

Figure S1, Related to Fig. 1: Distribution of ILC2s in cuffs of multiple non-barrier organs. (**A**). Schematic of IL-5Cre-tdtomato; R26-CAG-tdtomato (Ai14) lineage-tracker-mouse reporter line. (**B and C**) Flow cytometry analysis of mouse from (**A**) demonstrating >90% specificity (**B**) and ~90% sensitivity (**C**) of the reporter in lung, perigonadal adipose tissue (GAT), and spleen (Spln). (**D**) 2D thin-cut lung imaging of ILC2s (IL-5RFP⁺) in Collagen IV-negative arterial adventitial cuff. (**E**) 2D thin-cut imaging of ILC2s (IL-5 RFP⁺) and rare Th2 cells (IL-5 RFP⁺ CD4⁺, yellow arrowheads) in cuffs from pancreas, mesenteric adipose tissue (MAT), liver, uterus, small intestine (SI), and spleen. (**F**) 2D thin-cut imaging of (ILC2) IL-5 RFP⁺ cells and CCL19-Cre; R26-YFP⁺ SC in lung cuffs. (**B-F**) Images and flow cytometry plots are representative of three or more mice.



Figure S2, **Related to Fig. 1:** Distribution of ILC2s in epithelial, serosal body-cavity sites, and secondary lymphoid organs (SLOs). (**A-C**) 2D thin-cut imaging of ILC2 (IL- $5RFP^+$) in 'epithelial tissues', including (**A**) stomach at the boundary of the muscularis and mucosa layers, (**B**) small intestine (SI) villus and epithelial crypts (marked with Ki-67), and (**C**) skin hair follicles. (**D-E**) 2D thin cut-imaging of ILC2 (IL- $5RFP^+$) in SLOs, including (**D**) mesenteric lymph node (MLN) and (**E**) peyers patches of the SI. (**F-I**) 2D thin-cut imaging of ILC2 in serosal body cavity membranes, including (**F**) peritoneal membrane, (**G**) visceral pleura of the lung, (**H**) pericardium of the heart, or (**I**) peritoneal surface of the uterus. In some cases, PDGFR α GFP⁺ stromal cells were also visualized as noted. Images are representative of two or more mice.



Figure S3, Related to Figs 1-3: (A) 2D thin-cut imaging of ILC2 (IL-5RFP+ CD45+) and dendritic cells (DCs; CD45⁺ CD11c⁺ MHCII⁺) in gonadal adipose tissue (GAT), pancreas, stomach and lung. (B) 3D imaging from lung of a 200µm projection of (top) a large lung survey with PDGFRaGFP, SMA, IL-5 RFP, and Lyve1 showing airways, arteries, veins, and lymphatics. Images were analyzed by utilizing co-localization channels and surfacing in Imaris software. Surfaces were rendered on SMA⁺ airways, SMA⁺, Lyve1⁺ blood vessels, SMA⁻, Lyve1⁺ lymphatic vessels, GFP⁺ PDGFRa stromal cells, and IL-5 RFP⁺ ILC2s. (Bottom) A close up of an airway-artery bundle with surfaces in place. (C) Analysis of 3D images related to Fig 3A-D, demonstrating number of contacts per ILC2 (IL-5 RFP⁺) with the indicated stromal subset. (**D-G**) Localization of ILC2s, DC and Treg subsets, and IL-33+ ASCs in indicated tissues. (F) 3D rendering with zoom of a 200µm thick projection with IL-5 RFP+ ILC2, Tregs (FoxP3GFP+), and DCs (CD11c⁺ MHCII⁺). (D,G) 2D thin cut images of liver and (E) pancreas identified ILC2 (IL-5 RFP⁺) and Tregs (CD3²⁺ FoxP3GFP⁺) in proximity to IL-33⁺ ASCs. (H-K) 2Dthin cut images from tissues showing localization of PDGFRaGFP cells and IL-5RFP+ ILC2s during postnatal developmental time points indicated; (H) P5 lung, (I) P10 liver, or (K) P2 small intestine. (J) Quantification of localization of cuff (left) and sinusoidal (blood-borne, right) ILC2s (IL5 RFP⁺) with PDGFRaGFP⁺ SCs in P10 liver. Images are representative of three or more mice.



Figure S4, Related to Fig 4: IL-33mcherry reporter construction, validation, and analysis via imaging and flow cytometry (**A**) Diagram of the construction for the IL-33-H2B-mcherry nuclear localization IL-33 reporter (**B**) qPCR validation of *II33* transcript from sorted cells of the indicated IL-33mcherry⁺ (red) and cherry- (black) populations. (**C**) Flow cytometry analysis of lungs from IL-33mcherry heterozygous mice, demonstrating (top) subsets of epithelia, endothelia (blood BEC and lymphatic LEC), and stromal subsets, with (bottom) breakdown of all IL-33mcherry⁺ cells within these categories. (**D**) Flow cytometry from indicated tissues pre-gated on all PDGFR α GFP⁺ stromal cells and demonstrating IL-33mcherry subset expression (or reporter negative wild-type gating control). (**F**-**G**) Thin-cut imaging of IL-33 expression in adventitial cuffs of (**E**) uterus (non-pregnant), (**F**) spleen, and (**G**) pancreas revealed PDGFR α^+ IL-33⁺ cells present in the cuff. (**H**) Flow cytometry analysis of pancreas from IL-33mcherry homozygous (red) or wild-type control (grey) mice identified a dominant mesenchymal subset of Sca1⁺ IL-33⁺ ASCs. Images and flow cytometry plots are representative of 3 experiments. (**B**) pooled data from 3 independent experiments.



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ISNE_1

Figure S5, Related to Fig 5: scRNA seq of adipose tissue ASCs resemble lung ASCs. (A) Heatmap of mean expression values of top 50 genes in lung ASCs (cluster 5). Data is log-transformed. (B) Validation of Col1a1 and Ccl11 mRNA expression by qPCR on sorted IL-33⁺ and IL-33⁻ subsets of lung mesenchymal (PDGFR α^+), epithelial (EpCAM⁺) and endothelial (CD31⁺) cells. (C) Unsupervised clustering analysis of IL-33mcherry+ CD45- cells from perigonadal adipose tissue (GAT) visualized with t-SNE. Each dot indicates an individual cell (total cell number: 2037). (D) Heatmap of mean expression values of top 50 genes in GAT ASCs. Data is log- transformed. (E) Gene expression of selected genes projected onto tSNE plots. (F) Heatmap of mean expression values of top 50 genes in GAT ASCs projected on total lung CD45⁻ cell clusters. Data is logtransformed and row normalized. Stars indicate the three lung mesenchymal clusters (4, 5, and 9) and correspond to those described in Fig 5E. (G) Unsupervised clustering analysis of aggregated lung samples (IL-33mcherry⁺ CD45⁻ cells) from naïve and 4 weeks post N. Brasiliensis infection visualized with t-SNE. Each dot indicates an individual cell (total cell number: 5287). Cells are colored based on clusters (left panel) or sample origin (right panel).



Control ILC2 Deficient

Figure S6, Related to Fig 6: (A and B) Monolayers of ASCs (PDGFRa⁺Sca1⁺, orange) and parenchymal enriched stromal cells (PDGFRa⁺Sca⁻, blue) at (A) ILC2 seeding and (B) enumerated 7 days post-seeding. (C) Log-normalized Tslp reads per cell from lung IL-33mcherry⁺ scRNAseg clusters defined in Fig 5A. (**D**) ILC2s and (**E**) TSLP amounts in ASC-ILC2 7-day co-cultures from wild-type (WT), IL-33 deficient, or wild-type treated with anti-TSLP blocking antibody. (F) Day 5 ILC2 proliferation (CTV dilution) and apoptosis (annexin V) analysis from ILC2 supernatant cultures +/- anti-TSLP. (G and H) ASC expression of IL-13 receptor components, with (G) IL4R α , shown by flow cytometry, or (H) *II13ra*, shown as log-normalized reads from scRNAseg in Fig 5E. (I-J) ASCs IL-33mcherry expression (GMFI) and (K) culture supernatant TSLP amounts after co-culture with the indicated lymphocytes or treatment with the indicated cytokines at 10ng/mL for (I) 5 or (J and K) 3 days. (L) IL-33mCherry expression and frequency of eosinophils (percent of CD45⁺ cells) in lungs from IL-33mcherry heterozygous mice treated with IL-13 or IL-4complex i.n. for seven days, analyzed 20hrs post final dose. (M and N) Thin cut lung images from naive mice and day 28 post *N. brasiliensis* infection and stained for the indicated markers, demonstrating accumulation of cuff ILC2s (IL5RFP⁺ CD3e⁻), IL-5⁺ Th2 cells (IL5RFP+ CD3e⁺), and total CD45⁺ hematopoietic cells. (O) ILC2s (IL-5RFP⁺ CD3e⁻) in spleen day 28 post N. brasiliensis or day 6 post IL-33 treatment (500ng x3). (P-U) Flow cytometry analysis of lungs from the indicated time points post N. brasiliensis infection, demonstrating (P) relative percent of PDGFR α GFP¹⁰ cuff ASCs, (**Q**) total stromal cell IL-33mCherry expression, or (**R-U**) post-infection frequencies of (R) ILC2s, (S) Th2 cells, (T) GFP-dim ASCs, or (U) GFPdim GP38+ ASC subset from control (*II5-cre*) or ILC2 deficient (*II5-cre*;R26-DTA) mice. All images and flow cytometry plots are representative of three or more mice, bar graphs are pooled data from 2-3 experiments.



Figure S7, Related to Fig 7: (**A**) 2D thin-cut images of tamoxifen treated (x3 doses) *Gli1-cre*ERT2; R26-YFP from perigonadal adipose tissue (GAT), pancreas, spleen, and liver. (**B and C**) Percentage Gli1⁻YFP⁺ cells of indicated populations analyzed by flow cytometry from naive mice (black), IL-33 treated (x3 500ng doses over 6 days, grey), or day 12 post *N. brasiliensis* infection (blue). (**D**) qPCR analysis of *Gli1* from the indicated sorted SC subsets from lung (black, IL-33mcherry-negative; red, IL-33mcherry⁺). (**E and**

F) Flow cytometry analysis of ILC2 numbers (**E**) one week or (**F**) four weeks post tamoxifen induction of Gli1⁺ cell deletion. (**G-L**) Flow cytometry analysis of tamoxifen treated Gli1-deleter mice and littermate controls after (**J-L**) IL-33 treatment (500ng x3 and PBS controls or (**G-I**) *N. brasiliensis* infection PI D12-14. (**M**) qPCR on sorted stromal cells from lungs of *Gli1-cre*ERT2; IL-33 Flox mice. (**N**) 2D thin-cut imaging of lungs from *Gli1-cre*ERT2; IL-33 Flox and Cre-negative littermate controls, stained for IL-33. (**A**,**N**) Representative of three or more mice. (**B and C**) One representative of three experiments shown, (**D-M**) pooled data from three or more independent experiments, with individual mice shown. * p<0.05, ** p<0.01, or indicated.

Table S1, Related to Fig 5: Complete scRNAseq results for all clusters defined. Tabs for 1) lung IL-33mcherry⁺ 2) lung total CD45⁻ 3) GAT IL-33mcherry⁺ 4) Lung naïve and day 30 PI *N. brasiliensis* 5) DAVID GO analysis for total lung cluster 5 ASCs.

Movie S1, Related to Fig 1: Meningeal ILC2s: 3D

Movie S2, Related to Fig 1: Pancreas ILC2s: serial Z stack sections

Movie S3, Related to Fig 3: Lung ILC2s with cuff stromal cells: serial Z stack sections

Movie S4, Related to Fig 3: GAT ILC2s with IL-33 expressing stroma: 3D