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Supplemental Information

Transcriptional States and Chromatin Accessibility

Underlying Human Erythropoiesis

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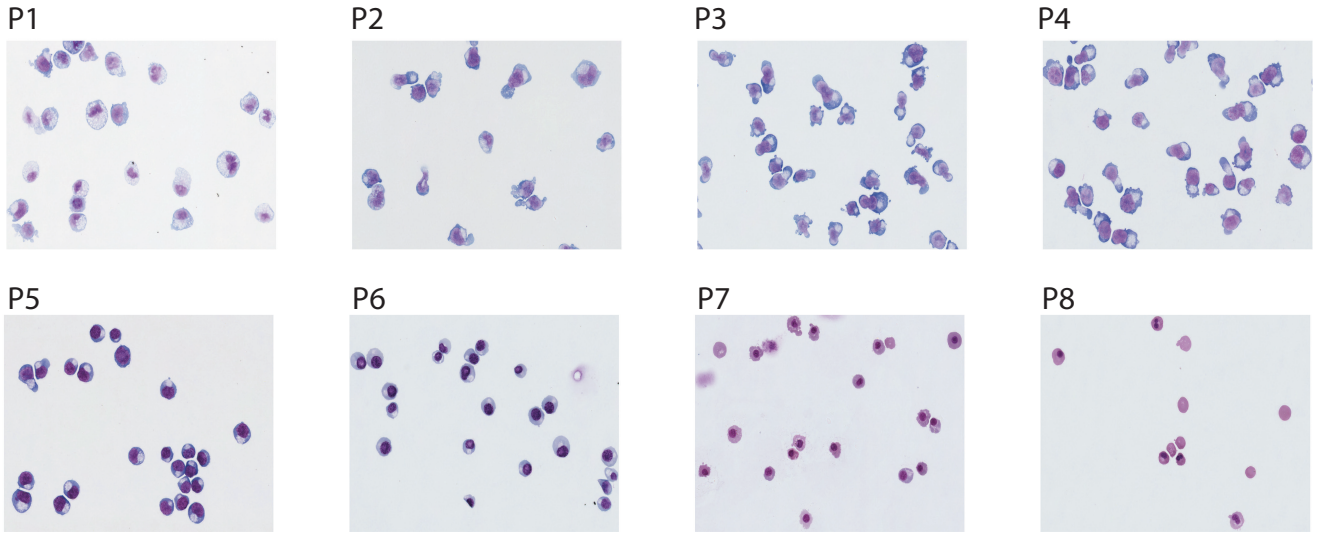
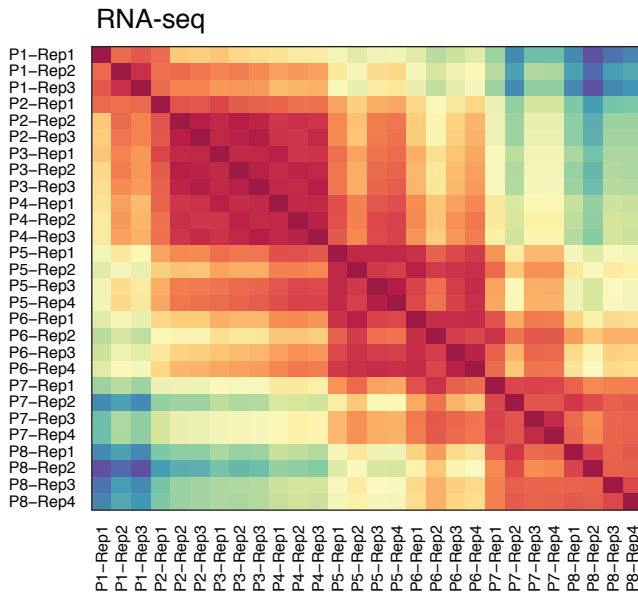
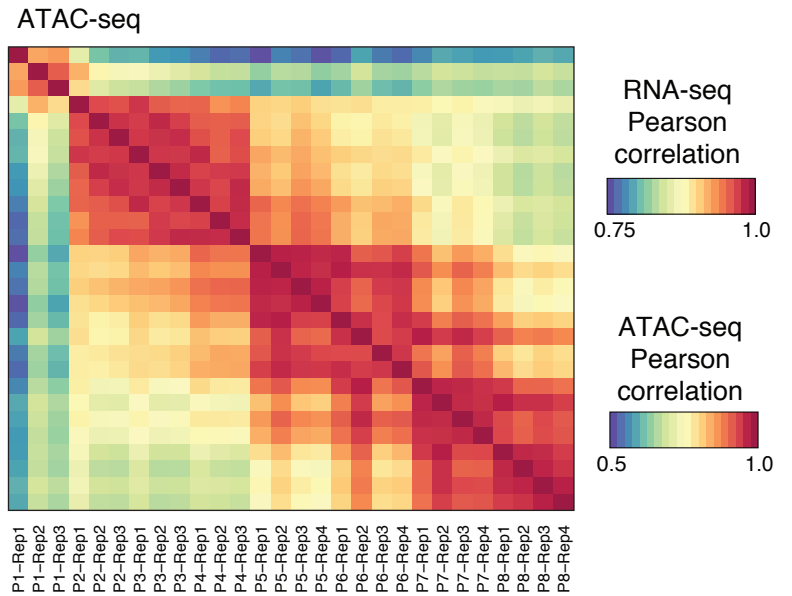
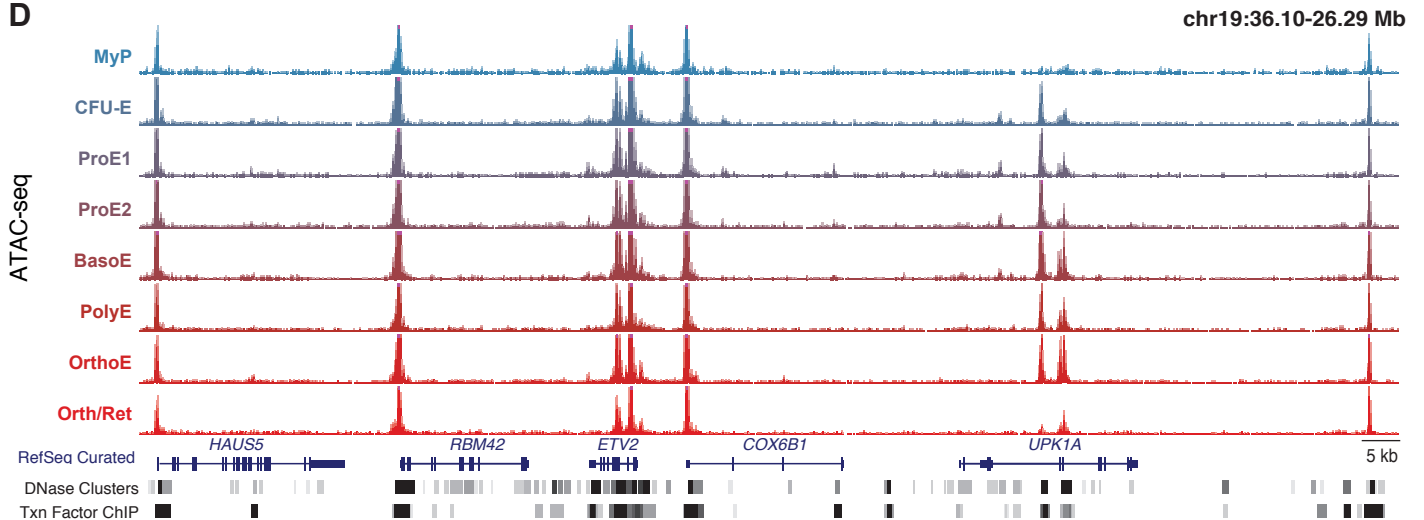
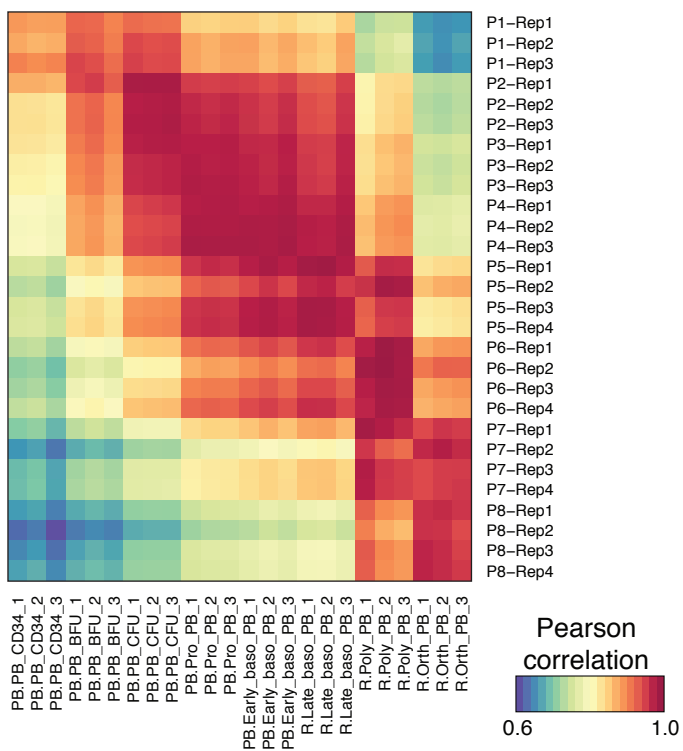
A**B****C****D**

Figure S1, related to Figure 1. A time course of human erythroid differentiation. (A) Images of MayGrunwald stained cytopspins of sorted populations P1-P8 (**Figure 1B**, 63x magnification). (**B,C**) Heatmap showing the correlation across individual populations and replicates of (**B**) RNA-seq and (**C**) ATAC-seq libraries generated in this study. Color bar: Pearson correlation. (**D**) *ETV2* locus with corresponding ATAC-seq data across indicated populations show high quality chromatin accessibility profiles at known promoter and gene regulatory regions.

A



B

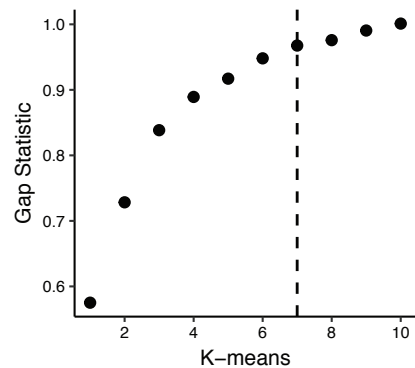
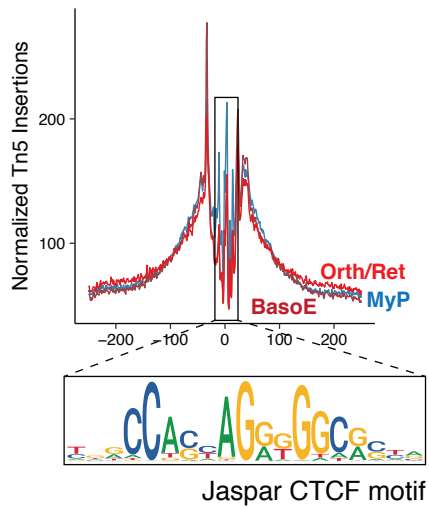


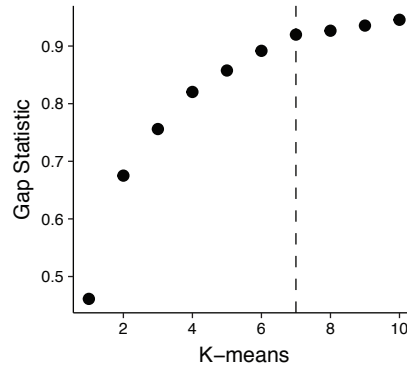
Figure S2, related to Figure 2. Transcriptomic landscape of human erythropoiesis.

(A) Heatmap showing the correlation across individual replicates of RNA-seq libraries generated in this study (y-axis) compared to previously published reports (x-axis). Color bar: Pearson correlation. **(B)** Gap statistic associated with the choice of $k=7$ clusters for the RNA-seq data.

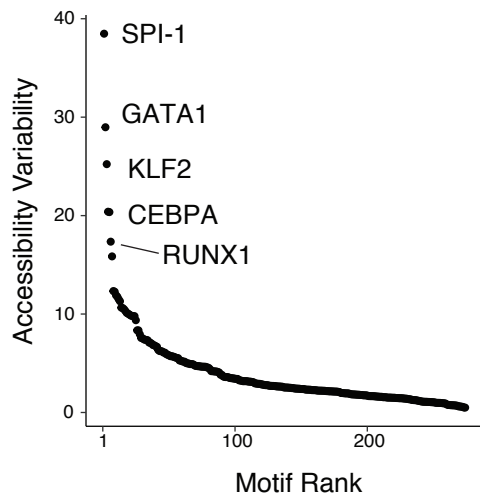
A



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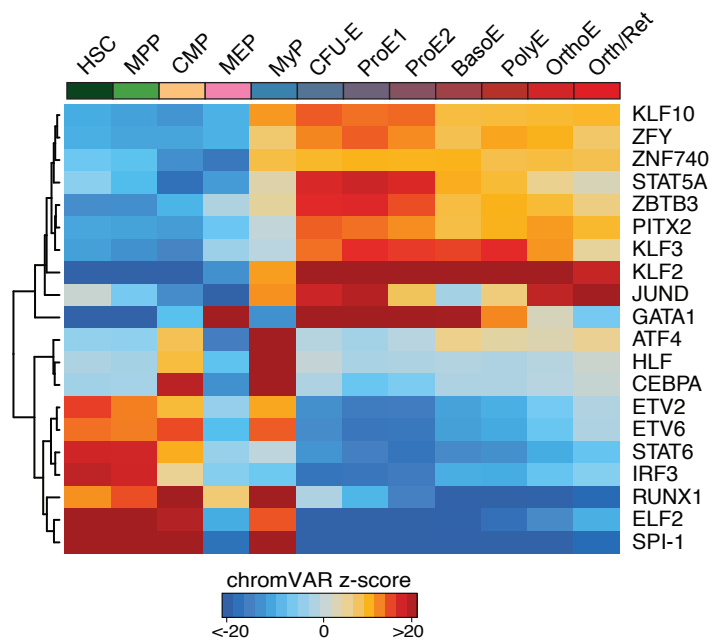


Figure S3, related to Figure 3. Open chromatin landscape of human erythropoiesis.

(A) ATAC-seq footprinting plot, showing that Tn5 insertion density near the CTCF motif is relatively constant throughout erythroid differentiation. Normalized insertions of MyP, BasoE and Orth/Ret are shown. **(B)** Gap statistic associated with the choice of $k=7$ clusters for the ATAC-seq peaks. **(C)** Rank order plot of transcription factor binding sites with greatest variability in chromatin accessibility across sampled cell populations as also indicated in panel D. **(D)** Heatmap showing temporal changes in chromatin accessibility for the top 20 TFs with greatest accessibility variability between populations. Color bar: chromVAR deviation z-score.

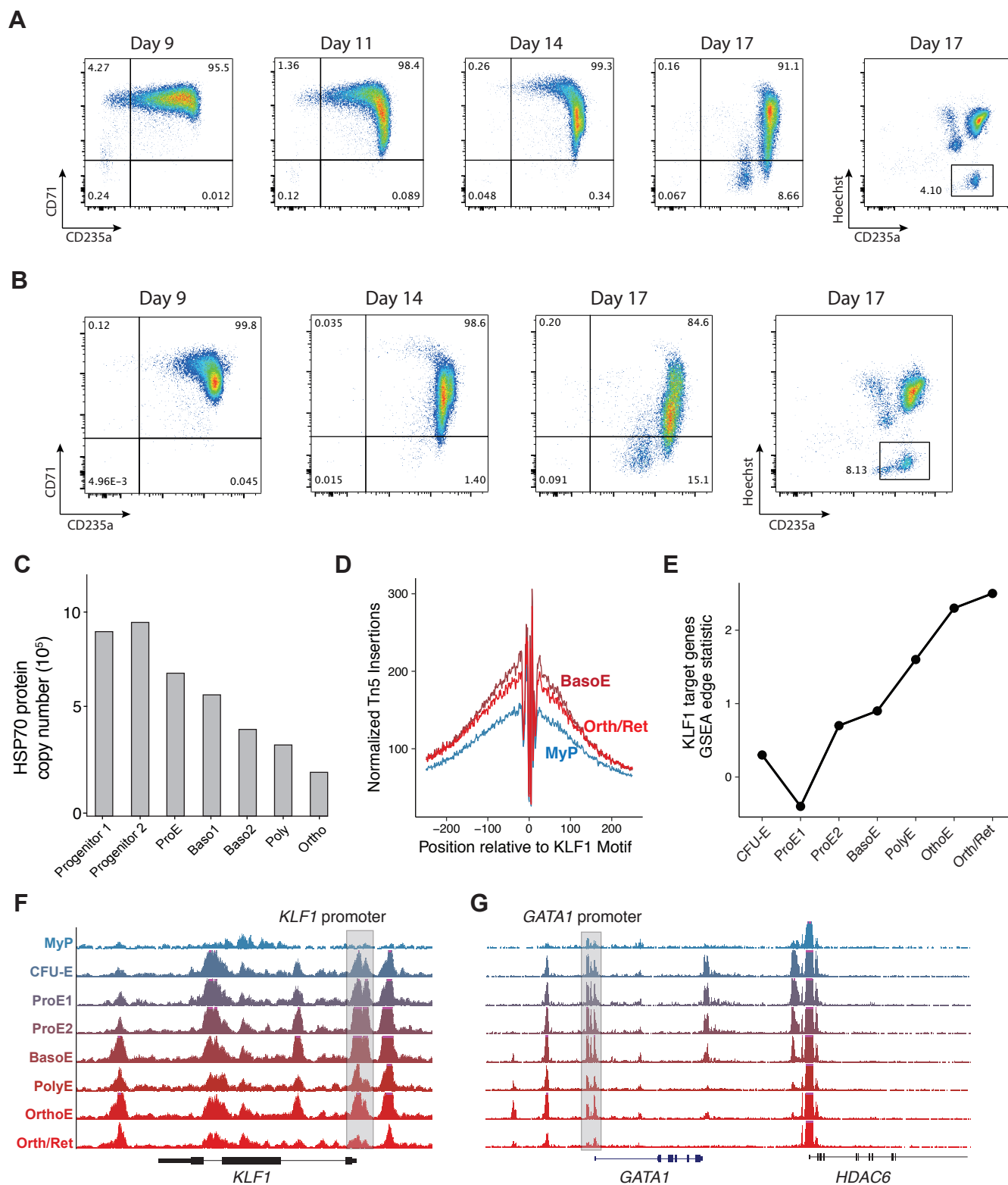


Figure S4, related to Figure 4. Regulatory dynamics of erythroid transcription factors. (A, B) Flow cytometry plots assessing erythroid differentiation using surface markers CD71 and CD235 at indicated time points of *in vitro* culture of primary hematopoietic cells. Hoechst staining was used to measure enucleation frequency. Plots from two cultures are shown. Samples shown in (A) correspond to the western blot shown in Figure 4C. Samples shown in (B) correspond to the western blot shown in Figure 4D. (C) Protein copy number for HSP70 across erythroid differentiation. (D) ATAC-seq footprinting plot, showing that Tn5 insertion density near the KLF1 motif is most “active” in BasoE, but remains high in Ortho/Ret. (E) *KLF1* transcriptional activity using GSEA edge statistics across differential gene expressions across indicated populations. (F,G) ATAC-seq peaks are shown in the *KLF1* and *GATA1* loci across indicated populations. Shaded regions highlight differential peaks in the promoter regions of both genes.

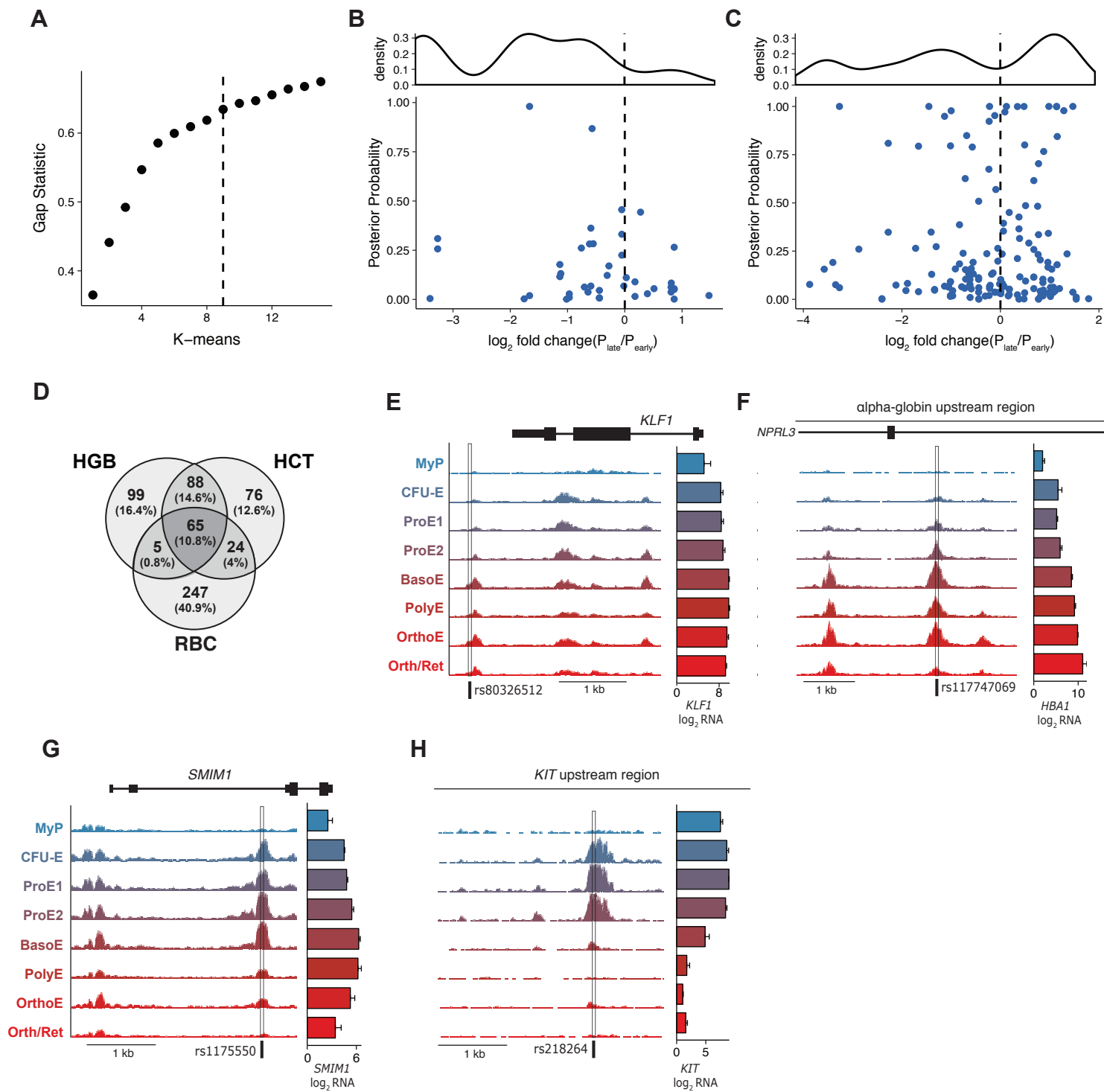


Figure S5, related to Figure 5. *Cis*-regulatory variation and dynamics in human erythropoiesis. (A) Gap statistic associated with the choice of $k=9$ clusters for the ATAC-seq peaks overlapping fine-mapped genetic variants with $PP > 0.10$. Fine-mapped GWAS variants associated with (B) HGB/HCT and (C) MCV/MCH/MCHC/RBC, plotted by posterior probability of causal association vs. \log_2 fold-change in ATAC-seq counts per million from CFU-E to ProE2 (P_{early}) to BasoE/PolyE (P_{late}). Density plots of variants are shown above, weighted by $PP \cdot \log_2$ fold-change. (D) Venn diagram showing the overlap of fine mapped variants associated with HGB, HCT and RBC with $PP > 0.10$. (E-H) ATAC-seq peaks are shown at indicated loci across indicated populations. Boxes indicate open chromatin regions overlapping with erythroid-trait associated variants. These open chromatin regions show different accessibility dynamics throughout differentiation, suggesting potential windows of active gene regulation. Bar graphs indicate mean (\pm SEM) \log_2 counts-per-million RNA-seq reads per-population.

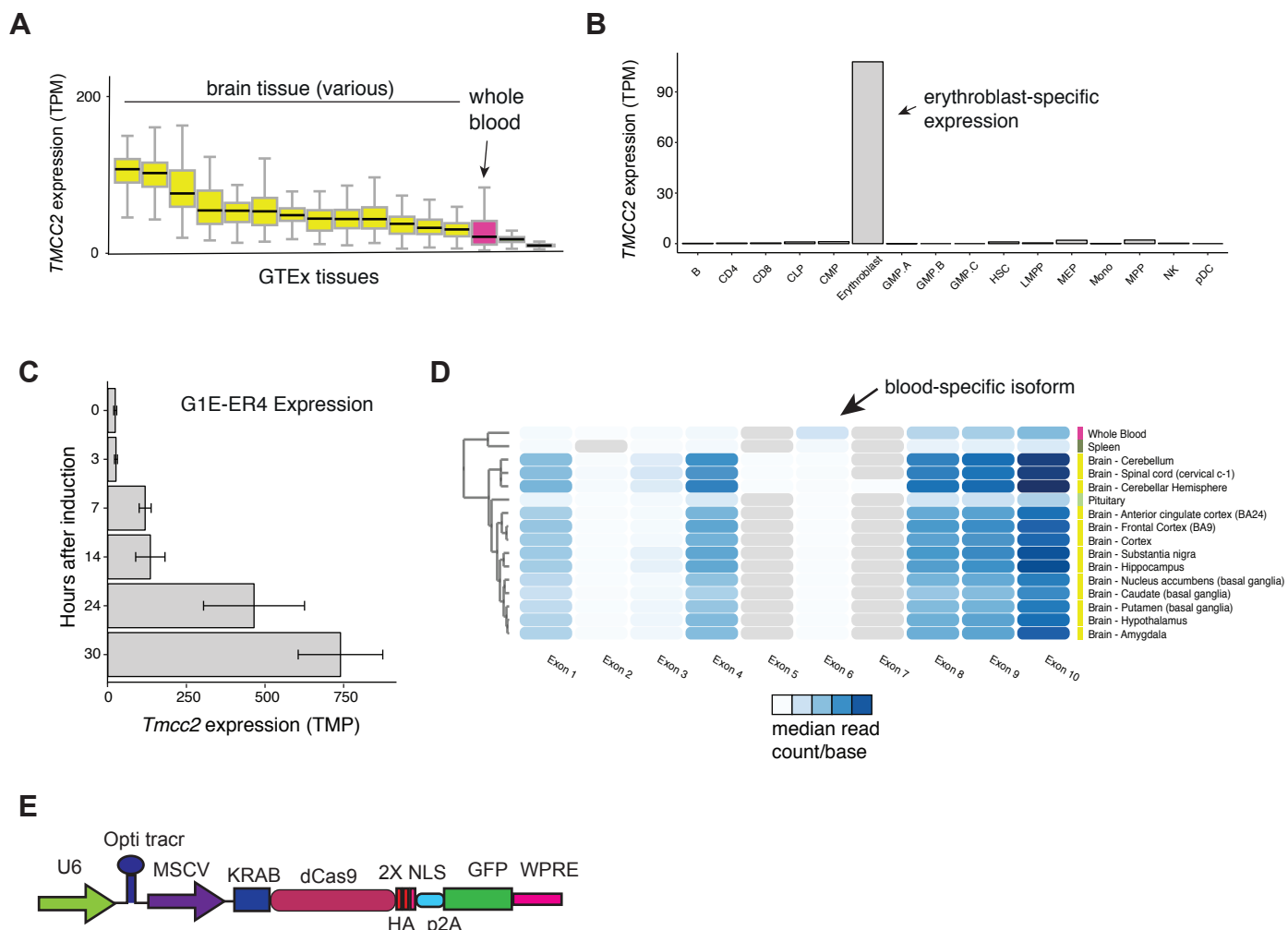


Figure S6, related to Figure 6. Erythroid-specific isoform expression and regulation of *TMCC2*. (A) *TMCC2* shows high transcript expression in the human brain and whole blood. Center line: median; box limits: upper and lower quartiles; whisker: 1.5x interquartile range. (B) *TMCC2* is highly expressed in erythroblasts among displayed hematopoietic cell types. (C) *Tmcc2* is strongly induced in differentiating G1E-ER4 cells. Error bars represent +/- 1 standard deviation between replicates. (D) *TMCC2* shows a blood-specific isoform including exon 6, which is absent in different regions of the human brain. Color bar: Median read count per base. (E) Schematic of the lentiviral vector construct expressing a guide RNA and the KRAB domain fused to dCas9.

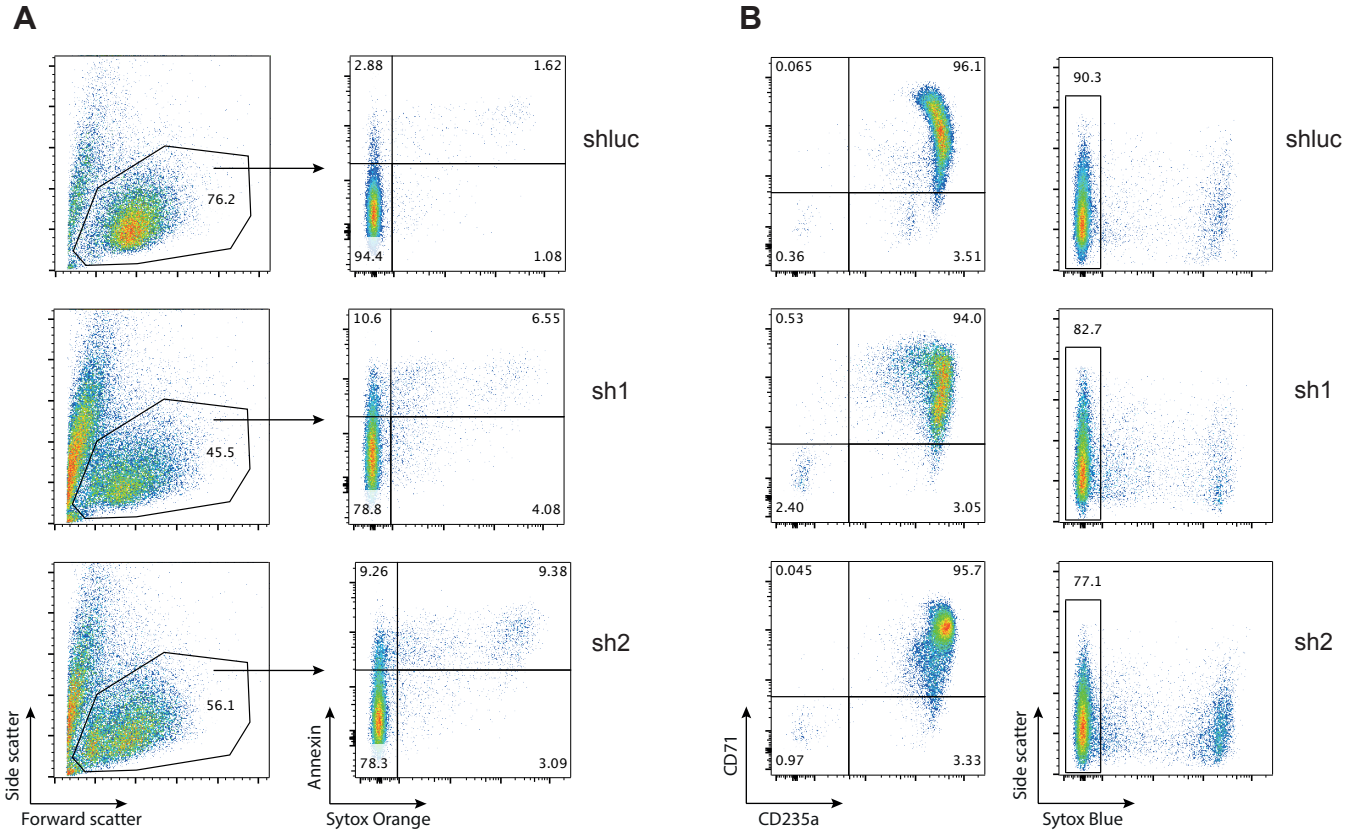


Figure S7, related to Figure 7. *TMCC2* is an essential regulator of terminal human erythropoiesis. (A) Flow cytometry plots showing forward and side scatter distributions (left) and Annexin vs. Sytox Orange staining (right) in *TMCC2* knockdown cells (sh1, sh2) compared to control cells (shluc). (B) Flow cytometry plots showing CD71 and CD235 surface marker expression (left) and side scatter vs. Sytox Blue staining (right) in *TMCC2* knockdown cells (sh1, sh2) compared to control cells (shluc).