

## Supplementary Tables

### **Supplementary Table 1. Primers used to clone LADL plasmids**

See attached .xls file.

### **Supplementary Table 2. Primers and matched plasmids for cloning LADL constructs.**

<b>Name of insert</b>	<b>Primers</b>	<b>PCR template</b>	<b>Size (bp)</b>
<i><b>LADL Anchor Control plasmid</b></i>			
EF1a	MRP177, MRP 188	Addgene #47457	1237
3XFLAG-dCas9	MRP189, MRP 190	dCas9 plasmid	4300
GS-CIBN	MRP198, MRP 199	Addgene #47457	649
2A-Puro	MRP200, MRP 182	Addgene #62987	681
<i><b>Empty anchor Control plasmid</b></i>			
EF1a	MRP036, JV002	Addgene #47457	1237
Puro	JV001, MRP 051	Addgene #62987	681
<i><b>Empty target Control plasmid</b></i>			
EF1a	MRP177, MRP 178	Addgene #47457	1247
CRY2PHR	MRP179, MRP 183	Addgene #47457	1700
2A-mCherry	MRP184, MRP 185	CRY2olig mut 2-1	681

**Supplementary Table 3. List of gRNA primer sequences**

<b>Primer number</b>	<b>gRNA Primer name</b>	<b>gRNA sequence</b>	<b>Backbone plasmid (single clone)</b>	<b>Multiplex</b>
129	<i>Klf4_Enh_1_F</i>	CACCGTACATGCAGTAGTACTAAGT	S12.1	<b>Desert gRNAs (<i>Klf4/ Zfp462</i>)</b>
130	<i>Klf4_Enh_1_R</i>	AAACACTTAGTACTACTGCATGTAC	S12.1	
135	<i>Klf4_Enh_2_F</i>	CACCGTTTGTGTTTTAGTG TAGATT	B1	
136	<i>Klf4_Enh_2_R</i>	AAACAATCTACACTAAAACACAAAC	B1	
115	<i>Zfp462_Prom_2_F</i>	CACCGTAAAGAAAAGTGT TTTATCGA	B2	
116	<i>Zfp462_Prom_2_R</i>	AAACTCGATAAAACACTTTTCTTTAC	B2	
117	<i>Zfp462_Prom_1_F</i>	CACCGAAGTGTTTATCGAGGGAAAG	B3	
118	<i>Zfp462_Prom_1_R</i>	AAACCTTTCCCTCGATAAAACACTTC	B3	
115	<i>Zfp462_Prom_2_F</i>	CACCGTAAAGAAAAGTGT TTTATCGA	S12.1	<b>Promoter only Target plasmid (<i>Zfp462</i> only)</b>
116	<i>Zfp462_Prom_2_R</i>	AAACTCGATAAAACACTTTTCTTTAC	S12.1	
117	<i>Zfp462_Prom_1_F</i>	CACCGAAGTGTTTATCGAGGGAAAG	B1	
118	<i>Zfp462_Prom_1_R</i>	AAACCTTTCCCTCGATAAAACACTTC	B1	

**Supplementary Table 4. List of plasmids with individual gRNAs without soluble CRY2**

<b>gRNA in plasmid</b>	<b>gRNA Primer name</b>	<b>BbsI digested plasmid backbone</b>	<b>Targeting region</b>	<b>Genomic feature of target</b>	<b>Refer to</b>
129	<i>Klf4_Enh_1_F</i>	S12.1	Engineered Loop Anchor 2	Desert near SE	<b>Supplementary Figure 2e</b>
135	<i>Klf4_Enh_2_F</i>	B1 (Addgene # 58778)	Engineered Loop Anchor 2	Desert near SE	<b>Supplementary Figure 2f</b>
115	<i>Zfp462_Prom_2_F</i>	B2 (Addgene # 58779)	Engineered Loop Anchor 1	Desert near <i>Zfp462</i> TSS	<b>Supplementary Figure 2g</b>
117	<i>Zfp462_Prom_1_F</i>	B3 (Addgene # 58780)	Engineered Loop Anchor 1	Desert near <i>Zfp462</i> TSS	<b>Supplementary Figure 2h</b>

**Supplementary Table 5. List of plasmids with multiplexed gRNAs without soluble CRY2**

<b>gRNAs in plasmid</b>	<b>gRNA Primer name</b>	<b>BsaI digest plasmid backbone</b>	<b>Targeting region</b>	<b>Multiplex Plasmid name</b>	<b>Refer to</b>
129 135 115 117	<i>Klf4_Enh_1_F</i> <i>Klf4_Enh_2_F</i> <i>Zfp462_Prom_2_F</i> <i>Zfp462_Prom_1_F</i>	S12.1	Desert near SE and Desert near <i>Zfp462</i> TSS	Empty bridge control plasmid	<b>Supplementary Figure 2i</b>

**Supplementary Table 6. List of plasmids with multiplexed gRNAs with soluble CRY2**

<b>gRNAs in plasmid</b>	<b>gRNA Primer name</b>	<b>BsaI digest plasmid backbone</b>	<b>Targeting region</b>	<b>Multiplex Plasmid name</b>	<b>Refer to</b>
129 135 115 117	<i>Klf4_Enh_1_F</i> <i>Klf4_Enh_2_F</i> <i>Zfp462_Prom_2_F</i> <i>Zfp462_Prom_1_F</i>	S12.1	Desert near SE and Desert near <i>Zfp462</i> TSS	LADL Bridge + Target	<b>Supplementary Figure 2l</b>
115 117	<i>Zfp462_Prom_2_F</i> <i>Zfp462_Prom_1_F</i>	S12.1	Desert near <i>Zfp462</i> TSS	One-sided guide Control	<b>Supplementary Figure 2m</b>

**Supplementary Table 7. List of primers used for RNA qRT-PCR**

<b>Name</b>	<b>Sequence</b>
Nanog_F_MR137	TGCCTGCAGTTTTTCATCCC
Nanog_R_MR138	TAGAAGAATCAGGGCTGCCTTG
Mouse_GAPDH_F_MR141	GCACAGTCAAGGCCGAGAAT
Mouse_GAPDH_R_MR142	GCCTTCTCCATGGTGGTGAA
Sox2_F_MR143	GCACATGAACGGCTGGAGCAACG
Sox2_R_MR144	TGCTGCGAGTAGGACATGCTGTAGG
<i>Klf4</i> _F_MR147	AGACCAGATGCAGTCACAAGTC
<i>Klf4</i> _R_MR148	TTTTGCCACAGCCTGCATAG
Oct4_F_MR151	TGTGGACCTCAGGTTGGACT
Oct4_R_MR152	TTTCATGTCCTGGGACTCCTC
MRP213_qPCR_ <i>Zfp462</i> _F	GCCAACTGATGTTGCCGAGGACAATG
MRP214_qPCR_ <i>Zfp462</i> _R	CCTGAAGTAGCGTACGCAGAACTTG
MRP215_qPCR_Zhang_dCas9_F	GCACAGCATCAAGAAGAACCTG
MRP216_qPCR_Zhang_dCas9_R	CGTTGCTGAAGATCTCTTGCGAG
MRP149_CRY2_qPCR_F	AATGCCTCGACATGTCCATC
MRP150_CRY2_qPCR_F	AGCGCGTTACTGGGTTTTTC
Nestin_Fwd_JB	AGGCCACTGAAAAGTTCCAG
Nestin_Rev_JB	TAAGGGACATCTTGAGGTGTGC

**Supplementary Table 8: Summary of external sequencing libraries analyzed in this study**  
See attached .xls file.

**Supplementary Table 9. List of primers used for Chromatin Immunoprecipitation**

<b>Primer number</b>	<b>ChIP Primer name</b>	<b>Sequence</b>	<b>Target region</b>
MRP217	MRP217_IP_Down_Y _ED_129to135_F	TGGGCCTACTTAGTACTACTGC	Engineered site at SE
MRP218	MRP218_IP_Down_Y _ED_129to135_R	GCTGGGTAAGTAGCCCTCTAC	Engineered site at SE
MRP221	MRP221_IP_Down_H _ED_115to117_F	AAGCCCCTTTCCCTCGATAAAC	Engineered site at <i>Zfp462</i> promoter
MRP222	MRP222_IP_Down_H _ED_115to117_R	ACACTAGGAGGATGGGGATAGTC	Engineered site at <i>Zfp462</i> promoter
MRP223	MRP223_IP_H_CTCF _149to155_F	GCTCTATGTTCTAACACCTCTCC	Negative control
MRP224	MRP224_IP_H_CTCF _149to155_R	CGTGCTTGTACACACACACAG	Negative control

**Supplementary Table 10. 5C Primer Sequences.**

See attached .xls file.

**Supplementary Table 11. 5C Primer Genomic Coordinates.**

See attached .xls file.

**Supplementary Table 12: Summary of mapped 5C sequencing reads**

See attached .xls file.

**Supplementary Table 13. Fluorescence-labeled oligonucleotide sequences for RNA FISH.**

See attached .xls file.