

Supplementary Materials for

Activation of P-TEFb by cAMP-PKA signaling in autosomal dominant polycystic kidney disease

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Supplementary Materials and Methods

Immunohistochemistry (IHC)

The kidney tissues were fixed in 10% formalin overnight and paraffin-embedded. Tissue sections were blocked and incubated with primary antibody against PCNA (Santa Cruz, sc-56). After primary antibody incubation, tissue sections were incubated with goat anti-mouse/rabbit-HRP secondary antibody followed by staining with DAB substrate.

IHC sections were imaged using a microscope (BX51, Olympus).

Embryonic kidney culture

Kidneys were dissected from CD1 mice at E13.5, and grown on 0.4 μm transparent Millicell organotypic cell culture inserts (Millipore) in 12-well plates. Embryonic kidneys were maintained in a 37°C incubator with 5% CO₂. A maximum of four explants were grown on each membrane. Kidneys were cultured in DMEM/F12 medium supplemented with 2 mM L-glutamine (Sigma, G7513); 10 mM HEPES (Sigma, H3375); 1/100 insulin-transferrin-selenium (Life, 41400-045); 1/40 penicillin/streptomycin (Life, 15140-122); 25 ng/ml prostaglandin E1 (Sigma, P5515); 32 pg/ml T3 (Sigma, T5516); 100 μM 8-Br-cAMP (Sigma, B5386). Appropriate amount of each drug or DMSO were added to each well. The medium was changed twice per day. Total area and cyst area measurement were evaluated using the NIH ImageJ software. The cyst index was calculated as the ratio of cyst area to total area.

MDCK I cell cyst growth assay

For cyst generation, 400 MDCK I cells were suspended in 400 μ l of MEM containing 10 mM HEPES; 3.1 mg/mL collagen I (PureCol) and 27 mM NaHCO₃. After gel formation by incubation for 90 minutes at 37°C, 1 mL of DMEM/F12 medium supplemented with 10% FBS and 10 μ M forskolin (Sigma, F3917) was added to each well. MDCK I cells were cultured up to twelve days in a 37°C incubator with 5% CO₂. The medium was changed once per day. Cells were treated with FP from day 4 to day 12. Cells were transfected either with scrambled or CDK9 D167N mutant before cyst generation. Cells were infected with lentivirus carrying either control shRNA or shCDK9 before cyst generation. Micrographs of the same cysts were taken on day 4, 6, 8, 10 and 12 after seeding. For analysis of cyst formation and growth, cyst numbers were counted, and diameters were measured using the NIH ImageJ software. CDK9 shRNA sequences :
CDK9 #1 (GCTTCTGAAACACGAGAATGT), CDK9 #2
(GCTGCAAAGGCAGTATATACC), CDK9 #3 (GCTCAACGGCCTCTACTATAT).

Antibodies used for WB

Cyclin T1 (Santa Cruz, sc-10750), CDK9 (Santa Cruz, sc-8338), RNAPII (Santa Cruz, sc-899), GST (Santa Cruz, sc-459), Polycystin-2 (Santa Cruz, sc-28331), EGFP (Santa Cruz, sc-8334), α -Tubulin (Proteintech, 10449-1-AP), MePCE (Protein- tech, 14917-1-AP), LARP7 (Proteintech, 17067-1-AP), RNAPII CTD S2 (Millipore, 04-1571),

RNAPII CTD S5 (Millipore, 04-1572), GAPDH (Millipore, MAB347), p-PKA Substrate (Cell Signaling Technology, 9624), Myc (Cell Signaling Technology, 2278), p-CREB (Cell Signaling Technology, 9198), HEXIM1 (Bethyl, A303-112A), FLAG-HRP (Sigma, F7425), His (Ruiying Biological, RLM3004) and p-HEXIM1(ser158) (Custom made). For p-HEXIM1(ser158) antibody: Rabbit polyclonal antibodies against p-HEXIM1 (ser158) were produced by Jingjie PTM Biolabs (Hangzhou). The peptides used for antibody generation were CKHRRRPpSKKKRHW (phosphorylated) and CKHRRRPSKKKRHW (unphosphorylated).

Primers used for RT-PCR

Ngal [GACTTCCGGAGCGATCAGTT (forward) and CTGATCCAGTAGCGACAGCC (reverse)], *p16* [ATGGGTTCGCAGGTTCTTGG (forward) and CGTGAACGTTGCCCATCATC (reverse)], *Mcp-1* [CAGGTCCCTGTTCATGCTTCT (forward) and CCCATTCCTTCTTGGGGTCA (reverse)], *Il-6* [AGTTGTGCAATGGCAATTCTGA (forward) and GGTACTCCAGAAGACCAGAGGA (reverse)], *Tgfb1* [AGCTGCGCTTGCAGAGATTA (forward) and AGCCCTGTATTCCGTCTCCT (reverse)], *Colla1* [TTCTCCTGGCAAAGACGGAC (forward) and CTCAAGGTCACGGTCACGAA (reverse)], *7SK snRNA* (Human) [GACATCTGTCACCCCATTGA (forward) and GCGCAGCTACTCGTATACCC (reverse)], *7SK snRNA* (Dog) [GCGACATCTGTCACCCCATT (forward) and

TTGGAGGTTCTAGCAGGGA(reverse)]

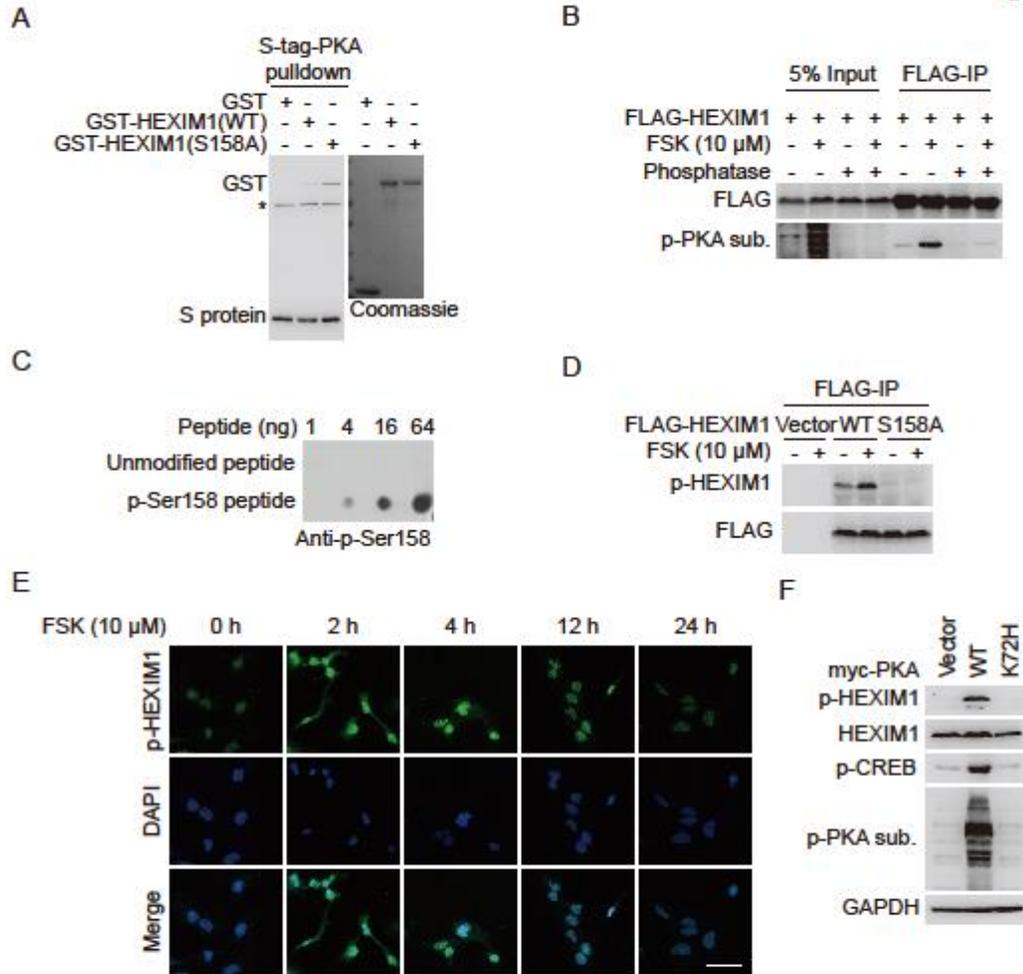


Fig. S1. PKA phosphorylates HEXIM1 at serine-158. (A) S-tag pulldown assays examining the interactions between S-tag-PKA and GST-HEXIM1 (WT and S158A). Asterisk indicates the non-specific bands. (B) FLAG-IP assays analyzing the phosphorylation status of FLAG-HEXIM1 in 293T cells treated with the indicated agents. (C) Dot blot analysis of p-HEXIM1. (D) FLAG-IP assays examining the phosphorylation status of FLAG-HEXIM1 (WT and S158A) using the anti-p-HEXIM1 antibody. (E) Immunofluorescence analysis of p-HEXIM1 in 293T cells treated with FSK. (F) Western blot analysis of p-HEXIM1 and p-CREB in 293T cells transfected with vector or myc-PKA (WT and K72H). Scale bar: 50 μ m.

fig. S2

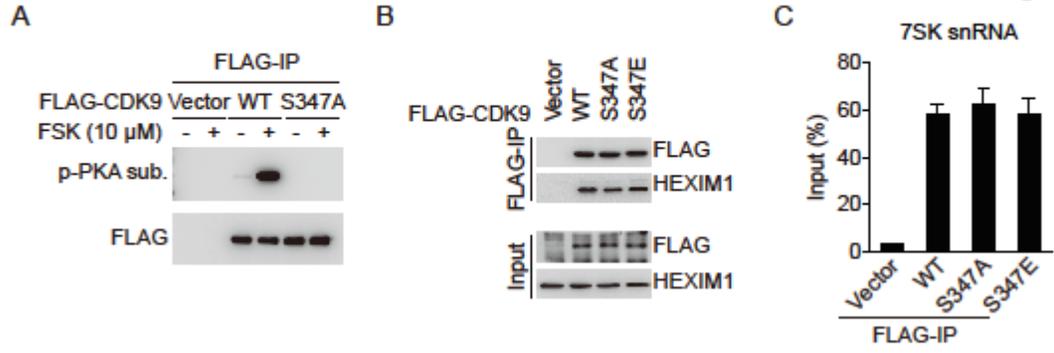


Fig. S2. Phosphorylation of CDK9 at serine-347 by PKA does not affect P-TEFb/HEXIM1/7SK

snRNP complex assembly. (A) FLAG-IP assays analyzing the phosphorylation status of FLAG-CDK9 (WT and S347A) in 293T cells treated with FSK. (B) Co-IP assays examining the interactions between FLAG-CDK9 (WT, S347A and S347E) and endogenous HEXIM1 in 293T cells. (C) RNA-IP assays examining the associations between the indicated FLAG-CDK9 and 7SK snRNA.

Data represent as mean \pm SEM of three replicates.

fig. S3

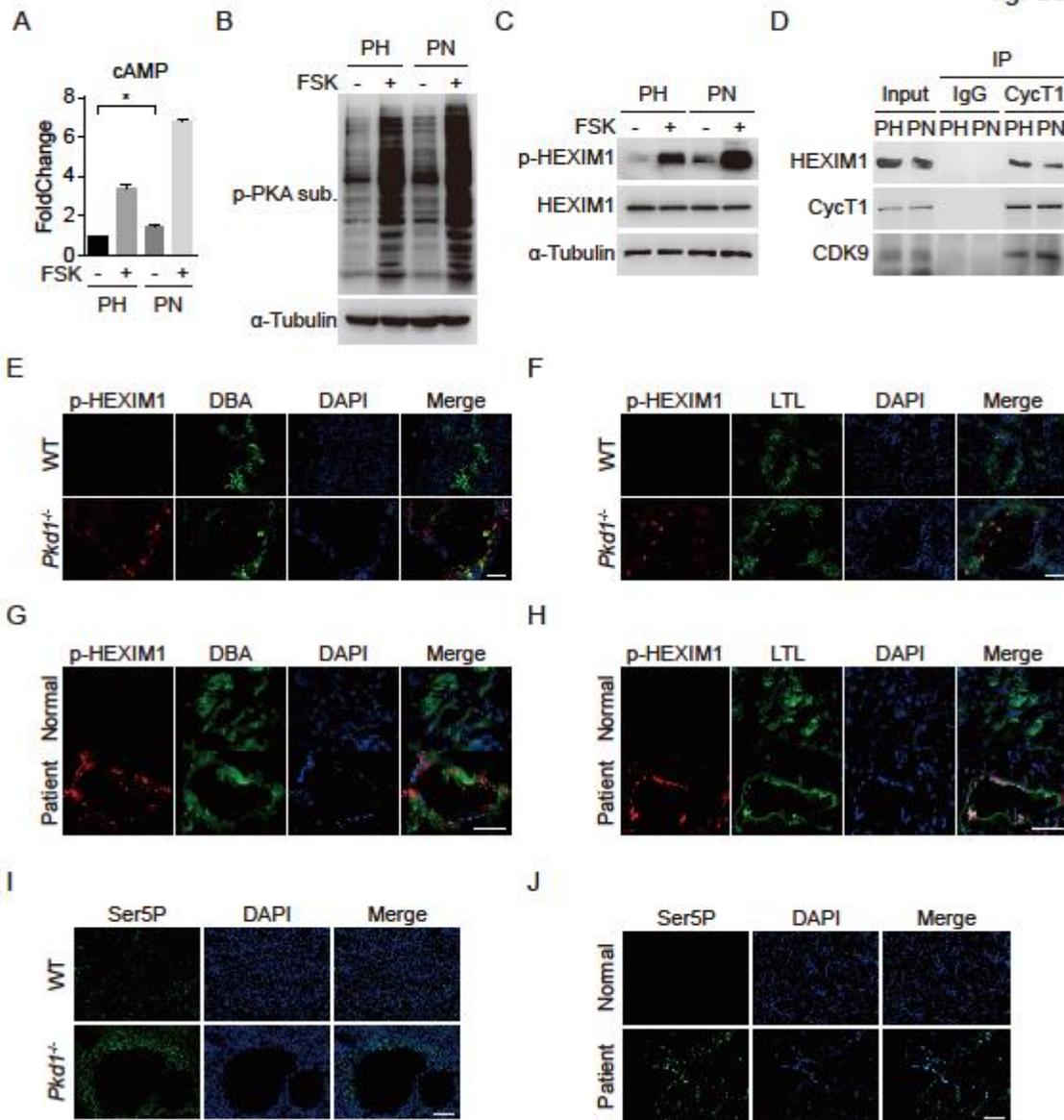
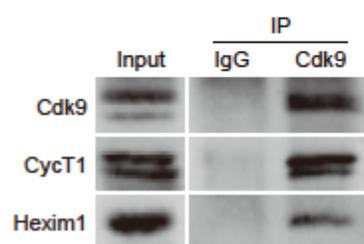


Fig. S3. The phosphorylation level of HEXIM1 is increased in cystic epithelial cells. (A) cAMP levels detection in WT and cystic cells with or without FSK treatment. (B and C) Western blot analysis of PKA activity (B) and p-HEXIM1 (C) in WT and cystic cells with or without FSK treatment. (D) Co-IP assays examining the interactions between P-TEFb and HEXIM1 in WT and cystic cells. (E and F) Immunofluorescence analysis of p-HEXIM1 levels in the DBA-positive (E) or LTL-positive (F) kidney tissues from WT and *Pkd1*^{-/-} mice. (G and H) Immunofluorescence analysis of p-HEXIM1 levels in the DBA-positive (G) or LTL-positive (H) tissues from human kidneys. (I) Immunofluorescence analysis of Pol II Ser5P levels in mice kidneys. (J) Immunofluorescence analysis of Pol II Ser5P levels in human kidneys. Scale bars: 50 μm.

A



B

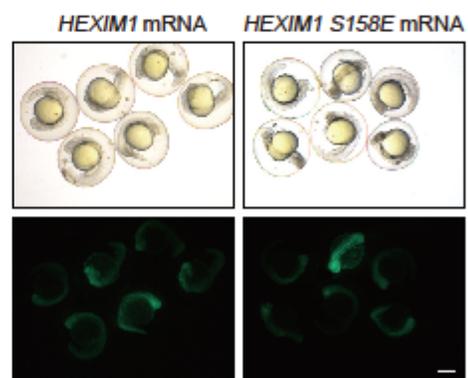


Fig. S4. The interaction between P-TEFb and Hexim1 is conserved in zebrafish. (A) Co-IP assays examining the interactions between zebrafish P-TEFb and Hexim1. (B) Live cell images showing EGFP-HEXIM1 expression in zebrafish embryos injected with human EGFP-HEXIM1 mRNA (WT and S158E). Scale bar: 300 μ m.

fig. S5

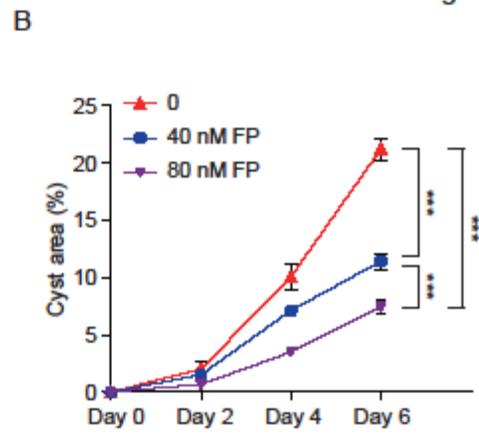
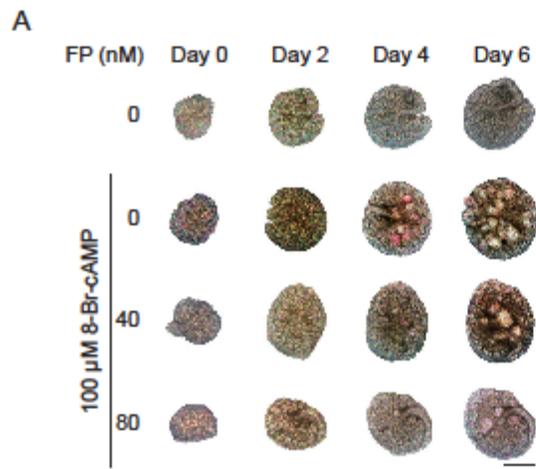


Fig. S5. FP inhibits cystogenesis in the embryonic kidney model. (A) Representative images of mouse embryonic kidneys (E13.5) treated with the indicated doses of Flavopiridol. (B) Quantification of the percentage of cyst area relative to total kidney area ($n \geq 3$). Scale bar: 1 mm. Two-way ANOVA with Tukey's post-hoc test was used for statistical analysis.

fig. S6

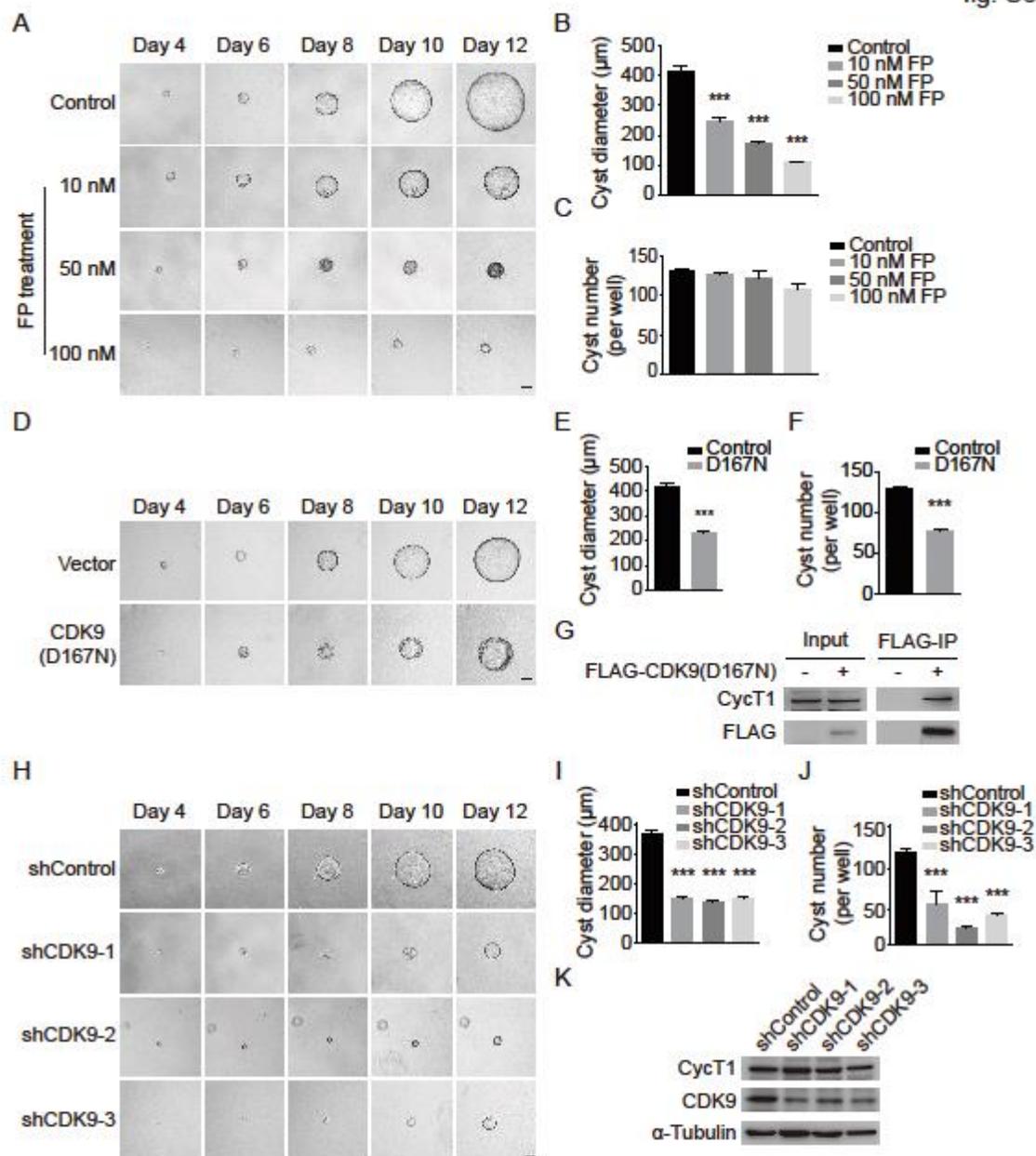


Fig. S6. Inhibition of CDK9 attenuates cyst development in the MDCK cyst model. (A)

Representative images of MDCK cysts treated with the indicated doses of flavopiridol. **(B and C)** Cyst diameter **(B)** and cyst numbers **(C)** of the indicated groups were measured on day 12. **(D)**

Representative images of MDCK cysts transfected with vector or human CDK9 (D167N). **(E and F)**

Cyst diameter **(E)** and cyst numbers **(F)** of the indicated groups were measured on day 12. **(G)**

Co-immunoprecipitation (Co-IP) assays examining the interactions between FLAG-CDK9 D167N and endogenous canine CycT1 in MDCK cells transfected with FLAG-CDK9 D167N. **(H)** Representative

images of MDCK cells infected with lentiviruses carrying control or CDK9 shRNAs. **(I and J)** Cyst diameter **(I)** and cyst numbers **(J)** of the indicated groups were measured on day 12. **(K)**

Immunoblotting analysis of CDK9 knockdown efficiencies in MDCK cells. Data represent as mean \pm SEM. *** $p < 0.001$. Scale bars: 100 μm . Two-tailed unpaired Student's t test was used for statistical analysis.

fig. S7

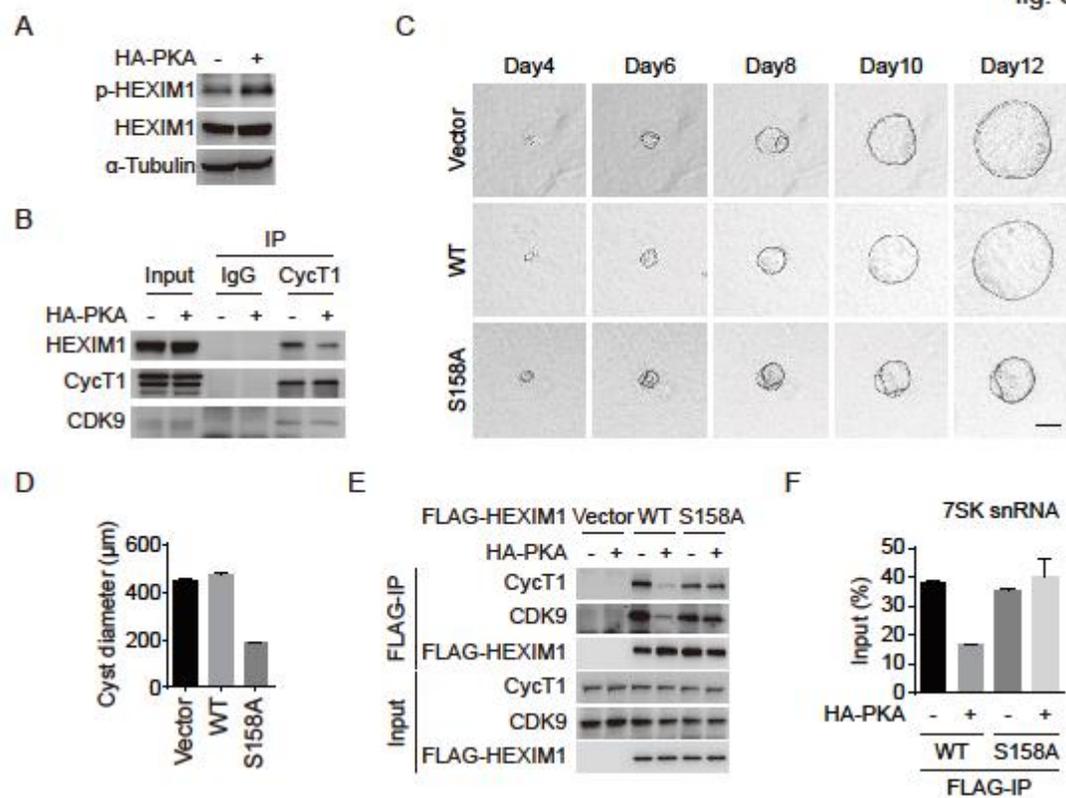


Fig. S7. HEXIM1 phosphorylation and P-TEFb/HEXIM1/7SK snRNP complex assembly in the MDCK cyst model. (A) Western blot analysis of p-HEXIM1 in MDCK transfected with or without PKA. (B) Co-IP assays examining the interactions between P-TEFb and HEXIM1 in MDCK transfected with or without PKA. (C) Representative images of MDCK cysts transfected with vector or human HEXIM1 (WT or S158A). (D) Cyst diameter of the indicated groups were measured on day 12. (E) Co-IP analysis of the interactions between the indicated FLAG-HEXIM1 and P-TEFb in MDCK cells transfected with or without PKA. (F) RNA-IP assays examining the associations between the indicated FLAG-HEXIM1 and 7SK snRNA in MDCK cells transfected with or without PKA. Scale bar: 100 μ m.

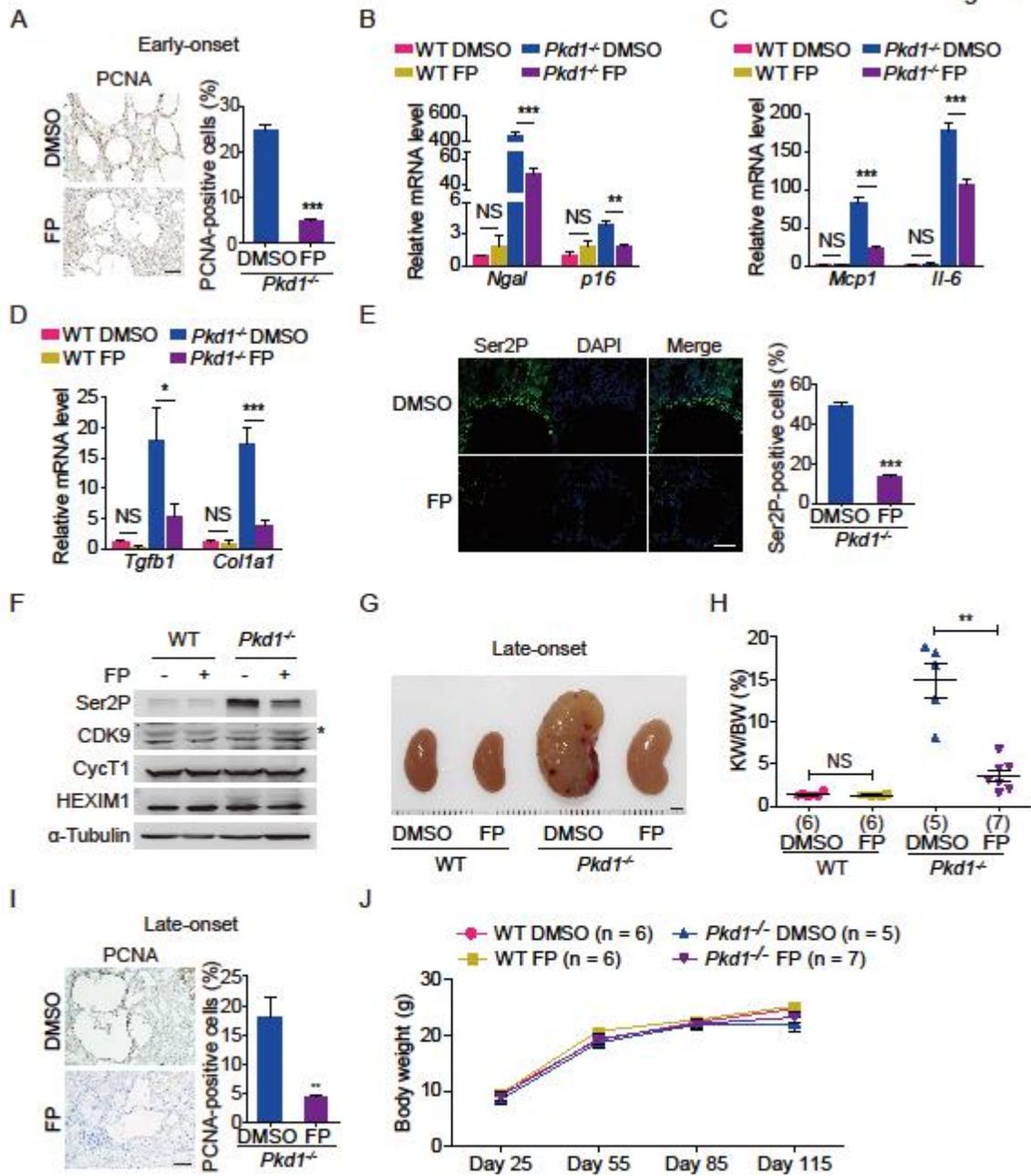


Fig. S8. Effect of FP on cystogenesis in the early- and late-onset ADPKD mouse models. (A)

Immunostaining of PCNA in P29 mice from the indicated groups. **(B-D)** Quantitative RT-PCR analysis of relative mRNA levels of injury markers **(B)**, inflammatory markers **(C)** and fibrotic markers **(D)** in kidneys from the indicated groups. **(E)** Immunofluorescence analysis of Ser2P Pol II expression in kidneys from *Pkd1*^{-/-} mice treated with DMSO or FP. **(F)** Western blot analysis of P-TEFb, Ser2P Pol II and HEXIM1 expression in kidneys from the indicated groups. **(G)** Representative images of P115 kidneys from the indicated groups. **(H)** Ratios of kidney-weight to body-weight in the indicated groups of P115 mice. **(I)** Immunostaining of PCNA in P115 kidneys from *Pkd1*^{-/-} mice treated with DMSO or FP. **(J)** Growth curve of mouse body weight in late-onset ADPKD model. *, non-specific bands. Scale bars: 100 μ m. Data represent the mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$. Two-tailed unpaired Student's t test was used for statistical analysis. (Photo credit: Zhiheng Liu, Tianjin Medical University).