

Supplementary information for:

Conserved residues control the T1R3-specific allosteric signaling pathway of the mammalian sweet taste receptor

Jean-Baptiste Chéron¹, Amanda Soohoo^{2,3}, Yi Wang^{2,4}, Jérôme Golebiowski¹, Serge Antonczak¹, Peihua Jiang^{2*}, Sébastien Fiorucci^{1*}

¹ Université Côte d'Azur, CNRS, Institut de Chimie de Nice UMR7272, 06108 Nice, France

² Monell Chemical Senses Center, 3500 Market Street, Philadelphia, PA, 19104, United States

³ Massachusetts General Hospital, Harvard Medical School, Boston, MA, 02114, United States

⁴ Department of Ecology and Hubei Key Laboratory of Cell Homeostasis, College of Life Sciences, Wuhan University, Wuhan, 430072, China

*Corresponding authors: Sebastien.Fiorucci@unice.fr, pjiang@monell.org

Content:

Figure s1: Trajectory analysis of the cyclamate entering the T1R3 transmembrane domain.....	2
Table s1: Structure-function relationships of residues involved in the cyclamate-specific signaling pathway.....	3
Table s2: Summary of molecular dynamics simulations.....	4
Figure s2: Trajectory analysis of the molecular dynamics simulations.....	5
Figure s3: Representative structures of wt-ago-s2 MD simulation.	6
Figure s4: Structural analysis of ligand-receptor interactions.	7
Figure s5: Structural analysis of key residues involved in the T1R3 molecular switch.	8
Figure s6: Reduced sensitivity to lactisole of sweet taste receptor R725E and R725A mutants.....	9
Figure s7: Dose-response curves for sweet taste receptor W775F mutant.	10
Table s3: Meta-analysis of T1R3 TMD site-directed mutagenesis data for allosteric and orthosteric agonists.....	11
Figure s8: Multiple sequence alignment of mammalian T1R3 transmembrane domains.	13
Figure s9: Comparison of allosteric binding pocket of class C GPCR.	14
References.....	15

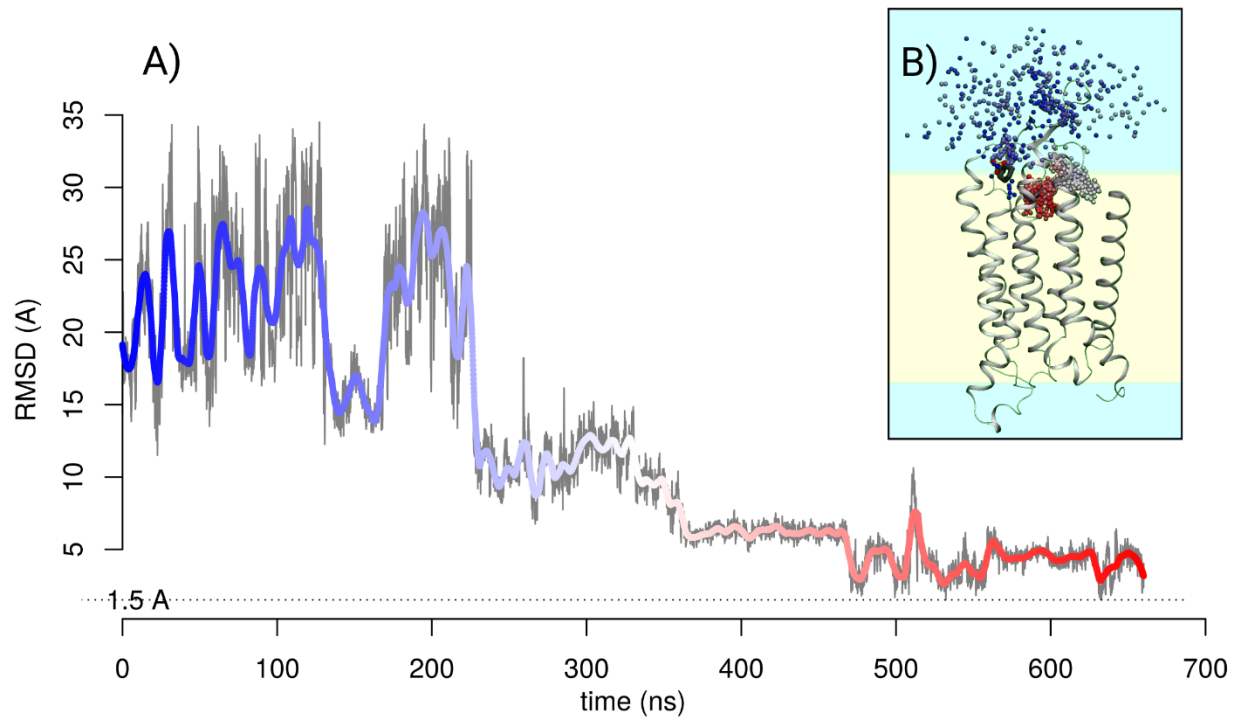


Figure s1: Trajectory analysis of the cyclamate entering the T1R3 transmembrane domain.

A) Root mean square deviation of the cyclamate molecule along the molecular dynamics simulations with respect to the docked position. B) View of the ligand entrance pathway from the bulk phase (in blue) to the binding cavity (in red).

Table s1: Structure-function relationships of residues involved in the cyclamate-specific signaling pathway.

Sequence conservation (less conserved residues in bold) has been calculated according to multiple sequence alignment of 41 mammalian organisms (Figure s8). Residue location is mentioned considering that the upper part of the binding site is oriented toward the extracellular medium.

Residue	Ballesteros Weinstein numbering	Conservation	Location in the allosteric binding site	Function
F624	2x56	97%	Upper part	cyclamate binding
Q636	3x32	97%	Upper part	cyclamate binding
Q637	3x33	94%	Upper part	cyclamate binding
S640	3x36	38%	Upper part	cyclamate binding
H641	3x37	100%	Upper part	cyclamate binding
L644	3x40	97%	Lower part	-
T645	3x41	97%	Lower part	-
L648	3x44	100%	Lower part	-
H721	ecl2	94%	Upper part	cyclamate binding
R723	ecl2	68%	Upper part	cyclamate binding
T724	ecl2	35%	Upper part	cyclamate binding
R725	ecl2	59%	Upper part	cyclamate binding
S726	ecl2	97%	Upper part	cyclamate binding
S729	5x39	88%	Upper part	cyclamate binding
F730	5x40	79%	Upper part	cyclamate binding
H734	5x44	100%	Lower part	-
N737	5x47	100%	Lower part	molecular switch
Y771	6x46	97%	Lower part	molecular switch
W775	6x50	100%	Lower part	molecular switch
F778	6x53	94%	Upper part	cyclamate binding
L782	6x57	62%	Upper part	cyclamate binding
R790	7x28	38%	Upper part	cyclamate binding
Q794	7x32	97%	Upper part	cyclamate binding
C801	7x39	100%	Lower part	-

Table s2: Summary of molecular dynamics simulations.

Molecular Dynamics (MD) simulation lengths and average Root Mean Square Deviations (RMSD) of wild-type (wt) and mutant (mut) protein in the presence (ago) or not (apo) of cyclamate. Two independent simulations (s1 and s2) have been performed for the apo and ago systems. The RMSD is computed using the backbone atoms of the protein and the heavy atoms of the cyclamate molecule after a structural superimposition of the receptor backbone atoms to the first frame of the MD trajectory.

		Simulation Time (μs)	Protein RMSD (\AA)	Ligand RMSD (\AA)
wt-ago	s1	4.12	2.0	3.4
	s2	3.76	2.9	3.8
wt-apo	s1	4.01	1.9	-
	s2	2.02	1.9	-
mut-ago	R725A	1.52	1.7	6
	W775H	1.00	1.9	4.7

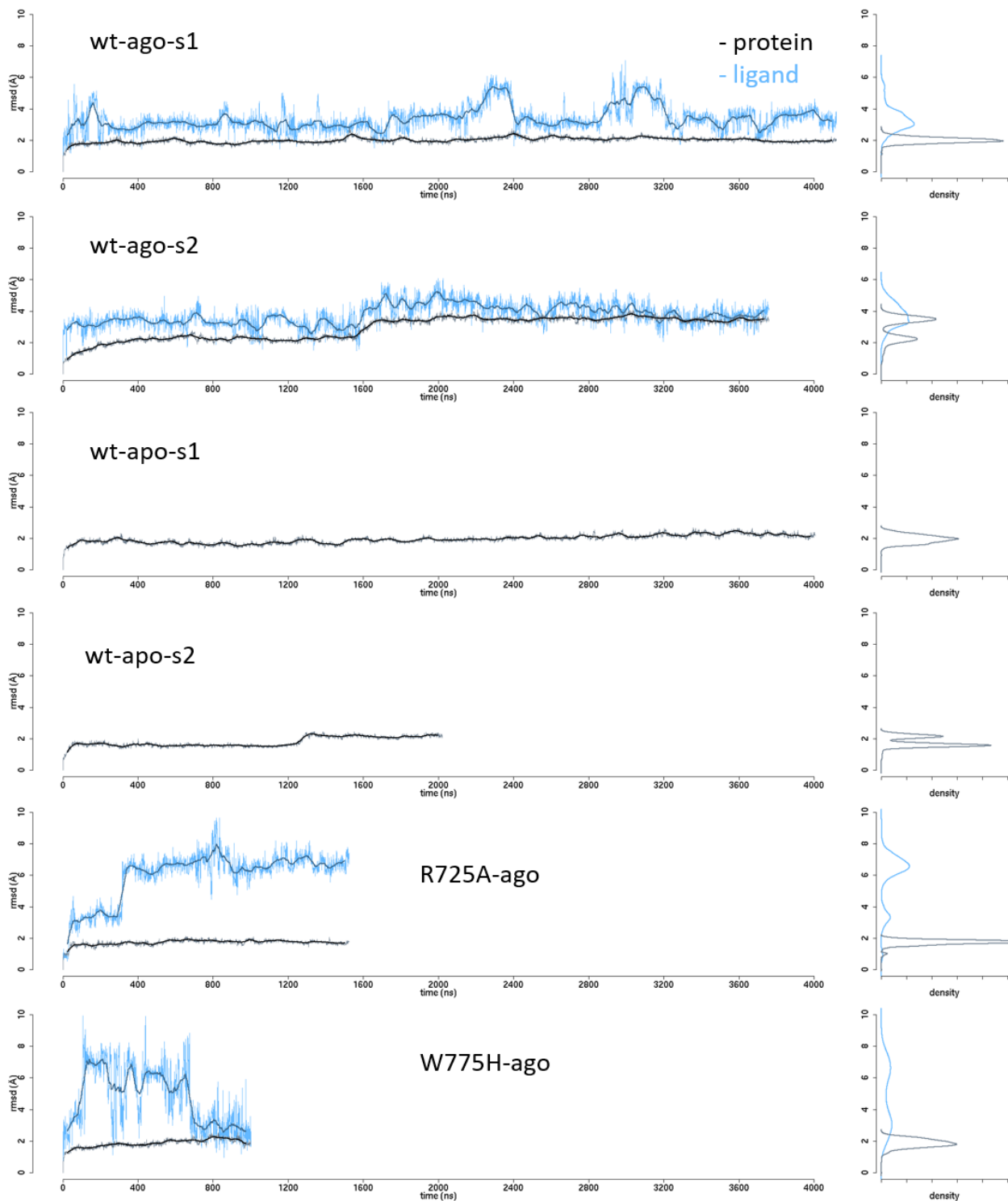


Figure s2: Trajectory analysis of the molecular dynamics simulations.

Time series plots and density distribution of protein (black) and ligand (blue) RMSD in multiple independent MD runs. Despite the high RMSD values, cyclamate remains within the T1R3 allosteric pocket all along the MD trajectories. It adopts alternate conformations and interacts with different charged or polar residues (Figure s4). See Table s2 for acronym details.

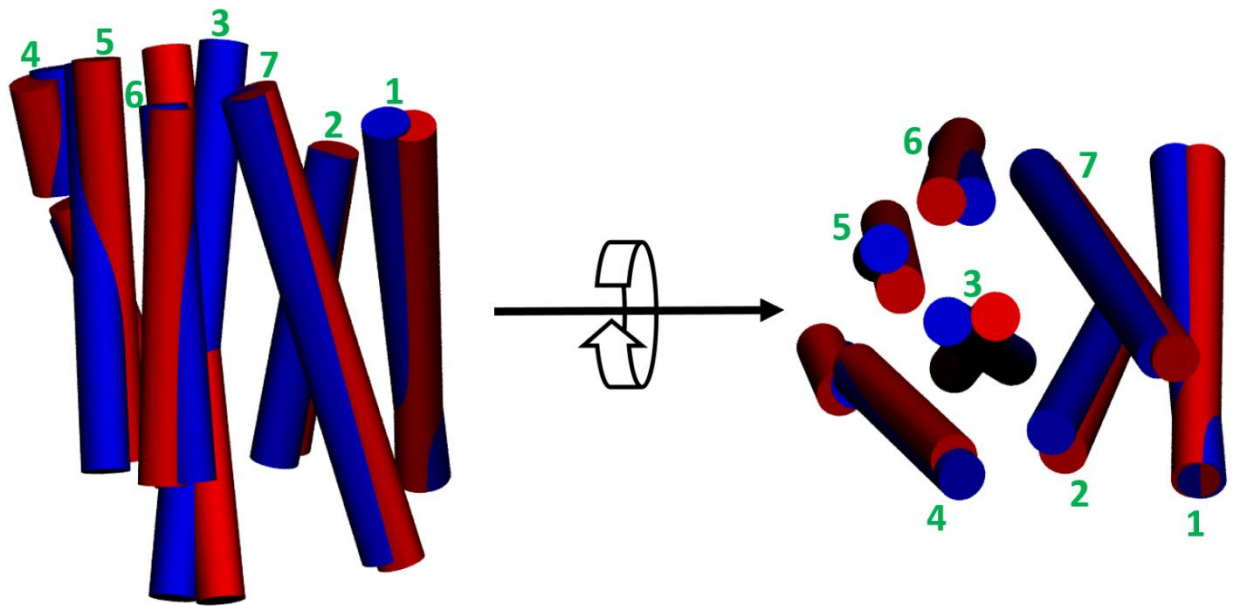


Figure s3: Representative structures of wt-ago-s2 MD simulation.

Representative structures of the wild-type receptor bound to cyclamate in the s2 MD simulation. Blue and red structures correspond to the first and the second part of the MD trajectory respectively. Major differences involve TM3, 5 and 6 of the receptor. They may correspond to conformational changes between inactive and active-like states.

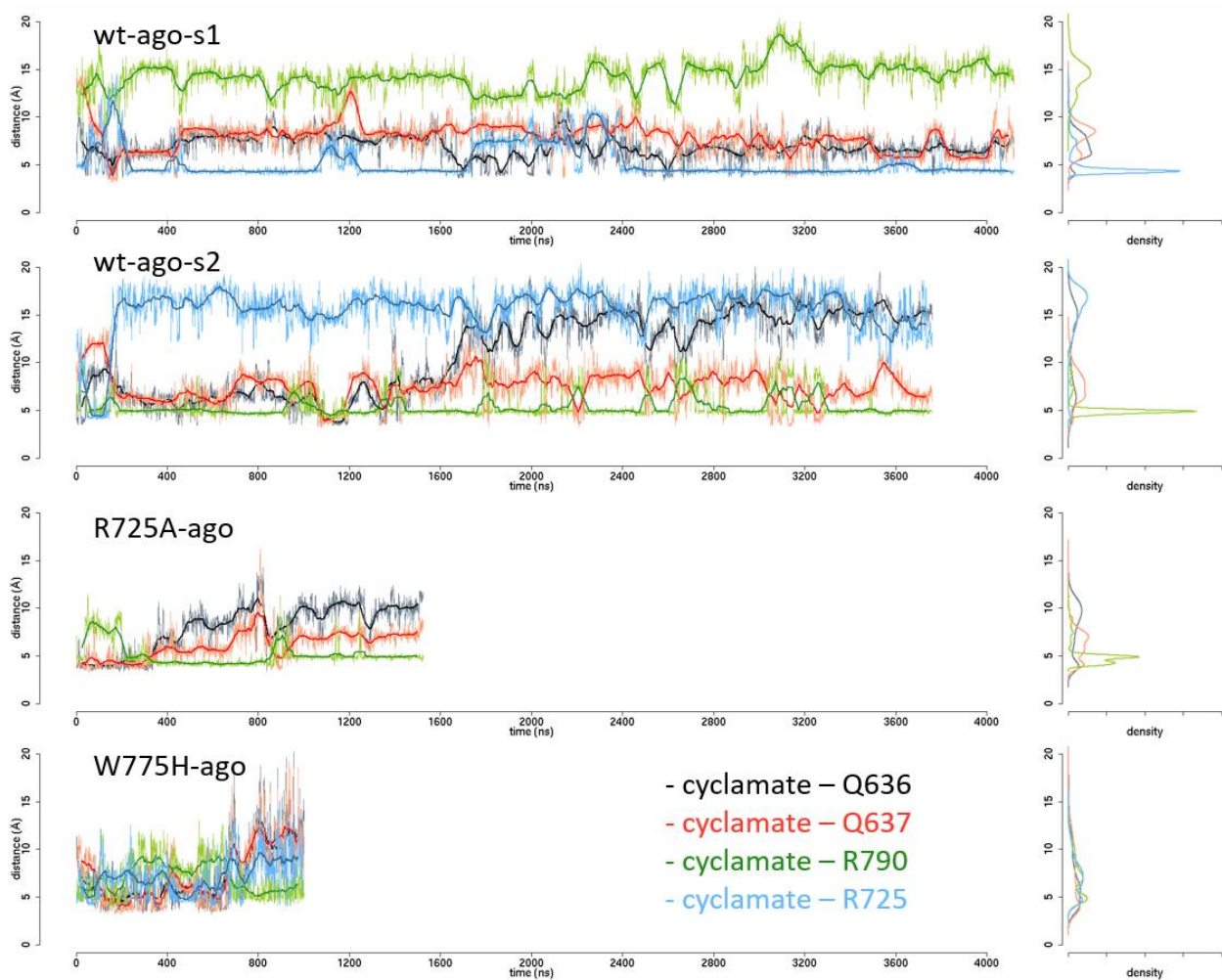


Figure s4: Structural analysis of ligand-receptor interactions.

Time series plots and density distributions of the ligand-receptor distances in *wt-ago* and *mut-ago* MD simulations. To avoid bias in the analysis due to the rotation of the cyclamate sulfamic acid functional group, the sulfur atom has been considered to calculate the ligand-receptor distance. For the same reason, the C^ε and N^{ε2} atoms have been considered for glutamine (Q636 and Q637) and arginine (R790 and R725) residues respectively. A distance in the range of 4.5 to 5 Å corresponds to a hydrogen bond interaction.

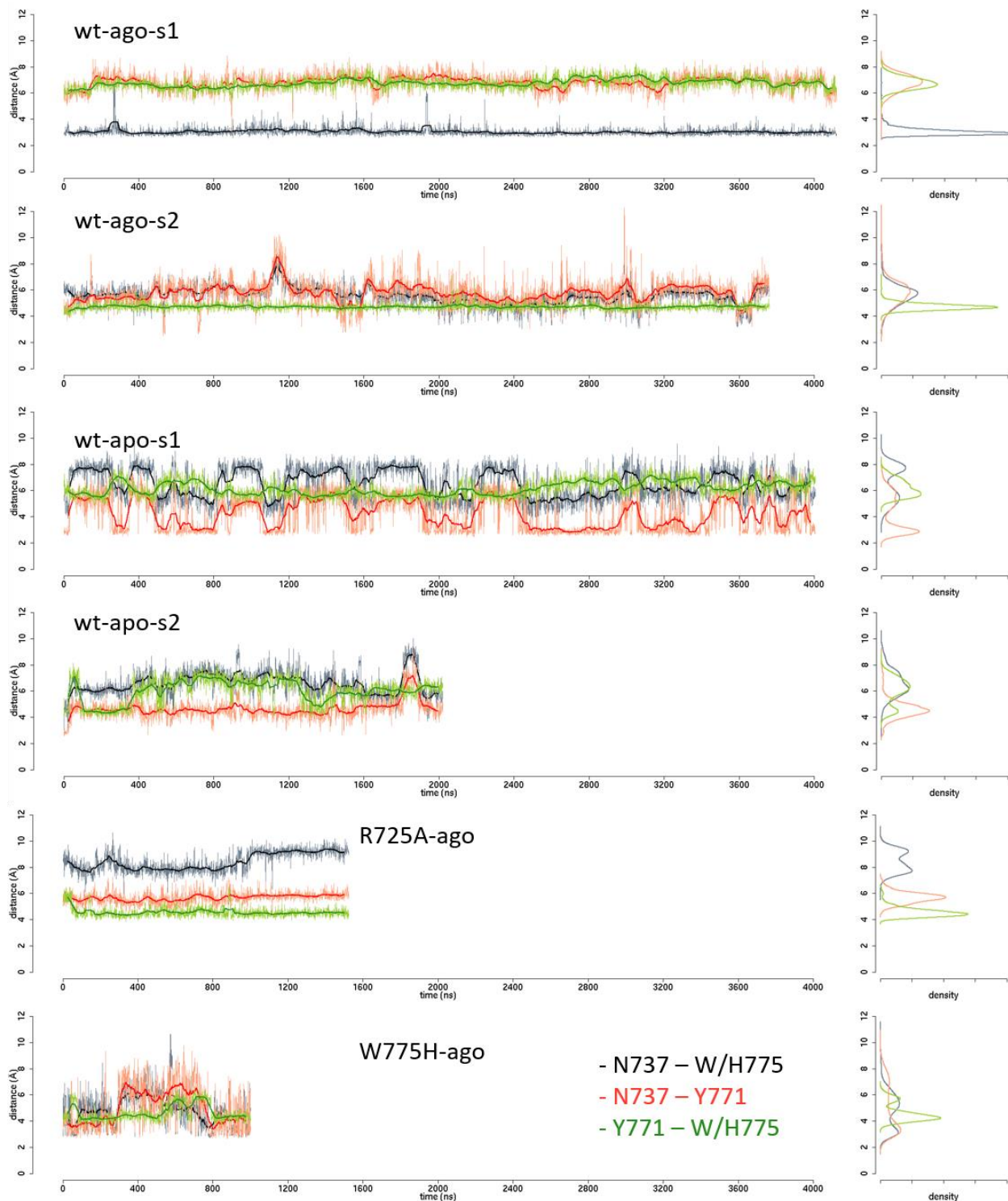


Figure s5: Structural analysis of key residues involved in the TIR3 molecular switch.

Time series plots and density distributions of N737^{5.47}-W/H775^{6.50} and N737^{5.47}-Y771^{6.46} distances and Y771^{6.46}-W/H775^{6.50} side-chain center-of-mass distance in multiple independent MD simulations (see Table s2 for acronym details). Distances have been calculated between the O^{δ1}, Oⁿ and N^{ε1} (or N^{ε2} in the case of W775H mutant) atoms for N737^{5.47}, Y771^{6.46} and W775^{6.50}, respectively. The Y771^{6.46} center of mass has been calculated considering the aromatic ring including the C^{ε1}, C^{ε2}, C^{δ1}, C^{δ2}, C^γ and C^ζ atoms. The W775^{6.50} center of mass has been calculated considering the pyrrole ring including the N^{ε1}, C^{ε2}, C^{δ1}, C^{δ2}, C^γ atoms.

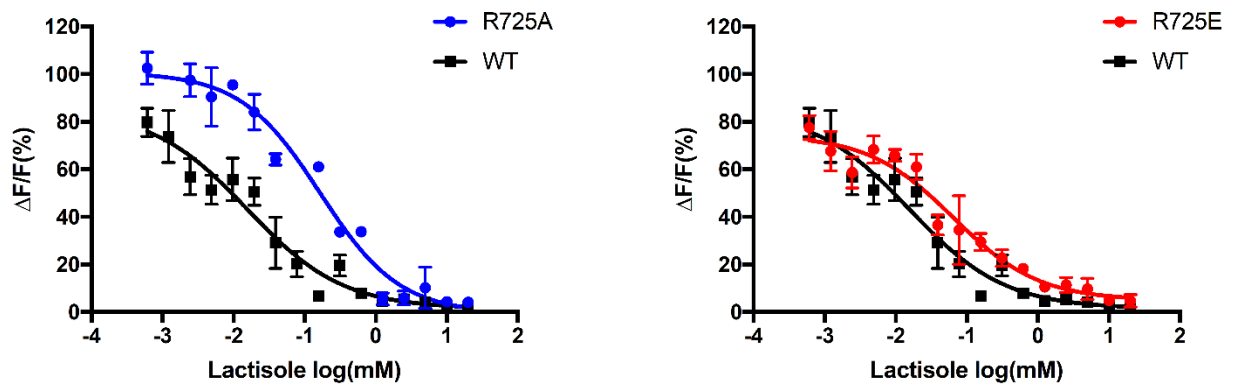


Figure s6: Reduced sensitivity to lactisole of sweet taste receptor R725E and R725A mutants.

Dose-response profiles toward D-tryptophan (10 mM) plus varying amounts of lactisole of (■) wild-type and (●) R725E/A mutant receptors. All transfections were conducted in triplicate.

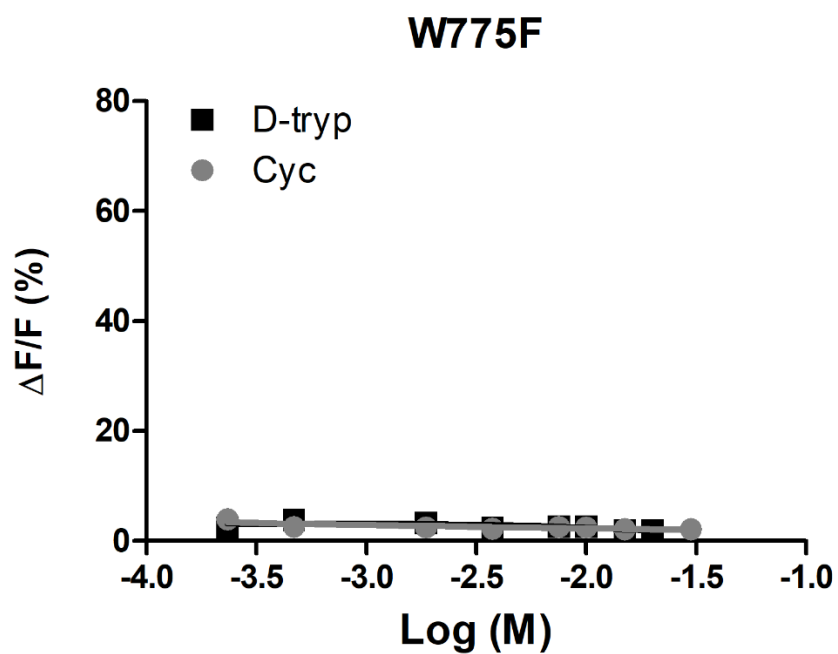


Figure s7: Dose-response curves for sweet taste receptor W775F mutant.

Cyclamate and D-tryptophan dose response curves obtained for the sweet taste receptor single point mutant of residue W775F. All transfections were conducted in triplicate; each experiment was repeated two to three times for D-tryptophan (■) and cyclamate (●).

Table s3: Meta-analysis of T1R3 TMD site-directed mutagenesis data for allosteric and orthosteric agonists.

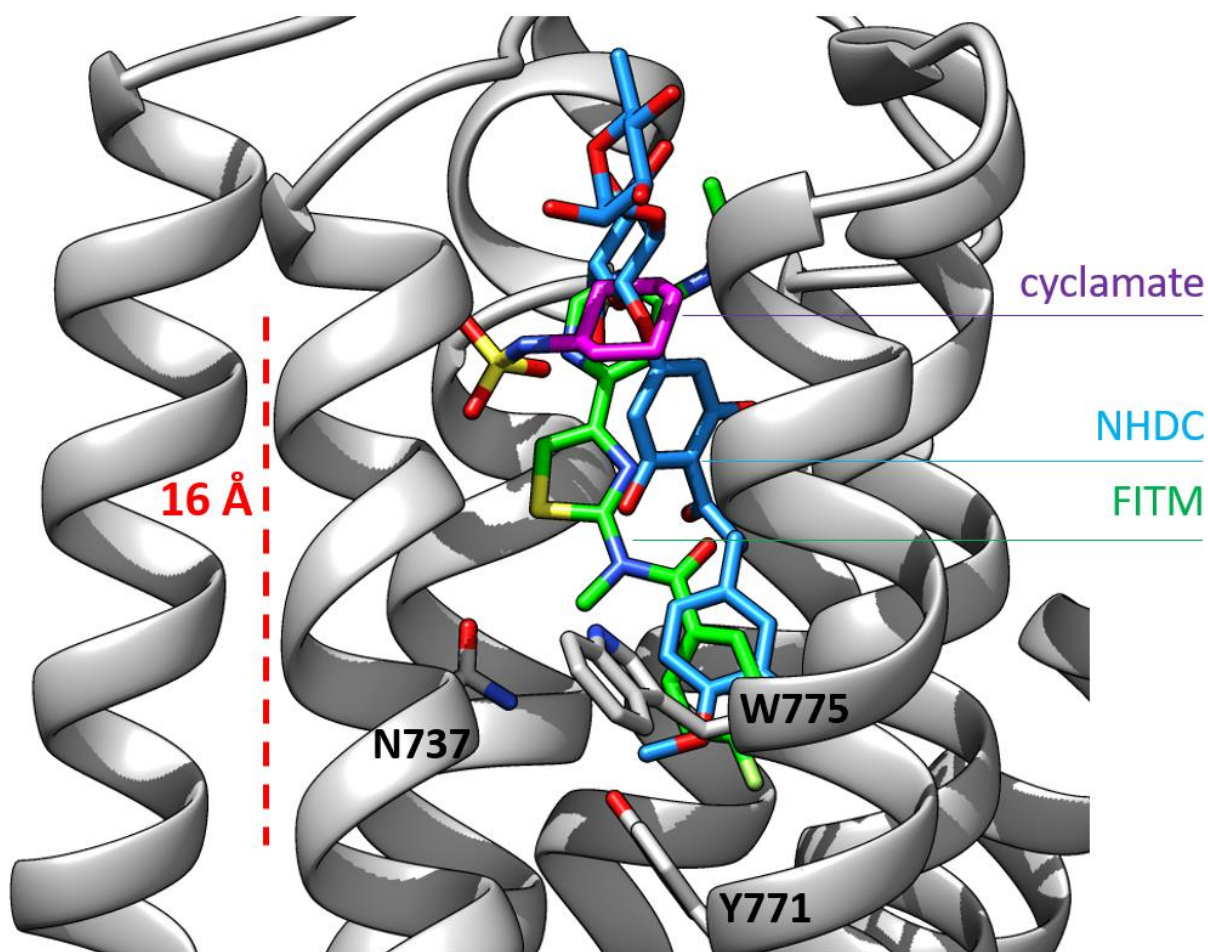
Experimental data come from Jiang *et al.* 2005(1) and Winnig *et al.* 2007(2). New site-directed mutagenesis experiments reported in the present study (see Figure 2c) are indicated by an asterisk. $\Delta(\text{mut-wt})$ indicates the mutation effects on the receptor response. D-Trp and Asp means D-tryptophan and aspartame respectively.

Position	Ballesteros Weinstein numbering	Mutation	$\Delta(\text{mut-wt})$		References
			D-Trp or Asp	Cyclamate	
620	2x52	S \Rightarrow A	-36%	-30%	(2)
621	2x53	V \Rightarrow I	-36%	-32%	(2)
621	2x53	V \Rightarrow L	-94%	-92%	(2)
624	2x56	F \Rightarrow L	-79%	-78%	(2)
636	3x32	Q \Rightarrow A	-39%	-86%	(2)
636	3x32	Q \Rightarrow A	-50%	-104%	(1)
637	3x33	Q \Rightarrow E	-48%	-81%	(2)
637	3x33	Q \Rightarrow E	-31%	-100%	(1)
640	3x36	S \Rightarrow A	-36%	-38%	(2)
640	3x36	S \Rightarrow A	-21%	-21%	(1)
641	3x37	H \Rightarrow A	-18%	-62%	(2)
641	3x37	H \Rightarrow A	-17%	-71%	(1)
644	3x40	L \Rightarrow A	-104%	-104%	(1)
645	3x41	T \Rightarrow A	-96%	-104%	(1)
699	4x50	Y \Rightarrow F	-42%	-49%	(2)
699	4x50	Y \Rightarrow L	-30%	-38%	(2)
721	ecl2	H \Rightarrow A	-39%	-81%	(2)
721	ecl2	H \Rightarrow A	-38%	-88%	(1)
723	ecl2	R \Rightarrow A	-30%	-49%	(2)
723	ecl2	R \Rightarrow A	-17%	-54%	(1)
724	ecl2	T \Rightarrow L	-30%	-41%	(2)
725	ecl2	R \Rightarrow A	-12%	-27%	*
725	ecl2	R \Rightarrow E	-16%	-58%	*
725	ecl2	R \Rightarrow M	-42%	-35%	(2)
726	ecl2	S \Rightarrow A	-48%	-76%	(2)
729	5x39	S \Rightarrow A	-27%	-38%	(2)
729	5x39	S \Rightarrow A	-8%	-29%	(1)
729	5x39	S \Rightarrow F	-14%	-15%	*
729	5x39	S \Rightarrow T	-41%	-30%	*
729	5x39	S \Rightarrow Y	-55%	-70%	*
730	5x40	F \Rightarrow A	-21%	-90%	(1)
730	5x40	F \Rightarrow C	-63%	-95%	(1)

730	5x40	F⇒L	-30%	-76%	(2)
730	5x40	F⇒L	-16%	-82%	(1)
730	5x40	F⇒Q	-32%	-95%	(1)
730	5x40	F⇒S	-37%	-95%	(1)
730	5x40	F⇒T	-47%	-95%	(1)
730	5x40	F⇒W	0%	-52%	(1)
730	5x40	F⇒Y	5%	19%	(1)
733	5x43	A⇒V	-6%	-30%	(2)
733	5x43	A⇒V	42%	5%	(1)
734	5x44	H⇒A	-30%	-95%	(2)
734	5x44	H⇒A	-83%	-96%	(1)
735	5x45	A⇒I	16%	5%	(1)
737	5x47	N⇒A	-64%	-85%	*
739	5x49	T⇒M	16%	18%	(1)
771	6x46	Y⇒A	-104%	-104%	(1)
771	6x46	Y⇒F	-90%	-83%	*
771	6x46	Y⇒L	-19%	-88%	*
775	6x50	W⇒A	-79%	-86%	(2)
775	6x50	W⇒A	-92%	-104%	(1)
775	6x50	W⇒F	-84%	-95%	*
775	6x50	W⇒H	-69%	-90%	*
775	6x50	W⇒Y	-88%	-90%	*
776	6x51	V⇒A	-46%	-38%	(1)
778	6x53	F⇒A	-18%	-70%	(2)
778	6x53	F⇒A	-17%	-88%	(1)
779	6x54	V⇒A	-45%	-30%	(2)
779	6x54	V⇒A	-63%	-54%	(1)
782	6x57	L⇒A	-6%	-81%	(2)
782	6x57	L⇒A	-46%	-100%	(1)
788	ec13	V⇒A	-14%	-18%	(1)
789	ec13	L⇒Y	50%	24%	(1)
790	7x28	R⇒A	-72%	-100%	(1)
790	7x28	R⇒E	-52%	-104%	(1)
790	7x28	R⇒H	-100%	-112%	(1)
790	7x28	R⇒K	-28%	-100%	(1)
790	7x28	R⇒Q	-27%	-81%	(2)
790	7x28	R⇒Q	0%	-94%	(1)
790	7x28	R⇒Y	-60%	-100%	(1)
794	7x32	Q⇒A	-100%	-100%	(1)
798	7x36	L⇒I	-18%	-5%	(2)
798	7x36	L⇒I	0%	-18%	(1)
800	7x38	L⇒F	-9%	-8%	(2)
800	7x38	L⇒V	7%	-12%	(1)
801	7x39	C⇒I	-91%	-89%	(2)
801	7x39	C⇒L	-48%	-89%	(2)
802	7x40	V⇒A	-7%	-18%	(1)
804	7x42	G⇒A	-55%	-43%	(2)
804	7x42	G⇒V	-79%	-76%	(2)
805	7x43	I⇒A	-100%	-104%	(1)
807	7x45	A⇒V	50%	24%	(1)
808	7x46	A⇒T	7%	-12%	(1)

Figure s9: Comparison of allosteric binding pocket of class C GPCR.

T1R3 allosteric binding cavity superimposed on the crystal structure of the mGluR1 (PDB code 4OR2). (Wu et al., 2014) Docked cyclamate and neohesperidine dihydrochalcone (NHDC) are represented by purple and blue sticks respectively while the ligand of mGluR1 (FITM) is represented by green sticks. Residues N737^{5.47}, Y771^{6.46} and W775^{6.50} forming the cradle of the allosteric binding site are represented by grey sticks.



References

Jiang P, Cui M, Zhao B, Snyder LA, Benard LMJ, Osman R, Max M, and Margolskee RF. 2005. Identification of the cyclamate interaction site within the transmembrane domain of the human sweet taste receptor subunit T1R3. *J Biol Chem.* 280:34296–34305.

Winnig M, Bufe B, Kratochwil NA, Slack JP, and Meyerhof W. 2007. The binding site for neohesperidin dihydrochalcone at the human sweet taste receptor. *BMC Struct Biol.* 7:66.

Wu H, Wang C, Gregory KJ, Han GW, Cho HP, Xia Y, Niswender CM, Katritch V, Meiler J, Cherezov V, et al. 2014. Structure of a class C GPCR metabotropic glutamate receptor 1 bound to an allosteric modulator. *Science.* 344:58–64.