# **SUPPORTING INFORMATION**

# **An unbiased lipid phenotyping approach to study the genetic determinants of lipids and their association with coronary heart disease risk factors**

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### **Supplementary Methods.** Technical discussion on annotation of lipid signals

Lipid signals were annotated to a molecular formula on the basis of the accurate mass. The resolution of 65,000 at 400 *m/z,* as used in this study, allowed for the baseline separation of, for instance, molecular formulae  $C_{41}H_{78}NPO_8 + H^+$  ( $m/z$  744.554) and C<sub>42</sub>H<sub>82</sub>NPO<sub>7</sub>+H<sup>+</sup> (*m/z* 744.590), but was unable to determine if the former was PC(33:2) or PE(36:2) as the species are isobaric. The average mass accuracy error in the measurement of the *m/z* was 0.85 ppm across all intact lipids, with the highest difference for detected lipids of 2.69 ppm for PC ae (37:4). However, this did not mean that only one lipid species contributed to a specific mass-to-charge ratio. For example, the ion that we identified as TG(52:2) with *m/z* 876.802 could be many different triglyceride lipids [e.g. TG(16:0/18:2/18:0), TG(14:0/16:0/22:2) or TG(16:0/18:1/18:1)], all of which have the same molecular weight. Interpretation of species at the chemical formula level allowed us to model changes within lipid pools according to biological processes such as chain elongation and desaturation. We assumed that a given signal peak was likely to be a combination of several lipid species. Our annotation was further based on fragmentation data for the most predominant ion through additional LC-MS/MS analyses.

Because the "open-profiling" approach did not predetermine which lipid species would be detected, it provided data on all ionisable molecules and was therefore very sensitive to contamination, especially of compounds with high ionization efficiencies. In all analyses, we found adipates (*m/z* 371.316) and organophosphates, such as Tris(ditertbutylphenyl)phosphite (*m/z* 647.459) and its oxidation products (*m/z* 663.454), that leached from plastics into the organic solvents. However, using glass-coated well-plates minimized the contact time of the samples with the plastics, and by using blanks and QCs at three different concentrations, we were able to exclude the contaminating ions (approximately half of all the signals) from the final data set. The use of glass-coated well plates was essential to obtain both precise and reliable data. Furthermore, as the method relied on nanoflow, contaminants had minimal impact on the ionization efficiency.

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The developed peak-picking algorithm enabled us to process the almost seven thousand data files using parallelization with a processing time of about four minutes per sample per core. The analysis time was greatly sped up by processing each file independently since there was no requirement to load all files jointly into memory to perform the alignment; the ability to perform analyses in parallel also greatly sped up the analysis time. This approach is only suitable to compare similar samples where the same lipids are expected, as it requires known lipids with their target *m/z*. The results required manual curation as in certain cases the target *m/z* was too close to an isotope or adduct of another lipid. We therefore confirmed the identity of all the ions that passed the QC filter, and selected samples were analysed by high-resolution LC-MS/MS to confirm lipid annotations.

**Supplementary Table.** Summary of genetic associations of principal components of

lipids







The association of the second, third, and fourth principal components of lipids with genetic variants is shown in order of *P*-value of significance within each principal component and genetic locus. The lead variant (SNP most strongly associated with each principal component) within each locus is highlighted. Abbreviations: EA = effect allele; GRCh37 = Genome Reference Consortium human genome build 37; NEA = non-effect allele; SE = standard error; SNP = single nucleotide polymorphism.

# **Supplementary Figure 1.** Extent of missing data according to the number of lipids and





(a) Percentage of lipids with missing data per participant. This subfigure shows that 5,328 out of 5,662 participants had less than 10% (0-9%) missing data, while 144 participants had 90-100% missing data. (b) Percentage of lipids with missing data per lipid. This subfigure shows that 427 out of 444 lipid peaks had less than 10% (0-9%) missing data, while there were 17 lipids that were missing in 10% or more of the lipids.



**Supplementary Figure 2.** Coefficients of variation and normalised relative intensities of

lipids

(a) The coefficients of variation for each lipid expressed against the relative intensity for the quality control samples. (b) Normalised relative intensities of lipids.

**Supplementary Figure 3.** Partial correlation coefficients of phosphatidylcholines and triacylglycerols with levels of major lipid markers



The partial correlations of (a) phosphatidylcholines and (b) triglycerides with total cholesterol, HDL-C, LDL-C, and total triacylglycerols are shown. All analyses were adjusted for age and sex.



**Supplementary Figure 4.** Association of lipids with smoking status and physical activity

All analyses were adjusted for age and sex. Out of the lipids that were associated with rs662799 in the *APOA5-APOC3* locus, results are shown for (a) the top twenty lipids that were most significantly associated with smoking status and (b) the top twenty lipids that were most significantly associated with physical activity.



# **Supplementary Figure 5.** Scree plot of eigenvalues from principal component analysis

**Supplementary Figure 6.** Scatter plots showing matrix loadings of normalised relative intensities of lipids from principal component analysis



Lipids are coloured according to overall lipid category. (a) Overall lipids. (b) Subset of triacylglycerols. Blue oval indicates triacylglycerols containing odd-chain fatty acids increased by dairy consumption; orange oval indicates triacylglycerols containing ω-3 and ω-6 polyunsaturated fatty acids increased by fish consumption; green oval indicates triacylglycerols containing even-chain fatty acids formed in part through *de novo* lipogenesis. Pink ovals represent one or more distinct categories. (c) Subset of lipids associated with rs662799 in the *APOA5-APOC3* locus. Scatter plot is shown for the subset of lipids that are significantly associated with rs662799 (chr11:11663707) in the *APOA5- APOC3* locus at *P* < 8.9 x 10-10.

**Supplementary Figure 7.** Association of established coronary heart disease risk factors

with principal components of lipid levels



All analyses were adjusted for age and sex. Odds ratios (OR) and 95% confidence intervals (CI) for each principal component are expressed per 1-SD increase in the loading scores of the lipids that make up that component. Definitions: Diabetes =  $HbA_{1c} \ge 6.5\%$ ; Hypertension = SBP  $\geq$  140 mm Hg or DBP  $\geq$  90 mm Hg; Obese = BMI  $\geq$  30 kg/m<sup>2</sup>; Overweight =  $BMI \geq 25$  kg/m<sup>2</sup>. Abbreviations: BMI = body mass index; DBP = diastolic blood pressure; SBP = systolic blood pressure.



**Supplementary Figure 8**. Manhattan and regional association plots of principal components of lipid levels

Regional association plots were produced using LocusZoom. (a) Second principal component derived from lipid levels. (b) Third principal component derived from lipid levels. (c) Fourth principal component derived from lipid levels.