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Abstract:	<p>With the decreasing cost of sequencing and the rapid developments in genomics technologies and protocols, the need for validated bioinformatics software that enables efficient large-scale data processing is growing. Here we present GenPipes, a flexible Python-based framework that facilitates the development and deployment of multi-step workflows optimized for High Performance Computing clusters and the cloud. GenPipes already implements 12 benchmarked and scalable pipelines for various genomics applications, including RNA-Seq, ChIP-Seq, DNA-Seq, Methyl-Seq, Hi-C, capture Hi-C, metagenomics and PacBio long read assembly. The software is available under a GPLv3 open source license and is continuously updated to follow recent advances in genomics and bioinformatics. The framework has been already configured on several servers and a docker image is also available to facilitate additional installations. In summary, GenPipes offers genomic researchers a simple method to analyze different types of data, customizable to their needs and resources, as well as the flexibility to create their own workflows.</p>	
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Response to Reviewers:	<p>Dear Editor,</p> <p>Thank you for the opportunity to submit a revised version of the manuscript GIGA-D-18-00198, which addresses the final points raised by the reviewers. Please find our point-by-point response below. New text that has been added to the revised manuscript is shown in red.</p> <p>Response to the Reviewers:</p> <p>Reviewer #2: I thank the authors for addressing several of my concerns. Two issues remain:</p> <p># Major comments</p> <p>* Regarding the feature table, the reasoning of the authors is acceptable (apart from my concern below), if the caption explicitly mentions that the table is "meant to provide the reader with an overview of the features of several tools in the field but not necessarily an exhaustive list".</p> <p>We have added the suggested text to Table 1's caption as follows:</p> <p>"... It is also worth noting that the following table is meant to provide the reader with an overview of the features of several tools in the field but not necessarily an exhaustive list. For a full description of each tool's capabilities, please consult their official documentation."</p> <p>* Regarding the answer "Bioconda offers a collection of packages and not an integrated system and can be quite heavy in memory requirements. Hence, we think that "package-manager-integration" is not necessarily an indication of the strength of the WMS. It is a specific choice, one that offers ease of installation but has its pitfalls as well. GenPipes does not use package managers by design. GenPipes manages its own libraries making sure there is no conflicting libraries in the process. For users who do not want to install GenPipes manually, we offer a Docker container that has also been tested with Singularity. We have updated the GenPipes' bitbucket documentation to highlight the availability of the GenPipes' Docker container":</p> <p>The way to provide the software stack for an analysis is very important for</p>

	<p>reproducibility and maintainability of a pipeline. Hence, this should definitely be a column of the feature table in any case. Certainly it is legitimate to decide against Conda or other package managers, but in above argument I do not find any reason to hide this aspect from the user in the feature table. Why not adding a column about package manager support and let the reader decide whether they need/want this feature or not?</p> <p>We have added a column, "Package Manager" , to Table1, as suggested by the reviewer.</p>
Additional Information:	
Question	Response
Are you submitting this manuscript to a special series or article collection?	No
<p>Experimental design and statistics</p> <p>Full details of the experimental design and statistical methods used should be given in the Methods section, as detailed in our Minimum Standards Reporting Checklist. Information essential to interpreting the data presented should be made available in the figure legends.</p> <p>Have you included all the information requested in your manuscript?</p>	Yes
<p>Resources</p> <p>A description of all resources used, including antibodies, cell lines, animals and software tools, with enough information to allow them to be uniquely identified, should be included in the Methods section. Authors are strongly encouraged to cite Research Resource Identifiers (RRIDs) for antibodies, model organisms and tools, where possible.</p> <p>Have you included the information requested as detailed in our Minimum Standards Reporting Checklist?</p>	Yes
<p>Availability of data and materials</p> <p>All datasets and code on which the conclusions of the paper rely must be</p>	Yes

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GenPipes: an open-source framework for distributed and scalable genomic analyses

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ABSTRACT

With the decreasing cost of sequencing and the rapid developments in genomics technologies and protocols, the need for validated bioinformatics software that enables efficient large-scale data processing is growing. Here we present GenPipes, a flexible Python-based framework that facilitates the development and deployment of multi-step workflows optimized for High Performance Computing clusters and the cloud. GenPipes already implements 12 validated and scalable pipelines for various genomics applications, including RNA-Seq, ChIP-Seq, DNA-Seq, Methyl-Seq, Hi-C, capture Hi-C, metagenomics and PacBio long read assembly. The software is available under a GPLv3 open source license and is continuously updated to follow recent advances in genomics and bioinformatics. The framework has been already configured on several servers and a docker image is also available to facilitate additional

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4 installations. In summary, GenPipes offers genomic researchers a simple method to analyze different
5 types of data, customizable to their needs and resources, as well as the flexibility to create their own
6 workflows.
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9 **Keywords:** genomics; workflow management systems; frameworks; workflow; pipeline; bioinformatics.
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12 13 14 **INTRODUCTION**

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16 Sequencing has become an indispensable tool in our quest to understand biological processes.
17 Moreover, facilitated by a significant decline in overall costs, new technologies and experimental
18 protocols are being developed at a fast pace. This has resulted in massive amounts of sequencing data
19 being produced and deposited in various public archives. For instance, a number of national initiatives,
20 such as *Genomics England* and *All of US*, plan to sequence hundreds of thousands of individual
21 genomes in an effort to further develop precision medicine. Similarly, a number of large initiatives, such
22 as ENCODE [1] and the International Human Epigenome Consortium (IHEC) [2], plan to generate
23 thousands of epigenomics datasets to better understand gene regulation in normal and disease
24 processes. Despite this rapid progress in sequencing, genomics technologies and available datasets,
25 processing and analyses have struggled to keep up. Indeed, the need for robust, open-source and
26 scalable bioinformatics pipelines has become a major bottleneck for genomics [3].
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34 Available bioinformatics tools for genomic data can be categorized into three different groups: 1)
35 analysis platforms/workbenches, 2) workflow management systems (WMS)/frameworks, and 3) individual
36 analysis pipelines/workflows. Platforms of the first type, like Galaxy [4] or DNA Nexus [5], provide a full
37 workbench for data upload and storage, and are accompanied with a set of available tools. While they
38 provide fast and easy user services, such tools can be inconvenient for large scale projects due to having
39 to move sizeable datasets to the platform. In the second type, WMSs such as Snakemake [6], Nextflow
40 [7], BPIPE [8], BigDataScript [9] and declarative workflow description languages, such as CWL or WDL
41 are dedicated to providing a customizable framework to build bioinformatics pipelines. Such solutions are
42 flexible and can help in pipeline implementation but rarely provide robust pre-built pipelines which are
43 ready for production analysis. Finally, tools of the third type are individual analysis pipelines for various
44 applications that have been validated and published. These are useful for specific applications but can
45 sometimes be challenging to implement, difficult to modify or scale-up. They have also rarely been tested
46 on multiple computing infrastructures.
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55 Here we present GenPipes, an open-source, Python-based WMS for pipeline development. As
56 part of its implementation, GenPipes includes a set of high-quality, standardized analysis pipelines,
57 designed for High Performance Computing (HPC) resources and cloud environments. GenPipes' WMS
58 and pipelines have been tested, benchmarked and used extensively over the past four years. GenPipes
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4 is continuously updated and is configured on several different HPC clusters with different properties. By
5 combining both WMS and extensively validated End-to-End analysis workflows, GenPipes offers turnkey
6 analyses for a wide range of bioinformatics applications in the genomics field while also enabling flexible
7 and robust extensions.
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10 11 12 13 14 15 16 17 18 **MATERIAL AND METHODS**

19 ***Overview of the GenPipes Framework***

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23 GenPipes is an object-oriented framework consisting of Python scripts and libraries which create
24 a list of jobs to be launched as Bash commands (Figure 1). There are four main objects that manage the
25 different components of the analysis workflow, namely, *Pipeline*, *Step*, *Job* and *Scheduler*. The main
26 object is the “*Pipeline*” object which controls the workflow of the analysis. Each specific analysis workflow
27 is thus defined as a specific *Pipeline* object. *Pipeline* objects can inherit from one another. The *Pipeline*
28 object defines the flow of the analysis by calling specific “*Step*” objects. The *Pipeline* instance could call
29 all steps implemented in a pipeline or only a set of steps selected by the user. Each step of a pipeline is a
30 unit block that encapsulates a part of the analysis (e.g., trimming or alignment). The *Step* object is a
31 central unit object which corresponds to a specific analysis task. The execution of the task is directly
32 managed by the code defined in each *Step* instance; some steps may execute their task on each sample
33 individually while other steps execute their task using all the samples collectively. The main purpose of
34 the *Step* object is to generate a list of “*Job*” objects which correspond to the consecutive execution of
35 single tasks. The *Job* object defines the commands that will be submitted to the system. It contains all the
36 elements needed to execute the commands, such as input files, modules to be loaded, as well as job
37 dependencies and temporary files. Each *Job* object will be submitted to the system using a specific
38 “*Scheduler*” object. The *Scheduler* object creates execution commands that are compatible with the
39 user’s computing system. Four different *Scheduler* objects have already been implemented (PBS,
40 SLURM, Batch and Daemon), see below.
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51 GenPipes’ object-oriented framework simplifies the development of new features and its
52 adaptation to new systems; new workflows can be created by implementing a *Pipeline* object which
53 inherits features and steps from other existing *Pipeline* objects. Similarly, deploying GenPipes on a new
54 system may only require the development of the corresponding *Scheduler* object along with specific
55 configuration files. GenPipes’ command execution details have been implemented using a shared library
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4 system which allows the modification of tasks by simply adjusting input parameters. This simplifies code
5 maintenance and makes changes in software versions consistent across all pipelines.
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10 ***Freely distributed and pre-installed on a number of HPC resources***

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12 GenPipes is an open-source framework freely distributed and open for external contributions from
13 the developer community. GenPipes can be installed from scratch on any Linux cluster supporting Python
14 2.7 by following the available instructions (<https://bitbucket.org/mugqic/genpipes/src/master/>). GenPipes
15 can also be used via a Docker image which simplifies the setup process and can be used on a range of
16 platforms, including cloud platforms. This allows system-wide installations, as well as local user
17 installations via the Docker image without needing special permissions.
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23 Through a partnership with the Compute Canada consortium (<https://www.computecanada.ca>),
24 the pipelines and third-party tools have also been configured on 6 different Compute Canada HPC
25 centers. It allows any Canadian researcher to use GenPipes along with the needed computing resources
26 by simply applying to the consortium [10]. To ensure consistency of pipeline versions and used
27 dependencies (such as genome references and annotation files) and to avoid discrepancy between
28 compute sites, pipeline setup has been centralized to one location which is then distributed on a real-time
29 shared file system: the CERN Virtual Machine File System [11].
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37 ***Running GenPipes***

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39 GenPipes is a command line tool. Its use has been simplified to accommodate general users. A
40 full tutorial is available [12]. Briefly, to launch GenPipes, the following is needed:
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- 44 • A readset file that contains information about the samples, indicated using the flag “-r”. GenPipes
45 can aggregate and merge samples as indicated by the readset file.
- 46 • Configuration/ini files that contain parameters related to the cluster and the third-party tools,
47 indicated using the flag “-c”. Configuration files are customizable, allowing users to adjust
48 different parameters.
- 49 • The specific steps to be executed, indicated by the flag “-s”.
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53 The generic command to run GenPipes is:
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55 <pipeline>.py -c myConfigurationFile -r myReadSetFile -s 1-X > Commands.txt && bash Commands.txt  
56  
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58 Where <pipeline> can be any of the 12 available pipelines and X is the step number desired.
59 Commands.txt contains the commands that the system will execute.
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4 Pipelines that conduct sample comparisons, like ChIP-Seq and RNA-Seq, require a design file that
5 describes each contrast. Custom sample groupings can be defined in the design file. Design files are
6 indicated by the flag “-d”. The tumour_pair pipeline requires normal-tumour pairing information provided in
7 a standard CSV file using the “-p” option. For more information on the design file and the content of each
8 file type, please consult the GenPipes tutorial and the online documentation.
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12 When the GenPipes command is launched, required modules and files will be searched for and
13 validated. If all required modules and files are found, the analysis commands will be produced. GenPipes
14 will create a directed acyclic graph (DAG) that defines job dependency based on input and output of each
15 step. For a representation of the DAG of each pipeline, refer to supplementary figures S1-14. Once
16 launched, the jobs are sent to the scheduler and queued. As jobs complete successfully, their dependent
17 jobs are released by the scheduler to run. If a job fails, all its dependent jobs are terminated and an email
18 notification is sent to the user. When GenPipes is re-run, it will detect which steps have successfully
19 completed, as described in section ‘Smart relaunch features’, and skip them but will create the command
20 script for the jobs that were not completed successfully. To force the entire command generation, despite
21 successful completion, the “-f” option should be added.
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28 **RESULTS**

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31 GenPipes was first released in 2014. Since then, it has grown to implement 12 pipelines and is
32 currently installed and maintained on 13 different clusters (Figure 2a-b). GenPipes has been actively used
33 for the last four years to quality control and analyze thousands of samples each year (Figure 2c). It has
34 also been used to analyze data for several large-scale projects such as IHEC [2] and eFORGE [13].
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41 ***Key features of GenPipes***

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43 GenPipes’ framework has been optimized to facilitate large scale data analysis. Several features
44 make this possible (Figure 2a):
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47 **Multiple schedulers**

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49 GenPipes is optimized for HPC processing. It can currently accommodate four different types of
50 schedulers:
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- 52 ● *PBSScheduler* creates a batch script that is compatible with a PBS (TORQUE) system.
- 53 ● *SLURMScheduler* creates a batch script that is compatible with a SLURM system.
- 54 ● *BatchScheduler* creates a batch script which contains all the instructions to run all the jobs one
55 after the other.
- 56 ● *DaemonScheduler* creates a log of the pipeline command in a JSON file.
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Job dependencies

In order to minimize the overall analysis time, GenPipes uses a dependency model based on input files, which is managed at the *Job* object level. A job does not need to wait for the completion of a previous step unless it is dependent on its output. Jobs thus become active and can be executed as soon as all their dependencies are met, regardless of the status of previous jobs or of other samples. Thus, when a pipeline is run on multiple samples, it creates several dependency paths, one per sample, each of which completes at its own pace.

Smart relaunch features

Large scale data analysis is subject to failure which could occur due to system failure (e.g. power outage, system reboot, etc...) or user failure (errors in set parameters, or resources). To limit the micro-management and time required to relaunch the pipeline from scratch, GenPipes includes a system of reporting which provides the status of every job in the analysis in order to facilitate the detection of jobs which have failed. Additionally, a relaunch system is implemented which allows restarting the analysis at the exact state before the failure. The relaunch system uses two features: md5sum hash and time stamps. When GenPipes is launched, a md5sum hash is produced for each command. Upon relaunch following a failure, the newly produced hash is compared to that of the completed job to detect changes in the commands. If the hashes are different, the job is relaunched. To detect updates in input files, GenPipes compares the time stamp on the input and output files of already completed jobs. If the date stamp on the input files is more recent than that on the output files then the job is relaunched. If neither the hash code nor the time stamp flag the job to be relaunched then it is considered complete and up-to-date and it will be skipped in the pipeline restart process.

Configuration files

Running large-scale analyses requires a very large number of parameters to be set. GenPipes implements a superposed configuration system to reduce the time required to set-up or modify parameters needed during the analysis. Configuration files, also referred to as "ini" files, are provided among the arguments of the GenPipes command. These files follow the standard INI format, which was selected for its readability and ease of use by non-expert users. Each pipeline reads all configuration files, one after the other, based on a user defined order. The order is of major importance as the system will overwrite a parameter each time it is specified in a new ini file. The system allows the use of the default configuration files provided in GenPipes alone or in combination with user specific configuration files. Configuration files provided with GenPipes are the result of years of experience along with intensive benchmarking. Additionally, several configuration files adjusted for different compute systems or different model organisms are available. The main advantage of this system is to reduce the users' task; only parameters that need to be modified (e.g. system parameters, genomic resources, user specific

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4 parameters) have to be adjusted during the set-up phase of the analysis. To track and enable
5 reproducibility, GenPipes always outputs a file containing the final list of parameters used for the analysis.
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8 **Choice among multiple inputs**

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10 GenPipes represents a series of *Step* objects that are interdependent based on inputs and
11 outputs. Many of the pipeline steps implemented in GenPipes, represent filtering, manipulation or
12 modification of specific genomics files share common formats (e.g. bam, fastq, vcf). To ensure more
13 flexibility in the analysis, a system of ordered list to be interpreted as input files is used. For a given *Step*,
14 each *Job* can be given a series of inputs. The *Job* will browse its list of possible inputs and will consider
15 them based on the order in the list. The first input file found either on disk or in the overall output list will
16 be chosen as input. The chosen input will determine the dependency of the *Job* to the other *Jobs* in the
17 pipeline. This system is really flexible and allows users to skip specific steps in the pipeline if they
18 consider them unnecessary.
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25 **Customizable workflows**

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27 Despite the benchmarking and testing made on the standard analysis procedures implemented in
28 GenPipes, some users may be interested in modifying pipelines. In order to make GenPipes more
29 flexible, a *protocol* system is used. The system allows the implementation of different workflows into a
30 single *Pipeline* object. As a result, one can replace specific steps by other user specific ones. In that
31 case, the user will only need to implement these new Steps and define an additional protocol which will
32 use part of the initial Steps and the newly developed ones. As an example, this has been used to
33 incorporate the Hi-C analysis workflow and the capture Hi-C analysis workflow into GenPipes' hicseq
34 pipeline. A flag (-t hic or -t capture) can be used to specify the workflow to be executed. This system has
35 been developed to reduce the amount of work for external users that decide to contribute to code
36 development and to limit the number of Pipeline objects to maintain. This will also allow us to provide
37 multiple workflows per pipeline to appeal to different tool preferences in each field.
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45 **Facilitating dependency installation**

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47 Genomic analyses require third party tools, as well as genome sequence files, annotation files
48 and indices. GenPipes comes configured with a large set of reference genomes and their respective
49 annotation files, as well as indices for most aligners. It also includes a large set of third party tools. If
50 GenPipes is being installed from scratch on new clusters, automatic bash scripts that download all tools
51 and genomes are included to ease the setup process. These scripts support local installations without the
52 need for super-user privileges. Tools and dependencies are versioned and are loaded by GenPipes in a
53 version-specific manner. This allows different pipelines to use different software versions based on need.
54 It also allows retention of the same parameters and tools for any given project for reproducibility.
55 GenPipes is also provided as a container version for which no dependency installation is required.
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7 ***Available workflows***
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9 GenPipes implements 12 standardized genomics workflows including: DNA-Seq, Tumour
10 Analysis, RNA-Seq, de novo RNA-Seq, ChIP-Seq, PacBio assembly, Methyl-Seq, Hi-C, capture Hi-C, and
11 Metagenomics (Figure 2c). All pipelines have been implemented following a robust design and
12 development routine by following established gold standards standard operating protocols (SOP). Below
13 we summarize GenPipes' workflows; more details are available in the GenPipes documentation. For more
14 details concerning computational resources used by each pipeline, refer to supplementary Table S1. All
15 workflows accept a bam or a fastq file as input.
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20 *DNA-Seq Pipeline:*
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23 DNA-Seq has been implemented optimizing the GATK best practices SOPs [14]. This procedure
24 entails trimming raw reads derived from whole genome or exome data followed by alignment to a known
25 reference, post alignment refinements and variant calling. Trimmed reads are aligned to a reference by
26 the Burrows-Wheeler Aligner, bwa-mem [15]. Refinements of mismatches near indels and base qualities
27 are performed using GATK indels realignment and base recalibration [14] to improve read quality post
28 alignment. Processed reads are marked as fragment duplicates using picard mark duplicates [14] and
29 SNP and small indels are identified using either GATK haplotype callers or samtools mpileup [16]. The
30 Genome in a Bottle [17] dataset was used to select steps and parameters minimizing the false positive
31 rate and maximizing the true positive variants to achieve a sensitivity of 99.7%, precision of 99.1% and
32 F1-score of 99.4% (For more details, refer to Supplementary Materials). Finally, additional annotations
33 are incorporated using dbNSFP [18] and/or Gemini [19] and quality control metrics are collected at
34 various stages and visualized using MulitQC [20]. This pipeline has two different protocols, the default
35 protocol based on the GATK variant caller, haplotype_caller, ("-t muggic", Figure 3) and one based on the
36 mpileup/bcftools caller ("-t mpileup", Figure S1). Another pipeline that is optimized for deep coverage
37 samples, dnaseq_high_coverage, can be found in Figure S2.
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47 *RNA-Seq Pipeline:*
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49 This pipeline aligns reads with STAR [21] 2-passes mode, assembles transcripts with Cufflinks
50 [22] and performs differential expression with Cuffdiff [23]. In parallel, gene-level expression is quantified
51 using htseq-count [24], which produces raw read counts that are subsequently used for differential gene
52 expression with both DESeq [25] and edgeR [26]. Several common quality metrics (rRNA content,
53 expression saturation estimation etc.) are also calculated through the use of RNA-SeQC [27] and in-
54 house scripts. Gene Ontology terms are also tested for over-representation using GOseq [28]. Expressed
55 short SNVs and indels calling is also performed by this pipeline, which optimizes GATK best practices to
56 reach a sensitivity 92.8%, precision 87.7% and F1-score 90.1%. A schema of pipeline steps can be found
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4 in Figure S3. Another pipeline, rnaseq_light, based on Kallisto [29] and used for quick quality control can
5 be found in Figure S4.
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7 De-Novo RNASeq Pipeline: 8

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10 This pipeline is adapted from the Trinity-Trinotate suggested workflow [30] [31]. It reconstructs
11 transcripts from short reads, predicts proteins and annotates leveraging several databases. Quantification
12 is computed using RSEM and differential expression is tested in a manner identical to the RNA-seq
13 pipeline. We observed that the default parameters of the Trinity suite are very conservative which could
14 result in the loss of low-expressed but biologically relevant transcripts. In order to provide the most
15 complete set of transcripts, the pipeline was designed with lower stringency during the assembly step in
16 order to produce every possible transcript and not miss low expressed mRNA. A stringent filtration step is
17 included afterward in order to provide a set of transcripts that make sense biologically. A schema of
18 pipeline steps can be found in Figure S5.
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24 ChIP-Seq Pipeline: 25

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27 The ChIP-Seq workflow is based on the ENCODE [1] workflow. It aligns reads using the Burrows-
28 Wheeler Aligner. It creates tag directories using Homer [32]. Peaks are called using MACS2 [33] and
29 annotated using Homer. Binding motifs are also identified using Homer. Metrics are calculated based on
30 IHEC requirements [34]. The ChIP-Seq pipeline can also be used for ATAC-Seq samples. However, we
31 are developing a pipeline that is specific to ATAC-Seq. A schema of pipeline steps can be found in Figure
32 S6.
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36 The Tumour Analysis Pipeline: 37

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39 The Tumour Pair workflow inherits the bam processing protocol from DNA-seq implementation to
40 retain the benchmarking optimizations but differs in alignment refinement and mutation identification by
41 maximizing the information utilizing both tumour and normal samples together. The pipeline is based on
42 an ensemble approach, which was optimized using both the DREAM3 challenge [35] and the CEPH
43 mixture datasets to select the best combination of callers for both SNV and SV detection. For SNVs,
44 multiple callers such as GATK mutect2, VarScan2 [36], bcftools and VarDict [37] were combined to
45 achieve a sensitivity of 97.5%, precision of 98.8% and F1-score of 98.1% for variants found in 2 or more
46 callers. Similarly, SVs were identified using multiple callers: DELLY [38], LUMPY [39], WHAM [40],
47 CNVkit [41] and Svaba [42] and combined using MetaSV [43] to achieve a sensitivity of 84.6%, precision
48 of 92.4% and F1-score of 88.3% for duplication variants found in the DREAM3 dataset (For more details,
49 refer to Supplementary Material). The pipeline also integrates specific cancer tools to estimate tumour
50 purity, tumour ploidy of sample pair normal-tumour. Additional annotations are incorporated to the SNV
51 calls using dbNSFP [18] and/or Gemini [19] and quality control metrics were collected at various stages
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4 and visualized using MultiQC [20]. This pipeline has 3 protocols (sv, ensemble or fastpass). Schemas of
5 pipeline steps for the three protocols can be found in Figures S7, 8 and 9.
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7 Whole Genome Bisulfite Seq Pipeline (WGBS or Methyl-Seq):

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10 The Methyl-Seq workflow is adapted from the Bismark pipeline [44]. It aligns paired-end reads
11 with bowtie2 default mode. Duplicates are removed with Picard and methylation calls are extracted using
12 bismark [44]. Wiggle tracks for both read coverage and methylation profile are generated for visualization.
13 Variants calls can be extracted from the WGBS data directly using bisSNP [45]. Bisulfite conversion rates
14 are estimated with lambda genome or from human non-CpG methylation directly. Several metrics based
15 on IHEC requirements are also calculated. Methyl-Seq can also process capture data if provided with a
16 capture bed file. A schema of pipeline steps can be found in Figure S10.
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22 Hi-C Pipeline:

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24 The HiC-Seq workflow aligns reads using HiCUP [46]. It creates tag directories, produces
25 interaction matrices, identifies compartments and significant interactions using Homer. It identifies
26 Topologically Associating Domains using TopDom [47] and RobustTAD (bioRxiv 293175). It also creates
27 “.hic” files using JuiceBox [48] and metrics reports using MultiQC [20]. The HiC-Seq workflow can also
28 process capture Hi-C data with the flag “-t capture” using CHICAGO [49]. Schemas for the HiC and
29 capture HiC protocols of this pipeline can be found in Figure S11 and Figure S12 respectively.
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34 The Metagenomic Pipeline (rRNA gene amplification analysis):

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36 This pipeline is based on the established Qiime procedure [50] for amplicon-based
37 metagenomics. It assembles read pairs using FLASH [51], detects chimeras with uchime [52] and picks
38 OTUs using vsearch [53]. OTUs are then aligned using PyNAST [54] and clustered with FastTree [55].
39 Standard diversity indices, taxonomical assignments and ordinations are then calculated and reported
40 graphically. A schema of pipeline steps can be found in Figure S13.
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48 The PacBio Pipeline:

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50 The PacBio whole genome assembly pipeline is built following the HGAP method [31], including
51 additional features, such as base modification detection
52 (<https://github.com/PacificBiosciences/Bioinformatics-Training/wiki/Methylome-Analysis-Technical-Note>)
53 and genome circularization [56]. De novo assembly is performed using PacBio's SMRT Link software
54 (<https://github.com/PacificBiosciences/SMRT-Link/wiki>). Assembly contigs are generated using HGAP4.
55 Alignments are then corrected and used as seeds by FALCON
56 (<https://github.com/PacificBiosciences/FALCON/wiki>) to create contigs. The resulting contigs are then
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4 polished and processed by “Arrow” (<https://github.com/PacificBiosciences/GenomicConsensus>) which
5 ultimately generates high quality consensus sequences. An optional step allowing assembly
6 circularization is integrated at the end of the pipeline. A schema of pipeline steps can be found in Figure
7 S14.
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10 11 12 13 **Comparison with other solutions for NGS analysis**

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16 Data collected for select tools modified from Griffith & Griffith et al. [57] (Table 1), shows that
17 GenPipes’ strength lies in its robust WMS that comes with one of the most diverse selection of analysis
18 pipelines which have been thoroughly tested. The pipelines in the framework cover a wide range of
19 sequencing applications (Figure 2a). The pipelines are end-to-end workflows running complete
20 bioinformatics analyses. While many available pipelines conclude with a bam file or run limited post-bam
21 analysis steps, the pipelines included in GenPipes are extensive, often having as many as 40 different
22 steps that cover a wide range of post-bam processing. It is important to note that GenPipes, as well as
23 several other WMSs, like Nextflow [58] and SnakeMake [59], support community-developed pipelines,
24 however, those have not been included in the comparison.
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30 GenPipes is compatible with HPC computing, as well as cloud computing [60] and includes a
31 workflow manager that can be adapted to new systems. GenPipes also provides job status tracking
32 through JSON files that can then be displayed on a web portal (an official portal for GenPipes will be
33 released soon). GenPipes’ available pipelines facilitate bioinformatics processing, while the framework
34 makes it flexible for modifications and new implementations.
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37 GenPipes developers offer continuous support through a Google forum page [61] and a help desk
38 email address (pipelines@computationalgenomics.ca). Since the release of version 2.0.0 in 2014, a
39 community of users has run GenPipes to conduct approximately 3000 analyses processing around
40 100,000 samples (Figure 2b-c).
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45 **DISCUSSION and CONCLUSION**

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48 GenPipes is a workflow management system that facilitates building robust genomic workflows.
49 GenPipes is a unique solution which combines both a framework for development and end-to-end
50 analysis pipelines for a very large set of genomics fields. The efficient framework for pipeline
51 development has resulted in a broad community of developers with over 30 active branches and more
52 than 10 forks of the GenPipes repository. GenPipes has several optimized features that adapt it to large
53 scale data analysis, namely:
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58 ● **Multiple schedulers:** GenPipes is optimized for HPC processing. It currently accommodates 4
59 schedulers.
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- **Job dependencies:** GenPipes establishes dependencies among its different steps. This enables launching all the steps at the same time and minimizes queue waiting time and management.
- **Smart relaunch:** GenPipes sets and detects flags at each successful step in the pipeline. This allows the detection of successfully completed steps and easy relaunch of failed steps.
- **Parameter encapsulation:** Genpipes uses a superposed configuration system to parse all required parameters from configuration files. This simplifies the use of the framework and makes it more flexible to user adjustments. Tested configuration files that are tailored to different clusters and different species are included with GenPipes.
- **Diverse inputs:** GenPipes has been developed to launch using different starting inputs, making it more flexible.
- **Flexible workflows:** GenPipes implements a workflow in steps. Users can choose to run specific steps of interest, limiting waste of time and resources.

GenPipes is under continuous development to update established pipelines and to create new pipelines for emerging technologies. For instance, new genomics pipelines are being developed for ATAC-Seq, single cell RNA-Seq and HiChIP. GenPipes is also being redeveloped to use the Common Workflow Language (CWL) to provide a cloud compatible version more seamlessly and more *Scheduler* objects, like DRMAA, are being added to expand compatibility with more platforms. GenPipes has become a reliable bioinformatics solution that has been used in various genomics publications for DNA-Seq [62-69], RNA-Seq [70] and ChIP-Seq [71] analyses. GenPipes is currently available as source code, as well as a Docker image for easy installation and use. GenPipes has been optimized for HPC systems but can run on a laptop computer on small datasets.

Availability and requirements

- Project name: GenPipes
- Project home page: <http://www.c3g.ca/genpipes>
- Operating system(s): Linux; Can be used on Windows and Mac OS using Docker
- Programming language: Python
- Other requirements: Workflow-dependant; detailed in documentation
- License: GNU GPLv3
- SciCrunch RRID: SCR_016376

Availability of Supporting Data

Snapshots of the code are available in the *GigaScience* GigaDB repository[72].

ABBREVIATIONS

CML: Common Workflow Language; DAG: directed acyclic graph; IHEC: International Human Epigenome Consortium; NGS: High Performance Computing; OTU: Operational Taxonomic Unit; WMS: workflow management system

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COMPETING INTERESTS

The Authors declare no competing interests.

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30 **TABLE AND FIGURES LEGENDS**

31 32 33 **Figure 1 - General workflow of GenPipes**

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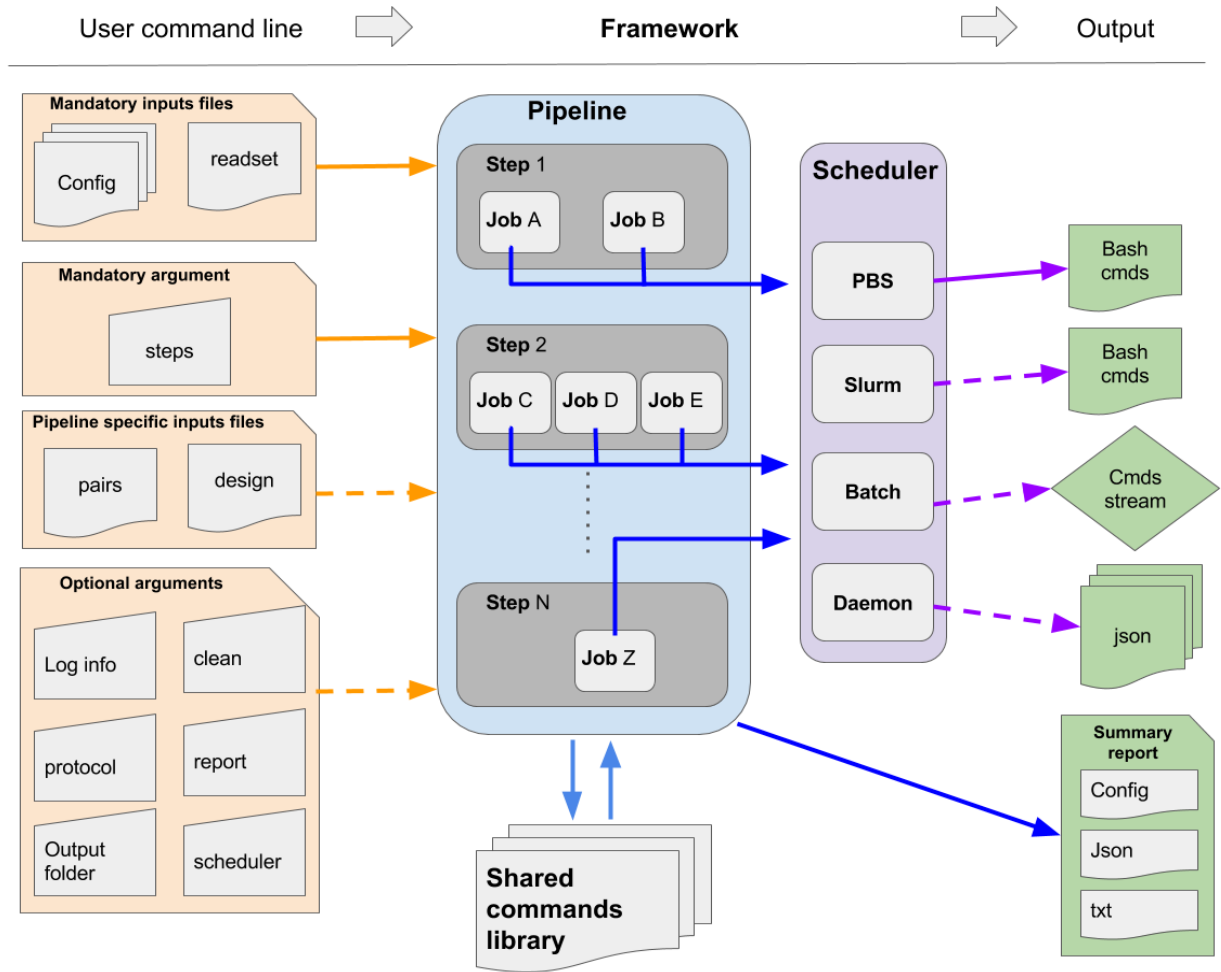
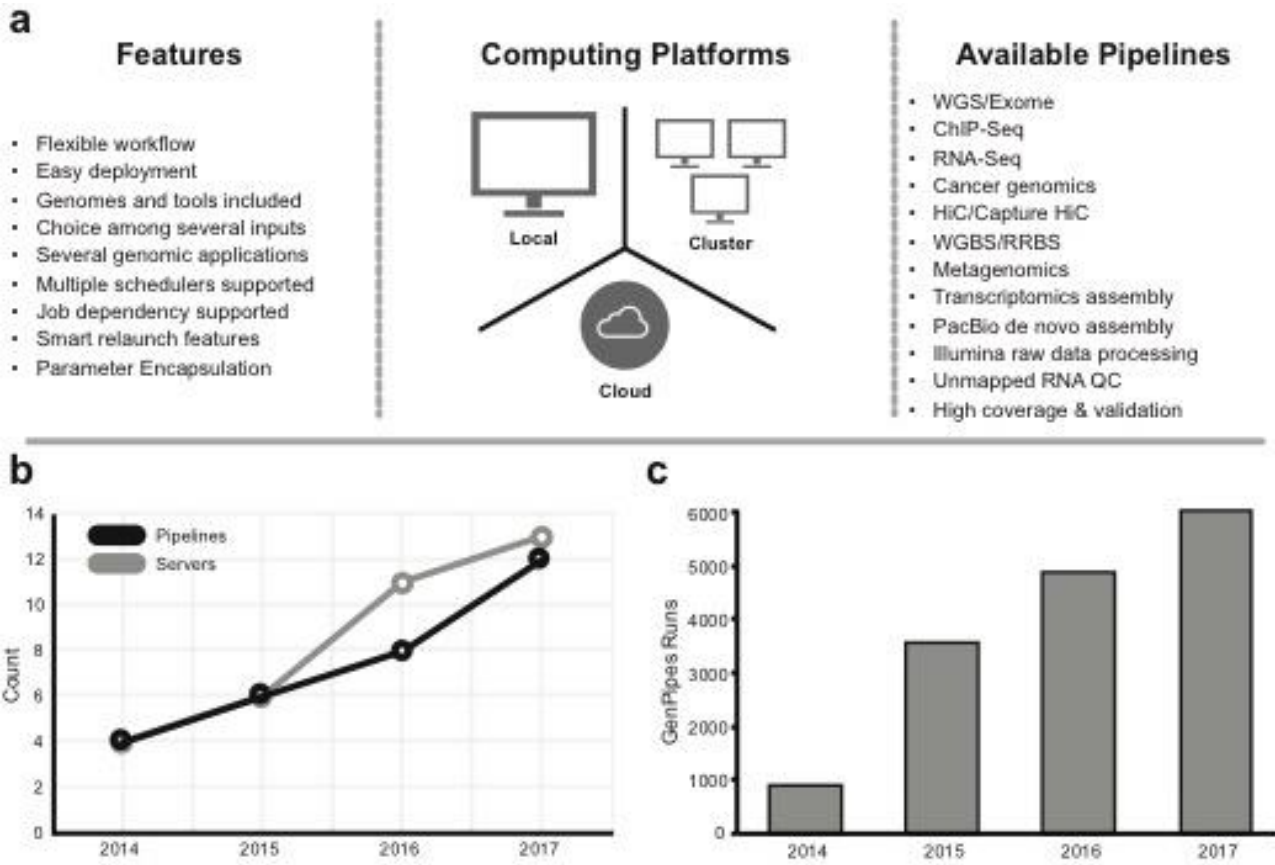


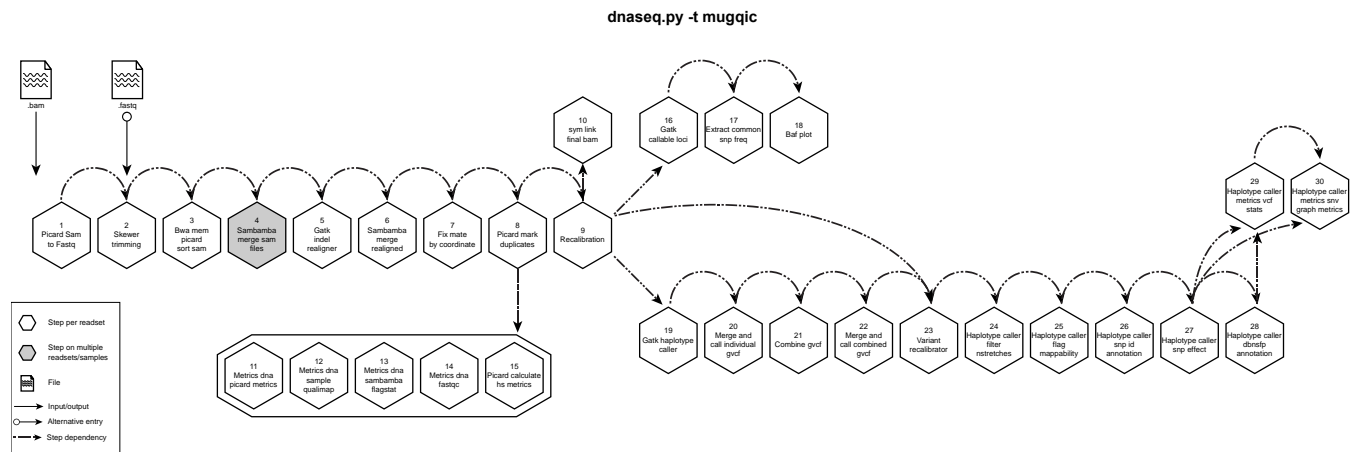
Diagram showing how the information flows from the user command line input through the 4 different objects (*Pipeline, Step, Job and Scheduler*) in order to generate system specific executable outputs.

Figure 2 - GenPipes properties



GenPipes' properties and growth. (a) Diagram showing GenPipes' features, compatible computing platforms and available pipelines. (b) GenPipes' available pipelines and maintained servers since the release of GenPipes in 2014. (c) Bar plot showing the number of GenPipes runs per year since its release.

Figure 3 – GenPipes DNaseSeq pipeline diagram



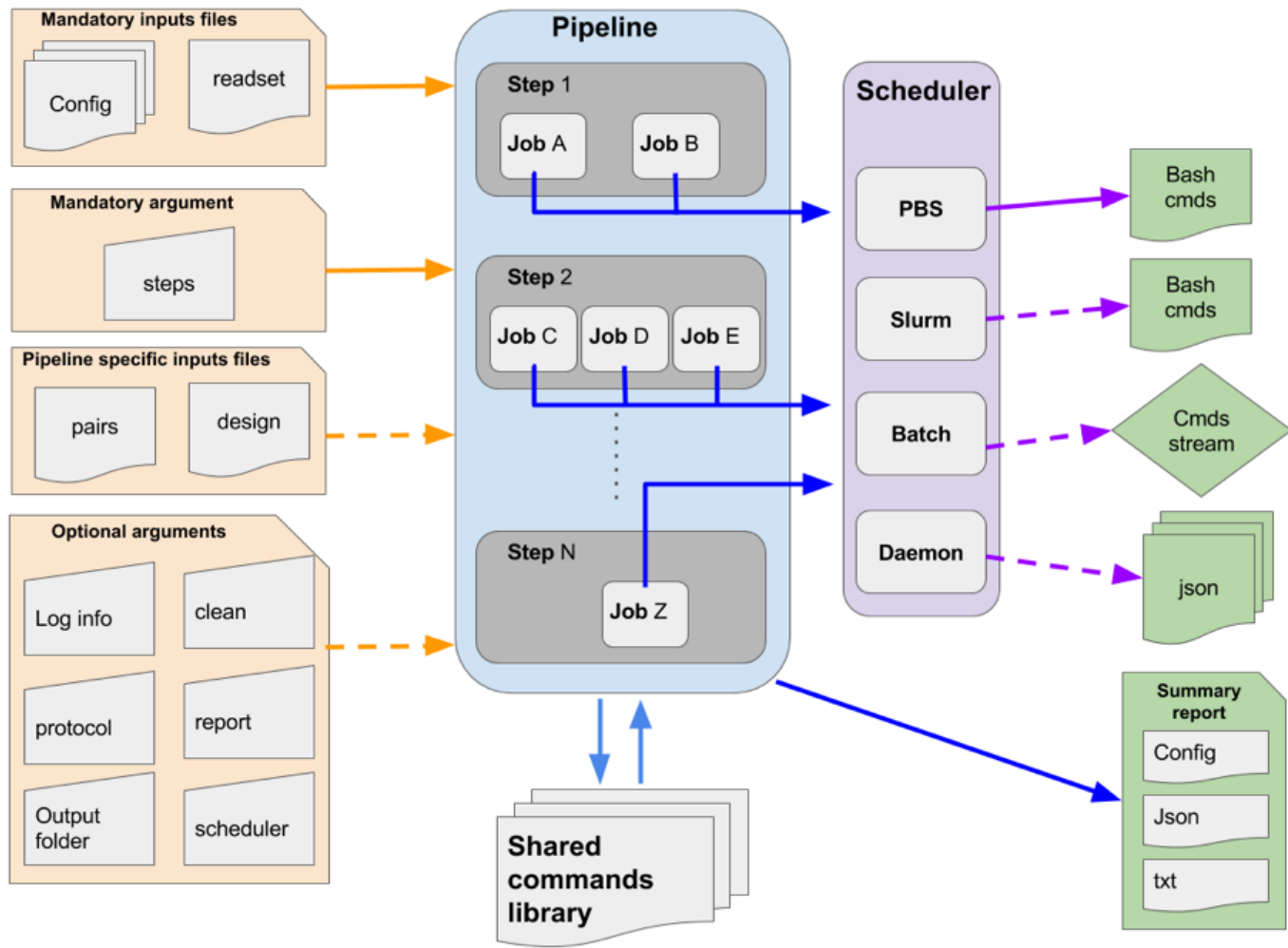
Schematic representation of GenPipes' dnaseq.py pipeline. Hexagons represent steps in the pipeline. White hexagons represent steps that process input from a single sample, while grey ones represent steps that process input from several samples. Arrows show step dependencies.

Table 1 - Comparison of available solutions for NGS analysis.

Solution	Features													Pipelines									
	Language	Software license	Published	Free	Open source	Cloud/Container	HPC	Workflow manager	Progress Monitoring	Package Manager	GUI	Reports	Config Validation	Germline	Somatic	RNA-Seq	RNA-Seq De novo	ChIP-seq	Metagenome	Methyl-Seq	Hi-C	PacBio assembly	
Genomics	Python	GNU LGPL	Pending	✓	✓	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
Genome Modeling System	Perl	GNU LGPLv3	[57]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
Galaxy	Python	Academic Free L3.0	[4]	✓	✓	✓	✓	✓	✓	✗	✗	✗	✗	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
bcbb-nextgen	Python	MIT License	No	✓	✓	✓	✓	✗	✗	✗	✗	✓	N/A	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
Omics Pipe	Python	MIT License	[72]	✓	✓	✓	✓	✗	✗	✗	✗	✓	✗	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
GenePattern	Java	Custom	[73]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✗	✓	N/A	✓	✗	✗	✗	✗	✗	✗	✗
Illumina BaseSpace	bash	Custom	No	✗	✗	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
BIANA Genomic Analysis	Java/Python	Custom	No	✗	✗	✓	✓	✓	✓	✗	N/A	N/A	N/A	✓	✓	✓	N/A	✗	✗	✗	✗	✗	✗
SeqWare	Java	GNU GPLv3	[74]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✓	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗
DNA Nexus Platform	Python/bash	Custom	No	✓	Partial	✓	✗	✓	✓	✓	✓	✓	N/A	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
gkno2	Python	MIT License	No	✓	✓	✓	✓	✗	✗	✗	✗	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
NGSANE	bash	BSD3	[75]	✓	✓	✓	✓	✗	✗	✗	✗	✓	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗
GATK Queue	Scala	MIT License & Broad Institute	No	Partial	Partial	✗	N/A	✓	✓	✗	✗	N/A	N/A	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
CGAS Pipeline	Java	N/A	No	✓	✗	N/A	✓	✓	✓	✗	✗	✓	✗	N/A	✓	✗	✗	✗	✗	✗	✗	✗	✗
MIT STAR	Python	GNU GPLv3	[76]	✓	✓	✓	✗	✗	✗	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
CRONOS/WDL	Scala	BSD 3-Clause	No	Partial	✓	✓	✓	✓	✓	✗	✗	✓	N/A	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
BigDataScript	BDS	Apache License V2	[9]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
Kronos	Python	MIT license	[77]	✓	✓	✓	✓	✗	✗	✗	✗	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
NextFlow	Java	GNU GPLv3	[7]	✓	✓	✓	✓	✗	✗	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
Snakemake	Python	MIT License	[8]	✓	✓	✓	✓	✓	✓	✓	✗	✓	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗

Modified from Griffith & Griffith et al. [57]. Note that community-built pipelines are not considered in the Pipelines section of the table. It is also worth noting that the following table is meant to provide the reader with an overview of the features of several tools in the field but not necessarily an exhaustive list. For a full description of each tool's capabilities, please consult their official documentation.

Solution	Language	Software license	Published	Features										Pipelines									
				Free	Open source	Cloud/Container	SPC	Workflow manager	Progress Monitoring	Package Manager	Git	Reports	Config generation	Genome	Genote	Met. Ass.	Met. Ass. (p. work)	CRP-ass	Metagenome	Met. Ass.	W-C	Facilitator	
GenoPipes	Python	Open GPL	Pending	✓	✓	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
Genome Modeling System	Perl	Open GPL v3	[17]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
Galaxy	Python	Business, Free, GPL	[8]	✓	✓	✓	✓	✓	✓	✗	✗	✗	✗	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
Inchigo-mergen	Python	MIT License	Yes	✓	✓	✓	✓	✓	✗	✗	✗	✗	✗	✓	✓	✓	✓	✗	✗	✗	✗	✗	✗
Omics Pipe	Python	MIT License	[10]	✓	✓	✓	✓	✓	✗	✓	✗	✓	✗	✓	✓	✓	✗	✓	✗	✗	✗	✗	✗
Gene Pattern	Java	Custom	[19]	✓	✓	✓	✓	✓	✓	✗	✓	✓	✗	✓	✗	✓	✗	✗	✗	✗	✗	✗	✗
Biological WorkflowSpace	Java	Custom	Yes	✗	✗	✓	✓	✓	✓	✗	✗	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
BNA Genomic Analysis	Java/Python	Custom	Yes	✗	✗	✓	✓	✓	✓	✗	✗	✗	✗	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗
SeqWare	Java	Open GPL v3	[14]	✓	✓	✓	✓	✓	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
DNA Nexus Platform	Python/Java	Custom	Yes	✓	Partial	✓	✗	✓	✓	✗	✗	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗
gNovo	Python	MIT License	Yes	✓	✓	✓	✓	✓	✗	✗	✗	✓	✓	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗
NGSANE	Java	GPL	[11]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗
GATK's Queue	Scala	MIT License & Broad Institute	Yes	Partial	Partial	✗	✗	✓	✓	✗	✗	✗	✗	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
CGA's Firehose	Java	MIT	Yes	✓	✗	✗	✗	✓	✓	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
MIT STAR	Python	Open GPL v3	[16]	✓	✓	✓	✗	✓	✗	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗	✗
OpenWBS/WDL	Scala	Open & Custom	Yes	Partial	✓	✓	✓	✓	✓	✗	✗	✗	✓	✓	✓	✗	✗	✗	✗	✓	✗	✗	✗
BigDataLifezyg	Scala	Apache License 2.0	[9]	✓	✓	✓	✓	✓	✓	✗	✗	✓	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗
Kronos	Python	MIT License	[11]	✓	✓	✓	✓	✓	✗	✓	✗	✓	✗	✓	✗	✗	✗	✗	✗	✗	✗	✗	✗
Nextflow	Java	Open GPL v3	[7]	✓	✓	✓	✓	✓	✗	✓	✗	✓	✗	✗	✗	✓	✗	✗	✗	✗	✗	✗	✗
Snakemake	Python	MIT License	[6]	✓	✓	✓	✓	✓	✓	✓	✗	✓	✓	✗	✓	✓	✗	✗	✗	✗	✗	✗	✗

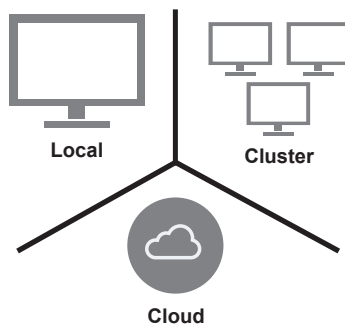


a

Features

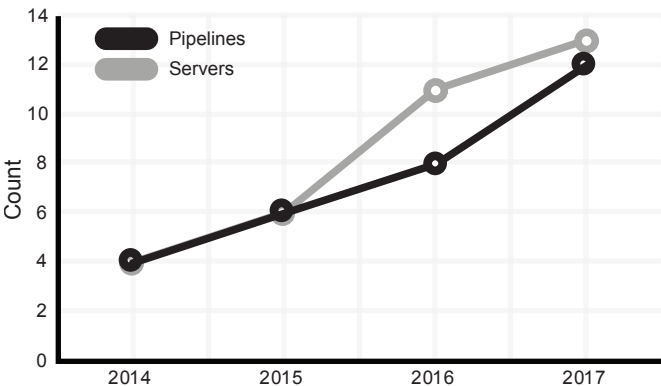
- Flexible workflow
- Easy deployment
- Genomes and tools included
- Choice among several inputs
- Several genomic applications
- Multiple schedulers supported
- Job dependency supported
- Smart relaunch features
- Parameter Encapsulation

Computing Platforms

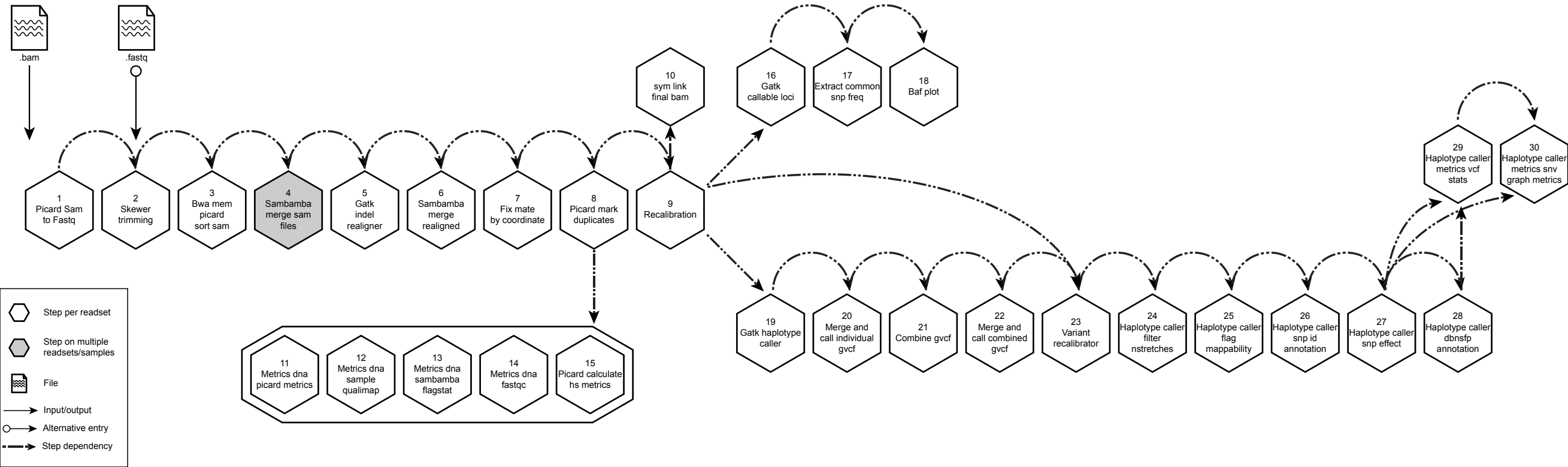


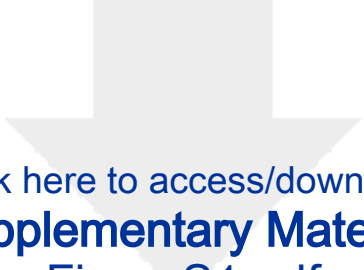
Available Pipelines

- WGS/Exome
- ChIP-Seq
- RNA-Seq
- Cancer genomics
- HiC/Capture HiC
- WGBS/RRBS
- Metagenomics
- Transcriptomics assembly
- PacBio de novo assembly
- Illumina raw data processing
- Unmapped RNA QC
- High coverage & validation

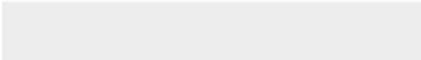
b**c**

dnaseq.py -t muggic





Click here to access/download
Supplementary Material
FigureS1.pdf



Dear Editor,

Thank you for the opportunity to submit a revised version of the manuscript GIGA-D-18-00198, which addresses the final points raised by the reviewers. Please find our point-by-point response below. New text that has been added to the revised manuscript is shown in red.

Response to the Reviewers:

Reviewer #2: I thank the authors for addressing several of my concerns. Two issues remain:

Major comments

* Regarding the feature table, the reasoning of the authors is acceptable (apart from my concern below), if the caption explicitly mentions that the table is "meant to provide the reader with an overview of the features of several tools in the field but not necessarily an exhaustive list".

We have added the suggested text to Table 1's caption as follows:

"... It is also worth noting that the following table is meant to provide the reader with an overview of the features of several tools in the field but not necessarily an exhaustive list. For a full description of each tool's capabilities, please consult their official documentation."

* Regarding the answer "Bioconda offers a collection of packages and not an integrated system and can be quite heavy in memory requirements. Hence, we think that "package-manager-integration" is not necessarily an indication of the strength of the WMS. It is a specific choice, one that offers ease of installation but has its pitfalls as well. GenPipes does not use package managers by design. GenPipes manages its own libraries making sure there is no conflicting libraries in the process. For users who do not want to install GenPipes manually, we offer a Docker container that has also been tested with Singularity. We have updated the GenPipes' bitbucket documentation to highlight the availability of the GenPipes' Docker container":

The way to provide the software stack for an analysis is very important for reproducibility and maintainability of a pipeline. Hence, this should definitely be a column of the feature table in any case. Certainly it is legitimate to decide against Conda or other package managers, but in above argument I do not find any reason to hide this aspect from the user in the feature table. Why not adding a column about package manager support and let the reader decide whether they need/want this feature or not?

We have added a column, "Package Manager", to Table1, as suggested by the reviewer.