

Assessing the Completeness of Reporting in Preclinical Oncolytic Virus Therapy Studies

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Irreproducibility of preclinical findings could be a significant barrier to the "bench-to-bedside" development of oncolytic viruses (OVs). A contributing factor is the incomplete and non-transparent reporting of study methodology and design. Using the NIH Principles and Guidelines for Reporting Preclinical Research, a core set of seven recommendations, we evaluated the completeness of reporting of preclinical OV studies. We also developed an evidence map identifying the current trends in OV research. A systematic search of MEDLINE and Embase identified all relevant articles published over an 18 month period. We screened 1,554 articles, and 236 met our a priori-defined inclusion criteria. Adenovirus (43%) was the most commonly used viral platform. Frequently investigated cancers included colorectal (14%), skin (12%), and breast (11%). Xenograft implantation (61%) in mice (96%) was the most common animal model. The use of preclinical reporting guidelines was listed in 0.4% of articles. Biological and technical replicates were completely reported in 1% of studies, statistics in 49%, randomization in 1%, blinding in 2%, sample size estimation in 0%, and inclusion/exclusion criteria in 0%. Overall, completeness of reporting in the preclinical OV therapy literature is poor. This may hinder efforts to interpret, replicate, and ultimately translate promising preclinical OV findings.

INTRODUCTION

Incomplete and non-transparent reporting of animal experiments¹ contributes to the irreproducibility of basic science studies.^{2,3} Such irreproducibility may help explain high attrition rates in drug development.⁴ To improve the level of reporting rigor and improve reproducibility, the NIH has identified essential items to be reported for all preclinical experimental animal research.⁵ This core set of preclinical reporting guidelines (NIH-PRG) lists items that should, at a minimum, be included in a preclinical publication.⁶ To properly evaluate, interpret, and reproduce an experiment's findings, readers need a

clear understanding and appreciation for how the experiment was conceived, executed, and analyzed. This information is necessary to judge the validity of findings. Incomplete and poor reporting does not allow for an accurate appraisal of the experiment's value, and important findings (positive or negative) could be missed. As demonstrated in clinical research, the quality and rigor of research improves when the reporting of key methodological items improves.^{7,8}

Cancer immunotherapy is a rapidly growing field.⁹ Oncolytic, or "cancer-killing," viruses (OVs) comprise a promising therapeutic platform that can elicit anti-cancer immune responses.^{10–13} Despite the potential benefits of OV therapy and a multitude of preclinical candidates, it should be noted that there is currently only one OV approved for human use by the U.S. Food and Drug Administration and European Medicines Agency (Imlygic for advanced melanoma, approved in 2015).¹⁴ The current state of OV preclinical study design and reporting is unknown, as is whether this could explain in part the relatively small number of approved candidates, despite the first reported studies of the modern OV era being published nearly 30 years ago.¹⁵

Our primary objectives were to review and assess the rigor in the design and completeness of reporting of preclinical *in vivo* OV therapy studies by applying the recommendations of the NIH-PRG. Furthermore, our study sought to produce an evidence map highlighting the recent global state of OV research and identify commonly studied viruses, cancers, and animal models. To address this, we used rigorous methods commonly used for systematic reviews, including



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Figure 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) Flow Diagram for Study Selection

developing an *a priori* protocol, a systematic search strategy to identify eligible articles, and screening of articles and extraction of data performed independently and in duplicate. Our appraisal of basic experimental design concepts and key reporting elements aims to increase transparency in preclinical OV research and to improve "bench-to-bedside" translation for OV immunotherapies.

RESULTS

Study Selection

Our search identified 1,554 records. Title and abstract screening excluded 1,244 records, with a subsequent full-text screening

excluding an additional 74 records. Study screening and selection are summarized in Figure 1. Two-hundred thirty-six articles were included in our review; the full list and individual data of the included studies can be found on Open Science Framework (OSF; https://osf. io/j2dwm/).

Epidemiology of Studies

Articles came from 20 different countries, based on the corresponding author's residency at the time of publication (Figure 2). Most common were the United States (n = 72, 31%) and China (n = 63, 27%). The articles were published in 2016 (n = 169,



Figure 2. Country of Publication

Information based on the corresponding author's residency at the time of the included article publication (image created using Tableau Software, Seattle, WA, USA).

72%) and the first 6 months of 2017 (n = 67, 28%) in 85 journals. The three most commonly acknowledged sources of research funding were government bodies (n = 196, 43%), academic institutions (n = 107, 24%), and foundations/charities (n = 94, 21%; total sources of funding N = 455). The most commonly reported outcome was tumor size/burden (n = 204, 32%), followed by animal survival (n = 116, 18%), viral infectivity (n = 109, 17%), and host antitumor response (n = 94, 15%; total number of reported outcomes N = 630). Seventy-six (32%) articles had titles that clearly indicated that they contained preclinical experiments. The full results for study design and publication characteristics can be found in Table S1.

In total, 26 different viral platforms were used. The most common was adenovirus (n = 104, 43%) followed by vaccinia virus (n = 34, 14%) and herpes simplex virus (n = 27, 11%; N = 241). The viral treatment most commonly used was intratumoral injection (n = 176, 68%)

of an actively replicating virus (n = 182, 77%) in a monotherapy regimen (n = 152, 59%).

Cancerous cell lines from 23 distinct types of tissue were used (N = 293). The most common were colorectal (n = 42, 14%), skin (n = 35, 12%), breast (n = 33, 11%), liver (n = 28, 10%), brain (n = 25, 9%), and lung (n = 25, 9%). This corresponds with the incidence of these cancers in humans.¹⁶ The majority of cancer models used heterotopic (n = 170, 66%; N = 259), xenograft (n = 153, 61%; N = 252), and implant (n = 235, 99%; N = 238) methods.

The animal models were based on four species, with the majority of experiments using mice (n = 231, 96%; N = 241). In total, 26,044 animals were used; 25,516 of these were mice. Explicit ethics approval for animal use was referenced in the majority of articles (n = 172, 73%). The housing and handling of animals was described in 104

articles (44%). The full results for viral, cancer, and animal model data can be found in Table S2.

Assessment of Reporting

Completeness of reporting against each of the seven NIH-PRG domains was assessed. Each section is labeled by the domains it addresses, and a brief summary of the items recommended by the NIH-PRG is provided to guide and orient the reader. The full results for our reporting assessment can be found in Figure 3 and Table S3.

The Listing of Reporting Guidelines

The use of community-based nomenclature and reporting standards are encouraged in the NIH-PRG.⁵ One article (0.4%) listed the use of reporting guidelines (Animal Research: Reporting *In Vivo* Experiments [ARRIVE])¹⁷ during study design and manuscript preparation.

Measurement Techniques

Authors are required to report on whether their findings were substantiated under a range of conditions (e.g., different viral dosages and different models).⁵ This was reported in 225 articles (95%). Furthermore, the number of subjects used and measurements performed should be sufficiently described, such that a distinction between biological and technical replicates can be made.⁵ One-hundred five articles (44%) reported the number of times a measurement was repeated for at least one outcome (e.g., tumor size was measured with calipers three times at each time point). Two articles (1%) reported the number of times a measurement was repeated for *all* experimental outcomes. Similarly, the number of animals used to measure each outcome was reported in 25 articles (17%).

Statistics and Sample Size Estimation

Statistical tests used, the exact value of n, and defined measures of central tendency and dispersion must be included.⁵ Statistical tests used were listed in 225 (95%) articles, the exact value of n in 148 (63%) articles, and measures of central tendency and dispersion in 203 (86%) and 206 (87%) articles, respectively. All four items were reported on in 116 articles (49%).

To determine an appropriate number of animals, an *a priori* power calculation is required.⁵ The primary outcome, sample size used, and the rationale for the sample size must be reported.⁵ A primary outcome was explicitly stated in 12 articles (5%). A sample size calculation was listed in 10 articles (4%); of these, 4 articles (2%) included the method of calculation (e.g., software package). No articles reported all items related to sample size estimation.

Randomization and Blinding

To reduce selection bias, the randomization of animals and the method of randomization must be reported.⁵ This is only applicable when a study includes multiple experimental arms, which occurred in 234 articles (99%). Random group assignment was stated in 103 articles (44%); of these, the method of randomization (e.g., use of a software package) was described in 2 articles (1%).

In addition, to reduce performance and selection bias, the NIH-PRG recommends that authors report whether experimenters were blinded to group assignment and outcome assessment, respectively.⁵ Blinding of personnel conducting experiments was listed in 7 articles (3%) and blinding of personnel assessing outcomes was listed in 15 articles (6%). Complete blinding (i.e., reporting of all items) was listed in 5 articles (2%).

Inclusion and Exclusion Criteria

Criteria for the exclusion of any data (e.g., animals, results) must be reported to minimize selection bias.⁵ Furthermore, authors are expected to include information regarding the total number of animals originally procured for the study.⁵ The total number of animals used was included in 7 articles (3%). The exclusion of any data, or lack thereof, was listed in 11 articles (5%). To address selective outcome reporting, the experimental design (as described in the Materials and Methods section) was compared to the reported results; 228 articles (97%) listed all the findings. Furthermore, the NIH-PRG requires any pilot or preliminary experiments, as well as any null or negative results to be reported.⁵ Pilot studies were reported in 27 articles (11%). Null or negative results were explicitly described in 96 articles (41%). No articles reported on all of the items recommended in the inclusion/exclusion criteria domain.

DISCUSSION

This review was a comprehensive appraisal of the recent preclinical *in vivo* OV immunotherapy literature. Included studies were published in nearly 100 journals and originated from 20 countries. Twenty-six different viral platforms (and hundreds of genetically modified strains) were used. Twenty-three cancer models (and hundreds of cell lines) were investigated.

Our reporting assessment indicated that basic components of experimental design, such as randomization and blinding, as well as key elements of study methodology, such as sample size estimation and *a priori*-defined inclusion/exclusion criteria, are poorly reported. Of the 21 items identified from the NIH-PRG recommendations, the vast majority were absent across included studies. The statistics domain was partially reported, with 49% of studies reporting all items. Across the other six NIH-PRG domains complete reporting was 2%, at most. Collectively, these results demonstrate that the quality of reporting in the preclinical *in vivo* OV therapy literature is poor and the publication of the NIH-PRG has not yet had a substantive influence.¹⁸

Numerous guidelines for thorough *in vivo* preclinical study design and reporting exist.¹⁹ This includes the extensive ARRIVE preclinical reporting guidelines that were published in 2010 (and included all elements of the NIH-PRG).¹⁷ Those guidelines were endorsed by hundreds of journals (including many in our sample), learned societies, and funders; however, adherence has been poor.^{20–22} In response, the simplified NIH-PRG were created in order to focus on seven key items to report; our study demonstrates that adherence to even these core reporting elements continues to be poor. Several studies have demonstrated common trends of poor reporting. Similar to

| | NIH-PRG | Level of Reporting | |
|--|--|--------------------|---------|
| Domain | Item | n | % (n/N) |
| Standards | Reporting guidelines listed | 1 | 99.6 |
| | | | |
| Replicates (biological vs. technical) | Results substantiated under a range of conditions | 225 | 95 5 |
| | Number of subjects per outcome | 25 | 11 89 |
| | Number of measurements per subject for one outcome | 105 | 44 56 |
| | Number of measurements per subject for all outcomes | 2 | 99 |
| | | | |
| | Total number of subjects used in each experimental group listed | 148 | 63 37 |
| 0 | All statistical tests used listed | 225 | 95 5 |
| Statistics | Definition of the measure of central tendency | 203 | 86 14 |
| | Definition of the measure of dispersion | 206 | 87 13 |
| | | | |
| Randomization | Random group assignment reported | 103 | 44 56 |
| | Method of randomization described | 2 | 99 |
| Blinding | Experimenters blinded during conduct of experiment | 7 | 3 97 |
| Diriding | Experimenters blinded during result assessment | 15 | 6 94 |
| | | | |
| | Description of primary outcome | 12 | 5 95 |
| Sample Size | Sample size reported | 10 | 4 96 |
| | Method of sample size determination described | 4 | 2 98 |
| | | | |
| | Total number of animals procured for the experiment reported | 7 | 3 97 |
| Inclusion and Exclusion Criteria | Criteria used to exclude any data or subjects reported | 11 | 5 95 |
| | Description of any outcomes that were measured and not reported in the results section | 228 | 97 3 |
| | Pilot or preliminary studies performed and listed | 27 | 89 |
| | Null or negative outcomes included in the results | 96 | 41 59 |

Figure 3. Reporting Assessment Results

Completeness of reporting across all included studies (N = 236) against the deconstructed NIH preclinical reporting guidelines (NIH-PRG) where n is the number of times each item was reported. Green and red correspond to an item being reported or not reported, respectively.

our study, Macleod et al.²³ showed that reporting both on randomization and the method of randomization was quite low (1.8%) in in vivo studies published by the Nature publication group; interestingly, after mandatory completion of a reporting checklist was instituted, the rate increased to 11.2% in these journals. In other studies, randomization was reported between 0% and 16% of included articles, depending on the journal and topic.^{20,22} Comparable results have also been found for blinding, with Macleod et al.²³ finding 4% of articles in their sample reporting on blinding of either experimenter or outcome assessors (which increased to 22.8% after institution of a mandatory checklist). Baker et al.²⁰ found approximately 20% of journals reported on blinding in their sample of studies. It is important to note that, even if a study did not employ randomization, blinding, sample size calculations, and other factors, the final publication could still easily adhere to the guidelines by transparently reporting that these methods were not used.

The Importance of Methodological Rigor and Transparent Reporting

The importance of rigorous study design and transparent reporting needs to be encouraged. Omission of key concepts in the preclinical setting has been associated with the exaggeration of effect sizes, type I (i.e., false positive) and type II (i.e., false negative) errors, and overestimation of the potential for successful bench-to-bedside translation.²⁴⁻²⁶ Incorporating methodological rigor into study design and reporting has been shown to reduce bias in the clinical setting.²⁷⁻³⁰ Accordingly, many commentators argue that fundamental concepts of clinical trial methodology, such as randomization and blinding, should be incorporated into preclinical experiments to increase study validity.³¹ These practices allow other researchers to evaluate the risk of bias in study reports: randomization addressing selection bias and blinding addressing performance bias. Within the cancer field, the importance of these design elements and reporting is evidenced by recent preclinical systematic reviews and meta-analyses of sunitinib³² and sorafenib.³³ These studies suggested that poor design and incomplete reporting of preclinical studies likely contributed to overestimated effect sizes in animal experiments relative to subsequent clinical studies in human patients. Within this context, our assessment of the OV literature is an important one that should be considered as efforts to clinically translate OV therapies continue. Within the present study, important methods to reduce bias, such as randomization and blinding were reported in only a minority of studies. It is unclear whether these items are not considered and are therefore omitted from study design or they are performed and simply not reported-a push toward complete reporting would shed light on this.

Strengths and Limitations

The choice of the NIH-PRG for our reporting tool and the method through which the recommendations were deconstructed and items operationalized into a checklist are supported by the expertise of the multidisciplinary research team, including basic scientists, who performed this study. Portions of this checklist were previously used by members of our group.³⁴ Furthermore, to ensure that the rele-

vant study characteristics were correctly collected, our team included several OV experts (C.I., J.S.D., R.A., N.E.-S., R.K., and B.A.K.).

Regarding potential limitations, the validity of included studies and their risk of bias was not evaluated; this limitation reflects reporting being the primary focus of our study. If risk of bias was evaluated, we would anticipate that most studies would be deemed "unclear risk of bias" in a majority of domains, due to a lack of reporting.³⁵ Second, it is possible that researchers are aware of reporting guidelines and rigorous study design, but interpreted items differently or simply did not report measures that were performed. Third, authors from countries outside of the United States and those who do not receive funding from the NIH may be unaware of these guidelines. Finally, a number of articles (n = 29, 12%) were published in *Oncotarget*, a journal that was recently delisted in MEDLINE.³⁶ However, when comparing the level of reporting in *Oncotarget* to all other articles, we found no meaningful differences (Table S4).

Future Steps

We believe the impetus for change will need to come through enforcement by external sources, such as funding agencies and journals (editorial boards, in particular).^{21,23} Examples include mandatory checklists at the time of submission³⁷ or requirement that the study be preregistered³⁸ on an open-source database prior to study commencement (similar to clinical trials).

Our study demonstrates that researchers as a community are either not yet aware of reporting guidelines or have not yet appreciated their significance. That is, they may be aware of some of these concepts (e.g., randomization and blinding), but may not appreciate the consequence that less rigorous design decisions have on results (e.g., exaggerating effect size). Here, we posit a few explanations. First, a contributing factor may be the lack of available formal training opportunities for basic scientists concerning these design and reporting elements. Clear guidelines on how these methods should be conducted and reported need to be established. Nonetheless, even in the absence of implementation of these methods, authors should be able to report that they were not used. Second, the impracticality of implementing these measures in some situations should be considered. For example, a single experimenter cannot be blinded to group allocation or it may be impossible to fully blind experiments (e.g., blinding may not be possible in animals treated with OVs, as they develop clearly visible signs associated with the virus). Again, in these situations, a lack of blinding could still be reported, for transparency. Third, studies adhering to a more rigorous design may require more resources (e.g., extra personnel to blind appropriately), access to which may vary widely among research groups and implementation of which may not be rewarded or compensated for by funding agencies at present. This, however, still does not interfere with transparent reporting regarding the application of these design elements. Last, it is possible that the lack of uptake of reporting guidelines, by other researchers and journal editors discourages any change of practices common in the laboratory. Without



Figure 4. Constructing the Reporting Checklist

The NIH Preclinical Reporting Guidelines (NIH-PRG) domain of randomization was deconstructed into two unidimensional items, and each was operationalized into a "yes" or "no" question.

tangible incentives, the time required for researchers to implement more rigorous study methodologies and fully report studies may be outweighed by pressures to publish rapidly.

Conclusions

This review is an extensive reporting assessment against an important set of published guidelines in a growing field of cancer research. Given the critical need for new cancer treatments, efficient allocation of resources, and reproducible experiments, it is crucial that preclinical *in vivo* OV research be conducted and reported in a transparent, methodical, and rigorous way. The items investigated here are fundamental to clinical research and have demonstrated benefits in preclinical research. Future work will need to determine how these guidelines and their underlying principles should be applied to preclinical research.

MATERIALS AND METHODS

Protocol

Using the Preferred Reporting Items for Systematic Review and Meta-Analysis Protocol (PRISMA-P) statement³⁹ as a guide, a study protocol was constructed and deposited on OSF (OSF; https://osf.io/ j2dwm/).³⁸ During completion of our review, there were no deviations from the protocol. The protocol was publicly posted concurrent with the commencement of data extraction. Although not a systematic review, the PRISMA statement⁴⁰ was used in the drafting of this manuscript.

Eligibility Criteria

We included all preclinical *in vivo* studies that used an OV therapy (prophylactic or interventional) to treat cancer, published between January 2016 and June 2017 (18 months). There were no limitations on viral platform, cancer model, animal model, comparisons, outcomes, or experimental design. Only original research, in the form of full-text English publications, was included. Abstracts, letters, reviews, and commentaries were excluded. Any *in vitro*, *ex vivo*, or clinical experiments were excluded.

Search Strategy and Article Screening

In consultation with an information specialist, a systematic search strategy was developed (OSF; https://osf.io/j2dwm/), and searches of MEDLINE and Embase were performed. Titles that met our eligi-

bility criteria were identified and uploaded to an audit-ready, cloudbased software (Distiller SR; Evidence Partners, Ottawa, ON, Canada). Two independent reviewers performed the process of study selection. Title and abstract screenings were performed independently and in duplicate using an accelerated screening method (one reviewer required to include, two reviewers required to exclude).⁴¹ A subsequent full-text screening was performed in duplicate by two independent reviewers (consensus required to exclude). Any conflicts were resolved through discussion with a third team member.

Data Extraction

Full-text articles which met our inclusion criteria were retrieved and uploaded to Distiller SR. All extractors (N.L.W., G.J.L., J.L.M., and I.C.) were affiliated with the Ottawa Hospital Research Institute. To ensure an adequate level of reviewer agreement, all extractors participated in a pilot training exercise. Each extractor was provided with a training document and independently extracted five articles. Feedback from a core group member (M.M.L.) was given, and a further five articles were independently extracted. At this point, the training exercise was completed, as inter-rater agreement met our pre-defined level of 80%. Two trained members of the extractor team independently extracted data from each article in duplicate. Data were collected concerning common study characteristics (e.g., country of corresponding author, journal, funding source, virus administered, cancer studied, and animal model used) and the level of reporting against the NIH-PRG domains (e.g., randomization and blinding). Conflicts were resolved through consensus discussions between extractors or with a senior investigator when needed. Online methods and supplemental information were retrieved whenever referenced.

Reporting Quality Assessment of Included Studies

The NIH-PRG recommendations are grouped into seven domains: (1) use of community-based reporting standards, (2) distinction between biological and technical replicates, (3) statistics, (4) sample size estimation, (5) randomization, (6) blinding, and (7) inclusion/ exclusion criteria.⁵ Each of these domains contains a multifaceted recommendation that addresses multiple points. To assess the level of reporting, each domain was deconstructed into unidimensional items. These items were operationalized into 21 "yes" or "no" questions, each addressing a single point. (The full list of questions is contained in our protocol.⁴²) For example, the domain of randomization was deconstructed into questions regarding whether random group assignment was reported and whether the method of randomization was described (Figure 4). All items mentioned in the NIH-PRG were considered relevant to our reporting assessment. This 21-item checklist served as our reporting assessment tool.

Data Synthesis

For each item addressed by the NIH-PRG recommendations, the level of reporting was expressed as the total number of times each item was reported (n) across all studies (N). This descriptive statistic, frequency count, is expressed nominally (n out of N) and as a percentage (n/N). No formal statistical analysis was performed.

SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at https://doi.org/10. 1016/j.omto.2019.05.004.

AUTHOR CONTRIBUTIONS

Conceptualization and methodology: M.M.L., J.P., K.D.C., J.-S.D., R.A., J.K., C.I., and D.A.F.; validation: N.E.-S., R.K., and B.A.K.; formal analysis: M.M.L., N.L.W., and D.A.F.; investigation: M.M.L., N.L.W., G.J.L., J.L.M., I.C., J.P., K.D.C., J.-S.D., R.A., J.K., N.E.S., R.K., B.A.K., C.I., and D.A.F.; resources: M.M.L. and D.A.F.; data curation: N.L.W., G.J.L., J.L.M., and I.C.; writing–original draft preparation: M.M.L., N.L.W., and D.A.F.; writing–review and editing: all authors; and funding acquisition: M.M.L. and D.A.F.

CONFLICTS OF INTEREST

The authors declare no competing interests.

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OMTO, Volume 14

Supplemental Information

Assessing the Completeness of Reporting

in Preclinical Oncolytic Virus Therapy Studies

Dean A. Fergusson, Neil L. Wesch, Garvin J. Leung, Jenna L. MacNeil, Isidora Conic, Justin Presseau, Kelly D. Cobey, Jean-Simon Diallo, Rebecca Auer, Jonathan Kimmelman, Natasha Kekre, Nader El-Sayes, Ramya Krishnan, Brian A. Keller, Carolina Ilkow, and Manoj M. Lalu

Supplementary Online Material

| Question | Response | Frequency |
|-------------------------------------|-----------------------------------|-----------|
| | | |
| | United States | 72 |
| | China | 63 |
| | Canada | 16 |
| | Japan | 15 |
| | Fililanu South Koroa | 10 |
| | United Kingdom | a IU |
| | Spain | 8 |
| | Germany | 7 |
| Indicate the corresponding author's | France | 6 |
| country of residence. | Russia | 5 |
| | Sweden | 3 |
| | Austria | 2 |
| | Denmark | 2 |
| | India Bolaium | 2 |
| | Ireland | 1 |
| | Netherlands | 1 |
| | Saudi Arabia | 1 |
| | Switzerland | 1 |
| | | |
| Indicate the year of publication. | 2016 | 169 |
| · · | 2017 | 67 |
| | Government | 196 |
| | Academic | 107 |
| Indicate all contributing forms of | Foundation / Charity | 94 |
| research funding | Industry | 39 |
| research randing. | Other (individual donations) | 12 |
| | No mention of funding | 5 |
| | Explicitly unfunded | 2 |
| | Operatoraat | 20 |
| | Melacular There are a constanting | 29 |
| | Molecular Therapy – Oncolytics | 23 |
| | Gene Therapy | 12 |
| | Molecular I herapy | 10 |
| | Cancer Gene Therapy | 10 |
| What is the journal of publication? | Oncolmmunology | 9 |
| | Clinical Cancer Research | 7 |
| | Molecular Cancer Therapeutics | 7 |
| | PLoS One | 6 |
| | Human Gene Therapy | 5 |
| | Neuro-Oncology | 5 |
| | Nature Communications | 4 |

Table S1. Study design and publication characteristics.

| Cancer Research | 4 |
|--|---|
| Vaccine | 4 |
| Cancer Letters | 4 |
| Journal of Controlled Release | 3 |
| Scientific Reports | 3 |
| International Journal of Oncology | 3 |
| International Journal of Cancer | 3 |
| Oncology Reports | 3 |
| Journal of Virology | 3 |
| Viruses | 3 |
| Cancer Biology | 2 |
| Acta Biomaterialia | 2 |
| Virus Research | 2 |
| Cancer Science | 2 |
| Journal of Cancer | 2 |
| ACS Nano | 2 |
| Journal of Translational Medicine | 2 |
| Blood | 2 |
| Journal of Pharmacological Sciences | 2 |
| Journal of General Virology | 2 |
| Cancer Immunology Immunotherapy | 2 |
| Apoptosis | 2 |
| Virology | 2 |
| Molecular Medicine Reports | 1 |
| Molecular Medicine | 1 |
| Medical Oncology | 1 |
| Journal of Cellular and Molecular Medicine | 1 |
| Journal of Gene Medicine | 1 |
| Journal of the National Cancer Institute | 1 |
| Cancer Immunology Research | 1 |
| Therapeutics and Clinical Risk Management | 1 |
| Surgery | 1 |
| The Journal of Immunology | 1 |
| The Journal of Nuclear Medicine | 1 |
| Virus Genes | 1 |
| Veterinary and Comparative Oncology | 1 |
| Applied Materials | 1 |
| Molecular Oncology | 1 |
| Drug Design, Development and Therapy | 1 |
| OncoTargets and Therapy | 1 |
| Molecular Therapy — Methods | 1 |

| | Acta Pharmacologica Sinica | 1 |
|--------------------------------------|--|-----|
| | Translational Oncology | 1 |
| | Breast Cancer Research | 1 |
| | American Journal of Cancer Research | 1 |
| | Stem Cell Reports | 1 |
| | Oncolytic Virotherapy | 1 |
| | Cellular Immunology | 1 |
| | Frontiers in Oncology | 1 |
| | BMC Cancer | 1 |
| | Reproductive Sciences | 1 |
| | Oncogene | 1 |
| | Biochemical and Biophysical Research Communications | 1 |
| | International Immunopharmacology | 1 |
| | Journal of Ovarian Research | 1 |
| | International Journal of Molecular Sciences | 1 |
| | Journal of Immunology Research | 1 |
| | Oral Oncology | 1 |
| | Tumor Biology | 1 |
| | Cellular Oncology | 1 |
| | Cell Reports | 1 |
| | EBioMedicine | 1 |
| | Journal of Experimental & Clinical Cancer Research | 1 |
| | The Canadian Journal of Veterinary Research | 1 |
| | Neurotherapeutics | 1 |
| | Head | 1 |
| | Molecular Genetics, Microbiology and Virology | 1 |
| | Current Gene Therapy | 1 |
| | Gut | 1 |
| | Russian Journal of Genetics: Applied Research | 1 |
| | International Journal of Clinical and Experimental Medicine | 1 |
| | Human Gene Therapy Clinical Development | 1 |
| | Iournal of Biomedical Science | 1 |
| | | - |
| | Cancer size/burden | 204 |
| Which of the following outcomes | Survival | 116 |
| were investigated? | Anti-tumor response | 94 |
| Ŭ | Cytokine response | 55 |
| | Tumor specificity/biodistribution | 52 |
| From the title alone, could you tell | Yes | 76 |
| this was a preclinical article? | No | 160 |
| - | - | |

| Adenovirus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccinia virus104 Vaccina virus11 Vesicular stomatitis virus11 Newcastle disease virus11 Newcastle disease virus11 Newcastle disease virus11 Newcastle disease virus9 Reovirus11 Newcastle disease virus11 Newcastle disease virus11 Newcastle disease virus13 Curus12 Curus13 Curus12 Curus13 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus12 Curus13 Curus13 Curus14 Curus16 Curus16 Curus16 Curus <th< th=""><th>Question</th><th>Response</th><th>Frequency</th></th<> | Question | Response | Frequency | |
|--|------------------------------------|------------------------------------|-----------|--|
| Adenovirus104Vaccinia virus34Herpes simplex virus27Vesicular stomatitis virus11Newcastle disease virus9Reovirus8Myxoma virus4Alphavirus3Cytomegalovirus3Maraba virus3Cytomegalovirus3Maraba virus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Human papilloma virus1Upphocytic choriomeningitis virus1Influenza virus1Influenza virus1Semliki forest virus1Semliki forest virus1No31Unsure2Yes182No31Unsure152What is the type of viral therapy?MonotherapyWhat is the route of viral administration?176Not intratumoural administration?176Unsure74Output77Unsure77 | | | | |
| Vaccinia virus34Herpes simplex virus27Vesicular stomatitis virus15Measles virus11Newcastle disease virus9Reovirus8Myxoma virus3Cytomegalovirus3Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Influenza virus1Lymphocytic choriomeningitis virus1Uymphocytic choriomeningitis virus1No31Uster virus1No31Unsure74Combination therapy74Combination therapy74What is the route of viral administration?177Unsure77Unsure77 | | Adenovirus | 104 | |
| Herpes simplex virus27Vesicular stomatitis virus15Measles virus11Newcastle disease virus9Reovirus8Myxoma virus4Alphavirus3Cytomegalovirus3Maraba virus3Cytomegalovirus3Maraba virus3Parvovirus3Parvovirus2Tanapoxvirus2Tanapoxvirus1Chicken infectious anemia virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Influenza virus1Influenza virus1Semliki forest virus1No31Unsure26No31Unsure26What is the route of viral therapy?MonotherapyWhat is the route of viralIntratumouralWhat is the route of viralIntratumouralAdministration?IntratumouralNo176Not intratumoural77Unsure7 | | Vaccinia virus | 34 | |
| Vesicular stomatitis virus15Measles virus11Newcastle disease virus9Reovirus8Myxoma virus3Cytomegalovirus3Cytomegalovirus3Cytomegalovirus3Parvovirus3Lentivirus2Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Chicken infectious anemia virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Influenza virus1Semiliki forest virus1Semiliki forest virus1Tobacco mosaic virus1Ves182No31Unsure26What is the route of viral therapy?MonotherapyWhat is the route of viralIntratumouralAdministration?IntratumouralWhat is the route of viralIntratumouralNo176Not intratumoural77Unsure74Combination therapy74Combination therapy74Not intratumoural77Unsure77Unsure77Unsure74Not intratumoural77Unsure74Not intratumoural77Unsure7Not intratumoural77Unsure7 | | Herpes simplex virus | 27 | |
| Measles virus11Newcastle disease virus9Reovirus8Myxoma virus4Alphavirus3Cytomegalovirus3Crytomegalovirus3Parvovirus3Parvovirus2Picornavirus2Picornavirus2Canine distemper virus1Chicken infectious anemia virus1Chicken infectious anemia virus1Chicken infectious anemia virus1Influenza virus1Influenza virus1Semliki forest virus1No3Is the virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viralIntratumouralWhat is the route of viralIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumouralNoIntratumoural | | Vesicular stomatitis virus | 15 | |
| Newcastle disease virus9Reovirus8Myxoma virus4Alphavirus3Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Picornavirus2Picornavirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Influenza virus1Lymphocytic choriomeningitis virus1Umphocytic choriomeningitis virus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?IntratumouralWhat is the route of viral administration?IntratumouralIntratumoural unsure77Unsure7 | | Measles virus | 11 | |
| Reovirus8Myxoma virus4Alphavirus3Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Semliki forest virus1Semliki forest virus1Semliki forest virus1Semliki forest virus1Ves182No31Unsure26What is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural Intratumoural Intratumoural Intratumoural Intratumoural Intratumoural | | Newcastle disease virus | 9 | |
| Myxoma virus4Alphavirus3Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Parvovirus3Lentivirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Influenza virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Semliki forest virus1Tobacco mosaic virus1Sten virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural 176What is the route of viral administration?Intratumoural 176 | | Reovirus | 8 | |
| Alphavirus3Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Influenza virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Mumps virus1Semliki forest virus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Ves182No31Unsure26What is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural IntratumouralNot intratumoural Unsure77Unsure7 | | Myxoma virus | 4 | |
| Cytomegalovirus3Maraba virus3Parvovirus3Lentivirus2Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Semliki forest virus1Tobacco mosaic virus1Semliki forest virus1Tobacco mosaic virus1Vhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural 177 UnsureWhat is the route of viral administration?Intratumoural 77 | | Alphavirus | 3 | |
| Maraba virus3What is the viral platform?Parvovirus3Lentivirus2Picornavirus2Picornavirus2Canine distemper virus1Chicken infectious anemia virus1Chicken infectious anemia virus1Chot-and-mouth disease virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus11Mengovirus1Semliki forest virus1Semliki forest virus111No3111Unsure26311What is the type of viral therapy?Monotherapy152What is the route of viral administration?Intratumoural77Unsure7Not intratumoural77Unsure7Nos intratumoural77Unsure7Nos intratumoural77 | | Cytomegalovirus | 3 | |
| What is the viral platform?Parvovirus3What is the viral platform?Picornavirus2Picornavirus2Canine distemper virus1Chicken infectious anemia virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Mengovirus1Semliki forest virus1Poliovirus1Semliki forest virus1Semliki forest virus1Vhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural No74Not intratumoural Unsure77Unsure7 | | Maraba virus | 3 | |
| What is the viral platform?Lentivirus2Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Mengovirus1Mengovirus1Semiliki forest virus1Tobacco mosaic virus1Ste virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural Not intratumoural Not intratumouralWhat is the route of viral administration?Intratumoural Tobacco | | Parvovirus | 3 | |
| Picornavirus2Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Mengovirus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural TobaccoNo152Not intratumoural Unsure77Unsure7 | What is the viral platform? | Lentivirus | 2 | |
| Tanapoxvirus2Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Semliki forest virus1Semliki forest virus1Tobacco mosaic virus1Semliki forest virus1Unsure26What is the type of viral therapy?Monotherapy + combination therapy 10What is the route of viral administration?Intratumoural 176 Not intratumoural 177 Unsure176 | | Picornavirus | 2 | |
| Canine distemper virus1Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesWhat is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural TobaccoWhat is the route of viral administration?Intratumoural Tobacco | | Tanapoxvirus | 2 | |
| Chicken infectious anemia virus1Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesMonotherapy152What is the type of viral therapy?Monotherapy + combination therapyWhat is the route of viralIntratumouralWhat is the route of viral176Not intratumoural176Not intratumoural77Unsure7 | | Canine distemper virus | 1 | |
| Foot-and-mouth disease virus1Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesMonotherapy152What is the type of viral therapy?Monotherapy + combination therapyWhat is the route of viral administration?Intratumoural 176What is the route of viral administration?Intratumoural 77 | | Chicken infectious anemia virus | 1 | |
| Human papilloma virus1Influenza virus1Lymphocytic choriomeningitis virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesMonotherapy152What is the type of viral therapy?Monotherapy + combination therapyWhat is the route of viral administration?Intratumoural T76 UnsureWhat is the route of viral administration?176 T77 T | | Foot-and-mouth disease virus | 1 | |
| Influenza virus1Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26What is the type of viral therapy?Monotherapy + combination therapyWhat is the route of viral administration?Intratumoural TroWhat is the route of viral administration?176 | | Human papilloma virus | 1 | |
| Lymphocytic choriomeningitis virus1Mengovirus1Mumps virus1Poliovirus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26What is the type of viral therapy?MonotherapyWhat is the route of viral administration?Intratumoural Not intratumoural Not intratumouralYhat is the route of viral administration?Intratumoural Yes | | Influenza virus | 1 | |
| Mengovirus1Mumps virus1Poliovirus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26What is the type of viral therapy?MonotherapyVhat is the route of viral administration?176Not intratumoural Unsure77Not intratumoural Unsure77Not intratumoural Unsure77 | | Lymphocytic choriomeningitis virus | 1 | |
| Mumps virus1Poliovirus1Poliovirus1Semliki forest virus1Tobacco mosaic virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26What is the type of viral therapy?MonotherapyWhat is the route of viral administration?77What is the route of viral administration?176 | | Mengovirus | 1 | |
| Poliovirus1Semliki forest virus1Tobacco mosaic virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26What is the type of viral therapy?MonotherapyMonotherapy + combination therapy74Combination therapy10What is the route of viral176Not intratumoural77Unsure7Unsure7 | | Mumps virus | 1 | |
| Semliki forest virus1Tobacco mosaic virus1Tobacco mosaic virus1Is the virus replicating?YesNo31Unsure26Unsure26What is the type of viral therapy?MonotherapyMonotherapy + combination therapy741010What is the route of viral administration?IntratumouralNot intratumoural administration?77Unsure7 | | Poliovirus | 1 | |
| Tobacco mosaic virus1Is the virus replicating?182Is the virus replicating?182No31Unsure26What is the type of viral therapy?MonotherapyMonotherapy + combination therapy74Combination therapy10What is the route of viral administration?176Not intratumoural Unsure77The route of viral administration?77 | | Semliki forest virus | 1 | |
| Yes182Is the virus replicating?No31Unsure26What is the type of viral therapy?Monotherapy152Monotherapy + combination therapy74Combination therapy10What is the route of viral176Not intratumoural77Unsure7Unsure7 | | Tobacco mosaic virus | 1 | |
| Yes182Is the virus replicating?No31Unsure26What is the type of viral therapy?Monotherapy152Monotherapy + combination therapy74Combination therapy10What is the route of viral176Not intratumoural77Unsure7 | | | | |
| Is the virus replicating?No31Unsure26What is the type of viral therapy?Monotherapy152Monotherapy + combination therapy74Combination therapy10What is the route of viral administration?176Unsure77Unsure7 | | Yes | 182 | |
| Unsure26Unsure26What is the type of viral therapy?MonotherapyMonotherapy + combination therapy74Combination therapy10Unsure176Not intratumoural77Unsure7 | Is the virus replicating? | No | 31 | |
| Monotherapy152What is the type of viral therapy?Monotherapy + combination therapy74Combination therapy10What is the route of viral administration?Intratumoural176Unsure77 | | Unsure | 26 | |
| Monotherapy152What is the type of viral therapy?Monotherapy + combination therapy74Combination therapy10What is the route of viral administration?Intratumoural176Unsure7 | | | | |
| What is the type of viral therapy?Monotherapy + combination therapy74Combination therapy10What is the route of viral administration?Intratumoural176Unsure7 | | Monotherapy | 152 | |
| Combination therapy 10 What is the route of viral administration? Intratumoural 176 Unsure 77 | What is the type of viral therapy? | Monotherapy + combination therapy | 74 | |
| What is the route of viral administration?Intratumoural Not intratumoural176 77 77Unsure77 | | Combination therapy | 10 | |
| What is the route of viral administration?Intratumoural Not intratumoural176 77 7 | | | | |
| administration? Not intratumoural 77 Unsure 7 | What is the route of viral | Intratumoural | 176 | |
| Unsure 7 | administration? | Not intratumoural | 77 | |
| | | Unsure | 7 | |

Table S2. Epidemiology of viral, cancer, and animal models.

| Question | Response | Frequency | |
|----------------------------------|-------------------|-----------|------------|
| | | | |
| | Colorectal | 42 | |
| | Skin | 35 | |
| | Breast | 33 | |
| | Liver | 28 | |
| | Brain | 25 | |
| | Lung | 25 | |
| | Ovary | 16 | |
| | Blood | 15 | |
| | Pancreas | 15 | |
| | Muscle | 10 | |
| | Prostate | 10 | |
| What type of cancer was induced? | Bladder | 7 | |
| | Bone marrow | 5 | |
| | Gastric | 5 | |
| | Kidney | 5 | |
| | Head and neck | 4 | |
| | Bone | 3 | |
| | Cervical | 3 | |
| | Esophagus | 2 | |
| | Mesothelium | 2 | |
| | Nervous system | 1 | |
| | Teratoma | 1 | |
| | Thyroid | 1 | |
| | | | |
| | Implantation | 235 | |
| What was the method of | Transgenic | 3 | |
| cancer/tumor induction? | Chemical | 0 | |
| | | | |
| | Xenograft | 153 | |
| Indicate the type of implant. | Syngeneic | 99 | |
| | | | |
| | Heterotopic | 170 | |
| What is the site of the tumor? | Orthotopic | 83 | |
| | Unsure | 6 | |
| Question | Response | Frequency | |
| | | | Number Use |
| | Mouse | 231 | 25516 |
| What was the type of animal(s) | Rat | 5 | 516 |
| used? | Hamster | 4 | NA |
| | Non-human primate | 1 | 12 |
| I | • | I | |

| | Immunodeficient | 134 |
|------------------------------------|-----------------------------|-----|
| What type of animal model was | Humanized | 3 |
| used? | Partially immunocompromised | 16 |
| | Unsure | 5 |
| | | |
| Was ethics approval for animal use | Yes | 172 |
| explicitly reported? | No | 64 |
| | | |
| Was the housing and handling of | Yes | 104 |
| | | |
| animals reported? | No | 132 |

Table S3. Reporting assessment results. The frequency of reporting of all 21 items

deconstructed and operationalized from the National Institutes of Health preclinical reporting

guidelines across the all included studies. These recommendations are separated into seven

distinct domains.

| Domain | Domain Item Description | |
|---|---|----------|
| Standards | Community based reporting guidelines listed | 1 (0.4) |
| | Results substantiated by repetition under a range of conditions | 225 (95) |
| Replicates (biological vs. technical) | Number of subjects stated for all experimental outcomes | 25 (17) |
| | Number of measurements per subject for one experimental outcome stated | 105 (44) |
| | Number of measurements per subject for all experimental outcomes stated | 2 (1) |
| | List of the total number of subjects used in each experimental group | 148 (63) |
| Statistics | List of all statistical tests used | 225 (95) |
| | Definition of the measure of central tendency | 203 (86) |
| | Definition of the measure of dispersion and precision | 206 (87) |
| Pondomization | Random group assignment reported | 103 (44) |
| Randomization | Description of the method of random group assignment | 2 (1) |
| Blinding | Experimenters blinded to group allocation during conduct of the experiment | 7 (3) |
| Dinding | Experimenters blinded to group allocation during result assessment | 15 (6) |
| | Description of an a priori primary outcome | 12 (5) |
| Sample Size Estimation | Sample size computed during study design | 10 (4) |
| | Description of the method of sample size determination | 4 (2) |
| Inclusion and Exclusion Criteria | Total number of animals procured for the experiment reported | 7 (3) |
| | Description of the criteria used for the exclusion of any data or subjects | 11 (5) |
| | Description of any outcomes that were measured and not reported in the results section | 228 (97) |
| | Pilot or preliminary studies performed and listed | 27 (11) |
| | Null or negative outcomes included in the results | 96 (41) |

Table S4. Exploratory comparative analysis. A comparison in the completeness of reporting between *Oncotarget*, which was delisted by MEDLINE in 2016,³³ compared to all other articles included in our review.

| | | Oncotarget N=29 | Other N=207 |
|--------------------|---|--------------------|----------------|
| Domain | Description | n (%) Reported | |
| Standards | Reporting guidelines | 0 (0) | 1 (0) |
| | Range of conditions | 29 (100) | 192 (93) |
| | Number of subject for one outcome | 15 (52) | 133 (64) |
| Replicates | Number of subjects for all outcomes | 4 (14) | 21 (10) |
| | Number of measurements for one outcome | 10 (34) | 95 (46) |
| | Number of measurements for all outcomes | 0 (0) | 2 (1) |
| | Statistical tests used | 25 (86) | 200 (97) |
| Statistics | Measure of central tendency | 24 (83) | 179 (86) |
| | Measure of dispersion | 23 (79) | 183 (88) |
| | Multiple experimental groups | 29 (100) | 205 (99) |
| Randomization | Random group assignment | 15 (52) | 89 (43) |
| | Method of randomization | 0 (0) | 2 (1) |
| Plinding | Group allocation blinding | 3 (10) | 4 (2) |
| Binding | Result assessment blinding | 2 (7) | 13 (6) |
| | Primary outcome | 2 (7) | 10 (5) |
| Sample Size | Sample size calculation | 2 (7) | 10 (5) |
| Louinduon | Mathematical method used | 0 (0) | 4 (2) |
| | Total number of subjects | 0 (0) | 7 (3) |
| | Data/subjects/results exclusion | 0 (0) | 11 (5) |
| Inclusion and | No result omission | 29 (100) | 199 (96) |
| Exclusion Criteria | Preliminary studies | 1 (3) | 26 (13) |
| | Null/negative results | 9 (31) | 87 (42) |