

Figure S1. *Stag3* loss leads to abnormal meiotic prophase and meiotic arrest in mouse testis. Related to Figure 1.

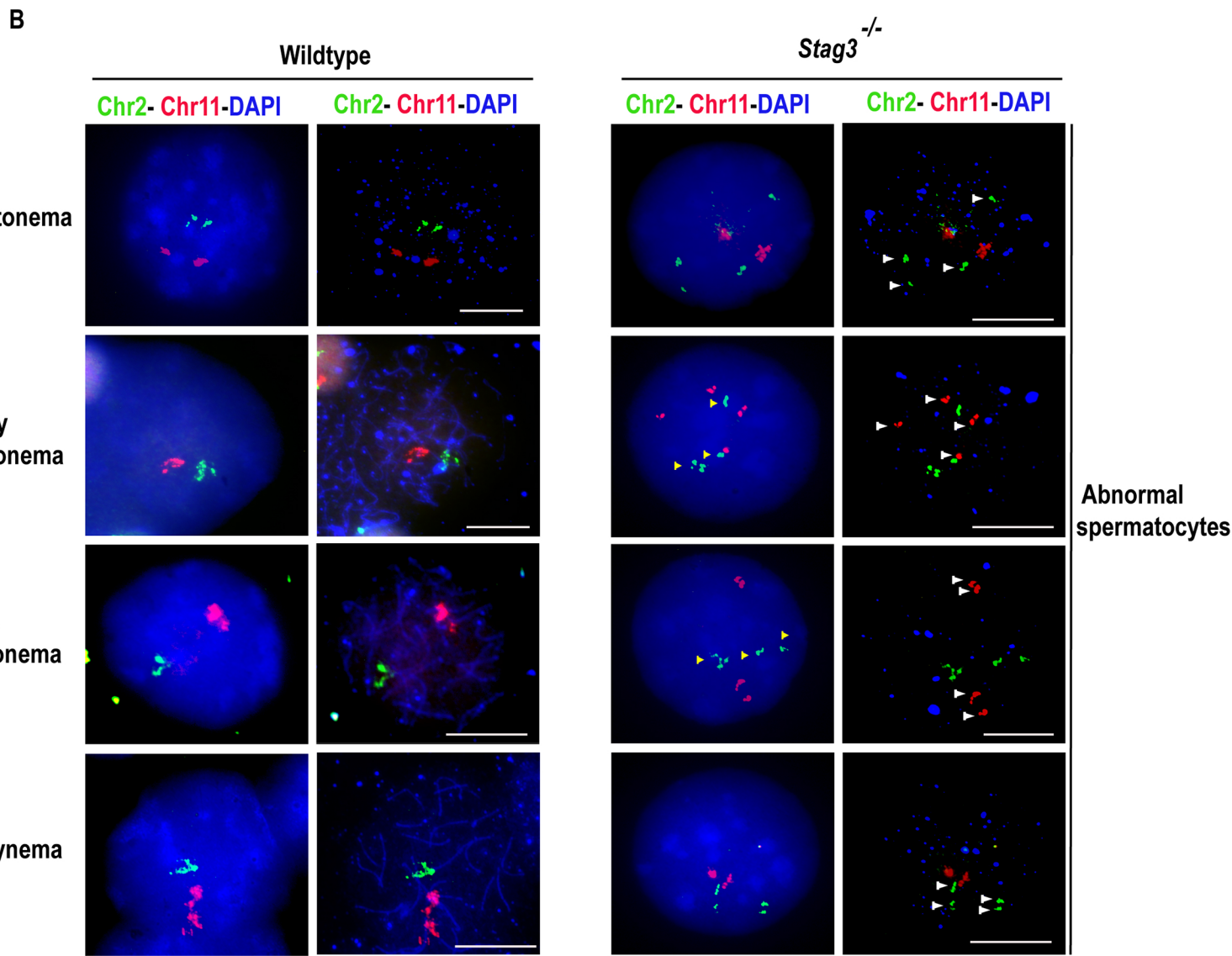
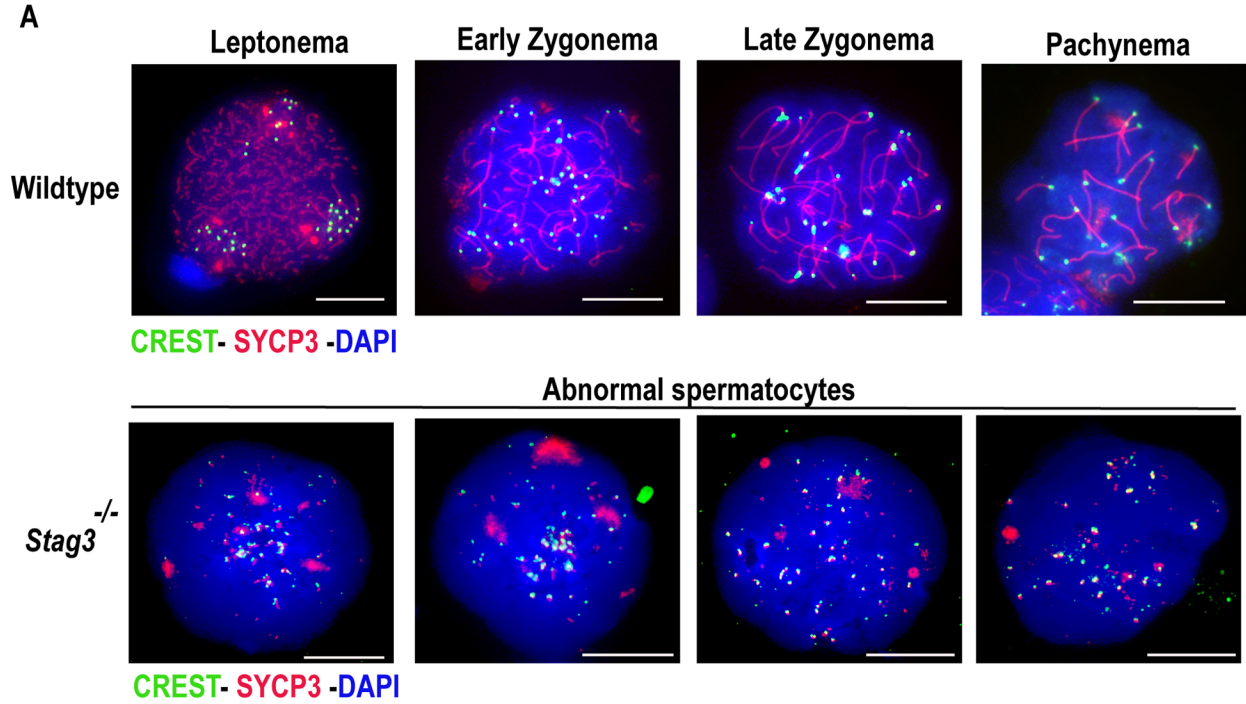
(A) Wildtype (WT) and *Stag3*^{-/-} spermatocyte chromatin spreads immunolabeled using antibodies against PRDM9 (green) and SC protein SYCP3 (red). DNA is stained with DAPI (grey), shown in the lower right corner of each image. Spermatocyte meiotic sub-stages are denoted on top of each image panel. Scale bars= 10μm.

(B) Stacked bar chart showing frequencies of different spermatocyte morphologies in wildtype (WT) and *Stag3*^{-/-} testes based on the staining patterns of three meiotic proteins: STRA8, SYCP3, and PRDM9, from Figure 1B. In pre-leptotene and leptotene cells STRA8, SYCP3 and PRDM9 co-express (blue bar), zygotene (-like) spermatocytes co-express SYCP3 and PRDM9 (red bar). Pachytene (-like) spermatocytes lack PRDM9 staining and stain only with SYCP3 (green bar).

(C) Immunofluorescence staining of spermatocyte chromatin spreads from wildtype (WT) and *Stag3*^{-/-} testes, stained with anti-SYCP3 (green) and γH2AFX (red); DNA is stained with DAPI (blue). Wildtype (WT) spermatocyte prophase I sub-stages are indicated, and examples of *Stag3*^{-/-} chromosome spreads showing different staining patterns are provided. Scale bars= 10μm.

(D) Terminal deoxynucleotidyl transferase (TdT) dUTP Nick-End Labeling (TUNEL) assay on wildtype (WT) and *Stag3*^{-/-} testis sections from 8 week old mice showing apoptotic germ cells (green); DNA is stained with DAPI (blue); scale bars=100μm.

(E) Average number of TUNEL-positive cells per testis seminiferous tubule cross-section. The *Stag3*^{-/-} tubules show significantly higher numbers of apoptotic cells than wildtype (B6N) mice ($P < 0.01$ for both comparisons; Mann–Whitney U test).

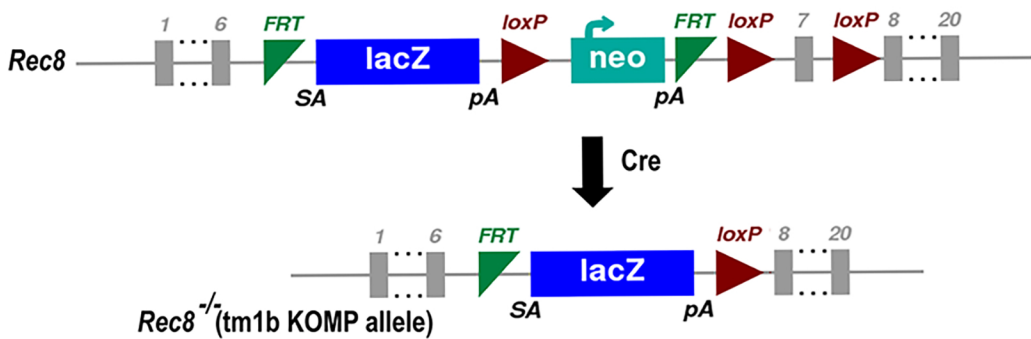


**Figure S2. Premature loss of sister chromatid cohesion in *Stag3*^{-/-} spermatocytes.
Related to Figure 1.**

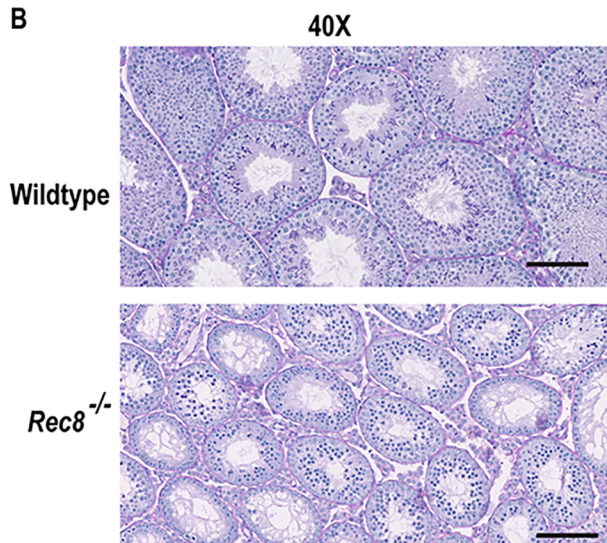
(A) Meiotic chromatin spreads from wildtype and *Stag3*^{-/-} spermatocytes, immunolabeled with antibodies against the centromere (green, CREST) and SC protein SYCP3 (red); DNA is stained with DAPI (blue). Scale bars=10μm.

(B) Wildtype and *Stag3*^{-/-} spermatocyte chromatin spreads stained with DAPI (blue) and hybridized with two 200-kilobase probes against Chrs 2 (green) and 11 (red) distal to the centromere. Note the lack of sister chromatid cohesion in *Stag3*^{-/-} spermatocytes (yellow arrows for 3 distinct FISH signals, white arrows for 4 distinct FISH signals, respectively). Scale bars= 10μm.

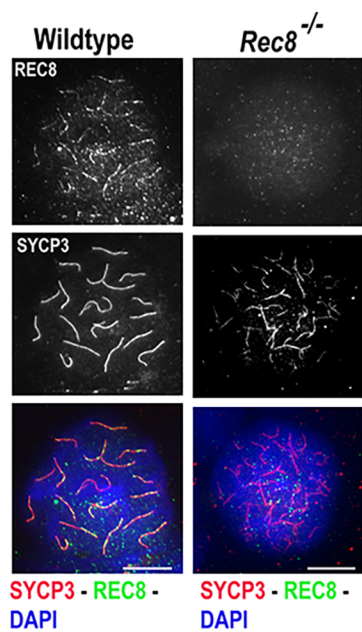
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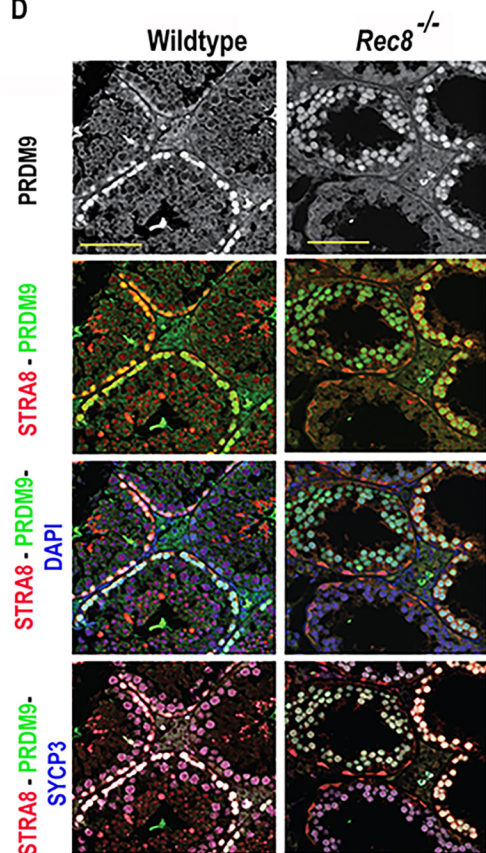


Figure S3. Features of the *Rec8* mutant used in this study. Related to Figure 2.

(A) A beta-galactosidase-containing cassette was introduced into the *Rec8* gene in C57BL/6N-derived embryonic stem (ES) cells. The *Rec8* knockout-first reporter allele was generated by a SOX2-Cre-recombinase system to delete exon 7 of the gene. For further details please refer to International Knockout Mouse Consortium (IKMC) website <https://www.komp.org/ProductSheet.php?cloneID=873812>.

(B) PAS-stained testis sections from 8 week old wildtype and *Rec8*^{-/-} males. Note meiotic arrest in *Rec8*^{-/-} testis. Scale bars=100µm.

(C) Spermatocyte chromatin spreads labeled with antibodies against SC proteins SYCP3 (grey/red) and cohesin subunit REC8 (grey/green). DNA is stained with DAPI (blue). *Rec8*^{-/-} spermatocytes arrest at a zygotene-like stage, lacking synapsis between homologous chromosomes, and exhibiting synapsis between sister chromatids. Scale bars=10µm.

(D) Immunofluorescence staining of testis sections with antibodies against PRDM9 (grey/green), STRA8 (red), SYCP3 (blue). Chromatin was stained with DAPI (blue); scale bars=100µm. Expression of PRDM9 is not obviously affected in *Rec8*^{-/-} testes.

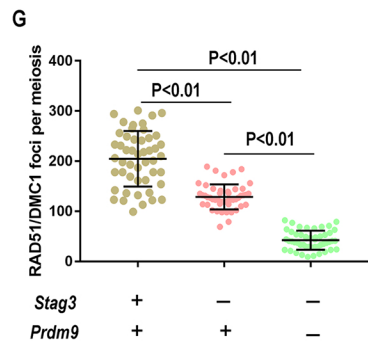
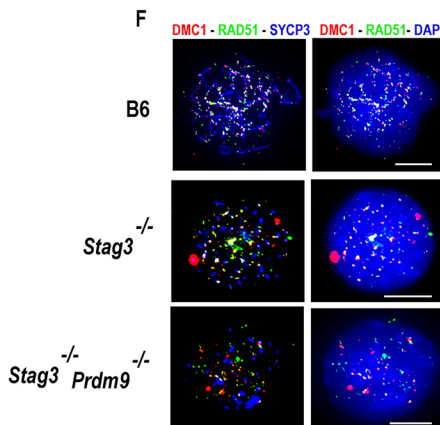
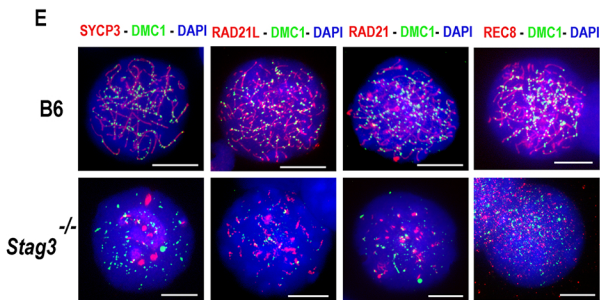
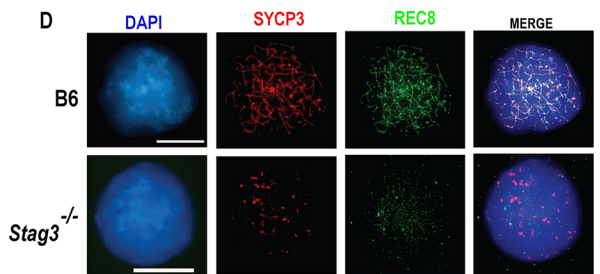
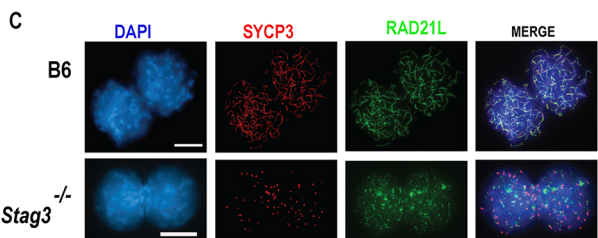
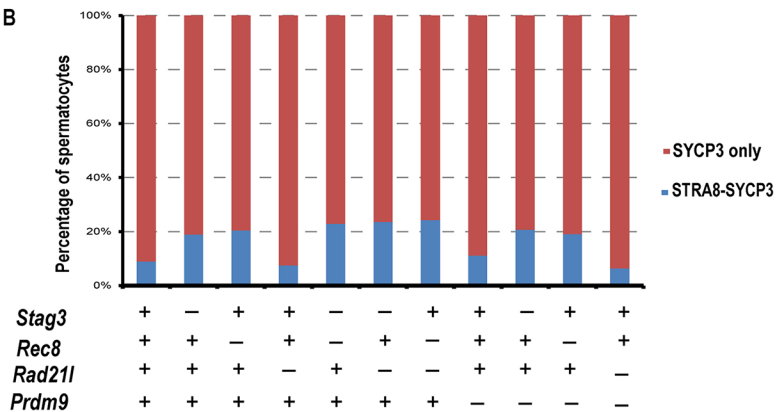
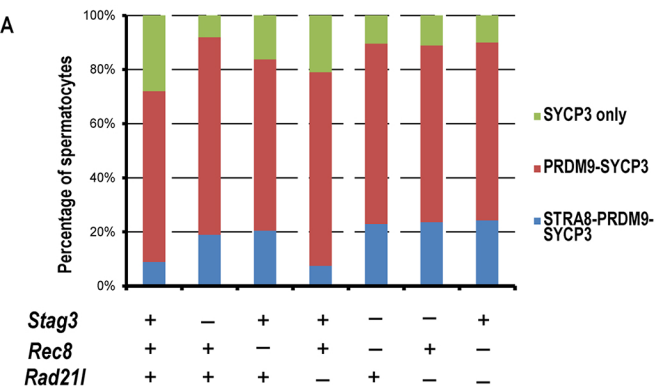


Figure S4. Spermatocyte profile in cohesin- and *Prdm9* deficient single and double mutants. Related to Figure 2.

(A) Stacked bar chart showing frequencies of different spermatocyte morphologies in wildtype (B6) and cohesin mutant testes based on the staining patterns of three meiotic proteins: STRA8, SYCP3, and PRDM9. In pre-leptotene and leptotene cells STRA8, SYCP3 and PRDM9 co-express (blue bar), zygotene (-like) spermatocytes co-express SYCP3 and PRDM9 (red bar). Pachytene (-like) spermatocytes lack PRDM9 staining and stain only with SYCP3 (green bar).

(B) Stacked bar chart showing frequencies of different spermatocyte morphologies in wildtype (B6), cohesin and *Prdm9* single and double mutant testes based on the staining patterns of two meiotic proteins: STRA8, SYCP3 and DAPI. Pre-leptotene and leptotene spermatocytes are STRA8, SYCP3 (blue bar), while other spermatocytes are only SYCP3 positive (red bar).

(C) Sparse chromosome axis-like structures with short linear profiles or punctate foci stained with RAD21L, REC8, and RAD21 in *Stag3*^{-/-} spermatocytes. Immunolabeled wildtype and *Stag3*^{-/-} spermatocytes stained with antibodies against SC protein SYCP3 (red) and cohesin subunit RAD21L (green). DAPI was used for chromatin staining (blue). Scale bars=10µm.

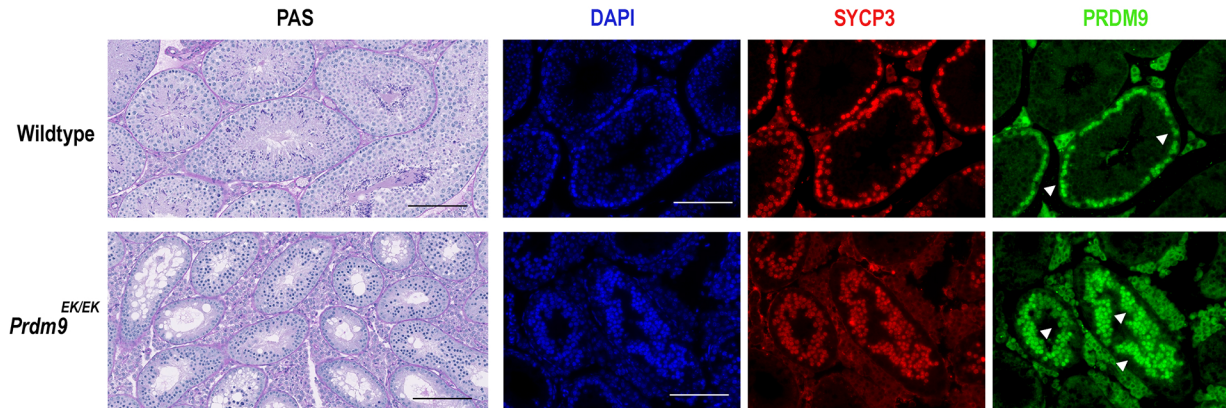
(D) Wildtype and *Stag3*^{-/-} spermatocytes stained with antibodies against SC protein SYCP3 (red) and cohesin subunit REC8 (green). DAPI was used to stain DNA (blue). Note the difference in staining pattern between RAD21L and REC8 in *Stag3*^{-/-} spermatocytes. Scale bars=10µm.

(E) Immunofluorescence staining of wildtype and *Stag3*^{-/-} spermatocyte chromatin spreads with antibodies against SC and axis protein SYCP3 and cohesin subunits RAD21L, RAD21 and REC8 (red) and DSB repair protein DMC1 (green). Note occasional co-localization between axis proteins and DMC1 signal in *Stag3*^{-/-} spermatocytes. DNA is stained using DAPI (blue). Scale bars=10µm.

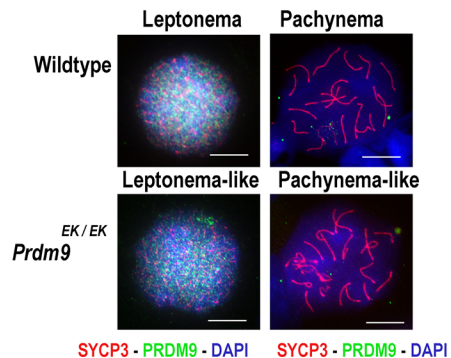
(F) Immunofluorescence detection of RAD51 (green) and DMC1 foci (red) in wildtype, *Stag3*^{-/-} and *Stag3*^{-/-} *Prdm9*^{-/-} spermatocyte chromatin spreads also labeled with antibody against SYCP3 (blue; right panel). Left panel: DNA is stained with DAPI (blue). Scale bars=10µm. Note co-occurrence between RAD51 and DMC1 signal in spermatocytes.

(G) Dot-plot showing quantification of Co-localized RAD51/DMC1 foci in wildtype, *Stag3*^{-/-} and *Stag3*^{-/-} *Prdm9*^{-/-} spermatocytes. Error bars=Standard deviation. P-values were calculated using Mann-Whitney U test with Tukey's multiple testing corrections.

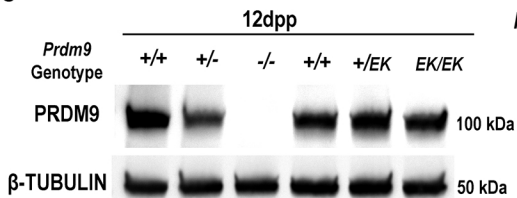
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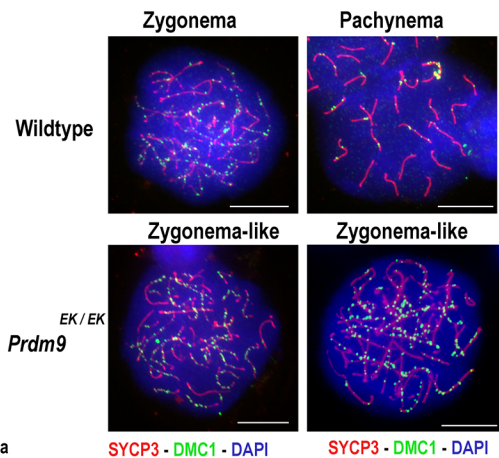
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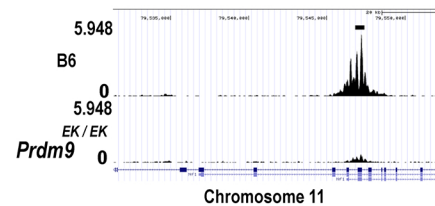
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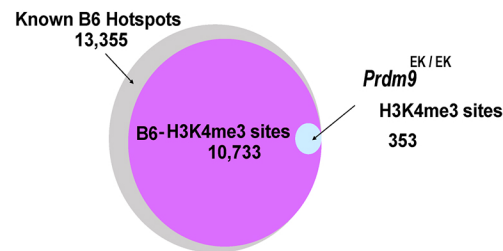


Figure S5. Meiotic arrest and failure of methyltransferase function in *Prdm9*^{EK/EK} spermatocytes. Related to Figure 2 and Methods S1.

(A) Left panel: Wildtype and *Prdm9*^{EK/EK} testis sections from adult mice stained with PAS. The *Prdm9*^{EK/EK} germ cells exhibit arrest at an abnormal late-zygotene-early-pachytene-like stage. Right 3 panels: Immunofluorescence staining of wildtype and *Prdm9*^{EK/EK} testis sections with antibodies against SC protein SYCP3 (red) and PRDM9 (green); DNA is stained with DAPI (blue). The high expression of PRDM9 in *Prdm9*^{EK/EK} spermatocytes suggests that the expression of PRDM9 is not affected by the E365K substitution mutation. Scale bars=100μm.

(B) Wildtype and *Prdm9*^{EK/EK} spermatocyte chromatin spreads immunolabeled using antibodies against PRDM9 (green) and SC protein SYCP3 (red). DNA is stained with DAPI (blue). Spermatocyte meiotic sub-stages are denoted on top of each image panel. Scale bars= 10μm.

(C) Immunoblot showing expression of PRDM9 and β-TUBULIN (loading control) in juvenile (12 d postpartum) testes from wildtype (“+/+”), *Prdm9*^{+ / EK} (“+ / EK”), *Prdm9*^{EK / EK} (“EK / EK”), and *Prdm9*^{- / -} (“- / -”) males. 50 μg of protein extract was loaded per lane.

(D) Wildtype and *Prdm9*^{EK/EK} spermatocytes immunolabeled with anti-SYCP3 (red) and anti-DMC1 antibodies; DNA is stained with DAPI (blue); scale bars=10μm. Almost all *Prdm9*^{EK/EK} spermatocytes fail to repair meiotic DSBs by the pachytene-like stage.

(E) Coverage profile of H3K4me3 from a strongly trimethylated *Prdm9*^{Dom2} hotspot on chromosome 11 in B6J (control) and *Prdm9*^{EK/EK} spermatocyte chromatin, revealing failure of methyltransferase activity of the mutated PRDM9.

(F) Venn diagram showing the number of recombination hotspots with a detectable H3K4me3 peak in wildtype (B6J) and *Prdm9*^{EK/EK} spermatocytes.

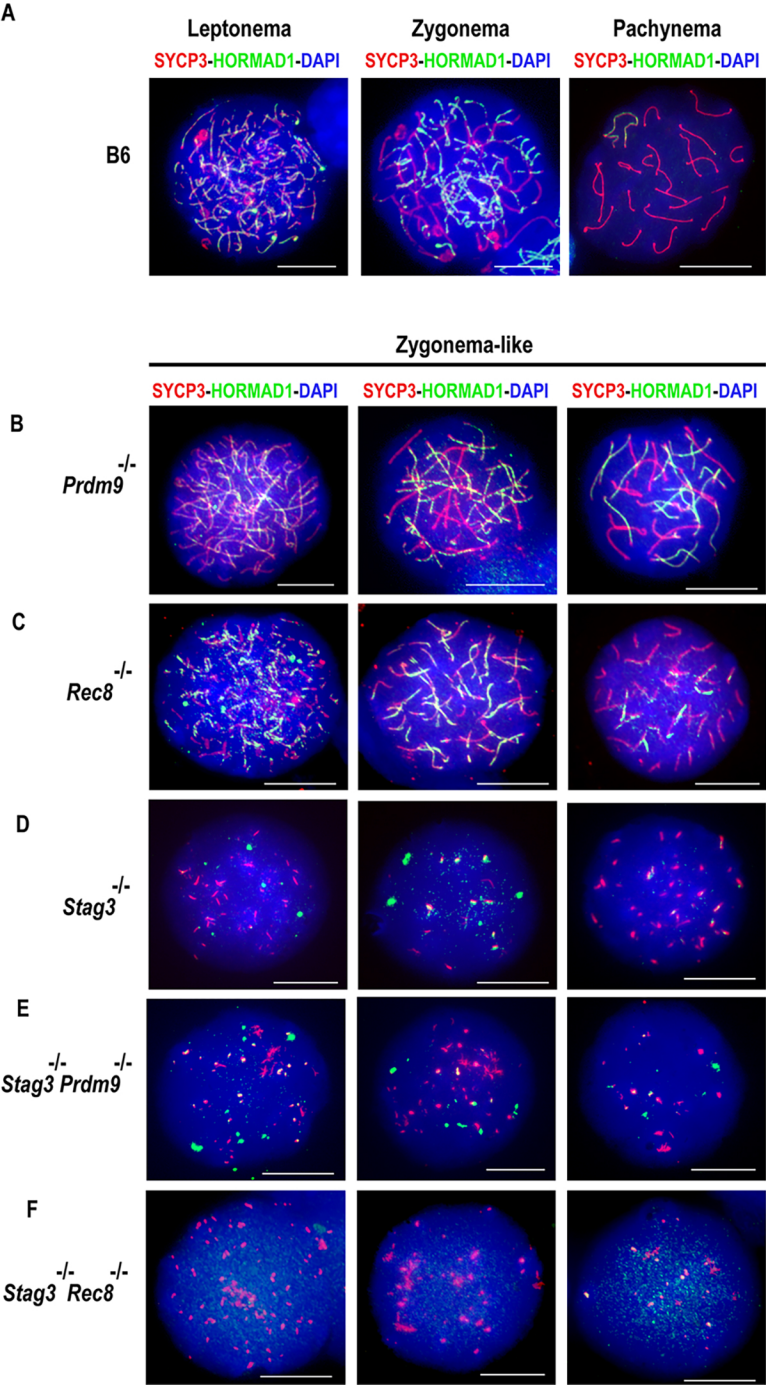


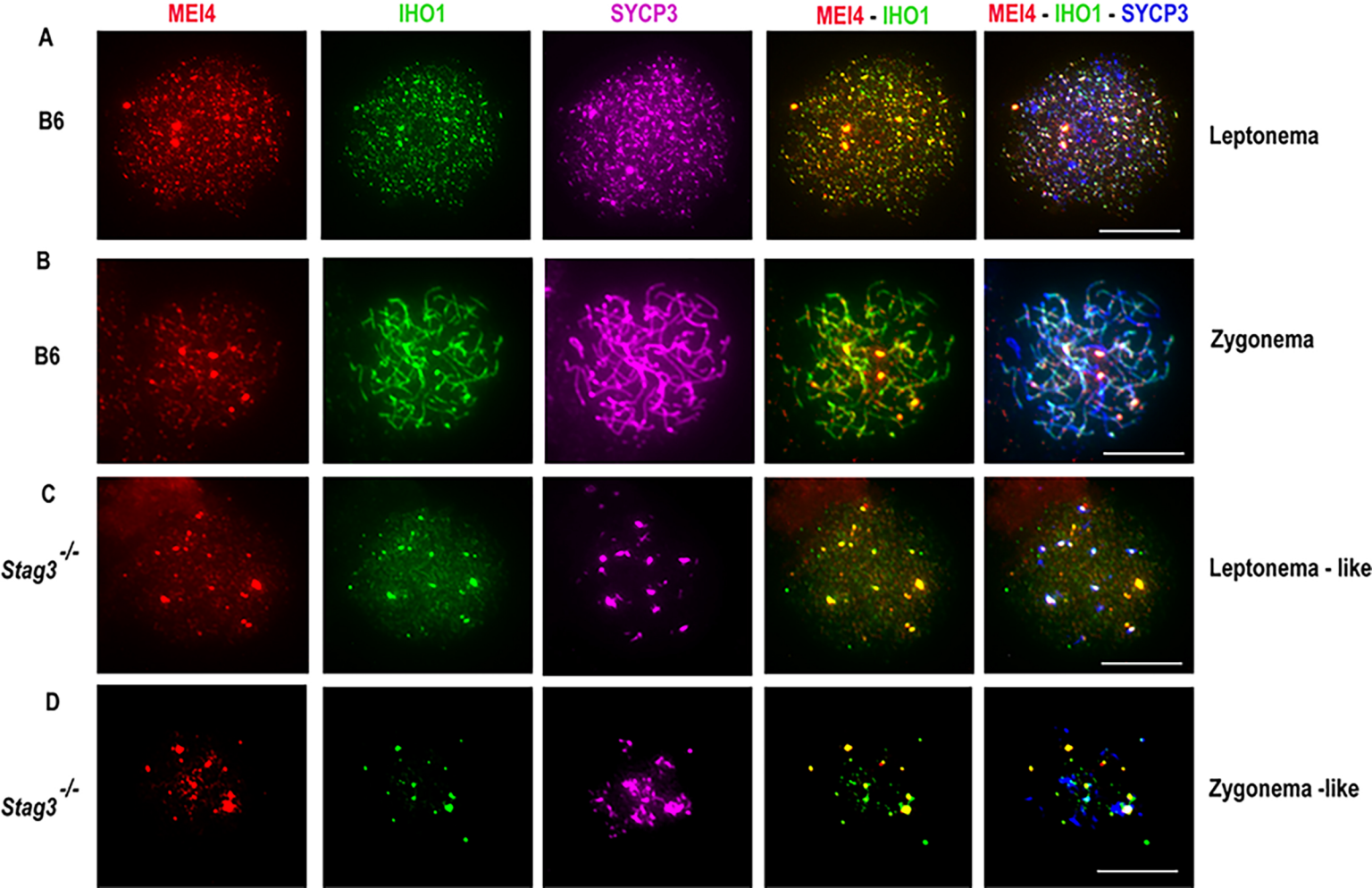
Figure S6. Cohesin proteins are essential for HORMAD1 localization on the chromosome axis. Related to Figure 5.

Immunofluorescence staining of wildtype and different mutant spermatocyte chromatin spreads using antibodies against SC protein SYCP3 (red) and axis protein HORMAD1 (green). DNA is stained with DAPI (blue). For each strain, three representative patterns of staining are provided to better capture the full phenotype.

(A & B) Immunolabeled chromatin spreads from B6N (wildtype) and *Prdm9*^{-/-} spermatocytes with HORMAD1 and SYCP3. Note co-localization of HORMAD1 with SYCP3 in wildtype and *Prdm9*^{-/-} spermatocytes on unsynapsed axes. Scale bars: 10µm.

(C) Immunostained meiotic chromatin spreads from *Rec8*^{-/-} spermatocytes. Note the intermittent staining of HORMAD1 signal on SYCP3 stained axis. Scale bars: 10µm.

(D, E & F) Immunofluorescence staining of *Stag3*^{-/-}, *Stag3*^{-/-}*Prdm9*^{-/-} and *Stag3*^{-/-}*Rec8*^{-/-} spermatocyte chromatin spreads. Both SYCP3 and HORMAD1 fail to form a linear axis-like profiles, but occasionally co-localize. Scale bars: 10µm.



**Figure S7. IHO1 and MEI4 co-localize in the absence of a meiotic cohesin axis.
Related to Figure 5.**

(A & B) B6 leptotene and zygotene spermatocyte chromatin spreads immunolabeled with antibodies against pre-DSB protein MEI4 (red), axis protein IHO1 (green), and SC protein SYCP3 (purple or blue). Note co-localization of IHO1 and MEI4. Scale bars: 10 μ m.

(C & D) Stag3^{-/-} leptotene and zygotene (-like) spermatocyte chromatin spreads stained with antibodies against pre-DSB protein MEI4 (red), axis protein IHO1 (green) and SC protein SYCP3 (purple or blue). Scale bars: 10 μ m. Note that IHO1 and MEI4 co-localize in Stag3^{-/-} spermatocytes in spite of the absence of a meiotic cohesin axis.

	PRDM9	STAG3	RAD21L	REC8	RAD21	SMC3	H3
Input	1.00	1.00	1.00	1.00	1.00	1.00	1.00
IgG	0.00	0.00	0.00	0.00	0.00	0.00	0.00
PRDM9-IP	3.08	0.23	0.00	0.20	0.00	0.74	0.30
PRDM9-IP + DNase	1.64	0.26	0.00	0.18	0.00	0.75	0.00

Table S1. PRDM9 interacts with cohesin complex subunits in mouse spermatocytes. Related to Figure 1.

Quantification of protein bands from Figure 1E. All values were normalized to input. Histone H3 IP was used to detect efficiency of DNase treatment.

	PRDM9	STAG3	RAD21L	REC8	RAD21	SMC3	H3
Input	1.00	1.00	1.00	1.00	1.00	1.00	1.00
IgG	0.00	0.00	0.00	0.00	0.00	0.00	0.01
SMC3-IP	0.45	2.01	9.74	0.54	1.75	1.27	0.14
SMC3-IP + DNase	0.43	1.18	6.52	0.28	1.78	1.02	0.01

Table S2. SMC3 interacts with PRDM9 and cohesin complex subunits in mouse spermatocytes. Related to Figure 1.

Quantification of protein bands from Figure 1F. All values were normalized to input. Histone H3 IP was used to detect efficiency of DNase treatment.

Genotyping Primers	Primer Sequence	Comments
<i>Stag3</i> Mt F	CGG TCG CTA CCA TTA CCA GT	
<i>Stag3</i> Wt F	TCT CAT CTC TAC AAT GCT TGC AC	
<i>Stag3</i> Common	AGA AGA GTA GCC CAA GAG AAG G	
<i>Rad21L</i> Wt F	TCT CAT GCT TCG TGC TTG AC	
<i>Rad21L</i> Wt R	ATG GCC ACT AAT GGG TTC AG	
<i>Rad21L</i> Mt R	TTC GTT CTT GGC CAT TCT CT	
<i>Rad21L</i> Mt F	CGG TCG CTA CCA TTA CCA GT	
<i>Prdm9_hayashi</i> Common	AGG AAT CTT CCT TCC TTG CTG TCG	
<i>Prdm9_hayashi</i> Wt R	ATT TCC CTG TAT CTT CTT CAG GAC T	
<i>Prdm9_hayashi</i> Mt R	CGC CAT TCA GGC TGC GCA ACT GTT	
<i>Prdm9_EK_F</i>	ACT GAA GTA GAG CTT TGA AAC TGG G	Sequencing based genotyping
<i>Prdm9_EK_R</i>	TGG TCC ATT TCT TGC TTC ACC	Sequencing based genotyping
<i>Rec8</i> Wt F	CAC TCA CAC TGG ATG TGG TAA TG	
<i>Rec8</i> Wt R	TGT TTG TAT GCA CCC TTG GA	
<i>Rec8</i> Mt F	TCC TGG GAT TAC TGA TGT CCA	
<i>Rec8</i> Mt R	CGG TCG CTA CCA TTA CCA GT	
<i>Spo11</i> Mt F	GCC AGA GGC CAC TTG TGT AG	
<i>Spo11</i> Wt F	TCA GGA CAG GGC ATA GCA GT	
<i>Spo11</i> Common	CTG CTC AGG GAG GAG AAC AC	

Table S3. Primers required for genotyping different mouse mutants. Related to STAR Methods.

In the table, “Wt” refers to wildtype and “Mt” refers to mutant-specific genotyping primers.