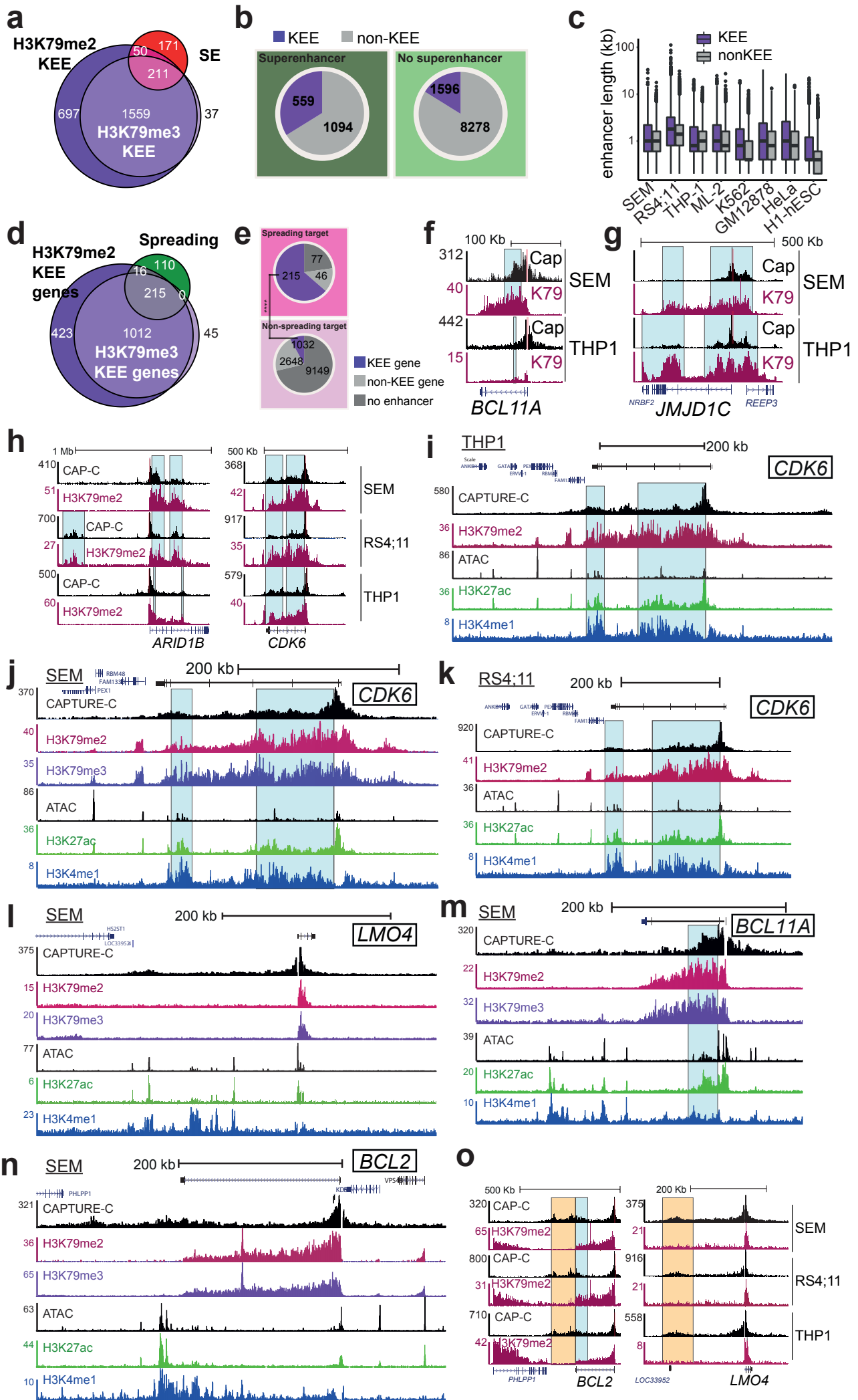


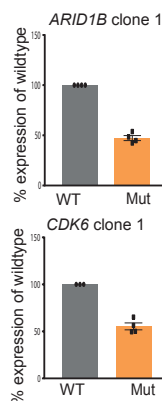
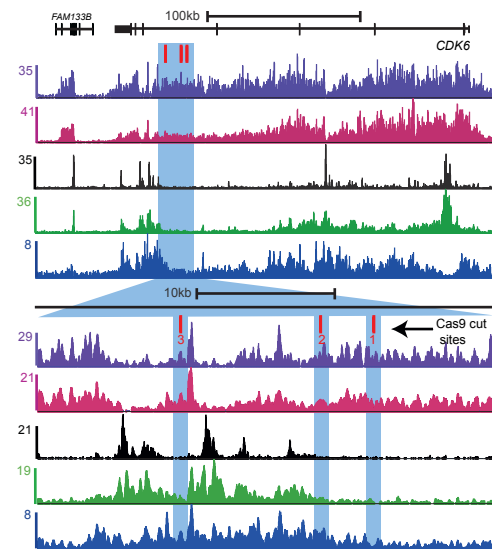
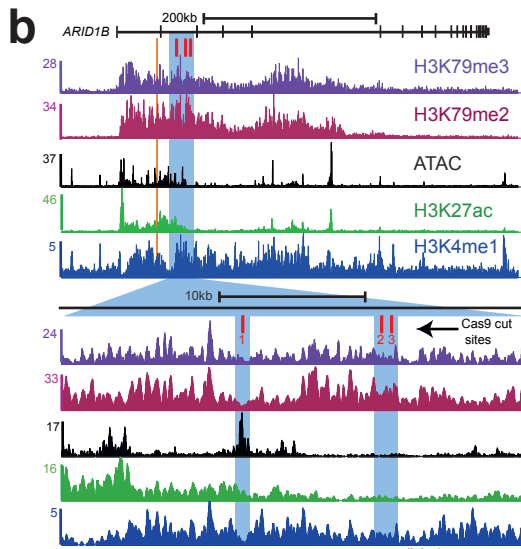
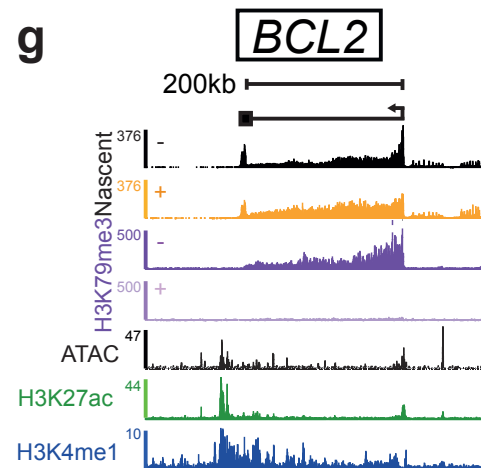
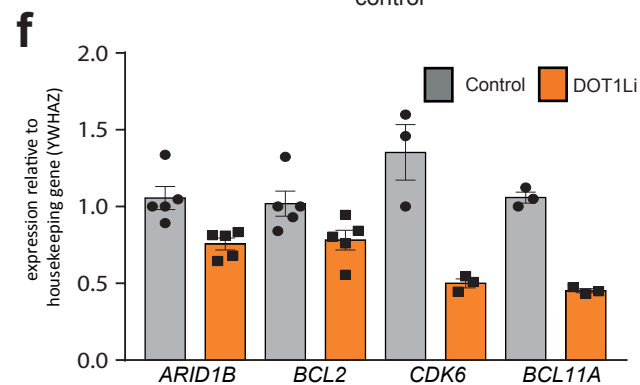
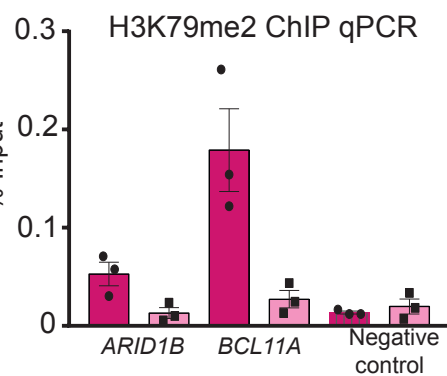
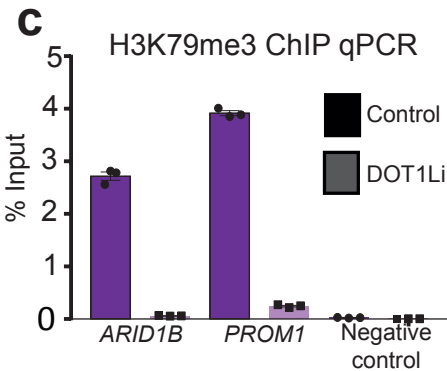
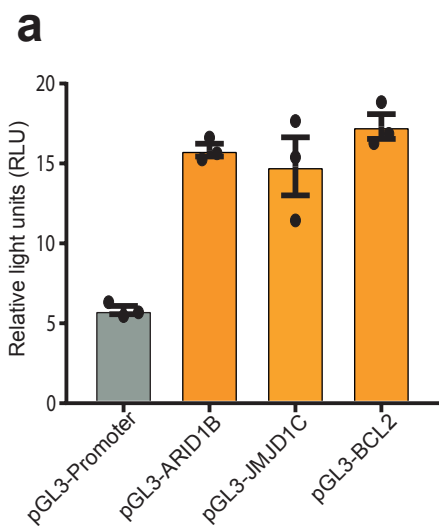
DOT1L inhibition reveals a distinct subset of enhancers dependent on H3K79 methylation

Supplementary information

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Supplementary Figure 1. H3K79me_{2/3} marks a subset of enhancers **(a)** Overlap between KEEs identified using H3K79me₂ and H3K79me₃ ChIP-seq in SEM cells and super-enhancers (SE). Numbers inside the red circle reflect the number of super-enhancers within each overlap group. **(b)** Number of KEEs and non-KEEs that overlap with super-enhancers or non-super-enhancers. **(c)** Lengths of KEEs (purple) and non-KEEs (gray) in different cell types. Box plots show interquartile range; center line represents the median value. **(d)** Overlap between KEE-genes identified using H3K79me₂ and H3K79me₃ ChIP-seq in SEM cells and spreading target genes. **(e)** Proportion of MLL-AF4 spreading genes that are also a KEE-gene (purple), non-KEE-gene (light gray), or not annotated with an enhancer (dark gray) (**** = $p < 0.0001$, Fisher's exact test) in SEM cells. **(f-g)** Capture-C signal and level of H3K79me₂ at *BCL11A* and *JMJD1C* in SEM and THP1 cells. Blue boxes represent KEE regions, red line shows location of Capture-C probe. **(h)** Capture-C and H3K79me₂ ChIP-seq in SEM, RS4;11 and THP1 cells at *ARID1B* and *CDK6*. Blue boxes represent KEE regions. **(i-k)** Capture-C, ChIP-seq and ATAC-seq at *CDK6* in THP1, SEM and RS4;11 cells. Blue boxes represent KEE regions **(l)** Capture-C, ChIP-seq and ATAC-seq at *LMO4* in SEM cells. **(m)** Capture-C, ChIP-seq and ATAC-seq at *BCL11A* in SEM cells. **(n)** Capture-C, ChIP-seq and ATAC-seq at *BCL2* in SEM cells. **(o)** Capture-C and H3K79me₂ ChIP-seq in SEM, RS4;11 and THP1 cells at *BCL2* and *LMO4*. Blue boxes represent KEE regions, orange boxes represent non-KEE regions.



ARID1B clone 1

WT GGCTGCTGGACTCACTGGTATTCCCCAGGCTGGAAGTGGCTCTCCGTTACAGGGCAGCCACAGCTGTCTGGTGA

mut allele 1 GGCTGCTGGACTCACTGGTATTCCCCAGGCTGGAAGTGGCTCTCCGTT

Cut site 2

WT ACAGAGTGAGAGAATCCGTATCAAAAAGCAGCCTGATGTTGTGACTCACACCCATAATCCAGCACTTTGGAGGCTGA

mut allele 1 ACAGAGTGAGAGAATCCGTATCAAAAAGCAGCCTGATGTTGTGACTCACACCCATAATCCAGCACTTTGGAGGCTGA

Cut site 3

WT GGAGGGGAGGAACATGGGGGAAATGCCCTTAAAACGGCTAAGCAGCAATAGATACCTTGAGTCATTTACCCCTCCA

mut allele 1 GGAGGGGAGGAACATGGGGGAAATGCCCTTAAAACGGCTAAGCAGCAATAGATACCTTGAGTCATTTACCCCTCCA

mut allele 2 GGAGGGGAGGAACATGGGGGAAATGCCCTTAAAACGGCTAAGCAGCAATAGATACCTTGAGTCATTTACCCCTCCA

CDK6 clone 1

WT GCAACATCCTCGTCAGCACTTCTGGTCCAGATTCAAAGGTATGCGTACTTCTTGGACCTGCTACTCTCCATCTTTTCT

mut allele 1 GCAACATCCTCGTCAGCACTTCTGGTCCAGATTCAAAGGTATGCGTACTTCTTGGACCTGCTACTCTCCATCTTTTCT

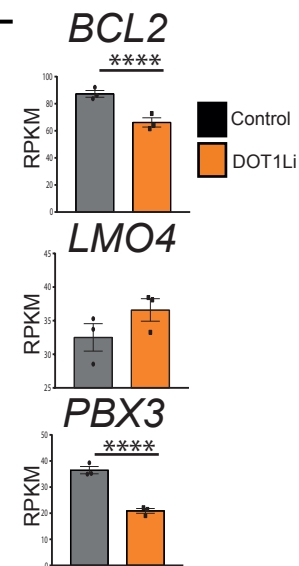
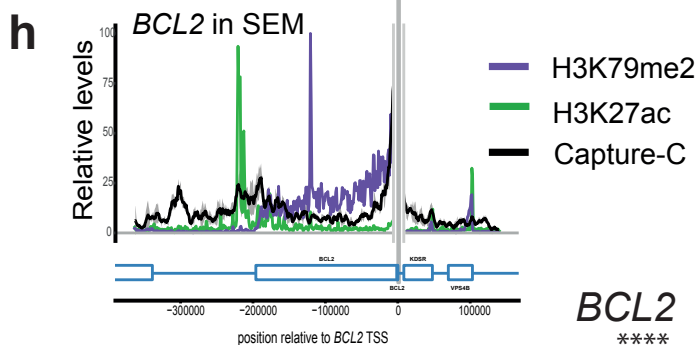
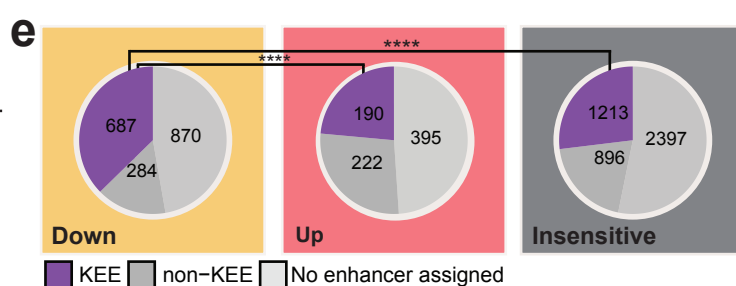
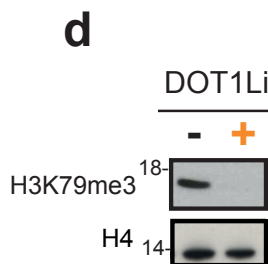
mut allele 2 GCAACATCCTCGTCAGCACTTCTGGTCCAGATTCAAAGGTATGCGTACTTCTTGGACCTGCTACTCTCCATCTTTTCT

Cut site 2

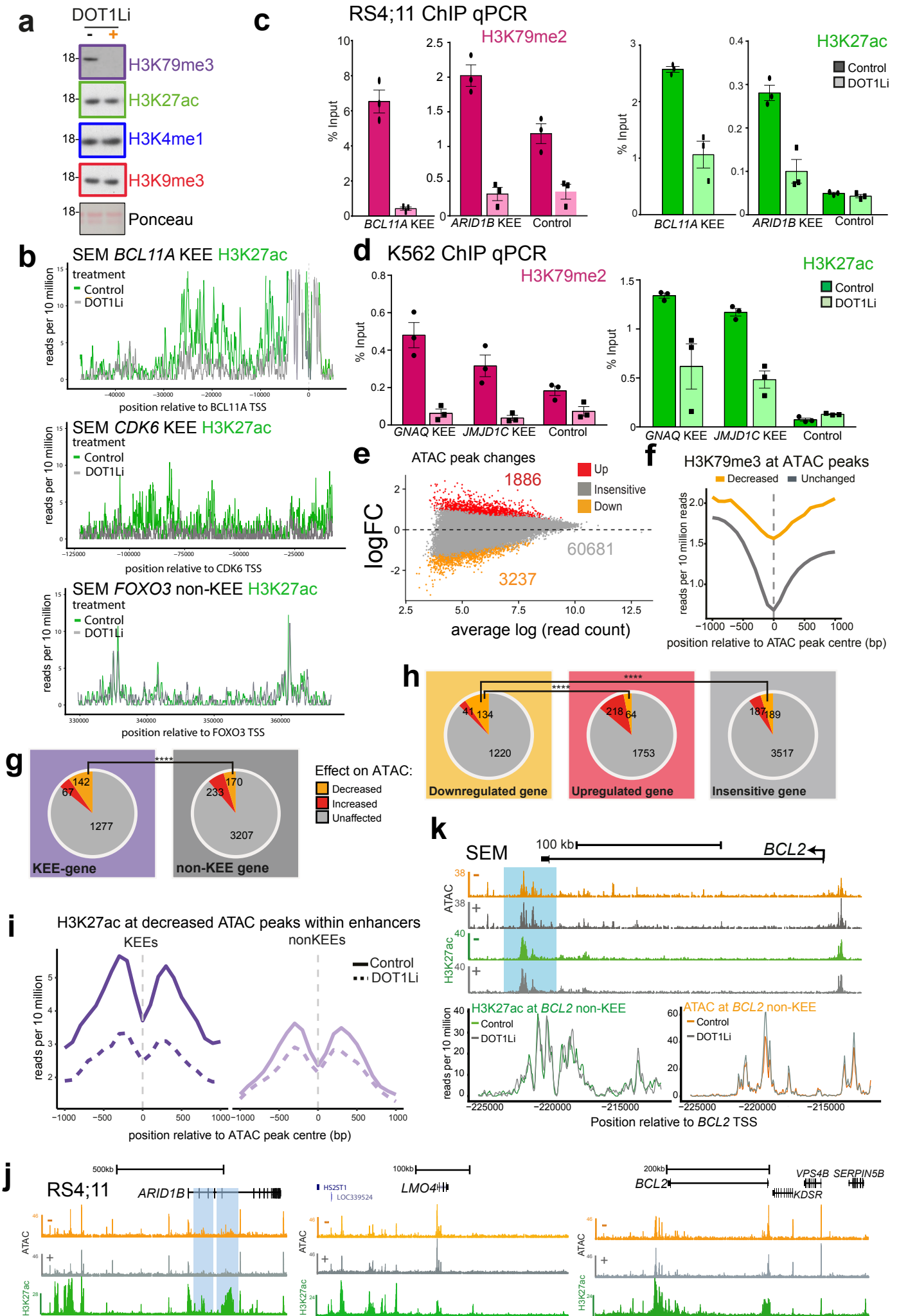
WT ATTTCTATACATCTCCTTTAAGAATGCCACAGGTTTTCCGGATTCTTATCTGGATGGAGCTCTCAATTTCTGAACCTTGACT

mut allele 1 ATTTCTATACATCTCCTTTAAGAATGCCACAGGTTTTCCGGATTCTTATCTGGATGGAGCTCTCAATTTCTGAACCTTGACT

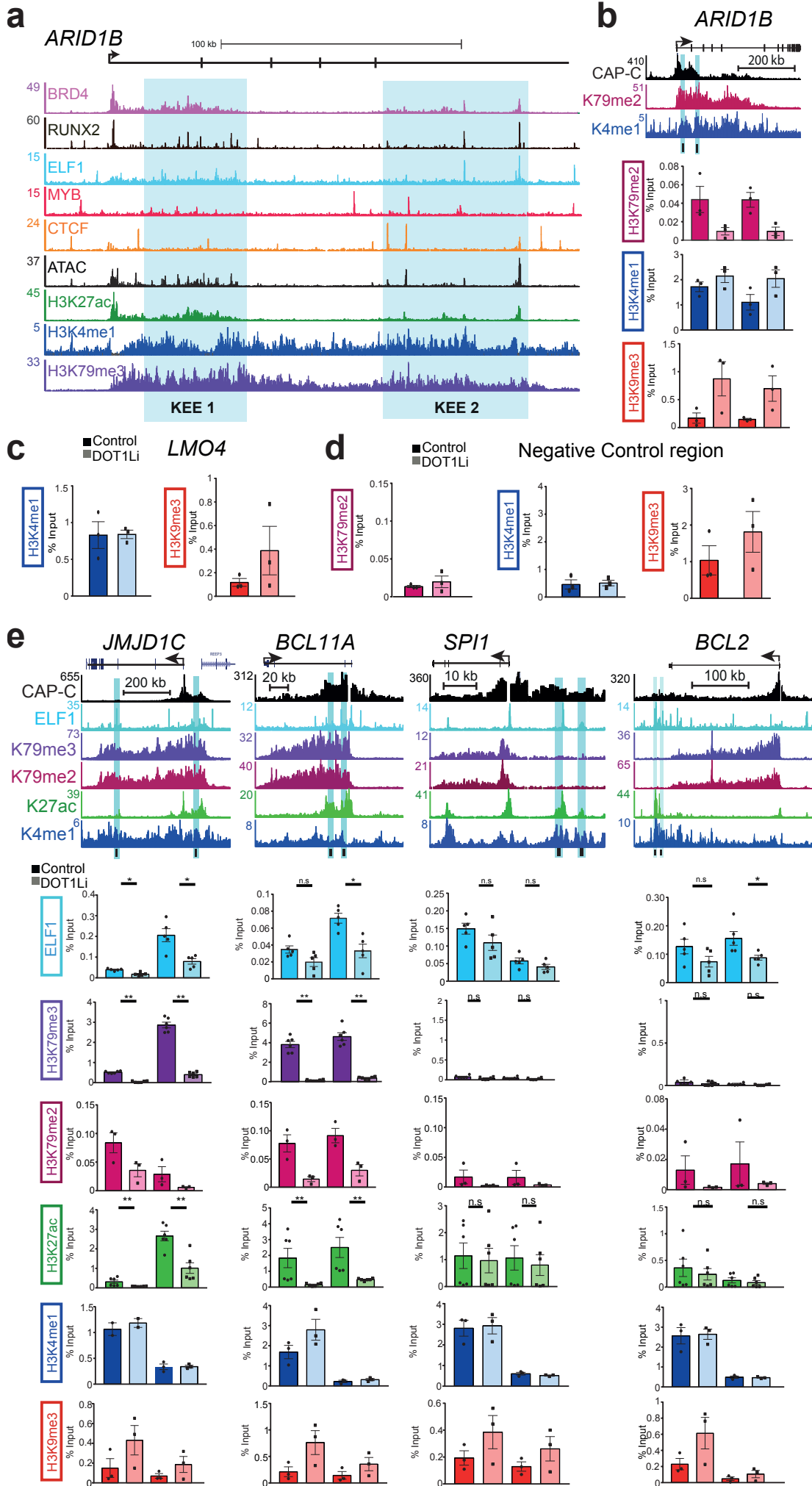
mut allele 2 ATTTCTATACATCTCCTTTAAGAATGCCACAGGTTTTCCGGATTCTTATCTGGATGGAGCTCTCAATTTCTGAACCTTGACT



Supplementary Figure 2. Loss of H3K79me_{2/3} leads to a reduction in transcription at KEE-genes **(a)** Luciferase assay for putative enhancer activity. pGL3 plasmid containing promoter alone, or sections of the *ARID1B* KEE, *JMJD1C* KEE or *BCL2* non-KEE were co-transfected with pRL-TK *Renilla* into 293T cells. Firefly luciferase activity was normalised to *Renilla* luciferase. Error bars represent s.e.m of three technical replicates. **(b)** Upper: Locations of gRNA cut sites at *ARID1B* and *CDK6* KEEs (red lines) and *ARID1B* luciferase enhancer (orange line). Lower: Sequence of an *ARID1B* mutant and *CDK6* mutant clone compared to WT, characterised using PCR and Sanger sequencing. Solid red bars below the ChIP-seq traces show the deleted regions. Left: RT-qPCR analysis of *ARID1B* and *CDK6* expression in the indicated clones relative to WT SEM cells, mean of at least three replicates. Error bars represent s.e.m. **(c)** H3K79me₃ and H3K79me₂ ChIP qPCR in control (dark shade) and DOT1Li (light shade) SEM cells at KEE genes and negative control locus. Error bars represent s.e.m (n=3). **(d)** Western blot analysis of H3K79me₃ and H4 in control (-) and DOT1Li (+) SEM cells. Representative of three biological replicates. **(e)** Proportion of DOT1Li downregulated, upregulated or insensitive genes associated with a KEE, non-KEE or no enhancer. **** = p<0.0001, Fisher's exact test. **(f)** RT-qPCR of total RNA in control (black) and DOT1Li (orange) SEM cells at *ARID1B*, *BCL2*, *CDK6* and *BCL11A*, normalised to the housekeeping gene *YWHAZ*. Error bars represent s.d (n=3). **(g)** Left: Nascent RNA-seq at *BCL2*, *LMO4* and *PBX3* in control (black, -) and DOT1Li (orange, +) conditions performed in SEM cells, H3K79me₃ ChIP-rx in control (purple, -) and DOT1Li (orange, +) conditions, ATAC-seq, H3K27ac and H3K4me₁ ChIP-seq. Right: Bar charts showing nascent RNA-seq RPKM values at *BCL2*, *LMO4* and *PBX3* in Control (black) and DOT1Li (orange) conditions. Error bars represent s.d from 3 biological replicates. For p-values see Supplementary Table 2. **(h)** Overlay of Capture-C with H3K79me₂ and H3K27ac ChIP-seq at *BCL2* in SEM cells. Gray bars represent location of Capture-C probe, +/-1 kb exclusion zone. Source data are provided as a source data file.

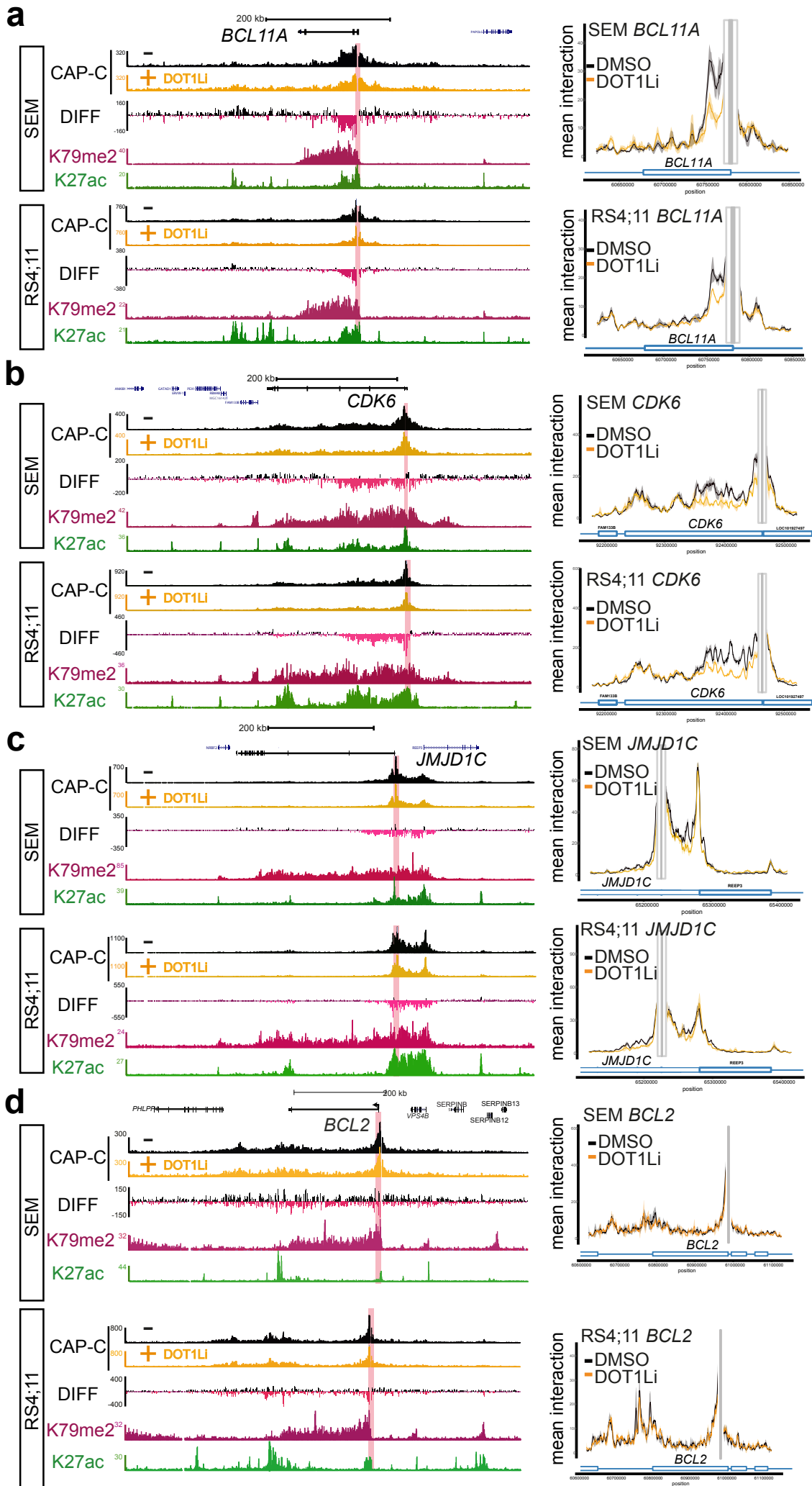


Supplementary Figure 3. Loss of H3K79me2/3 leads to reduction in active histone modifications and chromatin accessibility at KEEs **(a)** Western blot analysis of acid-extracted histones from SEM cells in control (-) and DOT1Li (+) conditions. **(b)** Overlay of H3K27ac ChIP-seq in control (green) and DOT1Li (gray) at *BCL11A*, *CDK6* KEE and *FOXO3* non-KEE region. **(c)** H3K79me2 and H3K27ac ChIP qPCR in RS4;11 cells at KEE-associated genes in control (darker shade) and DOT1Li (lighter shade) conditions. n=3, error bars represent s.e.m. **(d)** H3K79me2 and H3K27ac ChIP qPCR in K562 cells at KEE-associated genes in control (darker shade) and DOT1Li (lighter shade) conditions. n=3, error bars represent s.e.m. **(e)** MA plot representing up (red, 1886), down (orange, 3237) and insensitive (gray, 60681) ATAC peaks following DOT1Li, FDR <0.05, based on five biological replicates. **(f)** Metaplot of H3K79me3 ChIP-seq signal from control SEM cells centered around DOT1Li-decreased (orange) and unchanged (gray) ATAC peaks. **(g)** Proportion of ATAC peaks within KEE genes or non-KEE genes that show increases (red), decreases (orange) or no change (gray) following DOT1Li. **** = p <0.0001, Fisher's exact test. **(h)** Proportion of ATAC peaks within DOT1Li transcriptionally downregulated, upregulated and insensitive genes that show increases (red), decreases (orange) or no change (gray) following DOT1Li. **** = p <0.0001, Fisher's exact test. **(i)** Metaplot of H3K27ac ChIP-seq levels at DOT1Li-decreased ATAC peaks found within KEEs and non-KEEs. H3K27ac levels at these regions were measured from control (solid line) and DOT1Li (dashed line) SEM cells. **(j)** ATAC-seq in control (orange) and DOT1Li (gray) conditions and H3K27ac ChIP-seq (green) in RS4;11 cells at *ARID1B*, *LMO4* and *BCL2*. Blue boxes at *ARID1B* represent regions where reductions in ATAC correspond to those observed in SEM cells. **(k)** Upper: ATAC-seq and H3K27ac ChIP-seq in control (orange/green) and DOT1Li (gray) SEM cells at *BCL2*. Highlighted blue regions represent region represented in H3K27ac and ATAC-seq overlays below. Lower: Overlay of H3K27ac (left) and ATAC (right) signal at *BCL2* non-KEE in control (green/orange) and DOT1Li (gray) SEM cells. Source data are provided as a source data file.

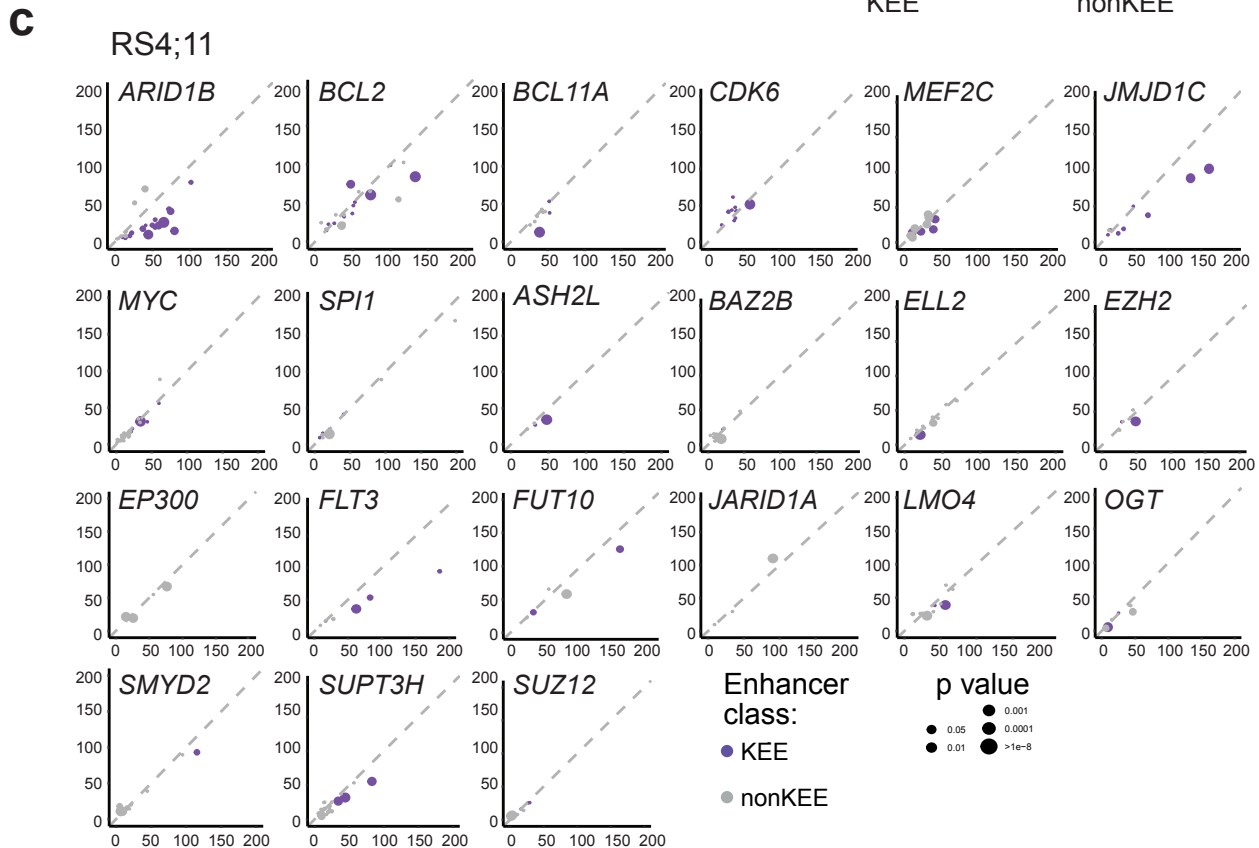
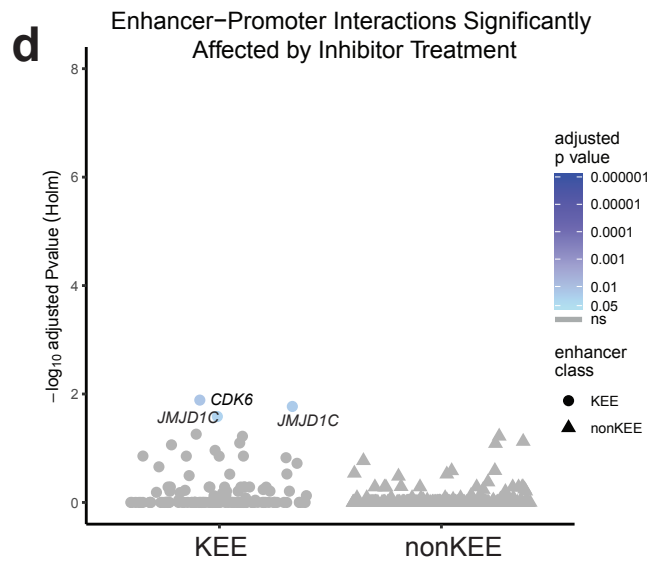
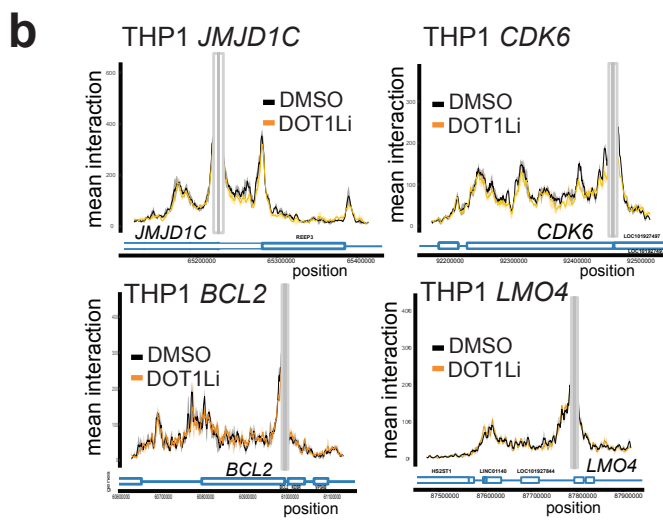
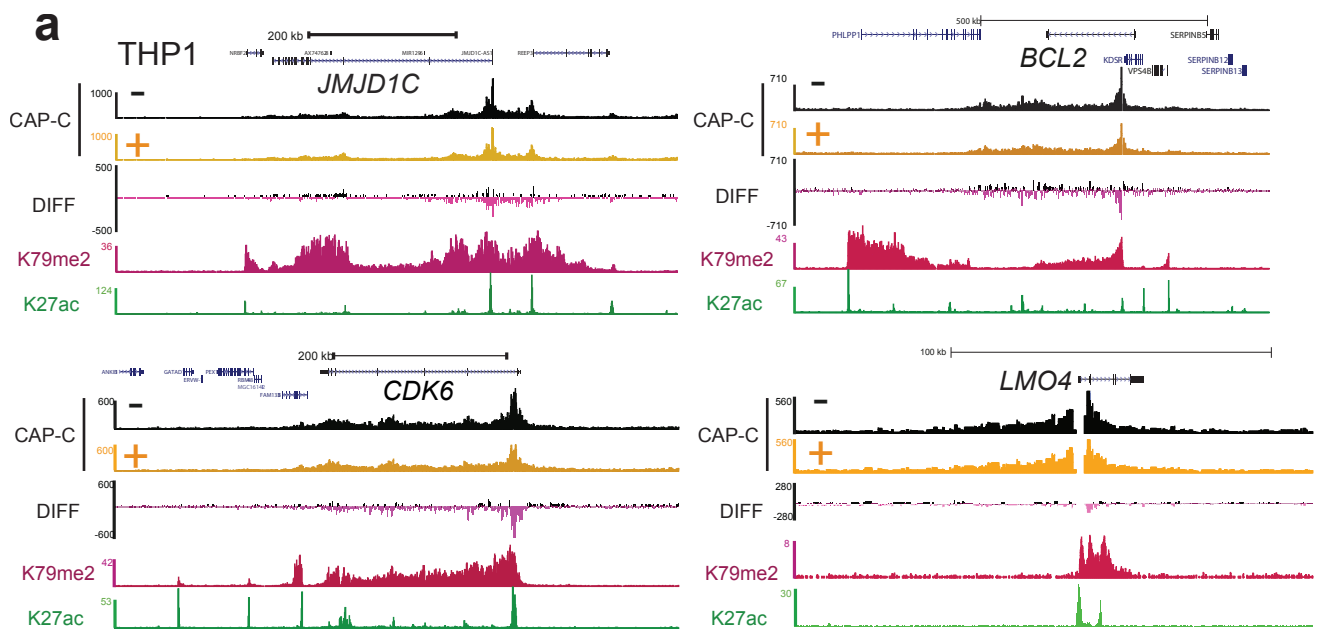


Supplementary Figure 4. Loss of H3K79me_{2/3} leads to a reduction in ELF1 binding at KEEs

(a) ChIP-seq at *ARID1B* demonstrating multiple transcription factor binding events at KEEs in SEM cells. **(b)** H3K79me₂, H3K4me₁ and H3K9me₃ ChIP qPCR at *ARID1B* in SEM cells in control (darker shade) and DOT1Li (lighter shade) conditions. **(c)** H3K4me₁ and H3K9me₃ ChIP qPCR at *LMO4* in control (darker shade) and DOT1Li (lighter shade) conditions. Error bars represent s.e.m **(d)** H3K79me₂, H3K4me₁ and H3K9me₃ ChIP qPCR at negative control region in control (darker shade) and DOT1Li (lighter shade) conditions. Error bars represent s.e.m **(e)** ChIP qPCR at KEEs and non-KEEs for ELF1, H3K79me₃, H3K79me₂, H3K27ac, H3K4me₁ and H3K9me₃ in control (darker shade) and DOT1Li (lighter shade) SEM cells. n=6, error bars represent s.e.m. Statistical test used Mann Whitney U test, * = p<0.05, ** = p<0.01, n.s. = non-significant. Source data are provided as a source data file.



Supplementary Figure 5. Loss of H3K79me_{2/3} leads to a reduction in KEE-promoter interactions at *ARID1B* but not non-KEE-promoter interactions at *LMO4* (**a-d**) Capture-C from the promoter of *BCL11A*, *CDK6*, *JMJD1C* and *BCL2* in control (black) and DOT1Li (orange) treatment, in SEM (upper) and RS4;11 (lower) cells, n=3. Left: Differential track demonstrates difference in signal between control and DOT1Li: increase (black), decrease (pink). ChIP-seq tracks for H3K79me₂ and H3K27ac. Right: overlay of control (black) and DOT1Li (orange) Capture-C signal across each gene. Gray bars represent location of Capture-C probe, +/-1 kb exclusion zone. Shaded area around Capture-C signal represents 1 s.d.



Supplementary Figure 6. Loss of H3K79me2/3 leads to specific reductions in KEE-promoter interactions at a large number of genes **(a)** Capture-C from the promoter of *JMJD1C*, *CDK6*, *BCL2* and *LMO4* in control (black) and DOT1Li (orange) treatment performed in triplicate in THP1 cells. Differential track (increases (black), decreases (pink)) demonstrates difference in signal between control and DOT1Li. ChIP-seq tracks for H3K79me2 and H3K27ac. **(b)** Overlay of control (black) and DOT1Li (orange) Capture-C signal (average of three biological replicates) across *JMJD1C*, *CDK6*, *BCL2* and *LMO4* in THP1 cells. Gray bars represent location of Capture-C probe, +/-1 kb exclusion zone. **(c)** Enhancer-promoter Capture-C interaction frequencies in Control (x-axis) and DOT1Li (y-axis) RS4;11 cells. Each point represents the interaction of a KEE (purple) or non-KEE (gray) with the indicated gene promoter. Size of dot corresponds to significance of change in interaction, using a Wilcoxon rank test. See Supplementary Table 6 for list of p values. **(d)** Statistical analysis of the significance of the change in enhancer-promoter Capture-C interactions following DOT1Li in THP1 cells. Each point represents the interaction of a KEE (circle) or non-KEE (triangle) with a gene promoter. Significantly affected interactions are labelled with the relevant gene promoter. Holm-Bonferroni adjusted p-values were calculated following a Wilcoxon rank test (n=3).

Primer name	Sequence
ARID1B KEE1 F primer	TTTCCAGTTCCGTCCAAGTATT
ARID1B KEE1 R primer	CTTTCTCCCAGCTTCCTGTTAG
ARID1B KEE2 F primer	CTAACAGGAAGCTGGGAGAAAAG
ARID1B KEE2 R primer	GCTTCTTGCTTAAGGCGTTTC
BCL11A KEE1 F primer	GCAAGCCAGGAAAGATGATAGA
BCL11A KEE1 R primer	AGAGACGCACAGCTCAATTT
BCL11A KEE2 F primer	ACACCCAGTGCCCAGAATTG
BCL11A KEE2 R primer	CGCGGGTCCTGAGATTCATT
JMJD1C KEE1 F primer	CTTCCAGCACGAGTCACATTA
JMJD1C KEE1 R primer	CTCGCCACTAGTCTGTCTTTG
JMJD1C KEE2 F primer	CAAGGTACGTCTGCGAGAG
JMJD1C KEE2 R primer	CGTGTCACACAGGGACAG
SPI-1 non-KEE1 F primer	TAACAGCTTCCGCCAAGAG
SPI1 non-KEE1 R primer	CGCAGAGTTGAGACCAGAG
SPI-1 non-KEE2 F primer	GCCAAACACTAGGTCAGAGAAA
SPI1 non-KEE2 R primer	CCTACTTTGGCCACCCTTAAA
BCL2 non-KEE1 F primer	CTTCACGCAGCATCCATTTG
BCL2 non-KEE1 R primer	CCAATTCAGCATCTAGTTCTCCT
BCL2 non-KEE2 F primer	CCGGCATCTGAGCTAATTACA
BCL2 non-KEE2 R primer	TCTGCCCAGTACCTCTTATCT
PROM1 intron 1 F primer	GGTGACACAATTCAACCCATAAC
PROM1 intron 1 R primer	CATGGGTCTTTAGTTGGTGTAGTA
GNAQ F primer	GTCCATCATGGCGTGCT
GNAQ R primer	CGGACGGTACTCACCGA
Negative control F primer	GGCTCCTGTAACCAACCACTACC
Negative control R primer	CCTCTGGGCTGGCTTCATTC
BCL2en luciferase assay F primer	ATGCATCGATGTCTTCACAGCCGTCATCCT
BCL2en luciferase assay R primer	ATGCGCGGCCGCGCCAATGACATACGACAGC
ARID1B KEE luciferase assay F	GGTGTGAGAAGTTGGGGCTA
ARID1B KEE luciferase assay R	GCTTGCTCCTAGGAAATACACC
JMJD1C KEE luciferase assay	AAATCGATAAGGATCCAGCTTGAAAATATCTTTCCATAATTTCTACTCTATTA CAC
JMJD1C KEE luciferase assay	AAGGGCATCGGTGATTGCTATAGGGATTTGGGTACTGATGC
YWHAZ Taqman probe mix	Hs03044281_g1
ARID1B Taqman probe mix	Hs00368175_m1
CDK6 Taqman probe mix	Hs01026371_m1
BCL2 Taqman probe mix	Hs00608023_m1
BCL11A Taqman probe mix	Hs01093197_m1

Supplementary Table 1. List of primers used.

Experiment	Cell line	Antibody	Accession Number
ChIP seq	SEM	H3K27ac	GSE74812
ChIP seq	SEM	H3K79me2	GSE74812
ChIP seq	SEM	H3K79me3	GSE74812
ChIP seq	SEM	H3K4me1	GSE74812
ChIP seq	SEM	H3K4me3	GSE74812
ChIP seq	SEM	BRD4	GSE83671
ChIP seq	RS4;11	H3K79me2	GSE38403
ChIP seq	RS4;11	H3K27ac	GSE71616
ChIP seq	RS4;11	H3K4me1	GSE71616
ChIP seq	RS4;11	H3K4me3	GSE71616
ChIP seq	ML-2	H3K79me2	GSE83671
ChIP seq	K562	H3K27ac	GSE29611
ChIP seq	K562	H3K79me2	GSE29611
ChIP seq	K562	H3K4me1	GSE31755
ChIP seq	K562	H3K4me3	GSE96303
ChIP seq	GM12878	H3K27ac	GSE29611
ChIP seq	GM12878	H3K79me2	GSE29611
ChIP seq	GM12878	H3K4me1	GSE29611
ChIP seq	GM12878	H3K4me3	GSE29611
ChIP seq	HeLa-S3	H3K27ac	GSE29611
ChIP seq	HeLa-S3	H3K79me2	GSE29611
ChIP seq	HeLa-S3	H3K4me1	GSE29611
ChIP seq	HeLa-S3	H3K4me3	GSE29611
ChIP seq	H1-hESC	H3K27ac	GSE16256
ChIP seq	H1-hESC	H3K79me2	GSE16256
ChIP seq	H1-hESC	H3K4me1	GSE16256
ChIP seq	H1-hESC	H3K4me3	GSE16256
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PolyA RNA seq	K562		GSE88351
PolyA RNA seq	GM12878		GSE88583
PolyA RNA seq	HeLa-S3		GSE33480
PolyA RNA seq	H1-hESC		GSE16256

Supplementary Table 2. GEO accession numbers: datasets from previous publications.