

Reporting Summary

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Statistical parameters

When statistical analyses are reported, confirm that the following items are present in the relevant location (e.g. figure legend, table legend, main text, or Methods section).

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- An indication of whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistics including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated
- Clearly defined error bars
State explicitly what error bars represent (e.g. SD, SE, CI)

Our web collection on [statistics for biologists](#) may be useful.

Software and code

Policy information about [availability of computer code](#)

Data collection

X-ray datasets were collected at the X06SA (PXI) beamline of the Swiss Light Source (Paul Scherrer Institute, Villigen, Switzerland) using an EIGER 16M detector (Dectris).

Data analysis

Structure determination:

Datasets were indexed and integrated with XDS and merged using BLEND of the CCP4 program suite. Scaling and averaging of symmetry-related intensities were performed by aP_scale with truncation of the data at the best high-resolution along h, k or l axis determined by AIMLESS. The STARANISO software (<http://staraniso.globalphasing.org/>) was applied to account for the anisotropy of the data. The SAD method was applied using the CRANK2 pipeline running with SHELX/D, REFMAC5, Parrot and Buccaneer in the CCP4 program suite. To obtain a final structure, iterative refinement and model building were performed by phenix.refine and Coot, respectively. The structure of bMCT with bound L-lactate was obtained by molecular replacement with Phaser using the bMCT structure with bound TSA as search model. The final structures were obtained after multiple rounds of model building with Coot and refinement with phenix.refine. For all the refinements, XYZ coordinates, individual B-factors, occupancies and TLS strategies were applied. The TLS groups were automatically assigned using Phenix. Figures representing structural information were prepared using Chimera or PyMol. Van der Waals pore radii were computed using HOLE. Electrostatic surface potentials were calculated by the Adaptive Poisson-Boltzmann Solver (APBS).

Analysis of transport assays:

Data of transport assays were analyzed using Prism 6.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers upon request. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Atomic models have been deposited in the Protein Data Bank under accession numbers 6G9X and 6HCL.

Field-specific reporting

Please select the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/authors/policies/ReportingSummary-flat.pdf](https://www.nature.com/authors/policies/ReportingSummary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size For transport studies, presented data originate from 3 independent experiments, each performed in triplicates.

Data exclusions In transport studies, no data was excluded from the analysis.

Replication Transport experiments were independently repeated 3 times.
For X-ray crystallography experiments more >1000 crystals were measured.

Randomization Randomization is not necessary and was thus not applied.

Blinding Blinding was not applied because it is not necessary in the field of X-ray crystallography.

Reporting for specific materials, systems and methods

Materials & experimental systems

n/a	Involvement	Included in study
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Unique biological materials
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Antibodies
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Palaeontology
<input type="checkbox"/>	<input checked="" type="checkbox"/>	Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Human research participants

Methods

n/a	Involvement	Included in study
<input checked="" type="checkbox"/>	<input type="checkbox"/>	ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/>	Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/>	MRI-based neuroimaging

Antibodies

Antibodies used: Commercial anti-pentaHis (Qiagen, catalogue number 34660) and goat anti-mouse IgG (H+L) HRP conjugate antibody (Biorad, catalogue number 172-1011).

Validation: Commercial antibodies

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals: Escherichia coli BL21(DE3) pLysS, Escherichia coli JA202

Wild animals: The study did not involve wild animals.

Field-collected samples: The study did not involve any field-collected samples.