

S1 Table. Strains and plasmids.

Strain or plasmid	Genotype or description	Reference
<i>A. baumannii</i>		
ATCC 17978	cerebrospinal fluid isolate	(1)
EGA336	ATCC 17978 <i>gyrA</i> (S81L)	This study
GVA41	ATCC 17978 <i>parC</i> (S84L)	This study
GVA37	ATCC 17978 <i>gyrA</i> (S81L) <i>parC</i> (S84L)	This study
EGA465	ATCC 17978 Δ <i>adeIJK</i>	(2)
GVA68	ATCC 17978 Δ <i>recN</i>	This study
GVA84	ATCC 17978 Δ <i>ctpA</i>	This study
EGA627	ATCC 17978 <i>pbp1a</i> (N178TfsX27)	This study
GVA61	ATCC 17978 Δ <i>gidA</i>	This study
YDA124	ATCC 17978 Δ P1 Δ P3	This study
<i>E. coli</i>		
DH5 α	<i>supE44</i> Δ <i>lacU169</i> (ϕ 80 <i>lacZ</i> Δ M15) <i>hsdR17 recA1 endA1 gyrA96 thi-1 relA1</i>	(3)
DH5 λ pir	DH5 α (λ pir) <i>tet::Mu recA</i>	(4)
TO60	DH5 α λ pir [F' <i>proAB lacI^qZ</i> Δ M15 Tn10 (Tc ^R)]	(5)
plasmids		
pUC18	<i>oriColE1</i> MCS Cb ^R	(6)
pSR47s	<i>oriTRP4 oriR6K</i> Km ^R	(7)
pDL1073	Tn10 (Km ^R) delivery plasmid, <i>ori pSC101</i> Cb ^R	This study
pWH1266	shuttle plasmid, Cb ^R Tet ^R	(8)
pEGE244	pWH1266 with <i>rrnB</i> & T7 tandem terminators replacing EcoRI-PstI fragment, Tet ^R	(2)
pGVE41	pWH1266 with <i>ctpA</i> replacing EcoRI-PstI fragment, Tet ^R	This study
pCC1	<i>recA</i> promoter-UTR(C::G)-mKate2 transcriptional fusion in pNLAC1 vector, Tet ^R	(9)
pCC7	<i>trpB</i> promoter-UTR(C::G)-mKate2 transcriptional fusion in pNLAC1 vector, Tet ^R	(9)

References

1. Bouvet P, Grimont P. 1986. Taxonomy of the Genus *Acinetobacter* with the Recognition of *Acinetobacter baumannii* sp. nov. *Acinetobacter haemolyticus* sp. nov. *Acinetobacter johnsonii* sp. nov. and *Acinetobacter junii* sp. nov. and Emended Descriptions of *Acinetobacter calcoaceticus* and *Acinetobacter lwoffii*. International Journal of Systematic Bacteriology 36:228-240.
2. Geisinger E, Mortman NJ, Vargas-Cuebas G, Tai AK, Isberg RR. 2018. A global regulatory system links virulence and antibiotic resistance to envelope homeostasis in *Acinetobacter baumannii*. PLoS Pathog 14:e1007030.
3. Hanahan D, Jessee J, Bloom FR. 1991. Plasmid transformation of *Escherichia coli* and other bacteria. Methods Enzymol 204:63-113.
4. Kolter R, Inuzuka M, Helinski DR. 1978. Trans-complementation-dependent replication of a low molecular weight origin fragment from plasmid R6K. Cell 15:1199-208.

5. O'Connor TJ, Adepoju Y, Boyd D, Isberg RR. 2011. Minimization of the *Legionella pneumophila* genome reveals chromosomal regions involved in host range expansion. Proc Natl Acad Sci U S A 108:14733-40.
6. Yanisch-Perron C, Vieira J, Messing J. 1985. Improved M13 phage cloning vectors and host strains: nucleotide sequences of the M13mp18 and pUC19 vectors. Gene 33:103-19.
7. Andrews HL, Vogel JP, Isberg RR. 1998. Identification of linked *Legionella pneumophila* genes essential for intracellular growth and evasion of the endocytic pathway. Infect Immun 66:950-8.
8. Hunger M, Schmucker R, Kishan V, Hillen W. 1990. Analysis and nucleotide sequence of an origin of DNA replication in *Acinetobacter calcoaceticus* and its use for *Escherichia coli* shuttle plasmids. Gene 87:45-51.
9. Ching C, Gozzi K, Heinemann B, Chai Y, Godoy VG. 2017. RNA-Mediated cis Regulation in *Acinetobacter baumannii* Modulates Stress-Induced Phenotypic Variation. J Bacteriol 199.