.0	0	0
Porcine spiral colon mucin	69.7	75	90
Porcine stomach mucin	30.9	59	85
Porcine cecum mucin	37.2	42	0
Mouse large intestine mucin	81.7	84	70
Mouse cecum mucin	94.0	93	95
Mouse stomach mucin	43.4	76	11
Mouse small intestine mucin	85.5	94	78
Rat ileum mucin	72.7	83	18
Rat duodenum & jejunum mucin	93.3	95	0
Rat cecum mucin	81.0	69	0
Rat stomach mucin	90.9	85	82
Asialofetuin	0.0	43	0
RNase B	95.2	96	49
Fetuin	0.0	37	49
Transferrin	0.0	0	0
Ovomucoid	97.1	96	41
Human α 1-acid glycoprotein	0.0	16	0



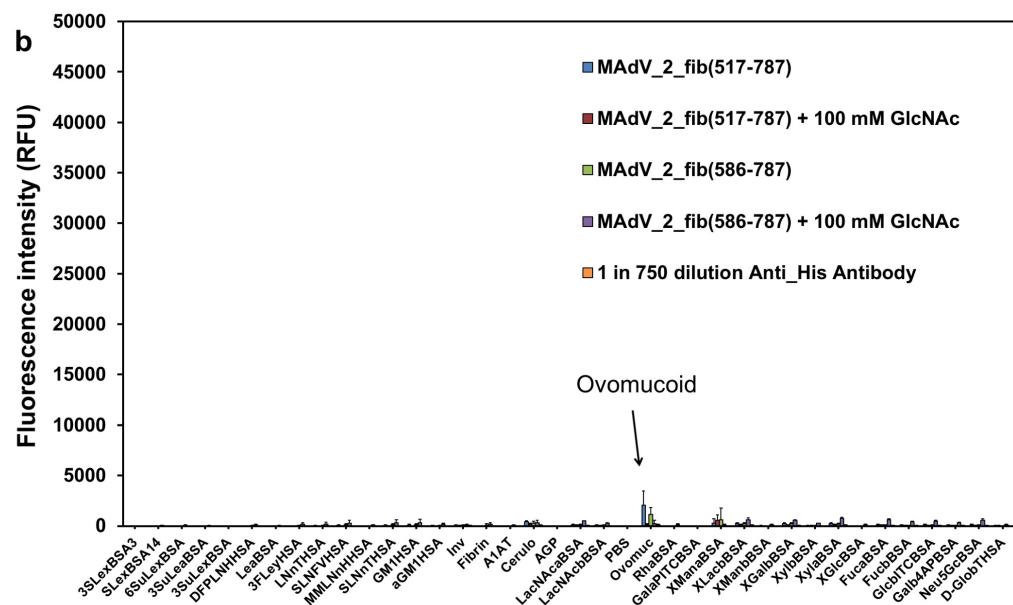
16
17 **Fig. S1.** Comparison of the MAdV-2, CAdV-2 and HAdV-5 fibre head structures. A. MAdV-2 fibre
18 head monomer (in green) superposed onto the CAdV-2 fibre head monomer (in blue). B. MAdV-2
19 fibre head monomer (in green) superposed onto the HAdV-5 fibre head monomer (in yellow). The
20 N-termini, C-termini and AB-, CD-, DG- and GH-loops are labelled.
21



50
51 **Fig. S3.** MAdV-2 fibre head is unlikely to bind CD46. Superposition of the MAdV-2 fibre head
52 domain (in green) onto the HAdV-11 and HAdV21 fibre head domains bound to CD46 (HAdV11:
53 PDB entry 3O8E, yellow; HAdV21: PDB entry 3I89; grey; CD46 in cyan). While the conformations
54 of the HI-loops of HAdV-11 and HAdV-21 are similar to each other; the HI-loop of MAdV-2 is much
55 shorter and apparently incompatible with CD46 binding. Arg279, Arg280, Asp284 and Glu285 of the
56 HAdV-11 are shown and labelled.
57

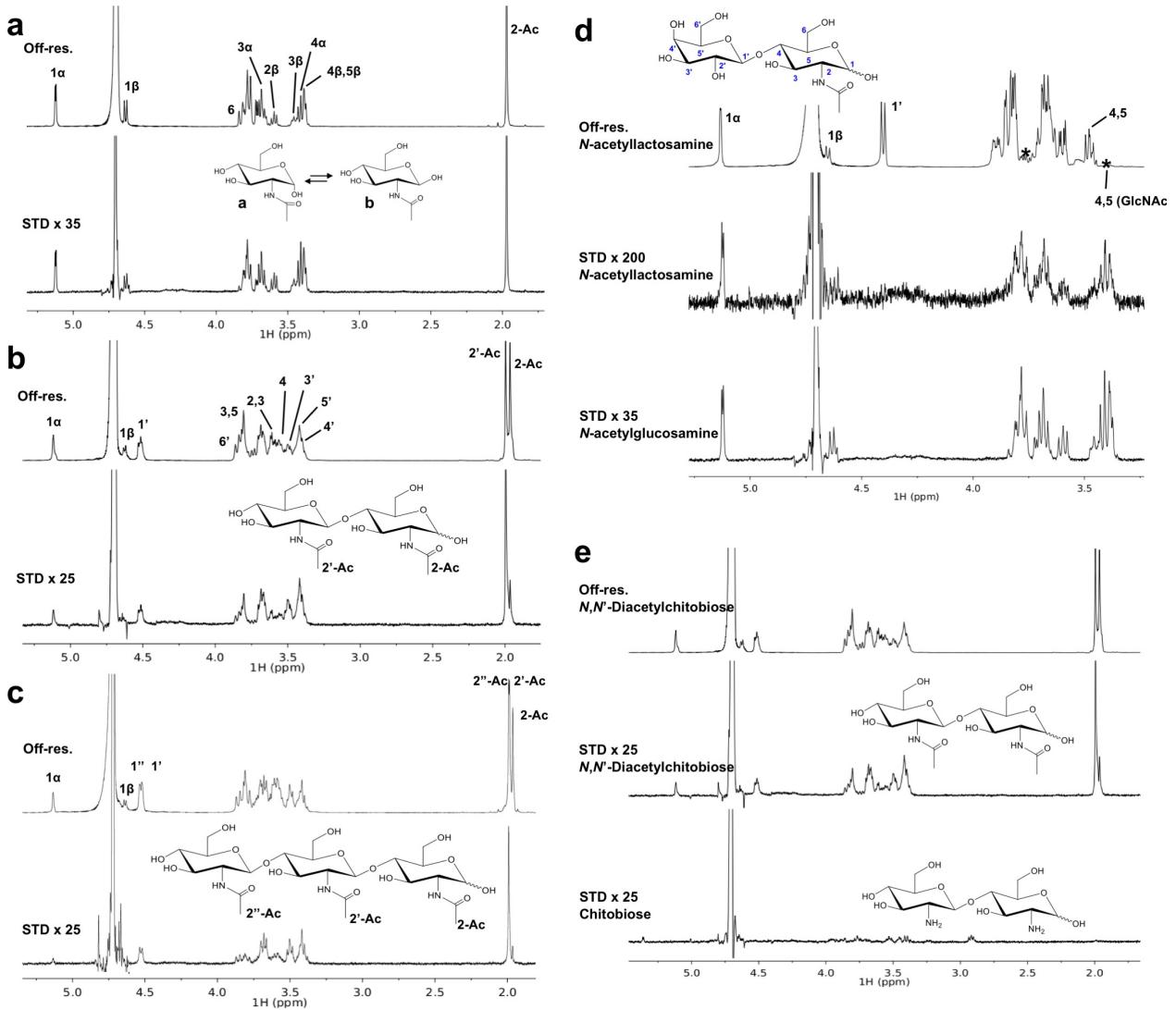


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59

60 **Fig. S4.** Binding profile of two isoforms of MAdV-2 fiber protein. Bar chart (divided in panels A and
61 B) representing the binding intensity of the long (MAdV-2-fib(517-787)) and short (MAdV-2-
62 fib(586-787)) forms of the MAdV-2 fiber to carbohydrates on a microarray surface. Binding was
63 detected using a fluorescently labelled anti-His antibody. The data represents the average of four
64 replicate experiments and the error bars depict one standard deviation of the mean calculated over
65 four microarray slides. The protein isoforms show comparable binding patterns with strong binding
66 to GlcNAc-BSA and lower binding to ovomucoid, both of which were inhibited by co-incubation
67 with 100 mM GlcNAc.

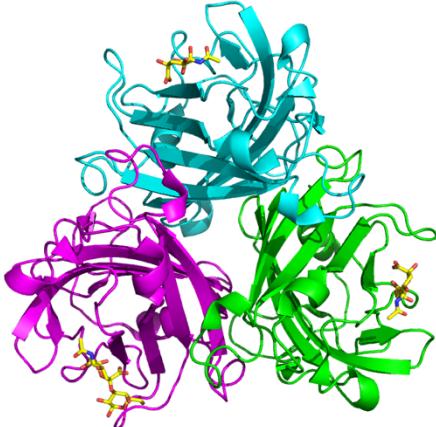


68
69

70 **Fig. S5.** A. *N*-acetyl-glucosamine binding to MAdV-2-fib(586-787) studied by STD-NMR. Top: Off-
71 resonance spectrum, with labels indicating the assignment for representative ligand signals. Bottom:
72 STD spectrum (up-scaled 35 \times). B. *N,N'*-diacetylchitobiose (GlcNAc- β -(1 \rightarrow 4)-GlcNAc) binding of
73 MAdV-2-fib(586-787). Top: Off-resonance spectrum, with labels indicating the assignment for
74 representative ligand signals. Bottom: STD spectrum (up-scaled 25 \times). C. Binding of *N,N',N''*-
75 triacetylchitotriose (GlcNAc- β -(1 \rightarrow 4)-GlcNAc- β -(1 \rightarrow 4)-GlcNAc) to MAdV-2-fib(586-787). Top
76 spectrum: Off-resonance (reference) spectrum. Bottom spectrum: STD spectra up-scaled 25 \times . D.
77 Effect of deacetylation on the binding of *N,N'*-diacetylchitobiose to MAdV-2-fib(586-787). Top
78 spectrum: Off-resonance spectrum of *N,N'*-diacetylchitobiose. Middle spectrum: STD spectrum of
79 *N,N'*-diacetylchitobiose. Bottom spectrum: STD spectrum of chitobiose. E. STD-NMR experiment
80 performed on *N*-acetyllactosamine (*Gal*- β -(1 \rightarrow 4)-GlcNAc), showing that the only species being
81 recognised in the sample are trace amounts of GlcNAc. Top spectrum: Off-resonance spectrum of *N*-
82 acetyllactosamine; middle spectrum: STD spectrum of the *N*-acetyllactosamine sample; bottom
83 spectrum: STD spectrum of GlcNAc.

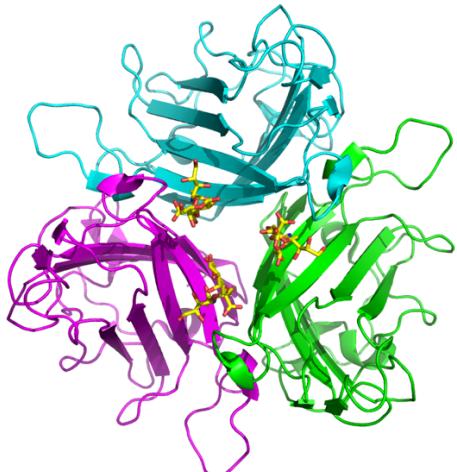
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a**b**

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c**d**

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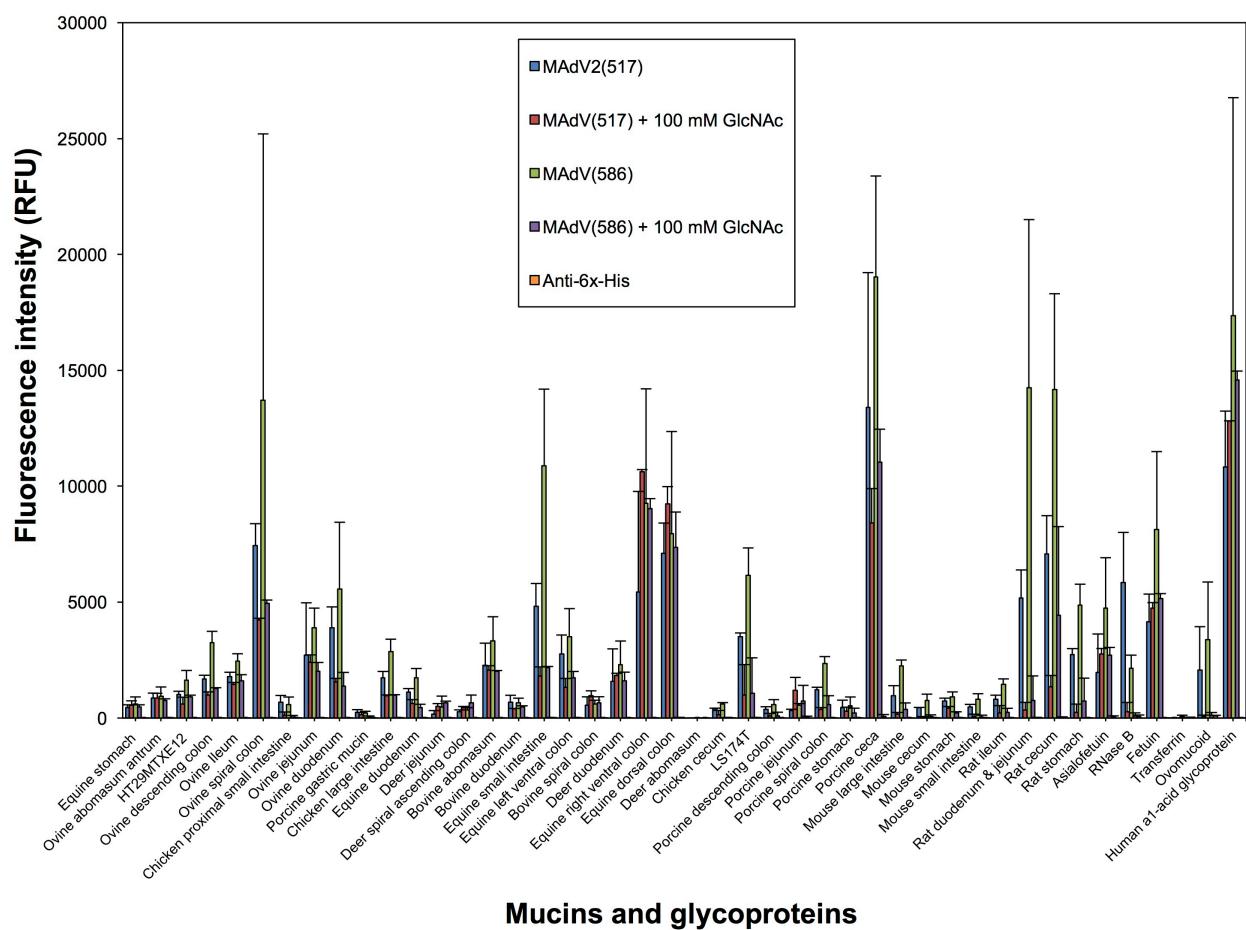
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Fig. S6. Comparison of the ligand binding sites of MAdV-2 fibre head bound to GlcNAc (PDB entry 5NC1; panel A), CAdV-2 fibre head bound to sialyllactose (PDB entry 2WBV; panel B), HAdV37 fibre head bound to sialyllactose (PDB entry 1UXA; panel C) and HAdV-52 fibre head bound to trisialic acid (PDB entry 6G47; panel D). Top views with the trimers in comparable orientations are shown, i.e. from the viral-distal end of the fibre.



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96 **Fig. S7.** MAdV-2 fibre binding to gastrointestinal tract mucins. Bar chart representing the binding
97 intensity of the MAdV-2fib(586-787) and MAdV-2fib(517-787) fibre proteins to mucins and
98 glycoproteins on a mucin microarray in the presence and absence of 100 mM GlcNAc. Binding was
99 detected using a fluorescently labelled anti-His antibody. The data for the uninhibited proteins
100 represents the mean of three technical replicate microarray slides and the data for the inhibited
101 proteins represent the mean of two technical replicates. Error bars represent one standard deviation
102 of the mean.