

Supplemental Materials

Molecular Biology of the Cell

Boudreau et al.

Figure S1 | Partial *dhc-1/lis-1* and *zyg-12/sun-1* RNAi affect centrosome separation through separate mechanisms. (A) Representative maximum intensity projections of embryonic confocal stacks. All RNAi treatments were performed with L4 stage worms expressing H2B:GFP and γ -tubulin:GFP for 24 hours via feeding unless otherwise noted. Scale bar = 10 μ m. (B)(C) Centrosome-centrosome distance and centrosome-male pronucleus distance were measured in 3D between respective centroids. Error bars are SEM and are represented as shaded areas around means. n of embryos analyzed per condition > 10. Statistics: *t* test where *P < 0.05. Time t = 0 s corresponds to centrosome separation.

Figure S2 | Cytoplasmic microtubule growth velocities prior to pronuclear migration. (A)(B)(C)(D)(E)(F) Frequency distributions of cytoplasmic microtubule growth velocities prior to pronuclear migration. The black dashed line represents a threshold over which microtubules are assumed to be accelerated. The grey dashed curves represent a gaussian distribution fit to control cytoplasmic microtubule growth velocities. n of microtubules analyzed per condition is > 50 across > 3 embryos. Frequency distributions between RNAi conditions were statistically non-significantly different ($p > 0.05$) using both Mann-Whitney and Kolmogorov-Smirnov tests. (G) Drawing illustrating the imaging plane, cytoplasmic microtubules in black with green plus ends representing EBP2, and representative single plane confocal image of embryos expressing EBP2:GFP. The yellow trajectory line represents a track for up to 10 frames (2.5 s) following the displayed frame using TrackMate. Scale bar = 10 μm .

Figure S3 | Quantification of endogenous DHC-1:mNeonGreen mean amounts on female and male pronuclear envelopes. (A)(B) Mean amounts of DHC-1:mNeonGreen (arbitrary units – a.u.) on the nuclear envelope in (A) 16 hour and 24 hour *dhc-1* RNAi treatments and (B) 24 hour *zyg-12* and *sur-6* RNAi treatments. Quantifications were performed using single slices of confocal stacks from embryos expressing mCherry:HIS-58 and endogenous DHC-1:mNeonGreen. n of embryos analyzed per condition is > 14. Time t = 0 s corresponds to NEBD.

Figure S4 | Pronuclear cross-sectional area measurements during pronuclear expansion. Pronuclear cross-sectional areas (μm^2) were measured using maximum intensity projections of mCherry:HIS-58 confocal stacks from embryos expressing mCherry:HIS-58 and endogenous DHC-1:mNeonGreen. Female and male pronuclei were analyzed in 24 hour *zyg-12* and *sur-6* RNAi treatments. n of embryos analyzed per condition is > 14. Time t = 0 s corresponds to NEBD.

Figure S5 | Cytosim-based computational simulations predict 24 hour *sur-6* RNAi effects on cortical Dynein motoring activity are not sufficient to affect centrosome separation. Predicted centrosome-centrosome distances in simulated control embryos and in embryos with a 50% reduction in cortical Dynein motoring velocity. Error bars are SEM and are represented as shaded areas around means of 10 simulation runs per condition. Time $t = 0$ s corresponds to centrosome separation.

FIGURE S1

H2B:GFP + γ -tubulin:GFP

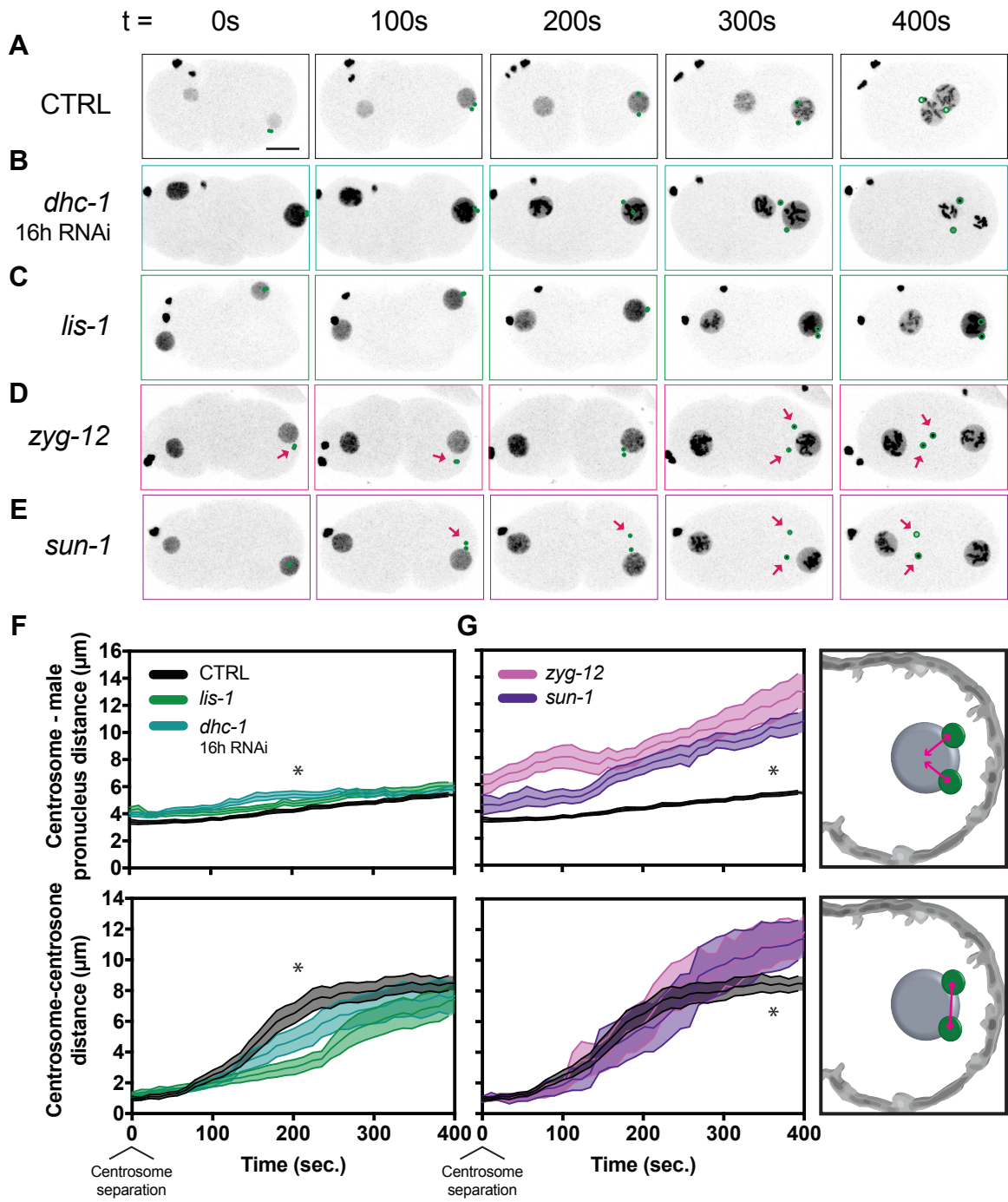


FIGURE S2

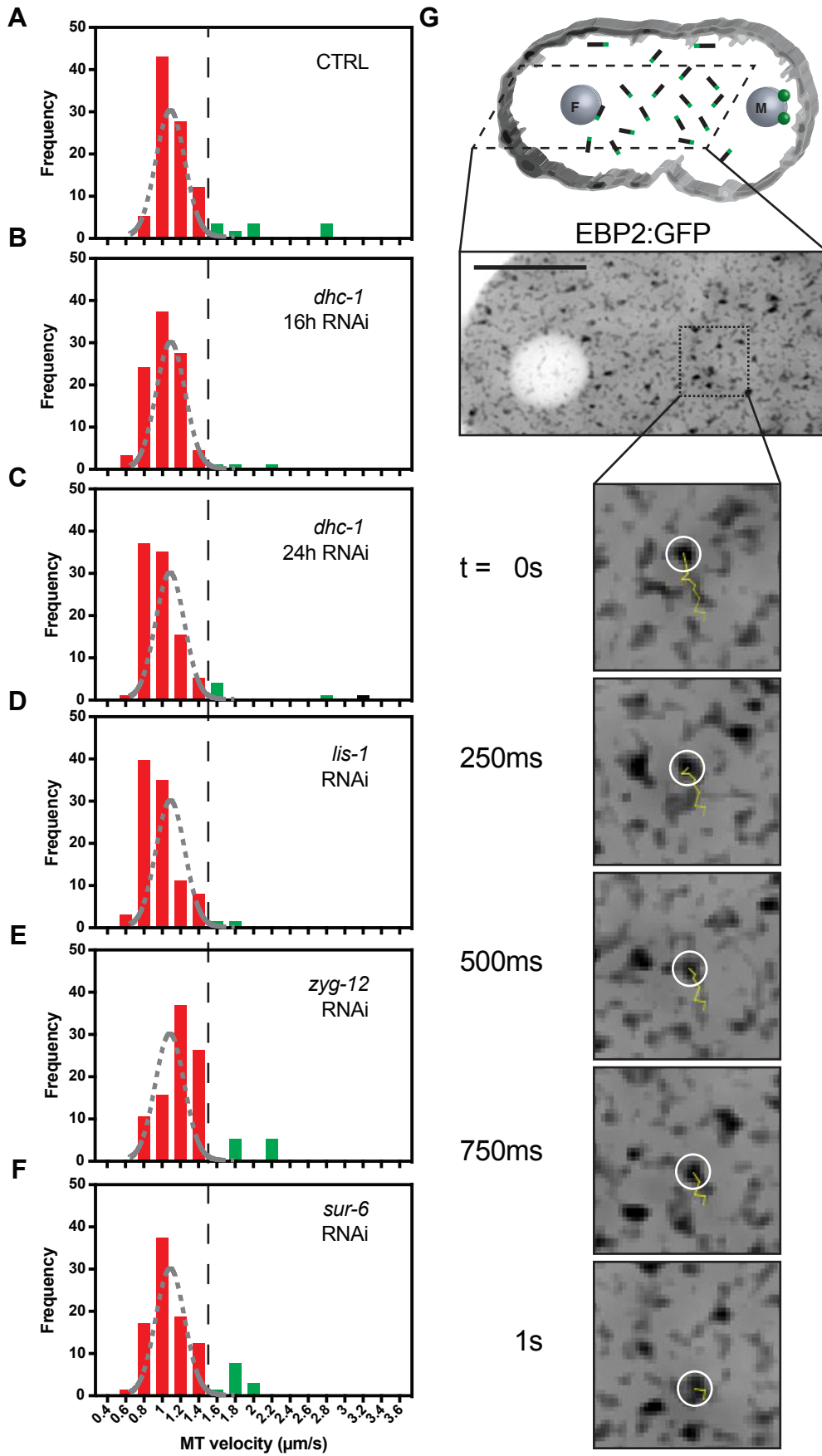


FIGURE S3

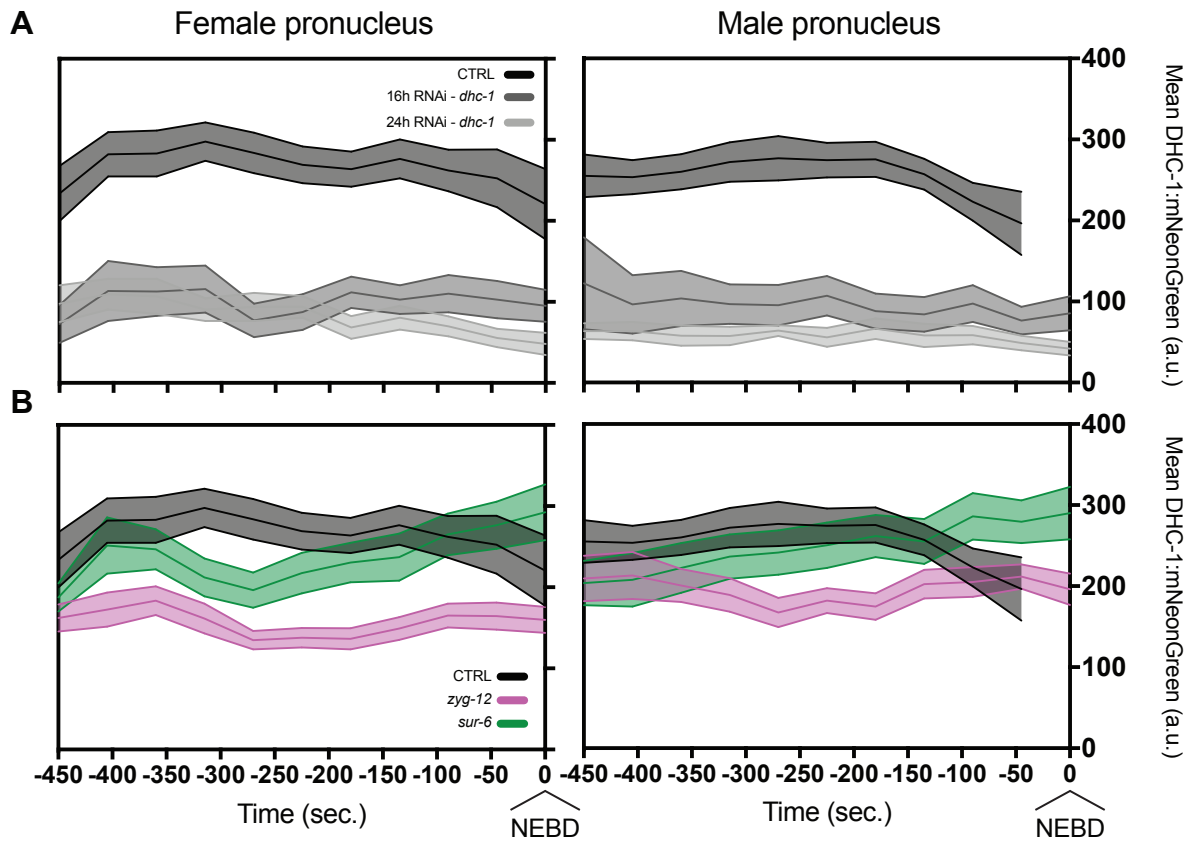


FIGURE S4

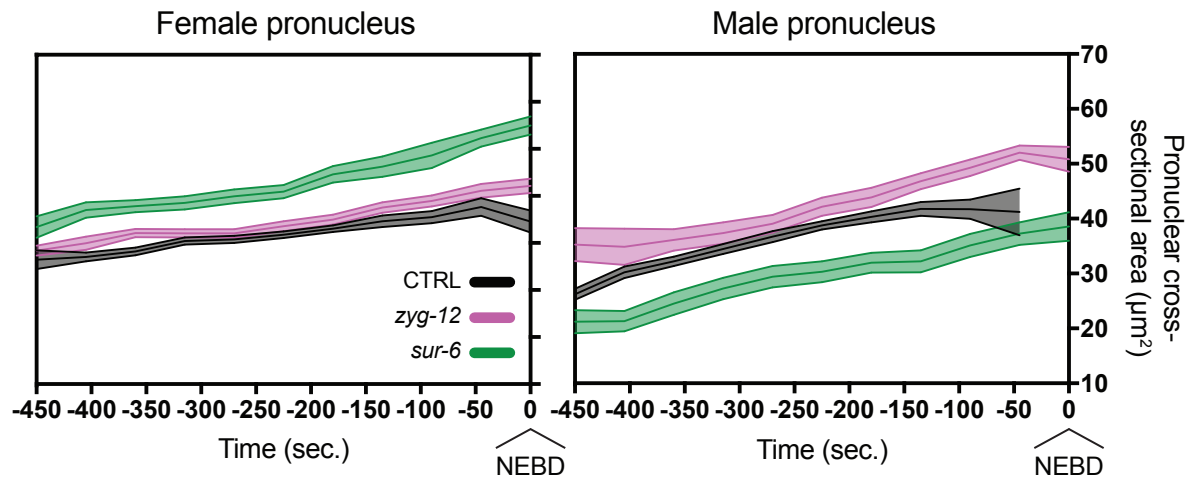


FIGURE S5

