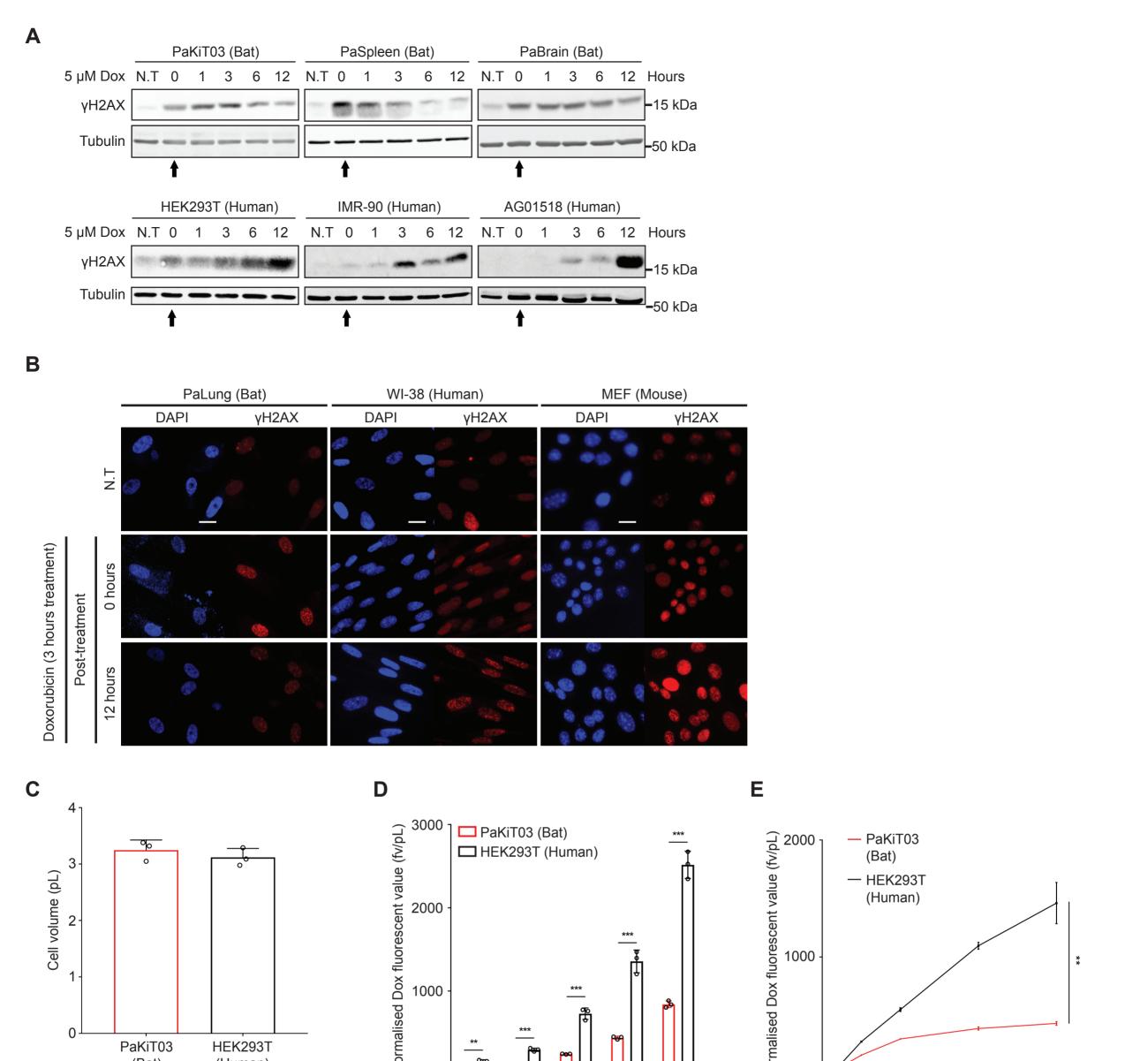


Supplementary Figure 1. Western blot analysis of γH2AX in *Pteropus alecto* and human cell lines treated with γ-irradiation

- (A) Western blot analysis of γH2AX in PaKiT03 and HEK293T cells exposed to 10 Gy of γ-irradiation. Protein lysates were harvested at the indicated time points. Tubulin was used as a loading control. N.T stands for no treatment. Blots are representative of three independent experiments.
- (B) Immunofluorescence staining of γH2AX for PaLung, WI-38 and MEF cells. Cells were treated with 10 Gy of γ-irradiation and fixed at the indicated time point. N.T stands for no treatment. Scale bars represent 20 μm. Results are representative of three independent experiments.
- (C) Western blot analysis of γH2AX in PaKiT03 and HEK293T cells. Cells were treated with 50 μM etoposide (Eto) for 3 hours, followed by drug-free medium up to 12 hours (starting at t=0 hours, indicated by arrow). Protein lysates were harvested at the indicated time points. Tubulin was used as a loading control. N.T stands for no treatment. Blots are representative of three independent experiments.



Supplementary Figure 2. Western blot analysis of yH2AX and intracellular accumulation of doxorubicin in *Pteropus alecto* and human cell lines

2.5

Dox concentration (μM)

120

Minutes

180

- (A) Western blot analysis of γH2AX. The indicated cells were treated with 5 μM doxorubicin (Dox) for 3 hours, followed by drug-free medium up to 12 hours (starting at t=0 hours, indicated by arrow). Protein lysates were harvested at the indicated time points. Tubulin was used as a loading control. N.T stands for no treatment. Blots are representative of three independent experiments.
- (B) Immunofluorescence staining of γH2AX for PaLung, WI-38 and MEF cells. Cells were treated with 5 μM doxorubicin for 3 hours, and fixed immediately (0 hour) or at 12 hours after culturing in a doxorubicin-free medium. N.T stands for no treatment. Scale bars represent 20 µm. Results are representative of three independent experiments.
- (C) The volume of PaKiT03 and HEK293T cells. Bars represent mean volume ± SD of three experiments.

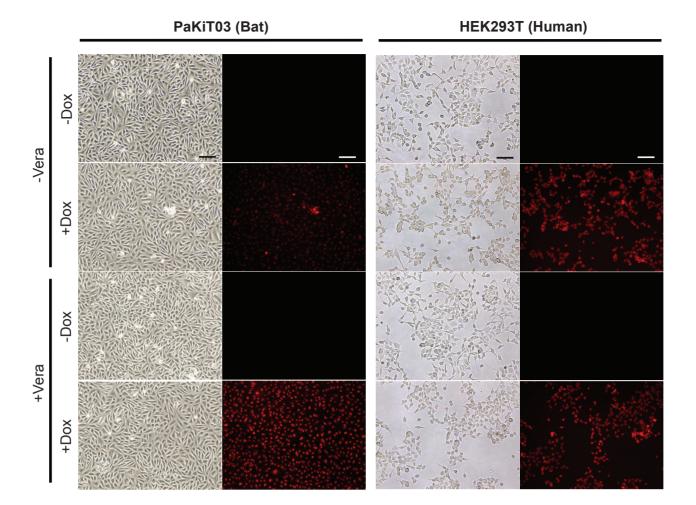
PaKiT03

(Bat)

HEK293T

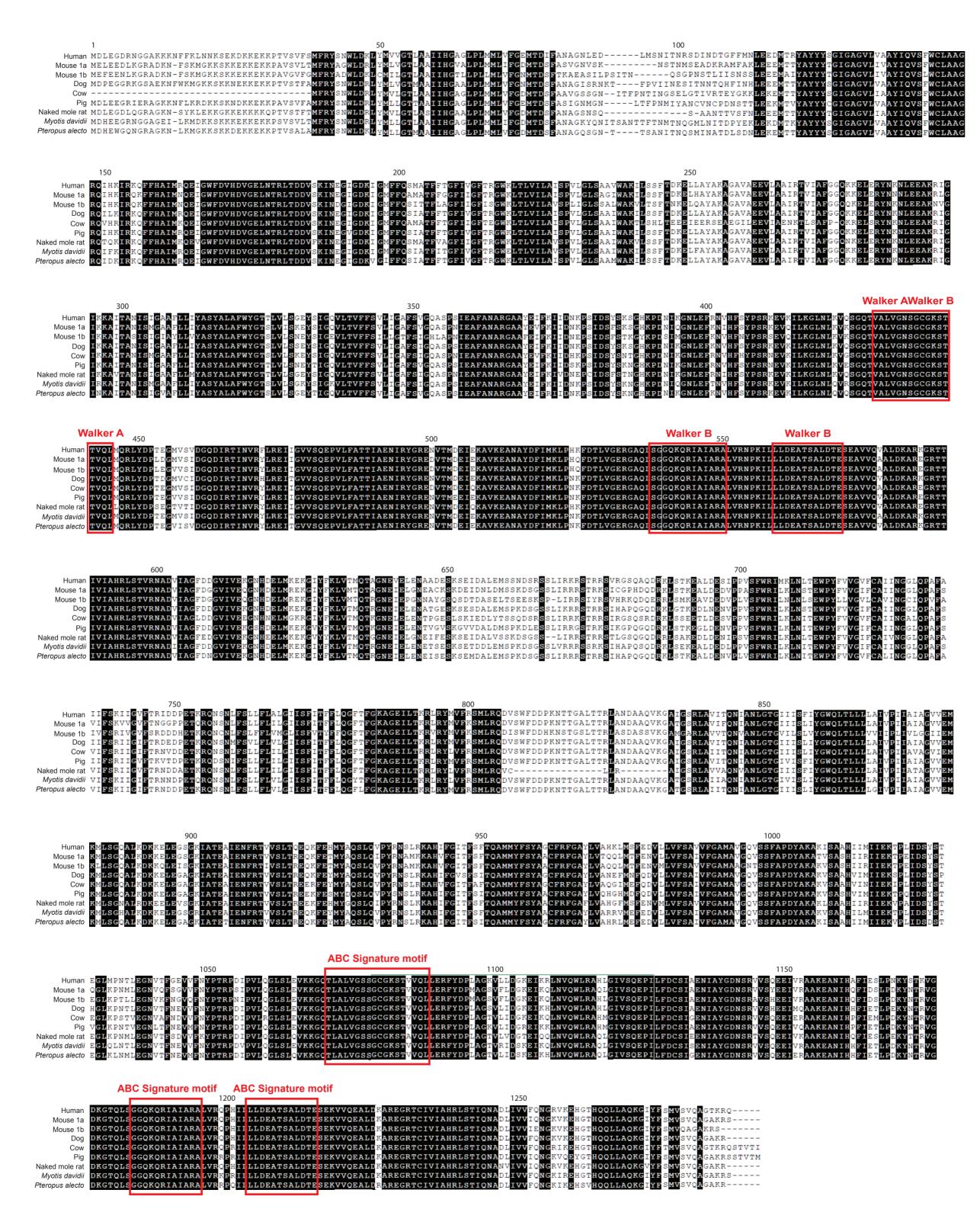
(Human)

- (D) Doxorubicin accumulation in PaKiT03 and HEK293T cells after 3 hours of incubation at the indicated concentration. The amount of accumulated doxorubicin in cells was analysed by flow cytometry. The mean fluorescence value (fv) of doxorubicin is normalised to cell volume (pL). Results were generated from three experimental repeats. Statistical significance of p < 0.01 was represented with ** and p < 0.001 with
- (E) Time course of doxorubicin accumulation in PaKiT03 and HEK293T cells. Cells were treated with 5 μM doxorubicin for the indicated time, and analysed by flow cytometry. The mean fluorescence value of doxorubicin accumulated in each cell line is normalised to cell size. Results were generated from three experimental repeats. Statistical significance of p < 0.01 was represented with **.



Supplementary Figure 3. Analysis of drug efflux capability via ABC transporters in PaKiT03 and HEK293T cells

Doxorubicin (Dox) fluorescence or phase contrast images of PaKiT03 and HEK293T cells. Cells were pre-treated without or with 5 μ M verapamil (Vera) for 30 minutes before the treatment with 10 μ M doxorubicin alone or together with verapamil for an additional 3 hours. Images were acquired with 10x objective lens and scale bar represents 100 μ m. All results were representative of at least three experimental repeats.



Supplementary Figure 4. A sequence alignment of ABCB1 protein

Amino acid sequences of ABCB1 from the indicated mammals are shown. nmr indicates naked mole rat. Amino acid residues conserved across species are highlighted in black. Walker A-, Walker B-, and ABC signature-motifs are marked with red rectangle.

Supplementary Table 1. Homology of ABCB1 protein sequence between human and other species

Mammalian species	Percentage conserved to Human ABCB1 protein (%)
Bat, Pteropus alecto	89.5
Bat, Myotis davidii	88.4
Rhesus monkey, <i>Macaca mulatta</i>	96.2
Mouse Abcb1a, Mus musculus	87.2
Mouse Abcb1b, Mus musculus	79.5
Dog, Canis lupus familiaris	90.0
Cow, Bos taurus	83.8
Pig, Sus scrofa	88.8
Cat, Felis catus	90.8
Naked mole rat, Heterocephalus glaber	84.8

Supplementary Table 2. Summary of individual or average maximum lifespan and body mass of bat species used in this study. All data are summarised from AnAge dataset, http://genomics.senescence.info/species/

Bat species	Maximum lifespan (years)	Body mass (g)
Pteropus alecto	19.7	672.1
Cynopterus brachyotis	10.1	45
Myotis species (Average)	22.13	10.4
Rhinolophus species (Average)	24.3	11.9

Supplementary Table 3. Summary of cell lines established and used in this study

Species	Cell lines	Description
Human	WI-38	Human fetal lung-derived fibroblast
	IMR-90	Human fetal lung-derived fibroblast
	AG01518	Normal human foreskin-derived fibroblast
	HEK293T	Human embryonic kidney epithelial cells, SV40 immortalized
	HepG2	Human hepatocellular carcinoma
	MDA-MB-231	Human adenocarcinoma
	HT-29	Human colorectal adenocarcinoma
	WiDr	Human colorectal adenocarcinoma
	U251MG	Human glioblastoma astrocytoma
	K562	Chronic myelogenous leukemia
	KCL22	Chronic myeloid leukemia in blast crisis
Mouse	NIH3T3	Mouse embryonic fibroblast, self-immortalized
	MEF	Mouse embryonic fibroblast, self-immortalized
Bats	PaLung	Pteropus alecto lung-derived fibroblast
	PaKidney	Pteropus alecto kidney-derived fibroblast
	PaBrain	Pteropus alecto brain-derived fibroblast, hTert immortalized
	PaSpleen	Pteropus alecto spleen-derived fibroblast, SV40 immortalized
	PaMarrow	Pteropus alecto bone marrow-derived cells, self-immortalized
	PaKiT03	Pteropus alecto kidney-derived fibroblast, SV40 immortalized
	C. brachyotis Lung	Cynoterus brachyotis primary lung-derived fibroblast,
		Pteropodidae family
	<i>M. davidii</i> Kidney	Myotis davidii kidney-derived cell line, self-immortalized,
		Vespertilionidae family
	M. muricola Lung	Myotis muricola Lung-derived cell line, self-immortalized,
		Vespertilionidae family
	R. lepidus Large Intestine	Rhinolopus lepidus primary large intestine-derived fibroblast,
		Rhinolophidae family

Figure 1A

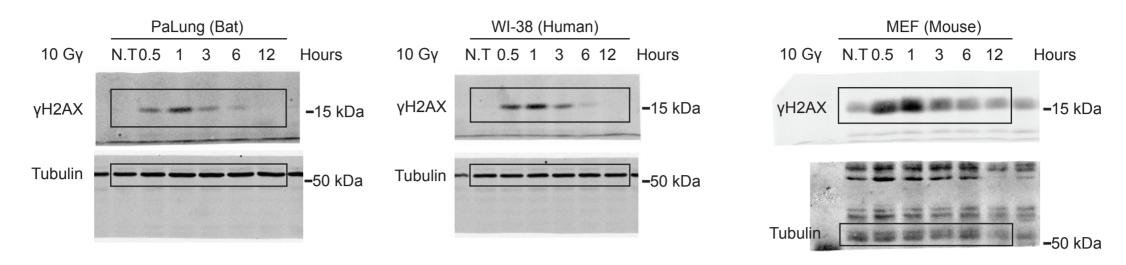
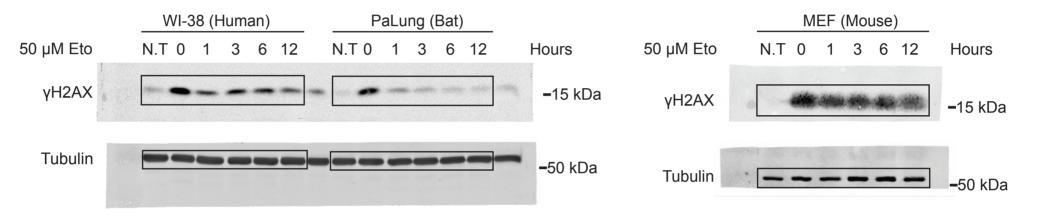


Figure 1C



Supplementary Figure 5. Uncropped Western blots for Figure 1A and 1C

Figure 2A

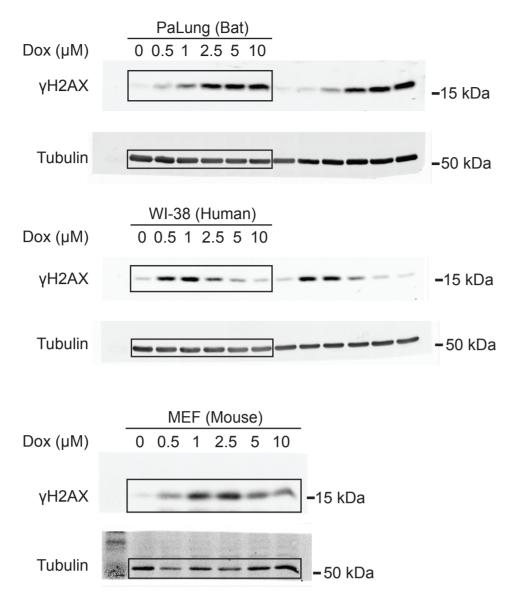
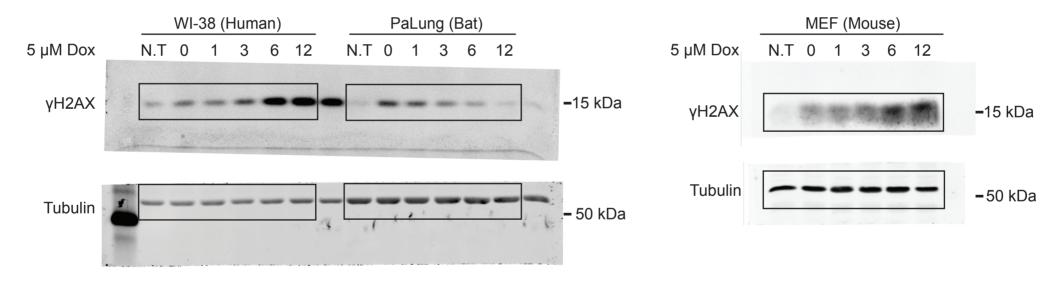
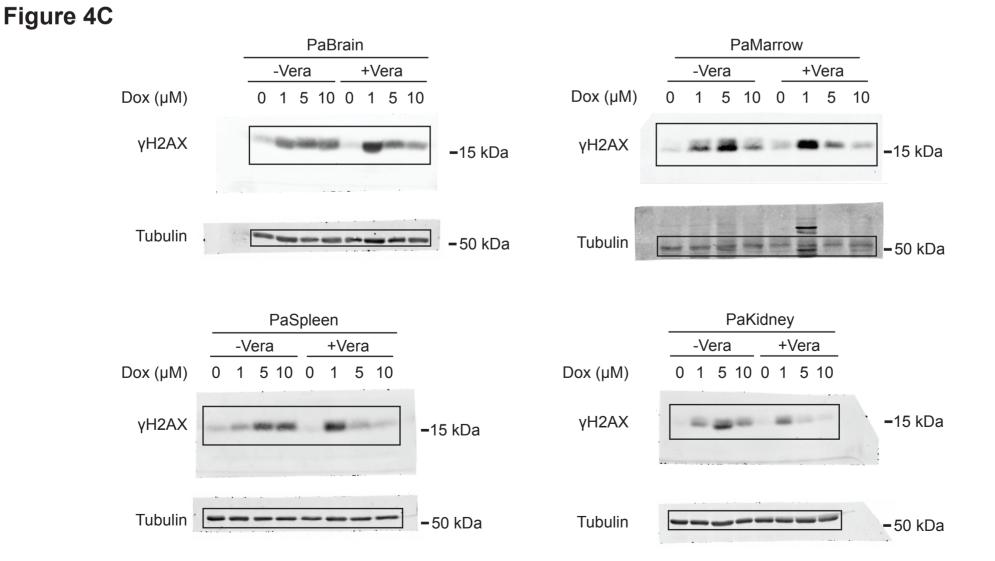


Figure 2B



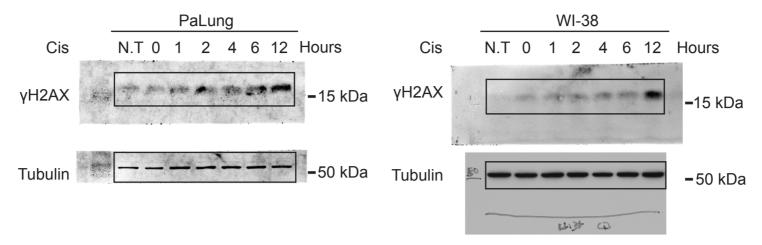
Supplementary Figure 6. Uncropped Western blots for Figure 2A and 2B.

Figure 4B Figure 4A WI-38 PaLung -CSA +CSA +Vera +Vera -Vera -Vera 0 1 5 10 0 1 5 10 Dox (µM) Dox (µM) 0 1 5 10 0 1 5 10 0 1 5 10 0 1 5 10 PaLung γΗ2ΑΧ **-**15 kDa γΗ2ΑΧ **−**15 kDa Tubulin Tubulin **−**50 kDa +CSA -CSA 0 1 5 10 0 1 5 10 Dox (µM) -Vera +Vera Dox (µM) 0 1 5 10 0 1 5 10 γΗ2ΑΧ **-**15 kDa WI-38 γΗ2ΑΧ **-**15 kDa PaKiT03 Tubulin Tubulin -CSA +CSA Dox (µM) 0 1 5 10 0 1 5 10 -Vera +Vera Dox (µM) 0 1 5 10 0 1 5 10 γΗ2ΑΧ PaKiT03 γΗ2ΑΧ **-**15 kDa HEK293T Tubulin Tubulin MEF HEK293T -CSA +CSA -CSA +CSA -Vera +Vera Dox (µM) 0 1 5 10 0 1 5 10 0 1 5 10 0 1 5 10 Dox (µM) 0 1 5 10 0 1 5 10 γΗ2ΑΧ γΗ2ΑΧ **–**15 kDa MEF Tubulin **-**50 kDa



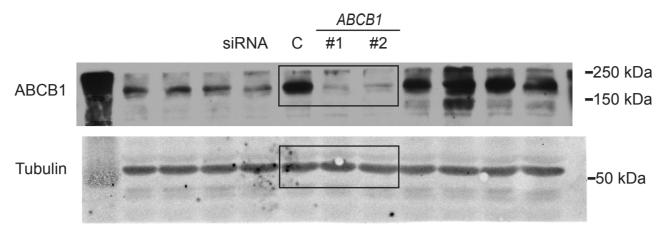
Supplementary Figure 7. Uncropped Western blots for Figure 4.

Figure 5D



Supplementary Figure 8. Uncropped Western blots for Figure 5D.

Figure 6A



Supplementary Figure 9. Uncropped Western blots for Figure 6A.

Figure 7B

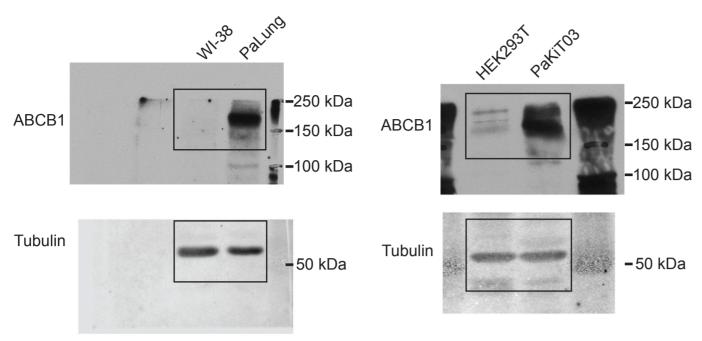
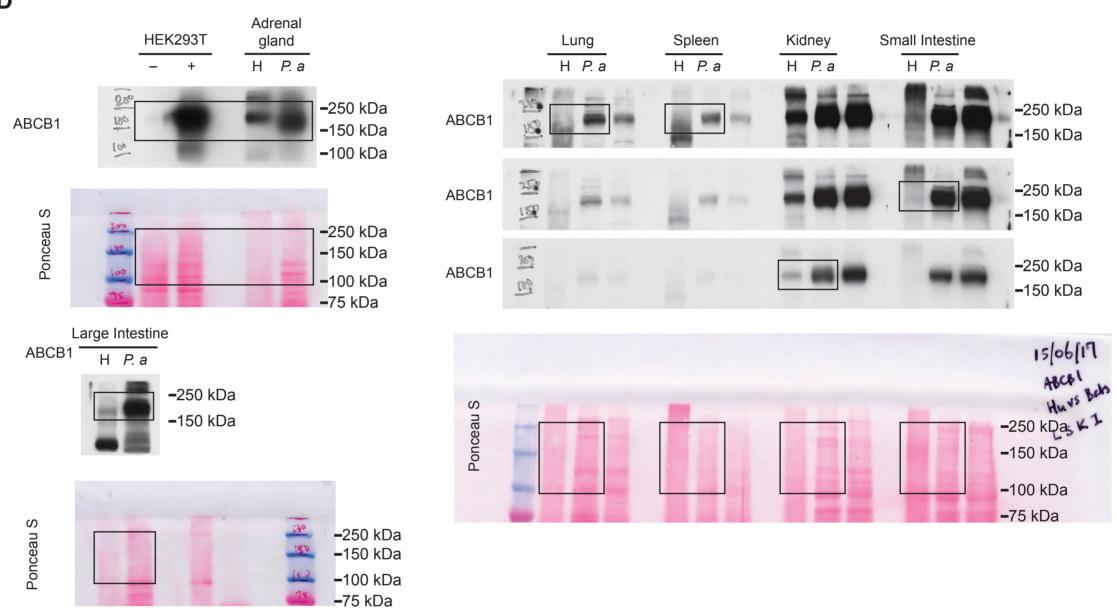


Figure 7D



Supplementary Figure 10. Uncropped Western blots for Figure 7B and 7D.

Figure 7E

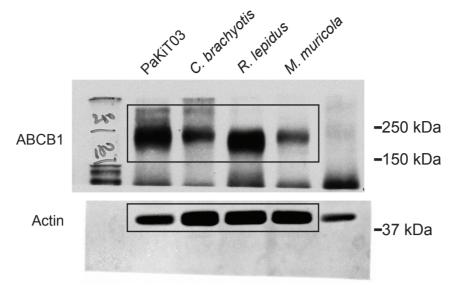
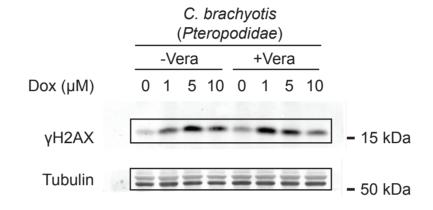
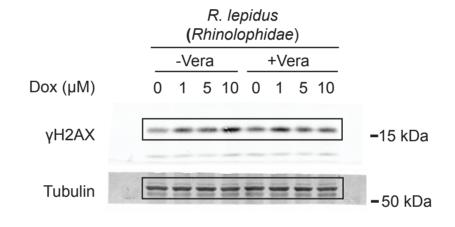
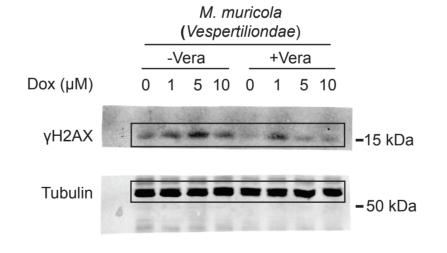


Figure 7G

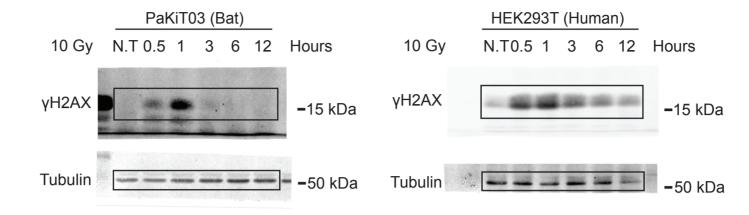




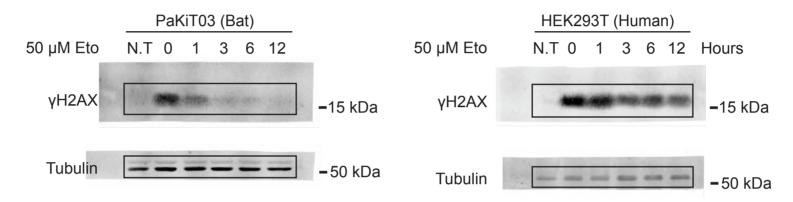


Supplementary Figure 11. Uncropped Western blots for Figure 7E and 7G.

Supplementary Figure 1A

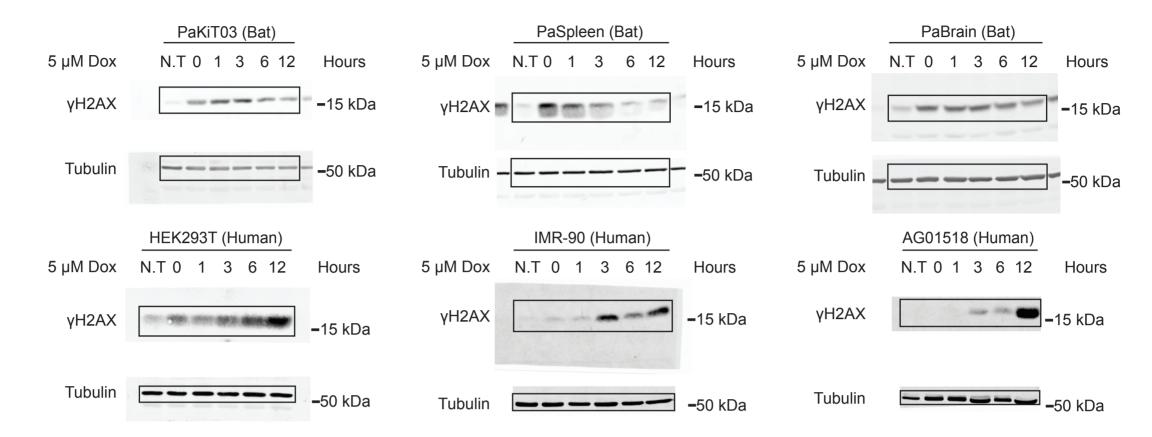


Supplementary Figure 1C



Supplementary Figure 12. Uncropped Western blots for Supplementary Figure 1A and 1C

Supplementary Figure 2A



Supplementary Figure 13. Uncropped Western blots for Supplementary Figure 2A