

# SUPPLEMENTAL INFORMATION

## Redefining the sRNA Transcriptome in *Streptococcus pneumoniae*

### Serotype 2 Strain D39

Dhriti Sinha<sup>1,2</sup>, Kurt Zimmer<sup>3</sup>, Todd A. Cameron<sup>2</sup>, Douglas B. Rusch<sup>4</sup>, Malcolm E. Winkler<sup>1#</sup>, and Nicholas R. De Lay<sup>2,5#</sup>

<sup>1</sup>Department of Biology, Indiana University Bloomington, Bloomington, IN 47405, USA

<sup>2</sup>Department of Microbiology and Molecular Genetics, McGovern Medical School, University of Texas Health Science Center, Houston, TX 77030, USA

<sup>3</sup>Indiana University School of Informatics, Computing and Engineering, Indiana University Bloomington, Bloomington, IN 47408, USA

<sup>4</sup>Center for Genomics and Bioinformatics, Indiana University Bloomington, Bloomington, IN 47405, USA

<sup>5</sup>MD Anderson Cancer Center UTHealth Graduate School of Biomedical Sciences, University of Texas Health Science Center, Houston, TX 77030, USA

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#Correspondence should be sent to Nicholas R. De Lay (Tel: +1 (713) 500-6293; Email: [nicholas.r.delay@uth.tmc.edu](mailto:nicholas.r.delay@uth.tmc.edu)) and Malcolm E. Winkler (Tel: +1 (812) 856-1318; E-mail: [winklerm@indiana.edu](mailto:winklerm@indiana.edu))

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TABLE S1: List showing the overlap between the sRNAs and riboswitches that were

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identified in *S. pneumoniae* D39V and D39W

sRNA ID in D39V	Coordinates in D39V	Flanking genes	sRNA ID in D39W	Coordinates in D39W	Identification in D39W
<i>ccnC</i> <sup>a</sup>	23967-24065 (+)	<i>spd_0024</i> , <i>spd_0025</i>	<i>ccnC</i>	23967-24065 (+)	dRNA-seq
<i>scRNA</i>	24632-24707 (+)	<i>spd_0026</i>	<i>scRNA</i>	24632-24707 (+)	
<i>srf-01</i>	29658-29873 (+)	<i>spd_0033</i> , <i>spd_0038</i>	ND	ND	ND
<i>srf-02</i> <sup>a</sup>	39980-40186 (+)	<i>spd_0047</i> , <i>spd_0048</i>	<i>spd_sr5</i>	39980-40082 (+)	sRNA-seq & dRNA-seq
<i>srf-03</i> <sup>a</sup>	41719-41886 (+)	<i>spd_0048</i> , <i>comA</i>	<i>spd_sr6</i>	41494-41559 (+)	dRNA-seq
<i>srf-04</i> <sup>b</sup>	13229-132308 (+)	<i>tmrU</i> , <i>spd_0128</i>	<i>spd_sr37</i>	131773-131842 (+)	dRNA-seq
<i>srf-05</i> <sup>b</sup>	149645 – 149877 (+)	<i>spd_0143</i> , <i>spd_0144</i>	<i>spd_sr14</i>	149223-149341 (+)	sRNA-seq
<i>srf-06</i>	150711-150914 (-)	<i>spd_0145</i> , <i>spd_0144</i>	ND	ND	ND
<i>ccnE</i> <sup>a</sup>	212734-212881 (+)	<i>spd_0221</i> , <i>spd_0222</i>	<i>ccnE</i>	212278 – 212425 (+)	sRNA-seq & dRNA-seq
<i>ccnA</i> <sup>b</sup>	231599 – 231691 (+)	<i>spd_0240</i> , <i>ruvB</i>	<i>ccnA</i>	231143 – 231235 (+)	dRNA-seq
<i>ccnB</i> <sup>a</sup>	231786-231883 (+)	<i>spd_0240</i> , <i>ruvB</i>	<i>ccnB</i>	231331 – 231427 (+)	dRNA-seq
<i>srf-07</i> <sup>a</sup>	232279-232354 (+)	<i>spd_0240</i> , <i>ruvB</i>	<i>spd_sr23</i>	231823 – 232091 (+)	sRNA-seq & dRNA-seq
<i>ccnD</i> <sup>a</sup>	234171-234264 (+)	<i>spd_0242</i> , <i>uppS</i>	<i>ccnD</i>	233715 – 233809 (+)	dRNA-seq
<i>srf-08</i>	282194-282479 (+)	<i>spd_0284</i> , <i>spd_0285</i>	ND	ND	ND
<i>rnpB</i>	344607-345007 (+)	<i>spd_0339</i> , <i>spd_0341</i>	<i>rnpB</i>	344150 – 344553 (+)	
<i>srf-10</i>	508697-508842 (+)	<i>spd_0500</i> , <i>bglG</i>	<i>spd_sr33</i>	508237 – 508335 (+)	sRNA-seq
<i>srf-11</i> <sup>a</sup>	587896-587989 (+)	<i>spd_0563</i> , <i>spd_0564</i>	<i>spd_sr42</i>	587439 – 587538 (+)	dRNA-seq
<i>srf-12</i> <sup>d</sup>	742022-742141	<i>spd_0726</i> , <i>spd_0729</i>	ND	ND	ND
<i>ssrA</i> <sup>a</sup>	781595-781939 (+)	<i>spd_0768</i> , <i>spd_0770</i>	<i>spd_sr46</i>	781128 – 781485 (+)	sRNA-seq & dRNA-seq
<i>srf-13</i> <sup>a,c</sup>	826260 – 826587 (+)	<i>spd_0808</i> , <i>cad</i>	<i>spd_sr49</i>	825802 – 826129 (+)	sRNA-seq & dRNA-seq
<i>srf-16</i>	1037649- 1037755 (-)	<i>spd_0914</i> , <i>spd_0915</i>	ND	ND	ND
<i>srf-17</i> <sup>b</sup>	1051910- 1052049 (-)	<i>spd_0899</i> , <i>asd</i>	<i>spd_sr17</i>	912572 – 912715 (+)	sRNA-seq & dRNA-seq

<b>srf-18<sup>c</sup></b>	1079561-1079658 (-)	<i>lacD, lacT</i>	<i>spd_sr60</i>	1079135 – 1079199 (-)	dRNA-seq
<i>srf-19</i>	1170746 – 1170923 (+)	<i>spd_1136, spd_1137</i>	<i>spd_sr63</i>	1170288 – 1170385 (+)	sRNA-seq & dRNA-seq
<b>srf-21<sup>a</sup></b>	1528520-1528643 (-)	<i>recG, spd_1506</i>	<i>spd_sr83</i>	1528044 – 1528186 (-)	sRNA-seq & dRNA-seq
<i>srf-22</i>	1548804-1549088 (-)	<i>spd_1527, spd_1526</i>	ND	ND	ND
<b>ssrS<sup>a</sup></b>	1598326 – 1598522 (+)	<i>spd_1580, spd_1581</i>	<i>spd_sr85</i>	1597869 – 1598142 (+)	sRNA-seq & dRNA-seq
<i>srf-24</i>	1873736-1873781 (-)	<i>mutS, spd_1902</i>	<i>spd_sr105</i>	1873276 - 1873324 (-)	dRNA-seq
<b>srf-25<sup>a</sup></b>	1892857-1893007 (-)	<i>spd_1924, spd_1923</i>	<i>spd_sr106</i>	1892400 – 1892550 (-)	sRNA-seq & dRNA-seq
<i>srf-26</i>	1949385-1949547(+)	<i>spd_1974, arcA</i>	ND	ND	ND
<i>srf-27<sup>d</sup></i>	1973172-1973570 (-)	<i>adcA, spd_1996</i>	<i>spd_sr110</i>	1973000 - 1973113 (-)	dRNA-seq
<i>srf-28</i>	1973509-1973913 (-)	<i>adcA, spd_1996</i>	<i>spd_sr112</i>	1973342 - 1973456 (-)	dRNA-seq
<i>srf-29</i>	2008242-2008356 (-)	<i>spd_2029, cbpD</i>	ND	ND	ND
<b>srf-30<sup>a,c</sup></b>	2020587-2020685 (-)	<i>spd_2043, rpsB</i>	<i>spd_sr116</i>	2020112-2020228 (-)	dRNA-seq
<b>riboswitch-1<sup>a</sup></b>	174291-174470 (-)	<i>ruvA, ribD</i>	<i>spd_sr22</i>	173841 – 174014 (-)	sRNA-seq & dRNA-seq
riboswitch-2	376842-377047 (+)	<i>spd_0371, spd_0372</i>	<i>spd_sr28</i>	376371 – 376530 (+)	sRNA-seq & dRNA-seq
riboswitch-3	381487-381662 (-)	<i>spd_0376, serS</i>	<i>spd_sr29</i>	381067 - 381205 (-)	sRNA-seq & dRNA-seq
riboswitch-4	441436-441663 (+)		ND	ND	ND
<b>riboswitch-5<sup>a</sup></b>	497356-497561 (+)	<i>spd_0490, spd_0491</i>	<i>spd_sr32</i>	496899 – 497104 (+)	dRNA-seq
riboswitch-6	513346-513544 (+)	<i>bglA-2, pheS</i>	<i>spd_sr34</i>	512889 – 513087 (+)	sRNA-seq & dRNA-seq
riboswitch-7	632244-632347 (+)	<i>spd_0436, spd_0437</i>	ND	ND	ND
<b>riboswitch-8<sup>a</sup></b>	644329-644497 (+)	<i>lctO, spd_0622</i>	<i>spd_sr43</i>	643872 – 644040 (+)	dRNA-seq
riboswitch-9	647211-647381 (+)	<i>thiE-1, spd_0625</i>	<i>spd_sr44</i>	646754 – 646926 (+)	sRNA-seq & dRNA-seq
riboswitch-10 <sup>d</sup>	653914-654012 (-)	<i>cspR, ribU</i>	ND	ND	ND
riboswitch-11	849418-849636 (+)	<i>spd_0831, spd_0833</i>	ND	ND	ND
riboswitch-12	864986-865160 (+)	<i>spd_0850, pyrK</i>	ND	ND	ND
<b>riboswitch-13<sup>a</sup></b>	996442-996678 (-)	<i>ppC, spd_0954</i>	<i>spd_sr12</i>	967941 – 968177 (+)	dRNA-seq

riboswitch-14 <sup>d</sup>	1052832-1052928 (-)	<i>spd_0897</i> , <i>spd_0898</i>	ND	ND	ND
<b>riboswitch-15<sup>a</sup></b>	1168835-1168970 (-)	<i>nth</i> , <i>pyrR</i>	<i>spd_sr62</i>	1168377-1168512 (-)	dRNA-seq
riboswitch-16	1175106-1175233 (+)	<i>gidB</i> , <i>uraA</i>	<i>spd_sr64</i>	1174647 – 1174790 (+)	sRNA-seq & dRNA-seq
<b>riboswitch-17<sup>a</sup></b>	1250018-1250194 (-)	<i>spd_1216</i> , <i>spd_1217</i>	<i>spd_sr70</i>	1249557 – 1249736 (-)	sRNA-seq & dRNA-seq
<b>riboswitch-18<sup>a</sup></b>	1326523-1326709 (-)	<i>spd_1308</i> , <i>spd_1307</i>	<i>spd_sr74</i>	1326066 – 1326252 (-)	sRNA-seq & dRNA-seq
riboswitch-19	1404449-1404618 (-)	<i>mntE</i> , <i>spd_1383</i>	ND	ND	ND
<b>riboswitch-20<sup>a</sup></b>	1459261-1459468 (+)	<i>spd_1441</i> , <i>spd_1442</i>	<i>spd_sr80</i>	1458804 – 1458975 (+)	sRNA-seq
riboswitch-21	1491275-1491462 (-)	<i>divIVA</i> , <i>ileS</i>	ND	ND	ND
<b>riboswitch-22<sup>a</sup></b>	1619505-1619652 (-)	<i>spd_1605</i> , <i>spd_1604</i>	<i>spd_sr88</i>	1619052 – 1619299 (-)	sRNA-seq & dRNA-seq
riboswitch-23	1642795-1642969 (+)	<i>spd_1626</i> , <i>xpt</i>	ND	ND	ND
<b>riboswitch-24<sup>a</sup></b>	2006750-2006885 (-)	<i>cbpD</i> , <i>spd_2027</i>	<i>spd_sr114</i>	2006263 – 2006428 (-)	sRNA-seq & dRNA-seq

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43     <sup>a</sup>sRNAs validated for expression in this study.

44     <sup>b</sup>sRNAs validated for expression by Tsui et al 2010 (1).

45     <sup>c</sup>sRNAs validated for expression by Slager et al 2018 (2).

46     <sup>d</sup>sRNAs predicted to be present in D39V by Slager et al 2018 (2).

47     ND = not detected.

**Table S2: List of sRNAs predicted in TIGR4, the coordinates of which are absent in D39W**

Mann et al (2012)			Acebo et al (2012)		
sRNA Name	Start	Stop	sRNA Name	Start	Stop
			<b>trn0027</b>	79336	79386
<b>F1<sup>a</sup></b>	91593	91664	<b>trn0036</b>	91593	91651
<b>F56</b>	158993	159090			
			<b>srn081</b>	273681	273799
			<b>trn0256</b>	436064	436160
			<b>srn135</b>		
<b>F12</b>	444624	444663		438151	438276
			<b>trn0290</b>	497393	497444
			<b>srn151</b>	501260	501364
<b>F57</b>	553923	553982			
<b>F24</b>	610528	610660			
<b>R13</b>	623327	623372			
			<b>trn0355</b>	673130	673199
<b>F29</b>	743792	743828			
<b>F62</b>	995726	995787			
			<b>srn266</b>	1035461	1035582
<b>F40</b>	1063101	1063151			
<b>F41<sup>a</sup></b>	1071112	1071214	<b>srn277</b>	1071102	1071212
<b>F46</b>	1454220	1454277			
			<b>trn0757</b>	1515505	1515588
<b>R16</b>	1227703	1227744			
<b>R12<sup>a</sup></b>	1731041	1731440	<b>trn0830</b>	1731046	1731098
			<b>srn448</b>	1901982	1902084
<b>R11</b>	2002164	2002195			

<sup>a</sup> sRNAs identified to be common in TIGR4 by Mann et al., 2012 (3) and Acebo et al., 2012 (4).

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**Table S3: Classification of *S. pneumoniae* D39W sRNAs into different functional categories**

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sRNA name	Start	End	Upstream Gene	Downstream Gene	Predicted/Putative Function
Spd_sr46	781128	781485	<i>spd_0768</i>	<i>spd_0770</i>	tmRNA <sup>a</sup> , defective in lung infection <sup>b</sup> , attenuated in invasive pneumonia model <sup>b</sup>
Spd_sr85	1597869	1598142	<i>spd_1580</i>	<i>spd_1581</i>	6S RNA <sup>a</sup>
Spd_sr48	862699	862827	<i>spd_0846</i>	<i>infC</i>	L20 leader <sup>a</sup>
Spd_sr55	999541	999605	<i>spd_0988</i>	<i>rplU</i>	L21 leader <sup>a</sup>
Spd_sr54	1215844	1215967	<i>spd_1189</i>	<i>rplJ</i>	L10 leader <sup>a</sup>
Spd_sr43	643872	644040	<i>lctO</i>	<i>spd_0622</i>	TPP-riboswitch <sup>a</sup>
Spd_sr44	646754	646926	<i>thiE-1</i>	<i>spd_0625</i>	TPP-riboswitch <sup>a</sup> , defective replication in bloodstream <sup>b</sup>
Spd_sr114	2006263	2006428	<i>cbpD</i>	<i>spd_2027</i>	TPP-riboswitch <sup>a</sup>
Spd_sr22	173827	173979	<i>ruvA</i>	<i>ribE</i>	FMN-riboswitch <sup>a</sup>
Spd_sr28	376371	376530	<i>spd_0371</i>	<i>spd_0372</i>	Glycine-riboswitch <sup>a</sup>
Spd_sr29	381067	381205	<i>spd_0376</i>	<i>serS</i>	T-box element <sup>a</sup>
Spd_sr32	496899	497104	<i>spd_0490</i>	<i>spd_0491</i>	T-box element <sup>a</sup>
Spd_sr34	512889	513087	<i>bglA-2</i>	<i>pheS</i>	T-box element <sup>a</sup> , defective in lung infection and nasopharynx colonization <sup>b</sup> , attenuated in invasive pneumonia model <sup>b</sup>
Spd_sr12	967941	968177	<i>ppC</i>	<i>spd_0954</i>	T-box element <sup>a</sup>
Spd_sr70	1249557	1249736	<i>potD</i>	<i>spd_1217</i>	T-box element <sup>a</sup>
Spd_sr74	1326066	1326252	<i>spd_1308</i>	<i>spd_1307</i>	T-box element <sup>a</sup>
Spd_sr80	1458804	1458975	<i>spd_1441</i>	<i>spd_1442</i>	T-box element <sup>a</sup>
Spd_sr88	1619052	1619299	<i>spd_1605</i>	<i>spd_1604</i>	T-box element <sup>a</sup> , defective in lung infection <sup>b</sup>
Spd_sr62	1168377	1168512	<i>nth</i>	<i>pyrR</i>	PyrR-regulated RNA
Spd_sr64	1174647	1174790	<i>gidB</i>	<i>uraA</i>	PyrR-regulated RNA <sup>c</sup> , defective in lung infection <sup>b</sup> , attenuated in invasive pneumonia model <sup>b</sup>
Spd_sr49	825802	826129	<i>spd_0808</i>	<i>cad</i>	defective in lung infection <sup>b</sup>
Spd_sr17	912572	912715	<i>spd_0899</i>	<i>asd</i>	defective in nasopharynx colonization <sup>b</sup>
Spd_sr111	1973304	1973394	<i>spd_1996</i>	<i>adcA</i>	Toxin module III <sup>c</sup> , defective in nasopharynx colonization <sup>b</sup>
CcnE	212278	212425	<i>spd_0221</i>	<i>spd_0222</i>	Stationary phase autolysis <sup>d</sup> , competence development <sup>e</sup> , attenuated in invasive pneumonia model <sup>b</sup>

<b>Spd_sr23</b>	231823	232091	<i>spd_0240</i>	<i>ruvB</i>	Toxin module I <sup>c</sup>
<b>Spd_sr24</b>	231853	232035	<i>ruvB</i>	<i>spd_0240</i>	Antitoxin sRNA I <sup>c</sup>
<b>Spd_sr109</b>	1972859	1973060	<i>spd_1996</i>	<i>adcA</i>	Toxin module II <sup>c</sup>
<b>Spd_sr110</b>	1973000	1973113	<i>adcA</i>	<i>spd_1996</i>	Antitoxin sRNA II <sup>c</sup>
<b>Spd_sr112</b>	1973342	1973456	<i>adcA</i>	<i>spd_1996</i>	Antitoxin sRNA III <sup>c</sup>

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<sup>a</sup>Function predicted from Rfam database.

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<sup>b</sup> D39 sRNAs correspond to the sRNAs mutants in TIGR4 strain, which exhibited virulence defect in different host niches as determined by (3).

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<sup>c</sup> D39 sRNAs in this functional category identified to be common between *S. pneumoniae* D39W (this study) and D39V (2). These specific functional categories are assigned to respective sRNAs by Slager et al., 2018 (2) in D39V.

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<sup>d</sup> sRNA function first determined in *S. pneumoniae* R6 (5).

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<sup>e</sup> sRNA function elucidated in D39 by Schnorpfeil et al 2013 (6) and Tsui et al 2010 (1).



TABLE S4: List of *S. pneumoniae* D39 genes with antisense expression<sup>a</sup>

SPD #	Gene	Sense	Antisense	Antisense/ Sense
<i>spd_0022</i>	pseudo gene	202	26121	129
<i>spd_0034</i>	IS1167, transposase, truncation	648	54856	85
<i>spd_0048</i>	IS1167, transposase, truncated	5233	56419	11
<i>spd_0085</i>	hypothetical protein	52	68430	1316
<i>spd_0100</i>	hypothetical protein	578	103989	180
<i>spd_0137</i>	IS1167, transposase, truncated	4512	36854	8
<i>spd_0142</i>	conserved hypothetical protein;	836	4311	5
<b><i>spd_0223</i></b>	iron(III) ABC transporter, permease protein	<b>1696</b>	<b>8697</b>	<b>5</b>
<b><i>spd_0224</i></b>	iron(III) ABC transporter, permease protein	<b>1009</b>	<b>5124</b>	<b>5</b>
<b><i>spd_0225</i></b>	ABC transporter, ATP-binding protein	<b>1161</b>	<b>7847</b>	<b>7</b>
<b><i>spd_0413</i></b>	hypothetical protein	<b>67</b>	<b>34000</b>	<b>507</b>
<b><i>spd_0415</i></b>	transposase, IS116/IS110/IS902 family, degenerate	<b>1313</b>	<b>315472</b>	<b>240</b>
<b><i>spd_0416</i></b>	transposase, IS116-IS110-IS902 family, degenerate	<b>542</b>	<b>106534</b>	<b>197</b>
<i>spd_0450</i>	type I restriction-modification system, S subunit, putative	20155	160108	8
<i>spd_0466</i>	conserved hypothetical protein	1262	16139	13
<b><i>spd_0470</i></b>	<b><i>blpC</i></b>	<b>278</b>	<b>19445</b>	<b>70</b>
<b><i>spd_0471</i></b>	<b><i>blpB</i></b>	<b>2082</b>	<b>361566</b>	<b>174</b>
<b><i>spd_0472</i></b>	<b><i>blpA</i></b>	<b>10387</b>	<b>288906</b>	<b>28</b>
<i>spd_0540</i>	amino acid ABC transporter, amino acid-binding protein, putative	10502	138547	13
<i>spd_0552</i>	hypothetical protein	673	10445	16
<i>spd_0607</i>	transposase, IS30 family, degenerate	186	30722	165
<b><i>spd_0615</i></b>	ABC transporter, substrate-binding protein, truncated	<b>3365</b>	<b>26180</b>	<b>8</b>
<b><i>spd_0616</i></b>	amino acid ABC transporter, ATP-binding protein	<b>3051</b>	<b>22951</b>	<b>8</b>
<b><i>spd_0618</i></b>	amino acid ABC transporter, permease protein	<b>991</b>	<b>8219</b>	<b>8</b>
<b><i>spd_0638</i></b>	transposase family protein	<b>650</b>	<b>10972</b>	<b>17</b>
<b><i>spd_0639</i></b>	conserved hypothetical protein	<b>285</b>	<b>2399</b>	<b>8</b>
<i>spd_0682</i>	hypothetical protein	1928	15842	8
<i>spd_0715</i>	MutT/nudix family protein	613	4792	8
<i>spd_0758</i>	IS1167 transposase, truncation	1960	801301	409
<i>spd_0770</i>	tRNA-Ser	168	845	5
<i>spd_0791</i>	conserved hypothetical protein	1064	27564	26

<i>spd_0794</i>	IS1381 transposase OrfA, truncated	819	19512	24
<i>spd_0810</i>	pilin gene inverting-related protein, authentic frameshift	1067	244038	229
<i>spd_0831</i>	conserved domain protein	1021	7845	8
<i>spd_0841</i>	<i>ald</i> (pseudo gene)	12126	124948	10
<i>spd_0895</i>	<i>hemH</i>	4218	66266	16
<i>spd_0938</i>	Tn5252, relaxase; truncated relative to TIGR4	1647	8488	5
<i>spd_0966</i>	IS1167, transposase, truncated	6053	108659	18
<i>spd_0975</i>	DNA repair protein <i>radC</i> homolog	6602	77356	12
<i>spd_0992</i>	conserved hypothetical protein	101	1968	19
<i>spd_0993</i>	conserved hypothetical protein	54	1013	19
<i>spd_1059</i>	hypothetical protein	44	784	18
<i>spd_1189</i>	conserved hypothetical protein	256	1472836	5753
<b><i>spd_1213</i></b>	membrane protein, putative	<b>9134</b>	<b>127449</b>	<b>14</b>
<b><i>spd_1214</i></b>	ABC transporter, ATP-binding protein	<b>4191</b>	<b>40066</b>	<b>10</b>
<i>spd_1261</i>	conserved hypothetical protein	1781	9001	5
<i>spd_1266</i>	membrane protein, putative	4490	22971	5
<i>spd_1275</i>	transcriptional regulator, GntR family	96	11619	121
<i>spd_1430</i>	<i>fer</i>	16179	79670	5
<b><i>spd_1452</i></b>	membrane protein, putative; truncated relative to TIGR4	<b>5519</b>	<b>61420</b>	<b>11</b>
<b><i>spd_1453</i></b>	pseudo gene	<b>2444</b>	<b>38771</b>	<b>16</b>
<b><i>spd_1454</i></b>	conserved hypothetical protein	<b>12049</b>	<b>118485</b>	<b>10</b>
<b><i>spd_1455</i></b>	conserved hypothetical protein	<b>155</b>	<b>2199</b>	<b>14</b>
<b><i>spd_1456</i></b>	conserved hypothetical protein	<b>204</b>	<b>3306</b>	<b>16</b>
<i>spd_1500</i>	ABC transporter, permease protein	44	754	17
<i>spd_1502</i>	ABC transporter, substrate-binding protein	280	1541	6
<i>spd_1578</i>	conserved hypothetical protein	2047	16927	8
<i>spd_1579</i>	conserved hypothetical protein	1676	28532	17
<i>spd_1593</i>	type IV prepilin peptidase, putative	1547	12407	8
<i>spd_1618</i>	conserved hypothetical protein	220	453455	2061
<b><i>spd_1628</i></b>	<b><i>xpt</i></b>	<b>2175</b>	<b>18732</b>	<b>9</b>
<b><i>spd_1629</i></b>	<b><i>pbuX</i></b>	<b>5962</b>	<b>61196</b>	<b>10</b>
<i>spd_1666</i>	IS1167 transposase, truncation	10284	124603	12
<i>spd_1681</i>	IS1167, transposase	606	35981	59
<i>spd_1770</i>	conserved hypothetical protein	156	33068	212
<i>spd_1795</i>	hypothetical protein	1029	7597	7
<i>spd_1901</i>	transposase, putative	323	59946	186
<i>spd_1906</i>	IS1381, transposase OrfB	4511	185846	41
<i>spd_1939</i>	hypothetical protein	539	91168	169

<i>spd_1955</i>	hypothetical protein	409	52774	129
<i>spd_1967</i>	IS1381, transposase OrfB	473	124528	263
<i>spd_1980</i>	IS3-Spn1, hypothetical protein, truncated	419	2448	6
<i>spd_1983</i>	hypothetical protein	122	36220	297
<i>spd_2001</i>	hypothetical protein	87	3345	38
<i>spd_2008</i>	group II intron, maturase, degenerate	346	6284	18
<i>spd_2038</i>	conserved hypothetical protein	293	6298	21

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<sup>a</sup>Bolded rows correspond to genes in an operon. The numbers in the sense and antisense

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column represent the RPKM (Reads Per Kilobase of transcript per Million mapped reads) values

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for the (+) and the (-) strand and the 'antisense/ sense' column represents the ratio between the

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respective RPKM values of two strands for each gene.

**Table S5: List of oligonucleotide probes used for Northern blot analysis**

<b>sRNA name</b>	<b>Oligonucleotide probe sequence in 5'-3' direction</b>
Spd_sr5 <sup>a</sup>	CAAATAAATCATTGATAGTAACACTTACGACATCC
Spd_sr6 <sup>a</sup>	GGAAATCATCGCCAAACTGGACTACGACGCCCCATC
Spd_sr8	CAAACCGCGTCAACGTCGCCTTGCCGTATAGATGTTACTG
Spd_sr13	CAGTGTTTTTGAGCAGCCCGCAGCTAGTTTCCTAGTTTG
Spd_sr16 <sup>a</sup>	GAAATATTTTAACGGTCATAACGGACTATTCCAGA
Spd_sr19	CATTCCATTTTGTGTCTAGATTGACCTTGATTGCGTTAC
Spd_sr20	GCCCCATATGACCTATAATGAAAAGCGTCTAACCAACTC
Spd_sr22	CATCCAGACTATACTGTCGGCTTTGGAATTTACC
Spd_sr23	CTTCGTATAGTCGATAAATGGCTAAACAAGGACAA
Spd_sr24	CCATCATGGGCTATCTCACCTCCTTTCGTAGGCGG
Spd_sr31	TAAAACCAAACCAGCTCCTAGTAAGGCTAGAAGGG
Spd_sr32	AACCAGTTTTGACACATTTGTGGACTTCTCAGCG
Spd_sr35	GGGATAACAAAGAGAATGACCGAAAGGGCAGAGAG
Spd_sr36-1 <sup>a,b</sup>	GCCGTCAAGCAACTCAAAGGAAAATAGGAAATCGA
Spd_sr36-2 <sup>b</sup>	CGGGCGATTAGCTAAATGCTTTACTAACTCTCTCG
Spd_sr42	GTTACGATAGTCTTGGTAAAATAGAATTGCCAATAAACC
Spd_sr43	CCCTTCGCCAGTCTTAACTGTATCAGTTCAATGGG
Spd_sr45	CTTCCATCAGCTAGCATTACCTAGATTGAGTCAGAGGG
Spd_sr47	GAAAACACCTCTGTGTTATACTTGTGTTCAACCACAAAC
Spd_sr49	ATCGTTTCACAAGATGACGTGTGCTTTCACCCACAC
Spd_sr54	GATACGTCTTGTCTCGGCAGGATATTTATGAGTCG
Spd_sr55	CCCATAAGATCTATAATGAAAAGCGACCAAACAACCTC
Spd_sr57	GTGCCTGATTTTCTCTCCTAATTGTTATTAGTAGC
Spd_sr58 <sup>a</sup>	GGATTTCCCCGATTTTTTGTGAAGACTCGCCTAGC
Spd_sr62	GACTTCACCCTTCTACTTTATCGCGCTCCTTGCCCTGCCTC
Spd_sr66	TTTCCCAACTAAAAGCACTCCAGTTACCGCAACGG
Spd_sr67	CTAAATCGCATTCCATCAACTTCATCTGTTTCGTC
Spd_sr70	CTCTGGACTAAGACAAGTGAAAATCAATTCTCAAC
Spd_sr71	CTACATGTGATGGCAATCAAGATATCAAGAATCATCCTAC
Spd_sr73	AAGAGTAAACTCAGCTAGTCCAACTAACTGAGTT
Spd_sr74	CTACAGAAAGCGCCAGCCCTTTATTTTGCCCTACT
Spd_sr78	CACTATCACAGGTTTCTGCCAGCTACTCCCTTGAG
Spd_sr80	GAGGGACACCTCTGATCGGCTCTAACGTGGCCACC
Spd_sr81	GGCAAATACATAAAAAGCTAACTGAACATTCTCGTATCC
Spd_sr82	CATTATCACACCTTTCTAAGGTGGTTTTTTTATCCCGT
Spd_sr83	TTACTTTAATCGTTACTGTCATATGAGAGTCCTCG
Spd_sr84	AATATGGGAGCCTTTACCGCCTCTAGTCTTTATCC
Spd_sr85	AGACGGCCTACATGCTGTTAGAAGACTTTTTGTTTC
Spd_sr88	CCAACCACCTATTCACAATCACCACAGGCTCCC
Spd_sr89	CTGATTGATCGTTCATTATGATTCCTTGACTGAACGAAC
Spd_sr96	GATTGCTCCAGTAATAAAACCATTGGGAATGAAGG
Spd_sr99	CATCAAGTGCTCCTGACGCAACATACCATAGTCCG
Spd_sr100 <sup>a</sup>	TTCTCTGTGTGTAGTGTACTTGCCACAATGCTTAC
Spd_sr106	ATGTGAGATAAAACGTGCTGTCGGTAAAAGCAAGC
Spd_sr108	GAAAATCAAAGTGCAAACCTAGGAAGCTAGCCGCAG
Spd_sr109	CTATGATAAGGCTCATTAGTTTCACCTCCTCTCAC

Spd_sr111	CACCTCCTCTCACGAACCCATAGGAACGTAATCGG
Spd_sr114	GTGCGGTCGAAGTTTAACTTCCTCTCAGACTGT
Spd_sr116	ACTTCAACTCGCAAACGACCCAAGGGCAACTGCTT
CcnC	TTTGTTACAACAAGTTAGGAGGTCTTCTTGTAACC
CcnB	CTCCACCTGATTGGGTGGAGTTAAGGGAGATTATTATG
CcnE	TAAAAGCCACCCATACAGGCGACTTTTGAAGGAG
CcnD	GGGAGATTATTATGAAAAAGTTTTAGGAGTTTAAGTTAAGG
F14	CAACTACTGTCCAAATCATACCAATTATGAACGG
F23	CGGTTATCTTCTACCTATCTATTTTTTCAACTCCT

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<sup>a</sup>Probes were designed within regions that were slightly upstream or downstream of the

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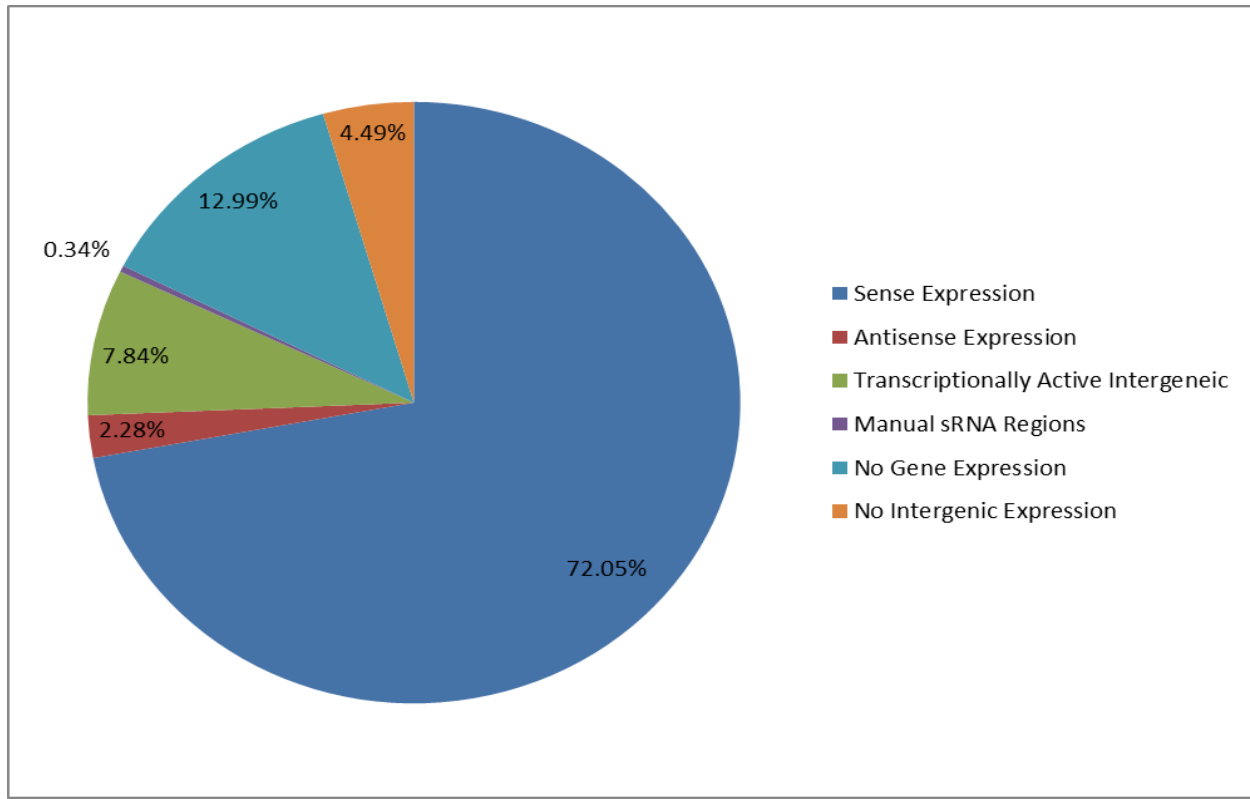
coordinates predicted for these sRNAs.

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<sup>b</sup>The presence of Spd\_sr36 can be validated with both the probes -Spd\_sr36-1 and Spd\_sr36-2.

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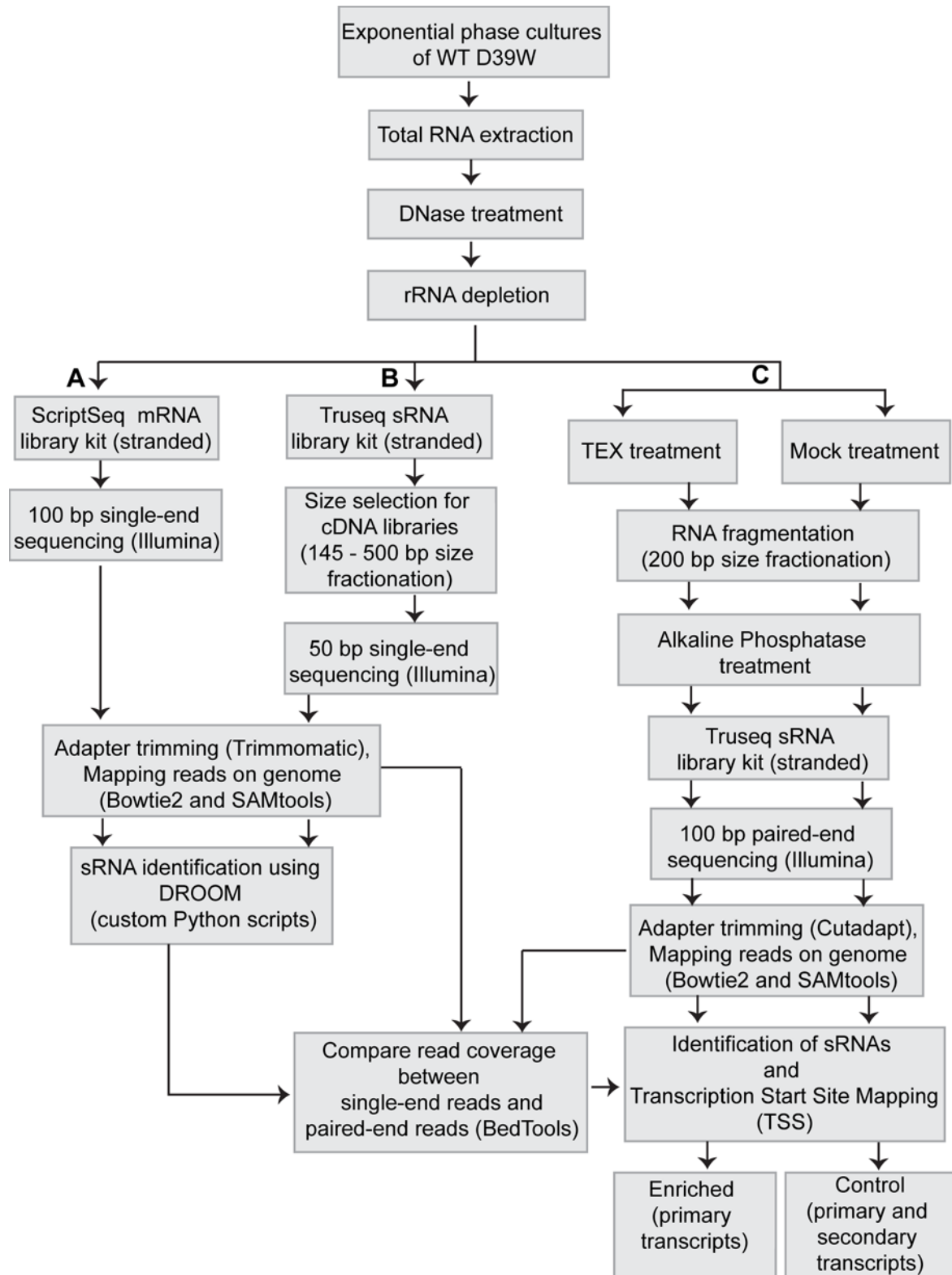
## SUPPLEMENTAL FIGURES



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**Fig. S1. Transcriptome map of *S. pneumoniae* D39.** Comprehensive map showing the distribution of expressed and unexpressed genes and intergenic regions, expressed sRNAs, and antisense transcripts of the *S. pneumoniae* D39W genome based on the minimum coverage threshold of eight read per base as determined for the mRNA-seq analysis.

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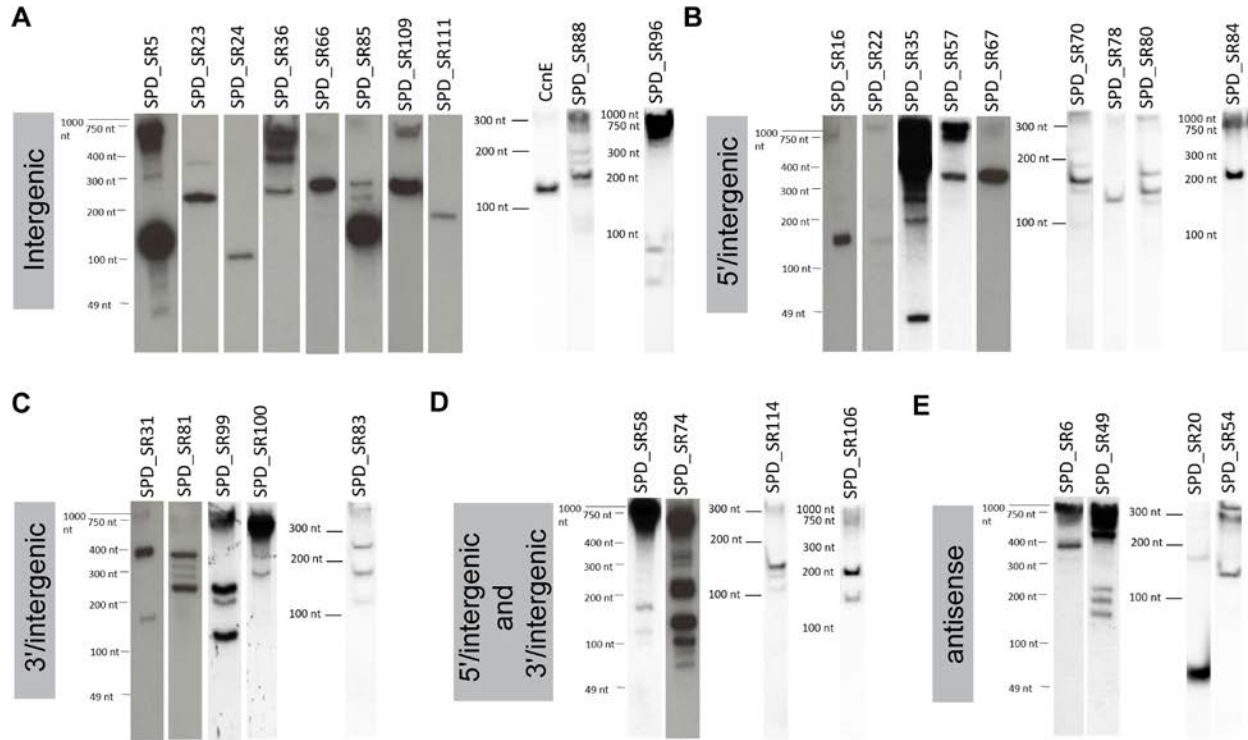
**Fig. S2. Experimental design and data analysis pipeline used to identify sRNAs in D39W.**

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Combined data from mRNA-seq (A) and sRNA-seq (B) analysis were used to identify sRNAs in

84 D39W. Transcription start sites of the identified sRNAs were mapped using differential RNA-seq  
85 analysis (C). Paired-end sequencing data from dRNA-seq were used to independently identify  
86 sRNAs in D39W. Analysis was performed with sequencing reads (single-end and paired-end)  
87 that mapped to the D39W reference genome. Details of the experimental design and analysis  
88 are described in Materials and Methods.





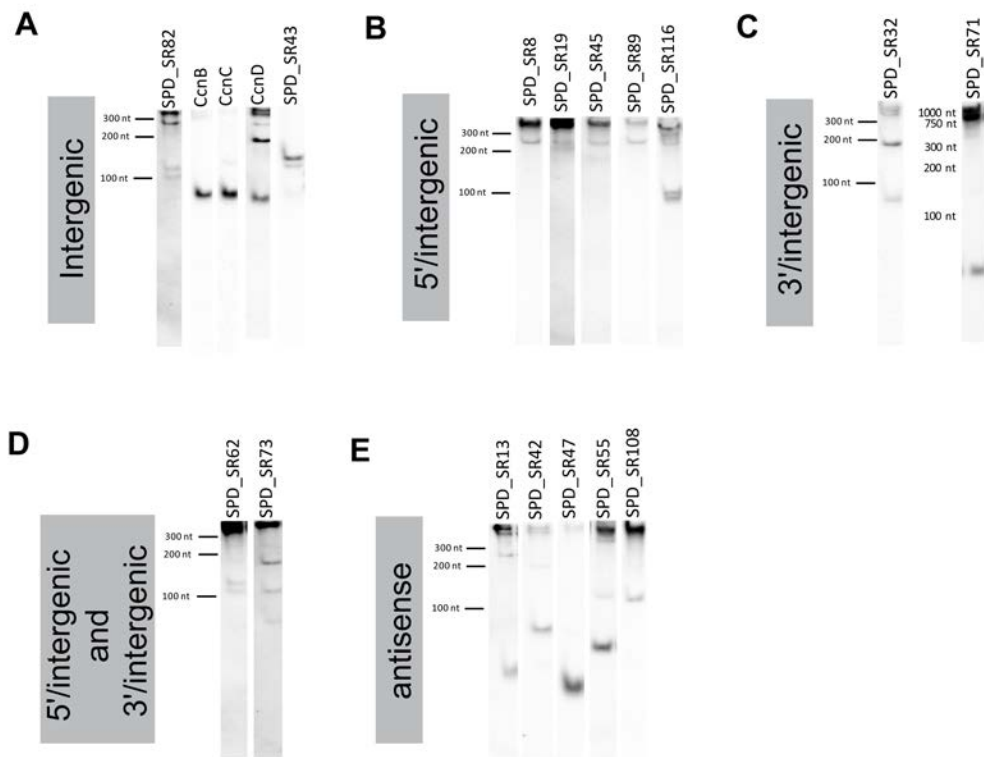
**Fig. S3. Northern blots corresponding to sRNAs identified in sRNA-seq.** Northern blot

validations corresponding to the sRNAs identified as intergenic, 5'/intergenic, 3'/intergenic,

5'/intergenic and 3'/intergenic or antisense by sRNA-seq analysis. Probes used for northern

blotting are listed in Table S5.

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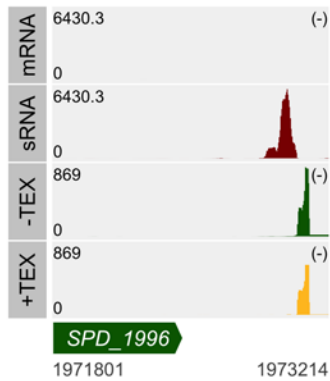


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**Fig. S4. Northern blots corresponding to sRNAs identified in dRNA-seq.** Northern blot

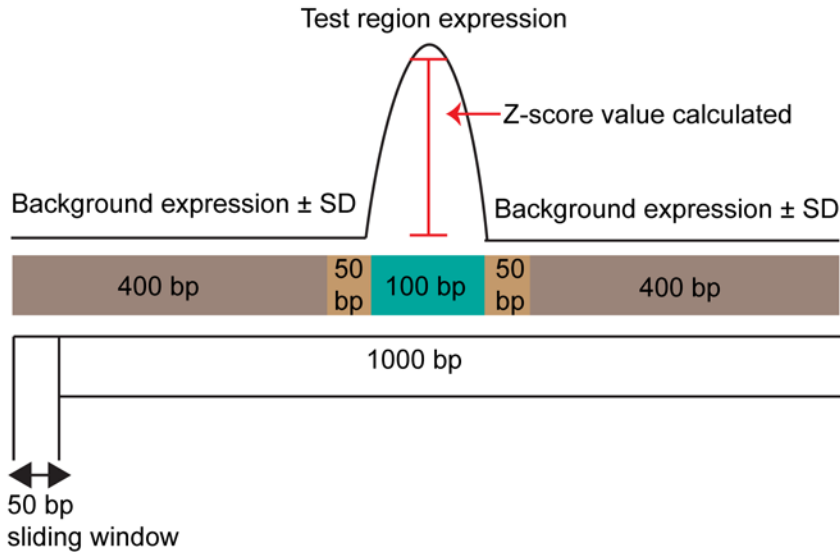
validations corresponding to the sRNAs identified as intergenic, 5'/intergenic, 3'/intergenic, 5'/intergenic and 3'/intergenic or antisense by dRNA-seq analysis. Probes used for northern blotting are listed in Table S5.

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**Fig. S5. Coverage graph of Spd\_sr110, identified as an sRNA in D39W.** Spd\_sr110 was

predicted to be present in D39V (2), but was not identified as a sRNA by RNA-seq analysis in D39V. Spd\_sr110 identified as a sRNA by dRNA-seq analysis in D39W represented by the peaks in –TEX (in green) and + TEX (in yellow) tracks. Track labels corresponding to read coverage maps are described in the legend of Figure 2. The peak in the sRNA track corresponds to Spd\_sr109 sRNA identified by sRNA-seq analysis on the (+) strand. Spd\_sr109 and Spd\_sr110 are predicted to constitute a toxin-antitoxin module in D39.



**Fig. S6. Schematic showing strategy used to identify sRNAs by RNA-seq.** A window of 1,000 base pairs in length was used. Predicted sRNAs identified as peaks in expression (100 bp test interval at the center in blue) relative to expression in neighboring portions of the genome (400 bp total to each side; flanking intervals in brown). The test area was flanked on either side by 50 bp of unused spacer sequence (in orange) to eliminate edge effects. The z-score of the test interval was determined relative to the mean and standard deviation (SD) observed in the flanking intervals. Analysis was performed genome-wide and z-scores calculated at 50 bp sliding intervals across the genome.

## SUPPLEMENTAL REFERENCES

1. **Tsui HCT, Mukherjee D, Ray VA, Sham LT, Feig AL, Winkler ME.** 2010. Identification and characterization of noncoding small RNAs in *Streptococcus pneumoniae* serotype 2 strain D39. *J Bacteriol* doi:10.1128/JB.01204-09.
2. **Slager J, Aprianto R, Veening J-W.** 2018. Deep genome annotation of the opportunistic human pathogen *Streptococcus pneumoniae* D39. *Nucleic Acids Res* **46**:9971-9989.
3. **Mann B, van Opijnen T, Wang J, Obert C, Wang Y-D, Carter R, McGoldrick DJ, Ridout G, Camilli A, Tuomanen EI, Rosch JW.** 2012. Control of virulence by small RNAs in *Streptococcus pneumoniae*. *PLoS pathog* **8**:e1002788-e1002788.
4. **Acebo P, Martin-Galiano AJ, Navarro S, Zaballos A, Amblar M.** 2012. Identification of 88 regulatory small RNAs in the TIGR4 strain of the human pathogen *Streptococcus pneumoniae*. *RNA* **18**:530-546.
5. **Halfmann A, Kovács M, Hakenbeck R, Brückner R.** 2007. Identification of the genes

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137  
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directly controlled by the response regulator CiaR in *Streptococcus pneumoniae*: five out of 15 promoters drive expression of small non-coding RNAs. Mol Microbiol **66**:110-126.

6. **Schnorpfeil A, Kranz M, Kovács M, Kirsch C, Gartmann J, Brunner I, Bittmann S, Brückner R.** 2013. Target evaluation of the non-coding csRNAs reveals a link of the two-component regulatory system CiaRH to competence control in *Streptococcus pneumoniae* R6. Mol Microbiol **89**:334-349.