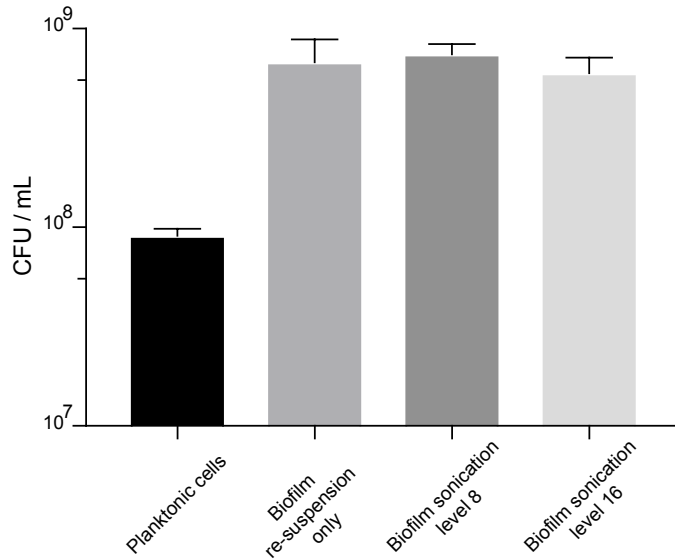


1 Supplemental Materials



2

3 **Supplemental Figure 1. Mab Biofilm Sonication.** CFUs of mature biofilms after
4 re-suspension and sonication. Planktonic cells were used as a control. Levels of
5 sonication represent numbers on the amplitude control knob of a Microson Ultra Sonic
6 Cell Disruptor XL.

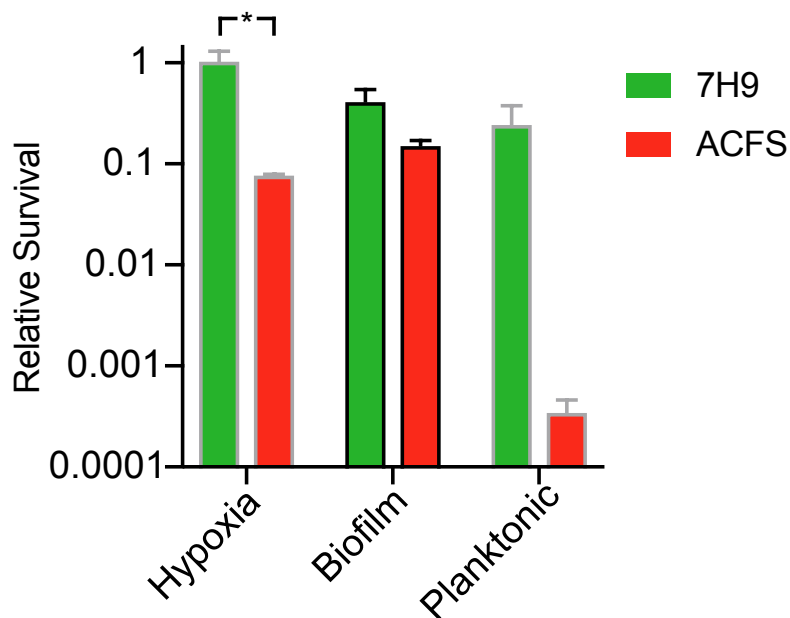
7

8 *Hypoxia.* We performed an assay in order to determine the extent to which hypoxia
9 contributes to antibiotic tolerance in *Mab*. It is known that anaerobic conditions can
10 induce antibiotic tolerance in *P.aeruginosa*, another common CF pathogen (1).
11 Therefore, we aimed to compare the antibiotic tolerance in *Mab* planktonic cells
12 subjected to anaerobic conditions, to antibiotic tolerance in mature biofilms at the
13 bottom of our 24-well plates. We sealed planktonic cultures and left them stationary for
14 13 days, then treated them with cefoxitin and measured survival. We see in
15 supplemental figure 2 that hypoxia does induce antibiotic tolerance in *Mab* when grown

16 in both 7H9 and ACFS media. These data indicate that hypoxia could be one of the
17 conditions that cause the biofilm-associated cells in our assay to be physiologically
18 different from the planktonic cells. The hypoxic cells in ACFS are more sensitive to
19 cefoxitin than those in 7H9 (student's t-test; P-value 0.01344). This implies that either
20 ACFS could retain more dissolved oxygen than 7H9 due to its greater viscosity, or that
21 the nutrient conditions affect the way that cells respond to hypoxia.

22

S2. Hypoxia control in ACFS and 7H9



23

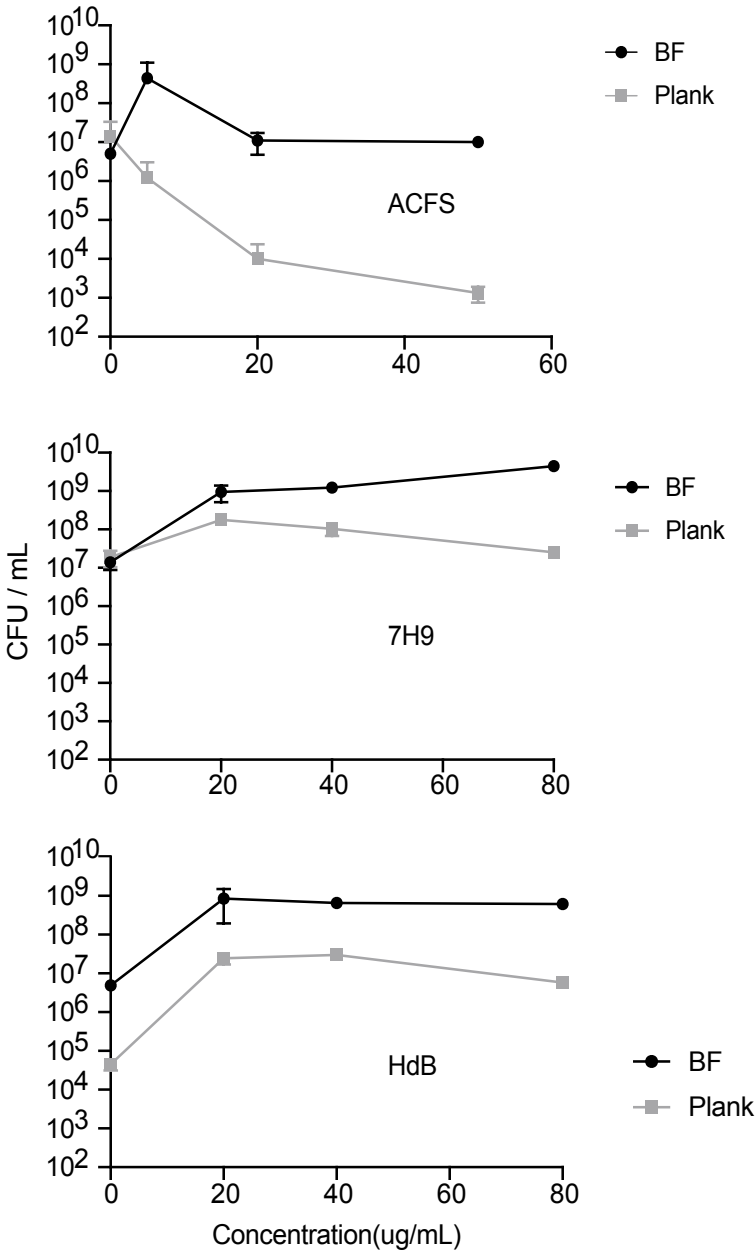
24 **Supplemental Figure 2. Antibiotic sensitivity of Hypoxic planktonic cells**

25 **compared to antibiotic sensitivity in mature biofilms and planktonic cells.** CFUs of
26 hypoxic cells 48 hours after treatment with 80ug/mL of cefoxitin (Hypoxia Assay). CFUs
27 of mature biofilm and planktonic cells 48 hours after treatment with 80ug/mL cefoxitin
28 (data from Fig. 6A, 6B).

29

30 *Antibiotic tolerance in the Mab rough morphotype.* We isolated a “rough” morphotype in
31 order to determine whether the media effects we observe on physiology are restricted to
32 the smooth strains. In supplemental figure 3 (S3) we performed an antibiotic treatment
33 course with mature biofilm and planktonic cells of rough *Mab*. Our original goal was to
34 test the susceptibility of the pellicle, biofilm, and planktonic cells of rough *Mab*, and
35 compare results to the smooth *Mab* data. Due to inconsistencies in pellicle formation
36 within wells, and pellicle dispersion post-treatment, the pellicle data cannot be included.
37 In addition, it must be noted that all results shown in S3 could be due to pellicle
38 dispersal into the biofilm and planktonic populations. Despite dispersal, results do show
39 that the trend in antibiotic susceptibility in smooth *Mab* holds in the rough *Mab*
40 morphotype.

S3. Antibiotic sensitivity in Mab rough strain



41

42

Supplemental Figure 3. Rough strain exhibits comparable antibiotic

43

susceptibility to smooth strain. CFUs of 6-day old biofilms and planktonic cells of

44

rough *Mab* treated with 80 $\mu\text{g/mL}$ of cefoxitin for 48 hours. Compare to cefoxitin-treated

45

smooth strain in all 3 medias (Fig. 6A, 6B, 6C). Pellicle dispersion post-treatment likely

46

causes the increase in CFU between no drug and 20 $\mu\text{g/mL}$.

47

48 **Reference**

- 49 1. **Borriello G, Werner E, Roe F, Kim AM, Ehrlich GD, Stewart PS.** 2004.
50 Oxygen Limitation Contributes to Antibiotic Tolerance of *Pseudomonas aeruginosa* in
51 Biofilms. *Antimicrobial Agents and Chemotherapy* **48**:2659–2664.