SUPPLEMENTAL MATERIAL

Supplementary Figures 1-3 and Supplementary Tables 1-5 for manuscript:

Novel Genes and Metabolite Trends in *Bifidobacterium longum* subsp. *infantis* Bi-26 Metabolism of Human Milk Oligosaccharide 2'-fucosyllactose

Bryan Zabel¹, Christian Clement Yde,² Paige Roos,³ Jørn Marcussen,² Henrik Max Jensen,² Krista Salli⁴, Johanna Hirvonen⁴, Arthur C. Ouwehand⁴, Wesley Morovic¹

¹Genomics & Microbiome Science, DuPont Nutrition & Health, Madison, WI, USA ²Advanced Analysis, DuPont Nutrition Biosciences ApS, Brabrand, Aarhus, Denmark

³Genomics Lab, DuPont Pioneer, Johnston, IA, USA

⁴Global Health and Nutrition Science, DuPont Nutrition and Health, Kantvik, Finland

	B. longum subsp. Infantis strain	B. longum subsp. Infantis strain
General genome feature	Bi-26	ATCC15697
Size	2,569,657	2,832,748
GC content	59.4%	59.9%
Predicted coding density	84.7%	87.3%
number of subsystems	286	284
number of coding sequences	2280	2637
number of RNAs	60	91
Subsystem features		
Amino Acids and Derivatives	201	201
Carbohydrates	146	145
Cell Division and Cell Cycle	19	19
Cell Wall and Capsule	61	56
Cofactors, Vitamins, Prosthetic Groups,		
Pigments	103	104
DNA Metabolism	63	57
Dormancy and Sporulation	2	2
Fatty Acids, Lipids, and Isoprenoids	35	33
Iron acquisition and metabolism	1	5
Membrane Transport	35	34
Matsholism of Aromatic Compounds	3	3
Plant	8	8
r lain Miscellaneous	8	8
Motility and Chemotavis	+ 2	4
Nitrogen Metabolism	2 7	0 7
Nucleosides and Nucleotides	, 57	56
Phages Prophages Transposable	51	50
elements. Plasmids	8	13
Phosphorus Metabolism	27	27
Potassium metabolism	4	4
Protein Metabolism	192	201
Regulation and Cell signaling	16	16
Respiration	2	2
RNA Metabolism	58	58
Secondary Metabolism	2	0
Stress Response	31	32
Sulfur Metabolism	9	9
Virulence, Disease and Defense	32	33

Supplementary Table 1: General genome feature comparison of *Bifidobacterium longum* subsp. *infantis* Bi-26 to *Bifidobacterium longum* subsp. *infantis* strain ATCC15697. Genomic assembly statistics along with subsystems (annotated from RAST) are displayed.

			Presence in B. longum subsp. infantis genomes								
Leloir pathway genes	Gene name	Enzyme	Bi-26	ATCC15697	BT1	Bifido_12	TPY12-1	IN-07	Bifido_04	IN-F29	EK1
galactokinase	galK	EC 2.7.7.6	+	+	+	+	+	+	+	+	+
UDP-glucose 4-epimerase	galE	EC 5.1.3.2	+	+	+	+	+	+	+	+	+
galactose-1-phosphate uridylyltransferase	galT	EC 2.7.7.10	+	+	+	+	+	+	+	+	+
phosphoglucomutase	pgm	EC 5.4.2.2	+	+	+	+	+	+	+	+	+
Main Fermentative pathway genes											
glucokinase glkA Blon_0565	glkA	EC 2.7.1.63	+	+	+	+	+	+	+	+	+
glucose-6-phosphate isomerase	gpi	EC 5.3.1.9	+	+	+	+	+	+	+	+	+
fructose-6-phosphate phosphoketolase	xfp	EC 2.6.1.16	+	+	+	+	+	+	+	+	+
transaldolase	tal	EC 2.2.1.2	+	+	+	+	+	+	+	+	+
transketolase	tkt	EC 2.2.1.1	+	+	+	+	+	+	+	+	+
ribose 5-phosphate isomerase	rpiA	EC 5.3.1.6	+	+	+	+	+	+	+	+	+
Ribulose-phosphate 3-epimerase	rpe	EC 5.1.3.1	+	+	+	+	+	+	+	+	+
xylulose-5-phosphate phosphoketolase	xfp	EC 4.1.2.9	+	+	+	+	+	+	+	+	+
glyceraldehyde-3-phosphate dehydrogenase	gap	EC 1.2.1.12	+	+	+	+	+	+	+	+	+
phosphoglycerate kinase	pgk	EC 2.7.2.3	+	+	+	+	+	+	+	+	+
phosphoglycerate mutase	pmg	EC 5.4.2.1	+	+	+	+	+	+	+	+	+
enolase	eno	EC 4.2.1.11	+	+	+	+	+	+	+	+	+
pyruvate kinase	pyk	EC 2.7.1.40	+	+	+	+	+	+	+	+	+
lactate dehydrogenase	idh	EC 1.1.1.27	+	+	+	+	+	+	+	+	+
Fucose genes											
Alpha-L-fucosidase	fucA	EC 3.2.1.51	+	+	+	+	+	+	+	+	+
Fucose permease	fucP	NA	+	+	+	+	+	+	+	+	+
	274	EC									
dehydrogenase clustered with L-fuconate utilization genes	NA	1.1.1.122	+	+	+	+	+	+	+	+	+
L-fuconate dehydratase		EC 4.2.1.68	+	+	+	+	+	+	+	+	+
L-fucose mutarotase	fucU	EC 5.1.3.29	+	+	+	+	+	+	+	+	+
lactaldehyde reductase	fucO	EC 1.1.1.77	+	+	+	+	+	+	+	+	+
triose-phosphate isomerase	tpiA	EC 5.3.1.1	+	+	+	+	+	+	+	+	+
Acetate gene											
acetate kinase	ackA	EC 2.7.2.1	+	+	+	+	+	+	+	+	+
Formate gene											
pyruvate formate-lyase	pfl	EC 2.3.1.54	+	+	+	+	+	+	+	+	+

Supplementary Table 2: Annotated genes related to 2'-FL utilization in Bi-26 and other *B. longum* subsp. *infantis* strains. Used in conjunction with Figure 1 to describe genes needed to make quantified metabolites. Included are gene name, KEGG enzyme class and pathway the gene is located in. Presence in other *B. longum* subsp. *infantis* strains is noted. Presence of genes was determined by searching for enzyme class number or annotations using Geneious. The strains selected were those that clustered with the type strain ATCC15697 in the National Center for Biotechnology Information (NCBI) *B. longum* dendrogram located at https://www.ncbi.nlm.nih.gov/genome/1833? Accessed 04/21/2019.



Supplementary Figure 1: Alignments of ATCC15697 and Bi-26 HMO cluster using Geneious. Genes are colored by their functionality. The middle black section of the figure shows the mismatches in translation. Gaps in Bi-26 for the IS element, transport proteins and secreted protein are where the genes are not present.



Supplementary Figure 2: Volcano plots of differential expression compared to absolute confidence for 2'-FL RNA-seq values compared to the other phases of growth on 2'-FL. Only values with a Log2 ratio above 1 and an absolute confidence above 1.30 (p-value 0.05) were considered significant.

A)	Phase(s)	Fucose related		Hypothetical		Trar	nsport relate	d Central n	Central metabolism			
'	All 3	9		14			29		31			
	Early only	0		8		4			13			
	Mid-log only	0		7		0			0			
	Late-log only	0		15		7			28			
	Early and mid	0		2		1			3		6	
	Early and late	0		1	16		4		19		47	
	Mid and late	0	0		1		1		2		4	
B)	Compar	Comparison		Early vs mid			Early	/s late	late Mid		vs late	
,	Phase		Ea	ırly	Mid-l	og	Early	Late	Mid-log	La	te	
	Fucose re	elated		0	0		0	0	0		3	
	Hypothe	Hypothetical		0	0		29	11	24	1	7	
	Transport	Transport related		1	0		26	16	28	2	3	
	Central Met	Central Metabolism		0	0		25	18	14	1	2	
	Othe	Other		0	0		14	15	7	1	2	
	Total			1	0		94	60	73	6	7	

Supplementary Table 3: (A) Comparison of genes functions upregulated in 2'-FL fermentation and at what part of the fermentation they were upregulated in. (B) Comparing 2'-FL early vs mid and late and 2'-FL mid vs late for gene differences. To look at the differences in major systems throughout fermentation.

Genes/ Functions	Log2 Ratio early	p-value early	Log2 Ratio mid	p-value mid	Log2 Ratio late	p-value late
Lactose Gene						
Beta-galactosidase (EC 3.2.1.23) CDS HMO cluster	1.76363239	2.23E-42	1.453036987	1.11E-05	1.521517602	2.05E-14
Leloir pathway genes						
Galactokinase (EC 2.7.1.6) CDS	0.126435896	0.288887344	0.750285825	0.062522351	0.544604571	3.85E-05
UDP-glucose 4-epimerase (EC 5.1.3.2) CDS	0.810049707	1.89E-10	0.45840775	0.145733665	0.372488235	0.005699352
Galactose-1-phosphate uridylyltransferase (EC 2.7.7.10) CDS	1.099644411	8.78E-21	0.823699827	0.030696562	0.687430616	4.70E-08
Phosphoglucomutase (EC 5.4.2.2) CDS	-0.576976916	9.06E-06	0.181997756	0.567249427	0.11699882	0.3739832
Main fermentation genes						
Polyphosphate glucokinase (EC 2.7.1.63) CDS	0.655557986	5.34E-08	0.762651769	0.047869446	0.860681059	2.22924E-09
Glucose-6-phosphate isomerase (EC 5.3.1.9) CDS	0.901957471	6.90E-13	0.933528033	0.041092786	0.385425846	0.008517778
Fructose-6-phosphate phosphoketolase (EC 4.1.2.22) CDS	0.391267047	0.001118049	0.670967696	0.114195627	0.30370557	0.017148752
Transaldolase (EC 2.2.1.2) CDS	0.554885296	2.25E-06	0.756569661	0.08854475	-0.023665187	0.850368066
Transketolase (EC 2.2.1.1) CDS	0.191512513	0.14128884	0.488204592	0.182984772	0.220792855	0.102939386
Ribose 5-phosphate isomerase A (EC 5.3.1.6) CDS	0.383782011	0.006770406	0.861195531	0.050840225	0.572597746	7.65931E-05
Ribulose-phosphate 3-epimerase (EC 5.1.3.1) CDS	0.413140195	0.001308143	0.205487583	0.555270292	0.113840979	0.37594078
NAD-dependent glyceraldehyde-3-phosphate dehydrogenase (EC 1.2.1.12) CDS	0.365239397	0.003791971	0.577401413	0.116716001	0.122090122	0.308817639
Phosphoglycerate kinase (EC 2.7.2.3) CDS	0.47453399	4.62E-05	0.594644044	0.089754822	0.444910069	0.001448169
Phosphoglycerate mutase (EC 5.4.2.1) CDS	0.287742191	0.034010109	0.319510881	0.337332752	0.140411506	0.307780837
Enolase (EC 4.2.1.11) CDS	0.412183735	0.000482006	0.67203813	0.112037536	0.072738769	0.58552877
Pyruvate kinase (EC 2.7.1.40) CDS	0.155276824	0.194011771	0.1794255	0.575430108	-0.03776668	0.796051926
L-lactate dehydrogenase (EC 1.1.1.27) CDS	0.651064527	7.94E-07	0.596209199	0.130691092	-0.102965481	0.468593789
Fucose genes						
Alpha-L-fucosidase (EC 3.2.1.51) CDS Non hmo cluster	3.687712028	6.66E-27	2.157112872	8.01E-07	4.281550579	9.41698E-29
Alpha-L-fucosidase (EC 3.2.1.51) CDS HMO cluster	1.32547242	2.89E-24	1.453024153	0.001622438	1.572076226	7.27592E-27
dehydrogenase clustered with L-fuconate utilization genes CDS	8.708998093	0	3.625277007	1.1182E-12	9.163268073	0
dehydrogenase clustered with L-fuconate utilization genes CDS	1.505194366	1.34E-33	1.67300627	0.000363805	2.013244319	4.81E-47
L-fuconate dehydratase (EC 4.2.1.68) CDS	7.118010773	0	4.00346757	2.14E-28	7.302836379	0
L-fuconate dehydratase (EC 4.2.1.68) CDS HMO cluster	6.113032129	2.20E-283	1.692842419	0.000300618	2.048002117	1.36E-48
L-fucose mutarotase CDS	6.113032129	2.20E-283	3.541064248	6.06E-27	6.546959726	5.27E-136
L-fucose mutarotase CDS HMO cluster	1.436944157	3.46E-22	1.597622702	0.000962691	1.701493459	1.47E-21
Lactaldehyde reductase (EC 1.1.1.77) CDS	1.439913964	2.10E-27	1.849089764	1.28E-08	2.530493203	4.72E-40
Triosephosphate isomerase (EC 5.3.1.1) CDS	0.603615877	3.19E-06	0.523593576	0.108366676	0.748135852	4.60E-09
Acetate gene						
Acetate kinase (EC 2.7.2.1) CDS	0.393547168	0.000835347	0.731456777	0.101646728	0.29486821	0.026206859
Formate gene		0.005.55				
PVrUVate tormate-IVase (F(2) + 1.54) (1)S	1 497771613	3 096-35	1 1 2 / 364895	0.000408604	1 473781291	

Supplementary Table 4: RNA-seq results of genes of interest in 2'-FL metabolism. Genes from Supplementary Table 2 log2 expression and p-value results from the early, mid-log and late phases are shown. Results are significant with a log2 of greater than 1 for 2'-FL samples and less than -1 for the lactose control samples with a p-value of less than 0.05. Green results are showing higher expression in 2'-FL samples while orange, yellow and red show higher expression in lactose samples or no change in expression.

Metabolite	Carb S	ource (mM)	Lactic a	cid (mM)	Acet	ate (mM)	Formate (mM)		Pyruv	Pyruvate (mM)		1,2-PD (mM)		Fucose (mM)	
Phase	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	
Initial	15.40	12.33	2.06	1.97	0.67	0.63	0.16	0.17	ND	ND	0.15	ND	0.49	ND	
Early	14.23	8.43	2.59	2.49	3.58	1.61	1.01	0.27	ND	ND	0.48	ND	0.72	ND	
Mid-log	12.65	7.27	4.48	3.59	8.43	2.81	1.73	0.39	ND	ND	1.27	ND	0.92	ND	
Late-log	4.03	ND	26.57	37.17	45.39	54.06	2.61	ND	4.23	ND	9.94	ND	1.34	ND	
	•		Rate of production												
Metabolite	Carb So	urce (mM)	Lactic a	cid (mM)	Acetate (mM)		Formate (mM)		Pyruvate (mM)		1,2-PD (mM)		Fucose (mM)		
Phase	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	
I→E	-0.17*	-0.56	0.08	0.07	0.42*	0.14	0.12*	0.01	NA NA		0.05*	NA	0.03*	NA	
E→M	-0.79*	-0.58	0.95*	0.55	2.43*	0.60	0.36*	0.06	NA NA		0.40*	NA	0.10*	NA	
M→L	-0.57*	NA	1.47*	2.24	2.46	3.42	0.06*	NA	NA	NA	0.58*	NA	0.03*	NA	
						F	ate of pr	oduction per	mM of ca	arbon source					
Metabolite	Carb com	sumed (mM)	Lactic a	icid (mM)	Acetate (mM)		Formate (mM)		Pyruvate (mM)		1,2-PD (mM)		Fucose (mM)		
Phase	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	2'FL	Lactose	
I→E	1.17*	3.9	0.45*	0.13	2.49*	0.25	0.73*	0.03	NA	NA	0.28*	NA	0.20*	NA	
E→M	1.58	1.16	1.20	0.95	3.07*	1.03	0.46*	0.10	NA	NA	0.50*	NA	0.13*	NA	
M→L	8.62	7.29	2.56*	4.61	4.29*	7.03	0.10*	NA	0.49*	NA	1.01*	NA	0.05*	NA	

*Significant difference compared to control

Supplementary Table 5: Overall metabolite results of each of the tested metabolites along with the carbon source are listed in the top table in mM values. The tables below contain the rates of production or consumption of each of the tested compounds. The middle table is valued with production over time while the lower table is valued at production per mM of carbon source consumed. Phases are labeled I=initial, E=early, M=mid-log, and L=late-log phases for the last two tables.



Supplementary Figure 3: Representative ¹H NMR spectra of a 24h 2'-FL fermentation sample. Selected quantified metabolites present at 24h 2'-FL fermentation have been labeled.