Thioredoxin-1 improves the immuno-metabolic phenotype of anti-tumor T cells

Paramita Chakraborty^{1#}, Shilpak Chatterjee^{1#}, Pravin Kesarwani^{1#}, Krishnamurthy Thyagarajan, Supinya lamsawat², Annika Dalheim³, Hung Nguyen², Shanmugam P. Selvam⁴, Patrick Nassare¹, Gina Scurti³, Gary Hardiman⁵, Nilanjana Maulik⁶, Lauren Ball⁷, Vamsi Gangaraju⁴, Mark P. Rubinstein¹, Nancy Klauber-DeMore¹, Elizabeth G. Hill⁸, Besim Ogretmen⁴, Xue-Zhong Yu², Michael I. Nishimura³, Shikhar Mehrotra^{*}

Departments of ¹Surgery, ²Microbiology & Immunology, ⁴Biochemistry & Molecular Biology, ⁵Nephrology, ⁷Pharmaceutical and Biomedical Sciences, ⁸Public Health, Hollings Cancer Center, Medical University of South Carolina, Charleston, 29425; ³Department of Surgery, Loyola University, Maywood, IL 60153; and ⁶Department of Surgery, University of Connecticut Health Center, 263 Farmington Avenue, Farmington, CT 06030.

[#]Shared first authors

Table of contents

- Supplementary Figure 1
- Supplementary Figure 2
- Supplementary Figure 3
- Supplementary Figure 4

Figure S1



Figure S1: Characterization of Pmel-Trx T cells. *A*). The gel picture shows PCR based genotyping for Pmel and Pmel-Trx mice. *B*) Thymocytes were prepared from the six-week old Pmel and Pmel-Trx mice and stained for flurochrome conjugated CD4 and CD8 antibody. The dot-plot represents distribution of CD4 and CD8 single positives (SP) and double positive (DP) thymocytes obtained from two different mice. *Lower panel* shows percent of each fraction (CD4 SP, CD8 SP, CD4CD8 DP) represented in bar diagram using the data from *upper panel* dot plots. ***p* value <0.01. *C*) Three day TCR activated T cells from Pmel and Pmel-Trx mice were restimulated with cognate antigen for 4 hr. and stained intracellularly using flurochrome conjuagetd anti-RIPK1 antibody.

Figure S2





Figure S2: Gene perturbation bar plot. All the genes in different pathways are ranked based on their absolute perturbation values computed by propagating the measured expression changes across the pathway topology. For each gene, the signed perturbation is represented with negative values in blue and positive values in red. The box and whisker plot on the left summarizes the distribution of all gene perturbations in this pathway. The box represents the 1st quartile, the median and the 3rd quartile, while circles represent the outliers.















Figure S3

Figure S3: Effect of rTrx on pS6 and iTreg generation. Splenic T cells obtained from Pmel transgenic mice were stimulated with cognate antigen gp100 for three days either alone or in presence of rTrx, GSH, or L-NAC before evaluating for the expression levels of: A) pS6, and B) Trx. Adjacent bar represents cumulative data from three repeat experiments. p value *< 0.05, **< 0.01. C) Purified CD4⁺ T cells from EGFP-FoxP3-knockin mouse splenocytes were used for in vitro differentiation to induced regulatory T cells (iTreg's) by activating with plate-bound anti-CD3 (5 µg/ ml), anti-CD28 (5 µg/ml) in presence TGF_β (4ng/ml), IL2 (100 unit/ml) for three days either in absence or presence of recombinant thioredoxin (rTrx). Left panel demonstrates expression of CD4 and FoxP3-GFP using BD FACS Accuri. Right panel represents iTreg's function in a T cell suppression assay that was performed using C57BL/6 mouse derived splenic CD4⁺CD25^{neg} T cells pre-labeled with CFSE dye (0.5µM) and co-cultured with Treg's at 1:1 ratio (100K each), and soluble anti-CD3 (2 µg/ml). After three days in co-culture the proliferation of CFSE labeled T cells was determined using FACS. Data was analyzed using FloJo software. Similar data was obtained in two independent experiments. D-F) C57BL/6 mice (n=3 mice/group) were inoculated (s.c) with 0.25×106 B16-F10 melanoma cells for 14 days, after which mice were adoptively transferred with 1×10⁶ three day gp100 activated Pmel T cells, Pmel cells treated with rTrx (10 µg/ml). After 12 days of T cells transfer, lymphocytes were retrieved from the excised tumor and the indicated lymphoid organs. FACS plot is shown for the retrieved lymphocytes that were stimulated overnight with aCD3 (2 µg/ ml) and α CD28 antibody (5 µg/ml) before staining with flurochrome-conjugated antibodies to determine intracellular IFNγ level (**D**), and Granzyme B (GzmB) (**E**) level. N=3. *p<0.05, **p<0.005. F) Upper panel shows the FACS plot obtained for determining the regulatory T cells by intracellular FoxP3 staining on CD4⁺CD25⁺ fraction using the retrieved lymphocytes (TILs, DLN, spleen). *Lower* panel represent the cumulative data from different recipient mice. N=3. *p<0.05, **p<0.005.



Figure S4: Characterization of the human T cells transduced with tyrosinase reactive TIL1383I TCR construct with Trx gene insert. Human T cells from normal healthy donor peripheral blood were retrovirally transduced with TIL1383I+Trx vector (*A*) or TIL1383I vector, and characterized for: *B*) transduction efficiency based on CD34 expression, *C*) CD62L, CD44, CD45RA, CD45RO expression, and *D*) Glucose uptake using 2NBDG assay. Cells were restimulated overnight with cognate hTyr peptide antigen using T2 cells for determining: *E*) Mitochondrial membrane potential using TMRM, and *F*) NO secretion using DAF. *G*) RNA from TCR transduced cells after REP was used to determine expression of 'stemness' genes using qPCR. *N=2. *p* value < 0.05; **p value < 0.01, ***p value < 0.005.