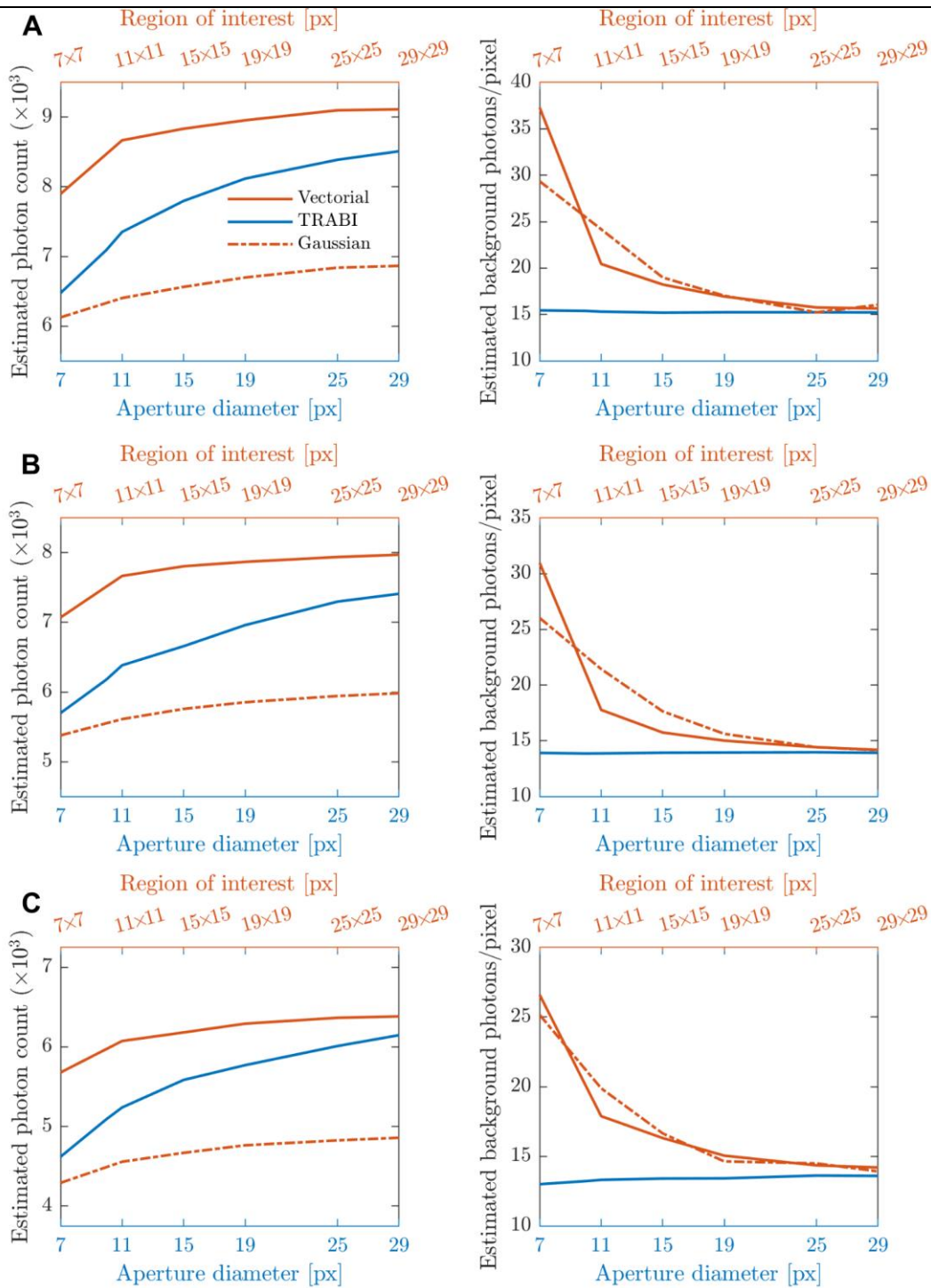


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Impact of optical aberrations on axial position determination by photometry

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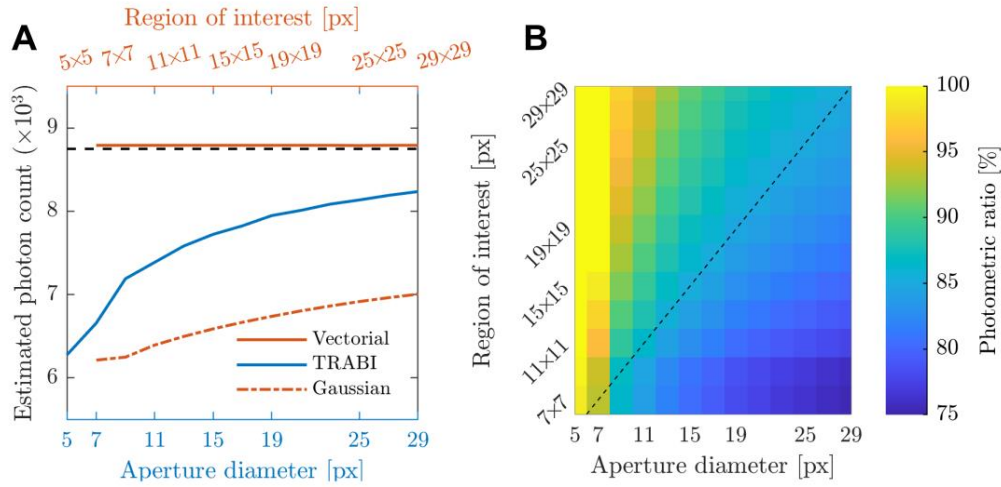


Supplementary Figure 1

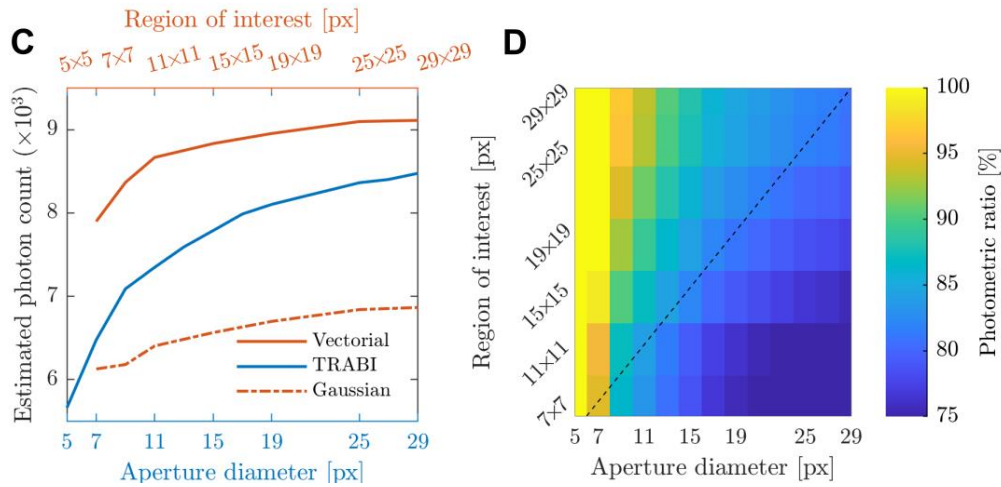
Reproducibility of photon count estimation.

(A-C) Estimated photon counts and background photons per pixel for three measurements of three separated 45-nm-diameter beads imaged with an aberration-corrected microscope as a function of analysis area on the camera; pixel size, 80 nm. The three lines show the count for fitting with a fully fledged vectorial, TRABI or a simple Gaussian PSF.

Zero aberration simulation



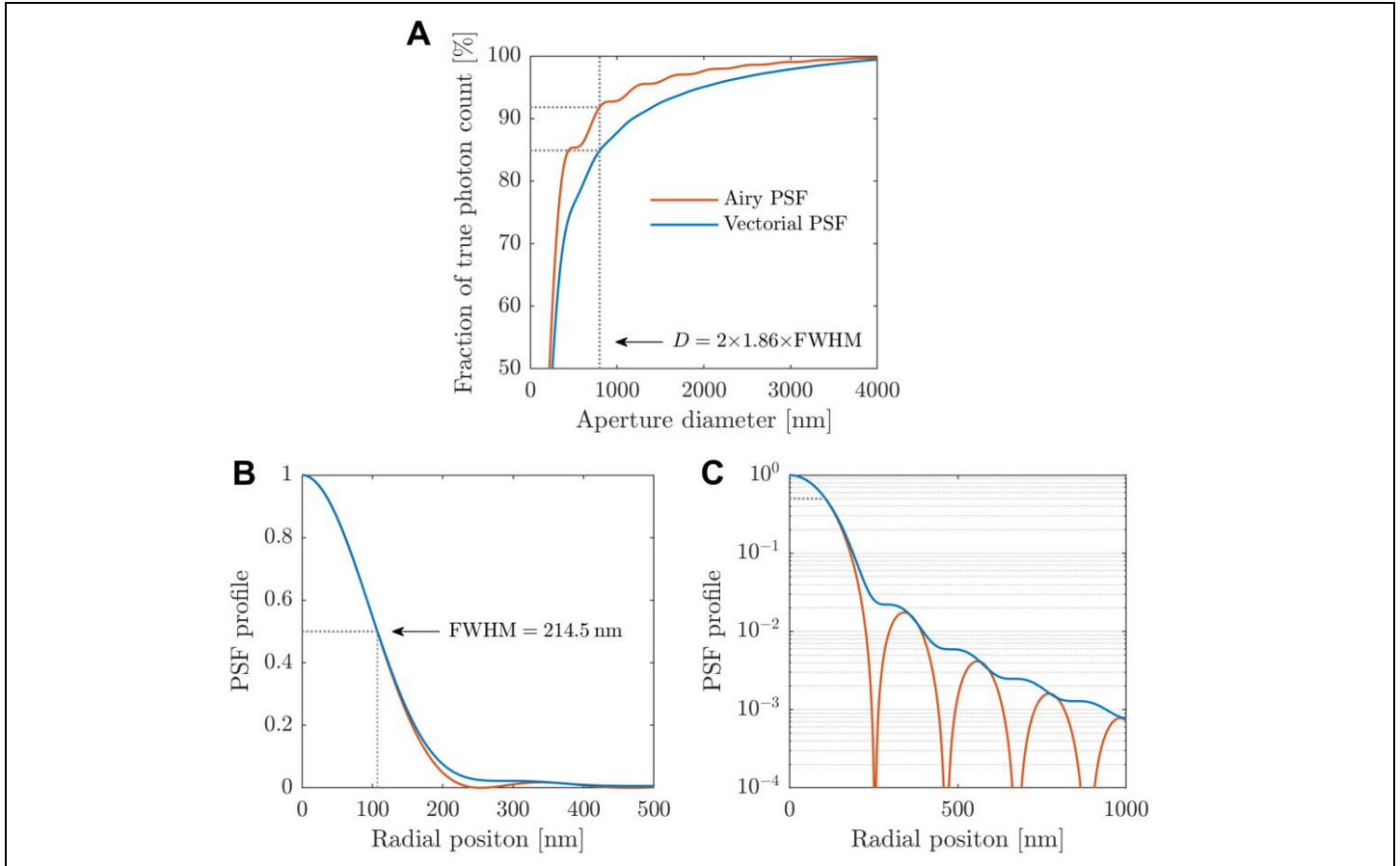
Aberration-corrected experiment



Supplementary Figure 2

Comparison of photon count estimations in simulation and experiment.

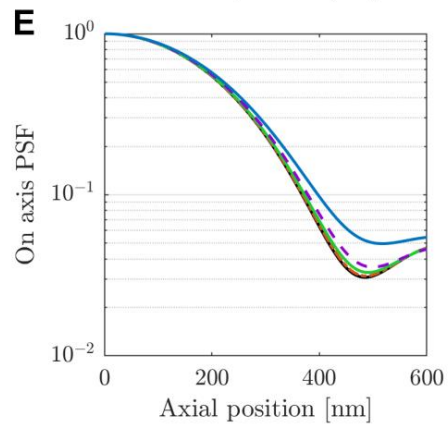
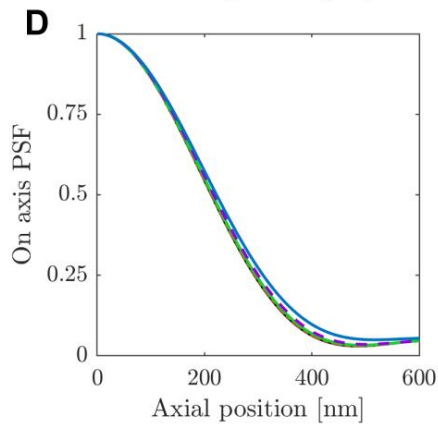
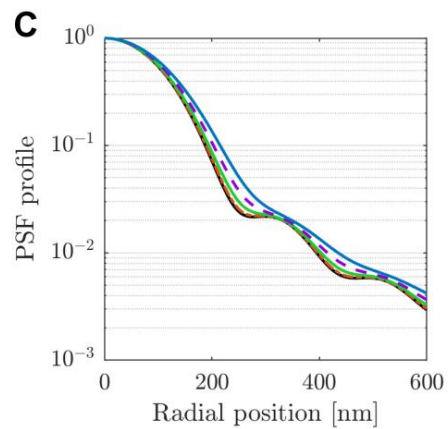
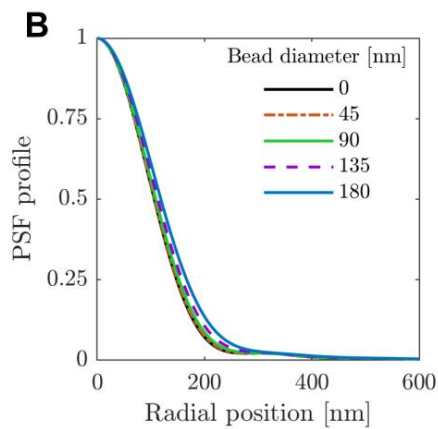
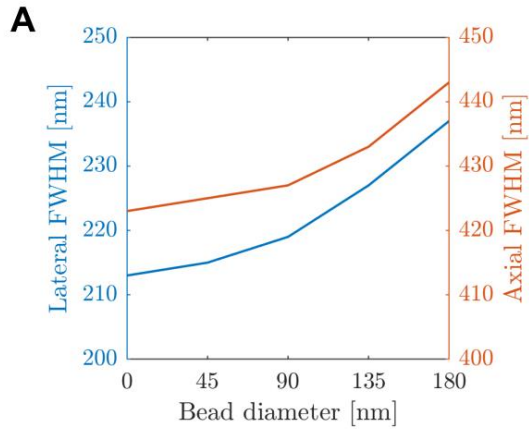
(A) Photon count estimation for a single simulated 45-nm bead PSF as a function of PSF image area on the camera for three different estimation algorithms: vectorial fit, TRABI, and Gaussian fit (8,750 signal photons, 15 background photons, same pixel size, NA and refractive index values as experiment). **(B)** The photometric ratio between Gaussian fit and TRABI photon count estimation as a function of PSF fit size. The dashed line indicates when the circular aperture size fits precisely within the square region of interest. **(C-D)** Same as panels (A,B) but for an experimentally recorded aberration-corrected 45-nm bead, indicating good quantitative agreement between experiment and simulation.



Supplementary Figure 3

TRABI photon count estimate as a function of aperture size and PSF model.

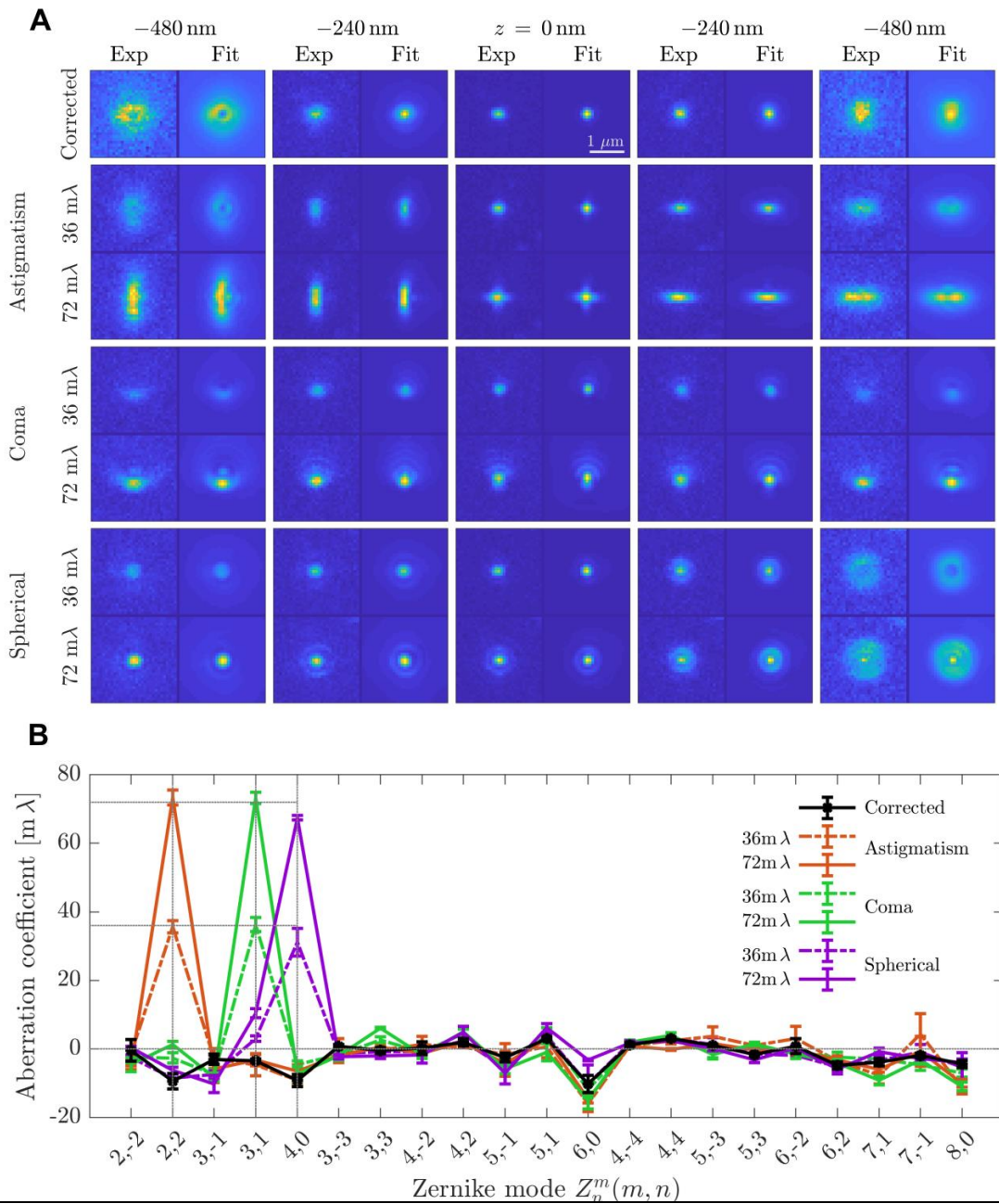
(A) Ratio of the estimated photon count by TRABI and simulated number of signal photons captured at the camera as a function of aperture diameter for two PSF models. TRABI aperture diameter (d) of $2 \times 1.86 \times \text{FWHM}$ ($\text{FWHM} = 214.5 \text{ nm}$) as suggested in Franke et al.¹ is indicated. For this aperture size, we find TRABI to estimate 92% and 85% of the total number of photons according to the low-NA scalar Airy PSF model and the high-NA vectorial PSF model, respectively. The fraction of the total energy contained within an aperture of the prescribed radii in the Airy PSF model ($\text{PSF}_{\text{Airy}}(\rho) = [2J_1(D)/(D)]^2$ where $D = 2\pi\rho(\text{NA}/\lambda)$; $\text{FWHM} = 0.514(\lambda/\text{NA})$) follows the analytically derived formula by Born & Wolf (ref. 4 in the Supplementary Methods reference list; Chapter 8; Fig. 8.13), in contrast to the 100% reported by Franke et al.¹ in their supplement. The PSF simulations were performed with 2,500 signal photons and no background photons and a pixel size of 1 nm (to exclude quantization effects). Otherwise, the simulation parameters were the same as the experimental parameters. **(B-C)** Characterization of the Airy and vectorial PSFs as a function of the radial position in a linear and log scale, respectively, showing the long tail of the PSF.



Supplementary Figure 4

The effect of bead size on PSF shape.

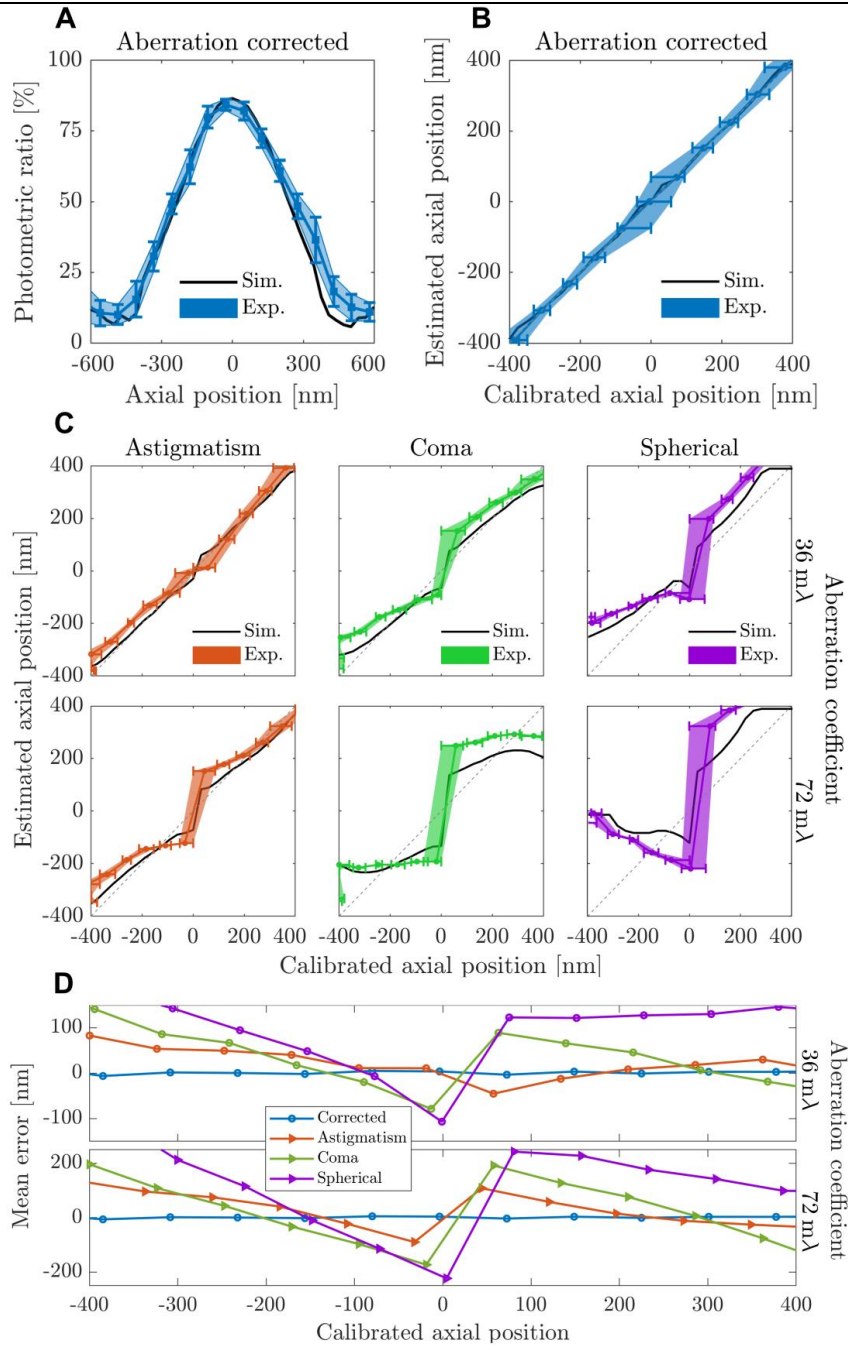
(A) The lateral and axial FWHM as a function of bead size. **(B-C)** Lateral average PSF in linear and log scale, respectively, as a function of the radial position. The PSFs are displayed for different bead sizes. **(D-E)** On-axis PSF in linear and log scale, respectively, as a function of the axial position. The 45-nm bead captures the full tail behavior in both the lateral and axial directions of an actual single-molecule emitter, and has an FWHM within a few nanometers of a single-molecule emitter, whereas PSF details clearly get lost with larger (>90 nm) beads.



Supplementary Figure 5

Quantification of aberration retrieval and correction.

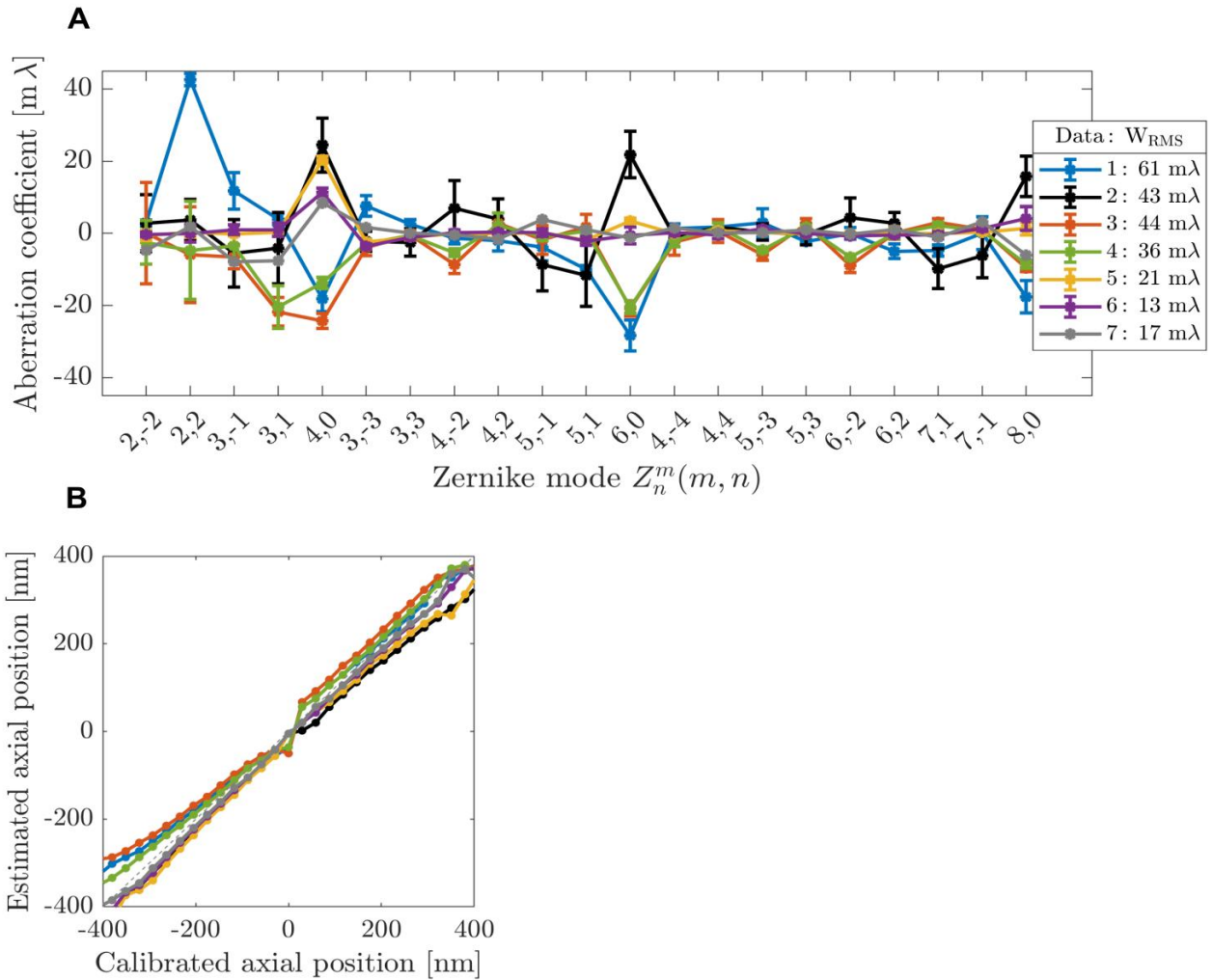
(A) Through-focus PSF image stacks of experimental and fitted PSFs after aberration correction and, subsequently, aberrated with a single primary Zernike mode: astigmatism (Z_2^2), coma (Z_3^1), and spherical aberration (Z_4^0), with aberration coefficients 36 mλ and 72 mλ (root-mean-square values). The region of interest for each PSF image is 31×31 pixels with a pixel size of 80 nm. All 4×4 sub-image pairs are contrast-stretched with the same factor for better visibility of spot shape. The estimated photon counts were within 9,500–21,000 signal photons and 15–18 background photons per pixel. Reproducibility as shown in **(B)**. **(B)** Fitted Zernike modes and retrieved aberration coefficients. The coefficients are averaged over six measurements with error bars indicating one s.d. The aberration fit routine includes all tertiary Zernike modes (all Z_n^m with $2 < n + |m| \leq 8$) and assumes optical parameters as described in the Supplementary Methods. (Horizontal dashed lines are used to guide the eye.)



Supplementary Figure 6

Axial calibration of the photometric ratio and axial estimation error caused by small single-mode aberrations.

(A) The photometric ratio over six 45-nm-bead measurements after aberration correction (Exp.) and simulated vectorial PSFs (Sim.) as a function of axial position (see Supplementary Methods). **(B)** The estimated axial position of aberration-corrected beads as a function of the calibrated axial position shown in **(A)**. Around focus the error is a bit larger as expected owing to the near-parabolic shape of the photometric ratio curve (see Supplementary Methods for a description of the error analysis). **(C)** Estimated axial position for single-mode aberrations on 45-nm beads using the calibrated axial position from the aberration-corrected data in **(A)**. The gray dashed line is used to guide the eye for the correct estimation. Experimental data (Exp.) in **A,B,C** are shown as mean \pm s.d. over six bead measurements. **(D)** Error of the estimated axial position as the distance between the mean estimated position and the true (calibrated) position.



Supplementary Figure 7

Typical microscope aberrations and their influence on the estimated axial position.

(A) Fitted Zernike modes and retrieved aberration coefficients for several microscopes, Data 1–7 (specifications are given in the Supplementary Methods). The coefficients are averaged over six and one bead measurement(s) for datasets 1–6 and 7, respectively, with error bars indicating one s.d., and W_{RMS} is the mean of the wavefront error. The aberration fit routine includes all tertiary Zernike modes and assumes optical parameters for each microscope as described in the Supplementary Methods. **(B)** Estimated axial position for simulated single-molecule PSFs with aberrations equaling the mean experimentally found microscope aberrations in **(A)** compared to the calibrated axial position using the aberration-corrected photometric ratio (see Supplementary Fig. 6a). Area of fit 7×7 pixels and aperture radius $1.86 \times FWHM$.

Supplementary Methods

Optical microscope setup

The experimental data is acquired with a setup that consists of a Nikon Ti-E microscope with a spatial light modulator (SLM) for aberration correction and PSF engineering, as described by us earlier¹. In short, beads are excited by a 488 nm laser (Sapphire 488-100 CW CDRH, Coherent) through a dichroic filter set (Ex: Semrock FF01-460/60-25, Di: Semrock Di02-R532-25X36, Em: Semrock FF01-545/55-25) and an objective lens (APO TIRF 100x/1.49, Nikon). The emission fluorescence passes (peak $\lambda = 552$ nm) through a relay system ($f_1 = 100$ mm, Thorlabs AC254-100-A and $f_2 = 200$ mm, Thorlabs AC508-200-A) resulting in a final magnification of 200x. The SLM (XY-series, 512x512, 15 μm pixel size, Meadowlark) is placed in the Fourier plane of the relay system. The fluorescence image signal is captured on an EMCCD camera (iXon Ultra - X987, 512x512, 16 μm pixel size, Andor) which has in effect a back-projected pixel size of 80 nm in the object plane.

Sample preparation

Fluorescent beads of 45 ± 8 nm diameter (FluoSpheres 0.04 μm 505/515, ThermoFisher) with peak emission at 552 nm through our filter set are used in the imaging experiments. The beads are immersed in a mounting medium of immersion oil ($n=1.518$ Type F, Nikon) to match the refractive index to the coverslip and immersion oil used for the objective lens. The beads are diluted to 10^{11} particles/mL in water, and 2.5 μL of the solution is drop-cast on a coverslip. After evaporation of the water, a drop of immersion oil is applied, and the coverslip is glued around the edges with nail varnish to a microscope slide. This step ensured the rigidity of the sample and is necessary for precise z-positioning.

Data acquisition

Fluorescent beads are imaged with 3.5 mW of excitation power (measured at the back aperture of the objective) and an exposure time of 1 second. For each set of aberration (type and magnitude), the experiment is repeated three times in the same field of view (FOV) to test for reproducibility. For each FOV, we select two beads located in regions with a low bead density to avoid crosstalk of beads. This is done to ensure a large region of interest with the no/minimal overlap of their respective PSF shoulders. This results in a total of six configurations, i.e. two beads measured three times. These six configurations are used to compute the mean and standard deviation of the data as shown as error bars in **Figure 1B,C** and **Supplementary Figure 5**. The data in **Figure 1A** and **Supplementary Figure 1** and **2B,C** are for individual beads.

Aberration retrieval and correction

Our setup contains an additional light path for pixel-wise calibration of the LCoS SLM to ≤ 20 m λ RMS wavefront aberration as described earlier¹. After calibration of the SLM, we acquire a z-stack of the fluorescent bead by moving the piezo stage in steps of 80 nm from -800 nm to +800 nm around focus. The z-stack is fitted with a 3D full vectorial PSF model in order to

estimate the aberrations of the optical system¹. The retrieved aberration coefficients are fed to the SLM with opposite sign in order to correct for the aberrations. Subsequently, another z-stack is acquired and fitted to verify the compensation of the aberrations. We then add aberrations of a desired type and magnitude deliberately in order to investigate the impact of aberrations in a well-controlled fashion.

Simulation of a realistic PSF model

We use the vectorial PSF model also for simulating the effect of arbitrary aberrations on photon counting⁷. In addition we take into account the non-zero size of a bead by convolving the PSF with the bead size. This fully-fledged PSF model takes the following effects into account: interfaces between media, polarization, dipole orientation (here freely rotating dipoles are assumed) and type and magnitude of aberrations. The simulation parameters are based on our optical setup: i.e., medium refractive index $n = 1.518$, numerical aperture $NA = 1.49$, wavelength $\lambda = 552$ nm, backprojected pixel size = 80 nm. We assume that the refractive indices between the imaging medium, coverslip, and immersion medium are matched (based on the immersion of the beads in oil). In **Supplementary Figure 3** and **4** we use a smaller sampling distance of 1 nm to exclude quantization effects. The bead diameter is 45 nm except in **Supplementary Figure 4** where the bead diameter is varied from 0 to 180 nm. Results for the Airy PSF model shown in **Supplementary Figure 3** follow directly from the derivation as detailed in Born & Wolf⁴ (Chapter 8). The simulated PSFs are normalized such that the sum over the detection plane is unity, then multiplied by the desired number of signal photons and a constant number of background photons per pixel is added before applying shot noise.

Photon count estimation

To determine the gain and offset, the EMCCD camera is calibrated using the procedure described by van Vliet et al.². For these experiments, we used an EM gain setting of 50 to reduce the read-noise to <1 e- RMS while retaining a good dynamic range and linear response (maximum intensity is kept under 5000 ADU - the linear regime for our camera settings). Excess noise in EMCCD cameras leads to an overestimation of the gain by a factor of two³, but this does not pose any problems in the further analysis of the data as only ratios of photon count estimates are of interest here.

The fitting procedure with a vectorial or Gaussian PSF model is implemented with a Levenberg-Marquardt iterative scheme for Maximum-Likelihood estimation (MLE) as described in Smith et al.⁶. The number of iterations is terminated by the tolerance in the residual. The tolerance limit is set to 10^{-6} and 10^{-4} for the vectorial and Gaussian fitter, respectively (maximum number of iterations is 75). Typically the number of iterations is less than 20.

TRABI⁵ is designed to take advantage of fluorescent on-off blinking of photoswitchable or photoactivatable fluorophores. TRABI can estimate both signal and background with a

“single aperture” by incorporating time information, i.e. estimating the background in the off-state and the foreground signal in the on state. Here we image non-blinking fluorescent beads, which does not allow us to apply the method directly. Instead, we estimate the number of background photons by an aperture located far away from the bead. For background averaging we use 7 successive frames in the through-focus stack as suggested by Franke et al⁵.

Estimation of error axial position estimation

The error in the axial position estimate from the photometric ratio is done in the following way. The experiments on different beads give the photometric ratio $PR(z)$ as a function of axial position z with uncertainty $\Delta PR(z)$ (**Fig. 1B,C**). The experimental aberration-corrected calibration curves $PR(z)$ and $PR(z) \pm \Delta PR(z)$ (**Supplementary Fig. 6A**) are up-sampled to 1 nm axial steps and stored in a look-up-table (LUT). Given a measured value PR_{exp} , the LUT will give the estimated axial position z_{exp} with uncertainty Δz_{exp} . This uncertainty represents statistical variations in the photometric ratio curves due to e.g. variations across the field-of-view of the microscope, and is on the order 25 nm away from focus to 65 nm close to focus (**Supplementary Fig. 6B**). The estimated axial position change in case aberrations are present as the photometric ratio curves change (**Supplementary Fig. 6C**). If calibration is done on an aberration-corrected setup, the application of the aberration-free LUT on photometric ratio measurements on aberrated spots will also lead to a bias in the axial position estimate (**Supplementary Fig. 6D**). These inaccuracies can amount up to 100 nm for aberration levels at 50% of the diffraction limit.

Microscope specifications for PSF retrieval of Supplement Figure 7

Dataset 1 is obtained with our setup described in **Supplementary Methods** prior to aberration correction.

Dataset 2 is acquired on a Leica DMI8 microscope. The sample (TetraSpeck Fluorescent Microspheres, 200 nm, well 4, RI: mounting medium 1.472) is excited by a Leica PL 6000, diode laser through a filter set (Chroma ZT/594rpc, Chroma 89100bs), and an objective lens (Leica Plan-Apochromat 63x/1.40 Oil DIC, RI: embedding medium 1.518). The fluorescence image signal (emission peak at $\lambda = 580$ nm) is captured on a sCMOS camera (Hamamatsu C11440 ORCA-flash 4.0, detector 13.312x13.12mm², pixel size 6.5x6.5 μm^2 , chip size 2048x2048).

Datasets 3,4,5,6 are acquired on a Nikon Eclipse Ti-E microscope. The sample (TetraSpeck Fluorescent Microspheres, 200 nm, well 4, RI: mounting medium 1.472) is excited by a Lumencor SPECTRA X, diode laser through a filter set (**dataset 3,5**: Ex.: SPECTRA X Chroma 470/24, Em.: SPECTRA X, ET515/30m Single Bandpass, and **dataset 4,6**: Ex.: SPECTRA X Chroma Excitation Filter 550/15, Em.: SPECTRA X, ET595/40m Single Bandpass) and an objective lens (**dataset 3,4**: CF160 TIRF Apo 60x/oil DIC 1.49 NA, **dataset 5,6**: CF160 SR/TIRF Apo 100x/oil 1.49 NA, RI: embedding medium 1.518). The fluorescence image signal

(emission peak **dataset 3,5**: $\lambda = 515$ nm, **dataset 4**: $\lambda = 570$ nm, **dataset 6**: $\lambda = 580$ nm) is captured on a sCMOS camera (Andor Zyla 5.5 sCMOS, detector 16.6x14 mm, pixel size 6.5x6.5 μm^2 , chip size 2560x2160).

Dataset 7 is acquired on an Olympus IX71 microscope. The sample (FluoSpheres F-8789, dark red, Invitrogen, 40 nm) is excited by a HL63133DG, Thorlabs, 637 nm diode laser through a filter (FF01-692/40-25, Semrock), dichroic mirror (650 nm, Semrock) and an objective lens (UAPON 150XOTIRF, Olympus America Inc., NA 1.45, RI: immersion medium 1.52). The fluorescence image signal (emission peak $\lambda = 690$ nm) is captured via a 2X magnification relay optics on an EMCCD camera (iXon 897, Andor, Andor Technologies PLC., CCD size 515x515, pixel size 12 μm). Setup details and sample preparation is described further by Liu et al.⁸.

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