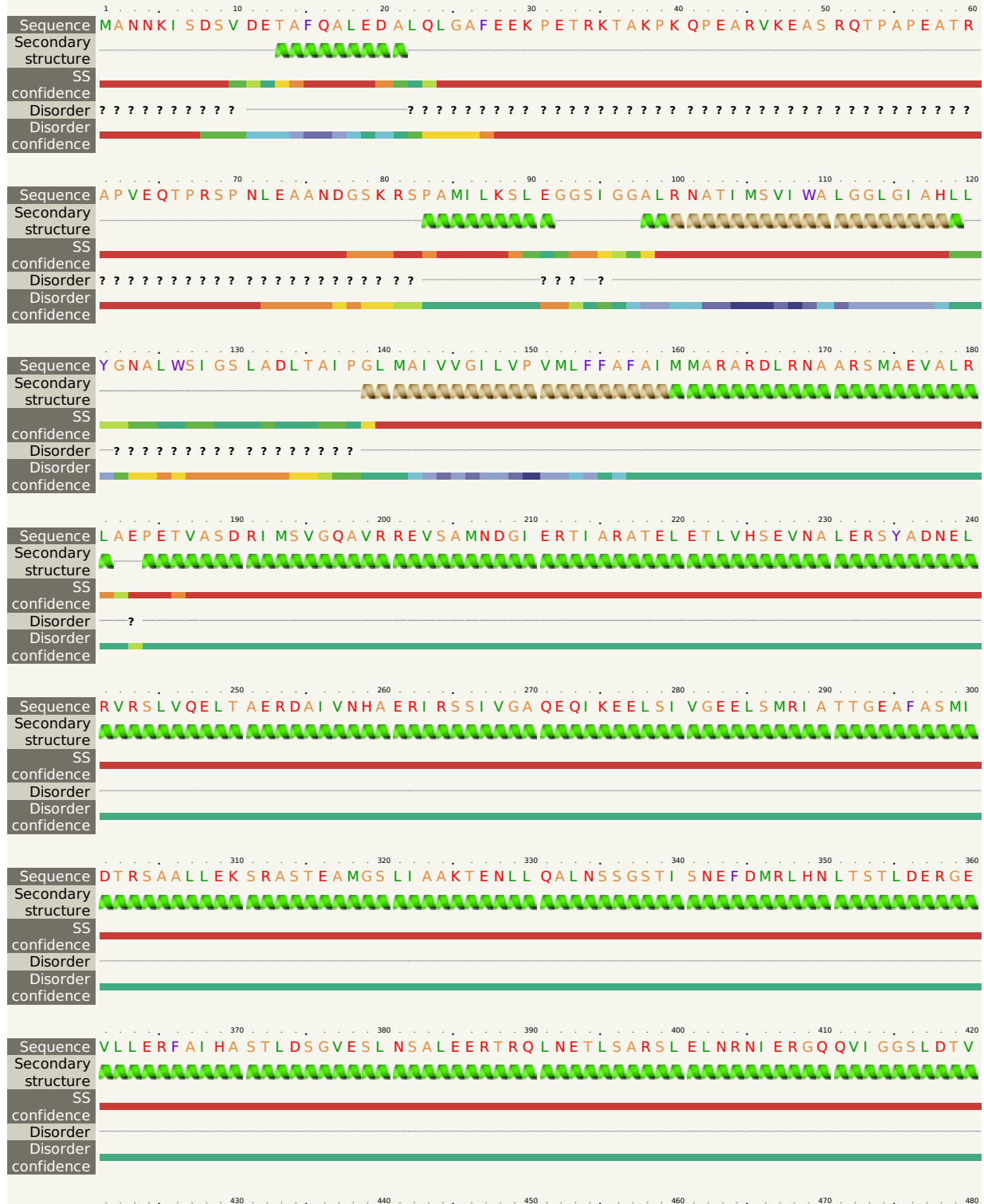
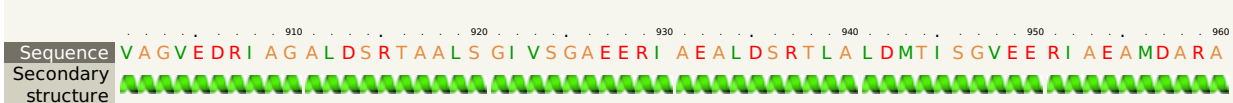
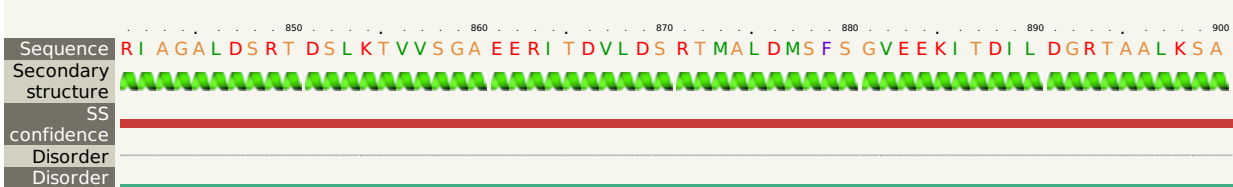
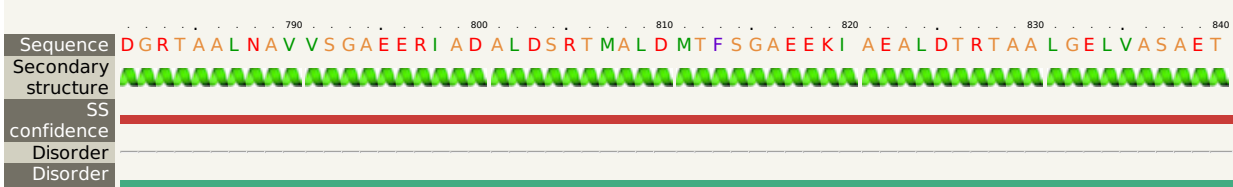
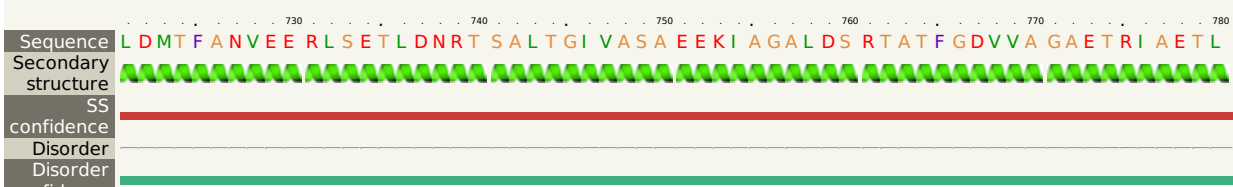
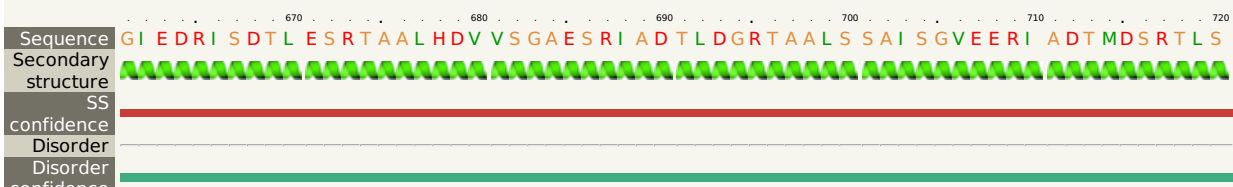
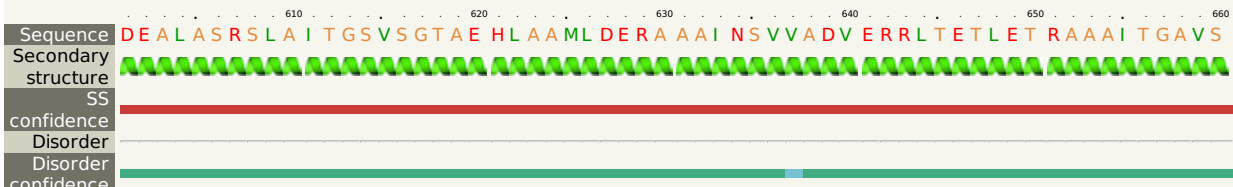
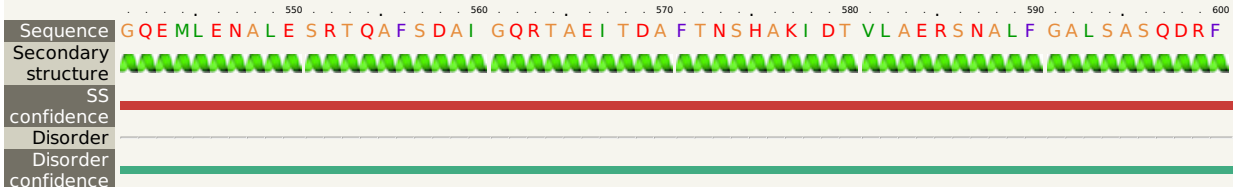
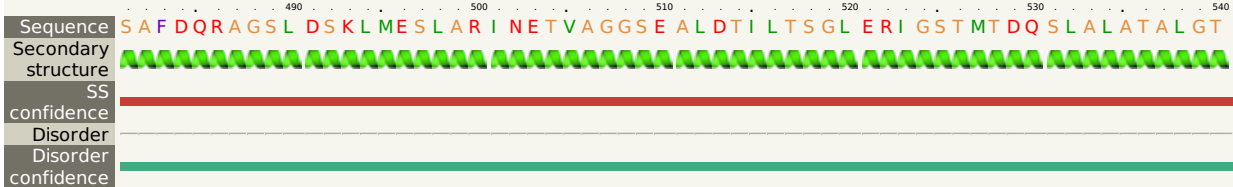


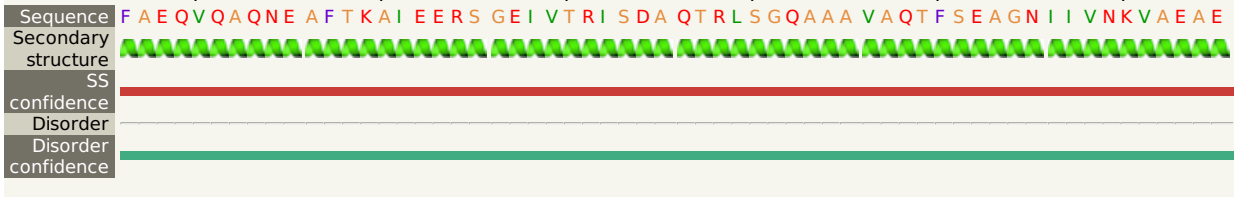
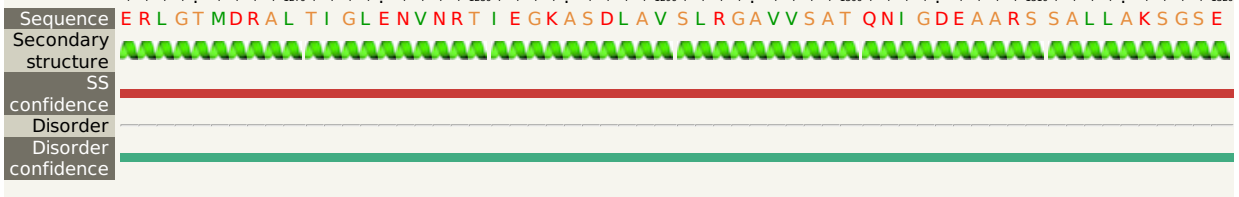
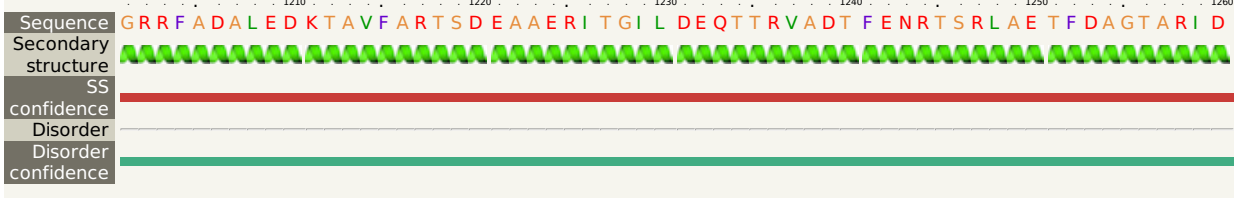
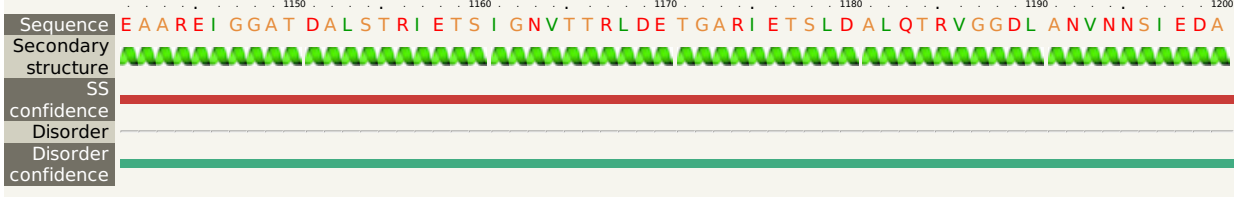
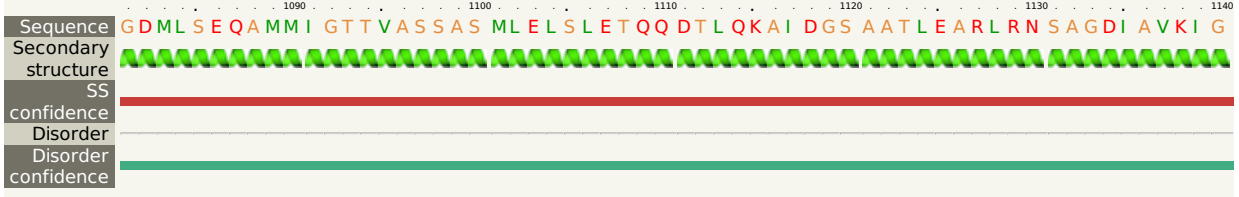
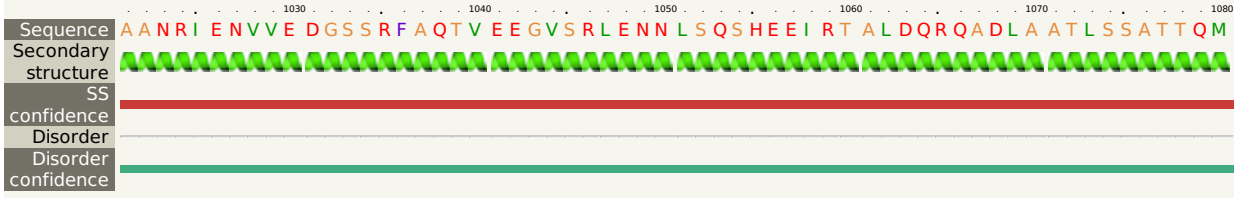
Supporting Information Appendix

Figure S1. Secondary structure predictions from PHYRE (1). A) *Agrobacterium tumefaciens* protein GPR. B) *Caulobacter crescentus* protein TipN.

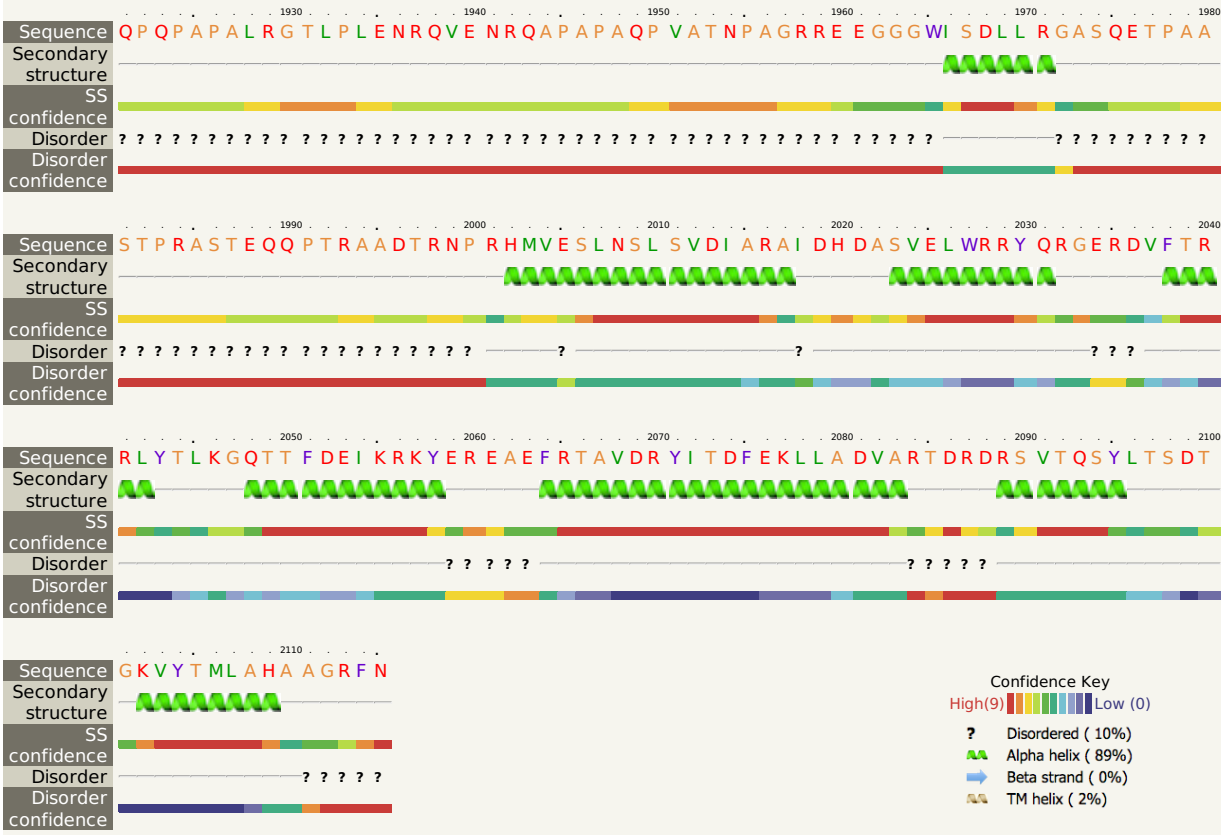
A.



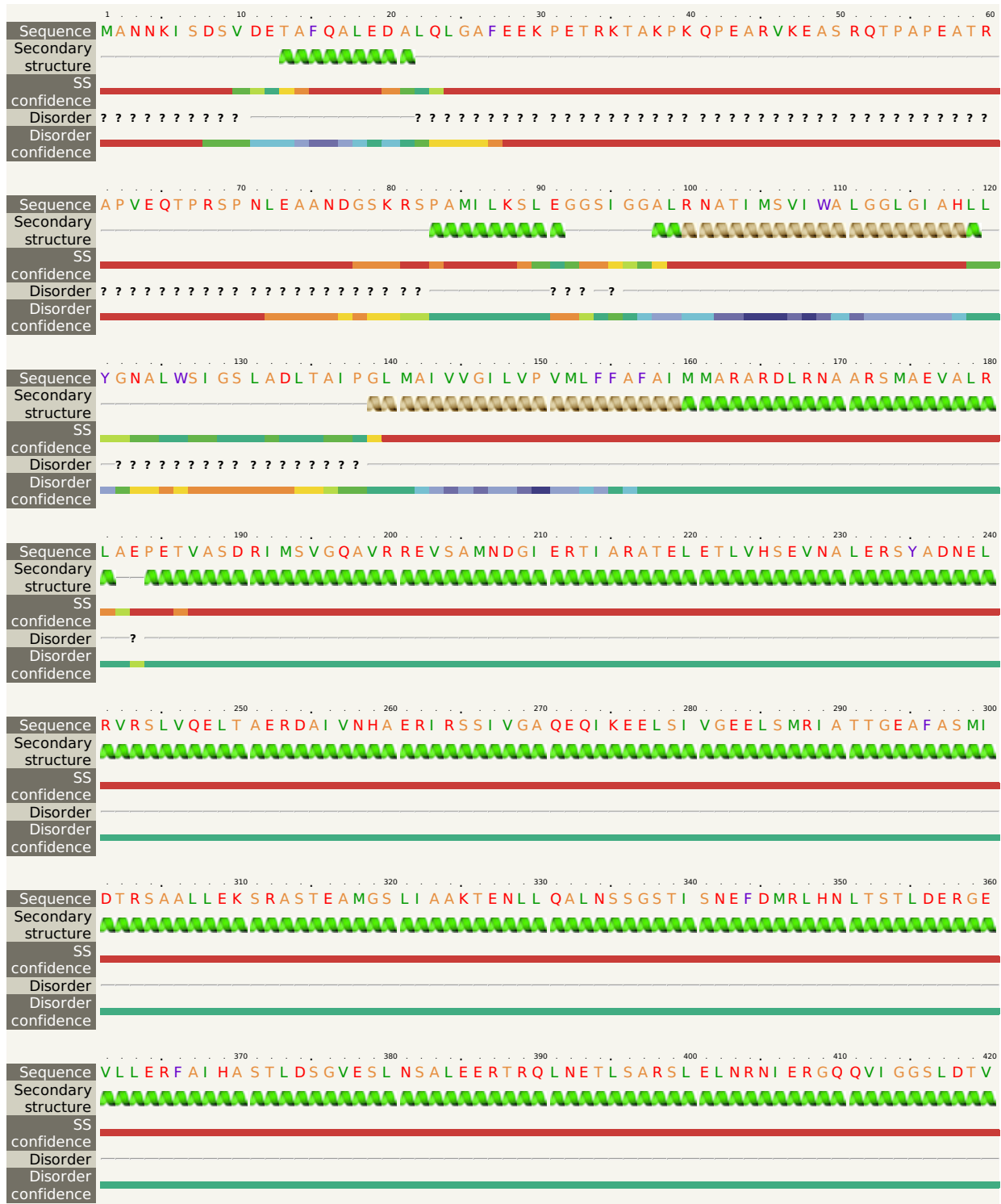


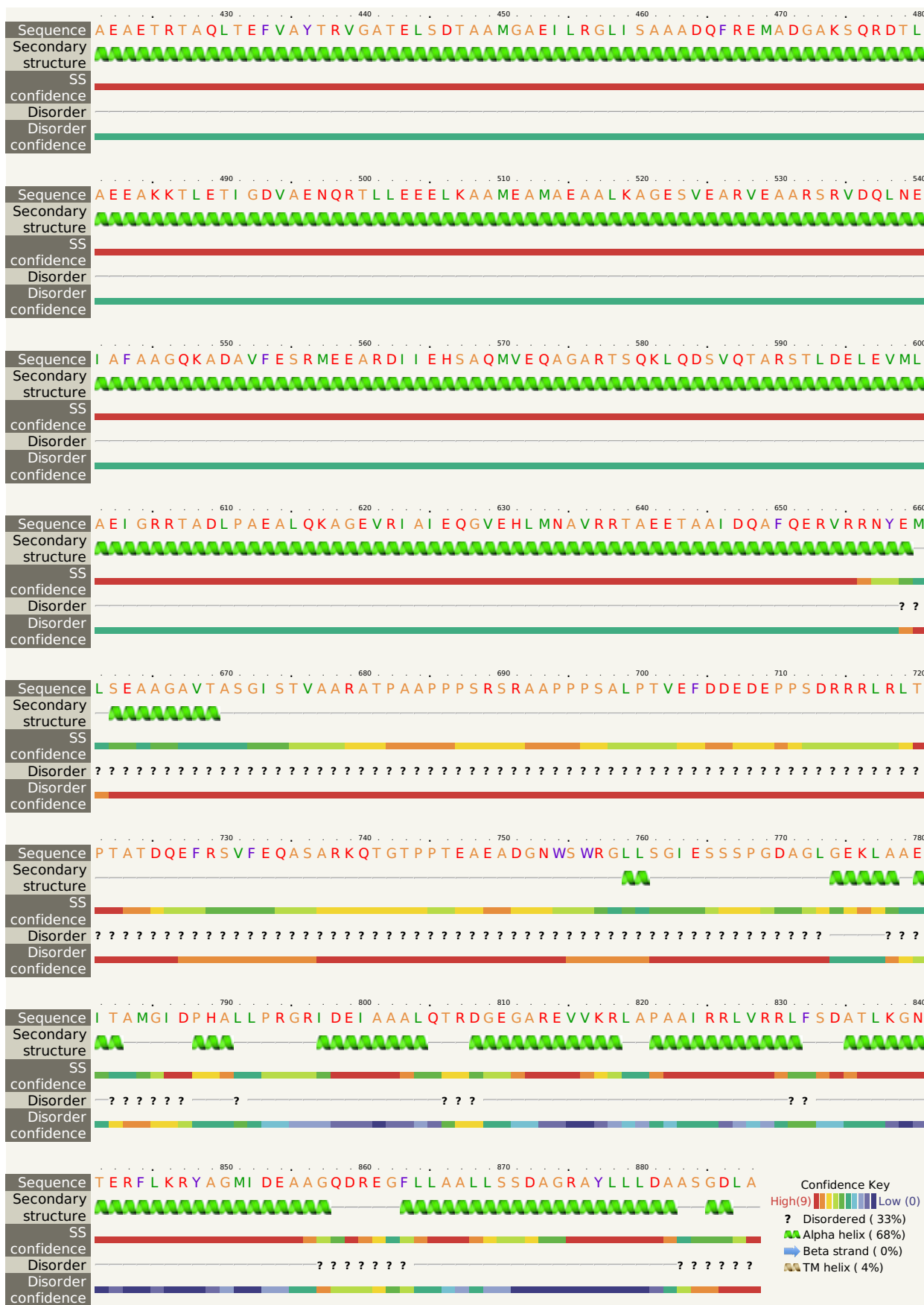






B.





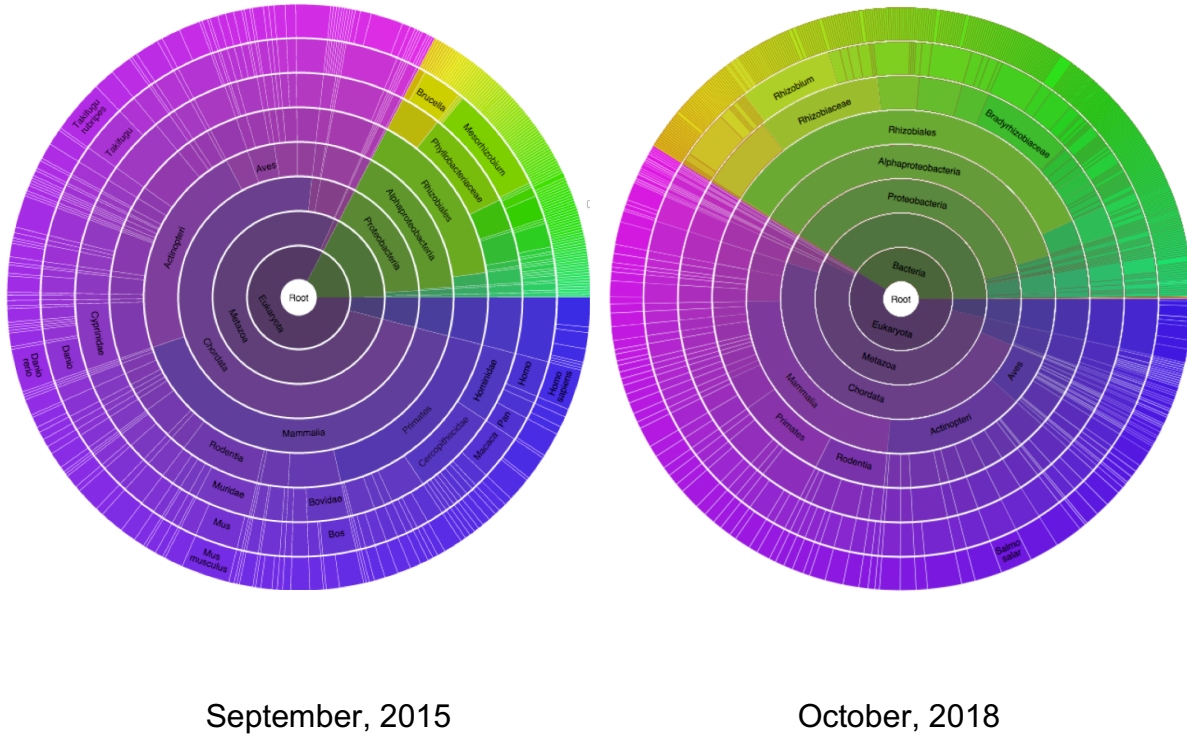


Figure S2. Change in species distribution of apolipoproteins (Pfam, PF01442, (2)) between Sept., 2015, and October, 2018.

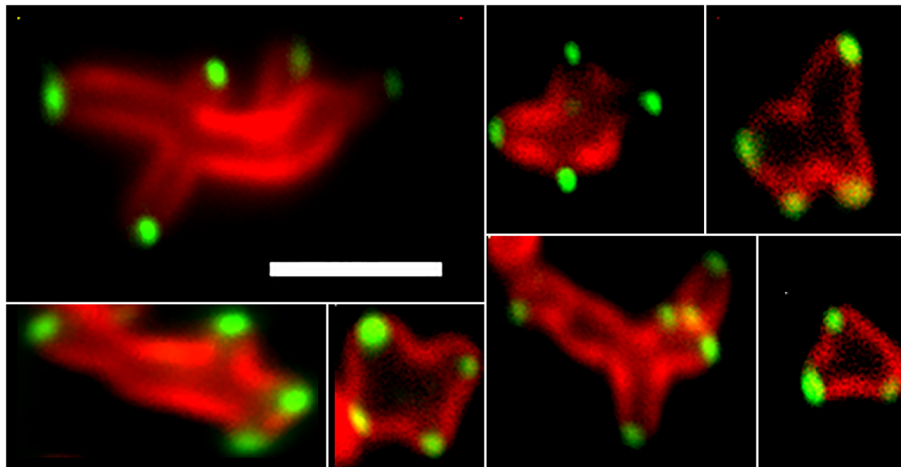


Figure S3. Expression of GPR-GFP *in trans* to WT GPR results in formation of ectopic poles. Cells are counterstained with FM4-64. Scale bar = 3 μ m.

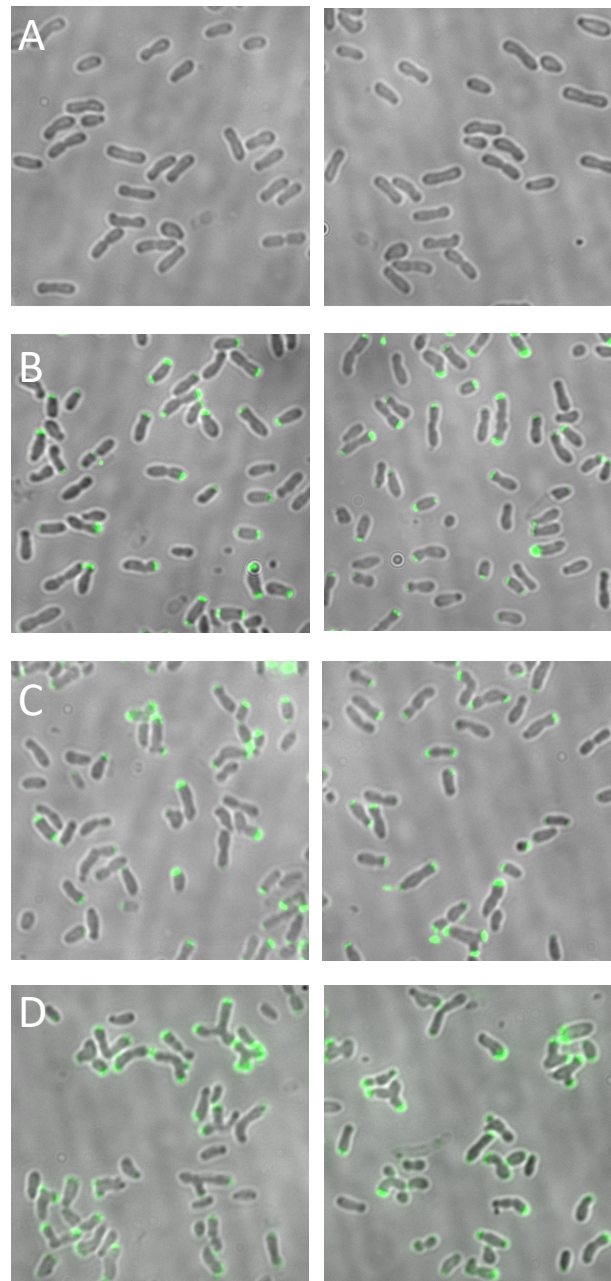


Figure S4. Induction of GFP-GPR in WT *Agrobacterium tumefaciens* with increasing concentrations of IPTG. Ectopic poles are formed at higher concentrations of IPTG. Cells were grown for 24 hours prior to imaging in the IPTG concentration indicated. A) 0mM IPTG, B) 0.25mM IPTG, C) 0.5mM IPTG, D) 1.0mM IPTG.

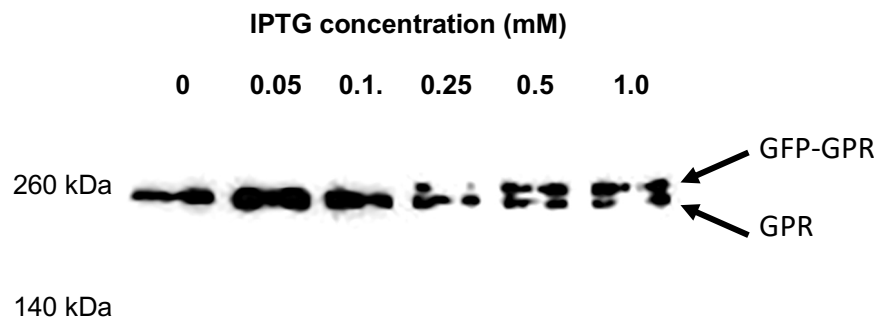


Figure S5. Western blot to detect WT GPR or GFP-GPR. Different concentrations of IPTG were used to induce GPR-GFP expression, cell extracts were separated by SDS-gel electrophoresis, and GPR was detected following Western blotting using antibodies to GPR.

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Figure S6. *gpr* deletion strategy. Atu1349 partial protein coding sequence in blue. Atu1348 (*GPR*) coding sequence in red. Intergenic region in black **bold**. Deleted sequence (underlined) comprises 238 bp upstream of the *gpr*-ATG and the first 2140 bp of the coding sequence for Atu1348 (total *gpr* protein coding sequence is 6345 bp).

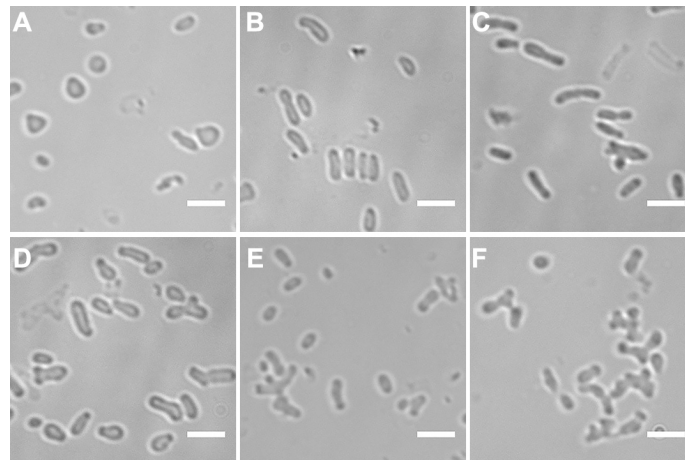


Figure S7. Δgpr complementation. Unipolar growth and cell division were restored in Δgpr following introduction of a plasmid carrying WT *gpr* under the control of the *lac* promoter ($P_{lac}::gpr$ (pSRKGm)). Cells were grown for 24 hours prior to imaging in the IPTG concentration indicated. A) 0 μ M IPTG, B) 25 μ M IPTG, C) 50 μ M IPTG, D) 100 μ M IPTG, E) 250 μ M IPTG, and F) 500 μ M IPTG. Scale bar = 3 μ m.

Supplementary Table 1. Apolipoprotein domains (Family PF01442) in GPR identified by www.pfam.xfam.org (2). ^aamino acid number

Position in GPR^a	Domain Length (amino acids)	e-value
290-450	161	9.7×10^{-1}
323-523	201	3.6×10^{-1}
416-612	197	1.5×10^{-2}
495-718	224	7.3×10^{-3}
625-824	201	7.2×10^{-5}
821-1008	188	5.5×10^{-2}
937-1126	190	7×10^{-6}
1115-1338	224	1.9×10^{-4}
1291-1439	149	13
1335-1542	208	6.5×10^{-4}
1583-1783	201	5.3×10^{-4}
1784-1878	95	290

Table S2. Strains and plasmids used in this study.

Strains	Relevant genotype and features	Source
XL1 Blue E. coli	cloning strain, endA1 gyrA96(nal ^R) thi-1 recA1 relA1 lac glnV44 F'[::Tn10 proAB+ lacIq Δ(lacZ)M15] hsdR17(rKmK+), Tet ^R	Lab stock
C58	wild-type <i>A. tumefaciens</i> strain C58	Lab stock
A185	C58 carrying pJZ251	This work
A187	C58 carrying pJZ253	This work
A202	C58 carrying pJZ253 and pJZ269	This work
A212	C58 with the native <i>gpr</i> ribosome binding (RBS) site substituted with a riboswitch, referred to as RS:: <i>gpr</i> in text	This work
A248	C58 with a deletion of 2378bp including 238bp upstream of the <i>gpr</i> ATG and the first 2140bp of the <i>gpr</i> coding sequence, referred to as Δ <i>gpr</i> in text	This work
A239	A212 carrying pJZ253	This work
A253	A248 carrying pJZ280	This work
A256	A212 carrying pRG001	This work
RESJ001	C58 carrying pJZ253 and pTC077	This work
Plasmids	Relevant genes and construction information	Source
pSRKGm	Broad host-range, <i>lacI</i> , gentamycin ^R	(3)
pSRKKm	Broad host-range, <i>lacI</i> , kanamycin ^R	(3)
pBluescript II SK	Phagemid, carbenicillin ^R	Stratagene
pJZ156	pBSKII+ with <i>sacB</i> , carbenicillin ^R	(4)
pJZ210	pSRKGm with P _{lac} :: <i>gfp</i>	This work
pJZ251	pSRKGm with P _{lac} :: <i>gpr</i> _{At} - <i>gfp</i>	This work

pJZ253	pSRKGm with $P_{lac}::gfp-gpr$	This work
pJZ269	pSRKKm with $P_{lac}::popZ_{At}-rfp$	This work
pJZ274	pJZ156 carrying 2kb sequence homologous to C58 genomic DNA in which the native <i>gpr</i> RBS is replaced with a riboswitch (Fig. S8)	This work
pJZ280	pSRKGm with $P_{lac}::gpr$	This work
pJZ298	pJZ156 carrying 1kb homologous to C58 genomic DNA on either side of the sequence to be deleted (Fig. S6)	This work
pRG001	pSRKGm with $P_{lac}::popZ_{At}-gfp$	(5)
pTC077	pDW029 with $P_{lac}::ftsZ-rfp$, streptomycin/spectinomycin ^R	(6)

SUPPLEMENTAL REFERENCES

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3. Khan SR, Gaines J, Roop RM, Farrand SK (2008) Broad-host-range expression vectors with tightly regulated promoters and their use to examine the influence of TraR and TraM expression on Ti plasmid quorum sensing. *Appl Env Microbiol* 74(16):5053–5062.
4. Anderson-Furgeson JC, Zupan JR, Grangeon R, Zambryski PC (2016) Loss of PodJ in *Agrobacterium tumefaciens* leads to ectopic polar growth, branching, and reduced cell division. *J Bacteriol* 198(13):1883–1891.
5. Grangeon R, Zupan JR, Anderson-Furgeson J, Zambryski PC (2015) PopZ identifies the new pole, and PodJ identifies the old pole during polar growth in *Agrobacterium tumefaciens*. *Proc Natl Acad Sci USA* 112(37):11666–11671.

6. Cameron TA, Anderson-Furgeson J, Zupan JR, Zik JJ, Zambryski PC (2014) Peptidoglycan synthesis machinery in *Agrobacterium tumefaciens* during unipolar growth and cell division. *MBio* 5(3):e01219–14.

Supplemental Movies

Movie S1, S2, S3, S4. *Agrobacterium* cells expressing GFP-GPR display a ring of foci just below the growth pole. GFP-GPR localization in 4 cells shown by rotational animation of 3-dimensional renderings from images obtained with structured illumination superresolution microscopy.

Movie S5. *Agrobacterium* cells expressing GFP-GPR and PopZ-RFP display a ring of green foci just below the growth pole and a single red focus. Localization shown in rotational animation of 3-dimensional rendering from images obtained with structured illumination superresolution microscopy.

Movie S6. Time lapse of *Agrobacterium* cell expressing GFP-GPR shown in Fig. 4.

Movie S7. Time lapse of 2 *Agrobacterium* cells expressing GFP-GPR show variability in persistence of GPR at old pole.

Movie S8. Time lapse of Δgpr cell in Fig. 5.

Movie S9. Δgpr cell division produces quartet of cells.