# Supporting Information: Long non-coding RNA *MIR31HG* is a bona fide prognostic marker with colorectal cancer cell-intrinsic properties

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1 Tables

Table S 1: Baseline characteristics of CIT [Marisa 2013], LICR [Jorissen 2009] and Oslo [Sveen 2018] cohorts included in the survival analyses. The numbers in parentheses indicate percentages. adj: adjuvant; CMS: consensus molecular subtypes; MSI/MSS; micro-satellite instable/stable; OS: overall survival; RFS: relapse-free survival

	total (N = 1097)	CIT (N = 562)	LICR (N = $126$ )	OSLO (N = 409)
age-years				
median	69	68	68	72
range	22-97	22-97	30-92	27-97
gender-no.				
male	581 (53)	309 (55)	70 (56)	202 (49)
female	516 (47)	253 (45)	56 (44)	207 (51)
site-no.				
left	510 (47)	342 (61)	46 (37)	122 (30)
right	444 (41)	220 (39)	51 (41)	173 (43)
rectum	138 (13)	0 (0)	27 (22)	111 (27)
stage-no.				
1	148 (13)	40 (7)	24 (19)	84 (21)
2	467 (43)	257 (46)	57 (45)	153 (37)
3	364 (33)	204 (36)	45 (36)	115 (28)
4	118 (11)	61 (11)	0 (0)	57 (14)
adjuvant chemo-no.				
yes	334 (34)	233 (41)	NA	101 (25)
no	637 (66)	329 (59)	NA	308 (75)
OS-months				
median	55	51	NA	60
range	0-200	0-200	NA-NA	0.66-120
RFS-months				
median	46	43	38	55
range	0-200	0-200	1.6-110	0.66-120
mutated-no.				
KRAS	349 (37)	216 (40)	NA	133 (33)
BRAF	118 (13)	49 (10)	NA	69 (17)
TP53	434 (57)	190 (54)	NA	244 (60)
MSS-no.				
MSI	144 (16)	72 (14)	NA	72 (18)
CMS-no.				
1:immune	173 (18)	89 (17)	20 (19)	64 (20)
2:canonical	413 (44)	232 (45)	42 (40)	139 (43)
3:metabolic	144 (15)	69 (13)	19 (18)	56 (17)
4:mesenchymal	215 (23)	126 (24)	25 (24)	64 (20)
not assigned	152 (14)	46 (8)	20 (16)	86 (21)

	crude (univariate)		adjusted (multivariate)			
	HR	CI95%	HR	CI95%	p-value	
MIR31 outlier	2.3	1.7 to 2.9	2.2	1.6 to 3.0	4.4e-06	
BRAF V600	1.1	0.8 to 1.5	1.4	0.9 to 2.2	9.0e-02	
CMS (CMS1)						
CMS2	1.1	0.8 to 1.6	1.7	1.1 to 2.6	2.5e-02	
CMS3	1.0	0.7 to 1.5	1.4	0.8 to 2.3	2.3e-01	
CMS4	1.9	1.4 to 2.7	2.2	1.4 to 3.3	4.5e-04	
stage (I)						
II	1.9	1.2 to 3.1	2.7	1.5 to 5.1	1.8e-03	
III	3.5	2.2 to 5.5	3.8	2.0 to 7.1	3.6e-05	
IV	11.4	7.1 to 18.5	13.1	6.9 to 24.9	3.8e-15	

Table S 2: Relapse-free survival modeled using Cox proportional univariate hazard function with dichotomized *MIR*31*HG* expression. Crude and adjusted values represent univariate and multivariate estimates respectively. MIR31 normal-like, BRAF wt, CMS1 and stage I were used as reference groups. R2=0.19; Wald test p-value<0.001.

	crude (univariate)		adjusted (multivariate)			
	HR	CI95%	HR	CI95%	p-value	
m31	1.3	1.2 to 1.4	1.2	1.1 to 1.3	4.3e-05	
BRAF V600	1.1	0.8 to 1.5	1.4	0.9 to 2.1	9.4e-02	
CMS (CMS1)						
CMS2	1.1	0.8 to 1.6	1.7	1.1 to 2.7	2.2e-02	
CMS3	1.0	0.7 to 1.5	1.4	0.9 to 2.4	1.7e-01	
CMS4	1.9	1.4 to 2.7	2.2	1.4 to 3.3	3.8e-04	
stage (I)						
Π	1.9	1.2 to 3.1	2.7	1.4 to 5.0	2.1e-03	
III	3.5	2.2 to 5.5	3.8	2.0 to 7.2	3.1e-05	
IV	11.4	7.1 to 18.5	12.6	6.6 to 24.0	1.0e-14	

Table S 3: Relapse-free survival modeled using Cox proportional univariate hazard function with continious MIR31HG expression (m31). Crude and adjusted values represent univariate and multivariate estimates respectively. BRAF wt, CMS1 and stage I were used as reference groups. R2=0.18; Wald test p-value<0.001.

Table S 4: Recurrence type according to MIR31HG dichotomized expression for stage II+III colorectal cancers. Numbers refer to cases and column percentages are indicated in parentheses. Data are for Oslo cohort only [Sveen 2018].

	MIR31 normal-like	MIR31 outlier
distant	32 (14%)	10 (28%)
localized	2 (0.86%)	0 (0%)
localized&distant	7 (3%)	3 (8.3%)
none	191 (82%)	23 (64%)

2 Figures and text



Figure S 1: *MIR31HG*, miR-31-5p and miR-31-3p are highly correlated in CRC cell lines and primary tumors (a) Scatter plot shows *MIR31HG*/mir-31-5p expression in CRC cell lines<sup>1</sup>. MSI/MSS samples are indicated with triangles/circles. Cell lines carrying mutations in indicated miRNA-processing genes are highlighted in red.  $\rho$  represents the corresponding Spearman's correlations. (b) Scatter plot shows *MIR31HG*/mir-31-5p expression in TCGA primaries<sup>2</sup>. MSI/MSS samples are indicated with triangles/circles and  $\rho$  is the Spearman's correlation. (c) Scatter plot show miR-31-3p/mir-31-5p expression in CRC cell lines<sup>1</sup>. (d) Scatter plot shows miR-31-3p/mir-31-5p expression in TCGA primaries<sup>2</sup>.  $\rho$  indicates the Spearman's rank correlation. mut; mutated; RPM: reads per millions miRNA mapped; RSEM: RNA-Seq by expectation maximization<sup>3</sup>; vst: variance stabilizing transformed<sup>4</sup>



Figure S 2: Colorectal cancer cell line miR-31-5p and *MIR31HG* expression estimates are consistent across research laboratories and technological platforms and present large ranges in expression values. (a) Scatter plot shows correspondence between in-house smRNA-seq reads per million (RPM) and Gaur *et al.*<sup>5</sup> qRT-PCR based cellular abundance estimates for 7 overlapping cell lines (black circles). Gray circles represent nonoverlapping cell lines with in-house RPM values and copies/cell estimates from the linear least squares regression model (dotted line). The Pearson's correlation coefficient (*r*) for overlapping samples are indicated. (b) Scatter plot depicts in-house Affymetrix microarray and CCLE<sup>6</sup> RNA-seq derived *MIR31HG* expression values for 21 overlapping CRC cell lines with the corresponding Pearson's correlation coefficient (*r*). A pseudo count of 1/10 was added to RNA-seq values to avoid *log* of zero. (c) Barplot illustrates miR-31-5p expression across the NCI-60 panel with CRC samples highlighted in blue. qRT-PCR based cellular abundance estimates are from Gaur *et al.*<sup>5</sup>.





patients (n = 16)

Figure S 3: MIR31 expression levels are maintained in xenografts indicating cancer cell-intrinsic expression. Letters indicate separate patients with black and red indicating primary tumor and PDX samples respectively. Data are from Linnekamp *et al.* and were retrieved from GEO with accession identifier GSE100480<sup>7</sup>. PDX: patient-derived xenograft

### 2.1 MIR31HG and miR-31-5p are analytically robust

To assess analytical robustness, we tested whether MIR31HG/miR-31-5p expression in CRC cell lines is either overtly responsive to environmental changes (*e.g.* differences in culturing between laboratories) or maintained across time and conditions (*i.e.* whether MIR31 activity represents a cell state marker). To answer this, we compared in-house miR-31-5p and *MIR31HG* expression levels against public datasets with overlapping cell lines. For miR-31-5p the Pearson's correlation among seven cell lines quantified by qRT-PCR as part of the NCI60 panel<sup>5</sup> and our smRNA-seq-derived estimates was 0.97 (Supplementary Fig.S2a)<sup>5</sup>. Similarly, the correlation in *MIR31HG* expression between in-house Affymetrix microarrays and the total RNA-seq data from Cancer Cell Line Encyclopedia (CCLE)<sup>6</sup> was 0.94 (*n*=21, Supplementary Fig.S2b)<sup>6</sup>. Thus, differences in *MIR31HG*/miR-31-5p expression between CRC cell lines are robustly recapitulated across molecular levels, research groups and technological platforms.



Figure S 4: *MIR31HG* expression is not associated with age, but MSI-status. (a) Scatterplot for age and *MIR31HG* expression. The *p*-value is from Pearson's correlation test. (b) Beanplot depicts distribution in *MIR31HG* expression for MSS and MSI samples. The horizontal bars represent the group-wise medians and the *p*-value is from Wilcoxon rank sum test.  $\Delta_{mean}$  states the mean  $log_2$  fold-change. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts. MSI/MSS: micro-satellite instable/stable



Figure S 5: MIR31 outlier status stratifies non-adjuvant treated stage II and III primary colorectal cancers. Kaplan-Meier plot shows relapse-free survival for non-adjuvant chemotherapy treated stage II+III pCRCs stratified according to stage and MIR31 status. The *p*-value is from Wald tests for Cox model. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 6: MIR31 outlier status stratifies subgroups of patients with primary colorectal cancers. (a) Kaplan-Meier plot shows relapse-free survival (RFS) for stage II+III pCRCs experiencing distant relapse stratified according to stage and MIR31 status. (b) Kaplan-Meier plot shows RFS for stage II/T4 pCRCs stratified according to MIR31 status. The *p*-value is from Wald tests for Cox model. Data are from Oslo<sup>10</sup> and CIT<sup>8</sup> cohorts. Recurrence type was available only for the Oslo cohort. LICR cohort lacked information on T-stage.



Figure S 7: Relapse-free survival characteristics are largely consistent across the Oslo, CIT and LICR cohorts. (a) Kaplan-Meier plot shows relapse-free survival for stage II+III samples stratified by cohort. (b) Plot visualizes pooled and per cohort hazard ratios with 95% confidence interval associated with MIR31 outlier status. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 8: MIR31 outliers present inferior relapse-free survival within important subgroups. Kaplan-Meier plots for different patient stage II+III subsets. The Kaplan-Meier titles indicate sample subset included. For instance, in the upper left plot, samples were stratified by MSS/MIR31. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 9: MIR31 outliers present inferior overall survival within important subgroups. Kaplan-Meier plots for different patient stage II+III subsets. The Kaplan-Meier titles indicate sample subset included. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 10: MIR31 status is associated with significantly shorter relapse-free survival for stage II–IV colorectal cancers. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 11: MIR31 status is associated with significantly shorter overall survival for stage II–III colorectal cancers. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 12: Kaplan-Meier plots for subgroups per dataset. The Kaplan-Meier titles indicate dataset while subset are indicated to the left of each row. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 13: Kaplan-Meier plots for subgroups per dataset. The Kaplan-Meier titles indicate dataset while subset are indicated to the left of each row. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.



Figure S 14: MIR31 outlier samples are predictive within the poor prognostic Isella *et al.* CRC intrinsic subtype B. (a) Barplot illustrates distribution of CRIS<sup>11</sup> and MIR31 outliers. The *p*-value is from  $\chi^2$ test. (b) Kaplan-Meier plot shows relapse-free survival for stage II+III pCRC patients stratified by CRIS and MIR31 outlier status. (c) Kaplan-Meier plot shows relapse-free survival for poor prognostic CRISB stage II+III pCRC patients stratified by MIR31 and stage. Data are from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts. CRIS: CRC intrinsic subtype; HR: hazard ratio; MAD: median absolute deviation; n/a: not assigned



Figure S 15: Plot visualizes odds ratios with 95% confidence intervals for cell line panel (*n*=78) and selected clinicopathological and molecular variables. *p*-values are from Fisher's exact tests. Panel is a merged set of Oslo<sup>1</sup>, CCLE<sup>6</sup> and Astra-Zeneca [no reference] cell lines. MSI: micro-satellite instable; mut: mutated; wt: wild type

### 2.2 Cell lines are representative models

As for pCRC, cell line MIR31HG expression distribution was heavily right-skewed with mode near microarray background. Using the same thresholding as applied to the pCRCs, 13/78 (17%) cell lines were classified as MIR31 outliers (Manuscript Fig.4a). For gene expression-based undifferentiated state, a proxy for low differentiation grade, 2/49 (4%) colon-like and 11/29 (39%) undifferentiated cell lines displayed MIR31 outlier expression (OR=11 [2.6–141],  $p=2 \times 10^{-4}$ , Fisher's exact test, Supplementary Fig.S15). Considering MSS only, 2/38 (5%) colon-like and 6/16 (38%) undifferentiated cell lines were MIR31 outliers (OR=10 [1.5-120],  $p=6 \times 10^{-3}$ ). Likewise, for 38 samples where details on anatomical sub-site were available, 6/7 (86%) MIR31 outliers were cell lines derived from right-sided tumors compared to 1/19 (5%) for non-outliers (OR=10 [1-525], p=0.03). With regards to MSI and BRAF-status, the highest median MIR31HG expression was observed in MSI/BRAF<sup>V600</sup> samples (Supplementary Fig.S16. However, many distinct MIR31HG outliers were both MSS and BRAF<sup>wt</sup>. Thus, BRAF mutation is unlikely to be the (sole) causal driver of MIR31HG activation. We have previously reported CRC cell line consensus molecular subtype (CMS) classification<sup>10</sup>. As expected based on the pCRCs and differences in differentiation-state, MIR31 status was also significantly associated with CMS (p=<2e-16, Fisher's exact test, Manuscript Fig.2b). Specifically, 0/23 CMS2-like and 20-29% of non-CMS2 subtypes were classified as MIR31 outliers. Summarized, the CRC cell line panel recapitulates MIR31 associations described for pCRCs.

TargetScan records 867 human transcripts with miR-31-5p seed matches whereof 464 show evidence of evolutionary conservation<sup>12</sup>. Despite the large difference in *MIR31HG*/miR-31-5p abundance between the two groups, there was no significant overall depletion of target transcripts in MIR31 outlier samples (Supplementary Fig.S17a). We further tested whether repression could be observed at the protein-level by taking advantage of mass spectroscopy derived protein abundances for 41 overlapping CRC cell lines<sup>13</sup>. As for the RNA-level, miR-31-5p targets showed no trend towards having lower relative protein expression in MIR31 outlier cell lines (Supplementary Fig.S17b).

expression and BRAF/MSI-status



Figure S 16: Boxplot shows *MIR31HG* expression stratified by *BRAF*-mutation and MSI-status. The dotted line represents the dichotomization threshold. Panel is a merged set of Oslo<sup>1</sup>, CCLE<sup>6</sup> and Astra-Zeneca [no reference] cell lines.



Figure S 17: No overall difference in expression of miR-31-5p targets between MIR31 outlier subgroups at either the RNA or protein levels were observed for cell lines. Barcodeplots show enrichment scores for ranked *t*-statistic from differential expression analysis comparing MIR31 outliers against remaining samples. Top panels include only genes with evolutionary conserved miR-31-5p binding sites while lower panels include all predicted targets. Gene lists are from TargetScan<sup>12</sup>. 78 and 41 cell lines were included in the mRNA and protein level analysis, respectively. RNA expression data are a merged set of Oslo<sup>1</sup>, CCLE<sup>6</sup> and Astra-Zeneca [no reference] cell lines. Protein expression data are from Supplementary Information in Roumeliotis *et al.*<sup>13</sup>.



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Figure S 18: *MIR31HG* is located within an interferon gene cluster and cell line *MIR31HG* expression is associated with IFNA responsive genes. (a) Figure shows interferon gene cluster with *MIR31HG* in immediate proximity. (b) Barcodeplot visualizes the differential expression enrichment of genes induced by IFN- $\alpha$ . Gene expression data are for Oslo<sup>1</sup>, CCLE<sup>6</sup> and Astra-Zeneca [no reference] cell lines. Moserle *et al.* HGU133p2 gene expression data of ovarian cancer cell lines exposed to IFN- $\alpha$  were used to identify genes upregulated upon exposure ( $log_2$ fold-change>1 and FDR adjusted-p<0.05, GEO identifier GSE10943)<sup>14</sup>. chr: chromosome; IFNA: interferon- $\alpha$ ; kb: kilobases



Figure S 19: MIR31 outlier samples present upregulation of interferon- $\alpha/\gamma$  signatures compared to non-CMS1 subgroups. Heatmap visualizes results from Camera<sup>15</sup> gene set analysis comparing CMS and MIR31 outliers. Color saturation indicates increasing significance and red and blue relative up- and downregulation, respectively. Gene signatures are from MSigDB Hallmarks<sup>16,17</sup> (v6.2). Gene expression data are from TCGA<sup>2</sup>. CMS: consensus molecular subtypes; EMT: epithelial mesenchymal transition; FDR: false discovery rate<sup>18</sup>; IFNA/G: interferon- $\alpha/\gamma$ ; MAD: median absolute deviation; MSS/MSI; micro-satellite stable/instable; pCRC; primary colorectal cancer



Figure S 20: MIR31 outlier tumors are less stromal than CMS4. (a) Beanplot shows MCPcounter<sup>19</sup> fibroblast infiltration score stratified by CMS and MIR31 outlier expression for TCGA cohort<sup>2</sup>. (b) Plot depicts fibroblast infiltration score for CIT cohort<sup>8</sup>. (c) Beanplot shows MCPcounter<sup>19</sup> cytotoxic T lymphocyte infiltration score stratified by CMS and MIR31 outlier expression. The *p*-values are from Wilcoxon rank sum tests. Gene expression data are from TCGA<sup>2</sup>.



Figure S 21: Among CMS4 tumors, MIR31 outliers exhibit more stromal infiltration than samples with normallike *MIR31HG* expression. (a) Beanplot shows MCPcounter<sup>19</sup> fibroblast infiltration score stratified by CMS and MIR31 outlier expression. (b). Beanplot depicts distributions in single sample gene set enrichment scores for normal colonic fibroblast TGF- $\beta$  response signature stratified by MIR31 outlier expression.<sup>20</sup>. (c) Beanplot shows MCPcounter<sup>19</sup> cytotoxic T lymphocyte infiltration score stratified by CMS and MIR31 outlier expression. The *p*-values are from Wilcoxon rank sum tests. Data include 226 CMS4 samples from CIT<sup>8</sup>, LICR<sup>9</sup> and Oslo<sup>10</sup> cohorts.

## 2.3 Survival models

Adding patient gender, patient age (<=70 vs >70), tumor localization (right vs left/rectum), adjuvant chemotherapy (yes/no), MSI, *CDX2* expression (dichotomized at 15.6 percentile[pilati\_cdx2\_2017]) and *KRAS*-status did not substantially improve the presented multivariable model. Replacing with CRC intrinsic subtypes (CRIS)<sup>11</sup> yielded comparable results (see output below).

```
Call:
```

```
survival::coxph(formula = survFit ~ mir + braf_mut + cms + survival::strata(stage) +
gender + agegrp + side + chemo + msi + cdxneg + kras_mut,
data = pool)
```

n= 735, number of events= 255

(530 observations deleted due to missingness)

	coef	exp(coef)	se(coef)	z	Pr(> z )	
mirMIR31 outlier	0.6549	1.9250	0.1867	3.51	4.5e-04	***
braf_mut1	0.4865	1.6267	0.2587	1.88	6.0e-02	
cmsCMS2	0.5259	1.6919	0.3036	1.73	8.3e-02	
cmsCMS3	0.2810	1.3244	0.3169	0.89	3.8e-01	
cmsCMS4	0.8372	2.3098	0.2773	3.02	2.5e-03	**
<pre>survival::strata(stage)2</pre>	1.0864	2.9635	0.3224	3.37	7.5e-04	***
<pre>survival::strata(stage)3</pre>	1.4975	4.4706	0.3346	4.48	7.6e-06	***
<pre>survival::strata(stage)4</pre>	2.7084	15.0056	0.3411	7.94	2.0e-15	***
gendermale	0.1815	1.1990	0.1329	1.37	1.7e-01	
agegrp(70,100]	0.0818	1.0853	0.1385	0.59	5.5e-01	
sideright	-0.0840	0.9194	0.1532	-0.55	5.8e-01	
chemoyes	-0.3597	0.6979	0.1581	-2.28	2.3e-02	*
msi	-0.1938	0.8239	0.2769	-0.70	4.8e-01	
cdxnegTRUE	0.1426	1.1533	0.2086	0.68	4.9e-01	
kras_mut1	0.0862	1.0900	0.1508	0.57	5.7e-01	
Signif. codes: 0 '***'	1e-03 '**	' 0.01 '*'	0.05 '.'	0.1 '	' 1	
	exp(coef)	) exp(-coe	ef) lower	.95 up	per .95	
mirMIR31 outlier	1.92	5 0.51	1.195	335	2.776	
braf_mut1	1.62	7 0.61	.48 0	980	2.701	
cmsCMS2	1.69	2 0.59	910 0.	933	3.068	
cmsCMS3	1.32	4 0.75	551 0.	712	2.465	
cmsCMS4	2.31	0 0.43	329 1.	341	3.977	
<pre>survival::strata(stage)2</pre>	2.96	3 0.33	374 1.	575	5.575	
<pre>survival::strata(stage)3</pre>	4.47	1 0.22	237 2.	320	8.614	
<pre>survival::strata(stage)4</pre>	15.00	6 0.06	66 7.	689	29.284	
gendermale	1.19	9 0.83	340 0.	924	1.556	
agegrp(70,100]	1.08	5 0.92	214 0.	827	1.424	
sideright	0.91	9 1.08	376 0.	681	1.242	
chemoyes	0.69	8 1.43	330 0.	512	0.951	
msi	0.82	4 1.21	.38 0.	479	1.418	
cdxnegTRUE	1.15	3 0.86	671 0.	766	1.736	
kras_mut1	1.09	0 0.91	.0.	811	1.465	

Concordance= 0.709 (se = 0.017 ) Rsquare= 0.195 (max possible= 0.987 ) Likelihood ratio test= 159 on 15 df, p=<2e-16 Wald test = 173 on 15 df, p=<2e-16 Score (logrank) test = 207 on 15 df, p=<2e-16 Call: survival::coxph(formula = survFit ~ mir + braf\_mut + cris + survival::strata(stage) + gender + agegrp + side + chemo + msi + cdxneg + kras\_mut, data = pool) n= 638, number of events= 236 (627 observations deleted due to missingness) coef exp(coef) se(coef) z Pr(>|z|) mirMIR31 outlier 0.57286 1.77333 0.20474 2.80 5.1e-03 \*\* 0.21801 1.24360 0.25641 0.85 4.0e-01 braf mut1 crisCRISB 0.09177 1.09611 0.21927 0.42 6.8e-01 crisCRISC -0.03360 0.96696 0.21747 -0.15 8.8e-01 crisCRISD 0.05960 1.06141 0.22278 0.27 7.9e-01 crisCRISE -0.17517 0.83932 0.21979 -0.80 4.3e-01 survival::strata(stage)2 0.65210 1.91956 0.28543 2.28 2.2e-02 \* survival::strata(stage)3 1.02528 2.78788 0.30192 3.40 6.8e-04 \*\*\* survival::strata(stage)4 2.38076 10.81308 0.30899 7.71 1.3e-14 \*\*\* 0.17557 1.19193 0.13607 1.29 2.0e-01 gendermale agegrp(70,100] 0.07321 1.07595 0.14711 0.50 6.2e-01 0.04576 1.04682 0.15614 0.29 7.7e-01 sideright -0.14765 0.86273 0.16885 -0.87 3.8e-01 chemoyes -0.631690.53169 0.27041 -2.34 1.9e-02 \* msi cdxnegTRUE 0.21914 1.24501 0.22236 0.99 3.2e-01 0.00858 1.00861 0.16156 0.05 9.6e-01 kras\_mut1 Signif. codes: 0 '\*\*\*' 1e-03 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1 exp(coef) exp(-coef) lower .95 upper .95 mirMIR31 outlier 1.773 0.5639 1.187 2.649 braf\_mut1 1.244 0.8041 0.752 2.056 1.685 crisCRISB 1.096 0.9123 0.713 crisCRISC 0.967 1.0342 0.631 1.481 crisCRISD 1.061 0.9421 0.686 1.643 crisCRISE 0.839 1.1914 0.546 1.291 survival::strata(stage)2 1.920 0.5210 1.097 3.359 survival::strata(stage)3 2.788 0.3587 1.543 5.038 survival::strata(stage)4 10.813 0.0925 5.901 19.813 gendermale 1.192 0.8390 0.913 1.556 agegrp(70,100] 1.076 0.9294 0.806 1.436 sideright 1.047 0.9553 0.771 1.422 chemoyes 0.863 1.1591 0.620 1.201 msi 0.532 1.8808 0.313 0.903 cdxnegTRUE 1.245 0.8032 0.805 1.925 kras\_mut1 1.009 0.9915 0.735 1.384 Concordance= 0.705 (se = 0.018 ) Rsquare= 0.195 (max possible= 0.989 ) Likelihood ratio test= 139 on 16 df, p=<2e-16 Wald test = 161 on 16 df, p=<2e-16Score (logrank) test = 195 on 16 df, p=<2e-16

# 3 R packages

Packages explicitly loaded include beanplot<sup>21</sup>, Biobase<sup>22</sup>, CMSclassifier<sup>23</sup>, genefilter<sup>24</sup>, Greg<sup>25</sup>, kableExtra<sup>26</sup>, knitr<sup>27</sup>, limma<sup>28</sup>, MCPcounter<sup>19</sup>, qwraps2<sup>29</sup>, RColorBrewer<sup>30</sup>; survival<sup>31</sup>, and sva<sup>32</sup>.

## 4 References

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