

Supplementary Materials for

Control of antiviral innate immune response by protein geranylgeranylation

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Supplementary Material

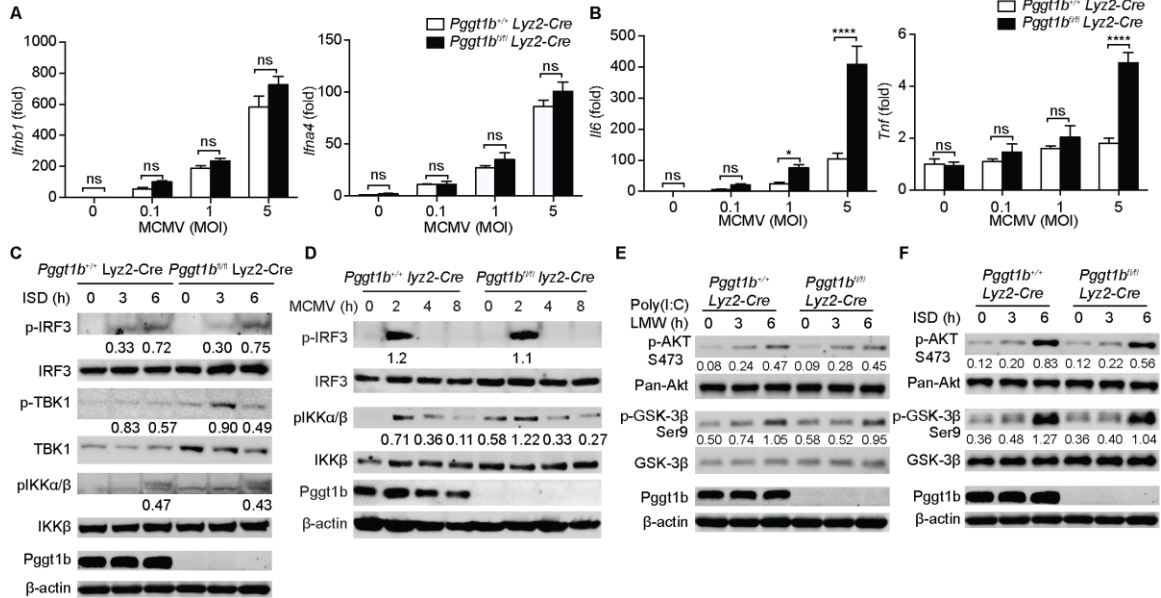


Figure S1

Fig. S1. Protein geranylgeranylation has no effect on DNA virus-induced antiviral innate immune response. (A and B) Quantitative RT-PCR analysis of transcript abundance of *Ifnb1*, *Ifna4*, *Il6* and *Tnf* in *Pggt1b^{+/+} Lyz2-Cre* and *Pggt1b^{fl/fl} Lyz2-Cre* BMDMs 4 h following infection with mouse cytomegalovirus (MCMV). (C and D) Immunoblot analysis of phosphorylated (p-) or total IRF3, TBK1, IKK and Pgg1b or β-actin in *Pggt1b^{+/+} Lyz2-Cre* and *Pggt1b^{fl/fl} Lyz2-Cre* BMDMs transfected with ISD (C) or infected with mouse Cytomegalovirus (MCMV, 1 MOI) (D). (E and F) Immunoblot with phosphor-Akt S473, pan-Akt, phosphor-Gsk3β Ser9, and Gsk3β antibody of BMDMs cell lysates transfected with poly I:C LMW (E) or ISD (F); Numbers below lanes indicate densitometry of phosphorylated AKT S473 or GSK-3β relative to that of total AKT S473 or GSK-3β respectively. *p: <0.05, ****p: <0.0001 (two-way ANOVA). Data are representative of three independent biological experiments.

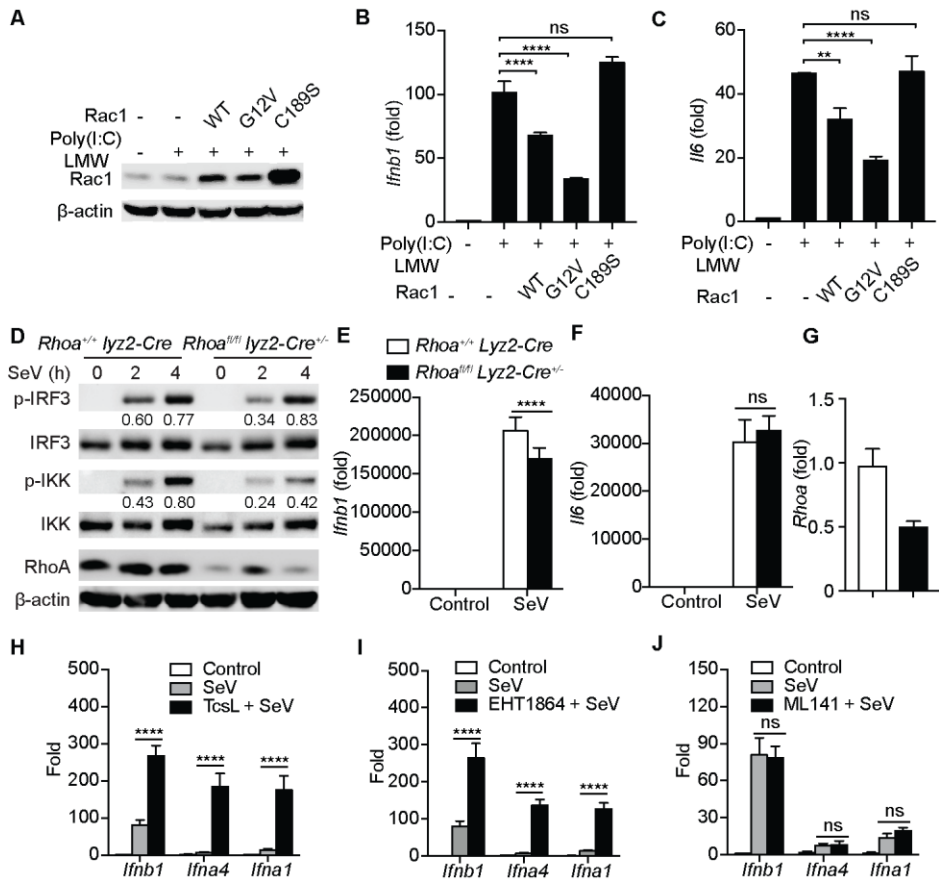


Figure S2

Fig. S2. Rac1 but not RhoA or Cdc42 suppresses MAVS signaling in a protein geranylgeranylation–dependent manner. (A to C) Immunoblot of Rac1 (A) and *Ifnb1* and *Il6* transcript abundance (B and C) in MEFs stably expressing wild-type (WT), constitutive active form (G12V), or C189S mutant *Rac1* after transfection with 1 μ g/mL of poly (I:C) LMW for 4 h. (D) Immunoblot of phospho-IRF3 and phospho-IKK in cell lysates of *Rhoa*^{+/+} *lyz2-Cre* and *Rhoa*^{fl/fl} *lyz2-Cre*^{+/-} BMDMs after SeV infection. (E to G) Quantitative RT-PCR analysis of transcript abundance of *Ifnb1*, *Il6* and *Rhoa* in *Rhoa*^{+/+} *lyz2-Cre* and *Rhoa*^{fl/fl} *lyz2-Cre*^{+/-} BMDMs 4 h after SeV infection. (H to J) Quantitative RT-PCR analysis of transcript abundance of *Ifnb1*, *Ifna4* and *Ifna1* in BMDMs pre-treated with 100 ng/mL TcsL toxin (H), 20 μ M Rac inhibitor EHT1864 (I), or 20 μ M Cdc42 inhibitor ML141 (J), followed by infection with SeV for 4 h. **p: < 0.01, ****p: <0.0001 (two-way ANOVA). Data are representative of three independent experiments.

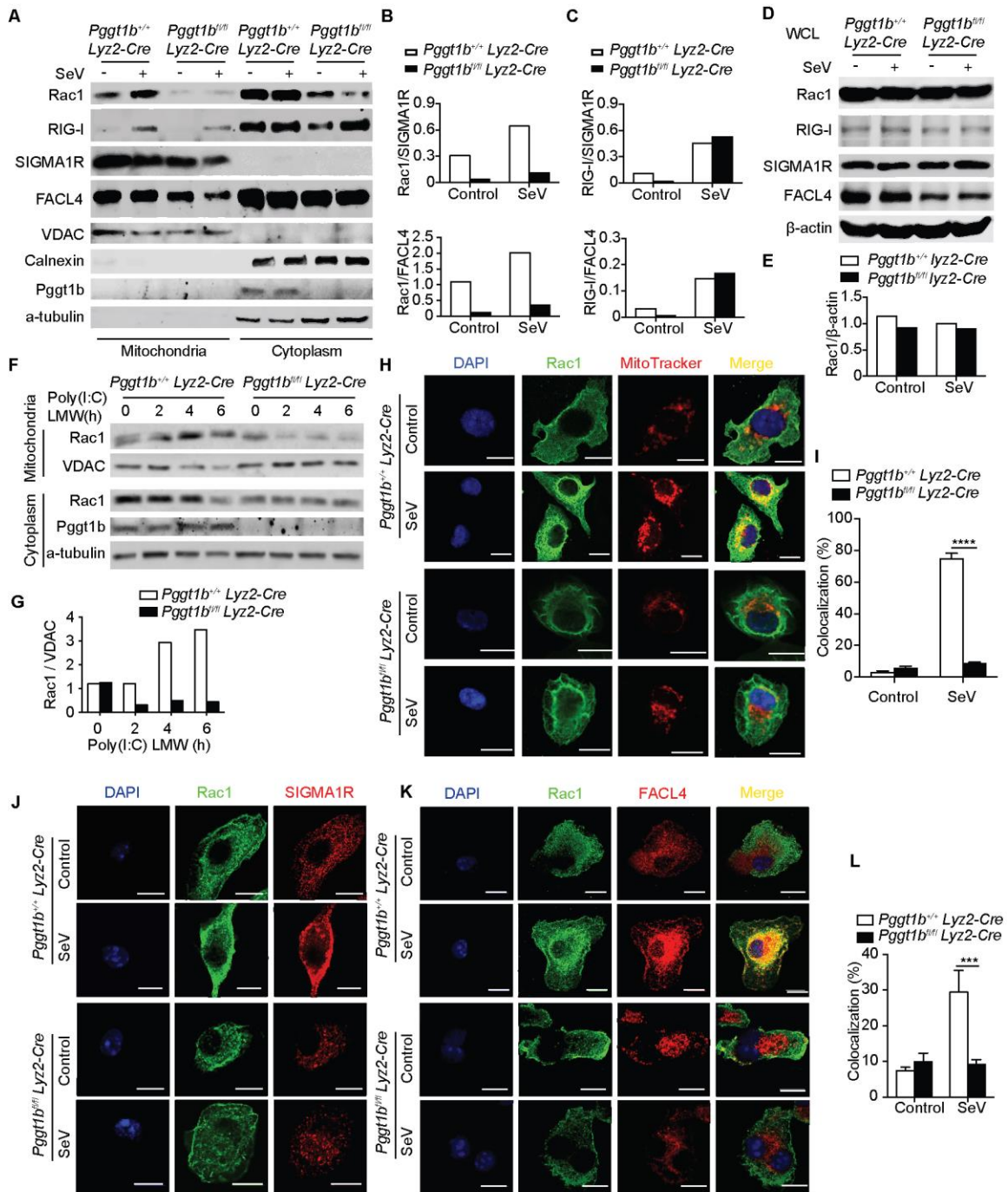


Figure S3

Fig. S3. Protein geranylgeranylation targets Rac1 to MERC ER membranes upon RLR engagement. (A) Immunoblot of crude mitochondria and cytoplasm fractions from *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs after SeV infection. (B and C) Abundance of Rac1 or RIG-I relative to MERC-ER marker SIGMA1R or FACL4 were quantitated using ImageJ. (D) Immunoblot of whole cell lysates (WCL) from *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs infected with SeV. (E) Densitometry of Rac1 relative to that of β -actin in WCL of (D). (F) Immunoblot of crude mitochondria and cytoplasm fractions from *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs transfected with poly (I:C) LMW. (G) Abundance of Rac1 relative to VDAC in (F) was quantitated using ImageJ. (H) Confocal microscopy of Rac1 in *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs after SeV infection. MitoTracker was used to label mitochondria. Scale bar is 10 μ m. (I) The percentage of colocalization was calculated using the ImageJ Software from at least 50 cells randomly selected from the immunostaining slides (H). (J) Single channels of DAPI, Rac1 and SIGMAR1 for Fig. 2I. (K and L) Confocal microscopy of immunofluorescent staining of Rac1 (green) and FACL4 (red) in *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs after SeV infection; Cells were counterstained with DAPI (blue); scale bar is 10 μ m (K); (L) The percentage of colocalization was calculated using the ImageJ software from at least 50 cells randomly selected from the immunostaining slides (K); ***p: < 0.001, ****p: < 0.0001 (two-way ANOVA). Data are representative of three independent experiments.

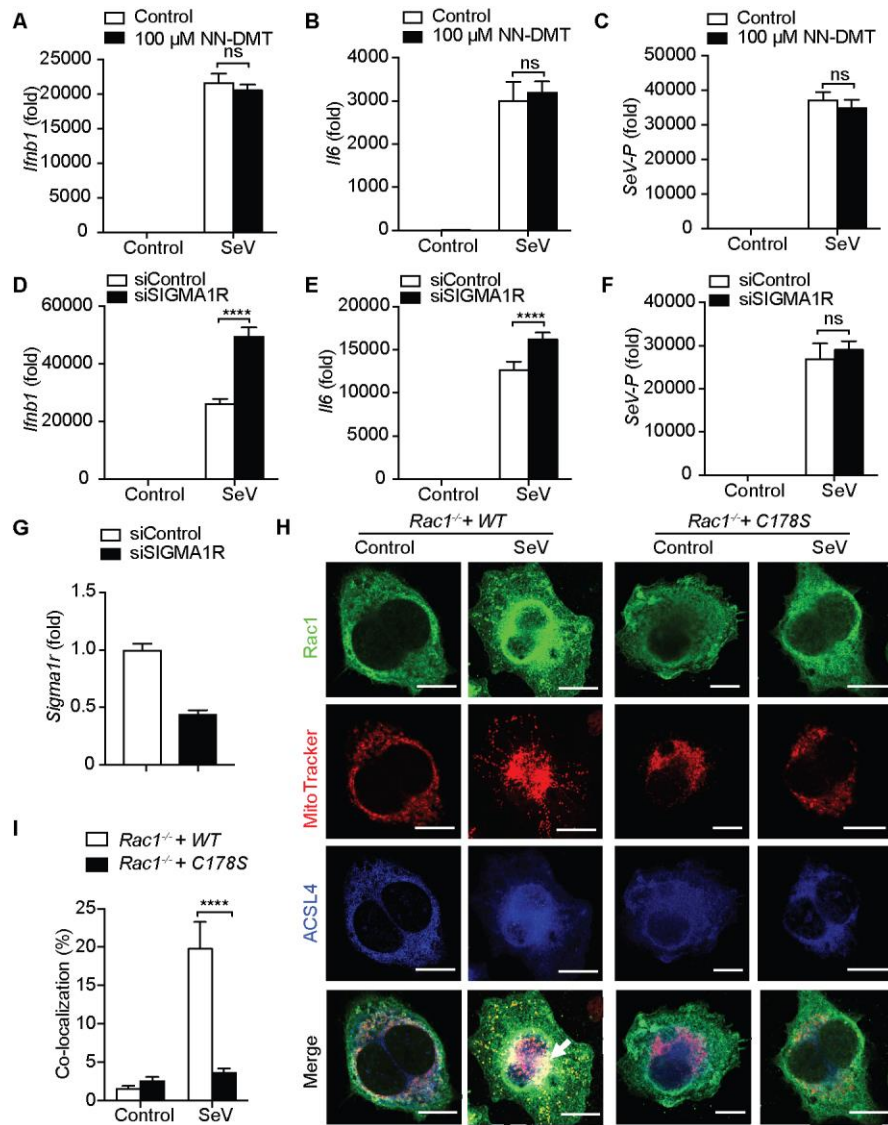


Figure S4

Fig. S4. Chaperon but not the signaling function of SIGMA1R and Rac1 palmitoylation is important for anchoring Rac1 to MERC ER membranes. (A to C) Quantitative RT-PCR analysis of transcript abundance of *Ifnb1*, *Il6* and *SeV-P* in BMDMs pre-treated with 100 μ M NN-DMT for 1 h, followed by SeV infection. **(D to G)** Quantitative RT-PCR analysis of transcript abundance of *Ifnb1*, *Il6*, *SeV-P* and *Sigma1r* in BMDMs transfected with 100 nM Sigma1r siRNA or scramble siRNA for 48 h, followed by SeV infection; ****p: <0.0001 (two-way ANOVA). **(H and I)** Confocal microscopy of colocalization of WT and C178S mutant form of Rac1 (green) together with MitoTracker (Red) and FACL4 (blue) in *Rac1*^{-/-} iBMDMs reconstituted with WT or C178S Rac1, scale bar is 10 μ m **(H)**; **(I)** The percentage of colocalization among Rac1, MitoTracker and FACL4 was calculated using ImageJ **(H)**; ****p: < 0.0001 (two-way ANOVA). Data represent the average and standard error of three independent experiments.

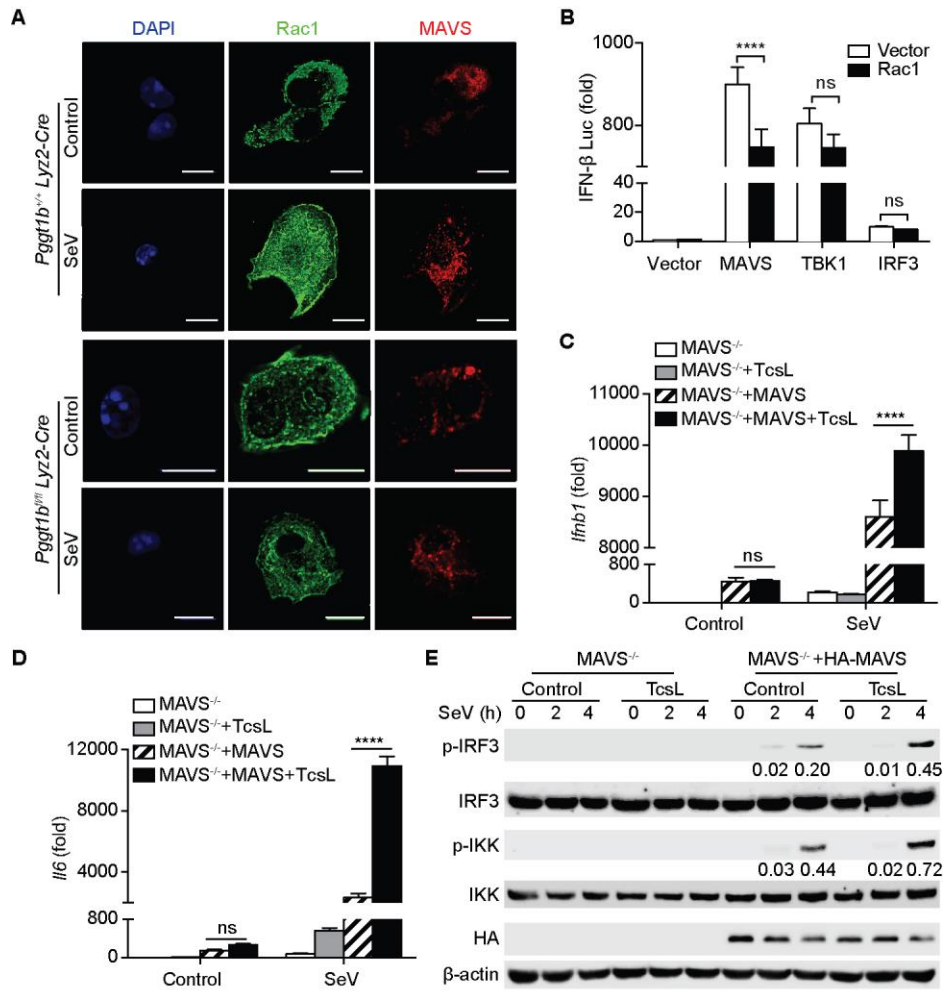


Figure S5

Fig. S5. Rac1 directly engages MAVS signalosome and inhibits MAVS signaling in a protein geranylgeranylation-dependent manner. (A) Single channels of DAPI, Rac1 and MAVS for Fig. 3B. (B) Luciferase activity 24 hours after IFN β -luciferase reporter, MAVS, TBK1 or IRF3 encoding plasmid were transfected together with wild-type Rac1 expressing plasmid into 293 cells. (C and D) Quantitative RT-PCR analysis of *Ifnb1* and *Ii6* in *MAVS*^{-/-} iBMDMs or *MAVS*^{-/-} iBMDMs reconstituted with wild-type MAVS pretreated with TcsL followed by SeV infection; ****p: <0.0001 (two-way ANOVA). (E) Immunoblot of phospho-IRF3 and phospho-IKK in cell lysates of immortalized *MAVS*^{-/-} iBMDMs stably reconstituted with HA-tagged Mavs (*MAVS*^{-/-} + HA-MAVS) pre-treated with TcsL followed by infection with SeV; Numbers below lanes indicate densitometry of phosphorylated IRF3 or IKK relative to that of total IRF3 or IKK respectively. ****p: <0.0001 (two-way ANOVA). Data are representative of three independent experiments.

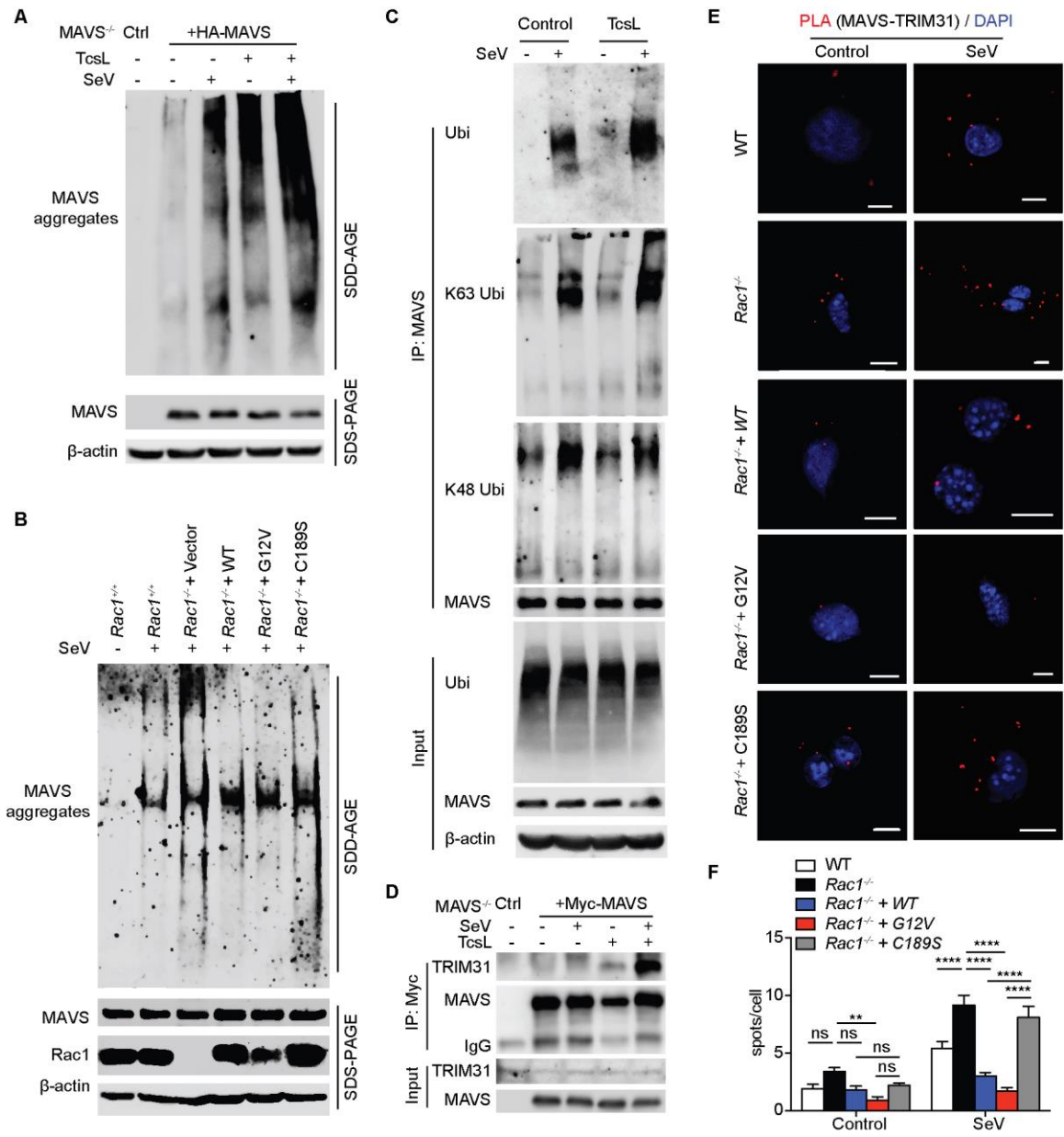


Figure S6

Fig. S6. MERC-localized Rac1 inhibits Trim31-MAVS interaction and subsequent MAVS Lys⁶³-linked ubiquitination, aggregation, and activation upon RNA virus infection. (A) Immunoblot of *Mavs*^{-/-} iBMDMs reconstituted with HA-tagged *Mavs* pre-treated with TcsL, followed by infection with SeV and resolved by SDD-AGE (top) or total cell lysates resolved by SDS-PAGE (bottom). (B) Anti-MAVS immunoblot of mitochondria extracts of wild-type, *Rac1*^{-/-} iBMDMs and *Rac1*^{-/-} iBMDMs reconstituted with WT, G12V, or C189S of *Rac1*, infected with SeV for 4 h and resolved by SDD-AGE (top) or total cell lysate of iBMDMs infected with SeV resolved by SDS-PAGE (bottom). (C) Immunoblot with pan-ubiquitin, K63-, or K48-specific ubiquitin antibody on anti-MAVS immunoprecipitates from cell lysate of wild type macrophages pre-treated with TcsL, followed by infection with Sendai virus. (D) Co-immunoprecipitation analysis of the interaction of Trim31 with MAVS in *MAVS*^{-/-} iBMDMs reconstituted with Myc-tagged *Mavs* pre-treated with TcsL followed by SeV infection. *Mavs*^{-/-} iBMDM was used as a control. (E and F) WT or *Rac1*^{-/-} iBMDMs reconstituted with WT, G12V, or C189S of *Rac1* were infected with SeV for 4 h and fixed, and endogenous MAVS and TRIM31 interaction were determined using PLA assay (red spots); Confocal images were quantified using ImageJ software, (n = 20 cells); **p: < 0.01, ****p: < 0.0001 (two-way ANOVA). Data are representative of three independent experiments.

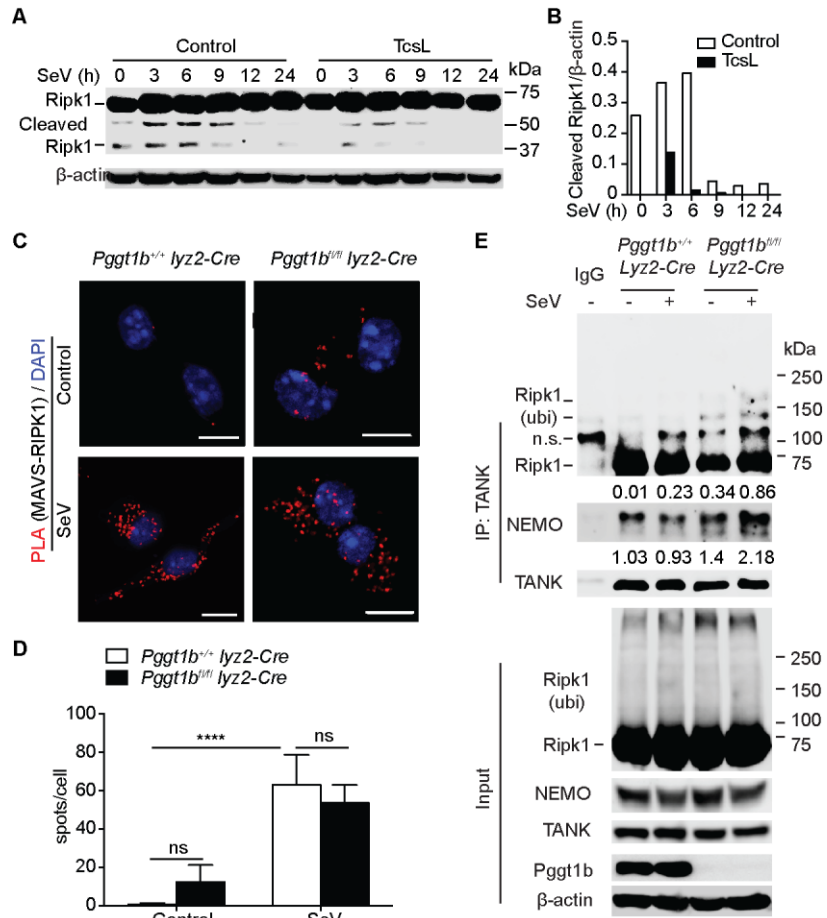


Figure S7

Fig. S7. MERC-localized Rac1 promotes cleavage of Ripk1 and the association of TANK with ubiquitinated Ripk1. (A) Immunoblot with anti-Ripk1 of cell lysate from TcsL pretreated BMDMs infected with SeV at different time points. (B) Densitometry of cleaved 37KD Ripk1 fragment relative to that of β -actin in the whole cell lysates (WCL) of (A). (C and D) *Pggt1b*^{+/+} *Lyz2-Cre* and *Pggt1b*^{fl/fl} *Lyz2-Cre* BMDMs were infected with SeV for 4 h and fixed, and endogenous MAVS and Ripk1 interaction were determined using PLA assay (red spots); Confocal images were quantified using ImageJ software, (n = 20 cells); ****p: < 0.0001 (two-way ANOVA). (E) Immunoblot of anti-TANK immunoprecipitates from the total cell lysates of BMDMs infected with SeV for 4 h. Data are representative of three independent biological experiments.

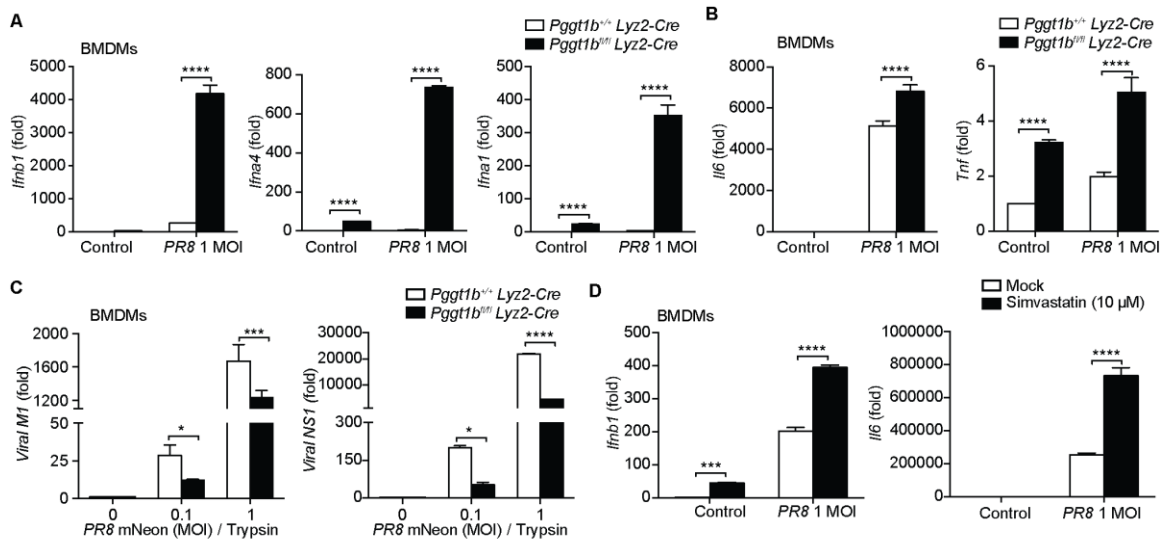


Figure S8

Fig. S8. *Pggt1b* deficiency and Statin pretreatment in BMDMs enhance antiviral innate immune response after influenza A virus *PR8* challenge. (A and B)

Quantitative RT-PCR analysis of *Ifnb1*, *Ifna4*, *Ifna1*, *Il6* and *Tnf* in BMDMs from *Pggt1b*^{+/+} *Lyz2-Cre* or *Pggt1b*^{fl/fl} *Lyz2-Cre* mice 9 h after acute infection with *PR8* (1 MOI). (C) Quantitative RT-PCR of IAV viral genes *M1* and *NS1* in BMDMs pretreated with 10 μ M simvastatin for 18 h followed by infection with IAV *PR8* mNeon (0.01 MOI) containing 1 μ g/mL TPCK-trypsin for 24 h. (D) Quantitative RT-PCR analysis of *Ifnb1* and *Il6* in primary BMDMs pretreated with or without 10 μ M simvastatin for 18 h followed by infection with 1 MOI *PR8*. *p: <0.05, ***p: <0.001, ****p: <0.0001 (two-way ANOVA). Data are representative of three independent biological experiments.

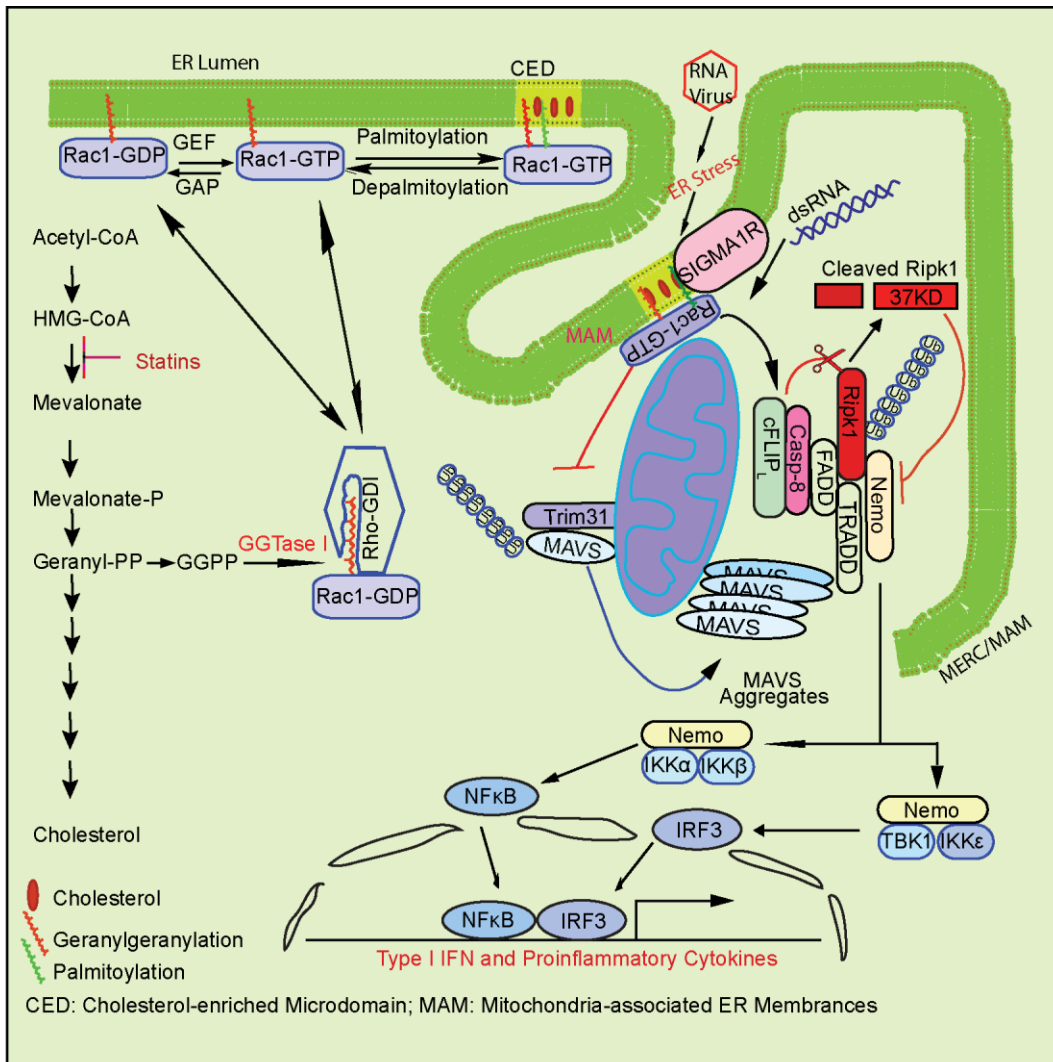


Figure S9

Fig. S9. Graphic summary.

Table S1. Primers for quantitative RT-PCR.

Gene	Forward	Reverse
<i>mIfnb1</i>	5'-CACAGCCCTCTCCATCAACTA-3'	5'-CATTTCCGAATGTTTCGTCCT-3'
<i>mIfna1</i>	5'-ACCCAGCAGATCCTGAACAT-3'	5'-AATGAGTCTAGGGTATTCC-3'
<i>mIfna4</i>	5'-TCAAGCCATCCTTGTGCTAA-3'	5'-GTCTTTTGTATGTGAAGAGGTTCAA-3'
<i>mIl6</i>	5'-GCTACCAAACCTGGATATAATCAGGA-3'	5'-CCAGGTAGCTATGGTACTCCAGAA-3'
<i>mTnf</i>	5'-TGCTGGGAAGCCTAAAAGG-3'	5'-CGAATTTTGTGAGAAGATGATCCTG-3'
<i>mRac1</i>	5'-AGATGCAGGCCATCAAGTGT-3'	5'-TCAAAGACGGTGGGGATGTA-3'
<i>mRac2</i>	5'-GACAGTAAGCCGGTGAACCTG-3'	5'-CTGACTAGCGAGAAGCAGATG-3'
<i>mRhoA</i>	5'-AACTGCATCCAGAACCTGT -3'	5'-TACCACAAGCTCCATCACCA -3'
<i>Vrial NS1</i>	5'-AGAAAGTGGCAGGCCCTCTTTGTA-3'	5'-TGTCCTGGAAGAGAAGGCAATGGT-3'
<i>Vrial M1</i>	5'-TCAGGCCCCCTCAAAGCCGA-3'	5'-GGGCACGGTGAGCGTGAACA-3'
<i>SeV-P</i>	5'-CAAAAGTGAGGGCGAAGGAGAA -3'	5'-CGCCCAGATCCTGAGATACAGA -3'
<i>mPgg1b</i>	5'CCTTCTGTGGCATTGCGTCA-3'	5'-CAACAAGGCGATCTTGAGTTG-3'
<i>mGapdh</i>	5'-TGGCAAAGTGGAGATTGTTGCC-3'	5'-AAGATGGTGATGGGCTTCCCG-3'
<i>hIFNB1</i>	5'- ATGACCAACAAGTGTCTCCTCC-3'	5'- GCTCATGGAAAGAGCTGTAGTG-3'
<i>hIFNA1</i>	5'-GCCTCGCCCTTTGCTTTACT-3'	5'- CTGTGGGTCTCAGGGAGATCA-3'
<i>hGAPDH</i>	5'- CATGAGAAGTATGACAACAGCCT-3'	5'-AGTCCTTCCACGATACCAAAGT-3'

m: murine; h: human