1 Supplementary Information for

2	Identification of metabolic vulnerabilities of receptor tyrosine
3	kinases-driven cancer
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5	Jin et al.
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15	a, Sensitivity of metabolism-targeted inhibitors in lung cancer cell lines bearing
16	indicated genetic alterations. Left, Diagram showing the inhibitors used for metabolic
17	pathways intervention. Right, Bar plot obtained by the unbiased hierarchical cluster
18	analysis of the cell lines according to cell growth inhibition rate of individual
19	metabolic inhibitors. Cells were treated with ETO (50 μ M), CB839 (10 μ M), Oxamate
20	(30 mM), DCA (10 mM) or 2DG (2.5 mM) for 72 hr. Cell growth inhibition rate was
21	measured using the CCK8 assay b , Heatmap obtained by cluster analysis using the
22	mRNA data of metabolic genes of 740 lung adenocarcinoma patients in the TCGA
23	data sets. The columns indicate different patients bearing indicated genetic alterations,
24	and the rows indicate different metabolic genes. c, Immunoblotting analysis. Cells
25	were treated with indicated RTK inhibitors (100 nM) for 1 hr before being subjected
26	to immunoblotting analysis using indicated antibodies. d, IL3 dependence analysis.
27	Cell growth fold changes with or without IL3 were plotted by counting cell numbers.
28	Data were means of triplicates; error bars represented SD. e, Cell sensitivity to RTK
29	inhibition. Cells were treated with AZD4547 at indicated concentrations for 72 hr and
30	cell viability was analyzed using CCK8 assay. Data were means of triplicates; error

31	bars represented SD. f, Oxygen consumption rate (OCR) and extracellular
32	acidification rate (ECAR) measurement using Seahorse XF96 analyzer. g. The impact
33	of IL3 on OCR. BAF3-RTK or the parental BAF3 cells were cultured in the presence
34	or absence of IL3. OCR were measured using Seahorse XF96 analyzer. Data in f, g
35	were means of five replicates; error bars represented SD. h, Metabolite set enrichment
36	analysis. The metabolome view showing the representative metabolic pathways
37	arranged by $p < 0.05$ (from pathway enrichment analysis) on Y-axis, and pathway
38	impact > 0.1 (from pathway topology analysis) on X-axis, carried out with
39	MetaboAnalyst 4.0. The node color was based on its p value and the node radius was
40	determined by its pathway impact value. The altered metabolite sets were arranged
41	according to 1.5-fold cutoff and $P < 0.01$ in relative to parental BAF3 cells. i, Time
42	courses of ¹³ C labeled intensities of metabolites from [U- ¹³ C ₆]-glucose flux analysis
43	measured by GC/MS. BAF3 and BAF3-RTK cells were cultured in the presence of
44	$[U^{-13}C_6]$ -glucose for 1, 3, 6 or 12 hr. The labeling curves were plotted by measuring
45	fractional ¹³ C enrichment at 1, 3, 6 and 12 hr respectively. j-k , ¹³ C labeled intensities
46	of metabolites from $[U^{-13}C_5]$ -glutamine or $[U^{-13}C_{16}]$ -palmitate. BAF3 and

47	BAF3-RTK cells were cultured for 24 hr in the presence of either $[U^{-13}C_5]$ -glutamine
48	or $[U^{-13}C_{16}]$ -palmitate. The intermediate metabolites in TCA cycle labeled with ^{13}C
49	isotopologue were measured by GC/MS. Data in j-k were means of triplicates; error
50	bars represented SEM. I, Glucose/glutamine dependency analysis. Cells were cultured
51	with indicated concentrations of glucose (GLC)/glutamine (GLN) for 4 days. Images
52	were acquired every 6 hr by automated real-time assessment using IncuCyte ZOOM.
53	Growth curves were plotted as the change in confluence percentage. Data were means
54	of six replicates; error bars represented SD. m-o, Cell sensitivity to targeted
55	metabolism inhibition. Cells were treated with indicated inhibitors for 72 hr. The
56	viability of BAF3 and BAF3-RTK cells were analyzed using CCK8 assay, and the
57	viability of cancer cells were analyzed using the SRB assay. The assays were
58	performed in biological triplicates, and error bars represented SD. p, Transcriptome
59	analysis. Heatmap of transcriptome profiling representing the mRNA levels of genes
60	performed by RNA-seq. The rows indicate different genes, and the columns indicate
61	different cells (n = 3 per cell line). q, KEGG pathway enrichment analysis the
62	metabolic genes between EGFR- and FGFR-activated tumors that displayed in b . The

63	significantly enriched metabolism-related KEGG pathways ($P < 0.05$) were presented.
64	For each KEGG pathway, the bar shows the enrichment score of the pathway
65	according to p value. Unless otherwise stated, BAF3 parental cells were cultured with
66	IL3 and BAF3-RTK cells were cultured without IL3. For all bar graphs, $***p < 0.001$,
67	$p^{**} = 0.01$, $p^{*} = 0.05$, n.s. ≥ 0.05 for Student's pairwise t test. Source data are provided
68	as a Source Data file.

70 Supplementary Figure 2





74	a, Cell sensitivity to PHGDH inhibition. Cell viability was measured using the CCK8
75	assay following the treatment with NCT503 (20 μM) for 72 hr. BAF3-EGFR/FGFR1
76	cells were cultured without IL3. The assays were performed in biological triplicates,
77	and error bars represented SD. b, Sensitivity of a panel of cancer cells to PHGDH
78	inhibition. Cancer cells with indicated genetic alterations were treated with CBR5884
79	at 6.25, 12.5 or 25 μM for 6 days and the inhibition rate of cell growth was
80	determined relative to untreated control. Data were means of duplicates and error bars
81	represented SD. c, d, Body weight change of PC9 xenograft and LU-01-0251 PDX
82	models. Mice were dosed with NCT-503 (40 mg/kg) or Gefitinib (5 mg/kg for PC9
83	and 1 mg/kg for LU-01-0251) daily for indicated days (n = 8 for PC9, n = 6 for
84	LU-01-0251). Data were means and error bars represented SEM. e, ¹³ C enrichment of
85	purine nucleotides. BAF3-EGFR/FGFR1 cells were cultured in the presence of
86	$[U^{-13}C_6]$ -glucose for 24 hr without IL3. f, Tracer scheme illustrating the flux of
87	glucose to glutathione via SSP determined by ¹³ C-labeled metabolites (Left) and the
88	¹³ C enrichment of glutathione (Right). Cells were cultured in the presence of
89	[U- ¹³ C ₆]-glucose for 24 hr, and the incorporation percentage of ¹³ C from glucose was

90	analyzed by QTOF-MS. Data were means of triplicates; error bars represented SEM.
91	g, ROS level measurement. PC9 and DMS114 cells were transfected with indicated
92	siRNAs for 72 hr. Data were means of duplicates; error bars represented SD. h,
93	Immunoblotting analysis. Cells were transfected with indicated siRNAs for 48 hr
94	before being subjected to immunoblotting using indicated antibodies. i,
95	Immunohistochemistry analysis of representative tumor tissues from NSCLC PDX
96	tumors with EGFR mutation or wildtype RTK. Shown are representative field from
97	one section per tumor tissue ($n = 3$ independent tumor tissues from each PDX model).
98	Scale bar, 20 μ m. j, Cell viability assay. DMS114 and RT112 cells were transfected
99	with indicated siRNAs for 72 hr and cell viability was analyzed by counting cell
100	numbers. The assays were performed in duplicates, and error bars represented SD. \mathbf{k} ,
101	Diagram depicting the metabolic reprogramming upon EGFR activation. For all bar
102	graphs, $^{***}p < 0.001$, $^{**}p < 0.01$, $^{*}p < 0.05$, n.s. ≥ 0.05 for Student's pairwise t test.
103	Source data are provided as a Source Data file.







109	a , Time courses of ¹³ C labeled intensities of metabolites from $[U-^{13}C_3]$ -lactate flux
110	analysis measured by GC/MS. The labeling curves were plotted by measuring
111	fractional ¹³ C enrichment at 0.5, 6 and 24 hr respectively. BAF3-EGFR/FGFR1 cells
112	were cultured without IL3. The assays were performed in biological triplicates, and
113	error bars represented SD. b, Serum fractional enrichment of lactate. Mice with flank
114	xenografts of H1581 cells were co-injected with [U- ¹³ C ₆]-glucose and [3- ¹³ C]-lactate
115	intravenously and the serum was collected at 30 min ($n = 6$ mice per group). Data
116	were means and error bars represented SEM. c, Left, OCR measurement. OCR was
117	measured following the treatment with Oxamate (10 mM, 6 hr), GSK2837808A (20
118	μ M, 6 hr) or Gefitinib (100 nM, 24 hr) using Seahorse XF96 analyzer. Right, OCR
119	change fold. The OCR values were normalized by the control group without treatment
120	(CON). The assays were performed in biological four replicates, and error bars
121	represented SD. d, Immunohistochemistry analysis of representative tumor tissues
122	from NSCLC PDX models with FGFR gene alteration or wildtype RTK. Shown are
123	representative fields from one section per tumor tissue ($n = 3$ independent tumor
124	tissues from each PDX model). Scale bar, 20 µm. e, Dependency score for metabolic

125	genes in cell lines with (Amp) or without (WT) FGFR amplification. Data were
126	extracted from public dataset Project Achilles. $*p < 0.05$ was considered to be
127	statistically significant. f, Left: tumor growth curve of NCI-H1581 xenograft model.
128	Mice bearing NCI-H1581 tumors were treated with Oxamate (750 mg/kg) and
129	AZD4547 (2.5 mg/kg) daily for 21 days (n = 5). Growth curve was plotted by
130	measuring the tumor volume three times per week. Right: Body weight change of
131	NCI-H1581 and SNU16 xenograft models. Data were means and error bars
132	represented SEM. g, Body weight of LU6429 PDX model. Oxamate (750 mg/kg),
133	Metformin (250 mg/kg), AZD4547 (10 mg/kg) or indicated combinations were given
134	daily (n = 6 except for Oxamate-treated group n = 7). Data were means and error bars
135	represented SEM. h, Tumor growth curve, grouped scatter plot of individual mice
136	relative tumor volume on Day 24 and body weight change of LU0743 PDX model.
137	Mice were treated with Oxamate (750 mg/kg), AZD4547 (5 mg/kg) or indicated
138	combination daily for 32 days ($n = 6$). Growth curve was plotted by measuring the
139	tumor volume three times per week. Data were means and error bars represented SEM.
140	i, Diagram depicting the metabolic reprogramming upon FGFR activation. j-k, Tumor

141	growth curve, grouped scatter plot of individual mice relative tumor volume at the
142	endpoint, and body weight change of A431 xenograft and three PDX models. Mice
143	bearing indicated tumors were treated with Oxamate (750 mg/kg) or NCT503 (40
144	mg/kg) daily for indicated days (n = 10 for A431, n = 5 for LU2071, n = 6 for
145	LU-01-0393 and LU-01-0416). Data were means and error bars represented SEM. For
146	all bar graphs, *** $p < 0.001$, ** $p < 0.01$, $p < 0.05$, n.s. ≥ 0.05 for Student's pairwise t
147	test. Source data are provided as a Source Data file.
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154 a, RT-qPCR transcript analysis. RT112 and PC9 cells were transfected with indicated siRNA for 48 hr and mRNA level of indicated genes was measured by RT-qPCR. 155 156 Data were means of triplicate wells; error bars represented SD. b, Analysis of glucose-derived serine and glycine in the medium of PC9 cells. Cells were transfected 157 with indicated siRNA for 48 hr followed by 24 hr-culture in the presence of 158 $[U^{-13}C_6]$ -glucose. Serine and glycine were measured by GC/MS. Gefitinib treatment 159 160 (100 nM, 24 hr) was used as a positive control. c, Analysis of glucose-derived purine nucleosides in PC9 cells. Cells were treated as in b and purine nucleoside was 161 measured by QTOF-MS. Data in b-c were means of triplicates; error bars represented 162 SEM. For all bar graphs, $^{***}p < 0.001$, $^{**}p < 0.01$, $^{*}p < 0.05$, n.s. ≥ 0.05 for Student's 163 pairwise t test. Source data are provided as a Source Data file. 164