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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see <u>Authors & Referees</u> and the <u>Editorial Policy Checklist</u>.

Statistics

For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	\square The exact sample size (<i>n</i>) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted Give P values as exact values whenever suitable.
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on statistics for biologists contains articles on many of the points above

Software and code

Policy information about <u>availability of computer code</u>					
Data collection	We used PrairieView and LabView to acquire the 2-photon Ca imaging data. For 2-photon targeted electrophysiology, ScanImage and Clampex were used for data acquisition. Python was used for visual stimuli presentation.				
Data analysis	We used Matlab and Clampfit to analyze data. Chronux package was used for coherence estimates.				

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The authors declare that all data supporting the findings of this investigation are available within the article, its Supplementary Information, and from the corresponding authors upon reasonable request.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences

Behavioural & social sciences

Ecological, evolutionary & environmental sciences For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.							
Sample size	Sample sizes were chosen based on the numbers typically presented in the published studies in the field. More specifically, sample size of Ca imaging was made sufficiently large (e.g. > 300 ROIs) but not explicitly chosen. For 2-photon targeted whole-cell recordings, we collected data from as many cells as possible. In addition, a specific number of every line of transgenic mice were provided.						
Data exclusions	Whole-cell data of some patched neurons were excluded from final analysis based on pre-established criteria indicating bad recording quality.						
Replication	All attempts at replication were successful.						
Randomization	Due to the usage of individual transgenic mouse lines, no randomization of samples/organisms/participants was necessary.						
Blinding	No blinding was done.						

Reporting for specific materials, systems and methods

Methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study	n/a	Involved in the study
\ge	Antibodies	\boxtimes	ChIP-seq
\boxtimes	Eukaryotic cell lines	\boxtimes	Flow cytometry
\boxtimes	Palaeontology	\ge	MRI-based neuroimaging
	Animals and other organisms		
\ge	Human research participants		
\times	Clinical data		

Animals and other organisms

Policy information about <u>studies involving animals</u> ; <u>ARRIVE guidelines</u> recommended for reporting animal research						
Laboratory animals	Mouse, various transgenic lines with C57BI/6 genetic background, 2 to 6 months old, both sex.					
Wild animals	No wild animals were used in this study.					
Field-collected samples	No field-collected samples were used in this study.					
Ethics oversight	None					

Note that full information on the approval of the study protocol must also be provided in the manuscript.