Supporting Information Part A

Preparation of Chiral Allenes through Pd-Catalyzed Intermolecular Hydroamination of Conjugated Enynes: Enantioselective Synthesis Enabled by Catalyst Design

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I. General Information

General Procedures. All reactions were carried out in oven- (120 °C) or flame-dried glassware under an inert atmosphere of dry N₂ unless otherwise noted. Oven-dried (60 °C or 120 °C) stainless steel cannulas and/or glass syringes (or N₂-flushed plastic syringes) were used for reagent transfer. Organic solutions were concentrated under reduced pressure using a rotary evaporator (Büchi). Flash column chromatography was performed using SiliCycle SiliaFlash[®] P60 Silica Gel.

Reagents.

Reagents purchased and used as received acetyl chloride (Sigma-Aldrich) allyl palladium chloride dimer (Strem) azepane (Sigma-Aldrich) benzofuran-2-carbaldehyde (Beantown) benzophenone imine (Chem-Impex) bis[rhodium($\alpha, \alpha, \alpha', \alpha'$ -tetramethyl-1,3-benzenedipropionic acid)] (Rh₂(esp)₂, Sigma-Aldrich) 2-bromobenzovl chloride (Oakwood) 1-bromo-4-ethynylbenzene (Matrix) (*Z*)-1-bromo-1-propene (Sigma-Aldrich) 2-bromopropene (Beantown) 2-bromo-3,3,3-trifluoro-1-propene (Alfa Aesar) *tert*-butyl(ethynyl)dimethylsilane (TCI) *n*-butyllithium (2.0 M in cyclohexane or 2.5 M in hexanes, Sigma-Aldrich) *t*-butyllithium (1.7 M in pentane, Sigma-Aldrich) citric acid (Fisher) copper (I) Iodide (Strem) cyclohexylacetylene (Alfa Aesar) cyclohexylamine (Sigma-Aldrich) diisoproyl azodicarboxylate (Alfa Aesar) (S)-N, α -dimethylbenzylamine (Acros) 1-dodecyne (Beantown) 4-ethynylaniline (Chem-Impex) 4-ethynylanisole (TCI) 2-ethynyltoluene (Sigma-Aldrich) 3-ethynyltoluene (Alfa Aesar) 1-ethynyl-4-(trifluoromethyl)benzene (TCI) ferrocene carboxylic acid (Chem-Impex) indoline (Acros) L-tert-leucinol (TCI) methanesulfonyl chloride (Alfa Aesar) *N*-methylbenzylamine (Sigma-Aldrich) methyl 4-ethynylbenzoate (Sigma-Aldrich) 1-methylpiperazine (TCI) (*R*)-2-methylpyrrolidine (Alfa Aesar) morpholine (Sigma-Aldrich) oxalyl chloride (Alfa Aesar) 4-pentyn-1-ol (Alfa Aesar) phenylacetylene (Alfa Aesar) 4-phenyl-1-butyne (Acros) piperidine (Sigma-Aldrich) 4-piperidone ethylene ketal (Alfa Aesar) *N*-propargylphthalimide (Alfa Aesar) *n*-propylamine (Alfa Aesar) pyridine (Alfa Aesar)

pyrrolidine (Sigma-Aldrich) silver tetrafluoroborate (Strem) sodium tetrafluoroborate (Acros) 1,2,3,4-tetrahydroquinoline (Alfa Aesar) Tetrabutylammonium fluoride (1.0 M in THF, Sigma-Aldrich) tetrakis[(R)-(-)-[(1R)-1-(4-bromophenyl)-2,2-diphenylcyclopropanecarboxylato]dirhodium(II) (Rh₂(*R*-BTPCP)₄, Strem) tetrakis[(R)-(+)-N-(p-dodecylphenylsulfonyl)prolinato]dirhodium(II) (Rh₂(*R*-DOSP)₄, Strem) tetrakis[(S)-(+)-(1-adamantyl)-(N-phthalimido)acetato]dirhodium(II) (Rh₂(S-PTAD)₄, Strem) tetrakis[5-t-butyl-phthaloyl-N-(S)-tert-leucinato]dirhodium (Rh₂(S-tert-PTTL)₄, Strem) tetrakis(triphenylphosphine)palladium(0) (Strem) *p*-toluenesulfonyl chloride (Alfa Aesar) trimethylsilylacetylene (Chem-Impex) vinvl bromide (1.0 M in THF, Sigma-Aldrich) Ligands purchased and used as received (R)-2,2'-bis[bis(3,5-trifluoromethylphenyl)phosphino]-4,4',6,6'-tetramethoxybiphenyl (R-BTFM-Garphos, Alfa Aesar) (S,S)-2,3-bis(*tert*-butylmethylphosphino)quinoxaline (S,S-QuinoxP^{*}, Strem) (-)-1,2-bis((2S,5S)-2,5-dimethylphospholano)ethane (S,S-Me-BPE, Alfa Aesar) (±)-2,2'-bis(diphenylphosphino)-1,1'-binaphthalene (*rac*-BINAP, Chem-Impex) (*R*)-2,2'-bis(diphenylphosphino)-1,1'-binaphthalene (*R*-BINAP, Alfa Aesar) 1,4-bis(diphenylphosphino)butane (dppb, Alfa Aesar) 4,5-bis(diphenylphosphino)-9,9-dimethylxanthene (xantphos, Strem) (+)-1,2-bis((2R,5R)-2,5-diphenylphospholano)ethane (S,S-Ph-BPE, Strem) (2S,4S)-2,4-bis(diphenylphosphino)pentane (S,S-BDPP, Strem) 1,3-bis(diphenylphosphino)propane (dppp, Sigma-Aldrich) (R_P) -1-[(R)-|A-(Dimethylamino)-2-(diphenylphosphino)benzyl]-2-diphenylphosphinoferrocene (R_P) -1-[(R)-|A-(Dimethylamino)-2-(diphenylphosphino)benzyl]-2-diphenylphosphinoferrocene ((R,R)-taniaphos, Strem) (R)-1-[(S_P) -2-(diphenylphosphino)ferrocenyl]ethyldicyclohexylphosphine (R,S-josiphos, Strem) ethylenebis(diphenylphosphine) (dppe, Strem) 1,1'-ferrocenediyl-bis(diphenylphosphine) (dppf, Sigma-Aldrich) (oxydi-2,1-phenylene)bis(diphenylphosphine) (DPEphos, Strem) triphenylphosphine (Alfa Aesar) Reagents distilled from CaH₂

diethylamine (Beantown)

1,2,3,4-tetrahydroisoquinoline (THIQ, Sigma-Aldrich)

tetramethylethylenediamine (TMEDA, Alfa Aesar)

triethylamine (Sigma-Aldrich)

The following alkynes were prepared by a previously described route through Sonogashira coupling of the corresponding aryl halides with trimethylsilylacetylene followed by deprotection.¹ 4-ethynylbenzaldehyde 3-ethynylbenzonitrile

2-ethynylnaphthalene
2-ethynylphenol
3-ethynylpyridine
2-ethynylbenzofuran was prepared by a Corey-Fuchs reaction from the corresponding aldehyde as previously described.²

Other reagents prepared by previously described methods 2-bromooct-1-ene³ (1-bromovinyl)cyclohexane³ chlorobis(perfluorophenyl)phosphane⁴ methyl 2-diazo-2-phenylacetate⁵ sodium tetrakis[3,5-bis(trifluoromethyl)phenyl]borate (NaBAr₄^F)⁶

Solvents. Solvents were sparged with dry N_2 and purified under a positive pressure of dry N_2 by an Innovative Technologies PureSolve solvent purification system: tetrahydrofuran (Sigma-Aldrich), dichloromethane (Sigma-Aldrich), toluene (Sigma-Aldrich), and diethyl ether (Sigma-Aldrich) were passed through two consecutive alumina columns. Hexanes (Fisher) and ethyl acetate (Fisher) were used for flash column chromatography and used as received. HPLC-grade hexanes (Sigma-Aldrich), isopropanol (Sigma-Aldrich), acetonitrile (Fischer), and methanol (Sigma-Aldrich) were used as received.

Instrumentation. ¹H NMR spectra were recorded on a Varian INOVA (400 MHz) or a Bruker (500 MHz) spectrometer. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance resulting from incomplete deuteration as the internal reference (CDCl₃: δ 7.24, CD₃OD: δ 3.29). Data are reported as follows: chemical shift, integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, br = broad, m = multiplet, app. = apparent), coupling constant(s) (Hz). ¹³C NMR spectra were recorded on a Varian/Agilent VNMRS (500 MHz) or a Bruker (500 MHz) spectrometer with complete proton decoupling. Chemical shifts are reported in ppm from tetramethylsilane with the solvent resonance as the internal reference (CDCl₃: δ 77.07, CD₃OD: δ 47.6, DMSO-d₆: δ 40.0). ¹⁹F NMR spectra were recorded on a Varian INOVA (400 MHz) spectrometer. ³¹P NMR spectra were recorded on a Varian INOVA (400 MHz) spectrometer. Enantiomer ratios (er) were determined by HPLC (PhenomenexTM Lux[®] Cellulose-3, Amylose-1, or Chiralpak 1A-3) in comparison with authentic racemic materials on a Shimadzu Prominence Modular HPLC. High-resolution mass spectrometry was performed on an Agilent (1200 Series) LCMS-TOF-DART at the Duke University Mass Spectrometry Facility. Elemental Analysis was performed at Atlantic Microlab. Specific rotation values were recorded on a Rudolph Autopol IV Polarimeter. Melting points were measured on an Electrothermal MelTemp[®] capillary melting point apparatus and are uncorrected. Infrared (IR) spectra were collected on a Nicolet 6700 FT-IR spectrometer, v_{max} in cm⁻¹. Bands are characterized as broad (br), strong (s), medium (m), or weak (w).

II. Supplemental Screening Data

		$\begin{array}{c} 2.5 \text{ mol}\% [Pd(\eta^3-\text{allyl})Cl]_2 \\ 5 \text{ mol}\% \text{ Ligand} \\ +/- 6 \text{ mol}\% \text{ additive} \\ +/- Et_3N \\ CH_2Cl_2, 22 ^\circ C, 3 \text{ h} \end{array} \qquad \begin{array}{c} Ph & & & & \\ Ph & & & & \\ \hline & & & & \\ \hline & & & & \\ \hline & & & &$				
_	entry	Ligand ac	ditive (6 mol%)	base (equiv)	yield 2a (%) ^b	
	1	none	none	none	0	
	2	DPEphos	none	none	29	
	3	DPEphos	NaBAr ₄ F	none	84	
	4	Xantphos	NaBAr ₄ F	none	83	
	5	rac-BINAP	NaBAr ₄ F	none	46	
	6	dppf	NaBAr ₄ F	none	63	
	7	dppe	NaBAr ₄ F	none	14	
	8	dppp	NaBAr ₄ F	none	44	
	9	dppb	NaBAr ₄ F	none	62	
	10	PPh ₃ (10 mol%) NaBAr ₄ F	none	44	
	11	DPEphos	NaBAr ₄ F	Et ₃ N (2.0)	84	
	12	DPEphos	NaBF ₄	none	86	
	13 ^c	DPEphos	AgBF ₄	none	85	
	14	Pd-1	none	none	88	

 Table S1. Optimization of Non-Enantioselective Reaction^a

^aReactions run with 0.20 mmol THIQ in 0.25 mL CH₂Cl₂. ^bIsolated yield of pure **2a**. ^cAgCl precipitate removed by filtration

$= HNR^{1}R^{2}$ $+$ Ph $1a$ $1.2 equiv$		5 mol% Pd-2 or Pd-3 +/- 2.0 equiv Et ₃ N solvent, 22 °C, time		Ph 2a +	$\mathbb{R}^{1}\mathbb{R}^{2}$ $\mathbb{R}^{1}\mathbb{R}^{2}$ $\mathbb{R}^{1}\mathbb{R}^{2}$	<i>t</i> -Bu Ar = 3 Pd-2	$Pd \div PAr_2$ $Pd \div Pd \div$	
entry	Pd	solvent	base (equiv)	time (h)	2a:9 ^b	yield 2a (%) ^c	er ^d	
1	Pd-2	CH ₂ Cl ₂	none	17	2:1	43	50:50	
2	Pd-2	CH ₂ Cl ₂	none	3	4:1	56	53.5:46.5	
3	Pd-2	CH_2CI_2	none	0.5	8:1	51	68:32	
4	Pd-2	Et ₂ O	none	3	-	<5%	-	
5	Pd-3	Et ₂ O	none	3	13:1	73	69.5:30.5	
6	Pd-2	CH_2CI_2	Et ₃ N (2.0)	3	19:1	71	63:37	

Table S2. Initial Investigations into Enantioselective Reaction with PHOX Ligands^a

^aReactions run with 0.20 mmol THIQ in 0.25 mL solvent. ^bDetermined by ¹H NMR analysis of unpurified reaction mixture. ^cIsolated yield of pure 2a. ^dDetermined by HPLC analysis of pure 2a in comparison with an authentic racemic standard.

3

3

>20:1

>20:1

23

65

80:20

84:16

Et₃N (2.0)

Et₃N (2.0)

7

8

Et₂O

 CH_2CI_2

Pd-3

Pd-3

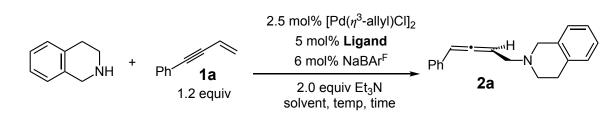


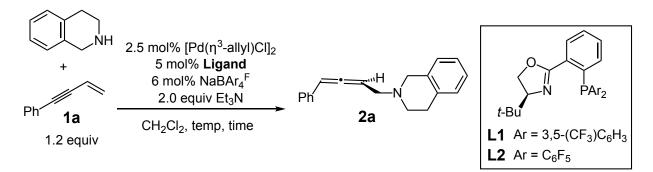
Table S3. Enantioselective Reaction Ligand Screening and Optimization^a

entry	ligand	solvent	temp (°C)	time (h)	yield 3a (%) ^b	er ^c
entry	ngana	30146111	temp (C)	une (n)	yielu Sa (70)	CI
1	(<i>R</i>)-BINAP	Et ₂ O	22	3	82	50:50
2	(R,S)-Josiphos	Et ₂ O	22	3	30	50:50
3	(S,S)-Ph-BPE	Et ₂ O	22	3	85	70:30
4	(S,S)-Me-BPE	Et ₂ O	22	3	<5	_
5	(S,S)-BDPP	Et ₂ O	22	3	82	50:50
6	(R,R)-Taniaphos	Et ₂ O	22	3	83	50:50
7	(R)-BTFM-Garphos	Et ₂ O	22	3	19	80:20
8	(S,S)-QuinoxP*	Et ₂ O	22	3	34	66:34
9	L4	Et ₂ O	22	3	63	50:50
10	L2	Et ₂ O	22	3	75	79.5:20.5
11	L5	Et ₂ O	22	3	77	75:25
12	L6	Et ₂ O	22	3	75	50:50
13	L3	Et ₂ O	22	3	73	84.5:15.5
14	L3	CH_2CI_2	22	3	66	86.5:13.5
15	L3	CH_2CI_2	22	19	78	82.5:17.5
16	L3	CH_2CI_2	0-4	3	33	93.5:6.5
17	L3	CH_2CI_2	0-4	19	63	92.5:7.5
18	L7	CH_2CI_2	0-4	3	37	92.5:7.5

^aReactions run with 0.20 mmol THIQ in 0.25 mL solvent. ^bIsolated yield of pure **3a**. ^cDetermined by HPLC analysis of pure **2a** in comparison with an authentic racemic standard.

 $\begin{array}{c} \begin{array}{c} & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\ & & \\$

Table S4. Experiments Performed in Duplicate or Triplicate to Compare Reaction Reproducibility with Different Ligand Electronics^{*a*}

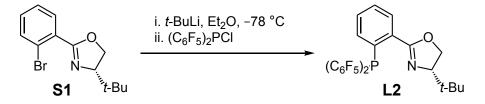


entry	ligand	solvent	temperature (°C)	time (h)	trial	yield 2a (%) ^b	er ^c
1	L1	Et ₂ O	22	3	1	73	74:26
					2	48	71.5:28.5
					3	70	51.5:48.5
2	L1	CH ₂ Cl ₂	22	3	1	65	82:18
					2	65	54:46
					3	70	76.5:23.5
3	L1	CH_2CI_2	22	20	1	68	61:39
					2	67	50:50
4	L1	CH ₂ Cl ₂	0-4	20	1	50	86:14
					2	62	71:29
5	L2	CH ₂ Cl ₂	22	3	1	67	82:18
					2	67	82.5:17.5
					3	73	82:18

^aReactions run with 0.20 mmol THIQ in 0.25 mL solvent. ^bIsolated yield of pure **2a**. ^cDetermined by HPLC analysis of pure **2a** in comparison with an authentic racemic standard.

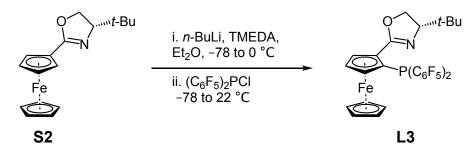
III. Ligand Synthesis

Ligands L1 and L4 were prepared as previously described.^{7,8}



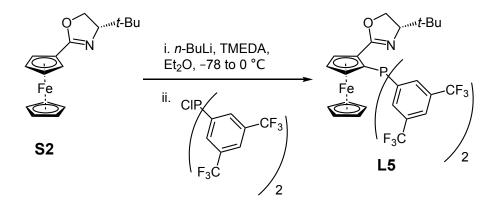
S1 was prepared by a known method.⁹

L2: To an oven dried 50-mL round-bottom flask equipped with a magnetic stirring rod was added S1 (847 mg, 3.00 mmol, 1.00 equiv) and Et₂O (15 mL). The solution was allowed to cool to -78 °C and t-BuLi (1.7 M in pentane, 2.30 mL, 3.9 mmol, 1.30 equiv) was added dropwise via syringe. reaction mixture was allowed to stir at -78 °C for 1 h and then The chlorobis(pentafluorophenyl)phosphane (80% pure by wt., 1.95 g, 3.90 mmol, 1.30 equiv) [note: for this material the remaining 20 wt.% was tris(pentafluorophenyl)phosphine and does not affect reactivity] was added as a solution in Et_2O (3 mL) via cannula transfer. The mixture was allowed to warm to ambient temperature and stir for 1 h. The reaction was quenched with sat aq NH_4Cl (50 mL). The aqueous layer was separated from the organics and was washed with Et_2O (20 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. L2 (1.37 g, 2.42 mmol, 80.7% yield) was obtained as a white solid after flash silica gel chromatography (99:1 to 95:5 hexanes: EtOAc). IR (neat, cm⁻¹) 2959 (w), 1650 (m), 1640 (m), 1513 (s), 1467 (s), 1381 (m), 1358 (m), 1309 (w), 1285 (m), 1256 (w), 1209 (w), 1138 (w), 1082 (s), 1051 (m), 1026 (w), 973 (s); ¹**H** NMR (400 MHz, CDCl₃) δ 7.99 (1H, ddd, J = 7.7, 5.0, 1.4 Hz), 7.52–7.47 (1H, m), 7.39 (1H, td, J = 7.7, 1.4 Hz), 7.15 (1H, dd, J = 7.4, 3.7 Hz), 4.34 (1H, dd, J = 10.1, 8.7 Hz), 4.16 (1H, dd, J = 10.1, 8.7 Hz), 4.16t, J = 8.7 Hz), 3.92 (1H, dd, J = 10.1, 8.8 Hz), 0.74 (9H, s); ¹³C NMR (125 MHz, CDCl₃) δ 162.2 (d, J = 5.0 Hz), 149.2-148.3 (m), 147.0-146.3 (m), 143.6-142.5 (m), 141.5-140.5 (m), 141.5-1138.9-138.1 (m), 136.9-136.0 (m), 132.4, 132.0 (d, J = 25.1 Hz), 131.3 (d, J = 22.1 Hz), 130.7, 129.6 (d, J = 1.8 Hz), 129.5, 112.2–111.4 (m), 77.1 (d, J = 1.5 Hz), 69.0, 33.5, 25.5 (note: aryl carbons show complex splitting due to ¹⁹F and ³¹P atoms); ¹⁹F NMR (376 MHz, CDCl₃) δ -129.61 --130.00 (2F, m), -130.19 --130.63 (2F, m), -151.09 --151.33 (1F, m), -151.53 --151.80 (1F, m), -160.95 - -161.16 (2F, m), -161.17 - -161.44 (2F, m); ³¹P NMR (162 MHz, CDCl₃) δ 75.2–74.2 (1P, m); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₅H₁₆F₁₀NOP: 568.0883, found: 568.0886: **MP** = 61–65 °C.

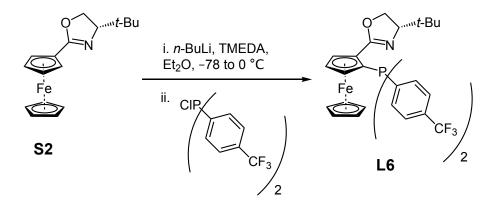


S2 was prepared from ferrocenecarboxylic acid and (*S*)-*tert*-leucinol by a previously reported method and spectral data match those previously reported.^{10,11} The synthesis of all ferrocene and ruthenocene based ligands is based on a previously reported method for related ligands and the stereoisomer depicted is based on the observed diastereoselectivity for this related procedure.¹⁰

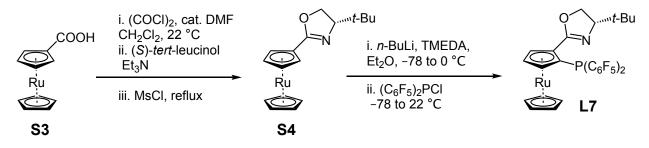
L3: To an oven-dried 50-mL round-bottom flask equipped with a magnetic stirring rod was added **S2** (622 mg, 2.00 mmol, 1.00 equiv), TMEDA (0.450 mL, 3.01 mmol, 1.51 equiv) and Et₂O (24 mL). The solution was then allowed to cool to -78 °C and subsequently n-BuLi (2.50 M in hexanes, 1.20 mL, 3.00 mmol, 1.50 equiv) was added dropwise via syringe. The mixture was allowed to stir at -78 °C for 0.5 h and then warm to 0 °C and stir for an additional 0.5 h. The resulting red solution was then allowed to cool to -78°C and chlorobis(pentafluorophenyl)phosphane (94% pure by wt., 1.11 g, 2.60 mmol, 1.30 equiv) was added as a solution in Et₂O (4.0 mL) via cannula transfer. The resulting solution was allowed to warm gradually to ambient temperature and stir for 1 h. The reaction mixture was then quenched with sat aq NaHCO₃ (30 mL). The aqueous layer was separated from organics and washed with Et₂O (30 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. L3 (986 mg, 1.46 mmol, 73.0% yield) was obtained as an orange solid after flash silica gel chromatography (98:2 to 95:5 hexanes:EtOAc). ¹H NMR analysis indicated L3 was formed as a single detectable diastereomer. **IR** (neat, cm⁻¹) 2957 (w) 1657 (m), 1638 (w), 1513 (s), 1470 (s), 1382 (m), 1365 (w), 1285 (m), 1256 (w), 1171 (w), 1151 (m), 1084 (s); ¹H NMR (400 MHz, CDCl₃) δ 4.88 (1H, dt, J = 2.7, 1.4 Hz), 4.41 (1H, t, J = 2.6 Hz), 4.22–4.14 (2H, m), 4.17 (5H, s), 3.92 (1H, br s), 3.66 (1H, dd, J = 9.9, 7.4 Hz), 0.87 (9H, s); ¹³C NMR (125 MHz, CDCl₃) δ 163.9 (d, J = 3.8 Hz), 149.5-149.0 (m), 148.3-147.8 (m), 147.5-147.1 (m), 146.2-141.7 (m),138.9-138.5 (m), 138.1-137.7 (m), 136.9-136.5 (m), 136.2-135.6 (m), 76.3 (d, J = 1.9 Hz), 73.3, 73.2, 72.4, 72.4, 70.9, 70.9, 70.9, 69.0, 33.5, 29.6 (note: aryl carbons show complex splitting due to ¹⁹F and ³¹P atoms); ¹⁹F NMR (376 MHz, CDCl₃) δ -127.0 – -127.5 (2F, m), -133.3 – -134.0 (2F, m), -148.7 - -148.9 (1F, m), $-153.9 (1F, t, J_F = 20.6 Hz)$, $-159.8 (2F, ddd, J_F = 23.8, 20.7, 9.3)$ Hz), -162.7 - -162.9 (2F, m); ³¹P NMR (162 MHz, CDCl₃) δ 68.0–67.0 (1P, m); HRMS (ESI⁺) $[M+H]^+$ calc'd for C₂₉H₂₀F₁₀[⁵⁶Fe]NOP: 676.0545, found: 676.0546; **MP** = 75-80 °C.



L5: To an oven-dried 100-mL round-bottom flask equipped with a magnetic stirring rod was added **S2** (966 mg, 3.20 mmol, 1.00 equiv), TMEDA (0.620 mL, 4.16 mmol, 1.30 equiv) and Et₂O (35 mL). The solution was then allowed to cool to -78 °C and subsequently *n*-BuLi (2.00 M in cyclohexane, 2.10 mL, 4.16 mmol, 1.30 equiv) was added dropwise via syringe. The mixture was allowed to stir at -78 °C for 1 h and then warm to 0 °C and stir for an additional 0.5 h. Chlorobis[3,5-bis(trifluoromethyl)phenyl]phosphine (2.05 g, 4.16 mmol, 1.30 equiv) was then added as a solution in Et_2O (5.0 mL) via cannula transfer. The resulting solution was allowed to warm gradually to ambient temperature and stir for 1 h. The mixture was then guenched with sat aq NH₄Cl (30 mL). The aqueous layer was separated from the organics and washed with Et_2O (30 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. L5 (2.07 g, 2.69 mmol, 84.1% yield) was obtained as an orange solid after flash silica gel chromatography (100% hexanes to 95:5 hexanes:EtOAc). ¹H NMR analysis indicated L5 was formed as a single detectable diastereomer. IR (neat, cm⁻¹) 1656 (m), 1353 (s), 1277 (s), 1171 (m), 1120 (s), 1093 (m), 1037 (w), 984 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.91 (1H, s), 7.88 (1H, s), 7.87 (1H, s), 7.78 (1H, s), 7.66 (1H, s), 7.64 (1H, s), 4.94 (1H, dt, J = 2.7, 1.4 Hz), 4.44 (1H, t, J = 2.6 Hz), 4.21(5H, s), 4.16 (1H, app. t, J = 8.9 Hz), 4.09 (1H, app. t, J = 7.5 Hz), 3.61 (1H, dd, J = 10.0, 7.4 Hz), 3.51-3.46 (1H, m), 0.88 (9H, s); ¹³C NMR (125 MHz, CDCl₃) δ 163 (d, J = 3.3 Hz), 142.4 (d, J= 20.1 Hz), 141.6 (d, J = 20.8 Hz), 134.4 (dd, J = 22.1, 2.5 Hz), 132.5 (dd, J = 19.6, 2.4 Hz), 131.7 (qd, J = 32.9, 6.2 Hz), 131.3 (qd, J = 32.7, 6.2 Hz), 123.3 (q, J = 271 Hz), 123.2 (q, J123.0 (t, J = 3.6 Hz), 122.3 (t, J = 3.7 Hz), 76.4, 76.3, 74.2 (d, J = 12.3 Hz), 73.3 (d, J = 5.3 Hz), 73.0 (d, J = 1.5 Hz), 71.4, 70.9, 68.6, 33.5, 25.7; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.89, -62.98, ³¹P NMR (162 MHz, CDCl₃) δ –16.6; HRMS (ESI⁺) [M+H]⁺ calc'd for C₃₃H₂₆F₁₂[⁵⁶Fe]NOP: 768.0983, found: 768.0984; **MP** = 125–127 °C.



L6: To an oven-dried 100-mL round-bottom flask equipped with a magnetic stirring rod was added **S2** (966 mg, 3.20 mmol, 1.00 equiv), TMEDA (0.620 mL, 4.16 mmol, 1.30 equiv) and Et₂O (35 mL). The solution was then allowed to cool to -78 °C and subsequently *n*-BuLi (2.00 M in cyclohexane, 2.10 mL, 4.16 mmol, 1.30 equiv) was added dropwise via syringe. The mixture was allowed to stir at -78 °C for 1 h and then warm to 0 °C and stir for an additional 0.5 h. Chlorobis[4-(trifluoromethyl)phenyl]phosphine (1.48 g, 4.16 mmol, 1.30 equiv) was then added as a solution in Et₂O (5.0 mL) via cannula transfer. The resulting solution was allowed to warm gradually to ambient temperature and stir for 1 h. The mixture was then guenched with sat ag NH₄Cl (30 mL). The aqueous layer was separated from the organics and washed with Et₂O (30 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. L6 (1.72 g, 2.73 mmol, 85.3% yield) was obtained as a vellow solid after flash silica gel chromatography (99:1 to 93:7 hexanes:EtOAc). ¹H NMR analysis indicated L6 was formed as a single detectable diastereomer. **IR** (neat, cm⁻¹) 2965 (w), 1652 (m), 1604 (w), 1483 (w), 1394 (m), 1364 (w), 1323 (s), 1279 (w), 1254 (w), 1209 (w), 1165 (s), 1121 (s), 1058 (s), 1007 (m), 985 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.61 (1H, d, J = 7.9 Hz), 7.56 (1H, t, J = 6.5 Hz), 7.47 (1H, d, J = 7.9 Hz), 7.29 (1H, t, J = 7.3Hz), 4.94 (1H, dt, J = 2.6, 1.2 Hz), 4.39 (1H, t, J = 2.5 Hz), 4.20 (5H, s), 4.16 (1H, dd, J = 9.8, 8.6 Hz), 3.97 (1H, app. t, J = 7.5 Hz), 3.69 (1H, dd, J = 9.9, 7.4 Hz), 3.59–3.55 (1H, m), 0.818 (9H, s); ¹³C NMR (125 MHz, CDCl₃) δ 163.6 (d, J = 2.9 Hz), 144.6 (d, J = 16.0 Hz), 143.1 (d, J = 17.0 Hz), 135.3 (d, J = 21.8 Hz), 132.5 (d, J = 19.3 Hz), 131.1 (q, J = 34.4 Hz), 130.0 (q, J = 32.0 Hz), 125.0 (dt, J = 10.3, 3.6 Hz), 124.7 (dt, J = 13.7, 3.7 Hz), 124.2 (q, J = 271 Hz), 124.0 (q, J = 271Hz), 76.2, 76.2, 76.1 (d, J = 3.4 Hz), 73.6 (d, J = 4.8 Hz), 72.6 (d, J = 1.5 Hz), 71.0, 70.8, 68.5, 33.6, 25.7; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.66, -62.75; ³¹P NMR (162 MHz, CDCl₃) δ -18.0; **HRMS** (ESI⁺) $[M+H]^+$ calc'd for $C_{31}H_{28}F_6[{}^{56}Fe]NOP$: 632.1235, found: 632.1238; **MP** = 197–199 °C (decomp.).



Ruthenocenecarboxylic acid (S3) was prepared from ruthenocene by a known method.¹²

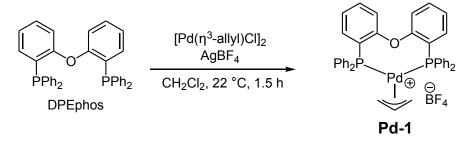
S4: To a dry 50-mL round-bottom flask equipped with a magnetic stirring rod was added ruthenocene carboxylic acid (S3, 743 mg, 2.70 mmol, 1.00 equiv), which was suspended in CH₂Cl₂ (7.5 mL) with one drop of DMF added. The suspension was then treated with oxalyl chloride (0.460 mL, 5.40 mmol, 2.00 equiv) and the resulting yellow solution was allowed to stir at ambient temperature for 1 h. Volatiles were then removed by mild vacuum distillation with gentle heating (warm water bath) to reveal the acid chloride as a vellow solid. The acid chloride was then dissolved in CH₂Cl₂ (7 mL) and added to a solution of (S)-tert-leucinol (333 mg, 2.84 mmol, 1.05 equiv) and Et₃N (1.90 mL, 13.5 mmol, 5.00 equiv) in CH₂Cl₂ (3 mL) via cannula transfer. The reaction mixture was allowed to stir at ambient temperature for 1.5 h, and then methanesulfonyl chloride (0.230 mL, 3.0 mmol, 1.10 equiv) was added. A reflux condenser was added to the flask and the reaction mixture was brought to reflux, which was maintained for 11 h. The reaction mixture was then allowed to cool to ambient temperature and was quenched with H₂O (20 mL). The aqueous layer was separated from the organics and washed with CH_2Cl_2 (3 X 10 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. S4 (580 mg, 1.63 mmol, 60.3% yield) was obtained as a white solid after flash silica gel chromatography (85:15 hexanes: EtOAc). ¹H NMR (400 MHz, CDCl₃) δ 5.16–5.13 (1H, m), 5.04–5.02 (1H, m), 4.67–4.61 (2H, m), 4.55 (5H, s), 4.13 (1H, dd, J = 9.9, 8.6 Hz), 4.07 (1H, dd, J = 8.6, 7.0 Hz), 3.81 (1H, dd, J = 9.9, 7.0 Hz), 0.88 (9H, s). Spectral data match those previously reported.¹³

L7: To an oven-dried 25-mL round-bottom flask equipped with a magnetic stirring rod was added S4 (178 mg, 0.500 mmol, 1.00 equiv), TMEDA (97.0 µL, 0.650 mmol, 1.30 equiv) and Et₂O (8 mL). The solution was then allowed to cool to -78 °C and subsequently *n*-BuLi (2.00 M in cyclohexane, 0.33 mL, 0.65 mmol, 1.30 equiv) was added dropwise via syringe. The mixture was allowed to stir at -78 °C for 1 h and then warm to 0 °C and stir for an additional 0.5 h. Chlorobis(pentafluorophenyl)phosphane (94% pure by wt., 277 mg, 0.650 mmol, 1.30 equiv) was then added as a solution in Et₂O (2.0 mL) via cannula transfer. The resulting solution was allowed to warm gradually to ambient temperature and stir for 1 h. The reaction mixture was then quenched with sat aq NH₄Cl (10 mL). The aqueous layer was separated from organics and washed with Et₂O (2 X 10 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. L7 (218 mg, 0.290 mmol, 58.1% yield) was obtained as a pale yellow solid after flash silica gel chromatography (95:5 hexanes:EtOAc). ¹H NMR analysis indicated L7 was formed as a single detectable diastereomer. IR (neat, cm⁻¹) 2953 (w), 1659 (m), 1638 (w), 1513 (s), 1468 (s), 1382 (m), 1365 (w), 1284 (m), 1254 (w), 1209 (w), 1170 (w), 1149 (m), 1083 (s), 1030 (w), 972 (s); ¹H **NMR** (400 MHz, CDCl₃) δ ; 5.21–5.19 (1H, m), 4.27–4.70 (1H, m), 5.43 (5H, s), 4.32–4.29 (1H, m), 4.16 (1H, app. t, J = 8.8 Hz), 4.11 (1H, dd, J = 8.7, 6.9 Hz), 3.65 (1H, dd, J = 9.6, 6.7 Hz), 0.81 (9H, s); ¹³C NMR (125 MHz, CDCl₃) δ 162.7 (t, J = 3.9 Hz), 149.3–148.8 (m), 148.0–147.6

(m), 147.3–146.8 (m), 146.1–145.5 (m), 142.1–141.3 (m), 140.4–139.6 (m), 138.3–138.2 (m), 138.0–137.5 (m), 136.8–136.2 (m), 136.1–135.6 (m), 80.8–80.4 (m), 76.1–75.9 (m), 75.9–75.8 (m), 73.8, 73.2, 72.8, 68.8, 33.9, 25.3 (note: aryl carbons show complex splitting due to ¹⁹F and ³¹P atoms); ¹⁹F NMR (376 MHz, CDCl₃) δ -127.67 – -127.88 (2F, m), -134.10 – -134.38 (2F, m), -148.95 – -149.15 (1F, m), -153.91 (1F, t, J_{CF} = 20.9 Hz), -159.99 – -160.21 (2F, m), -162.59 – 162.87 (2F, m); ³¹P NMR (162 MHz, CDCl₃) δ 67.8–66.5 (1P, m); HRMS (ESI⁺) [M+H]⁺ calc'd for; C₂₉H₂₀F₁₀NOP[¹⁰²Ru]: 722.0239, found: 722.0243; **MP** = 83–88 °C.

IV. Pd Complex Synthesis

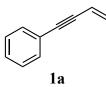
The synthesis of **Pd-2** and **Pd-3** have been reported previously.^{7,14} **Pd-1** was prepared by an analogous procedure.



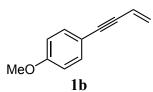
Pd-1: In an N₂-filled glovebox, to a dry 250-mL round-bottom flask equipped with a magnetic stirring rod was added (oxybis(2,1-phenylene))bis(diphenylphosphane) (DPEphos, 1.24 g, 2.30 mmol, 1.00 equiv), $[Pd(\eta^3-allyl)Cl]_2$ (420 mg, 1.15 mmol, 0.500 equiv), and CH_2Cl_2 (50 mL). The solution was allowed to stir at ambient temperature for 20 min and then AgBF₄ (493 mg, 2.53 mmol, 1.10 equiv) was added. Vigorous stirring was allowed to continue for 1 h during which time a white precipitate formed. The following work up took place outside of the glovebox. The heterogenous reaction mixture was filtered through a pad of celite eluting with CH₂Cl₂ (3 X 20 mL). The solution was then concentrated to reveal Pd-1 as an off-white powder (1.77 g, 2.06 mmol, 89.7% vield). ¹H NMR analysis revealed **Pd-1** to be formed as a 1:1 adduct with CH₂Cl₂. **IR** (neat, cm⁻¹); 3057 (w), 1567 (w), 1479 (w), 1465 (m), 1434 (s), 1262 (m), 1215 (s), 1187 (w), 1092 (m), 1052 (s), 1020 (s), 958 (m); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.56–7.29 (18H, m), 7.26-7.18 (4H, m), 7.09-7.00 (4H, m), 6.61-6.52 (2H, m), 6.14-6.00 (1H, m), 5.72 (2H, s, CH₂Cl₂), 4.06–3.93 (2H, m), 3.70–3.54 (2H, m); ¹³C NMR (125 MHz, DMSO- d_6) δ 158.2 (t, J = 3.8 Hz), 134.4, 133.9 (t, J = 6.6 Hz), 133.7, 133.4 (t, J = 6.4 Hz), 131.7, 131.5, 129.5 (q, J = 5.3Hz), 125.7, 123.7 (t, J = 5.0 Hz), 122.2, 122.0, 121.8, 121.3, 79.2, 79.1, 77.0, 55.4 (CH₂Cl₂); ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ-148.31, -148.36; ³¹P NMR (162 MHz, DMSO-*d*₆) δ 13.0; HRMS (ESI⁺) $[M+H]^+$ calc'd for C₃₉H₃₃OP₂[¹⁰⁶Pd]: 685.1036, found: 685.1055; **MP** = 255–260 °C (decomp.).

V. Substrate Synthesis

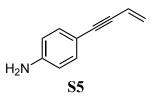
Enyne substrates were prepared by Sonogashira coupling¹⁵ from their corresponding terminal alkynes with vinyl bromide by a slightly modified method. <u>General Method A</u> is exemplified by the synthesis of 1a.



but-3-en-1-yn-1-ylbenzene (1a): General Method A: In an N₂-filled glovebox, to an oven-dried 250-mL round-bottom flask equipped with a magnetic stirring bar was added Pd(PPh₃)₄ (462 mg, 0.400 mmol, 2.00 mol%) and CuI (381 mg, 2.00 mmol, 10.0 mol%). The flask was sealed with a rubber septum and removed from the glovebox. Vinyl bromide (1.0 M in THF, 50.0 mL, 2.50 equiv) was added followed by freshly distilled and degassed Et₂NH (10.4 mL, 100 mmol, 5.00 equiv). The mixture was allowed to stir at ambient temperature for *ca*. 5 min and subsequently phenylacetylene (2.20 mL, 20.0 mmol, 1.00 equiv) was added as a solution in THF (10 mL) dropwise over 1 h. The mixture was then allowed to stir at ambient temperature overnight (15–20 h). The reaction contents were then filtered through a pad of celite, eluting with Et₂O (3 X 20 mL). The solution was then concentrated *in vacuo*. **1a** was obtained by flash silica gel chromatography (99:1 hexanes:Et₂O) to afford a colorless oil (2.43 g, 18.9 mmol, 94.6%). **¹H NMR** (400 MHz, CDCl₃) δ 7.48–7.41 (2H, m), 7.34–7.26 (3H, m), 6.01 (1H, dd, *J* = 17.6, 11.1), 5.73 (1H, dd, *J* = 17.6, 2.0 Hz), 5.54 (1H, dd, *J* = 11.1, 2.0 Hz). Spectral data match those previously reported.¹⁵

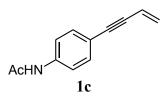


1-(but-3-en-1-yn-1-yl)-4-methoxybenzene (1b): Prepared by General Method A in 92.5% yield after flash silica gel chromatography (98:2 to 95:5 hexanes:EtOAc). ¹H NMR (400 MHz, CDCl₃) δ 7.39–7.34 (2H, m), 6.86–6.80 (2H, m), 5.99 (1H, dd, J = 17.5, 11.1 Hz), 5.68 (1H, dd, J = 17.5, 2.0 Hz), 5.48 (1H, dd, J = 11.1, 2.0 Hz), 3.80 (3H, s). Spectral data match those previously reported.¹⁶

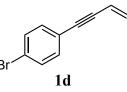


4-(but-3-en-1-yn-1-yl)aniline (S5): Prepared by General Method A in 89.0% yield after flash silica gel chromatography (80:20 hexanes:EtOAc). **IR** (neat, cm⁻¹) 3466 (m), 3376 (m), 3211 (w), 3033 (w), 3005 (w), 2212 (m), 2177 (m), 1618 (s), 1598(s), 1510 (s), 1432 (w), 1410 (w), 1284 (s), 1176 (s), 1127 (w), 1077 (w), 969 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.29–7.17 (2H, m),

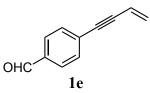
6.61–6.55 (2H, m), 5.99 (1H, dd, J = 17.5, 11.1 Hz), 5.64 (1H, dd, J = 17.5, 2.1 Hz), 5.44 (1H, dd, J = 11.1, 2.1 Hz), 3.79 (2H, br s); ¹³**C NMR** (125 MHz, CDCl₃) δ 146.7, 132.9, 125.4, 117.5, 114.7, 112.4, 90.8, 86.2; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₀H₉N: 144.0808, found: 144.0810.



N-(4-(but-3-en-1-yn-1-yl)phenyl)acetamide (1c): Prepared from S5 by the following procedure. To an oven-dried 25-mL round-bottom flask equipped with a magnetic stirring rod was add S5 (430 mg, 3.00 mmol, 1.00 equiv), pyridine (0.480 mL, 5.96 mmol, 1.99 equiv), and CH₂Cl₂ (5.0 mL). The mixture was allowed to cool to 0 °C and acetyl chloride (0.430 mL, 6.05 mmol, 2.02 equiv) was added. The mixture was then allowed to warm to ambient temperature and stir for 3 h. The reaction was then quenched with sat aq NaHCO₃ (20 mL). The aqueous layer was separated from the organics and washed with CH₂Cl₂ (2 X 20 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. **1c** was obtained as a pale yellow solid (462 mg, 2.49 mmol, 83.2% yield) after purification by flash silica gel chromatography (60:40 hexanes:EtOAc). **IR** (neat, cm⁻¹) 3252 (w), 3103 (w), 1660 (m), 1594 (s), 1530 (s), 1505 (s), 1403 (m), 1369 (m), 1319 (s), 1296 (w), 1256 (m), 1178 (w), 1111 (w), 1080 (w), 1040 (w), 1012 (w), 966 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.46 (2H, d, *J* = 8.6 Hz), 7.42–7.32 (3H, m), 5.99 (1H, dd, *J* = 17.5, 11.1), 5.70 (1H, dd, *J* = 17.5, 2.0 Hz), 5.51 (1H, dd, *J* = 11.1, 2.0 Hz), 2.16 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 168.7, 138.0, 132.3, 126.7, 119.5, 118.7, 117.1, 89.7, 87.8, 24.6; HRMS (ESI⁺) [M+H]⁺ calc'd for C₁₂H₁₁NO: 186.0913, found: 186.0917; **MP** = 144–147 °C.

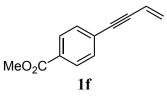


1-bromo-4-(but-3-en-1-yn-1-yl)benzene (1d): Prepared by General method A in 79.1% yield after flash silica gel chromatography (99:1 hexanes:Et₂O). ¹H NMR (400 MHz, CDCl₃) δ 7.46–7.40 (2H, m), 7.31–7.26 (2H, m), 5.98 (1H, dd, J = 17.6, 11.1 Hz), 5.73 (1H, dd, J = 17.6, 2.0 Hz), 5.55 (1H, dd, J = 11.1, 2.0 Hz). Spectral data match those previously reported.¹⁷

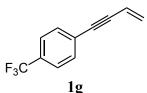


4-(but-3-en-1-yn-1-yl)benzaldehyde (1e): Prepared by General Method A in 86.6% yield after flash silica gel chromatography (95:5 hexanes:EtOAc). **IR** (neat, cm⁻¹) 2845 (w), 2744 (w), 2215 (w), 1693 (s), 1599 (s), 1556 (m), 1503 (w), 1413 (m), 1386 (m), 1300 (m), 1287 (m), 1265 (m), 1204 (s), 1163 (s), 1103 (m), 1013 (w), 1001 (w), 970 (m); ¹H NMR (400 MHz, CDCl₃) δ 9.99 (1H, s), 7.82 (2H, d, J = 8.2 Hz), 7.57 (2H, d, J = 8.2 Hz), 6.02 (1H, dd, J = 17.6, 11.2 Hz), 5.79 (1H, dd, J = 17.6, 1.9 Hz), 5.62 (1H, dd, J = 11.2, 1.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 191.4,

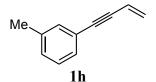
135.4, 132.0, 129.5, 129.4, 128.4, 116.7, 92.0, 89.0; **HRMS** (ESI⁺) $[M+H]^+$ calc'd for C₁₁H₈O: 157.0648, found: 157.0647.



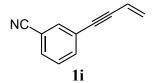
methyl 4-(but-3-en-1-yn-1-yl)benzoate (1f): Prepared by General Method A in 83.0% yield after flash silica gel chromatography (95:5 hexanes:EtOAc). **IR** (neat, cm⁻¹) 2950 (w), 2850 (w), 1707 (s), 1605 (s), 1436 (s), 1406 (m), 1307 (w), 1271 (s), 1193 (w), 1174 (m), 1105 (s), 1015 (m), 961 (m); ¹H NMR (400 MHz, CDCl₃) δ 8.00–7.94 (2H, m), 7.51–7.44 (2H, m), 6.01 (1H, dd, *J* = 17.5, 11.1 Hz), 5.76 (1H, dd, *J* = 17.5, 1.9 Hz), 5.59 (1H, dd, *J* = 11.1, 1.9 Hz), 3.89 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 166.5, 131.4, 129.4, 128.0, 127.8, 116.8, 90.9, 89.1, 52.2 (one sp² carbon missing due to overlap); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₂H₁₀O₂: 187.0754, found: 187.0756; **MP** = 34–36 °C.



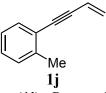
1-(but-3-en-1-yn-1-yl)-4-(trifluoromethyl)benzene (1g): Prepared by General Method A in 99.0% yield after flash silica gel chromatography (99:1 hexanes:Et₂O). ¹H NMR (400 MHz, CDCl₃) δ 7.56 (2H, AB_q, J_{AB} = 8.6 Hz), 7.52 (2H, AB_q, J_{AB} = 8.6 Hz), 6.01 (1H, dd, J = 17.5, 11.1 Hz), 5.78 (1H, dd, J = 17.5, 1.9 Hz), 5.60 (1H, dd, J = 11.1, 1.9 Hz); ¹⁹F NMR (376 MHz, CDCl₃) δ -62.9. Spectral data match those previously reported.¹⁶



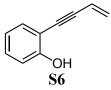
1-(but-3-en-1-yn-1-yl)-3-methylbenzene (1h): Prepared by General Method A in 88.8% yield after flash silica gel chromatography (99:1 hexanes:Et₂O). **IR** (neat, cm⁻¹) 3008 (w), 2920 (w), 2202 (w), 1840 (w), 1607 (m), 1578 (w), 1484 (m), 1450 (w), 1380 (w), 1281 (w), 1181 (w), 1094 (w), 1080 (w), 1040 (w), 969 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.29–7.22 (2H, m), 7.19 (1H, dd, J = 7.5, 7.5 Hz), 7.14–7.09 (1H, m), 6.00 (1H, dd, J = 17.5, 11.2 Hz), 5.71 (1H, dd, J = 17.5, 2.0 Hz) 5.52 (1H, dd, J = 11.2, 2.0 Hz), 2.32 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 138.0, 132.2, 129.2, 128.7, 128.2, 126.7, 122.9, 117.3, 90.2, 87.8, 21.2; **EA** calc' for C₁₁H₁₀: C = 92.91%, H = 7.09%, found: C = 92.65%, H = 7.12%.



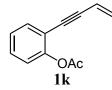
3-(but-3-en-1-yn-1-yl)benzonitrile (1i): Prepared by General Method A in 87.5% yield after flash silica gel chromatography (95:5 hexanes:EtOAc). **IR** (neat, cm⁻¹) 3070 (w), 3012 (w), 2230 (m), 1850 (w), 1722 (w), 1600 (m), 1571 (w), 1477 (s), 1417 (w), 1407 (w), 1319 (w), 1288 (w), 1267 (w), 1167 (w), 1095 (w), 1082 (w), 969 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.70 (1H, dd, J = 1.6,1.6 Hz), 7.63 (1H, ddd, J = 7.8, 1.4, 1.4 Hz), 7.57 (1H, ddd, J = 7.8, 1.4, 1.4 Hz), 7.42 (1H, dd, J = 7.8, 7.8 Hz), 5.99 (1H, dd, J = 17.5, 11.1 Hz), 5.78 (1H, dd, J = 17.5, 12.8, 90.4, 87.4; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₁H₇N: 154.0651, found: 154.0649.



1-(but-3-en-1-yn-1-yl)-2-methylbenzene (1j): Prepared by General Method A in 89.0% yield after flash silica gel chromatography (99:1 hexanes:Et₂O). **IR** (neat, cm⁻¹) 3009 (w), 2920 (w), 2212 (w), 1839 (w), 1605 (m), 1484 (m), 1455 (m), 1409 (w), 1379 (w), 1289 (w), 1256 (w), 1197 (w), 1158 (w), 1116 (m), 1042 (w), 968 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.40 (1H, d, J = 7.3 Hz), 7.23–7.16 (2H, m), 7.15–7.08 (1H, m), 6.05 (1H, dd, J = 17.5, 11.1 Hz), 5.72 (1H, dd, J = 17.5, 2.0 Hz), 5.53 (1H, dd, J = 11.1, 2.0 Hz), 2.44 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 140.2, 131.9, 129.4, 128.3, 126.5, 125.5, 122.9, 117.4, 92.0, 88.9, 20.7; **EA** calc' for C₁₁H₁₀: C = 92.91%, H = 7.09%, found: C = 91.58%, H = 6.89%.

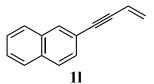


2-(but-3-en-1-yn-1-yl)phenol (S6): Prepared by General Method A in 61.2% yield after flash silica gel chromatography (95:5 hexanes:EtOAc). ¹H NMR (400 MHz, CDCl₃) δ 7.33 (1H, dd, J = 7.7, 1.6 Hz), 7.27–7.19 (1H, m), 6.94 (1H, dd, J = 8.3, 0.9 Hz), 6.86 (1H, td, J = 7.6, 1.1 Hz), 6.05 (1H, dd, J = 17.5, 11.2 Hz), 5.77 (1H, dd, J = 17.5, 1.8 Hz), 5.72 (1H, br s), 5.60 (1H, dd, J = 11.2, 1.8 Hz). Spectral data match those previously reported.¹⁸

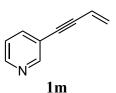


2-(but-3-en-1-yn-1-yl)phenyl acetate (1k): Prepared from **S6** by the following procedure. To an oven-dried 25-mL round-bottom flask equipped with a magnetic stirring rod was added **S6** (288 mg, 2.00 mmol, 1.00 equiv), pyridine (0.320 mL, 3.97 mmol 1.99 equiv), and CH₂Cl₂ (4.0 mL).

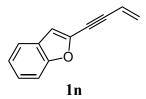
The solution was cooled to 0 °C and acetyl chloride (0.280 mL, 3.94 mmol, 1.97 equiv) was added. The solution was allowed to warm to ambient temperature and stir for 3 h. The reaction mixture was quenched with sat aq NaHCO₃ (20 mL) and the aquous layer was separated from the organics and washed with CH₂Cl₂ (2 X 20 mL). The combined organic fractions were dried over MgSO₄, filtered and concentrated. **1k** (342 mg, 1.84 mmol, 91.8% yield) was obtained as a colorless oil after flash silica gel chromatography (95:5 hexanes:EtOAc). **IR** (neat, cm⁻¹) 3010 (w), 1759 (s), 1590 (w), 1485 (m), 1445 (m), 1367 (m), 1290 (w), 1256 (w), 1207 (m), 1178 (s), 1106 (m), 1077 (w), 1034 (w), 1008 (m), 969 (w); ¹**H** NMR (400 MHz, CDCl₃) δ 7.47 (1H, dd, *J* = 7.7, 1.9 Hz), 7.33 (1H, ddd, *J* = 7.6, 7.6, 1.6 Hz), 7.19 (1H, ddd, *J* = 7.6, 7.6, 1.2 Hz), 7.08 (1H, dd, *J* = 8.1, 1.1 Hz), 6.00 (1H, dd, *J* = 17.6, 11.2 Hz), 5.71 (1H, dd, *J* = 17.6, 2.0 Hz), 5.55 (1H, dd, *J* = 11.2, 2.0 Hz), 2.33 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 168.9, 151.5, 133.0, 129.5, 127.5, 125.9, 122.3, 117.3, 116.9, 92.8, 84.7, 20.8; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₂H₁₀O₂: 187.0754, found: 187.0755.



2-(but-3-en-1-yn-1-yl)naphthalene (11): Prepared by General Method A in 79.5% yield after flash silica gel chromatography (99:1 hexanes:EtOAc). ¹H NMR (400 MHz, CDCl₃) δ 7.96 (1H, s), 7.86–7.70 (3H, m), 7.54–7.41 (3H, s), 6.06 (1H, dd, J = 17.5, 11.1 Hz), 5.78 (1H, dd, J = 17.5, 1.9 Hz), 5.57 (1H, dd, J = 11.1, 1.9 Hz). Spectral data match those previously reported.¹⁹

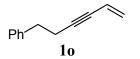


3-(but-3-en-1-yn-1-yl)pyridine (1m): Prepared by General Method A in 82.0% yield after flash silica gel chromatography (70:30 pentanes:Et₂O). **IR** (neat, cm⁻¹) 3030 (w), 1605 (m), 1578 (m), 1552 (m), 1475 (s), 1404 (s), 1327 (w), 1291 (w), 1268 (w), 1186 (m), 1121 (w), 1098 (w), 1021 (m), 969 (m); ¹H NMR (400 MHz, CDCl₃) δ 8.66 (1H, d, J = 1.8 Hz), 8.51 (1H, dd, J = 4.9, 1.6 Hz), 7.70 (1H, ddd, J = 7.9, 1.9, 1.9 Hz), 7.26–7.21 (1H, m), 6.01 (1H, dd, J = 17.6, 11.2 Hz), 5.77 (1H, dd, J = 17.6, 1.9 Hz), 5.60 (1H, dd, J = 11.2, 1.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 152.1, 148.5, 138.3, 128.0, 123.0, 120.3, 116.6, 91.2, 86.5; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₉H₇N 130.0651, found: 130.0653.

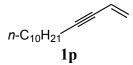


2-(but-3-en-1-yn-1-yl)benzofuran (1n): Prepared by General Method A in 76.1% yield after flash silica gel chromatography (100% hexanes). **IR** (neat, cm⁻¹) 3057 (w), 1854 (w), 1778 (w), 1608 (w), 1557 (w), 1473 (w), 1448 (m), 1408 (w), 1349 (w), 1305 (w), 1288 (w), 1267 (w), 1255 (m),

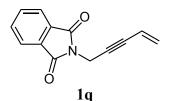
1200 (m), 1160 (m), 1143 (m), 1108 (m), 1073 (w), 966 (m); ¹**H** NMR (400 MHz, CDCl₃) δ 7.54 (1H, d, *J* = 7.6 Hz), 7.44 (1H, d, *J* = 8.2 Hz), 7.32 (1H, ddd, *J* = 7.3, 7.3, 1.3 Hz), 7.23 (1H, dd, *J* = 7.4, 7.4 Hz), 6.93 (1H, s), 6.05 (1H, dd, *J* = 17.6, 11.3 Hz), 5.84 (1H, dd, *J* = 17.6, 1.8 Hz), 5.65 (1H, dd, *J* = 11.3, 1.8 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 154.9, 138.6, 128.7, 127.7, 125.7, 123.3, 121.2, 116.1, 111.7, 111.2, 93.9, 80.2; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₂H₈O: 169.0648, found: 169.0645.



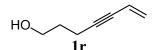
hex-5-en-3-yn-1-ylbenzene (10): Prepared by General Method A in 87.9% yield after flash silica gel chromatography (99:1 hexanes:Et₂O). ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.26 (2H, m), 7.25–7.18 (3H, m), 5.76 (1H, ddt, J = 17.6, 11.0, 2.0 Hz), 5.54 (1H, dd, J = 17.6, 2.2 Hz), 5.38 (1H, dd, J = 11.0, 2.2 Hz), 2.85 (2H, t, J = 7.6 Hz), 2.59 (2H, td, J = 7.6, 2.0 Hz). Spectral data match those previously reported.²⁰



tetradec-1-en-3-yne (1p): Prepared by General Method A in 95.0% yield after flash silica gel chromatography (100% hexanes). **IR** (neat, cm⁻¹) 2922 (s), 2853 (s), 2227 (w), 1608 (w), 1466 (m), 1378 (w), 1328 (w), 1162 (w), 971 (m); 911 (s); ¹H NMR (400 MHz, CDCl₃) δ 5.76 (1H, ddt, J = 17.5, 11.0, 2.0 Hz), 5.52 (1H, dd, J = 17.5, 2.2 Hz), 5.35 (1H, dd, J = 11.0, 2.2 Hz), 2.28 (2H, td, J = 7.1, 2.0 Hz), 1.56–1.47 (2H, m), 1.42–1.15 (14H, m), 0.86 (3H, t, J = 6.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 125.3, 117.6, 91.2, 79.3, 31.9, 29.6, 29.5, 29.3, 29.2, 28.9, 28.7, 22.7, 19.3, 14.1; **EA** calc' for C₁₄H₂₄: C = 87.42%, H = 12.58%, found: C = 87.44%, H = 12.65%.

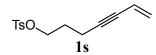


2-(pent-4-en-2-yn-1-yl)isoindoline-1,3-dione (1q): Prepared by General Method A in 82.8% yield after flash silica gel chromatography (80:20 hexanes:EtOAc). ¹H NMR (400 MHz, CDCl₃) δ 7.90–7.83 (2H, m), 7.75–7.69 (2H, m), 5.72 (1H, ddt, J = 17.6, 10.8, 1.8 Hz), 5.62 (1H, dd, J = 17.6, 2.4 Hz), 5.45 (1H, dd, J = 10.8, 2.4 Hz), 4.55 (2H, d, J = 1.9 Hz). Spectral data match those previously reported.²¹

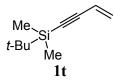


hept-6-en-4-yn-1-ol (1r): Prepared by General Method A in 92.2% yield after flash silica gel chromatography (70:30 to 50:50 pentanes:Et₂O). ¹H NMR (400 MHz, CDCl₃) δ 5.75 (1H, ddt, J = 17.5, 11.0, 2.0 Hz), 5.53 (1H, dd, J = 17.5, 2.2 Hz), 5.37 (1H, dd, J = 11.0, 2.2 Hz), 3.79–3.71

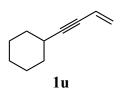
(2H, m), 2.42 (2H, td, J = 7.0, 2.0 Hz), 1.82–1.72 (2H, m), 1.55 (1H, br s). Spectral data match those previously reported.²²



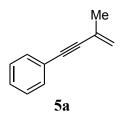
hept-6-en-4-yn-1-yl 4-methylbenzenesulfonate (1s): Prepared from 1r by the following procedure. To an oven-dried 50-mL round-bottom flask equipped with a magnetic stirring rod was added 1r (551 mg, 5.00 mmol, 1.00 equiv), pyridine (1.60 mL, 19.9 mmol, 3.97 equiv), and CH₂Cl₂ (5.0 mL). The solution was allowed to cool to 0 °C and then *p*-toluenesulfonyl chloride (1.43 g, 7.50 mmol, 1.50 equiv) was added portionwise over 10 min. The reaction mixture was allowed to stir at 0 °C for 1 h and then partitioned between H₂O (50 mL) and Et₂O (50 mL). The aqueous layer was separated from the organics and washed with Et₂O (20 mL). The combined organic fractions were dried over MgSO₄, filtered and concentrated. **1s** (1.14 g, 4.30 mmol, 86.0% yield) was obtained as a pale yellow oil after flash silica gel chromatography (90:10 hexanes:EtOAc). IR (neat, cm⁻¹) 2960 (w), 2227 (w), 1598 (m), 1495 (w), 1433 (w), 1357 (s), 1307 (w), 1291 (w), 1188 (m), 1173 (s), 1120 (w), 1096 (m), 973 (m), 921 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.81–7.75 (2H, m), 7.32 (2H, d, J = 8.3 Hz), 5.65 (1H, ddt, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz), 5.46 (1H, dd, J = 17.5, 11.0, 2.0 Hz2.2 Hz), 5.35 (1H, dd, J = 11.0, 2.2 Hz), 4.12 (2H, t, J = 6.1 Hz), 2.42 (3H, s), 2.35 (2H, td, J =6.9, 2.0 Hz), 1.84 (2H, dt, J = 6.4, 6.4 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 144.8, 132.8, 129.9, 127.9, 126.0, 117.2, 88.4, 80.3, 68.9, 27.8, 21.6, 15.5; HRMS (ESI⁺) [M+H]⁺ calc'd for: C₁₄H₁₆O₃S: 265.0893, found: 265.0896.



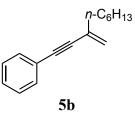
but-3-en-1-yn-1-yl(*tert*-butyl)dimethylsilane (1t): Prepared by General Method A in 84.0% yield after flash silica gel chromatography (100% pentanes). ¹H NMR (400 MHz, CDCl₃) δ 5.80 (1H, dd, J = 17.6, 11.1 Hz), 5.67 (1H, dd, J = 17.6, 2.4 Hz), 5.48 (1H, dd, J = 11.1, 2.4 Hz), 0.93 (9H, s), 0.11 (6H, s). Spectral data match those previously reported.¹⁷



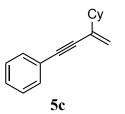
but-3-en-1-yn-1-ylcyclohexane (1u): Prepared by General Method A in 82.7% yield after flash silica gel chromatography (99:1 pentanes:Et₂O). ¹**H NMR** (400 MHz, CDCl₃) δ 5.78 (1H, ddd, J = 17.5, 11.0, 1.9 Hz), 5.52 (1H, dd, J = 17.5, 2.2 Hz), 5.35 (1H, dd, J = 11.0, 2.2 Hz), 2.88–2.52 (1H, m), 1.98–0.97 (10H, m). Spectral data match those previously reported.¹⁷



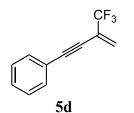
(3-methylbut-3-en-1-yn-1-yl)benzene (5a): Prepared by General Method A in 91.3% yield after flash silica gel chromatography (99:1 hexanes:EtOAc) using 2-bromopropene (2.5 equiv) in place of vinyl bromide. ¹H NMR (400 MHz, CDCl₃) δ 7.47–7.40 (2H, m), 7.34–7.27 (3H, m), 5.41–5.37 (1H, m), 5.31–5.27 (1H, m), 2.00–1.98 (3H, m). Spectral data match those previously reported.²³



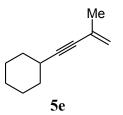
(3-methylenenon-1-yn-1-yl)benzene (5b): Prepared by General Method A in 40.8% yield after flash silica gel chromatography (99:1 hexanes:EtOAc) using 2-bromooct-1-ene (2.5 equiv) in place of vinyl bromide. ¹H NMR (500 MHz, CDCl₃) δ 7.50–745 (2H, m), 7.36–7.29 (3H, m), 5.44 (1H, ABq, J = 1.9 Hz), 5.32 (1H, ABq, J = 1.9 Hz), 2.27 (2H, t, J = 7.6 Hz), 1.68–1.57 (2H, m), 1.43–1.28 (6H, m), 0.94 (3H, t, J = 6.9 Hz). Spectral data match those previously reported.²⁴



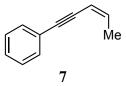
(3-cyclohexylbut-3-en-1-yn-1-yl)benzene (5c): Prepared by General Method A in 83.2% yield after flash silica gel chromatography (99:1 hexanes:EtOAc) using (1-bromovinyl)cyclohexane (2.5 equiv) in place of vinyl bromide. ¹H NMR (400 MHz, CDCl₃) δ 7.54–7.46 (2H, m), 7.39–7.30 (3H, m), 5.43 (1H, s), 5.34 (1H, s), 2.18 (1H, t, *J* = 10.9 Hz), 2.00–1.68 (4H, m), 1.49–1.15 (6H, m). Spectral data match those previously reported.²⁵



(3-(trifluoromethyl)but-3-en-1-yn-1-yl)benzene (5d): Prepared by General Method A in 88.0% yield after flash silica gel chromatography (99:1 hexanes:EtOAc) using 2-bromo-3,3,3-trifluoroprop-1-ene (2.5 equiv) in place of vinyl bromide. ¹H NMR (500 MHz, CDCl₃) δ 7.46–7.35 (2H, m), 7.30–7.20 (3H, m), 6.00 (1H, s), 5.84 (1H, s). Spectral data match those previously reported.²⁶



(3-methylbut-3-en-1-yn-1-yl)cyclohexane (5e): Prepared by General Method A in 49.8% yield after flash silica gel chromatography (99:1 hexanes:EtOAc) using 2-bromopropene (2.5 equiv) in place of vinyl bromide. ¹H NMR (400 MHz, CDCl₃) δ 5.20–5.16 (1H, m), 5.13–5.09 (1H, m), 2.49–2.37 (1H, m), 1.85 (3H, s), 1.81–1.62 (4H, m), 1.56–1.36 (4H, m), 1.33–1.21 (2H, m). Spectral data match those previously reported.²⁷



(Z)-pent-3-en-1-yn-1-ylbenzene (7): Prepared by General Method A in 91.1% yield after flash silica gel chromatography (99:1 pentane:Et₂O) using (Z)-1-bromo-1-propene (2.5 equiv) in place of vinyl bromide. ¹H NMR (400 MHz, CDCl₃) δ 7.49–7.41 (2H, m), 7.35–7.27 (3H, m), 6.04 (1H, dq, J = 10.7, 6.8 Hz), 5.70 (1H, dq, J = 10.7, 1.6 Hz), 1.96 (3H, dd, J = 6.8, 1.6 Hz). Spectral data match those previously reported.²⁸

VI. Substrate Scope

<u>General Method B (non-enantioselective method)</u>: In an N₂-filled glovebox, to a 2-dram vial equipped with a magnetic stirring rod were added successively: **Pd-1** catalyst (8.6 mg, 0.01 mmol, 5 mol %), CH₂Cl₂ (0.25 mL), appropriate enyne (0.240 mmol, 1.20 equiv), and lastly appropriate amine nucleophile (0.200 mmol, 1.00 equiv). The reaction vials were then capped with a PTFE lined cap and allowed to stir at ambient temperature for the specified amount of time. The reaction contents were then diluted with 1:1 hexanes:EtOAc (1 mL) and passed through a short plug of either silica gel or neutral alumina, eluting with 1:1 hexanes:EtOAc (*ca.* 20 mL, in some cases 2% Et₃N was added to the eluent). The solution was then concentrated. The pure products were obtained by flash silica gel chromatography as described for each compound.

<u>General Method C (*in situ* non-enantioselective method)</u>: In an N₂-filled glovebox, to a 2-dram vial equipped with a magnetic stirring rod were added successively: (1.8 mg, 5.0 μ mol, 2.5 mol %), DPEphos (5.4 mg, 0.010 mmol, 5.0 mol %), and NaBAr₄^F (10.6 mg, 0.012 mmol, 6.0 mol %). This mixture was dissolved in CH₂Cl₂ (0.25 mL) and then appropriate enyne (0.240 mmol, 1.20 equiv) was added followed by appropriate amine nucleophile (0.200 mmol, 1.00 equiv). The reaction vials were then capped with a PTFE lined cap and were allowed to stir at ambient temperature for the specified amount of time. The reactions were worked up and purified as described for General Method B.

<u>General Method D (enantioselective method for enyne scope)</u>: In an N₂-filled glovebox, to a 2dram vial equipped with a magnetic stirring rod were added successively: $[Pd(\eta^3-allyl)Cl]_2$ (1.8 mg, 5.0 µmol, 2.5 mol %), **L3** (6.8 mg, 0.010 mmol, 5.0 mol %), and NaBAr₄^F (10.6 mg, 0.012 mmol, 6.0 mol %). The mixture was dissolved in CH₂Cl₂ (0.25 mL) and allowed to stir for *ca*. 5 min, resulting in a deep orange/red solution, and then appropriate enyne (0.240 mmol, 1.20 equiv) was added. The reaction vials were then capped with a PTFE lined cap, removed from the glovebox, and allowed to cool in an ice water bath at 0–4 °C for *ca*. 15 minutes before Et₃N (56.0 µL, 0.400 mmol, 2.00 equiv) and THIQ (25.0 µL, 0.200 mmol, 1.00 equiv) were added. The reactions were allowed to stir at 4 °C for 20 h and then worked up and purified as described for General Method B.

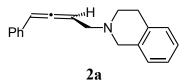
General Method E (enantioselecitve method for amine scope):

Catalyst stock solution: In an N₂-filled glovebox, to a 2-dram vial equipped with a magnetic stirring rod were added successively: $[Pd(\eta^3-allyl)Cl]_2$ (7.3 mg, 0.020 mmol, 1.0 mol %), L3 (27.0 mg, 0.040 mmol, 2.0 mol %), and NaBAr₄^F (44.3 mg, 0.050 mmol, 2.5 mol %). The mixture was dissolved in Et₂O (2.50 mL) and allowed to stir for *ca*. 5 min, resulting in a deep orange solution.

To a 2-dram vial equipped with a magnetic stirring rod was added 0.50 mL of the above catalyst stock solution and then enyne **2a** (65.0 μ L, 0.480 mmol, 1.20 equiv). The reaction vial was then capped with a PTFE lined cap, removed from the glovebox, and allowed to cool in an ice water bath at 0–4 °C for *ca.* 15 minutes before Et₃N (112 μ L, 0.800 mmol, 2.00 equiv) and appropriate amine nucleophile (0.400 mmol, 1.00 equiv) were added. The reaction mixtures were maintained at 4 °C for 3–5 h and then worked up and purified as described for General Method B.

Important Notes About Substrate Scope:

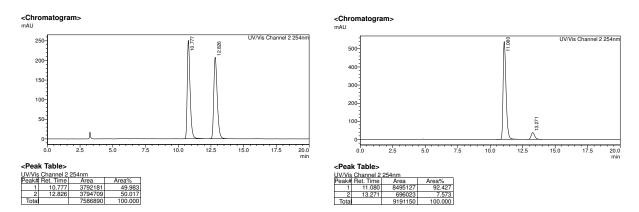
- The results for the substrate scope in the manuscript are an average of two runs, but for clarity, the results shown below are one of the two runs. In all cases the remainder of the mass balance is unreacted starting material and, unless otherwise noted, evidence of other competing side reactions was not observed.
- Some samples may contain ≤5% of the allylated starting amine (e.g. THIQ) that is generated upon pre-catalyst activation and which could not be separated from the desired product. Samples that contain this minor impurity are indicated with an asterisk (*) where the yields are reported.
- It was found early on that many of the products produced in this research exhibit only moderate stability if stored neat at ambient temperature. In most cases, products can be stored for up to 48 h at ambient temperature without detectable decomposition. It is advised that the allene products be stored in a -20 °C freezer, where they are generally stable for weeks. To avoid problems associated with decomposition and racemization, all products were purified immediately after workup and were analyzed by ¹H NMR and HPLC immediately after purification.



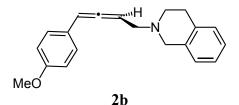
2-(4-phenylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2a): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2a** as a pale yellow oil (46.1 mg, 0.176 mmol, 88.2 % yield). **IR** (neat, cm⁻¹) 3026 (w), 2914 (w), 2787 (m), 2746 (w), 1947 (m), 1650 (w), 1597 (w), 1494 (m), 1456 (m), 1427 (w), 1360 (w), 1314 (w), 1277 (w), 1264 (w), 1232 (w), 1130 (m), 1089 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 7.36–7.30 (4H, m), 7.24–7.20 (1H, m), 7.15–7.09 (3H, m), 7.06–7.01 (1H, m), 6.24 (1H, dt, *J* = 6.4, 2.5 Hz), 5.71 (1H, ddd, *J* = 6.7, 6.7, 6.7 Hz), 3.79 (1H, AB_q, *J*_{AB} = 15.0 Hz), 3.73 (1H, AB_q, *J*_{AB} = 15.0 Hz), 3.36 (2H, dd, *J* = 7.2, 2.4 Hz), 2.98–2.91 (2H, m), 2.89–2.83 (2H, m); ¹³**C NMR** (125 MHz, CDCl₃) δ 206.3, 134.7, 134.3, 134.2, 128.7, 128.7, 127.0, 126.8, 126.6, 126.2, 125.6, 94.9, 91.7, 57.2, 55.6, 50.4, 29.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₁₉N: 262.1590, found: 262.1591.

<u>Scaled Up Reaction</u>: To an oven-dried 25-mL round-bottom flask equipped with a magnetic stirring rod was added [Pd(η^3 -allyl)Cl]₂ (18.3 mg, 0.0500 mmol, 1.00 mol %), DPEphos (53.9 mg, 0.100 mmol, 2.00 mol %), and NaBAr₄^F (102 mg, 0.115 mmol, 2.30 mol %). This mixture was dissolved in CH₂Cl₂ (6.25 mL) which was then allowed to stir an ambient temperature for *ca*. 5 min. To this solution was added enyne **1a** (811 μ L, 6.00 mmol, 1.20 equiv) and then 1,2,3,4-tetrahydroisoquinoline (634 μ L, 5.00 mmol, 1.00 equiv). The reaction mixture was allowed to stir at ambient temperature for 3 h and then concentrated. The concentrated material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to afford **2a** as a pale yellow oil (1.21 g, 4.61 mmol, 92.2% yield).

By General Method D: (*R*)-**2a** was isolated as a pale yellow oil (32.8 mg, 0.125 mmol, 62.7% yield). HPLC analysis indicated er = 92.5:7.5. $[\alpha]_D^{24} = -161.0$ (*c* = 1.0, CHCl₃) for a sample of 92.5:7.5 er.



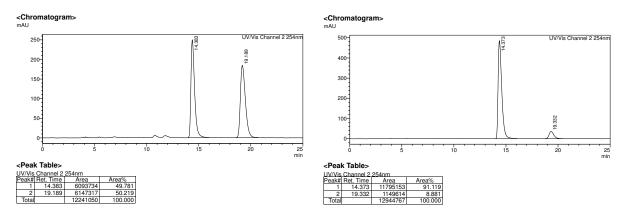
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 90:10 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 92.5:7.5.

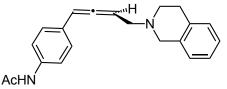


2-(4-(4-methoxyphenyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2b): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2b** as a yellow oil (49.8 mg, 0.171 mmol, 85.4% yield). **IR** (neat, cm⁻¹) 3003 (w), 2916 (m), 2832 (w), 2786 (m), 2748 (w), 1946 (w), 1650 (w), 1605 (s), 1580 (w), 1509 (s), 1462 (m), 1439 (m), 1385 (w), 1358 (w), 1336 (w), 1301 (m), 1282 (w), 1241 (s), 1169 (s), 1130 (s), 1082 (m), 1060 (w), 1032 (s); ¹**H NMR** (400 MHz, CDCl₃) δ 7.29–7.20 (2H, m), 7.17–7.08 (3H, m), 7.06–6.99 (1H, m), 6.90–6.84 (2H, m), 6.19 (1H, dt, J = 6.4, 2.3 Hz), 5.67 (1H, ddd, J = 6.8, 6.8, 6.8 Hz), 3.80 (3H, s), 3.77 (1H, AB_q, $J_{AB} = 14.8 \text{ Hz}$), 3.71 (1H, AB_q, $J_{AB} = 14.8 \text{ Hz}$), 3.34 (2H, dd, J = 7.2, 2.4 Hz), 2.98–2.89 (2H, m), 2.88–2.80 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 205.8, 158.8, 134.6, 134.2, 128.7, 127.9, 126.6, 126.5, 126.2, 125.6, 114.2, 94.3, 91.5, 57.5, 55.6, 55.3, 50.3, 29.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁NO: 292.1696, found: 292.1698.

<u>By General Method D:</u> (*R*)-**2b** was isolated as a pale yellow oil (37.0 mg, 0.127 mmol, 63.5% yield). HPLC analysis indicated er = 91:9. $[\alpha]_D^{25} = -116.3$ (*c* = 1.0, CHCl₃) for a sample of 91:9 er.

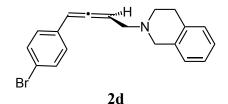
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 70:30 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 91:9.





2c (racemic)

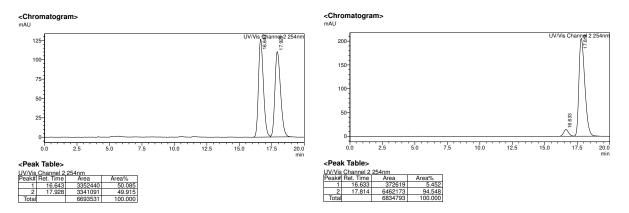
N-(4-(4-(3,4-dihydroisoquinolin-2(1*H*)-yl)buta-1,2-dien-1-yl)phenyl)acetamide (2c): Prepared by General Method B using Pd-1 at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield 2c as a yellow semisolid (45.5 mg, 0.143 mmol, 71.5% yield). **IR** (neat, cm⁻¹) 3257 (m), 3023 (m), 2918 (m), 2801 (m), 2244 (w), 1947 (w), 1664 (s), 1599 (s), 1532 (s), 1511 (s), 1427 (m), 1368 (m), 1313 (s), 1262 (m), 1195 (w), 1176 (w), 1129 (w), 1088 (w), 1036 (w), 1007 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.73 (1H, br s), 7.49–7.42 (2H, m), 7.26–7.19 (2H, m), 7.15–7.06 (3H, m) 7.04–6.98 (1H, m), 6.17 (1H, dt, J = 6.4, 2.5 Hz), 5.67 (1H, ddd, J = 6.7, 6.7, 6.7 Hz), 3.74 (1H, AB_q, $J_{AB} = 15.2$ Hz), 3.70 (1H, AB_q, $J_{AB} = 15.2$ Hz), 3.32 (2H, dd, J = 7.1, 2.1 Hz), 2.96–2.87 (2H, m), 2.86–2.78 (2H, m), 2.13 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 206.2, 168.5, 136.9, 134.6, 134.1, 130.2, 128.7, 127.3, 126.6, 126.2, 125.7, 120.2, 94.4, 91.7, 57.2, 55.6, 50.3, 29.1, 24.6; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₁H₂₂N₂O: 319.1805, found: 319.1807.

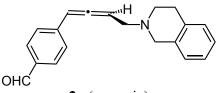


2-(4-(4-bromophenyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2d): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2d** as a yellow solid (59.4 mg, 0.175 mmol, 87.3% yield). **IR** (neat, cm⁻¹) 3019 (w), 2924 (m), 2789 (m), 2743 (m), 1946 (m), 1907 (w), 1583 (w), 1485 (s), 1452 (m), 1426 (m), 1377 (w), 1359 (m), 1330 (m), 1308 (m), 1226 (w), 1192 (w), 1129 (m), 1089 (s), 1037 (s), 1007 (s), 932 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 7.46–7.40 (2H, m), 7.20–7.14 (2H, m), 7.14–7.07 (3H, m), 7.04–6.99 (1H, m), 6.16 (1H, dt, *J* = 6.4, 2.4 Hz), 5.68 (1H, ddd, *J* 6.9, 6.9, 6.9 Hz), 3.74 (1H, AB_q, *J*_{AB} = 15.8 Hz), 3.70 (1H, AB_q, *J*_{AB} = 15.8 Hz), 3.38–3.29 (2H, m), 2.97–2.89 (2H, m), 2.85–2.79 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 206.3, 134.6, 134.1, 133.4, 131.8, 128.8, 128.3, 126.6, 126.2, 125.7, 120.6, 94.1, 92.2, 57.0, 55.6, 50.4, 29.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₁₈NBr: 340.0695, found: 340.0704; **MP** = 66–69 °C.

By General Method D: (*R*)-2d was isolated as a yellow solid (35.3 mg, 0.104 mmol, 51.9% yield). HPLC analysis indicated er = 94.5:5.5. $[\alpha]_{D}^{27} = -182.3$ (*c* = 1.0, CHCl₃) for a sample of 94.5:5.5 er.

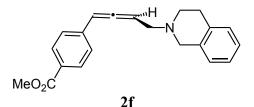
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 80:20 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 94.5:5.5.





2e (racemic)

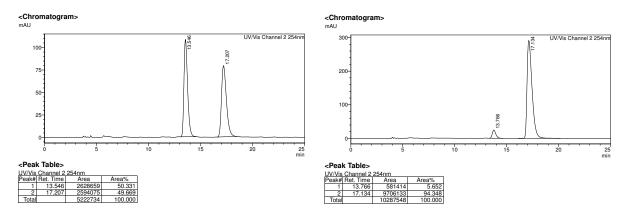
4-(4-(3,4-dihydroisoquinolin-2(1*H***)-yl)buta-1,2-dien-1-yl)benzaldehyde (2e):** Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (70:30 hexanes:EtOAc) to yield **2e** as a yellow oil (38.9 mg, 0.134 mmol, 67.2% yield). **IR** (neat, cm⁻¹) 3021 (w), 2917 (w), 2801 (m), 2744 (w), 1945 (m), 1690 (s), 1600 (s), 1570 (w), 1497 (w), 1428 (w), 1384 (w), 1334 (w), 1303 (m), 1210 (m), 1163 (m), 1130 (w), 1091 (m), 1056 (w), 1038 (w), 1013 (w); ¹H **NMR** (400 MHz, CDCl₃) δ 9.96 (1H, s), 7.86–7.79 (2H, m), 7.48–7.41 (2H, m), 7.17–7.06 (3H, m), 7.04–6.98 (1H, m), 6.28 (1H, dt, *J* = 6.4, 2.4 Hz), 5.77 (1H, ddd, *J* = 6.7, 6.7, 6.7 Hz), 3.75 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.73 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.43–3.33 (2H, m), 2.97–2.89 (2H, m), 2.87–2.79 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 207.6, 191.6, 141.1, 135.0, 134.4, 134.0, 130.1, 128.7, 127.2, 126.6, 126.3, 125.7, 94.6, 92.2, 56.7, 55.5, 50.3, 29.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₁₉NO: 290.1539, found: 290.1540.

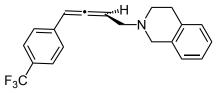


methyl 4-(4-(3,4-dihydroisoquinolin-2(1*H***)-yl)buta-1,2-dien-1-yl)benzoate (2f):** Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (70:30 hexanes:EtOAc) to yield **2f** as a pale yellow oil (55.0 mg, 0.172 mmol, 86.1 % yield). **IR** (neat, cm⁻¹) 3026 (w), 2912 (w), 2806 (w), 1952 (w), 1713 (s), 1605 (m), 1497 (w), 1435 (m), 1362 (w), 1331 (w), 1276 (s), 1235 (w), 1192 (w), 1177 (m), 1109 (m), 1091 (m), 1014 (w), 1039 (w); ¹H **NMR** (400 MHz, CDCl₃) δ 8.02–7.96 (2H, m), 7.39–7.32 (2H, m), 7.17–7.05 (3H, m), 7.04–6.98 (1H, m), 6.25 (1H, dt, J = 6.4, 2.3 Hz), 5.74 (1H, ddd, J = 6.7, 6.7, 6.7 Hz), 3.90 (3H, s), 3.76 (1H, AB_q, $J_{AB} = 14.9$ Hz), 3.70 (1H, AB_q, $J_{AB} = 14.9$ Hz), 3.36 (2H, dt, J = 7.1, 2.0 Hz), 2.97–2.88 (2H, m), 2.87–2.77 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 207.2, 166.9, 139.4, 134.5, 134.0, 130.0, 128.7, 128.5, 126.6, 126.2, 125.7, 94.5, 92.1, 56.8, 55.5, 52.1, 50.3, 29.1 (note: one sp² carbon could not be identified due to overlap); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₁H₂₁NO₂: 320.1645, found: 320.1645.

<u>By General Method D:</u> (*R*)-**2f** was isolated as a pale yellow oil (38.1 mg, 0.119 mmol, 59.6% yield). HPLC analysis standard indicated er = 95.5:4.5. $[\alpha]_D^{24} = -158.7$ (*c* = 1.0, CHCl₃) for a sample of 94.5:5.5 er.

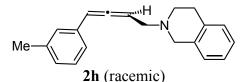
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 70:30 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 94.5:5.5.



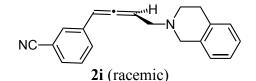


2g (racemic)

2-(4-(4-(trifluoromethyl)phenyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2g): Prepared by General Method B using Pd-1 at 22 °C for 3 h. The material was purified by flash silica gel chromatography (80:20 hexanes:EtOAc) to yield 2g as a yellow solid (58.7 mg, 0.178 mmol, 89.1 % yield). IR (neat, cm⁻¹) 3023 (w), 2925 (w), 2814 (w), 1951 (m), 1611 (m), 1581 (w), 1497 (w), 1464 (w), 1450 (w), 1438 (w), 1359 (w), 1322 (s), 1157 (s), 1113 (s), 1087 (m), 1063 (s), 1028 (w), 1014 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.67 (2H, d *J* = 8.2 Hz), 7.40 (2H, d, *J* = 8.2 Hz), 7.18–7.08 (3H, m), 7.06–6.99 (1H, m), 6.26 (1H, dt, *J* = 6.4, 2.3 Hz), 5.76 (1H, ddd, *J* = 6.8, 6.8, 6.8 Hz), 3.76 (1H, AB_q, *J*_{AB} = 15.7 Hz), 3.73 (1H, AB_q, *J*_{AB} = 15.7 Hz), 3.41–3.32 (2H, m), 2.99–2.90 (2H, m), 2.88–2.81 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 207.0, 138.3, 134.4, 134.0, 128.9 (q, *J*_{CF} = 32.0 Hz), 128.7, 126.9, 126.6 (q, *J*_{CF} = 3.2 Hz), 125.7, 125.6, 125.6, 124.2 (q, *J*_{CF} = 270.2 Hz), 94.1, 92.2, 56.8, 55.6, 50.3, 29.1; ¹⁹F NMR (376 MHz, CDCl₃) δ –62.4; HRMS (ESI⁺) [M+H]⁺ calc'd for C₂₀H₁₈F₃N: 330.1464, found: 330.1460; MP = 73–75 °C.

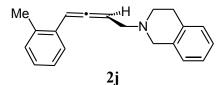


2-(4-(*m***-tolyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2h)**: Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2h** as a pale yellow oil (48.2 mg, 0.175 mmol, 87.5% yield). **IR** (neat, cm⁻¹) 3021 (w), 2915 (m), 2792 (m), 2746 (w), 1947 (m), 1584 (m), 1603 (w), 1488 (m), 1453 (m), 1380 (w), 1359 (w), 1332 (m), 1306 (w), 1276 (w), 1232 (w), 1130 (m), 1091 (m), 1056 (w), 1038 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.26–7.20 (1H, m), 7.18–7.10 (5H, m), 7.07–7.02 (2H, m), 6.21 (1H, dt, *J* = 6.4, 2.3 Hz), 5.70 (1H, ddd, *J* = 7.0, 7.0, 7.0 Hz), 3.80 (1H, ABq, *J*_{AB} = 14.8 Hz), 3.73 (1H, ABq, *J*_{AB} = 14.8 Hz), 3.36 (2H, dd, *J* = 7.2, 2.4 Hz), 2.99–2.92 (2H, m), 2.90–2.83 (2H, m), 2.36 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 206.4, 138.2, 134.6, 134.2, 134.1, 128.7, 128.6, 127.8, 127.5, 126.6, 126.2, 125.7, 124.0, 94.8, 91.4, 57.2, 55.5, 50.3, 29.1, 21.4; HRMS (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁N: 276.1747, found: 276.1749.



3-(4-(3,4-dihydroisoquinolin-2(1*H***)-yl)buta-1,2-dien-1-yl)benzonitrile (2i):** Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (65:35 hexanes:EtOAc) to yield **2i** as a pale yellow oil (53.8 mg, 0.188 mmol, 93.9% yield).* **IR** (neat, cm⁻¹) 3021 (w), 2917 (m), 2796 (m), 2229 (m), 1948 (m), 1597 (w), 1578

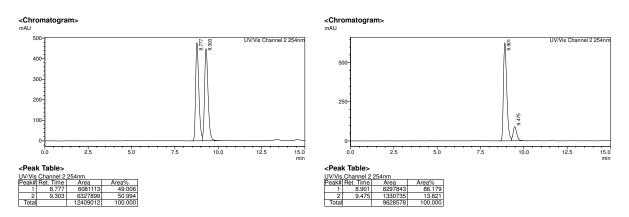
(w), 1497 (w), 1481 (m), 1463 (m), 1453 (m), 1434 (m), 1381 (w), 1359 (w), 1332 (m), 1305 (w), 1277 (w), 1226 (w), 1192 (w), 1130 (m), 1092 (m), 1056 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.57 (1H, s), 7.53–7.44 (2H, m), 7.40 (1H, dd *J* = 7.7, 7.7 Hz), 7.18–7.06 (3H, m), 7.05–6.99 (1H, m), 6.20 (1H, dt, *J* = 6.4, 2.4 Hz), 5.77 (1H, ddd, *J* = 6.9, 6.9, 6.9 Hz), 3.75 (1H, AB_q, *J*_{AB} = 15.2 Hz), 3.71 (1H, AB_q, *J*_{AB} =15.2 Hz), 3.42–3.30 (2H, m), 2.97–2.88 (2H, m), 2.86–2.78 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 206.6, 136.0, 134.4, 134.0, 130.9, 130.3, 130.1, 129.4, 128.7, 126.6, 126.2, 125.7, 118.7, 112.8, 93.5, 92.9, 56.8, 55.6, 50.4, 29.1; HRMS (ESI⁺) [M+H]⁺ calc'd for C₂₀H₁₈N₂: 287.1543, found: 287.1544.

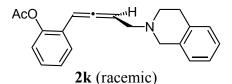


2-(4-(*o***-tolyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2j):** Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2j** as a pale yellow oil (43.0 mg, 0.156 mmol, 78.1% yield). **IR** (neat, cm⁻¹) 3021 (w), 2915 (m), 2786 (m), 2746 (w), 1945 (m), 1655 (w), 1600 (w), 1491 (m), 1462 (m), 1381 (w), 1358 (w), 1334 (m), 1302 (w), 1276 (w), 1192 (w), 1127 (m), 1095 (m), 1037 (w); ¹**H NMR** (400 MHz, CDCl₃) δ 7.41 (1H, d, J = 7.6 Hz), 7.22–7.09 (6H, m), 7.06–7.01 (1H, m), 6.43 (1H, dt, J = 6.4, 2.4 Hz), 5.67 (1H, ddd, J = 7.0, 7.0, 7.0 Hz), 3.77 (1H, AB_q, J_{AB} = 15.0 Hz), 3.72 (1H, AB_q, J_{AB} = 15.0 Hz), 3.36 (2H, dd, J = 7.1, 2.4 Hz), 2.98–2.91 (2H, m), 2.88–2.82 (2H, m), 2.38 (3H, s); ¹³**C NMR** (125 MHz, CDCl₃) δ 207.0, 135.0, 134.6, 134.1, 132.5, 130.5, 128.7, 127.3, 126.9, 126.6, 126.2, 126.1, 125.7, 92.1, 90.8, 57.3, 55.6, 50.4, 29.1, 19.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁N: 276.1747, found: 276.1752.

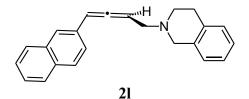
<u>By General Method D:</u> (*R*)-2j was isolated as a pale yellow oil (32.7 mg, 0.119 mmol, 59.4% yield). HPLC analysis indicated er = 86:14. $[\alpha]_D^{27} = -102.2$ (*c* = 1.0, CHCl₃) for a sample of 86:14 er.

HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 90:10 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 86:14.





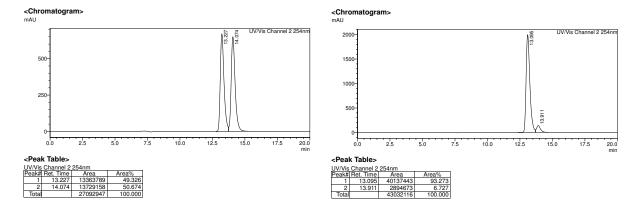
2-(4-(3,4-dihydroisoquinolin-2(1*H***)-yl)buta-1,2-dien-1-yl)phenyl acetate (2k):** Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2k** as a pale yellow oil (42.9 mg, 0.134 mmol, 67.2% yield). **IR** (neat, cm⁻¹) 3021 (w), 2917 (m), 2789 (w), 1948 (m), 1762 (s), 1649 (w), 1604 (w), 1580 (w), 1490 (m), 1453 (w), 1367 (m), 1334 (w), 1307 (w), 1277 (w), 1194 (s), 1171 (s), 1131 (m), 1091 (s), 1038 (w), 1008 (m); ¹H **NMR** (400 MHz, CDCl₃) δ 7.50–7.42 (1H, m), 7.29–7.18 (2H, m), 7.16–6.98 (5H, m), 6.28 (1H, dt, *J* = 6.5, 2.3 Hz), 5.70 (1H, ddd, *J* = 7.0, 7.0, 7.0 Hz), 3.75 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.71 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.40–3.29 (2H, m), 2.96–2.89 (2H, m), 2.87–2.78 (2H, m), 2.33 (3H, s); ¹³C **NMR** (125 MHz, CDCl₃) δ 207.0, 169.2, 147.5, 134.6, 134.1, 128.7, 128.3, 127.9, 126.6, 126.5, 126.2, 125.6, 122.8, 91.5, 88.7, 57.0, 55.6, 50.4, 29.1, 20.9 (one sp² carbon could not be identified due to overlap); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₁H₂₁NO₂: 320.1645, found: 320.1649.

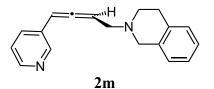


2-(4-(naphthalen-2-yl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2l): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2l** as a white solid (56.1 mg, 0.180 mmol, 90.1% yield). **IR** (neat, cm⁻¹) 2906 (w), 2808 (w), 1940 (w), 1596 (w), 1496 (w), 1448 (w), 1380 (w), 1356 (w), 1322 (w), 1270 (w), 1126 (w), 1082 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.89–7.73 (3H, m), 7.69 (1H, s), 7.58–7.39 (3H, m), 7.21–7.10 (3H, m), 7.10–6.98 (1H, m), 6.43 (1H, dt, *J* = 6.3, 2.6 Hz), 7.79 (1H, ddd, *J* = 6.9, 6.9, 6.9 Hz), 3.82 (1H, AB_q, *J*_{AB} = 14.8 Hz), 3.76 (1H, AB_q, *J*_{AB} = 14.8 Hz), 3.41 (2H, dt, *J* = 7.1, 2.1 Hz), 3.01–2.93 (2H, m), 2.92–2.84 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 206.9, 134.7, 134.2, 133.7, 132.7, 131.9, 128.8, 128.3, 127.8, 127.7, 126.7, 126.3, 126.2, 125.7, 125.7, 125.7, 124.7, 95.3, 92.0, 57.3, 55.7, 50.4, 29.2; HRMS (ESI⁺) [M+H]⁺ calc'd for C₂₃H₂₁N: 312.1747, found: 312.1744; **MP** = 101–104 °C.

By General Method D: (*R*)-21 was isolated as a white solid (40.0 mg, 0.128 mmol, 64.2% yield). HPLC analysis indicated er = 93.5:6.5. $[\alpha]_D^{26} = -198.8$ (*c* = 1.0, CHCl₃) for a sample of 93.5:6.5 er.

HPLC: Column: Chiralpak 1A-3 (3 μ m, 4.6 mm X 150 mm). Mobile phase: 98:2 *i*-PrOH:MeCN with 0.1% Et₂NH additive, 0.25 mL/min. Detection wavelength: 254 nm. Er = 93.5:6.5.

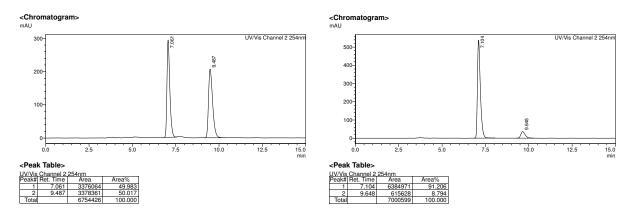


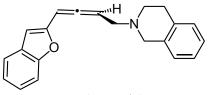


2-(4-(pyridin-3-yl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (**2m**): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (70:30 hexanes:EtOAc with an additional 2% Et₃N additive) to yield **2m** as a pale yellow oil (42.3 mg, 0.161 mmol, 80.6% yield).* **IR** (neat, cm⁻¹) 3022 (w), 2917 (m), 2796 (m), 1948 (m), 1643 (w), 1584 (w), 1570 (m), 1497 (w), 1480 (m), 1445 (m), 1427 (m), 1383 (w), 1325 (m), 1232 (w), 1180 (w), 1129 (m), 1093 (m), 1056 (w), 1024 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 8.52 (1H, d, J = 2.1 Hz), 8.42 (1H, dd, J = 4.8, 1.6 Hz), 7.58 (1H, ddd, J = 7.9, 1.8, 1.8 Hz), 7.25–7.19 (1H, m), 7.14–7.06 (3H, m), 7.03–6.98 (1H, m), 6.19 (1H, dt, J = 6.4, 2.4 Hz), 5.73 (1H, ddd, J = 6.9, 6.9, 6.9 Hz), 3.74 (1H, AB_q, $J_{AB} = 15.2$ Hz), 3.70 (1H, AB_q, $J_{AB} = 15.2$ Hz), 3.55 (2H, dd, J = 7.1, 2.4 Hz), 2.95–2.87 (2H, m), 2.86–2.79 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 206.5, 148.2, 148.0, 134.4, 134.0, 133.5, 130.3, 128.7, 126.6, 126.2, 125.7, 123.5, 92.4, 91.7, 56.8, 55.6, 50.3, 29.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₈H₁₈N₂: 263.1543, found: 263.1545.

<u>By General Method D:</u> (*R*)-2m was isolated as a pale yellow oil (16.0 mg, 0.0610 mmol, 30.5% yield). HPLC analysis indicated er = 91:9. $[\alpha]_D^{26} = -120.9$ (*c* = 1.0, CHCl₃) for a sample of 91:9 er.

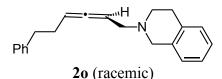
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 70:30 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 91:9.



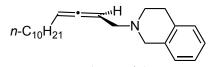


2n (racemic)

2-(4-(benzofuran-2-yl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2n): Prepared by General Method B using **Pd-1** at 22 °C for 15 h. The material was purified by flash silica gel chromatography (75:25 hexanes:EtOAc) to yield **2n** as a pale yellow oil (38.9 mg, 0.129 mmol, 64.5% yield). **IR** (neat, cm⁻¹) 3021 (w), 2916 (m), 2796 (m), 1950 (w), 1679 (w), 1582 (w), 1497 (w), 1453 (m), 1382 (w), 1361 (w), 1279 (w), 1253 (m), 1220 (w), 1164 (w), 1130 (m), 1090 (m), 1007 (w); ¹**H NMR** (400 MHz, CDCl₃) δ 7.53–7.47 (1H, m), 7.44 (1H, d, J = 8.0 Hz), 7.29–7.17 (2H, m), 7.16–7.08 (3H, m), 7.07–6.99 (1H, m), 6.59 (1H, s), 6.30 (1H, dt, J = 6.4, 2.3 Hz), 5.83 (1H, ddd, J = 6.9, 6.9, 6.9 Hz), 3.86 (1H, AB_q, J_{AB} = 14.9 Hz), 3.74 (1H, AB_q, J_{AB} = 14.9 Hz), 3.39 (2H, dd, J = 7.2, 2.4 Hz), 2.99–2.92 (2H, m), 2.92–2.86 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 207.1, 155.0, 150.8, 134.7, 134.2, 129.1, 128.8, 126.7, 126.2, 125.7, 124.1, 122.8, 120.5, 110.9, 104.0, 92.5, 85.9, 57.0, 55.6, 50.4, 29.2; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₁H₁₉NO: 302.1539, found: 302.1534.



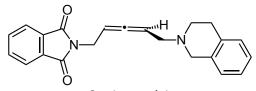
2-(6-phenylhexa-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (20): Prepared by General Method C at 22 °C for 3 h. The material was purified by flash silica gel chromatography (80:20 hexanes:EtOAc) to yield **20** as a pale yellow oil (50.3 mg, 0.174 mmol, 86.9% yield). **IR** (neat, cm⁻¹) 3023 (w), 2916 (m), 2789 (m), 2747 (w), 1961 (m), 1651 (m), 1603 (w), 1583 (w), 1496 (m), 1452 (m), 1385 (w), 1360 (w), 1333 (m), 1232 (w), 1192 (w), 1130 (m), 1091 (m), 1029 (w); **¹H NMR** (400 MHz, CDCl₃) δ 7.34–7.27 (2H, m), 7.24–7.17 (3H, m), 7.16–7.08 (3H, m), 7.05–7.00 (1H, m), 5.27–5.20 (2H, m), 3.69 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.64 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.18–3.12 (2H, m), 2.92 (2H, app. t, *J* = 5.9 Hz), 2.82–2.71 (4H, m), 2.42–2.31 (2H, m); **¹³C NMR** (125 MHz, CDCl₃) δ 205.5, 141.7, 134.7, 134.2, 128.7, 128.6, 128.3, 126.6, 126.1, 125.9, 125.6, 90.5, 88.0, 57.6, 55.5, 50.2, 35.5, 30.5, 29.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₁H₂₃N: 290.1903, found: 290.1902.



2p (racemic)

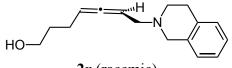
2-(tetradeca-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2p): Prepared by General Method C at 22 °C for 3 h. The material was purified by flash silica gel chromatography (85:15 hexanes:EtOAc) to yield **2p** as a pale yellow oil (53.0 mg, 0.163 mmol, 81.4% yield). **IR** (neat, cm⁻¹) 2921 (s), 2851 (s), 2791 (m), 2747 (w), 1961 (w), 1658 (w), 1585 (w), 1498 (w), 1454 (m), 1397 (w), 1360 (w), 1333 (m), 1303 (w), 1232 (w), 1192 (w), 1193 (w), 1092 (m); ¹**H NMR** (400

MHz, CDCl₃) δ 7.16–7.06 (3H, m), 7.05–6.99 (1H, m), 5.25–5.12 (2H, m), 3.71 (1H, AB_q, J_{AB} = 15.1 Hz), 3.66 (1H, AB_q, J_{AB} = 15.1 Hz), 3.19 (2H, dt, J = 6.9, 2.6 Hz), 2.94–2.89 (2H, m), 2.83–2.76 (2H, m), 2.07–1.95 (2H, m), 1.471.15 (16H, m), 0.872 (3H, t, J = 7.0 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.3, 134.7, 134.2, 128.7, 126.6, 126.1, 125.6, 91.3, 87.5, 57.8, 55.5, 50.2, 31.9, 29.7, 29.5, 29.4, 29.2, 29.1, 29.0, 28.7, 22.7, 14.1 (note: one sp³ carbon could not be identified due to overlap); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₃H₃₅N: 326.2842, found: 326.2841.



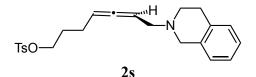
2q (racemic)

2-(5-(3,4-dihydroisoquinolin-2(1*H***)-yl)penta-2,3-dien-1-yl)isoindoline-1,3-dione (2q):** Prepared by General Method C at 22 °C for 3 h. The material was purified by flash silica gel chromatography (50:50 hexanes:EtOAc) to yield **2q** as a yellow oil (54.4 mg, 0.158 mmol, 79.0% yield).* **IR** (neat, cm⁻¹) 2921 (w), 2792 (w), 1966 (w), 1771 (m), 1709 (s), 1613 (w), 1498 (w), 1466 (w), 1425 (m), 1388 (s), 1315 (s), 1189 (w), 1170 (w), 1129 (w), 1109 (m), 1088 (m), 1007 (w); ¹**H NMR** (400 MHz, CDCl₃) δ 7.85–7.80 (2H, m), 7.71–7.66 (2H, m), 7.13–6.98 (3H, m), 6.92–6.84 (1H, m), 5.37–5.26 (2H, m), 4.34–4.23 (2H, m), 3.56 (1H, AB_q, *J*_{AB} = 15.5 Hz), 3.52 (1H, AB_q, *J*_{AB} = 15.5 Hz), 3.23–3.14 (1H, m), 3.12–3.02 (1H, m), 2.80–2.70 (2H, m), 2.61 (2H, app. t, *J* = 5.9 Hz); ¹³C **NMR** (125 MHz, CDCl₃) δ 204.8, 167.7, 134.5, 134.0, 134.0, 132.1, 128.6, 126.6, 126.1, 125.5, 123.3, 91.2, 87.2, 56.9, 55.4, 50.1, 36.6, 29.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₂H₂₀N₂O₂: 345.1598, found: 345.1600.

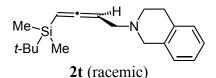


2r (racemic)

7-(3,4-dihydroisoquinolin-2(1*H***)-yl)hepta-4,5-dien-1-ol (2r):** Prepared by General Method C at 22 °C for 3 h. The material was purified by flash silica gel chromatography (50:50 hexanes:EtOAc to 100% EtOAc) to yield **2r** as a colorless oil (28.5 mg, 0.117 mmol, 58.5% yield). **IR** (neat, cm⁻¹) 3283 (br m), 2917 (m), 2794 (m), 1960 (m), 1584 (w), 1498 (w), 1451 (m), 1385 (w), 1360 (w), 1333 (m), 1304 (m), 1233 (w), 1192 (w), 1130 (m), 1057 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.13–7.06 (3H, m), 7.03–6.98 (1H, m), 5.26–5.13 (2H, m), 3.67 (2H, s), 3.61 (2H, t, *J* = 6.4 Hz), 3.21–3.11 (2H, m), 2.94–2.86 (2H, m), 2.81–2.73 (2H, m), 2.31 (1H, br s), 2.15–2.02 (2H, m), 1.72–1.57 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.4, 134.5, 134.1, 128.7, 126.6, 126.2, 125.6, 90.8, 87.9, 61.8, 57.5, 55.6, 50.4, 31.7, 28.9, 25.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₁NO: 244.1696, found: 244.1694.

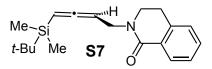


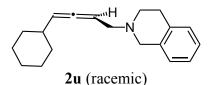
7-(3,4-dihydroisoquinolin-2(1*H***)-yl)hepta-4,5-dien-1-yl 4-methylbenzenesulfonate (2s):** Prepared by General Method C at 22 °C for 3 h. The material was purified by flash silica gel chromatography (50:50 hexanes:EtOAc) to yield **2s** as a pale yellow oil (59.4 mg, 0.149 mmol, 74.7% yield). **IR** (neat, cm⁻¹) 2918 (w), 2791 (w), 1962 (w), 1650 (w), 1597 (w), 1496 (w), 1449 (m), 1356 (s), 1306 (w), 1232 (w), 1188 (m), 1173 (s), 1095 (m), 1033 (w), 1009 (m), 927 (s); ¹**H NMR** (400 MHz, CDCl₃) δ 7.79–7.74 (2H, m), 7.31 (2H, d, *J* = 8.3 Hz), 7.13–7.04 (3H, m), 7.02–6.97 (1H, m), 5.26–5.15 (1H, m), 5.13–5.00 (1H, m), 4.06 (2H, t, *J* = 6.4 Hz), 3.65 (2H, s), 3.15 (2H, dd, *J* = 7.1, 2.4 Hz), 2.92–2.85 (2H, m), 2.79–2.71 (2H, m), 2.41 (3H, s), 2.10–2.00 (2H, m), 1.82–1.70 (2H, m); ¹³**C NMR** (125 MHz, CDCl₃) δ 205.3, 144.7, 134.4, 134.0, 133.1, 129.8, 128.7, 127.9, 126.6, 126.2, 125.6, 89.7, 88.5, 69.7, 57.5, 55.5, 50.2, 28.9, 28.1, 24.3, 21.6; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₃H₂₇NO₃S: 398.1784, found: 398.1783.



(*S*)-2-(4-(*tert*-butyldimethylsilyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2t): Prepared by General Method C at 22 °C for 15 h. The material was purified by flash silica gel chromatography (100% hexanes to 85:15 hexanes:EtOAc) to afford 2t (60.6 mg, 0.202 mmol, 50.6% yield) as a yellow oil. **IR** (neat, cm⁻¹) 2950 (m), 2926 (m), 2855 (m), 2803 (w), 1937 (s), 1657 (w), 1498 (w), 1462 (m), 1376 (w), 1360 (m), 1323 (m), 1190 (w), 1129 (w), 1096 (m), 1056 (w), 936 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.16–7.06 (3H, m), 7.04–6.99 (1H, m), 4.97 (1H, dt, J = 6.8, 2.9, Hz), 4.90 (1H, ddd, J = 7.2, 7.2, 7.2 Hz), 3.73 (1H, AB_q, $J_{AB} = 15.0 Hz$), 3.66 (1H, AB_q, $J_{AB} = 15.0 Hz$), 3.28–3.17 (2H, m), 2.95–2.87 (2H, m), 2.83–2.74 (2H, m), 0.903 (9H, s), 0.07 (6H, s); ¹³C NMR (125 MHz, CDCl₃) δ 211.1, 134.5, 134.1, 128.7, 126.6, 126.2, 125.7, 79.8, 79.6, 57.2, 55.3, 50.1, 29.0, 26.3, 17.0, –5.56, –5.65; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₂₉NSi: 300.2142, found: 300.2143.

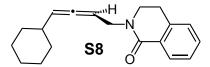
The formation of 2t was accompanied by the formation of *ca*. 5% of an oxidized side product that could not be separated and which we propose is S7. We have not fully characterized this minor side product and so cannot assign its structure with absolute certainty.

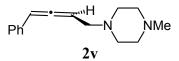




2-(4-cyclohexylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (2u): Prepared by General Method C at 22 °C for 15 h. The material was purified by flash silica gel chromatography (85:15 hexanes:EtOAc) to afford **2u** (84.6 mg, 0.316 mmol, 79.1% yield) as a yellow oil. **IR** (neat, cm⁻¹) 2920 (s), 2848 (s), 2789 (m), 2746 (w), 1959 (w), 1656 (m), 1605 (w), 1497 (w), 1479 (w), 1447 (s), 1385 (w), 1359 (w), 1329 (m), 1304 (m), 1231 (w), 1192 (w), 1130 (m), 1092 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 7.15–7.05 (3H, m), 7.04–6.97 (1H, m), 5.23 (1H, ddd, *J* = 13.5, 7.0, 3.0 Hz), 5.19–5.13 (1H, m), 3.71 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.66 (1H, AB_q, *J*_{AB} = 15.1 Hz), 3.24–3.14 (2H, m), 2.95–2.87 (2H, m), 2.83–2.75 (2H, m), 2.07–1.93 (1H, m), 1.83–1.55 (5H, m), 1.36–1.01 (5H, m); ¹³C NMR (125 MHz, CDCl₃) δ 204.2, 134.7, 134.2, 128.7, 126.6, 126.1, 125.6, 97.3, 88.5, 58.0, 55.6, 50.2, 37.2, 33.2, 33.1, 29.0, 26.2, 26.0 (note: one extra sp³ carbon was observed, but this has been noted before with related cyclohexyl-substituted compounds^{7,29}); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₂₅N: 268.2060, found: 268.2063.

The formation of 2u was accompanied by the formation of *ca*. 5% of an oxidized side product that could not be separated and which we propose is **S8**. We have not fully characterized this minor side product and so cannot assign its structure with absolute certainty.

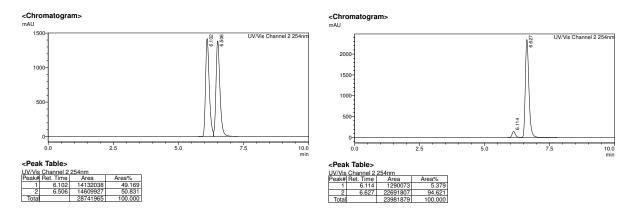


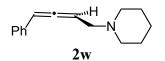


1-methyl-4-(4-phenylbuta-2,3-dien-1-yl)piperazine (2v): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 CH₂Cl₂:MeOH with 5% Et₃N) to yield **2v** as a pale yellow oil (41.4 mg, 0.144 mmol, 71.8% yield). **IR** (neat, cm⁻¹) 2933 (w), 2791 (m), 1949 (w), 1454 (s), 1330 (s), 1280 (s), 1161 (s), 1141 (m), 1009 (s), 690 (s), (625 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.32–7.22 (4H, m), 7.21–7.15 (1H, m), 6.16 (1H, dt, J = 6.4, 2.4 Hz), 5.59 (1H, ddd, J = 7.1, 7.1, 7.1 Hz), 3.18 (2H, dd, J = 7.2, 2.4 Hz), 2.82–2.26 (8H, m), 2.25 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 206.2, 134.2, 128.6, 126.9, 126.8, 94.8, 91.3, 57.4, 55.0, 52.4, 45.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₅H₂₀N₂: 229.1699, found: 229.1704.

<u>By General Method E:</u> (*R*)-2v was isolated as a pale yellow was (65.4 mg, 0.286 mmol, 71.6% yield). HPLC analysis indicated er = 94.5:5.5. $[\alpha]_D^{26} = -204.5$ (*c* = 1.0, CHCl₃) for a sample of 94.5:5.5 er

HPLC: Column: Chiralpak 1A-3 (3 μ m, 4.6 mm X 150 mm). Mobile phase: 60:40 MeOH:*i*-PrOH with 0.1% Et₂NH additive, 0.4 mL/min. Detection wavelength: 254 nm. Er = 94.5:5.5.

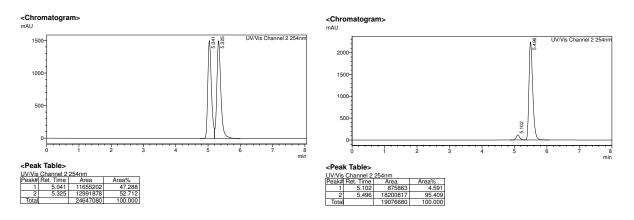


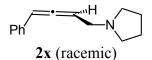


1-(4-phenylbuta-2,3-dien-1-yl)piperidine (2w): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (85:15 hexanes:EtOAc with 5% Et₃N) to yield **2w** as a pale yellow oil (35.6 mg, 0.167 mmol, 83.4% yield). **IR** (neat, cm⁻¹) 2933 (s), 2788 (w), 1949 (m), 1457 (w), 1338 (w), 1109 (m), 789 (s), 769 (s), 690 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.23 (4H, m), 7.21–7.14 (1H, m), 6.14 (1H, dt, J = 6.2, 2.3 Hz), 5.61 (1H, ddd, J = 7.1, 7.1, 7.1 Hz), 3.15–3.13 (2H, m) 2.59–2.40 (4H, m), 1.68–1.56 (4H, m), 1.49–1.38 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 206.1, 134.5, 128.6, 126.9, 126.8, 94.5, 91.7, 58.3, 54.1, 26.1, 24.3; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₅H₁₉N: 214.1590, found: 214.1591.

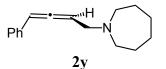
By General Method E: (*R*)-2w was isolated as a pale yellow oil (59.8 mg, 0.280 mmol, 70.1% yield). HPLC analysis indicated er = 95.5:4.5. $[\alpha]_D^{25} = -238.9$ (*c* = 1.0, CHCl₃) for a sample of 95.5:4.5 er.

HPLC: Chiralpak 1A-3 (3 μ m, 4.6 mm X 150 mm). Mobile phase: 100% MeOH with 0.1% Et₂NH additive, 0.5 mL/min. Detection wavelength: 254 nm. Er = 95.5:4.5.





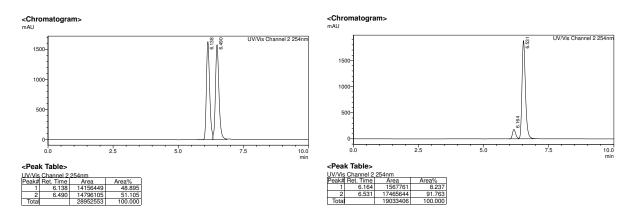
1-(4-phenylbuta-2,3-dien-1-yl)pyrrolidine (2x): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (80:20 hexanes:EtOAc with 5% Et₃N) to yield **2x** as a pale yellow oil (28.9 mg, 0.145 mmol, 72.5% yield). **IR** (neat, cm⁻¹) 2962 (m), 2783 (m), 1949 (w), 1494 (w), 1458 (m), 1365 (m), 1344 (m), 1140 (m), 870 (m), 773 (s), 691 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.26–7.20 (4H, m), 7.15–7.10 (1H, m), 6.12 (1H, dt, J = 6.4, 2.5 Hz), 5.61 (1H, ddd, J = 6.9, 6.9, 6.9 Hz), 3.37–3.19 (2H, m), 2.73–2.52 (4H, m), 1.92–1.72 (4H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.8, 134.3, 128.6, 126.9, 126.7, 94.8, 92.5, 54.6, 53.7, 23.6; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₄H₁₇N: 200.1434, found: 200.1435.

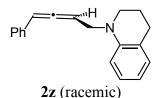


1-(4-phenylbuta-2,3-dien-1-yl)azepane (2y): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (70:30 hexanes:EtOAc with 5% Et₃N) to yield **2y** as a pale yellow oil (35.3 mg, 0.155 mmol, 77.6% yield). **IR** (neat, cm⁻¹) 2919 (m), 2580 (w), 1945 (m), 1457 (m), 1315 (w), 1123 (w), 1077 (m), 871 (m), 771 (s), 689 (s), 630 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.32–7.25 (4H, m), 7.20–7.15 (1H, m), 6.15 (1H, dt, J = 6.4, 2.4 Hz), 5.62 (1H, ddd, J = 6.9, 6.9, 6.9 Hz), 3.30 (2H, dd, J = 7.0, 2.4 Hz), 2.79–2.64 (4H, m), 1.75–1.48 (8H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.9, 134.6, 128.6, 126.8, 126.7, 94.5, 92.1, 57.8, 55.2, 28.3, 26.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₁N: 228.1747, found: 228.1749.

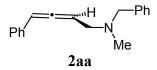
<u>By General Method E:</u> (*R*)-2y was isolated as a pale yellow oil (54.1 mg, 0.238 mmol, 59.5% yield). HPLC analysis indicated er = 92:8. $[\alpha]_D^{26} = -219.9$ (*c* = 1.0, CHCl₃) for a sample of 92:8 er.

HPLC: Column: Chiralpak 1A-3 (3 μ m, 4.6 mm X 150 mm). Mobile phase: 60:40 MeOH:*i*-PrOH with 0.1% Et₂NH additive, 0.4 mL/min. Detection wavelength: 254 nm. Er = 92:8.





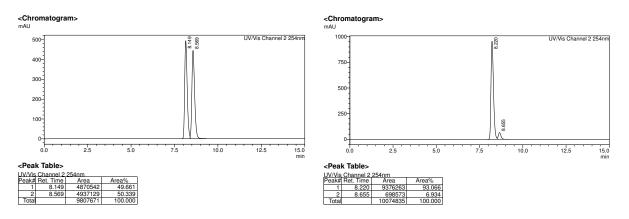
1-(4-phenylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroquinoline (2z): Prepared by General Method B using **Pd-1** at 22 °C for 20 h. The material was purified by flash silica gel chromatography (99:1 hexanes:EtOAc with 5% Et₃N) to yield **2z** as a pale yellow oil (31.9 mg, 0.122 mmol, 61.0% yield). **IR** (neat, cm⁻¹) 2842 (w), 1942 (w), 1704 (w), 1599 (m), 1494 (m), 1456 (m), 1264 (m), 732 (s), 699 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.20 (4H, m), 7.20–7.14 (1H, m), 7.06–7.00 (1H, m), 6.96 (1H, d, J = 7.3 Hz), 6.69 (1H, d, J = 8.2 Hz), 6.62 (1H, dd, J = 7.3, 7.3 Hz), 6.17 (1H, dt, J = 5.8, 2.7 Hz), 5.58 (1H, ddd, J = 6.3, 6.3, 6.3 Hz), 4.02 (2H, dd, J = 6.2, 2.7 Hz), 3.34–3.29 (2H, m), 2.73 (2H, t, J = 6.4 Hz), 2.01–1.90 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.7, 144.7, 134.3, 129.2, 128.6, 127.1, 126.9, 126.8, 123.0, 116.3, 111.5, 95.7, 90.6, 50.3, 49.3, 28.1, 22.3; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₁₉N: 262.1590, found: 262.1594.



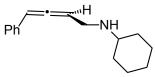
N-benzyl-N-methyl-4-phenylbuta-2,3-dien-1-amine (2aa): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (hexanes with 5% Et₃N) to yield **2aa** as a pale yellow oil (44.6 mg, 0.179 mmol, 89.4% yield). **IR** (neat, cm⁻¹) 3027(w), 2783 (w), 1946 (m), 1597 (w), 1493 (m), 1452 (s), 1205 (s), 772 (s), 780 (s), 690 (s), 619 (m). ¹H NMR (400 MHz, CDCl₃) δ 7.37–7.23 (9H, m), 7.22–7.16 (1H, m), 6.19 (1H, dt, J = 6.4, 2.4 Hz), 3.62 (1H, AB_q, $J_{AB} = 13.1$ Hz), 3.58 (1H, AB_q, $J_{AB} = 13.1$ Hz), 3.25–3.20 (2H, m), 2.31 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 206.2, 138.9, 134.5, 129.1, 128.7, 128.3, 127.1, 126.9, 126.8, 94.7, 91.7, 61.2, 56.1, 41.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₈H₁₉N: 250.1590, found: 250.1592.

By General Method E: (*R*)-2aa was isolated as a clear oil (53.7 mg, 0.215 mmol, 53.8% yield). HPLC analysis indicated er = 93:7. $[\alpha]_{D}^{25} = -209.1$ (*c* = 1.0, CHCl₃) for a sample of 93:7 er.

HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 99:1 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 93:7.

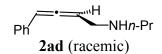


N,N-diethyl-4-phenylbuta-2,3-dien-1-amine (2ab): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (90:10 hexanes:EtOAc with 5%Et₃N) to yield **2ab** as a pale yellow oil (31.1 mg, 0.154 mmol, 77.2% yield). **IR** (neat, cm⁻¹) 2967 (m), 1947 (w), 1494 (w), 1457 (m), 1199 (w), 1089 (m), 868 (w), 763 (s), 713 (s), 629 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.46–7.13 (5H, m), 6.08 (1H, dt, J = 6.4, 2.3 Hz), 5.57 (1H, ddd, J = 6.8, 6.8, 6.8 Hz), 3.33 (2H, dd, J = 7.2, 2.0 Hz), 2.61 (4H, q, J = 7.2 Hz), 1.06 (6H, t, J = 7.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.9, 134.5, 128.6, 126.8, 126.7, 94.4, 90.6, 51.4, 46.7, 12.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₄H₁₉N: 202.1590, found: 202.1591.

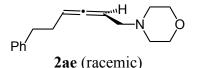


2ac (racemic)

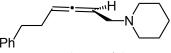
N-(4-phenylbuta-2,3-dien-1-yl)cyclohexanamine (2ac): Prepared by General Method B using 10 mol% **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (90:10:5 hexanes:EtOAc:Et₃N) to yield **2ac** as a colorless oil (26.6 mg, 0.117 mmol, 58.3% yield). **IR** (neat, cm⁻¹) 2925 (s), 2852 (m), 1944 (w), 1462 (w), 1450 (m), 1010 (w), 983 (w) 756 (m), 669 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.27–7.21 (4H, m), 7.16–7.10 (1H, m), 6.15 (1H, dt, *J* = 6.1, 2.8 Hz), 5.57 (1H, ddd, *J* = 6.3, 6.3, 6.3 Hz), 3.34 (2H, dd, *J* = 6.3, 2.9 Hz), 2.53–2.44 (1H, m), 1.87–1.73 (2H, m), 1.71–1.58 (2H, m), 1.56–1.50 (1H, m), 1.25–0.92 (6H, m); ¹³C NMR (125 MHz, CDCl₃) δ 204.8, 134.5, 128.6, 126.9, 126.7, 95.7, 94.6, 55.7, 45.2, 33.6, 33.4, 26.1, 25.1, 25.0 (note: two extra sp³ carbon was observed, but this has been noted before with related cyclohexyl-substituted compounds^{7,29}); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₁N: 228.1747, found: 228.1749.



4-phenyl-N-propylbuta-2,3-dien-1-amine (2ad): Prepared by General Method B using 10 mol% **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (90:10:5 hexanes:EtOAc:Et₃N) to yield **2ad** as a colorless oil (22.1 mg, 0.118 mmol, 59.0% yield). **IR** (neat, cm⁻¹) 2957 (m), 2930 (m), 1946 (m), 1494 (s), 1458 (m), 1072 (m), 910 (w), 773 (m), 690 (s); ¹H **NMR** (500 MHz, CDCl₃) δ 7.24–7.17 (4H, m), 7.14–7.08 (1H, m), 6.09–6.05 (1H, m), 5.52 (1H, ddd, J = 6.9, 6.9, 6.9 Hz), 3.35–3.30 (2H, m), 2.53–2.49 (2H, m), 1.98 (1H, br. s), 1.50–1.41 (2H, m), 0.82 (3H, t, J = 7.9 Hz); ¹³C **NMR** (125 MHz, CDCl₃) δ 206.1, 134.4, 128.6, 126.8, 126.7, 94.6, 91.2, 54.9, 52.4, 20.4, 11.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₃H₁₇N: 188.1434, found: 188.1431.

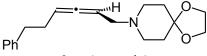


1-(6-phenylhexa-2,3-dien-1-yl)morpholine (2ae): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (2:1 hexanes:EtOAc with 5% Et₃N) to yield **2ae** as a pale yellow oil (40.1 mg, 0.165 mmol, 82.4% yield). **IR** (neat, cm⁻¹) 2917 (w), 2851 (w), 2799 (w), 1945 (w), 1452 (m), 1295 (w), 1114 (s), 1004 (m), 861 (s), 697 (s); ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.21 (2H, m), 7.20–7.14 (3H, m), 5.18–5.10 (2H, m), 3.03–2.85 (2H, m), 2.70 (2H, t, *J* = 7.8 Hz), 2.52–2.21 (6H, m), 1.69–1.49 (4H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.6, 141.5, 128.5, 128.3, 125.9, 90.4, 87.4, 66.9, 58.4, 53.2, 35.5, 30.5; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₁NO: 244.1696, found: 244.1696.



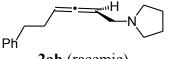
2af (racemic)

1-(6-phenylhexa-2,3-dien-1-yl)piperidine (2af): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (80:20 hexanes:EtOAc with 5% Et₃N) to yield **2af** as a pale yellow oil (36.4 mg, 0.151 mmol, 75.4% yield). **IR** (neat, cm⁻¹) 2930 (m), 1949 (w), 1495 (w), 1452 (m), 1440 (w),1336 (w), 1299 (w), 1108 (m), 743 (m), 696 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.23–7.16 (2H, m), 7.14–7.08 (3H, m), 5.17–5.10 (2H, m), 2.94–2.81 (2H, m), 2.64 (2H, app. t, J = 8.4 Hz), 2.34 (4H, br. s), 2..28–2.18 (2H, m), 1.52 (4H, app. p, J = 5.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.3, 141.7, 128.5, 128.3, 125.9, 90.2, 88.0, 58.7, 53.9, 35.5, 30.5, 25.9, 24.3; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₇H₂₃N: 242.1903, found: 242.1903.



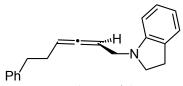
2ag (racemic)

8-(6-phenylhexa-2,3-dien-1-yl)-1,4-dioxa-8-azaspiro[4.5]decane (2ag): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (90:10 hexanes:EtOAc with 5% Et₃N) to yield **2ag** as a pale yellow oil (49.7 mg, 0.183 mmol, 91.6% yield). **IR** (neat, cm⁻¹) 2930 (w), 2850 (w), 2804 (w), 1947 (w), 1496 (m), 1451 (m), 1354 (m), 1268 (m), 1022 (s), 864 (s), 716 (s), 698 (s); ¹H NMR (500 MHz, CDCl₃) δ 7.25–7.17 (2H, m), 7.14–7.08 (3H, m), 5.14–5.02 (2H, m), 3.88 (4H, s), 2.98–2.87 (2H, m), 2.64 (2H, t, *J* = 7.7 Hz), 2.54–2.42 (4H, m), 2.28–2.20 (2H, m), 1.68 (4H, t, *J* = 5.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.3, 141.7, 128.5, 128.3, 125.9, 107.2, 90.4, 88.1, 64.2, 57.7, 50.8, 35.5, 34.9, 30.5; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₂₅NO₂: 300.1598, found: 300.1594.



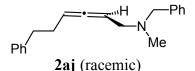
2ah (racemic)

1-(6-phenylhexa-2,3-dien-1-yl)pyrrolidine (2ah): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (2:1 hexanes:EtOAc with 5% Et₃N) to yield **2ah** as a pale yellow oil (33.9 mg, 0.122 mmol, 61.1% yield). **IR** (neat, cm⁻¹) 2925 (w), 2775 (w), 1943 (w), 1495 (w), 1453 (w), 1365 (w), 1123 (w), 866 (w), 743 (m), 697 (s); ¹H NMR (500 MHz, CDCl₃) δ 7.25–7.18 (2H, m), 7.13–7.10 (3H, m), 5.16–5.06 (2H, m), 3.05–2.95 (2H, m), 2.65 (2H, t, *J* = 7.2 Hz), 2.51–2.39 (4H, m) 2.34–2.18 (2H, m), 1.77–1.63 (4H, m); ¹³C NMR (125 MHz, CDCl₃) δ 204.8, 141.8, 128.5, 128.3, 128.9, 90.5, 89.2, 55.2, 53.6, 35.5, 30.5, 23.6; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₁N: 228.1747, found: 228.1747.

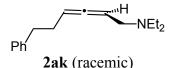


2ai (racemic)

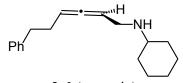
1-(6-phenylhexa-2,3-dien-1-yl)indoline (2ai): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (hexanes with 5% Et₃N) to yield **2ai** as a pale red oil (41.6 mg, 0.151 mmol, 75.5% yield).* **IR** (neat, cm⁻¹) 3024 (w), 2918 (w), 2827 (w), 1959 (w), 1605 (m), 1486 (m), 1382 (w), 1331 (w), 906 (s), 695 (s), 648 (w); ¹H **NMR** (500 MHz, CDCl₃) δ 7.24–7.19 (2H, m), 7.14–7.09 (3H, m), 7.02–6.96 (2H, m), 6.58 (1H, dd, J = 7.2, 7.2 Hz), 6.41 (1H, d, J = 7.7 Hz), 5.15–5.09 (1H, m), 5.09–5.03 (1H, m), 3.67–3.57 (2H, m), 3.34–3.23 (2H, m), 2.87 (2H, t, J = 8.3 Hz), 2.68–2.58 (2H, m), 2.30–2.17 (2H, m); ¹³C **NMR** (125 MHz, CDCl₃) δ 205.2, 151.7, 141.7, 130.4, 128.6, 128.3, 127.2, 125.9, 124.5, 117.7, 107.6, 91.2, 86.8, 52.6, 48.0, 35.4, 30.5, 28.5; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁N: 276.1747, found: 276.1753.



N-benzyl-N-methyl-6-phenylhexa-2,3-dien-1-amine (2aj): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (hexanes with 5% Et₃N) to yield **2aj** as a pale yellow oil (50.2 mg, 0.181 mmol, 90.4% yield). **IR** (neat, cm⁻¹) 3022 (w), 2818 (w), 2703 (w), 1962 (m), 1612 (m), 1440 (m), 1212 (w), 912 (s), 884 (s), 698 (s); ¹H NMR (500 MHz, CDCl₃) δ 7.26–7.07 (10H, m), 5.23–5.17 (2H, m), 3.43 (2H, s), 2.98–2.90 (2H, m), 2.65 (2H, t, *J* = 7.7 Hz), 2.30–2.20 (2H, m), 2.13 (3H, s); ¹³C NMR (125 MHz, CDCl₃) δ 205.3, 141.7, 139.0, 129.1, 128.5, 128.3, 128.2, 126.9, 125.9, 90.3, 88.2, 61.1, 56.6, 41.8, 35.5, 30.6; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₃N: 276.1903, found: 276.1899.

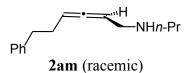


N,N-diethyl-6-phenylhexa-2,3-dien-1-amine (2ak): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (80:20 hexanes:EtOAc with 5% Et₃N) to yield **2ak** as a pale yellow oil (34.8 mg, 0.152 mmol, 75.9% yield). **IR** (neat, cm⁻¹) 2968 (w), 2932 (w) 2807 (w), 1965 (w), 1453 (w), 1264 (m), 1199 (w), 1065 (w), 733 (s), 697 (s); ¹H NMR (500 MHz, CDCl₃) δ 7.24–7.17 (2H, m), 7.14–7.08 (3H, m), 5.10–5.00 (2H, m), 3.05 (2H, dd, J = 7.1, 2.4 Hz), 2.65 (2H, t, J = 8.2 Hz), 2.47 (4H, q, J = 7.2 Hz), 2.30–2.19 (2H, m), 0.96 (6H, t, J = 7.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.1, 141.7, 128.5, 128.3, 125.9, 90.0, 87.1, 51.8, 46.5, 35.5, 30.6, 11.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₆H₂₃N: 230.1903, found: 230.1904.

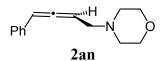


2al (racemic)

N-(6-phenylhexa-2,3-dien-1-yl)cyclohexanamine (2al): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc with 5% Et₃N) to yield **2al** as a pale yellow oil (28.2 mg, 0.124 mmol, 62.1% yield). **IR** (neat, cm⁻¹) 2998 (w), 2828 (m), 1956 (w), 1435 (m), 1234 (m), 1146 (s), 763 (s), 622 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.24–7.19 (2H, m), 7.14–7.08 (3H, m), 5.18–5.08 (2H, m), 3.13 (2H, dd, J = 6.3, 2.9 Hz), 2.66 (2H, t, J = 7.7 Hz), 2.44–2.37 (1H, m), 2.30–2.23 (2H, m), 1.82–1.50 (4H, m), 1.24–0.90 (6H, m), 0.87 (1H, br. s); ¹³C NMR (125 MHz, CDCl₃) δ 203.8, 141.7, 128.5, 128.3, 125.9, 91.5, 90.9, 55.7, 45.4, 35.4, 33.6, 33.5, 30.4, 26.2, 25.1 (note: one extra sp³ carbon was observed, but this has been noted before with related cyclohexyl-substituted compounds^{7,29}); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₈H₂₅N: 256.2060, found: 256.2064.

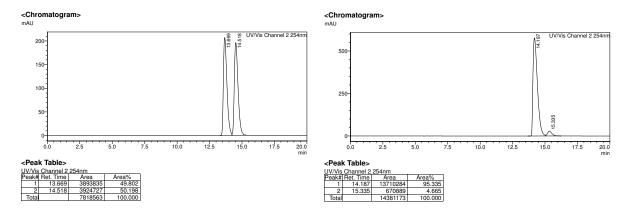


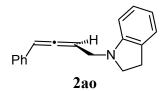
6-phenyl-N-propylhexa-2,3-dien-1-amine (2am): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc with 5% Et₃N) to yield **2am** as a pale yellow oil (22.0 mg, 0.109 mmol, 54.8% yield). **IR** (neat, cm⁻¹) 2956 (m), 1959 (w), 1495 (m), 1453 (s), 1334 (w), 1076 (m), 868 (w), 745 (m), 697 (s); ¹H **NMR** (400 MHz, CDCl₃) δ 7.25–7.15 (2H, m), 7.13–7.08 (3H, m), 5.13–4.97 (2H, m), 3.09–2.99 (2H, m), 2.65 (2H, t, *J* = 7.7 Hz), 2.39–2.31 (2H, m), 2.29–2.19 (2H, m), 1.47–1.33 (2H, m), 0.80 (3H, t, *J* = 7.4 Hz); ¹³C **NMR** (125 MHz, CDCl₃) δ 205.2, 141.7, 128.5, 128.3, 125.9, 90.1, 87.8, 54.7, 52.8, 35.5, 30.6, 20.3, 12.0; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₅H₂₁N: 216.1747, found: 216.1741.



(*R*)-4-(4-phenylbuta-2,3-dien-1-yl)morpholine (2an): Prepared by General Method E. The material was purified by flash silica gel chromatography (70:30 hexanes:EtOAc with an additional 2% Et₃N additive v/v) to yield (*R*)-2an as a pale yellow oil (56.8 mg, 0.264 mmol, 66.0% yield). HPLC analysis indicated er = 95.5:4.5. IR (neat, cm⁻¹) 3030 (w), 2956 (m), 2852 (m), 2801 (m), 1949 (m), 1713 (w), 1596 (w), 1494 (m), 1453 (m), 1344 (m), 1330 (m), 1315 (m), 1287 (m), 1270 (m), 1204 (w), 1114 (s), 1070 (m), 1034 (w), 1004 (s); ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.23 (4H, m), 7.22–7.16 (1H, m), 6.18 (1H, dt, *J* = 6.4, 2.4 Hz), 5.58 (1H, ddd, *J* = 7.0, 7.0, 7.0 Hz), 3.74 (4H, t, *J* = 4.6 Hz), 3.15 (2H, dd, *J* = 7.2, 2.4 Hz), 2.59–2.48 (4H, m); ¹³C NMR (125 MHz, CDCl₃) δ 206.3, 134.2, 128.6, 127.0, 126.7, 94.8, 91.2, 67.0, 57.8, 53.2; HRMS (ESI⁺) [M+H]⁺ calc'd for C₁₄H₁₇NO: 216.1383, found: 216.1381; [*α*]_D²⁶ = –185.6 (*c* = 1.0, CHCl₃) for a sample of 95.5:4.5 er.

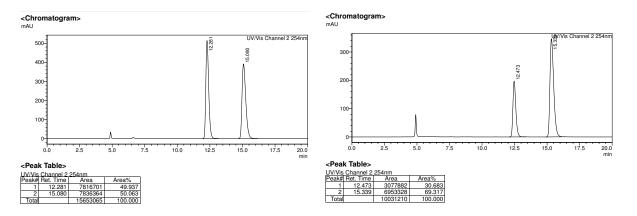
HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 99.9:0.1 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 95.5:4.5.

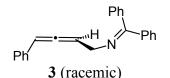




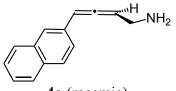
1-(4-phenylbuta-2,3-dien-1-yl)indoline (2ao): Prepared by General Method E. The material was purified by flash silica gel chromatography (100% hexanes to 98:2 hexanes:EtOAc) to yield (*R*)-**2ao** as a pale yellow oil (29.5 mg, 0.119 mmol, 59.6% yield).* HPLC analysis indicated er = 69.5:30.5. **IR** (neat, cm⁻¹) 3028 (w), 2920 (w), 2945 (w), 2846 (m), 1940 (m), 1604 (s), 1485 (m), 1469 (m), 1453 (m), 1384 (m), 1334 (m), 1305 (m), 1280 (m), 1261 (m), 1230 (m), 1199 (m), 1171 (m), 1151(m), 1140 (m), 1084 (m), 1068 (m), 1054 (m), 1020 (m), 985 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 7.32–7.23 (4H, m), 7.22–7.17 (1H, m), 7.10–7.03 (2H, m), 6.66 (1H, t, *J* = 7.4 Hz), 6.53 (1H, d, *J* = 7.8 Hz), 6.19 (1H, dt, *J* = 6.4, 2.4 Hz), 5.60 (1H, app. q, *J* = 6.7 Hz), 3.92 (1H, ddd, *J* = 14.7, 6.6, 2.5 Hz), 3.87 (1H, ddd, *J* = 14.8, 6.9, 2.5 Hz), 3.50–3.39 (2H, m), 2.98 (2H, t, *J* = 8.3 Hz); ¹³C **NMR** (125 MHz, CDCl₃) δ 206.1, 151.5, 134.3, 130.5, 128.7, 127.4, 127.1, 127.0, 124.6, 118.0, 107.8, 95.4, 90.4, 52.8, 47.9, 28.7; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₈H₁₇N: 248.1434, found: 248.1432; **[\alpha]_p**²⁷ = -59.3 (*c* = 1.0, CHCl₃) for a sample of 69.5:30.5 er.

HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 90:10 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 69.5:30.5.





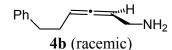
1,1-diphenyl-N-(4-phenylbuta-2,3-dien-1-yl)methanimine (3): To an oven-dried 2-dram vial equipped with a magnetic stirring rod was added Pd-1 (42.9 mg, 0.0500 mmol, 5.00 mol%) and CH_2Cl_2 (1.25 mL). To this suspension was added enyne **1a** (162 μ L, 1.20 mmol, 1.20 equiv), Et₃N (0.28 mL, 2.0 mmol, 2.0 equiv), and finally benzophenone imine (168 μ L, 1.00 mmol, 1.00 equiv). This mixture was allowed to stir at ambient temperature for 24 h. During this time the suspension gradually became a homogenous solution. The reaction mixture was then passed through a short plug of neutral alumina eluting with 1:1 hexanes: EtOAc (ca. 20 mL) and the solution concentrated. The product was purified by flash silica gel chromatography (hexanes with 2% Et₃N additive v/v to 95:5 hexanes: EtOAc with 2% Et₃N additive v/v) to afford **3** as a yellow solid (273 mg, 0.882 mmol, 88.2% yield). IR (neat, cm⁻¹) 3053 (w), 2902 (w), 1947 (w), 1620 (m), 1596 (w), 1574 (w), 1488 (w), 1459 (w), 1444 (m), 1425 (w), 1315 (m), 1287 (m), 1273 (m), 1258 (m), 1176 (w), 1154 (w), 1073 (w), 1024 (w), 996 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.63–7.57 (2H, m), 7.43–7.24 (10H, m), 7.21–7.12 (3H, m), 6.20 (1H, dt, J = 6.4, 2.9 Hz), 5.85 (1H, ddd, J = 6.4, 6.4, 6.4 Hz), 4.16-4.12 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.5, 169.5, 139.8, 136.5, 134.5, 130.1, 128.6, 128.6, 128.5, 128.1, 127.9, 126.9, 126.9, 95.8, 94.4, 52.8 (note: one sp² carbon is missing due to overlap); **HRMS** (ESI⁺) $[M+H]^+$ calc'd for C₂₃H₁₉N: 310.1590, found: 310.1595; **MP** = 71–74 °C.



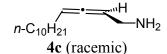
4a (racemic)

4-(naphthalen-2-yl)buta-2,3-dien-1-amine (4a): To an oven-dried 2-dram vial equipped with a magnetic stirring rod was added Pd-1 (42.9 mg, 0.0500 mmol, 5.00 mol%) and CH₂Cl₂ (1.25 mL). To this suspension was added envne 11 (206 μ L, 1.20 mmol, 1.20 equiv), Et₃N (0.28 mL, 2.0 mmol, 2.0 equiv), and finally benzophenone imine (168 μ L, 1.00 mmol, 1.00 equiv). This mixture was allowed to stir at ambient temperature for 24 h. During this time the suspension gradually became a homogenous solution. After this time, the reaction mixture was transferred to a 50-mL roundbottom flask, diluted with THF (5 mL), and then treated with 10% ag citric acid (10 mL). The reaction mixture was allowed to stir vigorously for 6 h. TLC analysis revealed that hydrolysis was complete after this time. The reaction mixture was diluted with EtOAc (30 mL) and basified with 2 M aq NaOH (30 mL). The aqueous fraction was separated from the organics and washed with EtOAc (2 X 10 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. The product was purified by flash silica gel chromatography (95:5 CH₂Cl₂:MeOH to 95:5 CH₂Cl₂:MeOH with 2% Et₃N additive v/v) to afford 4a as a yellow solid (136 mg, 0.696 mmol, 69.6% yield). IR (neat, cm⁻¹) 3300–2500 (br), 1944 (m), 1595 (m), 1506 (s), 1413 (w), 1364 (m), 1275 (m), 1250 (m), 1212 (m), 1144 (w), 1124 (w), 1006 (m), 949 (m); ¹H NMR (400 MHz, DMSO-*d*6) δ 7.87–7.80 (3H, m), 7.75 (1H, s), 7.53–7.39 (3H, m), 6.53 (1H, dt, *J* = 6.3, 2.8 Hz), 5.80 (1H, ddd, J = 6.1, 6.1, 6.1 Hz), 3.52 (2H, br s), 3.38–3.26 (2H, m); ¹³C NMR (125 MHz, DMSO-d6) § 205.4, 133.7, 132.7, 131.8, 128.6, 128.1, 128.1, 126.9, 126.3, 126.0, 125.2, 96.9,

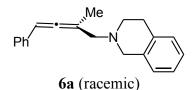
94.6 (note: one sp³ carbon missing due to overlap with DMSO-*d6*); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₄H₁₃N: 196.1121, found: 196.1123; **MP** = 170–176 °C (decomp.).



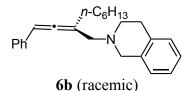
6-phenylhexa-2,3-dien-1-amine (4b): To an oven-dried 2-dram vial equipped with a magnetic stirring rod was added Pd-1 (42.9 mg, 0.0500 mmol, 5.00 mol%) and CH₂Cl₂ (1.25 mL). To this suspension was added envne 10 (202 μ L, 1.20 mmol, 1.20 equiv), Et₃N (0.28 mL, 2.0 mmol, 2.0 equiv), and finally benzophenone imine (168 μ L, 1.00 mmol, 1.00 equiv). This mixture was allowed to stir at ambient temperature for 24 h. During this time the suspension gradually became a homogenous solution. After this time, the reaction mixture was transferred to a 50-mL roundbottom flask, diluted with THF (5 mL), and then treated with 10% ag citric acid (10 mL). The reaction mixture was allowed to stir vigorously for 6 h. The reaction mixture was diluted with Et₂O (20 mL) and basified with 2 M aq NaOH (30 mL). The aqueous fraction was separated from the organics and washed with Et₂O (20 mL). The combined organic fractions were washed with sat aq brine (30 mL), dried over MgSO₄, filtered, and concentrated. The product was purified by flash silica gel chromatography (100% CH₂Cl₂ to 90:10 CH₂Cl₂:MeOH) to afford 4b as a pale yellow oil (100 mg, 0.579 mmol, 57.9% yield). NOTE: It is important that 4b be purified immediately after work up since this compound is not stable for long in its unpurified form. IR (neat, cm⁻¹) 3025 (w), 2918 (m), 2853 (m), 1962 (m), 1602 (m), 1495 (m), 1453 (m), 1333 (w), 1278 (w), 1078 (w), 1029 (w), 863 (m); ¹H NMR (500 MHz, CDCl₃) δ 7.30–7.23 (2H, m), 7.21–7.14 (3H, m), 5.31-5.19 (2H, m), 3.16 (2H, dd, J = 5.3, 3.3 Hz), 2.80-2.65 (2H, m), 2.43-2.25 (2H, m), 2.14(2H, br s); ¹³C NMR (125 MHz, CDCl₃) δ 203.4, 141.5, 128.6, 128.3, 126.0, 93.3, 92.1, 40.0, 35.2, 30.3; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₂H₁₅N: 174.1277, found: 174.1281.



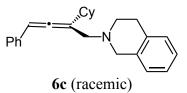
tetradeca-2,3-dien-1-amine (4c): To an oven-dried 2-dram vial equipped with a magnetic stirring rod was added Pd-1 (42.9 mg, 0.0500 mmol, 5.00 mol%) and CH₂Cl₂ (1.25 mL). To this suspension was added enyne 1p (288 μ L, 1.20 mmol, 1.20 equiv), Et₃N (0.28 mL, 2.0 mmol, 2.0 equiv), and finally benzophenone imine (168 μ L, 1.00 mmol, 1.00 equiv). This mixture was allowed to stir at ambient temperature for 24 h. During this time the suspension gradually became a homogenous solution. After this time, the reaction mixture was transferred to a 50-mL roundbottom flask, diluted with THF (5 mL), and then treated with 10% aq citric acid (10 mL). The reaction mixture was allowed to stir vigorously for 6 h. The reaction mixture was diluted with Et₂O (20 mL) and basified with 2 M aq NaOH (30 mL). The aqueous fraction was separated from the organics and washed with Et₂O (20 mL). The combined organic fractions were washed with sat aq brine (20 mL), dried over MgSO₄, filtered, and concentrated. The product was purified by flash silica gel chromatography (100% CH₂Cl₂ to 90:10 CH₂Cl₂:MeOH) to afford 4c as a pale yellow wax (119 mg, 0.570 mmol, 57.0% yield). NOTE: It is important that 4c be purified *immediately* after work up since this compound is not stable for long in its unpurified form. IR (neat, cm⁻¹) 2921 (s), 2852 (s), 1962 (w), 1598 (w), 1516 (w), 1465 (m), 1377 (w), 872 (m); ¹H NMR (500 MHz, CDCl₃) δ 5.30–5.20 (2H, m), 3.28 (2H, dd, J = 5.2, 3.4 Hz), 2.87 (2H, dd, J = 5.2, 3.4 Hz), 2.87 (2H, br s), 2.04–1.96 (2H, m), 1.42–1.19 (16 H, m), 0.86 (3H, t, J = 6.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 203.3, 94.0, 91.4, 40.3, 31.9, 29.6, 29.6, 29.5, 29.3, 29.2, 29.1, 28.8, 22.7, 14.1; HRMS (ESI⁺) [M+H]⁺ calc'd for C₁₄H₂₇N: 210.2216, found: 210.2219.



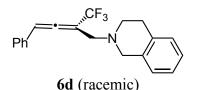
2-(2-methyl-4-phenylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinolineamine (6a): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc) to yield **6a** as a pale yellow oil (33.7 mg, 0.122 mmol, 61.1% yield).* **IR** (neat, cm⁻¹) 3030 (w), 2931 (s), 2808 (m), 1956 (m), 1644 (m), 1451 (m), 1303 (w), 988 (s), 824 (s), 773 (s), 628 (w); ¹H NMR (500 MHz, CDCl₃) δ 7.27–7.19 (4H, m), 7.14–7.09 (1H, m), 7.06–6.99 (3H, m), 6.95–6.89 (1H, m), 6.08–6.02 (1H, m), 3.63 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.59 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.16 (1H, ABX_q, *J_{AB}* = 12.8 Hz, *J_{BX}* = 2.2 Hz), 3.12 (1H, ABX_q, *J_{AB}* = 12.8 Hz, *J_{BX}* = 2.2 Hz), 2.81 (2H, t, *J* = 5.9 Hz), 2.76–2.64 (2H, m), 1.81 (3H, d, *J* = 2.9 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 204.1, 135.4, 134.9, 134.5, 128.7, 128.6, 126.8, 126.7, 126.6, 126.0, 125.6, 100.9, 93.7, 61.8, 56.2, 50.5, 29.1, 17.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁N: 276.1747, found: 276.1748.



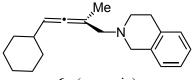
2-(2-(2-phenylvinylidene)octyl)-1,2,3,4-tetrahydroisoquinoline (6b): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc) to yield **6b** as a pale yellow oil (45.8 mg, 0.134 mmol, 67.0% yield). **IR** (neat, cm⁻¹) 3042 (m), 2899 (s), 1977 (m), 1503 (m), 1434 (m), 1206 (m), 1188 (m), 969 (s), 845 (m), 712 (s), 680 (w); ¹**H NMR** (500 MHz, CDCl₃) δ 7.27–7.21 (4H, m), 7.15–7.10 (1H, m), 7.06–7.02 (3H, m), 6.95–6.90 (1H, m), 6.17–6.08 (1H, m), 3.62 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.57 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.17 (1H, ABX_q, *J_{AB}* = 12.8 Hz, *J_{BX}* = 1.9 Hz), 3.12 (1H, ABX_q, *J_{AB}* = 12.8 Hz, *J_{BX}* = 1.9 Hz), 3.12 (1H, ABX_q, *J_{AB}* = 12.8 Hz, *J_{AX}* = 1.9 Hz), 2.86–2.78 (2H, m), 2.77–2.62 (2H, m), 2.19–2.03 (2H, m), 1.50–1.36 (2H, m), 1.32–1.14 (4H, m), 0.86–0.71 (5H, m); ¹³C NMR (125 MHz, CDCl₃) δ 203.6, 135.6, 135.1, 134.5, 134.4, 128.7, 128.6, 126.8, 126.6, 126.0, 125.5, 105.9, 95.1, 60.9, 56.3, 50.5, 31.7, 30.7, 29.2, 29.1, 27.7, 22.7, 14.1; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₅H₃₁N: 346.2526, found: 346.2529.



2-(2-cyclohexyl-4-phenylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (6c): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc) to yield **6c** as a pale yellow oil (38.0 mg, 0.106 mmol, 53.2% yield). **IR** (neat, cm⁻¹) 3027 (w), 2910 (s), 2848 (m), 1944 (w), 1597 (w), 1495 (m), 1446 (m), 1333 (m), 1086 (m), 933 (s), 820 (m), 738 (s), 692 (s), 649 (m), 626 (w); ¹**H NMR** (500 MHz, CDCl₃) δ 7.26–7.18 (4H, m), 7.14–7.07 (1H, m), 7.04–6.98 (3H, m), 6.93–6.87 (1H, m), 6.16–6.12 (1H, m), 3.62 (1H, ABq, *J*_{AB} = 14.9 Hz), 3.53 (1H, ABq, *J*_{AB} = 14.9 Hz), 3.18 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.13 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.13 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.13 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.13 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.13 (1H, ABXq, *J*_{AB} = 13.0 Hz, *J*_{BX} = 2.2 Hz), 3.25, 29.2, 26.5, 26.4 (note: one extra sp³ carbon was observed, but this has been noted before with related cyclohexyl-substituted compounds^{7,29}); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₅H₂₉N: 344.2373, found: 344.2377.



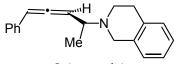
2-(4-phenyl-2-(trifluoromethyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (6d): Prepared by General Method B using Pd-1 at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc) to yield 6d as a pale yellow oil (42.2 mg, 0.123 mmol, 61.4% yield). IR (neat, cm⁻¹) 3024 (w), 2922 (w), 2798 (w), 1957 (w), 1497 (m), 1462 (m), 1279 (m), 1174 (s), 1073 (w), 935 (w), 742 (s), 691 (s), 652 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.38–7.27 (5H, m), 7.13–7.01 (3H, m), 6.96–6.91 (1H, m), 6.70–6.63 (1H, m), 3.75 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.68 (1H, AB_q, *J_{AB}* = 14.9 Hz), 3.46 (1H, ABX_q, *J_{AB}* = 15.2 Hz, *J_{BX}* = 3.0 Hz), 3.41 (1H, ABX_q, *J_{AB}* = 15.2 Hz, *J_{BX}* = 3.0 Hz), 2.89–2.72 (4H, m); ¹³C NMR (125 MHz, CDCl₃) δ 205.6 (q, *J_{CF}* = 4.3 Hz), 134.5, 134.2, 131.9, 131.5, 129.0, 128.7, 127.5, 126.6, 126.2, 125.7, 125.3 (q, *J_{CF}* = 273 Hz), 101.3, 100.0 (q, *J_{CF}* = 33.5 Hz), 55.8, 55.1, 50.3, 29.0; ¹⁹F NMR (376 MHz, CDCl₃) -62.46 (3F, d, *J* = 3.2 Hz); HRMS (ESI⁺) [M+H]⁺ calc'd for C₂₀H₁₈F₃N: 330.1464, found: 330.1470.



6e (racemic)

2-(4-cyclohexyl-2-methylbuta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (6e): Prepared by General Method B using **Pd-1** at 22 °C for 3 h. The material was purified by flash silica gel chromatography (95:5 hexanes:EtOAc) to yield **6e** as a pale yellow oil (43.9 mg, 0.156 mmol,

77.8% yield). **IR** (neat, cm⁻¹) 2902 (s), 2849 (m), 2791 (w), 1938 (w), 1497 (w), 1447 (m), 1367 (w), 1128 (w), 1089 (m), 933 (w), 908 (w), 739 (s); ¹**H NMR** (500 MHz, CDCl₃) δ 7.09–6.98 (3H, m), 6.98–6.91 (1H, m), 5.03–4.94 (1H, m), 3.57 (1H, AB_q, *J*_{AB} = 15.5 Hz), 3.53 (1H, AB_q, *J*_{AB} = 15.5 Hz), 3.03–2.96 (2H, m), 2.82 (2H, t, *J* = 5.9 Hz), 2.71–2.59 (1H, m), 1.76–1.60 (7H, m), 1.31–0.97 (6H, m); ¹³**C NMR** (125 MHz, CDCl₃) δ 201.7, 135.2, 134.6, 128.7, 126.6, 125.9, 125.5, 97.3, 95.9, 62.6, 56.2, 50.3, 37.6, 33.3, 29.2, 26.3, 26.1, 22.7, 17.6 (note: one extra sp³ carbon was observed, but this has been noted before with related cyclohexyl-substituted compounds^{7,29}); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₇N: 282.2216, found: 282.2214.



8 (racemic)

5-phenylpenta-3,4-dien-2-yl)-1,2,3,4-tetrahydroisoquinoline (8): Prepared by General Method C (0.40 mmol scale) at 22 °C for 20 h. ¹H NMR analysis of the unpurified material revealed a 2:1 mixture of diastereomers. The material was purified by flash silica gel chromatography (85:15 hexanes:EtOAc) to yield **8** as a pale yellow oil (28.2 mg, 0.102 mmol, 25.6% yield). The diastereomers could not be separated and all characterization data is for material with 2:1 dr. **IR** (neat, cm⁻¹) 3027 (w), 2970 (w), 2918 (w), 2799 (w), 1944 (m), 1597 (m), 1495 (m), 1455 (m), 1374 (w), 1333 (w), 1279 (w), 1267 (w), 1232 (w), 1210 (w), 1193 (w), 1155 (w), 1134 (m), 1099 (m), 1078 (w), 1027 (w), 934 (m); ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.29 (4H, m), 7.24–7.18 (1H, m), 7.15–7.09 (3H, m), 7.07–7.02 (1H, m), 6.25 (0.33 H, dd, *J* = 6.4, 2.2 Hz), 6.22 (0.66 H, dd, *J* = 6.4, 1.9 Hz), 5.72–5.65 (1H, m), 3.89–3.75 (2H, m), 3.60–3.52 (1H, m), 3.00–2.77 (4H, m), 1.37 (1H, d, *J* = 6.7 Hz), 1.34 (2H, d, *J* = 6.7 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 205.0, 135.2, 135.1, 134.6, 134.6, 234.5, 134.5, 128.8. 128.7, 128.7, 126.9, 126.9, 126.8, 126.7, 126.7, 126.1, 125.6, 95.9, 95.6, 95.5, 95.3, 59.1, 59.1, 52.0, 51.9, 46.7, 46.7, 29.7, 17.6, 17.2 (note: one sp, two sp², and one sp³ carbon were not detected due to overlap from complex mixture of diastereomers); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₀H₂₁N: 276.1747, found: 276.1753.

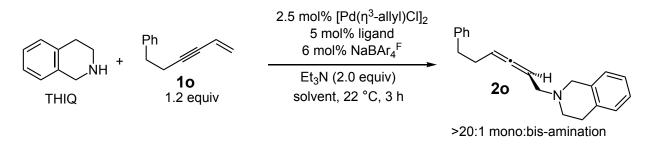
Following the procedure above (0.40 mmol scale) with addition of Et₃N (112 μ L, 0.80 mmol, 2.0 equiv) to the reaction mixture, **8** is isolated as a pale yellow oil (65.3 mg, 0.237 mmol, 59.3% yield). ¹H NMR analysis of the unpurified material revealed a 1:1 mixture of diastereomers. These diastereomers were not separated by chromatography.

VII. Supplemental Substrate Screening

1. Enantioselective Reaction Screening with Alkyl-Substituted Enynes

Efforts to expand the enantioselective reaction scope to alkyl-substituted enynes was met with limited success. Although PHOX ligands, promote the reactions, yields and er are poor (Table S5).

Table S5. Investigation of Alkyl-Substituted Enynes in the Enantioselective Reaction

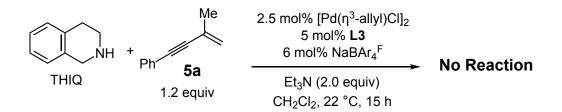


entry	ligand	solvent	yield (%) ^b	er ^c
1	L1	Et ₂ O	49	52:48
2	L2	CH_2CI_2	40	61:39
3	L3	CH_2CI_2	35	59:41

^aReactions run with 0.20 mmol THIQ in 0.25 mL solvent. ^bIsolated yield of pure **20**. ^cDetermined by HPLC analysis of pure **20** in comparison with an authentic racemic standard.

2. Enantioselective Reaction Screening with 1,3-Disubstituted 1,3-Enynes

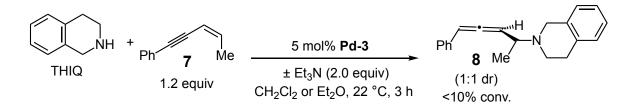
Efforts to expand the enantioselective reaction scope to 1,3-disubstituted enynes was met without success. Under standard reaction conditions, THIQ failed to add to enynes **5a**. Starting materials were recovered from this experiment and <5 % desired product formation was observed by ¹H NMR of the unpurified material.



3. Enantioselective Reaction Screening with 1,4-Disubstituted 1,3-Enynes

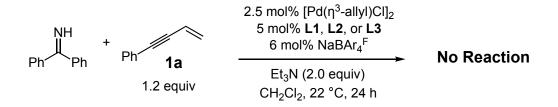
Efforts to expand the enantioselective reaction scope to 1,4-disubstituted enynes was met with little success. In the presence of a Pd-PHOX catalyst, the addition of THIQ to enyne 7 resulted in

< 10% conversion by ¹H NMR of the unpurified reaction mixture to a 1:1 mixture of diastereomers of **8**. The er of this product was not determined.



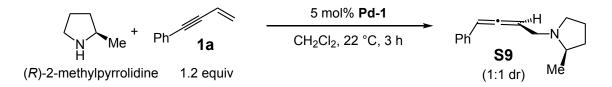
4. Enantioselective Reaction Screening with Benzophenone Imine

Efforts to expand the enantioselective reaction scope to benzophenone imine was met without success. Under standard reaction conditions with PHOX ligands L1, L2 or L3, benzophenone imine failed to add to enyne 1a. Starting materials were recovered from these experiments and <5% desired product formation was observed by ¹H NMR of the unpurified material.



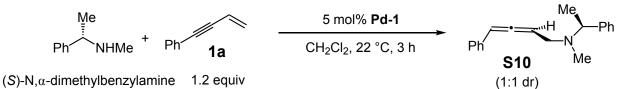
5. Addition of a Chiral Amine to Enyne 1a Under Racemic Reaction Conditions

Experiments to determine if the chirality of the amine nucleophile can influence the stereochemistry of the allene in the product is presented below. When (*R*)-2-methylpyrrolidine (>99:1 er) or (*S*)-N, α -dimethylbenzylamine (>99:1 er) was added to enyne 1a in the presence of 5 mol % Pd-1, the resulting products S9 and S10 were produced as 1:1 mixtures of diastereomers. Therefore, the chirality of amine nucleophiles has little to no influence on the stereochemistry of the allene.



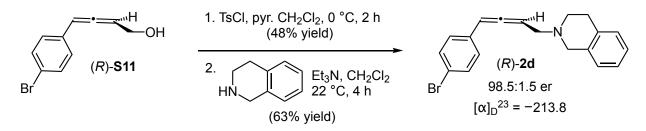
(*R*)-2-methyl-1-(4-phenylbuta-2,3-dien-1-yl)pyrrolidine (S9): Prepared by General Method B using Pd-1 at 22 °C for 3 h. ¹H NMR analysis of the unpurified material revealed a 1:1 mixture of diastereomers. The material was purified by flash silica gel chromatography (60:40 hexanes:EtOAc with 2% v/v Et₃N additive) to yield S9 as a pale yellow oil (34.7 mg, 0.163 mmol, 81.3% yield). The diastereomers could not be separated and all characterization data is for material

with 1:1 dr. **IR** (neat, cm⁻¹) 3030 (w), 2961 (m), 2869 (w), 2790 (m), 1949 (m), 1597 (w), 1495 (m), 1458 (m), 1375 (m), 1353 (w), 1308 (w), 1267 (w), 1220 (w), 1196 (w), 1167 (m), 1139 (m), 1071 (w), 1028 (w), 910 (m); ¹**H NMR** (400 MHz, CDCl₃) δ 7.32–7.25 (4H, m), 7.21–7.15 (1H, m), 6.19–6.11 (1H, m), 5.68–5.60 (1H, m), 3.62–3.52 (1H, m), 3.22 (0.5 H, ddd, *J* = 8.8, 8.8, 2.7 Hz), 3.13 (0.5 H, ddd, *J* = 9.0, 9.0, 2.8 Hz), 3.01 (0.5 H, ddd, *J* = 13.4, 8.0, 2.0 Hz), 2.95 (0.5 H, ddd, *J* = 13.0, 8.4, 1.6 Hz), 2.50–2.37 (1H, m), 2.36–2.25 (1H, m), 1.99–1.88 (1H, m), 1.84–1.62 (2H, m), 1.50–1.38 (1H, m), 1.13 (1.5 H, d, *J* = 6.1 Hz), 1.11 (1.5 H, d, *J* = 6.1 Hz); ¹³C **NMR** (125 MHz, CDCl₃) δ 205.8, 205.7, 134.6, 134.5, 128.6, 128.6, 126.8, 126.8, 126.7, 94.6, 94.5, 92.0, 91.9, 58.7, 58.4, 53.7, 53.5, 52.2, 51.8, 32.9, 32.9, 21.7, 21.6, 19.0, 18.9; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₅H₁₉N: 214.1590, found: 214.1596.



N-methyl-4-phenyl-*N*-((*S*)-1-phenylethyl)buta-2,3-dien-1-amine (S10): Prepared by General Method B using Pd-1 at 22 °C for 3 h. ¹H NMR analysis of the unpurified material revealed a 1:1 mixture of diastereomers. The material was purified by flash silica gel chromatography (85:15 hexanes:EtOAc) to yield S10 as a pale yellow oil (35.5 mg, 0.135 mmol, 67.4% yield). The diastereomers could not be separated and all characterization data is for material with 1:1 dr. **IR** (neat, cm⁻¹) 3082 (w), 3061 (w), 3029 (w), 2973 (m), 2936 (w), 2874 (w), 2839 (w), 2782 (m), 1946 (m), 1599 (w), 1495 (m), 1458 (m), 1449 (m), 1418 (w), 1404 (w), 1369 (w), 1346 (w), 1313 (m), 1281 (w), 1200 (m), 1155 (w), 1122 (w), 1071 (m), 1054 (w), 1028 (m), 1013 (w), 999 (w), 954 (w); ¹H NMR (400 MHz, CDCl₃) δ 7.36–7.15 (10H, m), 6.20–6.12 (1H, m), 5.64–5.54 (1H, m), 3.72–3.61 (1H, m), 3.39–3.25 (1H, m), 3.18–3.06 (1H, m), 2.30 (1.5 H, s), 2.30 (1.5 H, s), 1.39 (1.5 H, d, *J* = 6.5 Hz), 1.38 (1.5 H, d, *J* = 6.6 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 206.0, 206.0, 144.1, 144.0, 134.6, 134.5, 128.6, 128.3, 127.6, 127.6, 127.0, 126.9, 126.8, 126.8, 126.7, 94.5, 91.6, 62.7, 62.6, 53.7, 53.6, 39.0, 38.9, 19.7, 19.6 (note: four sp² carbons could not be identified due to complex overlap of diastereomer peaks); **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₁₉H₂₁N: 264.1747, found: 264.1754.

VIII. Stereoproof

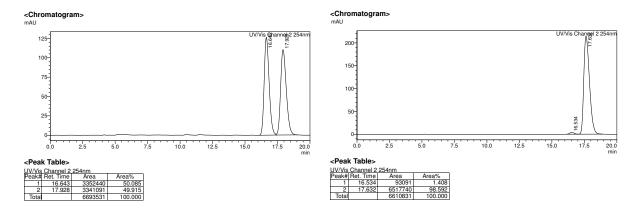


(*R*)-S11 was prepared by a previously described method where the absolute configuration is known.³⁰

(*R*)-4-(4-bromophenyl)buta-2,3-dien-1-yl 4-methylbenzenesulfonate (S12): To an oven-dried 2-dram vial equipped with a magnetic stirring rod was added (*R*)-S11 (225 mg, 1.00 mmol, 1.00 equiv) and CH₂Cl₂ (1.0 mL). The reaction mixture was allowed to cool to 0 °C in an ice water bath, and then pyridine (161 μ L, 2.00 mmol, 2.00 equiv) was added followed by *p*-TsCl (286 mg, 1.50 mmol, 1.50 equiv). The reaction mixture was allowed to stir at 0 °C for 2 h. The reaction contents were then partitioned between water (10 mL) and EtOAc (5 mL), and the aqueous layer was separated from the organics and washed with EtOAc (5 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. The product was then purified by flash silica gel chromatography (90:10 to 85:15 hexanes:EtOAc) to yield the tosylate (*R*-S12) as a colorless oil (183 mg, 0.482 mmol, 48.2 % yield). ¹H NMR (400 MHz, CDCl₃) δ 7.83–7.75 (2H, m), 7.44–7.37 (2H, m), 7.31 (2H, d, *J* = 8.2 Hz), 7.11–7.04 (2H, m), 6.19 (1H, dt, *J* = 6.4, 2.1 Hz), 5.64 (1H, app. q, *J* = 6.9 Hz), 4.67–4.58 (2H, m), 2.43 (3H, s). This product was found to be particularly unstable and so was used immediately in the next step.

(R)-2-(4-(4-bromophenyl)buta-2,3-dien-1-yl)-1,2,3,4-tetrahydroisoquinoline (*R*-2d): The tosylate (R)-S12 prepared above (114 mg, 0.300 mmol, 1.00 equiv) was added to an oven-dried 2dram vial equipped with a magnetic stirring rod and dissolved in CH₂Cl₂ (1.0 mL). To this solution was added Et₃N (84.0 μ L, 0.600 mmol, 2.00 equiv) and 1.2,3,4-tetrahydroisoquinoline (THIQ, 57.0 μ L, 0.450 mmol, 1.50 equiv). The mixture was allowed to stir at ambient temperature for 4 h. The reaction contents were then partitioned between sat aq NaHCO₃ (10 mL) and EtOAc (5 mL). The aqueous layer was separated from the organics and washed with EtOAc (2 X 3 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. Flash silica gel chromatography (75:25 hexanes: EtOAc) then afforded (R)-2d as a yellow solid (64.0 mg, 0.188 mmol, 62.7% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.46–7.40 (2H, m), 7.20–7.14 (2H, m), 7.14–7.07 (3H, m), 7.04–6.99 (1H, m), 6.16 (1H, dt, J = 6.4, 2.4 Hz), 5.68 (1H, ddd, J 6.9, 6.9, 6.9 Hz), 3.74 (1H, AB_q, $J_{AB} = 15.8$ Hz), 3.70 (1H, AB_q, $J_{AB} = 15.8$ Hz), 3.38–3.29 (2H, m), 2.97-2.89 (2H, m), 2.85-2.79 (2H, m); $[\alpha]_{D}^{23} = -213.8$ (c = 1.0, CHCl₃) for a sample of 98.5:1.5 er. The spectral data match those from (R)-2d as prepared by envne hydroamination. The direction of the specific rotation and comparison of HPLC retention times confirm that the major enantiomer formed by enantioselective hydroamination of the corresponding 1,3-envne with L3 is the (R)-enantiomer. The absolute stereochemistry for all other allene products is based on the stereochemical assignment for (R)-2d.

HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 80:20 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 98.5:1.5

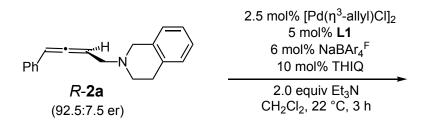


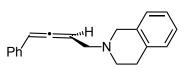
IX. Transamination/Reaction Reversibility Studies

The following experiments were carried out to elucidate aspects of the mechanism pertaining to reaction reversibility and racemization of allene products, and were conducted based on the below general procedure.

<u>General Procedure F (Reversibility Studies)</u>: In an N₂-filled glovebox, to a 2-dram vial equipped with a magnetic stirring rod were added successively: $[Pd(\eta^3-allyl)Cl]_2$ (1.8 mg, 5.0 μ mol, 2.5 mol %), appropriate ligand (0.010 mmol, 5.0 mol %), and NaBAr₄^F (10.6 mg, 0.012 mmol, 6.0 mol %). To a separate 2-dram vial was added (*R*)-**2a** (52.3 mg, 0.200 mmol, 1.00 equiv, 92.5:7.5 er) and dissolved in CH₂Cl₂ (50 μ L). The solution containing (*R*)-**2a** was added by micro syringe to the solution containing the Pd pre-catalyst (rinsing with an addition 50 μ L CH₂Cl₂). To the resulting solution was added Et₃N (56 μ L, 0.40 mmol, 2.0 equiv) followed by appropriate amine nucleophile (THIQ = 2.5 μ L, 0.020 mmol, 10 mol % *or* morpholine = 17.0 μ L, 0.200 mmol, 1.00 equiv). The solutions were allowed to stir at ambient temperature for 3 h and then worked up and purified as described for **2a** and **2an** as appropriate (see Substrate Scope section of SI).

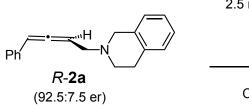
1. Reaction Reversibility Study with L1



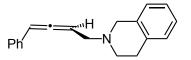


Trial 1: 79% recovery, 51:49 er Trial 2: 82% recovery, 53:47 er **Avg: 81% recovery, 52:48 er**

2. Reaction Reversibility Study with L3

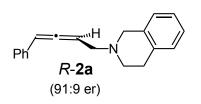


2.5 mol% $[Pd(\eta^{3}-allyl)Cl]_{2}$ 5 mol% L3 6 mol% NaBAr₄^F 10 mol% THIQ 2.0 equiv Et₃N CH₂Cl₂, 22 °C, 3 h

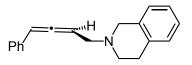


Trial 1: 68% recovery, 70.5:29.5 er Trial 2: 69% recovery, 73.5:26.5 er **Avg: 69% recovery, 72:28 er**

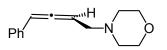
3. Transamination Study with L1 to Elucidate Mechanism of Racemization



2.5 mol% $[Pd(\eta^3-allyl)Cl]_2$ 5 mol% L1 6 mol% NaBAr₄^F 1.0 equiv morpholine 2.0 equiv Et₃N CH₂Cl₂, 22 °C, 3 h



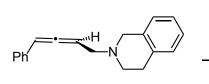
Trial 1: 57% recovery, 57.5:42.5 er Trial 2: 59% recovery, 54.5:45.5 er **Avg: 58% recovery, 56:44 er**



+

Trial 1: 24% yield, 59:41 Trial 2: 19% yield, 52:48 er **Avg: 22% yield, 55.5:44.5 er**

4. Prolonged Reaction Times for Transamination Experiments Results in Hydroamination of Allenes to Afford Bis-Amination Products.^{*a*}

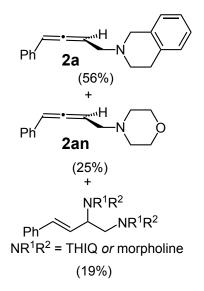


2a (racemic)

 $\begin{array}{c} \text{2.5 mol\%} \ [\text{Pd}(\eta^3\text{-allyl})\text{Cl}]_2\\ 5 \ \text{mol\%} \ \textbf{L1}\\ 6 \ \text{mol\%} \ \text{NaBAr_4}^\text{F}\\ 1.0 \ \text{equiv} \ \text{morpholine} \end{array}$

2.0 equiv Et₃N CH₂Cl₂, 22 °C, 15 h

^aYields in parentheses refer to the ratio of products determined by ¹H NMR of unpurified reaction mixture.



5. Transamination with DPEphos Derived Pd Catalyst Reveals Approximately 1:1 Ratio of Morpholine: THIQ Adducts Under a Variety of Conditions (Table S6).

2.5 mol% [Pd(η^3 -allyl)Cl]₂ ١H Ph 5 mol% DPEphos 1.0 equiv 2an 6 mol% NaBAr₄^F + base (2.0 equiv) ١H solvent, 22 °C, 3 h Ph Ph 2a 2a (racemic)

Table S6. Solvent and Base Screening of Transamination Reaction with DPEphos Pd Catalyst

entry	solvent	base	2an:2a ^a
1	Et ₂ O	Et₃N	1.5:1
2	CH ₂ Cl ₂	Et ₃ N	1.3:1
3	THF	Et ₃ N	1.4:1
4	hexanes	Et ₃ N	1.2:1
5	cyclohexane	Et ₃ N	1.4:1
6	dioxane	Et ₃ N	1.3:1
7	toluene	Et ₃ N	1.3:1
8	MeCN	Et ₃ N	1.3:1
9	acetone	Et ₃ N	1.3:1
10	Et ₂ O	<i>i</i> Pr ₂ NEt	1.5:1
11	Et ₂ O	DABCO	1.3:1
12	Et ₂ O	DBU ^b	>20:1
13	Et ₂ O	TMP	1.3:1
14	Et ₂ O	DMAP	0.9:1
15	Et ₂ O	TMG ^{b,c}	>20:1

^aDetermined by ¹H NMR of the unpurified reaction mixture. ^bBase additive does not promote the hydroamination of 1,3-enynes.

^cTMG = 1,1,3,3-tetramethylguanidine

To better understand the kinetics of enantiomerization of our allene products, we performed experiments aimed at monitoring the er over time for several different amine products. We chose to study indoline (pK_a conjugate acid = 5.6 in H₂O),³¹ morpholine (pK_a conjugate acid = 9.2 in DMSO and 8.5 in H₂O),³² and piperidine (pK_a conjugate acid = 10.9 in DMSO and 11.1 in H₂O)³² adducts **2ao**, **2an**, and **2w**, respectively, because these cover a broad range in terms pK_a and Lewis basicity.

1. Enantiomerization of Allenes in the Forward Progression of the Reaction

We aimed to discover if the rate and extent of er deterioration over time for the optimized reaction. A general procedure for these experiments is found below:

<u>General Procedure F:</u> In an N₂-filled glovebox, to a 2-dram vial equipped with a magnetic stirring rod were added successively: $[Pd(\eta^3-allyl)Cl]_2$ (3.7 mg, 10.0 μ mol, 1.0 mol %), L3 (13.5 mg, 0.020 mmol, 2.0 mol %), and NaBAr4^F (22.1 mg, 0.025 mmol, 2.5 mol %). The mixture was dissolved in Et₂O (1.25 mL) and allowed to stir for ca. 5 min, resulting in a deep orange/red solution, and then envne 1a (162 μ L, 1.20 mmol, 1.20 equiv) was added. The reaction vials were then capped with a PTFE lined cap, removed from the glovebox, and allowed to cool in an ice water bath at 0-4 °C for ca. 15 minutes before Et₃N (279 µL, 2.00 mmol, 2.00 equiv) and appropriate amine nucleophile (1.00 mmol, 1.00 equiv) were added. The reactions were allowed to stir at 4 °C for 24 h with aliquots (0.15 mL) taken at: 30 min, 1 h, 2 h, 4 h, 8 h, 16 h, and 24 h. These aliquots were passed through a short plug of silica gel eluting with a solvent system appropriate for each amine (2ao = ca. 20 mL 90:10 hexanes: EtOAc, 2an = ca. 20 mL 60:40 hexanes: EtOAc, and 2w =ca. 20 mL 60:40 hexanes: EtOAc with 2% v/v Et₃N additive). The aliquots were then concentrated under reduced pressure and dissolved in HPLC grade solvents (2ao and 2an = 10 mL 80:20 hexanes: IPA, 2w = 10 mL MeOH) and analyzed promptly (within 2 h) by HPLC for enantiomeric ratio. For HPLC methods and representative chromatograms for each compound, see the "Substrate Scope" section for 2ao, 2an, and 2w.

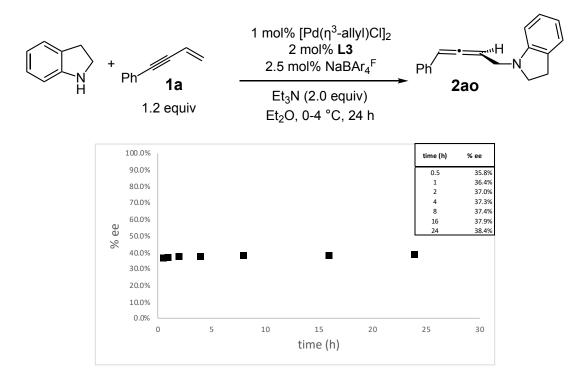
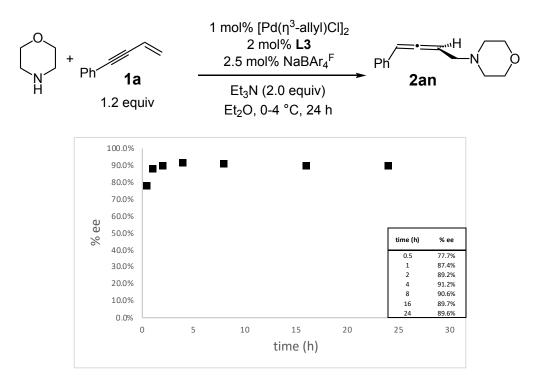


Chart S1. Enantiomeric Ratio of 2ao During Course of Reaction

Chart S2. Enantiomeric Ratio of 2an During Course of Reaction



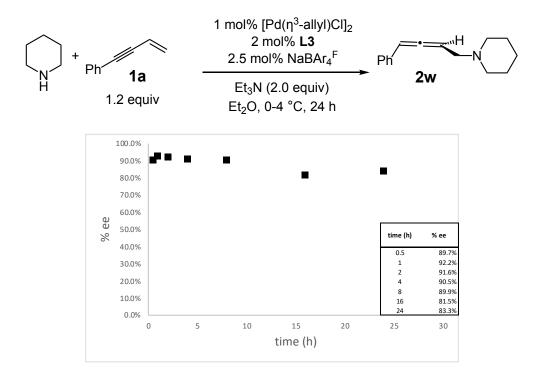
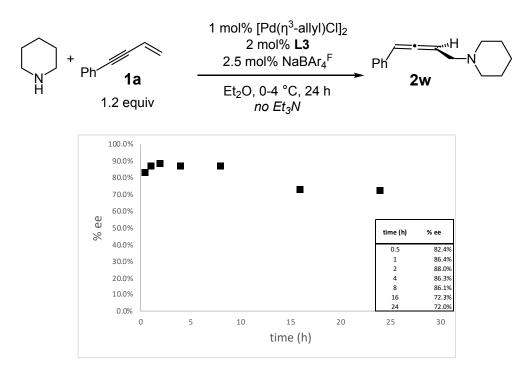


Chart S3. Enantiomeric Ratio of 2w During Course of Reaction

We additionally performed an experiment identical to Chart S3 but without Et₃N (following General Procedure F but without addition of Et₃N).

Chart S4. Enantiomeric Ratio of 2w During Course of Reaction without Et₃N



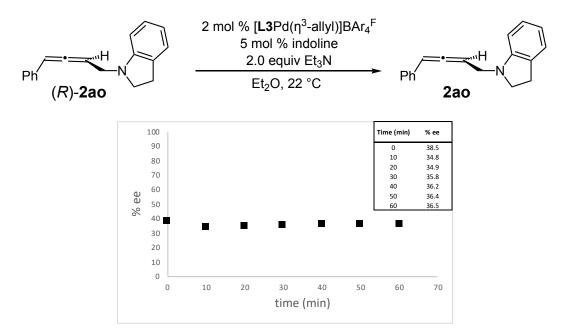
From the data above, we can conclude that an insignificant amount of enantiomerization occurs for indoline (**2ao**) and morpholine (**2an**) adducts (smaller pK_a values), whereas piperidine adduct **2w** demonstrates a very slow rate of enantiomerization (higher pK_a). It is unclear from the data above if Et₃N plays a significant role in the rate of enantiomerization, although slightly faster enantiomerization of **2w** occurs in its absence.

2. Isolated 2ao Resubjected to Reaction Conditions

Given the unusually low enantioselectivity of indoline addition, we performed an experiment where (R)-**2ao** (69:31 er) was resubjected to the reaction conditions to measure the rate of enantiomerization.

In an N₂-filled glovebox, to a 1-dram vial was added [L3Pd(η^3 -allyl)]BAr₄^F (3.4 mg, 2.0 μ mol, 2.0 mol %). This vial was capped with a rubber septum and removed from the glovebox. To this vial was added a solution of (*R*)-**2ao** (24.7 mg, 0.100 mmol, 1.00 equiv), indoline (25 μ L of a freshly prepared 0.20 M solution in Et₂O, 5.0 μ mol, 5.0 mol %), and Et₃N (28.0 μ L, 0.200 mmol, 2.00 equiv) in Et₂O (2 x 0.50 mL) via cannula transfer at ambient temperature. Experiments that indicate they were performed in the absence of Et₃N did not include this component in above solution. Aliquots (*ca.* 50 μ L) were taken at 10 min intervals during the reaction. These aliquots were immediately passed through a short plug of silica gel eluting with 80:20 hexanes:EtOAc (*ca.* 5–10 mL) and subsequently concentrated under reduced pressure. The concentrated aliquots were dissolved in HPLC grade 80:20 hexanes:IPA (*ca.* 1–2 mL) and analyzed promptly (within 2 h) by HPLC for enantiomeric ratio. For HPLC methods and representative chromatograms, see the "Substrate Scope" section for **2ao**.

Chart S5. Initial Rate of Enantiomerization of 2ao



An additional aliquot was taken after 46 h, but the er remained unchanged (36.8% ee).

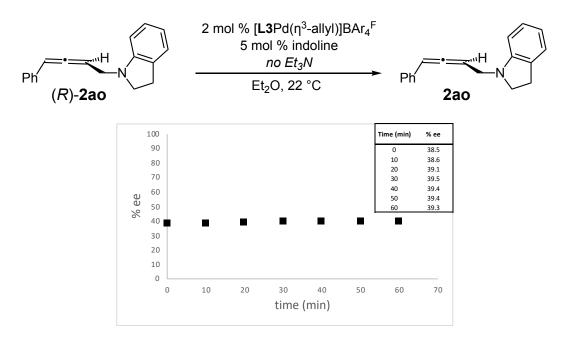


Chart S6. Initial Rate of Enantiomerization of 2ao without Et₃N

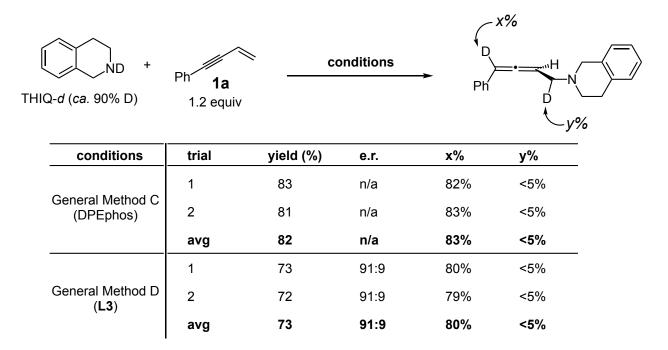
An additional aliquot was taken after 46 h, but the er remained unchanged (38.0% ee).

From the data in Charts S5 and S6 above, we can conclude that indoline adduct **2ao** does not significantly enantiomerize in the presence of a Pd catalyzed derived from L3. Therefore, the low enantioselectivity observed with indoline is likely due to kinetics of nucleophile addition and not post-reaction enantiomerization.

XI. Deuterium Labeling Studies

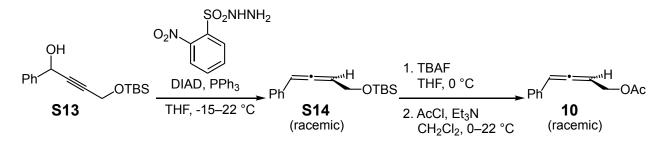
Deuterium labeling studies were carried out under standard non-enantioselective reaction conditions (General Method C) and enantioselective conditions (General Method D). All reactions performed on a 0.20 mmol scale with respect to THIQ-*d* and were otherwise carried out as described in "Substrate Scope" section of the SI (see above for **2a**). THIQ-*d* (*ca*. 90% deuterium incorporation) was prepared as previously described. Deuterium incorporation was measured based on disappearance of peaks in the ¹H NMR spectra of purified products using the AB quartet centered at *ca*. δ 3.76 (2H) in CDCl₃ as a reference for every measurement. An example ¹H NMR spectrum of *d*-**2a** can be found in the Supplementary Information Part B. Recovered starting material enyne from the reactions indicated <5% deuterium incorporation.

Table S7. Deuterium Labeling Experiments



XII. Allylic Substitution Experiments

The preparation of **S13** has been reported previously.³³

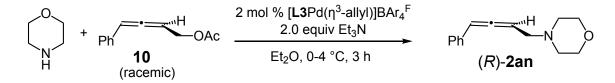


tert-butyldimethyl((4-phenylbuta-2,3-dien-1-yl)oxy)silane (S14): Prepared by the method of Myers and co-workers.³⁴ To a 250-mL round-bottom flask equipped with a magnetic stirring rod was added triphenylphosphine (3.75 g, 14.3 mmol, 1.30 equiv) and dissolved in THF (40 mL). This solution was cooled to -15 °C and diisopropyl azodicarboxylate (2.82 mL, 14.3 mmol, 1.30 equiv) was added dropwise via syringe. After 10 min of stirring at this temperature, S13 (3.04 g, 11.0 mmol, 1.00 equiv) was added as a solution in THF (40 mL) via cannula transfer. After an additional 10 min of stirring, 2-nitrobenzenesulfonohydrazide (3.11 g, 14.3 mmol, 1.30 equiv) was added as a solution in THF (40 mL) via cannula transfer. The resulting yellow solution was allowed to stir at -15 °C for 1 h and then warm to ambient temperature. Stirring was allowed to continue at ambient temperature for 13 h. After this time, the reaction mixture was concentrated under reduced pressure. Purification by flash silica gel chromatrography (99:1 hexanes:Et₂O) afforded S14 as a yellow oil (1.20 g, 4.59 mmol, 41.7% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.30–7.25 (4H, m), 7.21–7.15 (1H, m), 6.21 (dt, J = 6.4, 2.7 Hz), 5.67 (ddd, J = 6.3, 6.3, 6.3 Hz), 4.31–4.26 (2H, m), 0.89 (9H, s), 0.08 (3H, s), 0.07 (3H, s). Spectral data match those previously reported.³³

4-phenylbuta-2,3-dien-1-ol (S15): To a 100-mL round-bottom flask equipped with a magnetic stirring rod was added **S14** (1.04 g, 4.00 mmol, 1.00 equiv) and dissolved in THF (12 mL) and then cooled to 0 °C. TBAF (1.0 M in THF, 6.0 mL, 1.5 equiv) was added dropwise via syringe and stirring was allowed to continue at 0 °C for 3 h. The reaction mixture was partitioned between sat aq brine (50 mL) and Et₂O (50 mL). The aqueous layer was separated from organics and washed with Et₂O (30 mL). The combined organic fractions were dried over Na₂SO₄, filtered, and concentrated. Purification by flash silica gel chromatography (80:20 to 70:30 hexanes:Et₂O) afforded **S15** as a yellow oil (464 mg, 3.17 mmol, 79.3% yield).). ¹**H NMR** (400 MHz, CDCl₃) δ 7.33–7.26 (4H, m), 7.23–7.17 (1H, m), 6.31 (1H, dt, *J* = 6.4, 2.9 Hz), 5.78 (1H, ddd, *J* = 6.0, 6.0, 6.0 Hz), 4.28–4.20 (2H, m), 1.59–1.54 (1H, m). Spectral data match those previously reported.³⁵

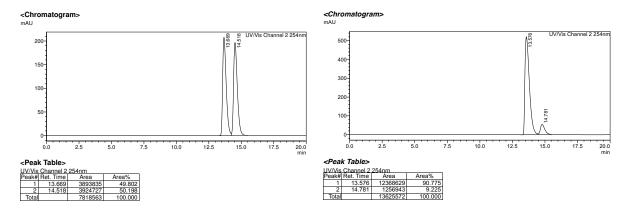
4-phenylbuta-2,3-dien-1-yl acetate (10): To a 50-mL round-bottom flask equipped with a magnetic stirring rod was added **S15** (421 mg, 2.88 mmol, 1.00 equiv) and dissolved in CH_2Cl_2 (10 mL). This solution was cooled to 0 °C and Et_3N (0.80 mL, 5.8 mmol, 2.0 equiv) was added followed by acetyl chloride (0.41 mL, 5.8 mmol, 2.0 equiv) dropwise via syringe. The reaction mixture was allowed to warm to ambient temperature and allowed to stir for 30 min. The reaction mixture was then poured into sat aq NaHCO₃ (20 mL). The aqueous layer was separated from organics and washed with CH_2Cl_2 (2 X 5 mL). The combined organic fractions were dried over MgSO₄, filtered, and concentrated. Purification by flash silica gel chromatography (90:10

hexanes:Et₂O) afforded pure **10** as a pale yellow oil (471 mg, 2.50 mmol, 86.9% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.33–7.25 (4H, m), 7.23–7.18 (1H, m), 6.28 (1H, dt, *J* = 6.4, 2.5 Hz), 5.70 (1H, ddd, *J* = 6.5, 6.5, 6.5 Hz), 4.66 (2H, dd, *J* = 6.6, 2.5 Hz), 2.06 (3H, s). Spectral data match those previously reported.³⁶



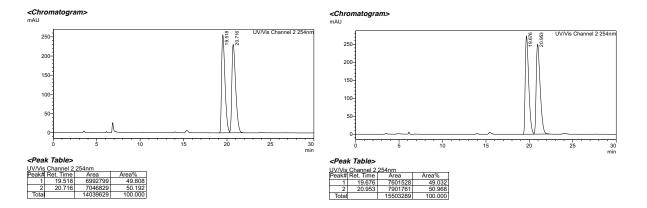
(*R*)-4-(4-phenylbuta-2,3-dien-1-yl)morpholine (2an): In an N₂-filled glovebox, to a dry 2-dram vial equipped with a magnetic stirring rod was added [(L3)Pd(η^3 -allyl)]BAr₄^F (6.7 mg, 4.0 μ mol, 2.0 mol %) and dissolved in Et₂O (0.50 mL). To this solution was added 10 (42.5 μ L, 0.240 mmol, 1.20 equiv). At this point, the vial was capped with a PTFE lined cap and removed from the glovebox. The vial was allowed to cool in an ice bath (0 °C) for *ca*. 10 min before Et₃N (56 μ L, 0.40 mmol, 2.0 equiv) was added followed by morpholine (17.5 μ L, 0.200 mmol, 1.00 equiv). The reaction mixture was allowed to stir at 0–4 °C for 3 h and then passed through a plug of silica gel eluting with 1:1 hexanes:EtOAc with 2 % v/v Et₃N addition (*ca*. 20 mL) and concentrated under reduced pressure. Purification by flash silica gel chromatography (70:30 hexanes:EtOAc with 2% v/v Et₃N additive) afforded **2an** as a colorless semi-solid (18.8 mg, 0.0873 mmol, 43.7 %). HPLC analysis revealed this material to be 91:9 er with the same major enantiomer as obtained through hydroamination of **1a**.

HPLC: Column: Cellulose-3 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 99.9:0.1 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 91:1.



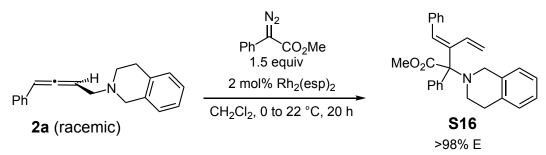
The starting material **10** was recovered from the above experiment (26.6 mg, 58.9% recovery based on 1.20 equiv used). This material was additionally analyzed by HPLC and determined to be racemic.

HPLC: Column: Cellulose-3 (3 μm, 4.6 mm X 250 mm). Mobile phase: 99.9:0.1 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm.

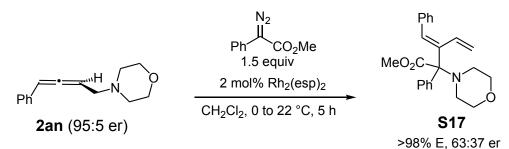


HPLC traces above show starting material allenyl acetate **10** (left) and **10** recovered from above allylic substitution reaction with morpholine (right). The recovered **10** is still a racemate indicating that kinetic resolution does not take place.

XIII. Derivatization of Allene Products

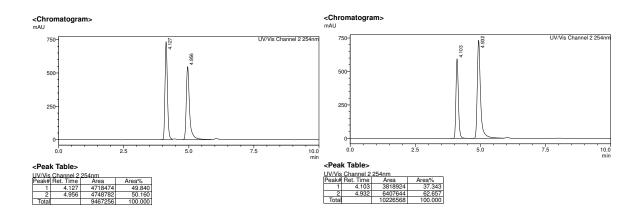


methyl (E)-3-benzylidene-2-(3,4-dihydroisoquinolin-2(1H)-yl)-2-phenylpent-4-enoate (S16): To an oven-dried 2-dram vial equipped with a magnetic stirring rod were added rac-2a (52.3 mg, 0.200 mmol, 1.00 equiv) and Rh₂(esp)₂ (3.0 mg, 4.0 µmol, 2.0 mol %). CH₂Cl₂ (0.50 mL) was added and the resulting solution was allowed to cool to 0 °C. Methyl 2-diazo-2-phenylacetate (53 mg, 0.30 mmol, 1.5 equiv) was added as a solution in CH_2Cl_2 (0.50 mL) dropwise via syringe. The reaction mixture was allowed to warm gradually to ambient temperature and then stir for 20 h. The reaction mixture was then passed through a short plug of neutral alumina, eluting with 1:1 hexanes: EtOAc (ca. 20 mL) and the solution was concentrated. ¹H NMR analysis of the unpurified reaction mixture indicated that the product was >98% E. The product was then purified by flash silica gel chromatography (90:10 hexanes: EtOAc) to afford **S16** as a white solid (68.1 mg, 0.166 mmol, 83.2% yield). IR (neat, cm⁻¹) 3021 (w), 2947 (w), 2829 (w), 2246 (w), 1720 (s), 1599 (w), 1492 (m), 1445 (m), 1429 (m), 1382 (w), 1226 (s), 1175 (s), 1077 (m), 1035 (m), 1010 (m), 906 (s); ¹**H** NMR (400 MHz, CDCl₃) δ 7.65–7.58 (2H, m), 7.37–7.08 (12H, m), 6.96 (1H, d, J = 7.3 Hz), 6.41 (1H, dd, J = 18.0, 11.8 Hz), 5.05 (1H, dd, J = 18.0, 1.5 Hz), 5.01 (1H, dd, J = 12.0, 1.5 Hz), 3.83 (3H, s), 3.80 (1H, AB_q, $J_{AB} = 15.0$ Hz), 3.73 (1H, AB_q, $J_{AB} = 15.0$ Hz), 3.14–2.89 (3H, m), 2.76–2.68 (1H, m); ¹³C NMR (125 MHz, CDCl₃) δ 169.5, 139.2, 138.6, 137.4, 135.9, 134.8, 133.3, 129.8, 129.7, 129.3, 128.8, 128.1, 127.5, 127.1, 127.0, 126.7, 126.1, 125.6, 119.3, 79.1, 51.3, 51.2, 47.2, 30.3; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₈H₂₇NO₂: 410.2115, found: 410.2120; $MP = 40-44 \ ^{\circ}C.$

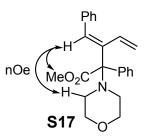


methyl (*E*)-3-benzylidene-2-morpholino-2-phenylpent-4-enoate (S17): To an oven-dried 2dram vial equipped with a magnetic stirring rod were added (*R*)-2an (32.3 mg, 0.150 mmol, 1.00 equiv, 95:5 er) and Rh₂(esp)₂ (2.3 mg, 3.0 μ mol 1, 2.0 mol %). CH₂Cl₂ (0.35 mL) was added and the resulting solution was allowed to cool to 0 °C. Methyl 2-diazo-2-phenylacetate (40 mg, 0.23 mmol, 1.5 equiv) was added as a solution in CH₂Cl₂ (0.40 mL) dropwise via syringe. The reaction mixture was allowed to warm gradually to ambient temperature and then stir for 5 h. The reaction mixture was then passed through a short plug of neutral alumina, eluting with 1:1 hexanes:EtOAc (*ca.* 20 mL) and the solution was concentrate. ¹H NMR analysis of the unpurified reaction mixture indicated that the product was >98% *E*. The product was then purified by flash silica gel chromatography (90:10 hexanes:EtOAc) to afford **S17** as a colorless viscous oil (46.1 mg, 0.127 mmol, 84.5% yield). **IR** (neat, cm⁻¹) 3021 (w), 2951 (m), 2889 (w), 2848 (m), 2246 (w), 1721 (s), 1599 (w), 1492 (m), 1445 (m), 1432 (w), 1390 (w), 1291 (w), 1266 (m), 1227 (s), 1193 (s), 1113 (s), 1070 (w), 1008 (m), 908 (s); ¹**H NMR** (400 MHz, CDCl₃) 7.56–7.49 (2H, m), 7.34–7.17 (8H, m), 7.04 (1H, s), 6.36 (1H, dd, J = 18.0, 11.9 Hz), 5.05 (1H, d, J = 17.9 Hz), 4.99 (1H, d, J = 11.8 Hz), 3.80 (3H, s), 3.80–3.73 (4H, m), 2.76–2.60 (2H, m), 2.57–2.42 (2H, m); δ ¹³**C NMR** (125 MHz, CDCl₃) δ 169.5, 138.3, 138.0, 137.2, 133.3, 130.6, 129.6, 129.3, 128.1, 127.5, 127.2, 127.1, 119.1, 78.8, 67.8, 51.2, 49.5; **HRMS** (ESI⁺) [M+H]⁺ calc'd for C₂₃H₂₅NO₃: 364.1907, found: 364.1914.

HPLC: Column: Amylose-1 (3 μ m, 4.6 mm X 250 mm). Mobile phase: 90:10 hexanes:*i*-PrOH, 1 mL/min. Detection wavelength: 254 nm. Er = 62.5:37.5 er



The olefin stereochemistry of **S17** was confirmed by NOESY analysis. Key nOe signals are depicted below:



Efforts to realize a stereoconvergent reaction starting from racemic **2an** with a chiral catalyst were largely unsuccessful. The relevant data are presented in Table S8.

 N_2 1.5 equiv Ph Ph CO₂Me 1 mol% [Rh] MeO₂C Ph Ρh 2an (racemic) solvent, 0-22 °C, 5-9 h S17 solvent time (h) yield 7b (%)^b entry [Rh] erc 1 Rh₂(S-PTAD)₄ CH₂Cl₂ 9 (10)ND 2 Rh₂(R-BTPCP)₄ CH_2CI_2 9 <2 _ 3 Rh₂(S-tert-PTTL)₄ CH_2CI_2 9 34 51:49 4 Rh₂(R-DOSP)₄ CH_2CI_2 9 72 53:47

pentane

Table S8. Screening of Chiral Rh-Based Catalysts with Racemic 2an^a

Rh₂(R-DOSP)₄

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^aReactions run with 0.20 mmol **2an** in 1.0 mL solvent. ^bIsolated yield of pure **S17**. Yields in parenthesis are conversions based on ¹H NMR of unpurified reaction mixture. ^cDetermined by HPLC analysis of pure **S17** in comparison with an authentic racemic standard. ND = not determined.

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65.5:34.5

XIV. References

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