

**Supplemental Information**

**Spatial organization of single mRNPs at different stages along the gene expression pathway**

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Rissland and Daniel Zenklusen\*

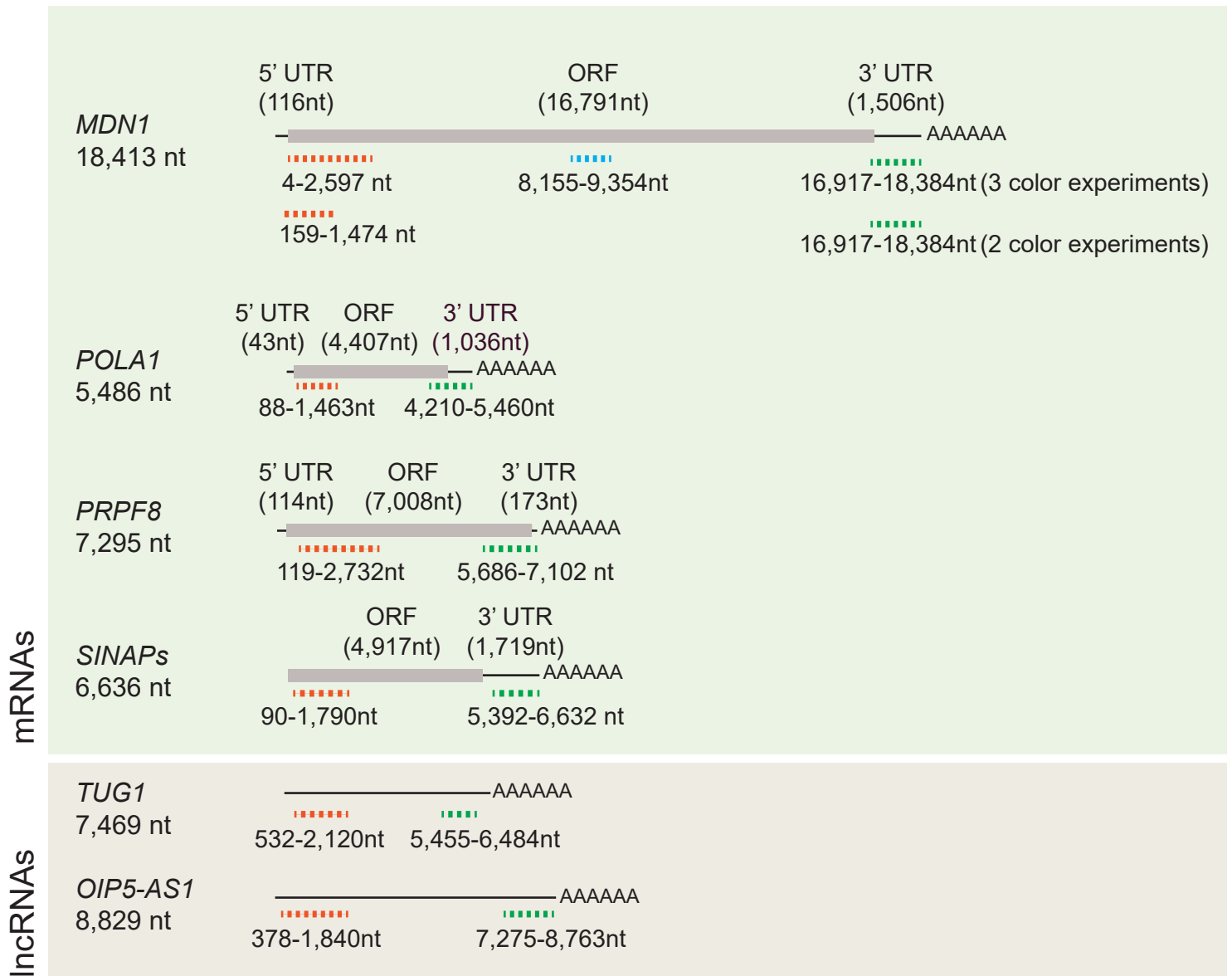
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## **Inventory of Supplemental Information**

Supplemental Figures 1-7

Supplemental Tables 1-3

**Supplementary Figures**



**Figure S1**

**Figure S1: Positions of smFISH probes used in this study.** Cartoons illustrating the positions of the probes used for the different genes used. See Supplemental Table 1 for probe sequences. The transcripts sequences were obtained from ensembl – *MDNI* (ENST00000369393), *POLA1* (ENST00000379068) and *PRPF8* (ENST00000304992) mRNAs and *TUG1* (ENST00000644773) and *OIP5-AS1* (ENST00000500949) lncRNAs.



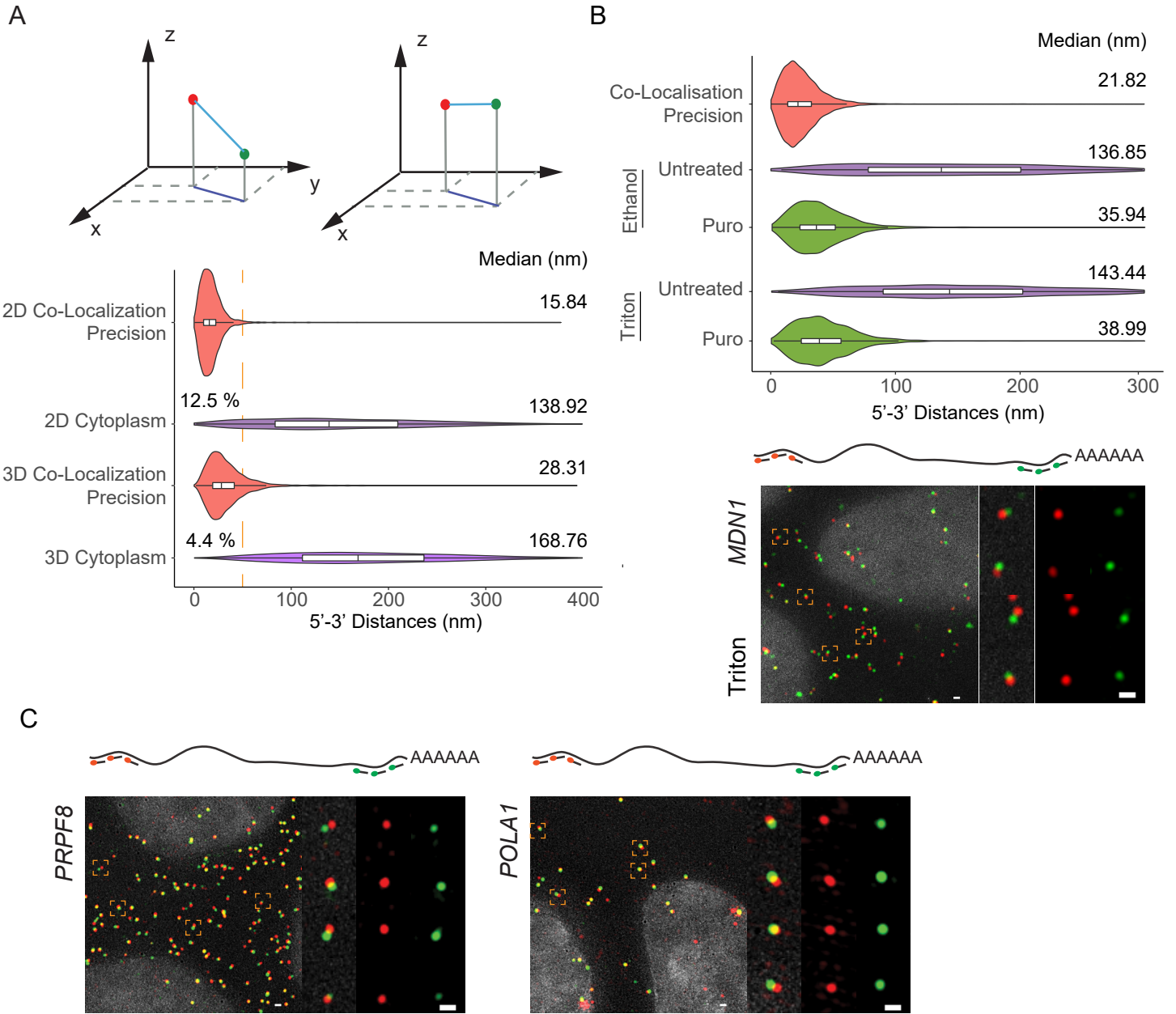
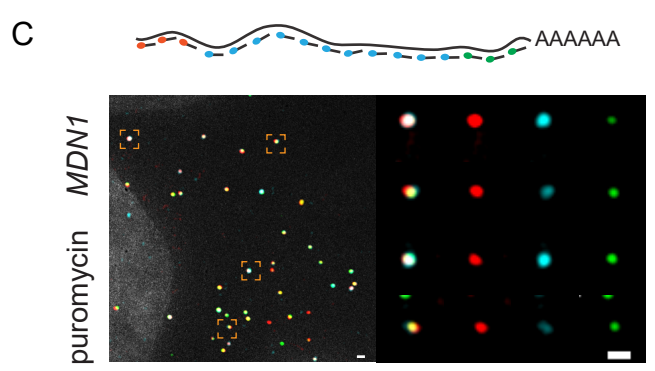
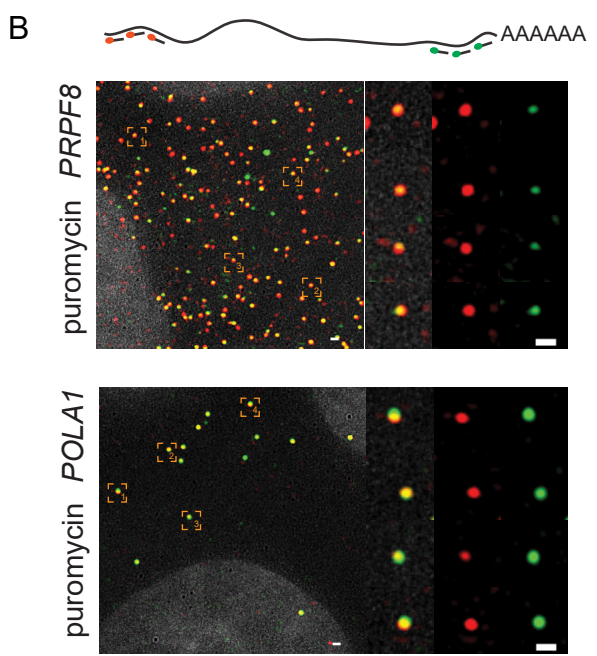
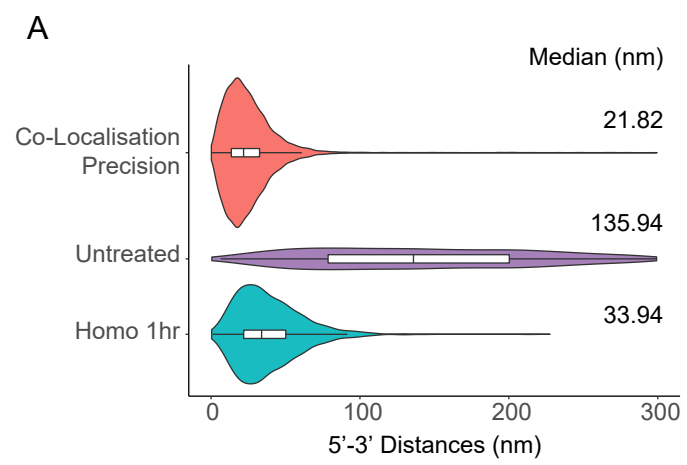


Figure S2

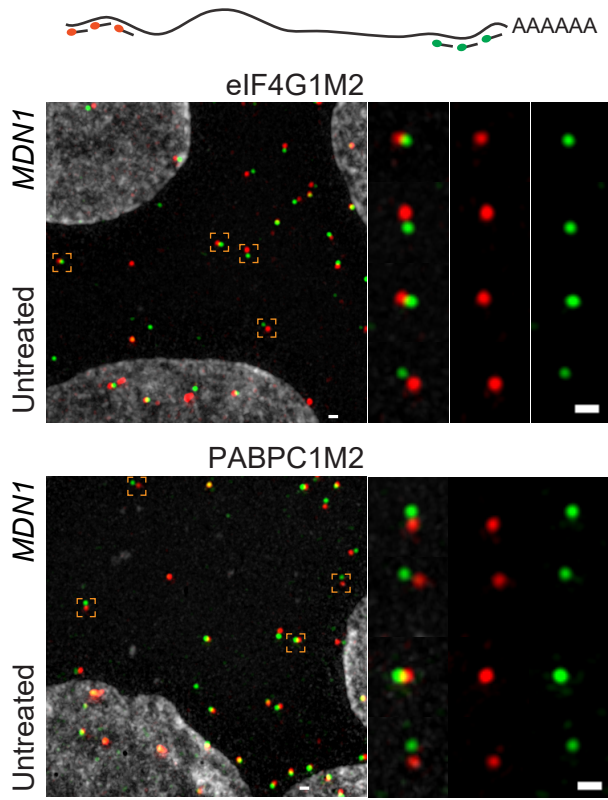
**Figure S2 Related to Figure 1: Visualizing mRNP conformations for *MDN1*, *POLA1* and *PRPF8* mRNAs and measurement of 5'-3' distances for *MDN1* mRNAs in 2D, 3D and in cells permeabilized using either TritonX or Ethanol** (A) Cartoon illustrating how 2D projection alters 5'-3' distances measured (above) and violin plots showing distance distribution of co-localization precision and 5'-3' distances for *MDN1* mRNAs calculated in 2D and 3D. Dotted line delineates the percentage of *MDN1* mRNAs with 5'-3' distances less than 50nm. (B) Violin plots showing distance distribution of co-localization precision (Probe Set#1, Table S3) and 5'-3' distances for *MDN1* mRNAs (Probe Set#2, Table S3) in cells permeabilized with either Ethanol or TritonX-100 (above) and smFISH images using hybridizing to the 5' and 3' ends of *MDN1* mRNAs in cells permeabilized with TritonX-100 (below), (C). smFISH images using probes hybridizing to the 5'(red) and 3'(green) ends of *PRPF8* (Probe Set#5, Table S3) and *POLA1*(Probe Set#6, Table S3) mRNAs in paraformaldehyde fixed HEK293 cells. Scale bars, 500 nm). Nuclei are visualized by DAPI staining (grey). Magnified images of individual RNAs marked by dashed squares are shown on the right. Schematic position of probes shown on top. White box plot inside the violin plot shows first quartile, median and third quartile. Median distances are shown on the right.



**Figure S3**

**Figure S3 Related to Figure 2: 5'-3' distance measurements for *MDN1* mRNA upon treatment with homoharringtonine and visualizing mRNP conformation of single *POLA1*, *PRPF8* and *MDN1* mRNAs when treated with puromycin.** (A) Violin plots showing distance distribution of co-localization precision (Probe Set#1, Table S3) and 5'-3' distances for *MDN1* mRNAs (Probe Set#2, Table S3) determined by Gaussian fitting from untreated and homoharringtonine (1hr) treated cells (B) smFISH images using probes hybridizing to the 5' and 3' ends of *PRPF8* (Probe Set#5, Table S3) and *POLA1* (Probe Set#6, Table S3) mRNAs in paraformaldehyde fixed HEK293 cells treated with puromycin (10 min, 100 µg/ml) (C) smFISH using 5' (red), 3' (green), and tiling (cyan) for *MDN1* mRNA (Probe Set#3, Table S3) in paraformaldehyde fixed HEK293 cells treated with puromycin (10 min, 100 µg/ml). Nuclei are visualized by DAPI staining (grey). Magnified images of individual RNAs marked by dashed squares are shown on the right. Schematic position of probes shown on top. Scale bars, 500 nm. White box plot inside the violin plot shows first quartile, median and third quartile. Median distances are shown on the right.

A



B

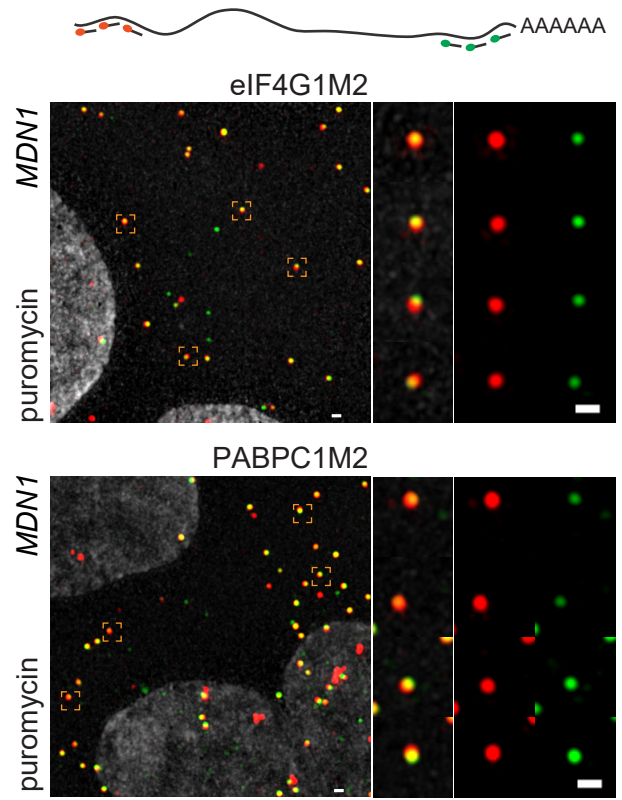


Figure S4

**Figure S4 Related to Figure 3: Visualizing mRNP conformations in mutant cell lines of PABPC1 and eIF4G1.** (A) smFISH images using probes hybridizing to the 5' (red) and 3'(green) ends (Probe Set#2, Table S3) of *MDN1* mRNA in mutant cell lines of eIF4G1 and PABPC1 in paraformaldehyde fixed HEK 293 cells. (B) smFISH images using probes hybridizing to the 5' (red) and 3' (green) ends of *MDN1* mRNA in mutant cell lines of eIF4G1 and PABPC1 in HEK 293 cells treated with puromycin (10 min, 100  $\mu$ g/ml). Nuclei are visualized by DAPI staining (grey). Magnified images of individual RNAs marked by dashed squares are shown on the right. Schematic position of probes shown on top. Scale bars, 500 nm.

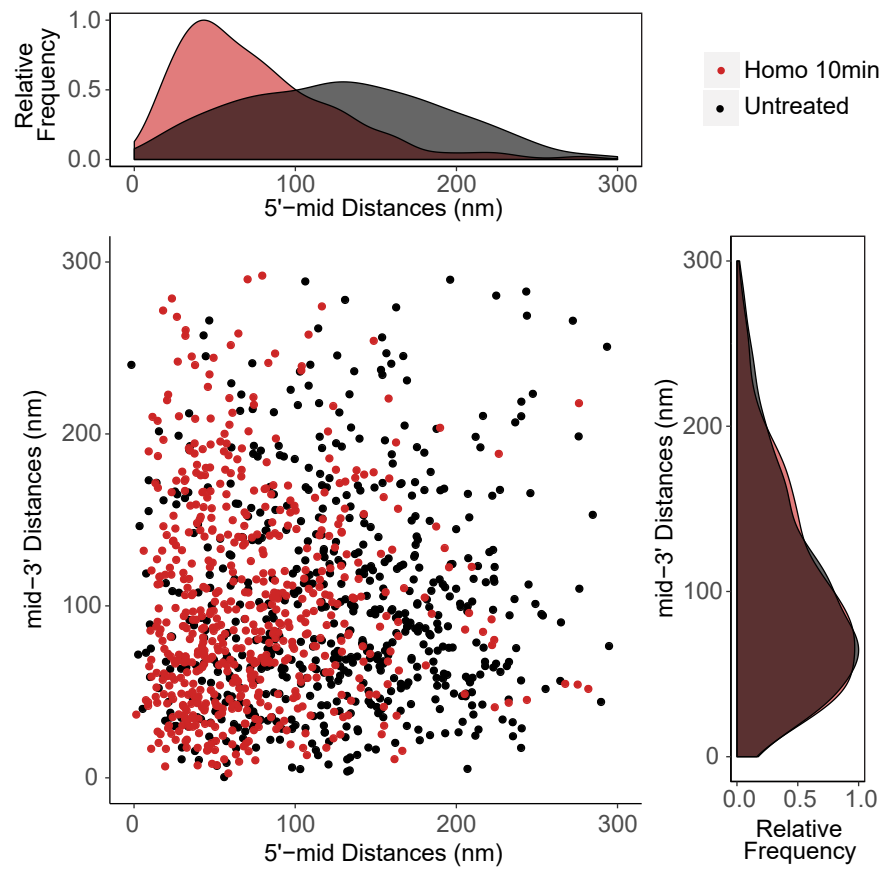
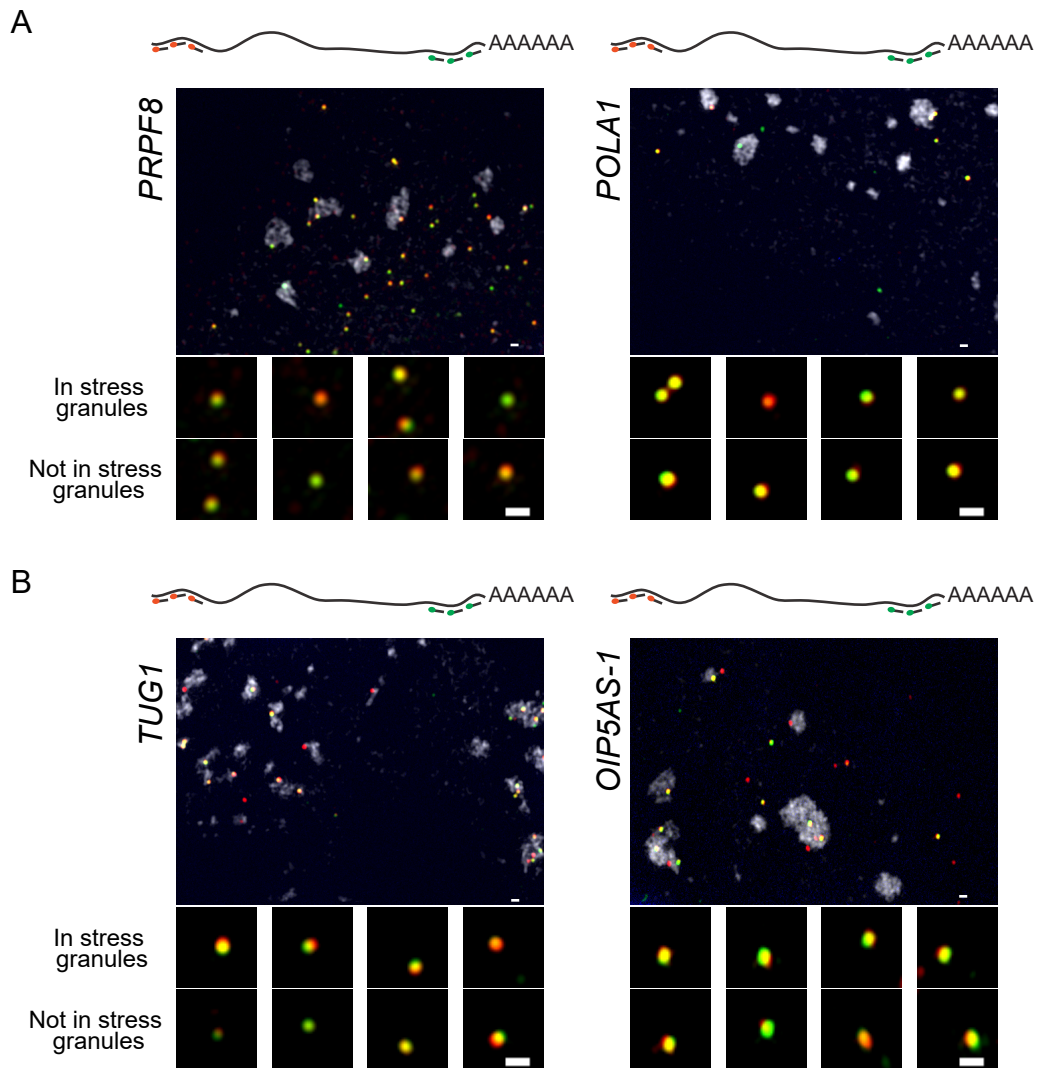


Figure S5

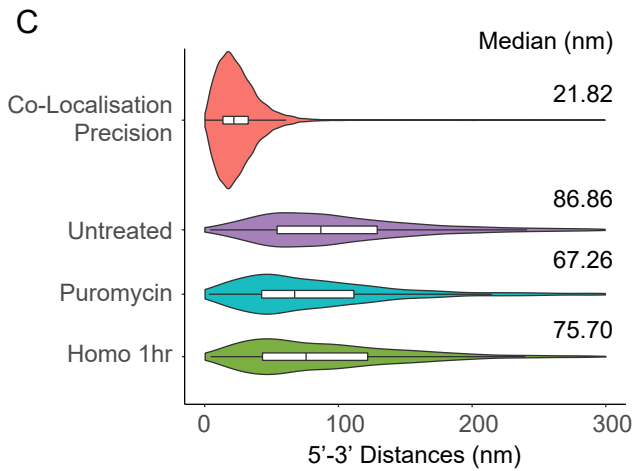
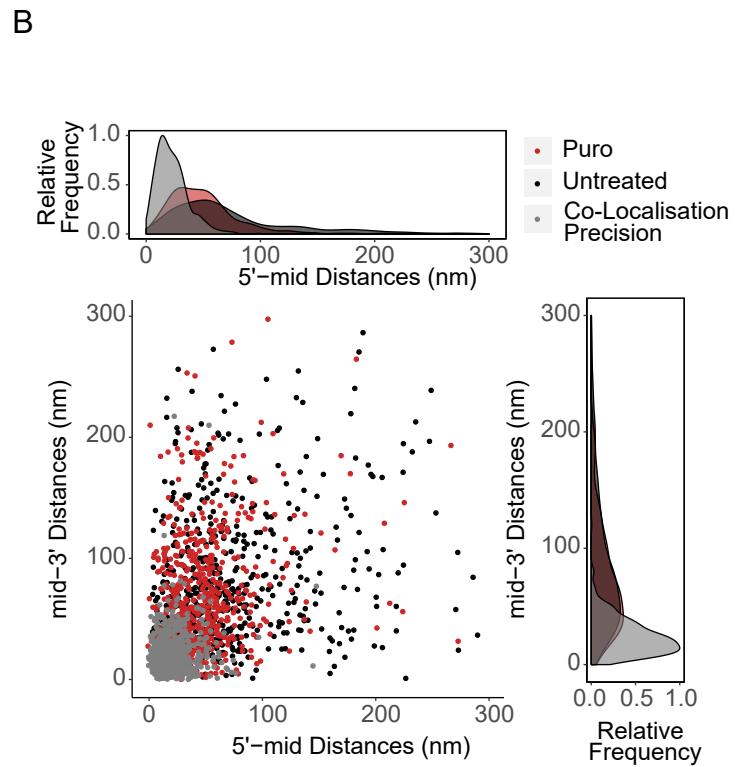
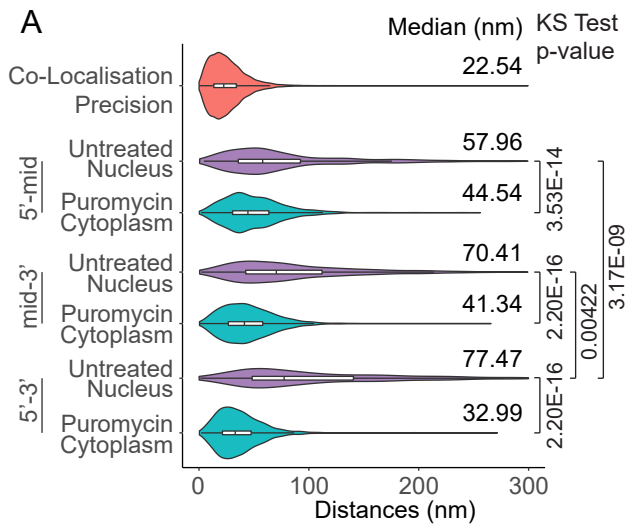
**Figure S5 Related to Figure 4: Compaction of the 5' end is altered upon a pulsed homoharringtonine treatment for 10min.** Scatter plot showing 5'-mid and mid-3' distances for individual cytoplasmic *MDN1* mRNAs from untreated cells (black) and cells treated with homoharringtonine (100 $\mu$ g/ml, 10min) (red). Frequency distribution are shown on top and on the right.





**Figure S6**

**Figure S6 Related to Figure 5: mRNA and lncRNA compaction and accumulation in stress granules.** smFISH visualizing 5' and 3' ends of *PRPF8* (Probe Set#11, Table S3) and *POLAI* (Probe Set#12, Table S3) mRNAs (A) or *TUG1* (Probe Set#13, Table S3) and *OIP5-ASI* (Probe Set#14, Table S3) lncRNAs (B) in U2OS cells treated with arsenite (1-hour, 2 mM). Only a selected cytoplasmic region of cells is shown. Stress granules are visualized using an oligo dT probe (grey). Magnified images of individual RNAs localized inside or outside of stress granules are shown on the bottom of the images. Schematic position of probes shown on top. For *POLAI* mRNAs and *OIP5-ASI* lncRNAs, not all magnified single RNAs shown in the bottom are from the corresponding image above. Scale bars, 500 nm.



**Figure S7**

**Figure S7 Related to Figure 6: Compaction of nuclear MDN1 mRNA upon puromycin or homoharringtonine treatment.** (A) Violin plots showing distance distribution of co-localization precision (Probe Set#1, Table S3) and 5'-3' distances for *MDN1* mRNAs (Probe Set#2, Table S3) determined by Gaussian fitting from nuclear mRNAs in untreated cells and cytoplasmic mRNAs in cells treated with puromycin (100µg/ml, 10 min) (Probe Set#4, Table S3). (B) Scatter plot showing 5'-mid and mid-3' distances for individual nuclear *MDN1* mRNAs from untreated cells (black) and cells treated with puromycin (100µg/ml, 10min) (red). Co-localization precision is shown in grey. Frequency distribution are shown on top and on the right. (C) Violin plots showing distance distribution of co-localization precision (Probe Set#1, Table S3) and 5'-3' distances for *MDN1* (Probe Set#2, Table S3) mRNAs determined by Gaussian fitting from untreated, puromycin (100µg/ml, 10min) or homoharringtonine (100µg/ml, 1 hour) treated HEK293 cells. White box plot inside the violin plot shows first quartile, median and third quartile. Median distances and p-values calculated using Kolmogorov-Smirnov test are shown on the right.

## Supplementary Tables

**Table S1 Related to Figure 3:** Primers used for making CRISPR/Cas9 cells lines

Name	Sequence (5'-3')	Purpose
PABPC1 M161A gRNA 1 Fw	CACCGAATCTGTTAGCCATCTAAC C	Guide RNA targeting PABPC1; annealed and ligated into pX330
PABPC1 M161A gRNA 1 Rev	AAACGGTTAGATGGCTAACAGAT T C	
PABPC1 region Fw NotI	GCGCACTAGCGGCCGCGAGGAAG CGTTCAACTGTGA	To amplify 5' arm region of PABPC1
PABPC1 Not I 5' arm Rev guide 1 XhoI	GCGCACTAGCGGCCGCCTCGAGA CCTGGATATTTGTGAAATAAAG	
PABPC1 BamHI 3' arm Fw guide 1	CGCGGATCCTAGATGGCTAACAG ATTGTCTCTC	To amplify 3' arm region of PABPC1
PABPC1 region Rev BamHI	CGCGGATCCTTGGTCAGGCTGGTC TCAA	
PABPC1 mutation Fw	GAAAGAGCTATTGAAAAAATGAA TGGAGCGCTCCTAAATGATCGCA AAGTATTTGTTGG	Internal primers for stitch PCR to make PABPC1 mutation
PABPC1 mutation Rev	CCAACAAATACTTTGCGATCATT AGGAGCGCTCCATTCATTTTTTCA ATAGCTCTTTC	
PABPC1 region Fw	GGCGAGAGATTGCGTCAAGAA	Screening for puromycin cassette insertion/excision
PABPC1 region Rev	CCCTGGTAACAGGCATTTGTGAG	
PABP seq Fw1	GCAATATGGAATTCTTTTATATG	To sequence PCR products from PABPC1 mutant cell line screening
PABP seq Fw2	GGAAGTGTGCAGTAATGGATATC	
PABP seq Rev1	CAATCTTGTCGCCCAGACTGG	
eIF4G1 mutant gRNA 1 Fw	CACCGTGCTGCTGGGACATTGTGC	Guide RNA targeting eIF4G1; annealed and ligated into pX330
eIF4G1 mutant gRNA 1 Rev	AAACGCACAATGTCCCAGCAGCA C	
eIF4G1 5' HR Arm Fw NotI	GCGCACTAGCGGCCGC GAGACAGGAAGTAGACTCAAG	To amplify 5' arm region of eIF4G1
eIF4G1 5' HR Arm Rev NotI XhoI	GCGCACTAGCGGCCGC CTCGAG CAATGTCCCAGCAGCACCTGACC	
eIF4G1 3' HR Arm Fw BamHI	CGC GGATCC TGCCGGAAAGAGCAGTACTTG	To amplify 3' arm region of eIF4G1
eIF4G1 3' HR Arm Rev BamHI	CGC GGATCC GGCACCTATTCTGGGCACC	
eIF4G1 mutation Fw	TGCAGCCGCTGCCGCGGGAGCAA TCTGGGGTGGCTGGTTC	Internal primers for stitch PCR to make eIF4G1 mutation
eIF4G1 mutation Rev	CGCCAGTGGGAAACTGCTGCACCCCTTGG GCTGGATAGTAGG	

eIF4G1 region Fw	GTAGTCGCACAGTCTTGGCTC	Screening for puromycin cassette insertion/excision
eIF4G1 region Rev	GAGTCCAGGGCAGAACAGAC	
4G seq Rev1	CACCCCTCGTAGGCAGGCACTC	To sequence PCR products from eIF4G mutant cell line screening
4G seq Fw 1	CAGAGTATGTGTGTACATGTTG	
4G seq Rev2	CTTCCTCGCTAGGCACTTCAG	
4G seq Rev 3	CCAGCAGTCCCCAAGTCAGTGG	

**Table S2 Related to Figure S1:** List of smFISH probes used (see attached excel file)

**Table S3 Related to Figure S1 and Table S2: Probe and antibody combinations used**

<b>Probe Set #</b>	<b>Experiment</b>	<b>Combination of probes/antibodies and dyes used (From Table S2)</b>
1	MDN1 middle alternating probes	MDN1 middle odd – Cy5 MDN1 middle even – Cy3
2	MDN1 5'-3'	MDN1 5' -Cy5 MDN1 3' - Dy550
3	MDN1 5'-tiling-3'	MDN1 5'+ MDN1 5' additional – Dy488 MDN1 tiling – Cy5 MDN 3' – Dy550
4	MDN1 5'-middle-3'	MDN1 5'+ MDN1 5' additional – Dy488 MDN1 middle – Cy5 MDN 3' – Dy550
5	PRPF8 5'-3'	PRPF8 5' -Cy5 PRPF8 3' - Cy3
6	POLA1 5'-3'	POLA1 5' -Cy3 POLA1 3' - Cy5
7	TUG1 5'-3'	TUG1 5' -Cy5 TUG1 3' - Cy3
8	OIP5-AS1 5'-3'	OIP5-AS1 5' -Cy5 OIP5-AS1 3' - Cy3
9	MDN1 5'-3'-dT	MDN1 5' -Cy5 MDN1 3' - Dy550 dT – Cy2
10	MDN1 tiling-3'-dT	MDN1 tiling + MDN1 tiling additional – Cy5 MDN1 3' – Dy550 dT - Cy2
11	PRPF8 5'-3'-dT	PRPF8 5' -Cy5 PRPF8 3' - Cy3 dT – Cy2
12	POLA1 5'-3'-dT	POLA1 5' -Cy3 POLA1 3' - Cy5 dT – Cy2
13	TUG1 5'-3'-dT	TUG1 5' -Cy5 TUG1 3' - Cy3 dT – Cy2
14	OIP5-AS1 5'-3'-dT	OIP5-AS1 5' -Cy5 OIP5-AS1 3' - Cy3 dT – Cy2
15	SINAPs 5'-3'	SuntagV4 5' – Quasar 570 MS2v5 3' – Cy5 Chicken anti-GFP Antibody Goat-Anti Chicken Alexa 488