

Supplementary Information

N^ε-acetyl-lysine derivatives with Zinc binding groups as novel HDAC inhibitors

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General information and materials

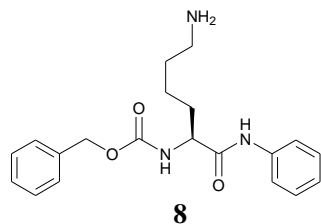
General: The following materials were obtained from commercial sources for the compound preparation, and were used as received without further treatment, and all reagents and solvents were commercial high purity quality. The purity of all tested compounds was over 95% by HPLC.

Materials: Energy Chemical ShangHai: N-alpha-Cbz-L-lysine, N-methylmorpholine (NMM), Isobutyl chloroformate (IBCF), Trifluoroacetic acid (TFA), Triethylamine (TEA), Tetrahydrofuran (DMF), 4-dimethylamino pyridine (DMAP), tert-butyl alcohol, 2-(1H-benzotriazole-1-yl)-1,1,3,3-tetramethylaminium hexafluorophosphate (HBTU), N,N-Diisopropylethylamine (DIEA), Hydroxylamine hydrochloride, o-Phenylenediamine, Succinic anhydride, Maleic anhydride, Nicotinic acid, 3-Indoleformic acid, 6-Methylpyridine-3-carboxylic acid, 3-(3-pyridyl) acrylic acid), Pyrrole-2-carboxylic acid, 2-Furoic acid, 2-Thiophenecarboxylic acid, Mercaptoacetic acid, 3-Quinolinecarboxylic acid, 6-Quinolinecarboxylic acid, 3-Indoleformic acid, 1H-Indene-3-carboxylic acid, Indole-3-carboxaldehyde, 1-Benzothiophene-3-carboxylic acid, Benzofuran-3-carboxylic acid, (2,4-Dioxo-1,3-thiazolidin-5-yl)acetic acid, N-[(tert-Butoxy)carbonyl]-L-tryptophan, Suberic acid, Monomethyl ester, Monomethyl Malonate, Monomethyl adipate. Aladdin ShangHai: LiOH. Boc-Lys(Ac)-AMC .

¹H and ¹³C-NMR were performed on JEOL ESC-400 spectrometers.

Synthesis and characterization of new compounds

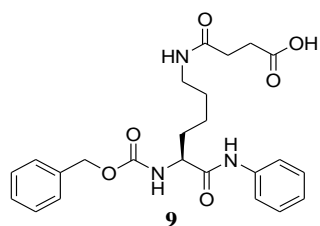
(8): 2-(benzyloxycarbonyl)-6-(tert-butoxycarbonyl)hexanoic acid (5.3g, 13.9mmol), DIEA



(4.8mL, 27.8mmol) and HBTU (7.9g, 20.85mmol) was added in anhydrous THF (100mL), and then the aniline (1.9mL, 20.85mmol) was added, after that the reaction was stirred 3h. The THF was removed under reduced pressure, the reaction mixture was diluted

with CH₂Cl₂, and saturated NaCl solution, the organic layer was washed with saturated NaCl, and dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography (5.95g, 93%). The compound **7** purified above (5.95g, 13.07mmol) was added in anhydrous CH₂Cl₂ (60mL) and cooled (0°C), and then the TFA (30mL) was added slowly, stirred for 2h. Then, the TFA was removed under reduced pressure until there was no TFA, the residue was purified by silica gel column chromatography (4.4g, 95%). ¹H NMR (400 MHz, CD₃OD) δ 7.51 (d, *J* = 7.2 Hz, 2H), 7.27 (dt, *J* = 33.0 Hz, 18.4, 7H), 7.07 (t, *J* = 7.2 Hz, 1H), 5.13 – 5.01 (m, 2H), 4.21 (d, *J* = 17.2 Hz, 1H), 2.87 (t, *J* = 7.3 Hz, 2H), 1.92 – 1.58 (m, 4H), 1.56 – 1.33 (m, 2H). ¹³C NMR (100 MHz, CD₃OD) δ 171.76, 157.43, 137.96, 136.86, 128.51, 127.90, 124.39, 120.35, 66.71, 55.31, 39.63, 32.02, 27.40, 22.25. HRMS (ESI) *m/z* calculated for C₂₀H₂₅N₃O₃⁺ (M+H)⁺ 356.1974, found 356.1982.

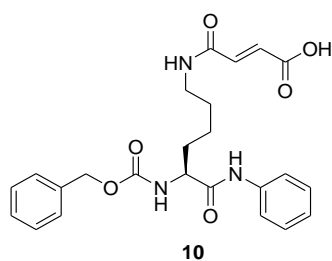
(9): Succinic anhydride (56mg, 0.84mmol) was dissolved in anhydrous THF (2mL) and TEA



(233ul, 1.68mmol), and the compound **8** (200mg, 0.56mmol) was added slowly, the mixture was stirred for 3 hours at RT. Then, the THF was removed under reduced pressure, the residue was purified by silica gel column chromatography (230mg, 90%). ¹H NMR (400

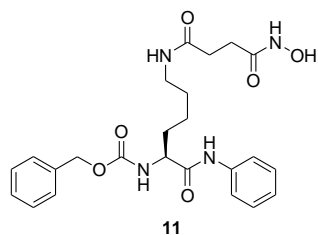
MHz, DMSO-*d*₆) δ 12.03 (s, 1H, -COOH), 9.95 (d, *J* = 22.5 Hz, 1H, -CONH-), 7.81 – 7.71 (m, 1H, -CONH-), 7.56 – 7.42 (m, 3H), 7.35 – 7.14 (m, 7H), 6.99 (dd, *J* = 19.7Hz, 12.3, 1H), 5.01 – 4.88 (m, 2H), 4.11 – 3.98 (m, 1H), 2.97 (d, *J* = 5.5 Hz, 2H), 2.38 – 2.29 (m, 2H), 2.25 – 2.17 (m, 2H), 1.57 (dd, *J* = 11.2 Hz, 6.8, 2H), 1.38 – 1.22 (m, 4H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 174.64, 171.76, 171.25, 156.63, 139.40, 137.30, 129.22, 128.94, 128.27, 123.75, 119.66, 66.12, 55.71, 31.95, 30.45, 29.70, 29.47, 23.64. HRMS (ESI) *m/z* calculated for C₂₄H₃₀N₃O₆⁺ (M+H)⁺ 456.2135, found 456.2193.

(10): The Maleic anhydride (50mg, 0.5mmol) was dissolved in anhydrous THF (2mL) and TEA



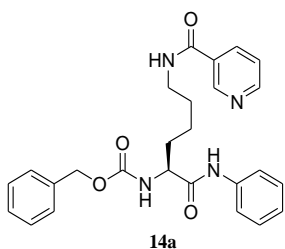
(250ul, 1.5mmol), and the compound **8** (180mg, 0.5mmol) was added slowly, the mixture was stirred for 3 hours at RT. Then, the THF was removed under reduced pressure, the residue was purified by silica gel column chromatography (205mg, 90%). ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.98 (d, *J* = 11.7 Hz, 1H, -CONH-), 9.10 (d, *J* = 5.8 Hz, 1H, -CONH-), 7.60 – 7.47 (m, 3H), 7.36 – 7.12 (m, 8H), 7.03 – 6.95 (m, 1H), 6.38 – 6.30 (m, 1H), 6.19 (dd, *J* = 15.1 Hz, 9.6, 1H), 4.98 (d, *J* = 12.2 Hz, 2H), 4.07 (s, 1H), 3.10 (dd, *J* = 12.0, 6.0 Hz, 2H), 1.68 – 1.51 (m, 2H), 1.50 – 1.25 (m, 4H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 171.59, 165.83, 156.58, 139.68, 137.52, 133.68, 132.33, 129.28, 128.87, 128.32, 128.25, 123.49, 120.18, 66.59, 55.64, 31.68, 28.37, 23.62. HRMS (ESI) *m/z* calculated for C₂₄H₂₈N₃O₆³⁺ (M+H)³⁺ 456.2135, found 456.2192.

(11): The compound **9** (70mg, 0.154mmol) was added in anhydrous THF (3mL), and cooled to 0°C and then the TEA(43μL, 0.308mmol), IBCF (30μL, 0.2316mmol) was added slowly, the mixture was stirred for 1 hours at 0°C to rt, and added Hydroxylamine hydrochloride (53mg, 0.77mmol) in Methanol, after that the reaction was stirred 3h. The THF was removed under reduced pressure, the reaction mixture was diluted with CH₂Cl₂, and saturated NaCl solution, the organic layer was washed with saturated NaCl, and dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography (30mg, 41%). ¹H NMR (400 MHz, CD₃OD) δ 7.56 – 7.41 (m, 2H), 7.37 – 7.11 (m, 7H), 7.05 (dd, *J* = 20.2, 12.8 Hz, 1H), 5.19 – 4.96 (m, 2H), 4.26 – 4.03 (m, 1H), 3.20 – 3.07 (m, 2H), 2.43 (dd, *J* = 13.0, 6.1 Hz, 2H), 2.31 (dd, *J* = 15.9 Hz, 8.7, 2H), 1.91 – 1.61 (m, 2H), 1.62 (s, 4H). ¹³C NMR (100 MHz, CD₃OD) δ 173.10, 172.03, 170.07, 157.45, 138.25, 136.59, 128.14, 124.27, 120.12, 66.61, 55.64, 38.62, 32.05, 30.66, 29.04, 28.50, 27.66, 24.93, 22.16. HRMS (ESI) *m/z* calculated for C₂₄H₃₁N₄O₆⁺ (M+H)⁺ 471.2244, found 471.2232.



0°C and then the TEA(43μL, 0.308mmol), IBCF (30μL, 0.2316mmol) was added slowly, the mixture was stirred for 1 hours at 0°C to rt, and added Hydroxylamine hydrochloride (53mg, 0.77mmol) in Methanol, after that the reaction was stirred 3h. The THF was removed under reduced pressure, the reaction mixture was diluted with CH₂Cl₂, and saturated NaCl solution, the organic layer was washed with saturated NaCl, and dried over Na₂SO₄ and concentrated. The residue was purified by silica gel column chromatography (30mg, 41%). ¹H NMR (400 MHz, CD₃OD) δ 7.56 – 7.41 (m, 2H), 7.37 – 7.11 (m, 7H), 7.05 (dd, *J* = 20.2, 12.8 Hz, 1H), 5.19 – 4.96 (m, 2H), 4.26 – 4.03 (m, 1H), 3.20 – 3.07 (m, 2H), 2.43 (dd, *J* = 13.0, 6.1 Hz, 2H), 2.31 (dd, *J* = 15.9 Hz, 8.7, 2H), 1.91 – 1.61 (m, 2H), 1.62 (s, 4H). ¹³C NMR (100 MHz, CD₃OD) δ 173.10, 172.03, 170.07, 157.45, 138.25, 136.59, 128.14, 124.27, 120.12, 66.61, 55.64, 38.62, 32.05, 30.66, 29.04, 28.50, 27.66, 24.93, 22.16. HRMS (ESI) *m/z* calculated for C₂₄H₃₁N₄O₆⁺ (M+H)⁺ 471.2244, found 471.2232.

(14a): The Nicotinic acid (52mg, 0.42mmol) was added in anhydrous THF (3.5mL), and then the

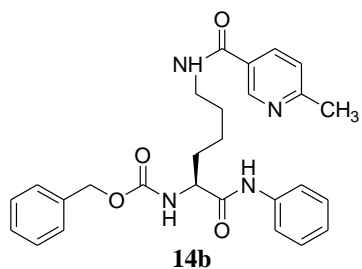


DIEA (98 μ L, 0.56mmol), HBTU(160mg, 0.42mmol), and The compound **8** (100mg, 0.28mmol), which was dissolved in THF was added, after that the reaction was stirred 4h. The THF was removed under reduced pressure, the reaction mixture was diluted with CH_2Cl_2 , and saturated NaCl solution, the organic layer was washed

with saturated NaCl, and dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography (81mg, 62%). ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.96 (s, 1H, -CONH-), 8.97 – 8.93 (m, 1H, -CONH-), 8.66 – 8.63 (m, 1H, -CONH-), 8.60 (t, $J = 5.0$ Hz, 1H), 8.12 – 8.09 (m, 1H), 7.53 (dd, $J = 16.6, 8.0$ Hz, 3H), 7.43 (dd, $J = 7.6, 5.1$ Hz, 1H), 7.33 – 7.21 (m, 7H), 7.00 (dd, $J = 11.0, 3.7$ Hz, 1H), 4.99 (s, 2H), 4.11 (dd, $J = 13.5, 8.0$ Hz, 1H), 3.23 (dd, $J = 12.3, 6.2$ Hz, 2H), 1.69 – 1.58 (m, 2H), 1.54 – 1.30 (m, 4H). ^{13}C NMR (100MHz, $\text{DMSO-}d_6$) δ 171.82, 165.11, 156.80, 152.16, 149.01, 139.44, 137.36, 135.52, 130.61, 129.23, 128.87, 128.33, 128.26, 123.93, 119.72, 66.00, 55.99, 32.06, 29.31, 23.64. HRMS (ESI) m/z calculated for $\text{C}_{26}\text{H}_{29}\text{N}_4\text{O}_4^+$ ($\text{M}+\text{H}$) $^+$ 461.2189, found 461.2233.

The synthesis of compounds **14b-p**, **15a-c** and **17a-c** were similar to the synthesis of compound **14a**. The yield of these compounds was about 62%~95%.

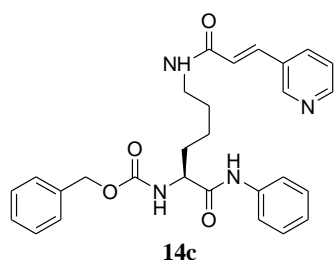
(14b): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.95 (s, 1H, -CONH-), 8.83 (d, $J = 2.0$ Hz, 1H, -CONH-),



8.51 (t, $J = 5.5$ Hz, 1H, -CONH-), 8.00 (dd, $J = 8.1, 2.3$ Hz, 1H), 7.52 (dd, $J = 17.6, 7.8$ Hz, 3H), 7.33 – 7.23 (m, 8H), 7.00 (t, $J = 7.1$ Hz, 1H), 4.99 (s, 2H), 4.10 (dd, $J = 13.5, 8.2$ Hz, 1H), 3.22 (dd, $J = 12.8, 6.5$, 2H), 2.47 (s, 5H), 1.72 – 1.59 (m, 2H), 1.54 – 1.29 (m, 4H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.91, 165.26,

161.24, 156.68, 148.67, 139.62, 137.51, 135.63, 129.23, 128.87, 128.33, 128.25, 127.83, 123.14, 119.72, 66.08, 55.83, 31.98, 29.16, 24.48, 23.64. HRMS (ESI) m/z calculated for $\text{C}_{27}\text{H}_{31}\text{N}_4\text{O}_4^+$ ($\text{M}+\text{H}$) $^+$ 475.2345, found 475.2423.

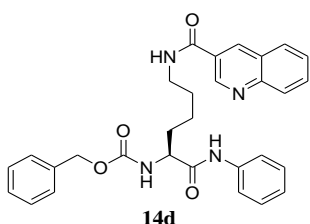
(14c): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.96 (s, 1H, -CONH-), 8.69 (d, $J = 1.5$ Hz, 1H, -CONH-),



8.50 (dd, $J = 4.7, 1.2$ Hz, 1H, -CONH-), 8.13 (t, $J = 5.5$ Hz, 1H),
7.91 (d, $J = 8.0$ Hz, 1H), 7.55 (d, $J = 7.9$ Hz, 2H), 7.50 (d, $J =$
7.8 Hz, 1H), 7.42-7.36 (m, 2H), 7.31 (d, $J = 4.3$ Hz, 4H),
7.27-7.21 (m, 3H), 6.99 (t, $J = 7.4$ Hz, 1H), 6.64 (s, 1H), 4.99 (s,
2H), 4.09 (dd, $J = 13.3, 8.1$ Hz, 1H), 3.13 (dd, $J = 12.6, 6.5$ Hz,

2H), 1.67 – 1.57 (m, 2H), 1.40 (dd, $J = 19.0, 4.6$ Hz, 4H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ
171.59, 164.99, 156.50, 150.79, 149.22, 141.67, 139.16, 137.26, 131.61, 129.44, 128.76, 128.50,
124.68, 123.73, 119.99, 65.54, 31.49, 29.66, 23.93. HRMS (ESI) m/z calculated for $\text{C}_{28}\text{H}_{31}\text{N}_4\text{O}_4^+$
($\text{M}+\text{H}$) $^+$ 487.2345, found 487.2393.

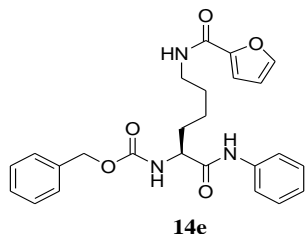
(14d): ^1H NMR (400 MHz, CD_3OD) δ 11.32 (s, 1H, -CONH-), 9.99 (d, $J = 26.6$ Hz, 1H,



-CONH-), 8.03 – 7.81 (m, 1H, -CONH-), 7.54 (t, $J = 9.8$ Hz, 3H),
7.38 – 7.09 (m, 7H), 6.97 (dt, $J = 10.7, 4.9$ Hz, 1H), 6.80 – 6.61 (m,
2H), 6.03 – 5.98 (m, 1H), 5.08 – 4.86 (m, 2H), 4.14 – 3.99 (m, 1H),
3.21 – 3.06 (m, 2H), 1.71 – 1.53 (m, 2H), 1.53 – 1.28 (m, 4H). ^{13}C

NMR (100 MHz, CD_3OD) δ 171.54, 161.06, 156.43, 153.37, 149.00, 139.57, 138.20, 129.10,
128.57, 128.25, 126.65, 123.32, 121.42, 119.74, 109.81, 108.74, 65.70, 32.06, 29.31, 23.56.
HRMS (ESI) m/z calculated for $\text{C}_{30}\text{H}_{31}\text{N}_4\text{O}_4^+$ ($\text{M}+\text{H}$) $^+$ 511.2345, found 511.2377.

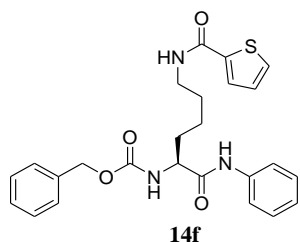
(14e): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.95 (s, 1H, -CONH-), 8.09 (dd, $J = 10.9, 3.0$ Hz, 2H),



7.64 (t, $J = 5.7$ Hz, 1H, -CONH-), 7.52 (dd, $J = 18.2, 7.8$ Hz, 3H),
7.34 – 7.22 (m, 7H), 7.00 (t, $J = 7.3$, 1H), 6.78 (s, 1H), 4.99 (s, 2H),
4.09 (d, $J = 5.2$ Hz, 1H), 3.14 (d, $J = 5.9$ Hz, 2H), 1.69 – 1.58 (m,
2H), 1.40 (ddd, $J = 37.4, 19.4, 10.9$ Hz, 4H). ^{13}C NMR (100 MHz,

$\text{DMSO-}d_6$) δ 171.69, 161.96, 156.57, 145.42, 144.42, 139.56, 137.53, 129.31, 128.87, 128.26,
123.34, 119.72, 109.45, 65.99, 55.94, 38.89, 32.08, 29.63, 23.65. HRMS (ESI) m/z calculated for
 $\text{C}_{25}\text{H}_{28}\text{N}_3\text{O}_5^+$ ($\text{M}+\text{H}$) $^+$ 450.2029, found 450.2073.

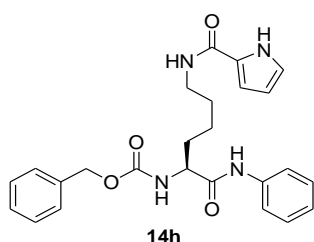
(14f): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 10.01 (s, 1H, -CONH-), 8.28 (t, $J = 5.4$ Hz, 1H, -CONH-),



8.05 (dd, $J = 12.2, 11.2$ Hz, 1H, -CONH-), 7.60 – 7.51 (m, 4H), 7.46 (d, $J = 5.0$ Hz, 1H), 7.31 (dt, $J = 12.6, 5.9$ Hz, 7H), 7.03 (t, $J = 7.3$ Hz, 1H), 5.01 (s, 2H), 4.12 (dd, $J = 13.3, 8.0$ Hz, 1H), 3.20 (dd, $J = 12.4, 6.3$ Hz, 2H), 1.75 – 1.58 (m, 2H), 1.56 – 1.31 (m, 4H). ^{13}C

NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.75, 162.54, 156.62, 139.45, 138.49, 137.52, 129.24, 128.88, 128.85, 128.34, 128.27, 127.30, 127.10, 123.77, 119.72, 65.87, 55.92, 32.20, 29.50, 23.82.

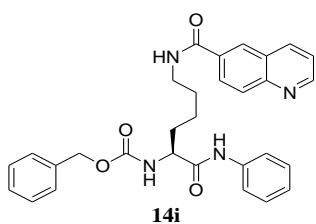
(14h): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.99 (d, $J = 10.0$ Hz, 1H, -CONH-), 9.23 (t, $J = 3.0$ Hz,



1H, -CONH-), 8.84 – 8.70 (m, 2H, -CONH-), 8.10 – 7.97 (m, 2H), 7.82 (ddd, $J = 8.4, 6.9, 1.4$ Hz, 1H), 7.71 – 7.63 (m, 1H), 7.63 – 7.44 (m, 3H), 7.41 – 7.10 (m, 7H), 7.06 – 6.92 (m, 1H), 5.13 – 4.85 (m, 2H), 4.18 – 4.06 (m, 1H), 3.40 – 3.20 (m, 6H), 1.82 –

1.52 (m, 4H), 1.50 – 1.33 (m, 2H). ^{13}C NMR (100MHz, $\text{DMSO-}d_6$) δ 171.42, 165.02, 156.49, 149.51, 148.70, 139.64, 137.51, 135.61, 131.61, 129.72, 129.22, 128.69, 128.39, 127.59, 123.61, 119.86, 65.94, 56.01, 32.26, 29.32, 25.05, 23.72, 22.68. HRMS (ESI) m/z calculated for $\text{C}_{25}\text{H}_{29}\text{N}_4\text{O}_4\text{Na}^+$ ($\text{M}+\text{H}$) $^+$ 471.2008, found 471.2006.

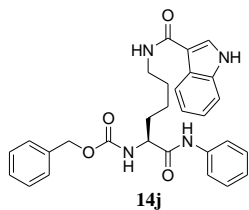
(14i): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.96 (s, 1H, -CONH-), 8.93 (s, 1H, -CONH-), 8.66 (s, 1H,



-CONH-), 8.45 – 8.37 (m, 2H), 8.11 (d, $J = 8.8$ Hz, 1H), 8.01 (d, $J = 8.7$ Hz, 1H), 7.55 (d, $J = 8.2$ Hz, 4H), 7.25 (dd, $J = 19.3, 12.1$ Hz, 7H), 6.98 (d, $J = 6.9$ Hz, 1H), 4.98 (s, 2H), 4.12 (d, $J = 5.1$ Hz, 1H), 3.30 – 3.26 (m, 2H), 1.65 (d, $J = 17.3$ Hz, 2H), 1.59 – 1.31 (m, 4H).

^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.91, 166.26, 156.85, 152.53, 149.28, 139.56, 137.70, 132.88, 129.44, 129.22, 128.81, 128.32, 128.25, 127.64, 123.77, 122.66, 119.71, 65.95, 55.96, 32.02, 29.36, 23.78. HRMS (ESI) m/z calculated for $\text{C}_{30}\text{H}_{31}\text{N}_4\text{O}_4^+$ ($\text{M}+\text{H}$) $^+$ 511.2345, found 511.2404.

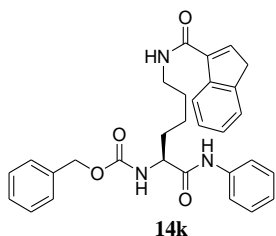
(14j): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 11.50 (s, 1H, -CONH-), 10.02 (d, $J = 6.2$ Hz, 1H,



-CONH-), 8.13 (t, $J = 7.1$ Hz, 1H, -CONH-), 8.00 – 7.95 (m, 1H, -CONH-), 7.87 (d, $J = 5.8$ Hz, 1H), 7.58 (d, $J = 5.9$ Hz, 3H), 7.43 – 7.24 (m, 8H), 7.07 (dtd, $J = 21.8, 14.5, 7.1$ Hz, 3H), 5.02 (d, $J = 6.7$ Hz, 2H), 4.18 – 4.09 (m, 1H), 3.24 (d, $J = 3.9$ Hz, 2H), 1.68 (s, 2H), 1.56 – 1.32

(m, 4H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.80, 165.20, 156.79, 139.50, 137.47, 136.52, 129.23, 128.87, 128.32, 128.24, 127.97, 126.65, 123.77, 122.27, 121.58, 120.72, 119.73, 112.19, 111.32, 65.92, 55.94, 32.15, 30.00, 23.64.

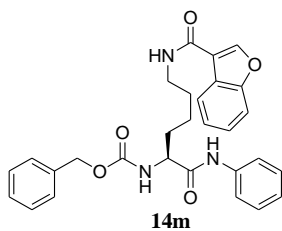
(14k): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.99 (d, $J = 8.0$ Hz, 1H, -CONH-), 8.28 (t, $J = 5.7$ Hz,



1H, -CONH-), 7.93 – 7.84 (m, 1H, -CONH-), 7.65 – 7.49 (m, 3H), 7.43 (t, $J = 7.5$ Hz, 1H), 7.41 – 7.06 (m, 10H), 7.04 – 6.95 (m, 1H), 5.13 – 4.86 (m, 2H), 4.20 – 4.02 (m, 1H), 3.66 – 3.51 (m, 1H), 3.46 (s, 2H), 3.37 – 3.27 (m, 4H), 3.27 – 2.99 (m, 3H), 1.79 – 1.56 (m, 2H), 1.56 – 1.32 (m, 4H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.82,

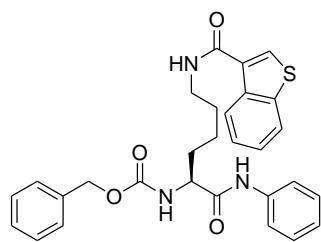
164.66, 156.90, 143.93, 142.26, 139.46, 137.54, 129.23, 128.71, 128.18, 126.49, 125.36, 124.27, 123.78, 122.32, 119.86, 65.94, 55.96, 53.76, 42.94, 31.89, 30.52, 29.15, 23.60. HRMS (ESI) m/z calculated for $\text{C}_{30}\text{H}_{33}\text{N}_3\text{O}_4\text{Na}^+$ ($\text{M}+\text{Na}$) $^+$ 520.2212, found 520.2208.

(14m): ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 9.97 (s, 1H, -CONH-), 8.50 – 8.38 (m, 1H, -CONH-),



8.34 – 8.19 (m, 1H, -CONH-), 8.02 (dd, $J = 7.2, 1.0$ Hz, 1H), 7.63 – 7.44 (m, 4H), 7.39 – 7.05 (m, 9H), 7.00 (t, $J = 7.3$ Hz, 1H), 5.08 – 4.85 (m, 2H), 4.17 – 4.00 (m, 1H), 3.22 (d, $J = 5.9$ Hz, 2H), 1.77 – 1.26 (m, 6H). ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 171.49, 162.69, 156.67,

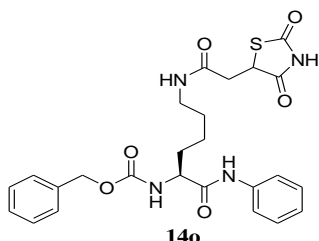
155.01, 147.87, 139.64, 137.60, 129.06, 128.20, 125.67, 123.96, 122.55, 120.26, 117.13, 112.27, 100.03, 65.84, 56.19, 29.70, 23.73. HRMS (ESI) m/z calculated for $\text{C}_{29}\text{H}_{30}\text{N}_3\text{O}_5^+$ ($\text{M}+\text{H}$) $^+$ 500.2185, found 500.2174.



14n

(14n): ^1H NMR (400 MHz, DMSO-*d*₆) δ 9.97 (s, 1H, -CONH-), 8.51 – 8.34 (m, 2H), 8.26 – 8.19 (m, 1H, -CONH-), 8.03 – 7.93 (m, 1H), 7.54 (t, *J* = 9.0 Hz, 3H), 7.43 – 7.35 (m, 2H), 7.35 – 7.17 (m, 7H), 7.00 (t, *J* = 7.4 Hz, 1H), 5.12 – 4.89 (m, 2H), 4.19 – 4.03 (m, 1H), 3.22 (ddt, *J* = 20.4, 13.5, 6.9 Hz, 2H), 1.77 – 1.27 (m, 6H). ^{13}C NMR (100 MHz, DMSO-*d*₆) δ 171.40, 163.73, 156.05, 140.14,

139.28, 137.56, 132.15, 130.73, 129.33, 128.79, 128.24, 125.34, 123.39, 119.69, 65.65, 55.96, 32.06, 29.22, 23.29. HRMS (ESI) *m/z* calculated for C₂₉H₃₀N₃O₄S⁺ (M+H)⁺ 516.1957, found 516.1941.

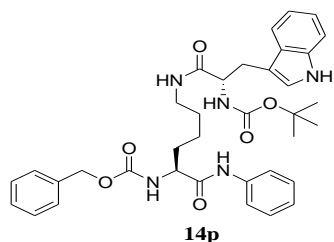


14o

(14o): ^1H NMR (400 MHz, DMSO-*d*₆) δ 11.90 (s, 1H, -CONH-), 9.95 (s, 1H, -CONH-), 8.00 (t, *J* = 4.9 Hz, 1H, -CONH-), 7.51 (dd, *J* = 29.6, 7.8 Hz, 3H), 7.35 – 7.21 (m, 7H), 7.00 (t, *J* = 7.3 Hz, 1H), 4.99 (s, 2H), 4.53 (dt, *J* = 9.2, 3.7 Hz, 1H), 4.08 (dd, *J* = 13.5, 8.1

Hz, 1H), 2.97 (t, *J* = 11.8 Hz, 2H), 2.94 (s, 1H), 2.75 – 2.66 (m, 1H), 1.70 – 1.53 (m, 2H), 1.31 (d, *J* = 38.8 Hz, 4H). ^{13}C NMR (100 MHz, DMSO-*d*₆) δ 176.53, 173.47, 171.60, 168.77, 156.74, 139.41, 137.51, 129.23, 128.87, 128.34, 128.27, 123.80, 119.80, 65.93, 55.78, 47.55, 37.54, 32.00, 29.21, 23.42. HRMS (ESI) *m/z* calculated for C₂₅H₂₉N₄O₆S⁺ (M+H)⁺ 513.1808, found 513.1883.

(14p): ^1H NMR (400 MHz, DMSO-*d*₆) δ 10.76 (d, *J* = 17.1 Hz, 1H, -CONH-), 10.00 (d, *J* = 18.7

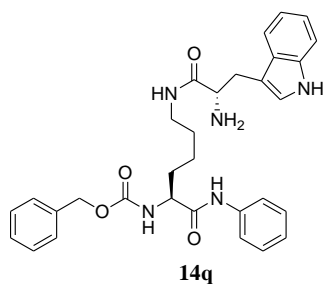


14p

Hz, 1H, -CONH-), 7.94 – 7.81 (m, 1H, -CONH-), 7.63 – 7.48 (m, 4H), 7.28 (ddd, *J* = 21.2, 14.8, 7.1 Hz, 8H), 6.99 (ddd, *J* = 28.8, 27.1, 11.8 Hz, 4H), 6.67 (dd, *J* = 18.5, 8.1 Hz, 1H), 4.98 (t, *J* = 15.7 Hz, 2H), 4.11 (s, 2H), 3.00 (d, *J* = 14.0 Hz, 3H), 2.91 – 2.79

(m, 1H), 1.62 (s, 2H), 1.46 – 1.21 (m, 13H). ^{13}C NMR (100 MHz, DMSO-*d*₆) δ 172.29, 171.69, 156.69, 155.58, 139.59, 137.63, 136.56, 129.24, 129.20, 128.86, 128.82, 128.27, 128.23, 127.85, 124.20, 123.79, 121.37, 119.69, 119.06, 118.59, 111.69, 110.71, 78.45, 65.85, 56.00, 55.67, 31.97, 29.19, 28.67, 28.62, 28.52, 28.47, 28.20, 23.34. HRMS (ESI) *m/z* calculated for C₃₆H₄₄N₅O₆⁺ (M+H)⁺ 642.3292, found 642.3389.

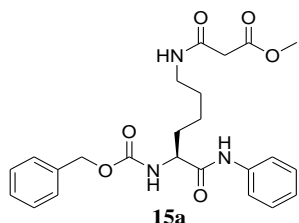
(14q): The compound **14p** (100mg, 0.155mmol) was added in anhydrous CH₂Cl₂ (4mL) and



cooled (0°C), and then the TFA (2mL) was added slowly, stirred for 2h. Then, the TFA was removed under reduced pressure until there was no TFA, the residue was purified by silica gel column chromatography (50mg, 60%). ¹H NMR(400MHz, DMSO-*d*₆) δ 10.79 (s, 1H, -CONH-), 9.98 (s, 1H, -CONH-), 7.83 (t, *J* = 5.5 Hz,

1H, -CONH-), 7.58 (d, *J* = 7.9 Hz, 2H), 7.52 (t, *J* = 6.8 Hz, 2H), 7.34 – 7.24 (m, 8H), 7.11 (s, 1H), 7.02 (t, *J* = 6.9 Hz, 2H), 6.94 (t, *J* = 7.1 Hz, 1H), 5.01 (s, 2H), 4.10 (d, *J* = 5.3 Hz, 1H), 3.40 (dd, *J* = 8.1, 4.7 Hz, 1H), 3.06 – 2.98 (m, 3H), 2.70 (dd, *J* = 14.1, 8.3 Hz, 1H), 1.62 (d, *J* = 5.4 Hz, 2H), 1.32 (dd, *J* = 18.2, 9.9 Hz, 4H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 174.89, 171.58, 156.87, 139.38, 137.76, 136.37, 128.62, 124.18, 123.63, 121.12, 119.44, 118.90, 111.95, 111.12, 65.90, 55.96, 33.22, 31.54, 29.87, 23.23, 22.67, 20.76. HRMS (ESI) *m/z* calculated for C₃₁H₃₆N₅O₄⁺ (M+H)⁺ 542.2767, found 542.2854.

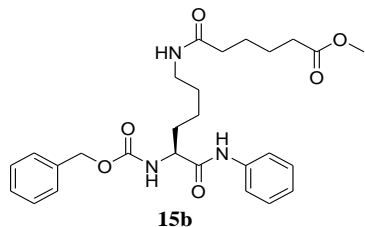
(15a): ¹H NMR (400 MHz, CD₃OD) δ 7.49 (dd, *J* = 24.8, 10.2 Hz, 2H), 7.36 – 7.14 (m, 7H), 7.07



(t, *J* = 7.3 Hz, 1H), 5.11 – 5.02 (m, 2H), 4.23 – 4.11 (m, 1H), 3.70 – 3.62 (m, 4H), 3.28 (dd, *J* = 3.1, 1.5 Hz, 1H), 3.19 (dq, *J* = 12.2, 6.3 Hz, 2H), 1.86 – 1.61 (m, 2H), 1.57 – 1.34 (m, 4H). ¹³C NMR (100 MHz, CD₃OD) δ 173.56, 172.20, 168.66, 167.30, 157.45, 138.33,

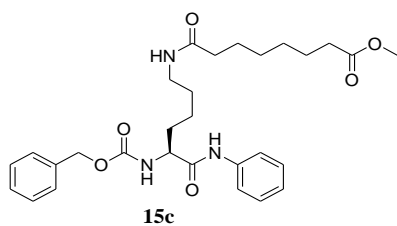
136.68, 128.47, 128.19, 127.12, 123.82, 120.27, 66.18, 55.79, 51.43, 38.87, 31.77, 30.95, 27.94, 23.06. HRMS (ESI) *m/z* calculated for C₂₄H₃₀N₃O₆Na⁺ (M+Na)⁺ 478.1954, found 478.1978.

(15b): ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.95 (s, 1H, -CONH-), 7.99 – 7.86 (m, 1H, -CONH-),



7.69 (d, *J* = 16.8 Hz, 1H, -CONH-), 7.52 (dd, *J* = 28.1, 6.3 Hz, 2H), 7.38 – 7.14 (m, 7H), 7.00 (t, *J* = 6.7 Hz, 1H), 4.97 (d, *J* = 14.1 Hz, 2H), 4.03 (dd, *J* = 24.4, 20.6 Hz, 2H), 3.62 – 3.49 (m, 3H), 2.97 (s, 2H), 2.23 (d, *J* = 3.2 Hz, 2H), 2.03 – 1.95 (m, 2H),

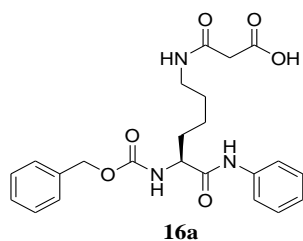
1.71 – 1.52 (m, 2H), 1.47 – 1.26 (m, 8H). HRMS (ESI) *m/z* calculated for C₂₇H₃₆N₃O₆⁺ (M+H)⁺ 498.2604, found 498.2583.



(15c): $^1\text{H NMR}$ (400 MHz, CD_3OD) δ 7.49 (t, $J = 20.2$ Hz, 2H), 7.42 – 7.13 (m, 7H), 7.05 (dd, $J = 17.3, 10.3$ Hz, 1H), 5.19 – 4.94 (m, 2H), 4.30 – 4.08 (m, 1H), 3.61 (s, 3H), 3.13 (t, $J = 6.0$ Hz, 2H), 2.26 (t, $J = 7.3$ Hz, 2H), 2.18 – 2.03 (m, 2H), 1.92 – 1.60 (m, 2H), 1.60 (s, 8H), 1.28 (d, $J = 19.5$ Hz, 4H).

$^{13}\text{C NMR}$ (100 MHz, CD_3OD) δ 174.67, 172.11, 157.10, 138.00, 136.98, 128.68, 128.19, 127.71, 123.90, 119.68, 66.39, 55.63, 50.86, 38.27, 35.51, 33.50, 28.73, 25.74, 24.51, 22.98. HRMS (ESI) m/z calculated for $\text{C}_{29}\text{H}_{39}\text{N}_3\text{O}_6\text{Na}^+$ ($\text{M}+\text{Na}$) $^+$ 548.2737, found 548.2731.

(16a): The compound **15a** (150mg, 0.32mmol) was added in $\text{THF}:\text{H}_2\text{O}=3:1$ (4mL) and cooled

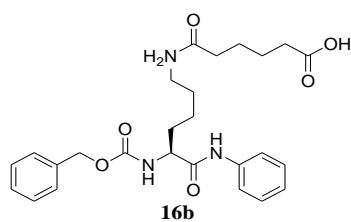


(0°C), and then the LiOH ($\text{THF}:\text{H}_2\text{O}=3:1$, 0.5mL) was added slowly, stirred for 30min. Then, added in 10% Citric acid solution, the reaction mixture was diluted with CH_2Cl_2 and saturated NaCl solution, the organic layer was washed with saturated NaCl , and

dried over Na_2SO_4 and concentrated. The residue was purified by silica gel column chromatography (110mg, 78%). $^1\text{H NMR}$ (400 MHz, CD_3OD) δ 7.50 (t, $J = 11.5$ Hz, 2H), 7.39 – 7.12 (m, 7H), 7.07 (t, $J = 6.9$ Hz, 1H), 5.14 – 5.01 (m, 2H), 4.16 (ddd, $J = 30.7, 8.3, 4.5$ Hz, 1H), 3.34 – 3.27 (m, 1H), 3.19 (d, $J = 6.8$ Hz, 3H), 1.87 – 1.63 (m, 2H), 1.60 – 1.36 (m, 4H). $^{13}\text{C NMR}$ (100 MHz, CD_3OD) δ 171.98, 170.28, 167.75, 162.43, 157.41, 138.14, 136.68, 128.58, 128.28, 127.50, 124.09, 120.16, 66.45, 55.52, 38.77, 31.46, 28.14, 23.07. HRMS (ESI) m/z calculated for $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}_6^+$ ($\text{M}+\text{H}$) $^+$ 442.1978, found 442.1961.

The synthesis of compounds **16b-c** were similar to the synthesis of compound **16a**. The yield of these compounds was about 70%~85%.

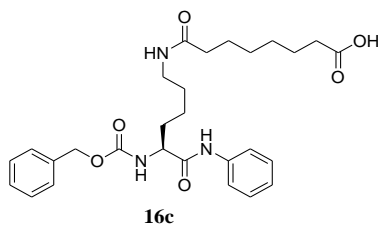
(16b): $^1\text{H NMR}$ (400 MHz, CD_3OD) δ 7.62 – 7.41 (m, 2H), 7.39 – 7.12 (m, 7H), 7.06 (dd, $J =$



16.2, 8.8 Hz, 1H), 5.19 – 4.96 (m, 2H), 4.25 – 4.06 (m, 1H), 3.21 – 3.08 (m, 2H), 2.33 – 2.18 (m, 2H), 2.18 (s, 2H), 1.88 – 1.63 (m, 2H), 1.63 – 1.33 (m, 8H). $^{13}\text{C NMR}$ (100 MHz, CD_3OD) δ 175.73, 174.31, 138.13, 137.02, 128.66, 124.39,

120.36, 66.47, 55.57, 38.36, 35.45, 33.46, 31.49, 29.19, 25.41, 23.99, 22.87. HRMS (ESI) m/z calculated for $C_{26}H_{34}N_3O_6^+$ ($M+H$)⁺ 484.2448, found 484.2426.

(16c): ¹H NMR (400 MHz, CD₃OD) δ 7.62 – 7.41 (m, 2H), 7.40 – 7.12 (m, 7H), 7.11 – 7.01 (m,

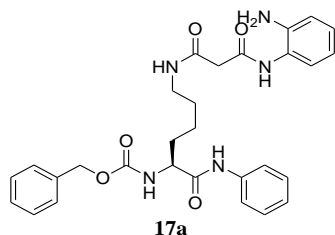


1H), 5.18 – 4.99 (m, 2H), 4.24 – 4.08 (m, 1H), 3.18 – 3.10 (m, 2H), 2.27 – 2.20 (m, 2H), 2.11 (t, $J = 7.5$ Hz, 2H), 1.87 – 1.65 (m, 2H), 1.60 – 1.36 (m, 8H), 1.34 – 1.18 (m, 6H).

¹³C NMR (100 MHz, CD₃OD) δ 176.06, 174.91, 172.07,

157.09, 138.36, 136.93, 128.53, 127.66, 123.94, 120.19, 66.65, 55.95, 38.72, 37.85, 35.54, 33.81, 28.58, 25.16, 23.98, 22.85. HRMS (ESI) m/z calculated for $C_{28}H_{38}N_3O_6^+$ ($M+H$)⁺ 512.2761, found 512.2750.

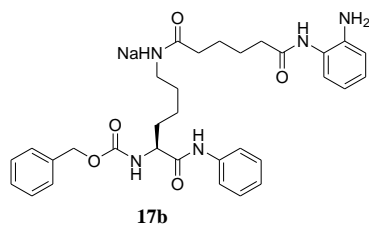
(17a): ¹H NMR (400 MHz, CD₃OD) δ 7.65 – 7.41 (m, 3H), 7.40 – 7.15 (m, 7H), 7.12 – 6.96 (m,



2H), 6.81 (dt, $J = 8.0, 1.8$ Hz, 1H), 6.67 (td, $J = 7.6, 1.3$ Hz, 1H), 5.15 – 4.99 (m, 2H), 4.18 (dd, $J = 19.3, 13.9$ Hz, 1H), 3.28 (dt, $J = 3.2, 1.6$ Hz, 2H), 3.24 – 3.11 (m, 2H), 1.91 – 1.63 (m, 2H), 1.61 – 1.36 (m, 4H). ¹³C NMR (100MHz, CD₃OD) δ 176.03,

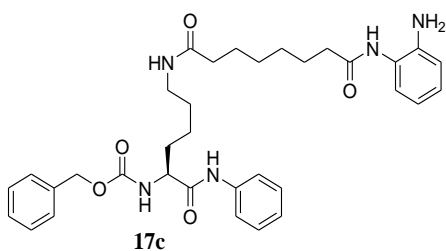
168.23, 138.26, 127.93, 127.07, 125.93, 124.29, 123.15, 120.07, 118.12, 116.98, 66.63, 65.17, 33.87, 31.32, 30.48, 29.64, 28.79, 28.55, 24.92, 22.67, 22.08. HRMS (ESI) m/z calculated for $C_{29}H_{34}N_5O_5^+$ ($M+H$)⁺ 532.2560, found 532.2545.

(17b): ¹H NMR (400 MHz, CD₃OD) δ 7.58 – 7.42 (m, 2H), 7.40 – 7.11 (m, 7H), 7.11 – 7.01 (m,



2H), 7.01 (s, 1H), 6.80 (dt, $J = 10.8, 5.4$ Hz, 1H), 6.68 (ddt, $J = 19.1, 13.0, 6.5$ Hz, 1H), 5.17 – 4.96 (m, 2H), 4.19 (dt, $J = 23.7, 11.8$ Hz, 1H), 3.22 – 3.04 (m, 2H), 2.48 – 2.32 (m, 2H), 2.25 – 2.10 (m, 2H), 1.89 – 1.60 (m, 6H), 1.46 (dddd, $J = 10.6,$

10.1, 8.4, 5.3 Hz, 2H). ¹³C NMR (100 MHz, CD₃OD) δ 174.64, 173.25, 172.11, 157.00, 141.57, 137.96, 136.85, 128.46, 127.07, 126.76, 125.61, 123.98, 119.75, 118.37, 117.22, 66.32, 55.69, 38.33, 35.26, 34.10, 31.32, 30.77, 29.32, 28.54, 25.16, 23.22, 22.62. HRMS (ESI) m/z calculated for $C_{32}H_{40}N_5O_5Na^+$ ($M+Na$)⁺ 596.2849, found 596.2839.

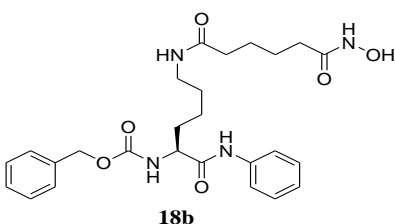


(17c): ^1H NMR (400 MHz, CD_3OD) δ 7.61 – 7.41 (m, 2H), 7.42 – 7.11 (m, 7H), 7.10 – 6.94 (m, 3H), 6.80 (dt, J = 10.8, 5.4 Hz, 1H), 6.71 – 6.64 (m, 1H), 5.07 (q, J = 12.6 Hz, 2H), 4.19 (dd, J = 8.4, 5.5 Hz, 1H), 3.20 – 3.04 (m, 2H), 2.39 (dt, J = 14.9, 7.5 Hz, 2H), 2.10 (dt, J = 21.1, 7.2 Hz, 2H), 1.88 – 1.44 (m, 8H), 1.43 – 1.29 (m,

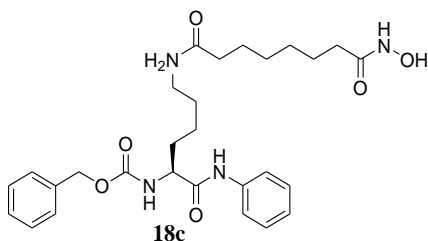
6H). ^{13}C NMR (100 MHz, CD_3OD) δ 175.12, 173.96, 142.07, 137.86, 127.70, 127.13, 125.67, 124.02, 120.06, 118.04, 117.19, 66.68, 38.45, 35.35, 31.63, 29.14, 28.28, 25.72, 22.92. HRMS (ESI) m/z calculated for $\text{C}_{34}\text{H}_{43}\text{N}_5\text{O}_5\text{Na}^+$ ($\text{M}+\text{Na}$) $^+$ 624.3162, found 624.3138.

The synthesis of compounds **18b-c** were similar to the synthesis of compound **11**. The yield of these compounds was about 42%.

(18b): ^1H NMR (400 MHz, CD_3OD) δ 7.58 – 7.44 (m, 2H), 7.39 – 7.12 (m, 7H), 7.06 (dd, J = 16.3, 8.9 Hz, 1H), 5.19 – 4.97 (m, 2H), 4.27 – 4.10 (m, 1H), 3.16 (p, J = 7.4 Hz, 2H), 2.20 – 1.99 (m, 4H), 1.88 – 1.63 (m, 2H), 1.63 – 1.35 (m, 8H). ^{13}C NMR (100MHz, CD_3OD) δ 174.48, 172.15, 171.28, 157.30, 138.04, 136.92, 128.17,



124.39, 120.30, 66.39, 55.63, 38.45, 35.25, 32.33, 28.83, 24.73, 23.28. HRMS (ESI) m/z calculated for $\text{C}_{26}\text{H}_{35}\text{N}_4\text{O}_6^+$ ($\text{M}+\text{H}$) $^+$ 499.2557, found 499.2554.



(18c): ^1H NMR (400 MHz, CD_3OD) δ 7.62 – 7.43 (m, 2H), 7.37 – 7.13 (m, 7H), 7.12 – 7.01 (m, 1H), 5.15 – 4.96 (m, 2H), 4.27 – 4.10 (m, 1H), 3.21 – 3.08 (m, 6H), 2.18 – 2.00 (m, 4H), 1.87 – 1.63 (m, 2H), 1.61 – 1.34 (m, 8H). ^{13}C NMR (100 MHz, CD_3OD) δ 174.59, 172.08, 157.11,

138.26, 136.85, 127.84, 123.89, 119.63, 66.38, 55.43, 38.75, 35.41, 32.33, 31.75, 30.36, 29.21, 28.10, 25.24, 23.00. HRMS (ESI) m/z calculated for $\text{C}_{28}\text{H}_{40}\text{N}_4\text{O}_6^+$ ($\text{M}+\text{H}$) $^+$ 527.2870, found 527.2854.

HDACS inhibition assay in vitro

In vitro HDACs inhibition assays were conducted as previously described^[1]. In brief, 4 μL of HeLa nuclear extract (9mg/mL) was mixed with various concentrations of tested compounds (1 μL), then fluorogenic substrate Boc-Lys(acetyl)-AMC (100 μM) and Buffer A (50 mM Tris-HCl pH=8.0, 150 mM NaCl, 30 mM KCl, 10 mM MgCl_2) were added. After incubation at 37 $^\circ\text{C}$ for 1h, the mixture was stopped by the addition of 50 μL of developer containing trypsin and 1 μM SAHA. And then incubation at 37 $^\circ\text{C}$ for 1h, the mixture was stopped by the addition of 50 μL of Buffer B (200mM HCl/300 mM Acetic acid), fluorescence intensity was measured using a microplate reader at excitation and emission wavelengths of 360 nm and 460 nm. The inhibition ratios were calculated from the fluorescence intensity readings of tested wells relative to those of control wells, and the IC₅₀ values (Concentrations of tested compounds **SAHA**: 500 nM, 100 nM, 50 nM, 25 nM, 12.5 nM, 6.25 nM; **11**: 100 μM , 75 μM , 50 μM , 25 μM , 12.5 μM ; **18b**: 50 μM , 10 μM , 1 μM , 500 nM, 100 nM, 50nM; **18c**: 50 μM , 10 μM , 1 μM , 500 nM, 100 nM, 50nM) were calculated using a regression analysis of the concentration/inhibition data. All experiments were done in triplicate.

Table S1. Selective investigation of N ϵ -acetyl lysine derivatives as HDAC inhibitors.

Inhibitor	HDAC Inhibition (%)		
	Hela nuclear extract		
	HDACI	HDACIIa	HDAC8
SAHA	87.05 \pm 2.193	15.51 \pm 3.084	41.19 \pm 0.180
18c	61.06 \pm 1.064	16.65 \pm 2.690	38.27 \pm 1.215

*1 μM

Cell Proliferation Assay

The rate of cell survival under the action of test substances was evaluated by an improved MTS assay as previously described^[2]. In brief, A549 (5000 cells / well) , HepG2 (5000 cells / well), HEK293 (5000 cells / well) and K562 (1 \times 10⁴ cells/well) cell lines were seeded into 96-well plates in 100 μL medium. After overnight incubation, each well was added with different concentrations of the compounds **SAHA**, **16c** and **18c**, then incubated for another 48 hours. After 48 hours 5ul of MTS reagent solution was added into each well and the cells were incubated for another 4 hours at 37 $^\circ\text{C}$ under 5% CO₂ environment. The absorbance was measured at 490 nm in

a Microplate to determine the cell viability. The absorbance was directly proportional to the number of viable cells. All experiments were done in triplicate.

Table S2. The IC₅₀s of **18c** and **SAHA** against both tumor cells and normal cells, then calculate and compare their selective index.

	K562	A549	HepG2	HEK293	selectivity index(SI)
SAHA	4.58±0.17	17.81±1.25	4.23±0.63	6.09±0.84	0.34~1.43
18c	41.18±1.73	134.10±2.13	158.40±2.20	>500	3.16~12.14
16c	>500	>500	>500	>500	/

Western Blot Methods

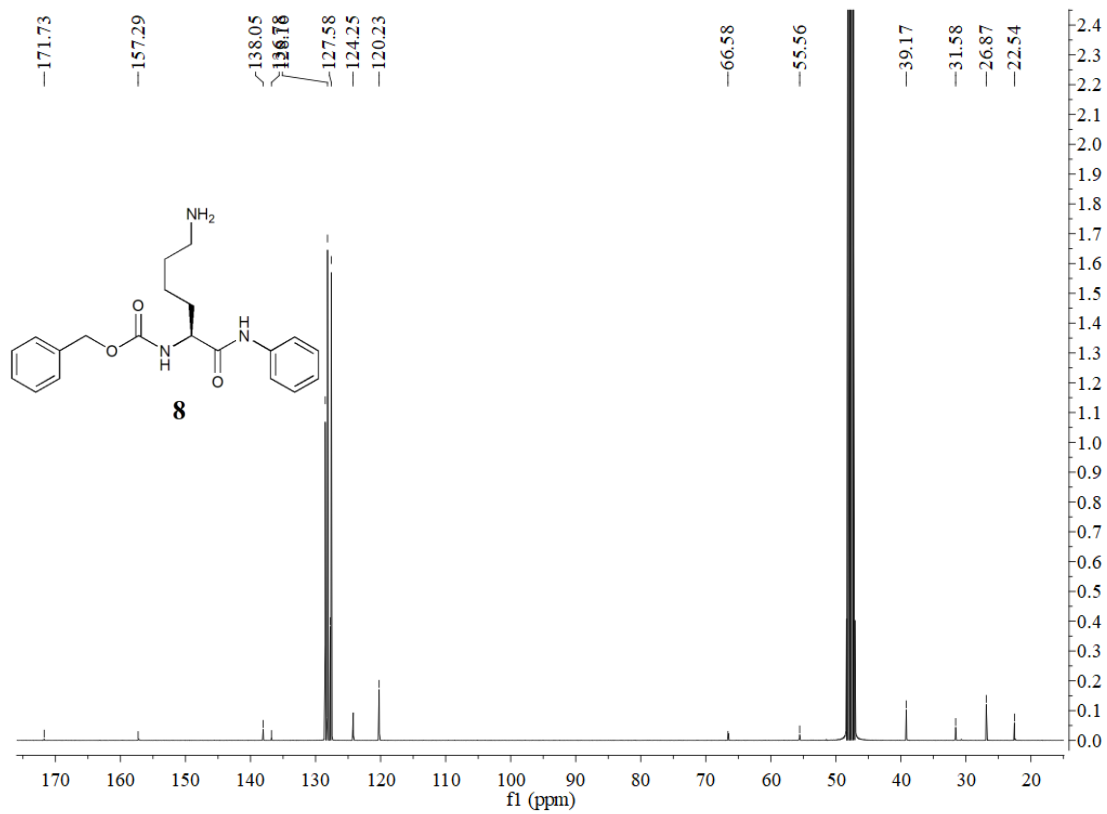
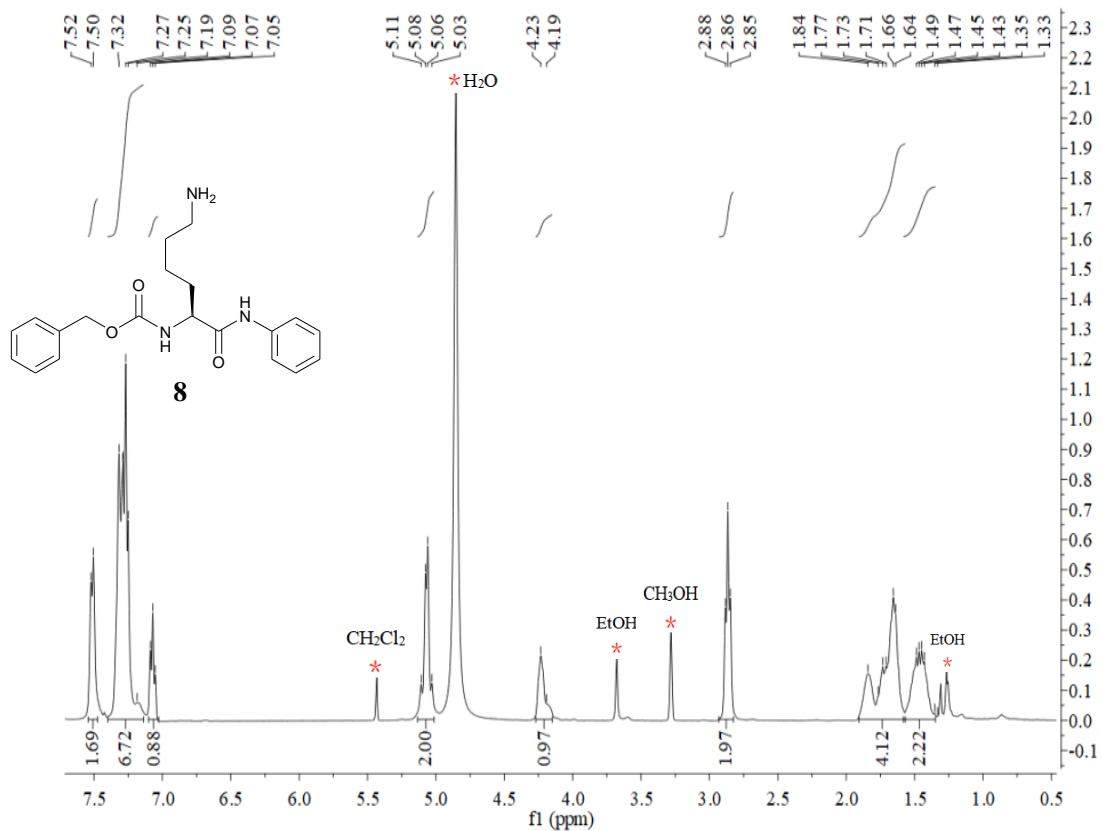
Respectively, A549 and K562 cells were harvested with three different concentrations of compound **18c** (50 μM, 100 μM, 200 μM) and concentrations of **SAHA** (10 μM) under 8 mL DMEM with 10% FBS (fetal bovine serum) for 24 hours. DMSO was used as the control. Later the cell pellets were collected by centrifuge at 1000 rpm for 5 minutes and then washed by ice-cold PBS (phosphate-buffered saline) twice. The PBS solution was drained and the cell pellets were re-suspended with ice-cold cell lysate buffer (50 mM Tris-HCl PH=7.4, 150 mM NaCl, 1 mM EDTA, 10% glycerin, 1% Triton 100 and protease inhibitors cocktail) and Vortex blending five times for 30 seconds each over a period of 30 minutes kept on ice. The samples were later centrifuged at 12,000×g for 10 minutes at 4 °C and the supernatants in each tube were collected and placed in a fresh tube kept on ice. BCA (bicinchoninic acid) assay was used to determine the total protein amount. SDS polyacrylamide gel electrophoresis (PAGE) was added into each sample and the mixture was boiled at 100°C for 10 minutes for denaturation. Around 60 μg protein was loaded into each well and the gels will be submerged in migration buffer which normally contains 25 mM Tris base, 190 mM glycine and 0.1% SDS. Run the gel for 20 minutes under 80 V/30 minutes under 120 V/25 minutes under 200 V. The proteins were then immobilized on a nitrocellulose membrane following electrophoretic transfer from the gel at 60 minutes under 100 V/350mA at 4 °C via wet transfer process. Non-protein binding areas on the membrane were blocked to prevent non-specific binding of antibodies by 5% BSA in TBST (10 mM Tris, PH=7.4, 150 mM NaCl and 0.5% Tween 20) at room temperature for 2 hour and the membranes were incubated with primary antibodies respectively (acetyl-α-Tubulin, sc-23950, Santa; anti-α-Tubulin, cat:M1000130, Solarbio) that specifically bound to the protein of interest. Unbound antibodies

were removed by washing and a secondary antibody conjugated to an enzyme, a fluorophore was used for detection. The detected signal from the protein: antibody: antibody complex was proportional to the amount of protein on the membrane. Later the membranes were striped and re-probed with loading control primary antibodies respectively (anti- α -Tubulin, acetyl- α -Tubulin) and repeated the previous steps to detect the signal.

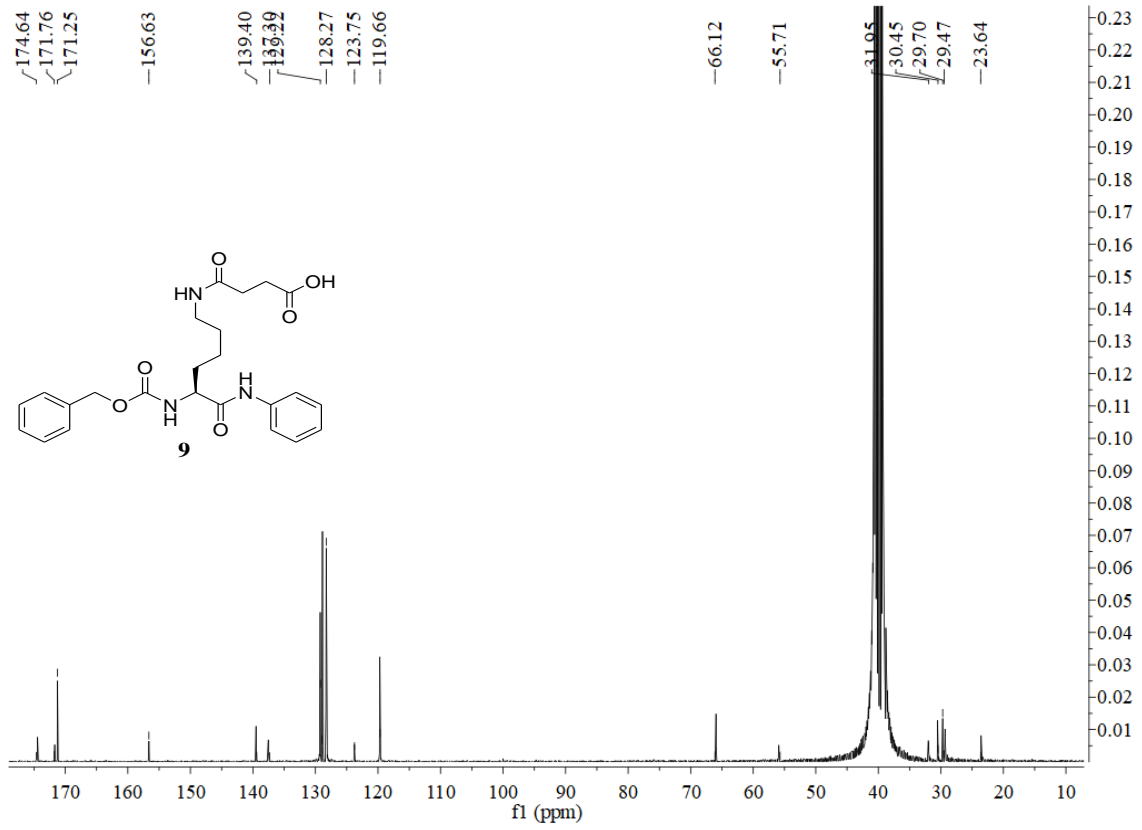
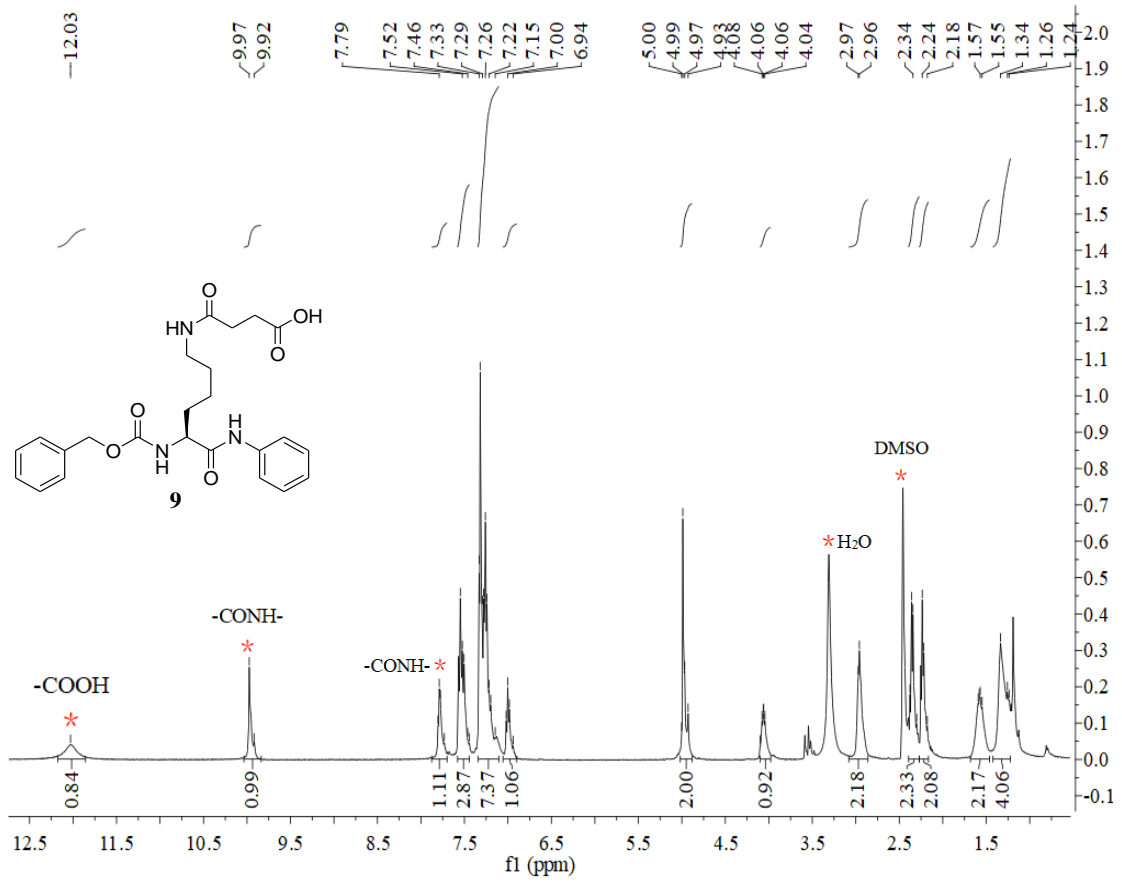
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1. Xiaoyang Li, Yuri K. Peterson, Elizabeth S. Inks, Richard A. Himes, Jiaying Li, Yingjie Zhang, Xiujie Kong, and C. James Chou, *Journal of Medicinal Chemistry*, 2018, **61**, 2589-2603.
2. Jinyu Yang, Gaoliang Cheng, Qihao Xu, Shenglin Luan, Shuxiang Wang, Dan Liu, Linxiang Zhao, *Bioorganic & Medicinal Chemistry*, 2018, **26**, 1418-1425.
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4. Xiaoyang Li, Elizabeth S. Inks, Xiaoguang Li, Jinning Hou, C. James Chou, Jian Zhang, Yuqi Jiang, Yingjie Zhang, and Wenfang Xu, *Journal of Medicinal Chemistry*, 2014, **57**, 3324-3341.

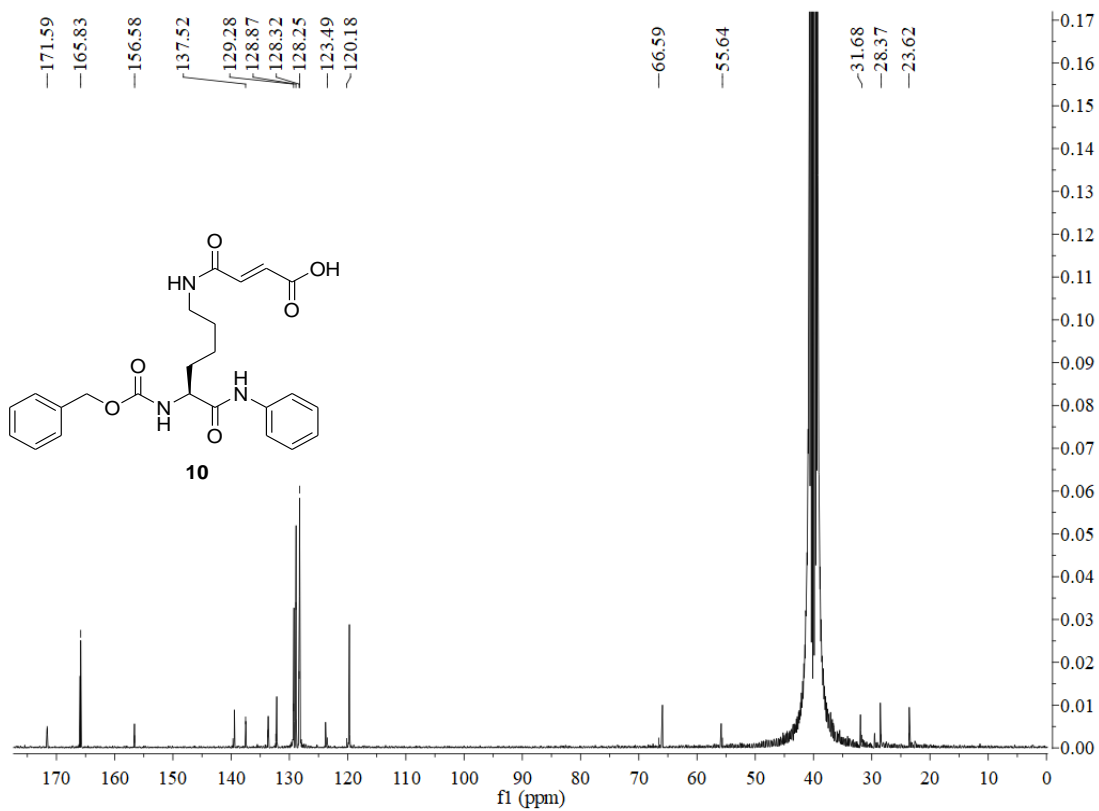
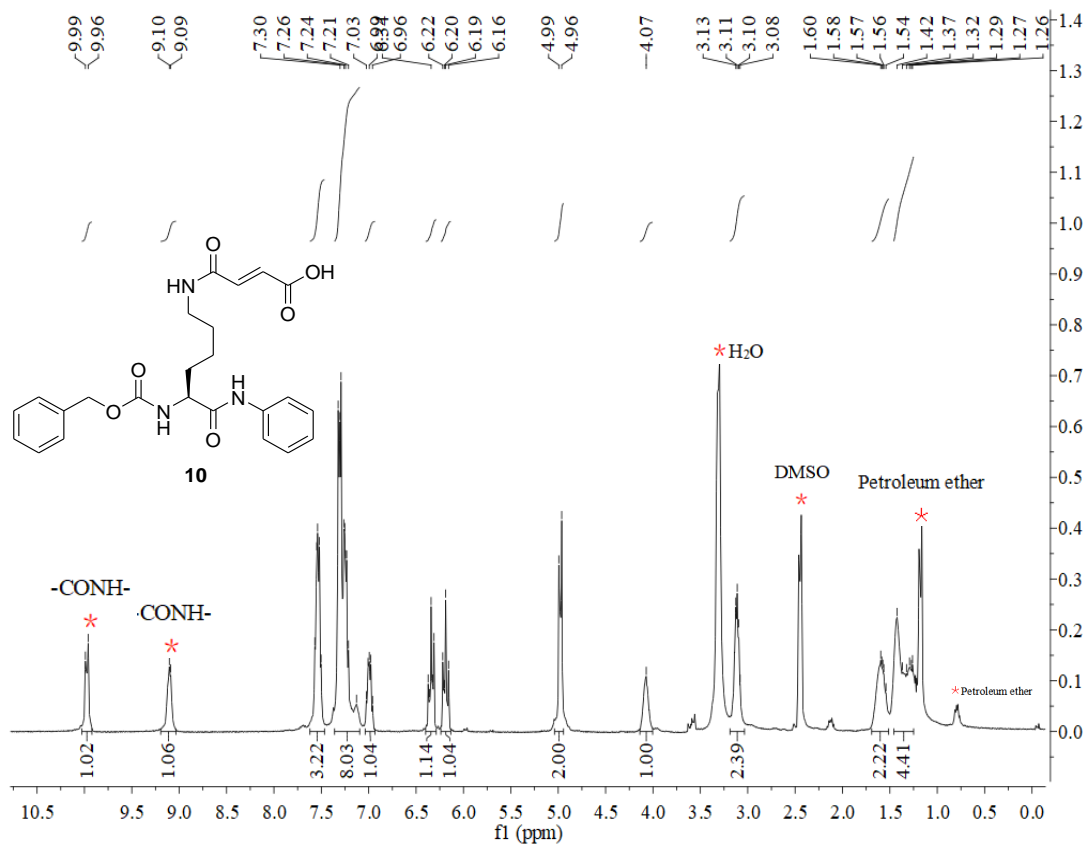
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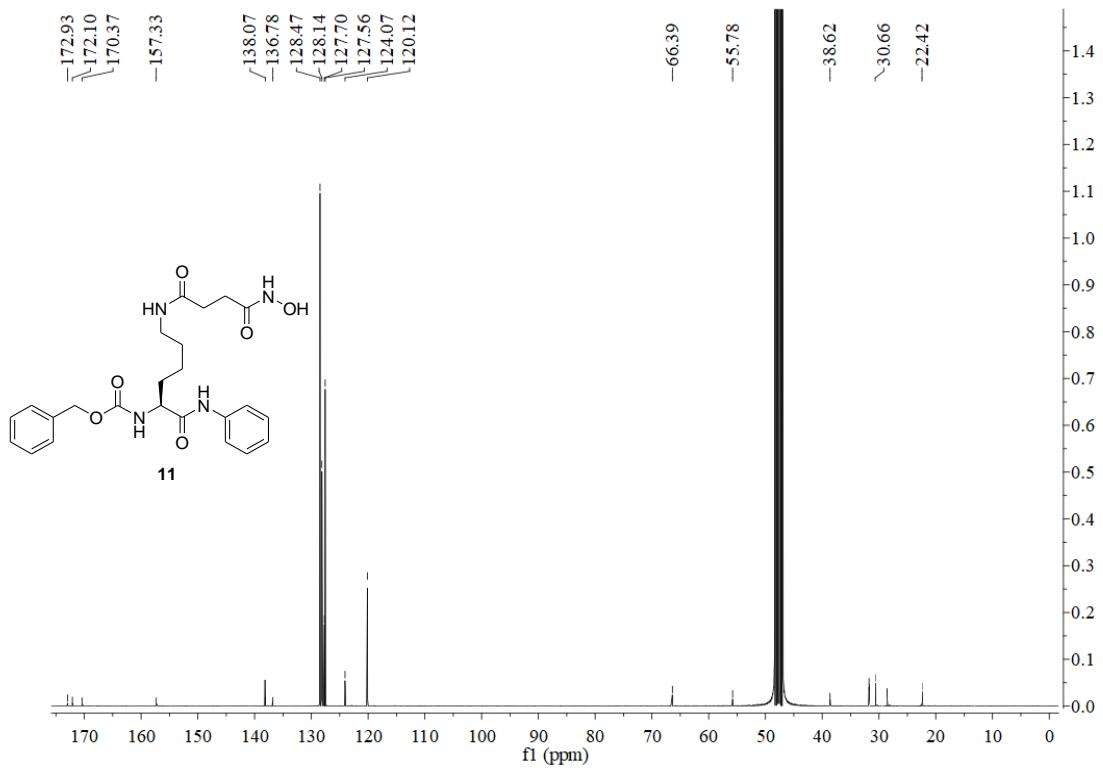
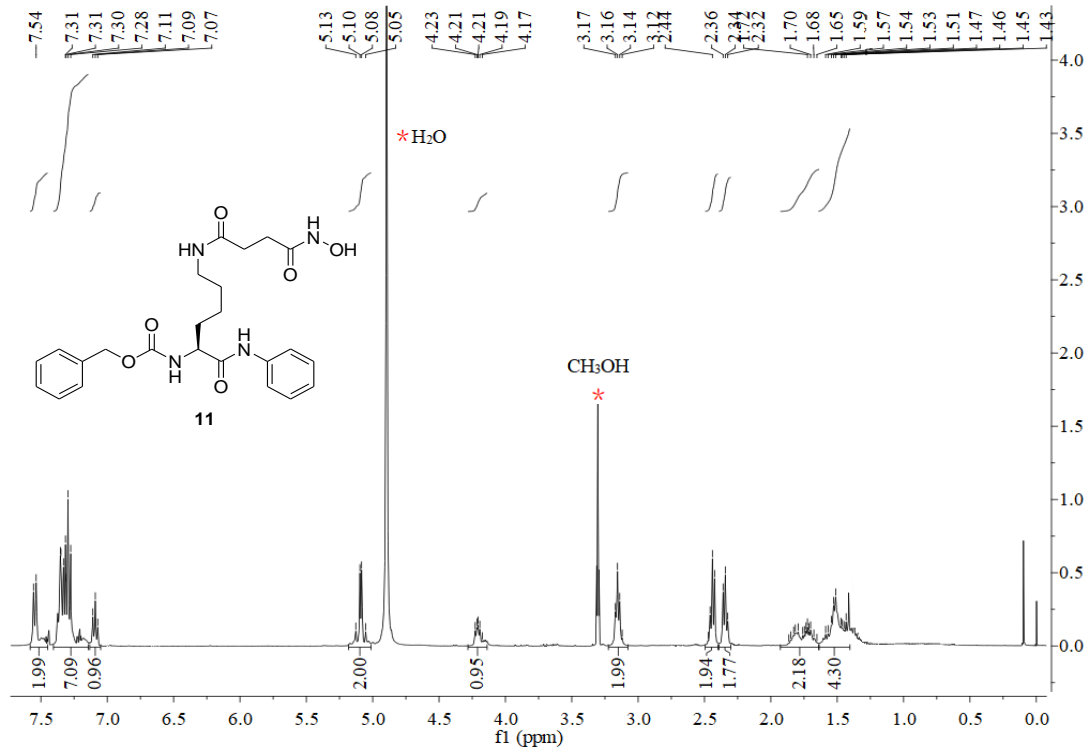
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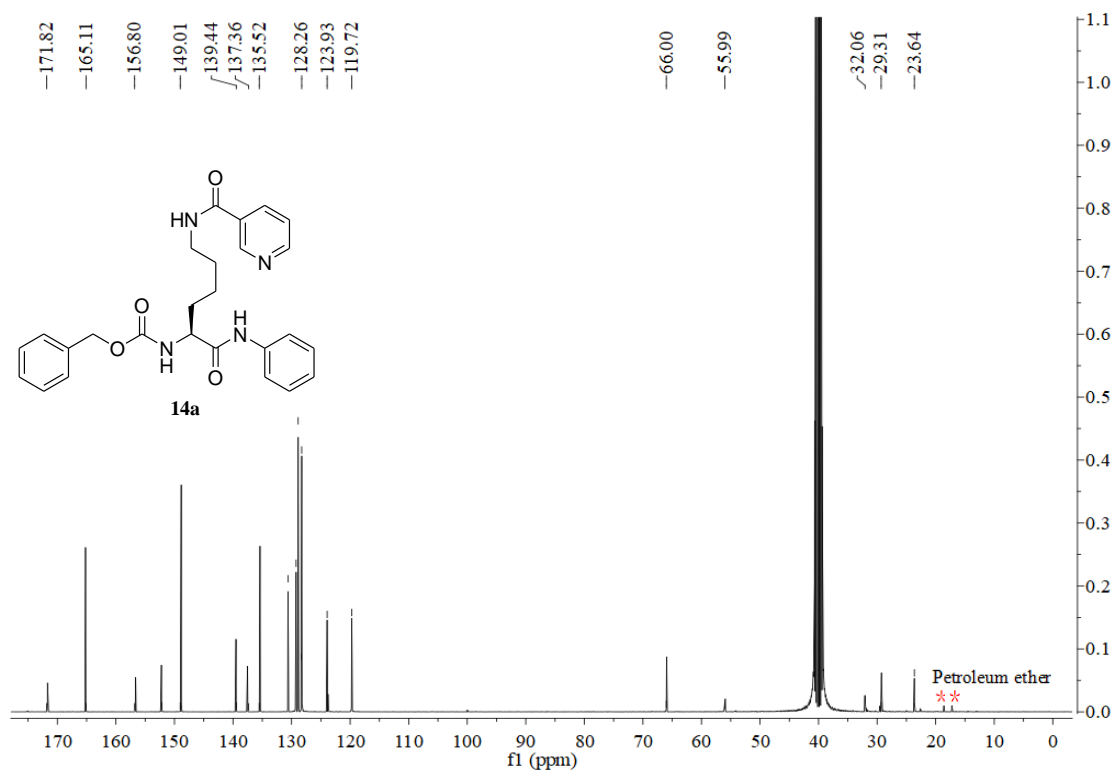
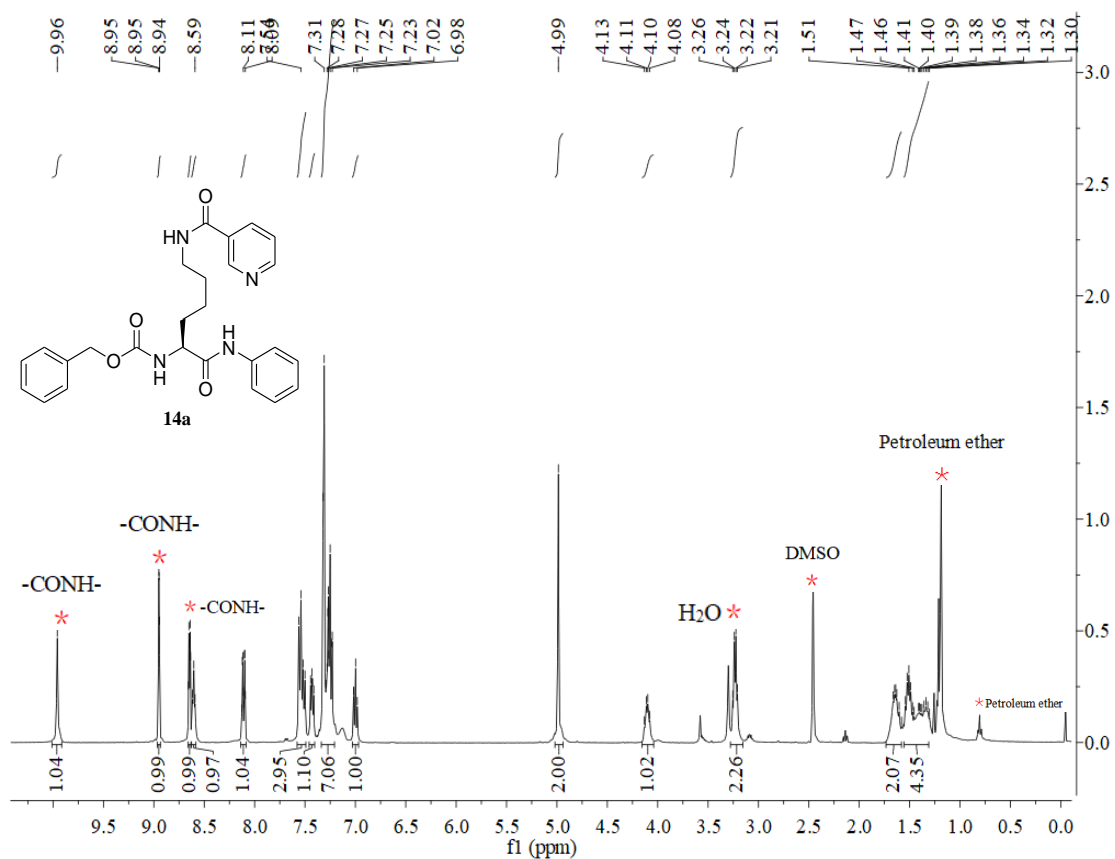
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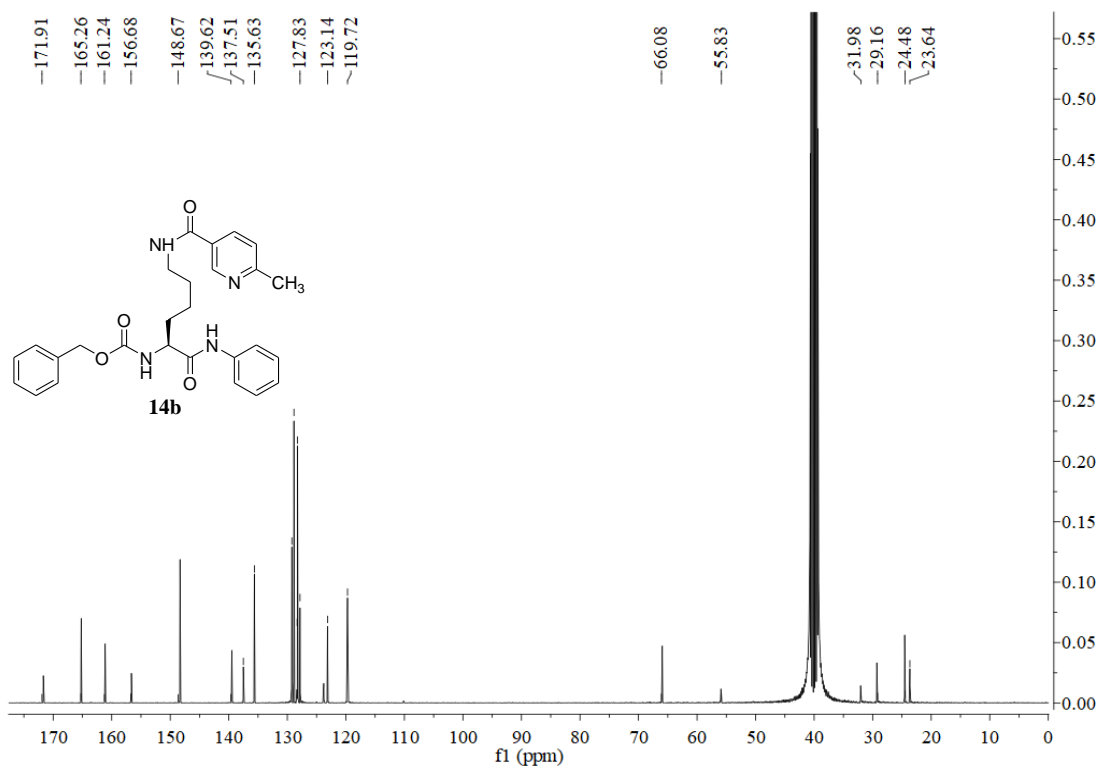
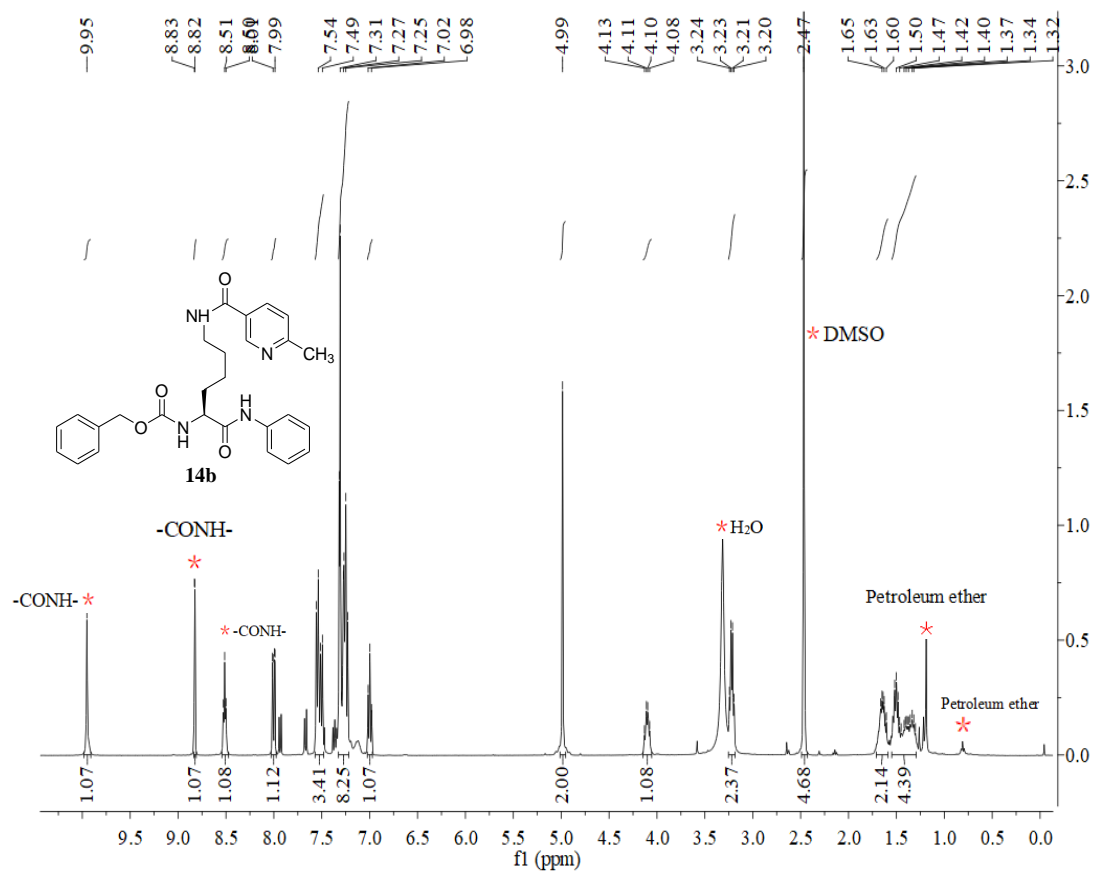
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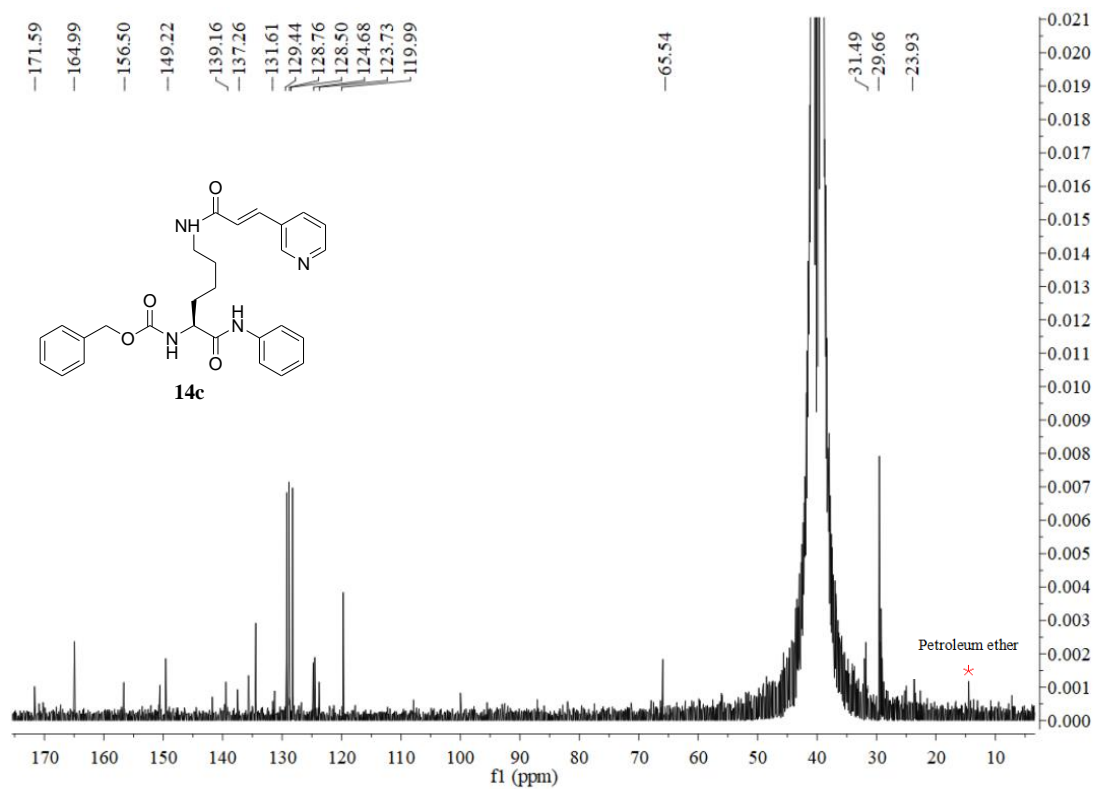
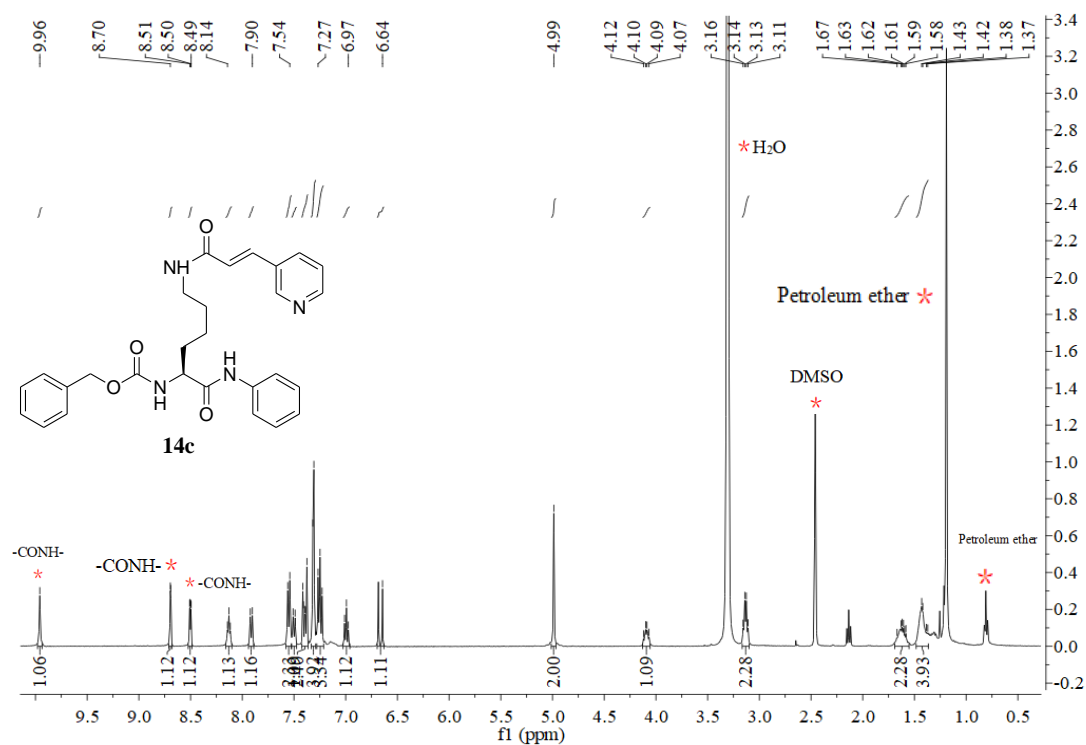
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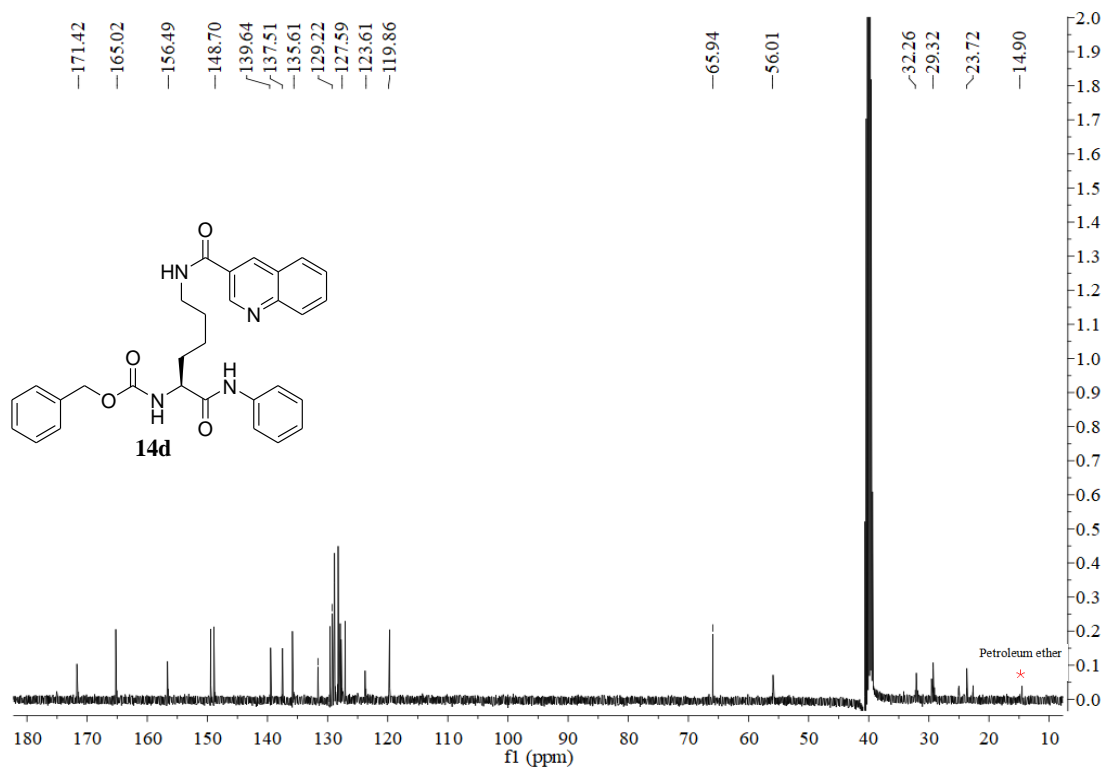
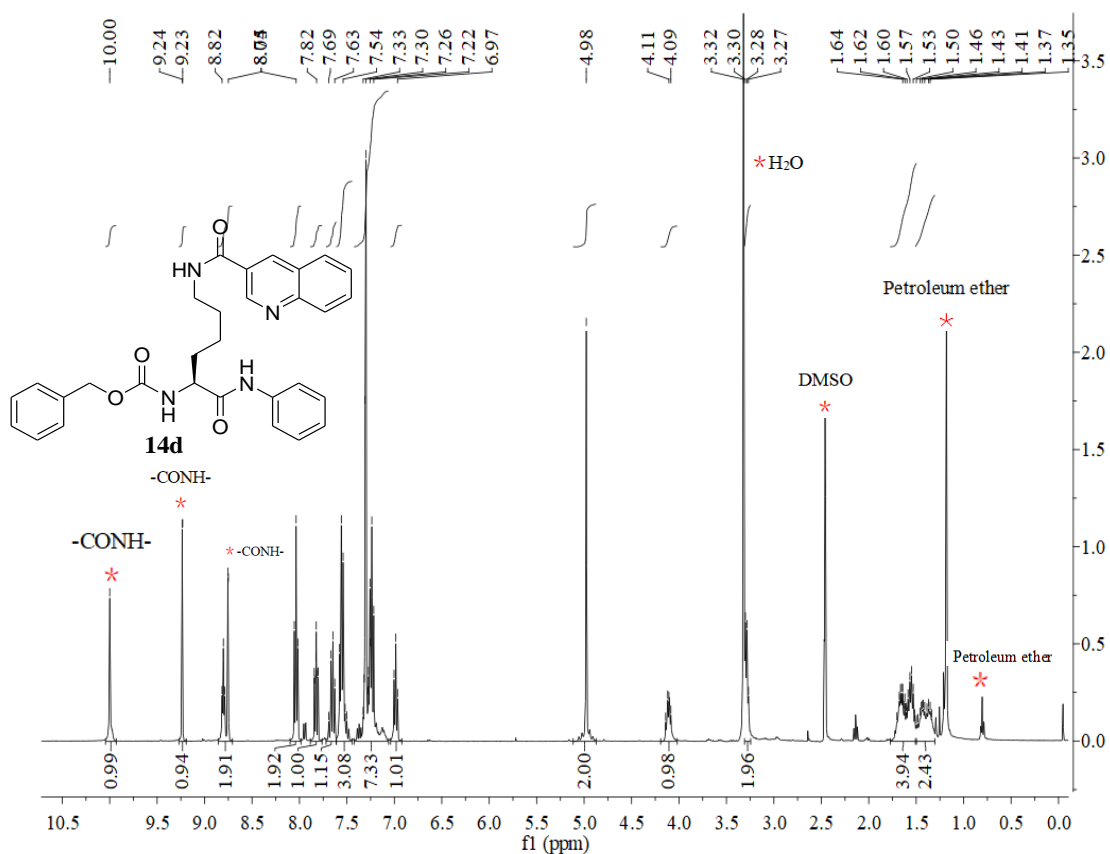
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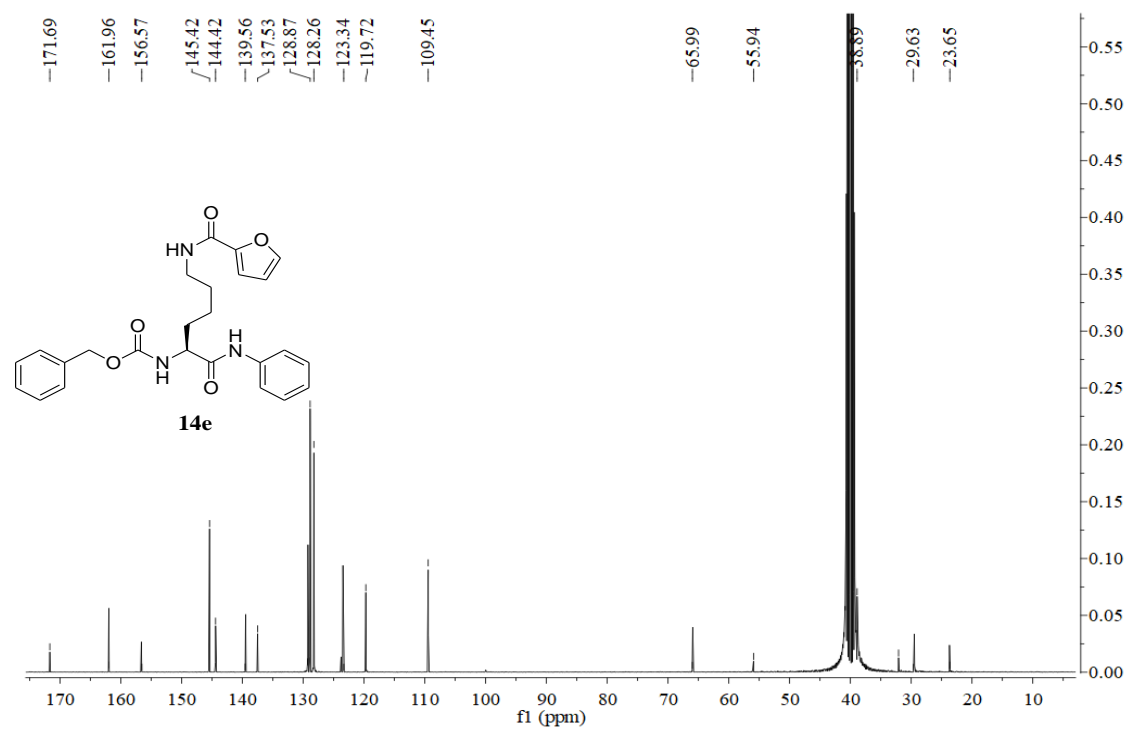
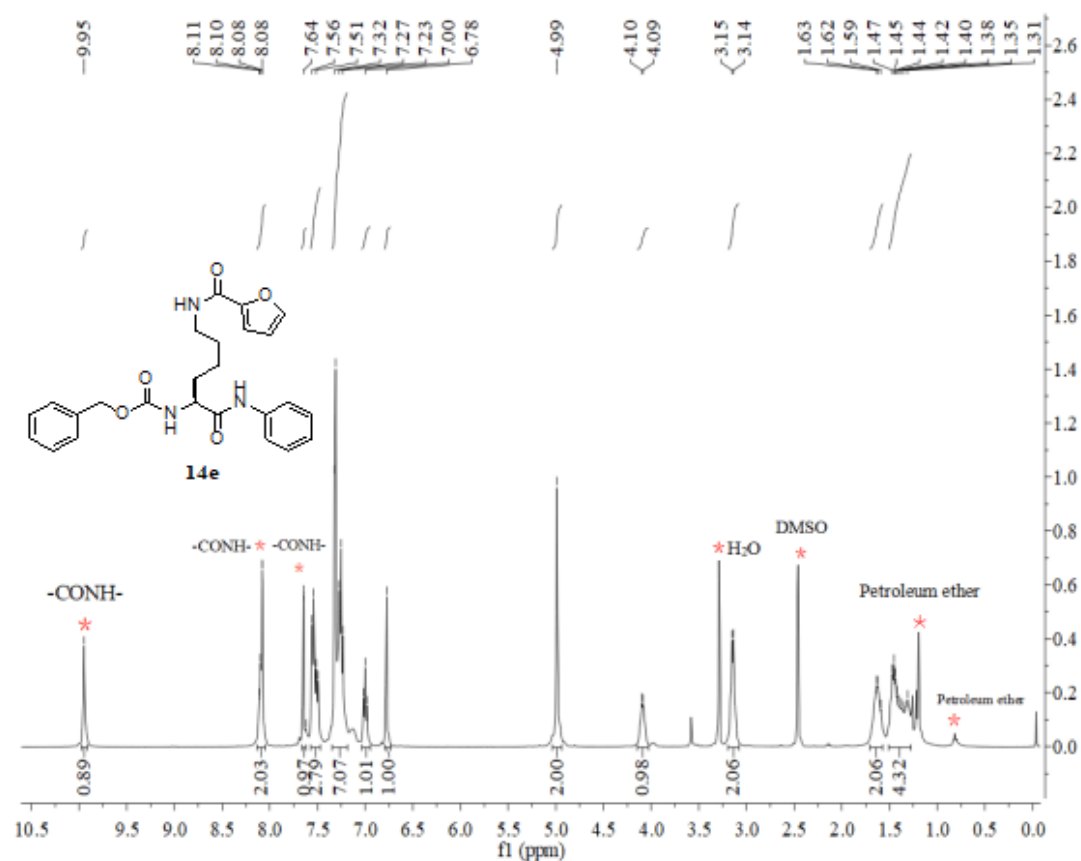
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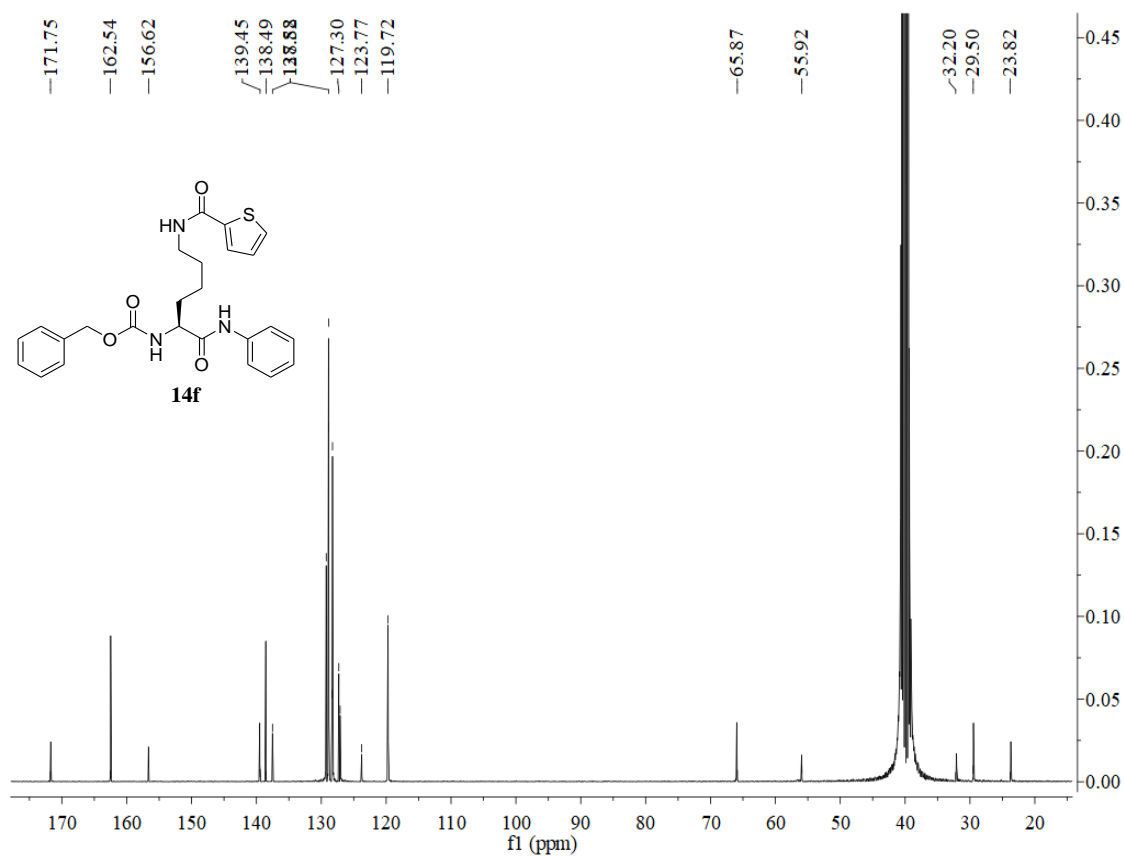
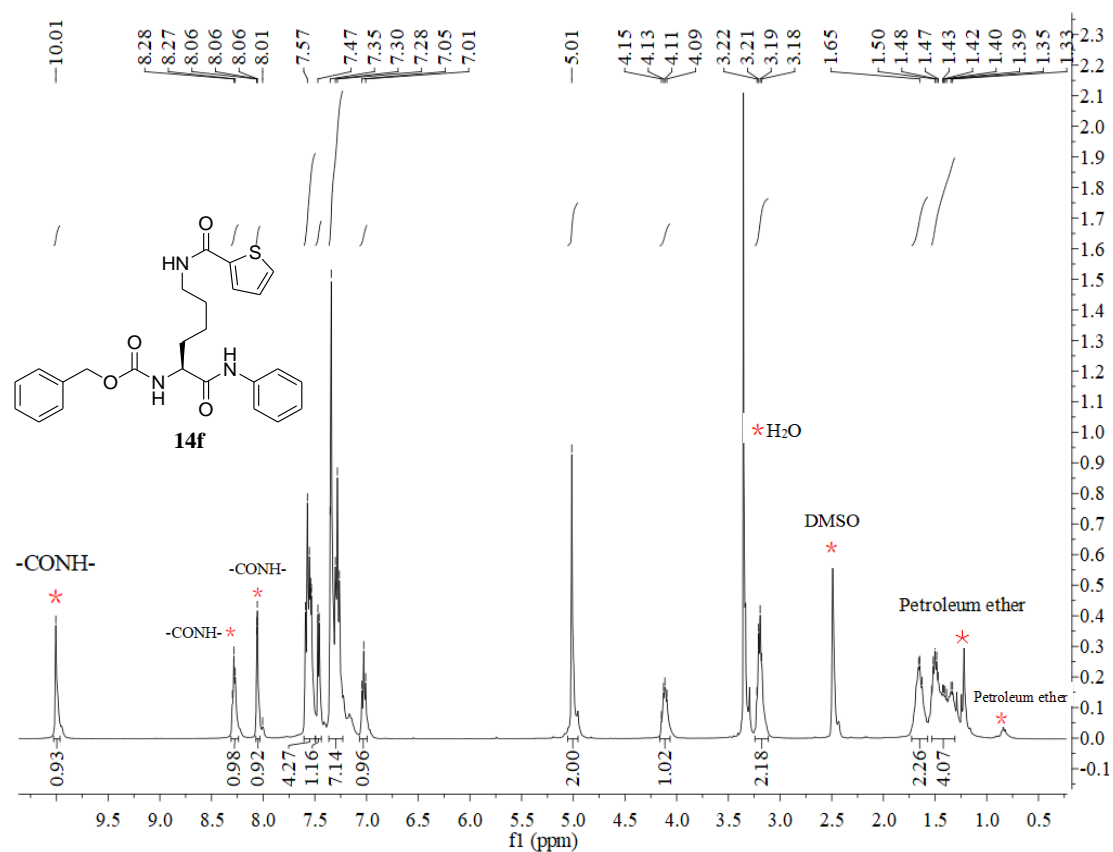
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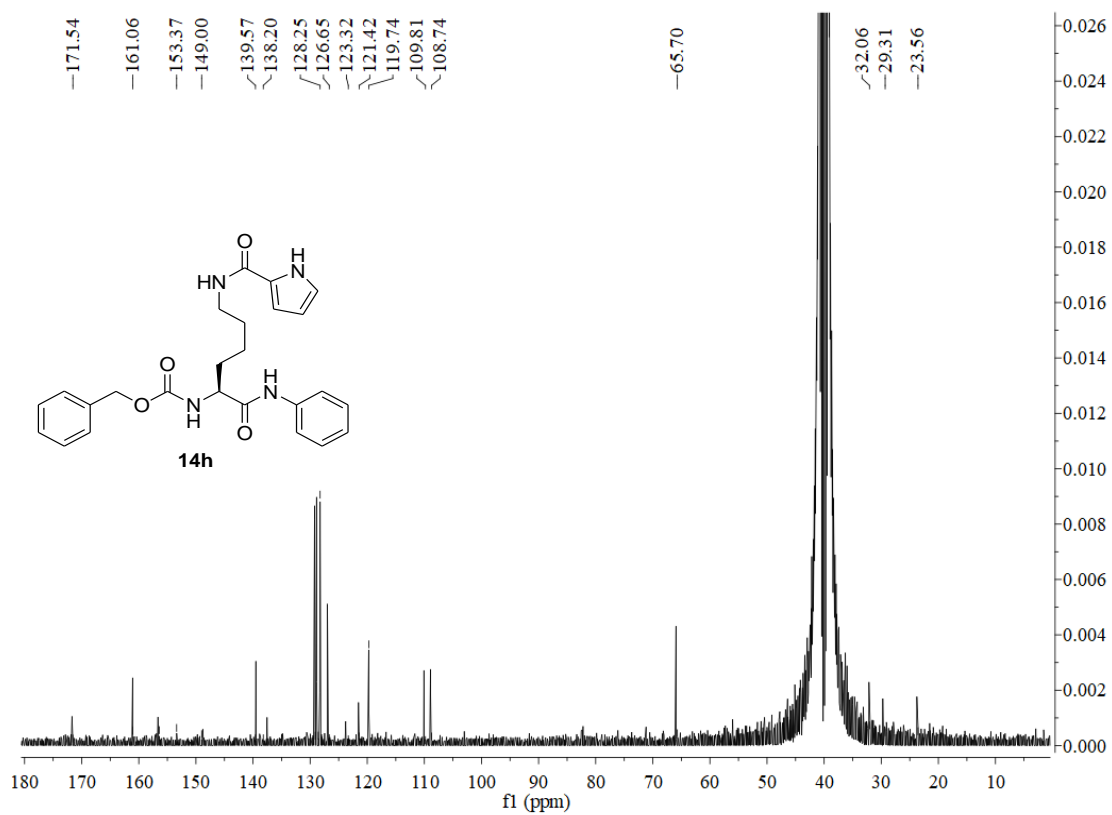
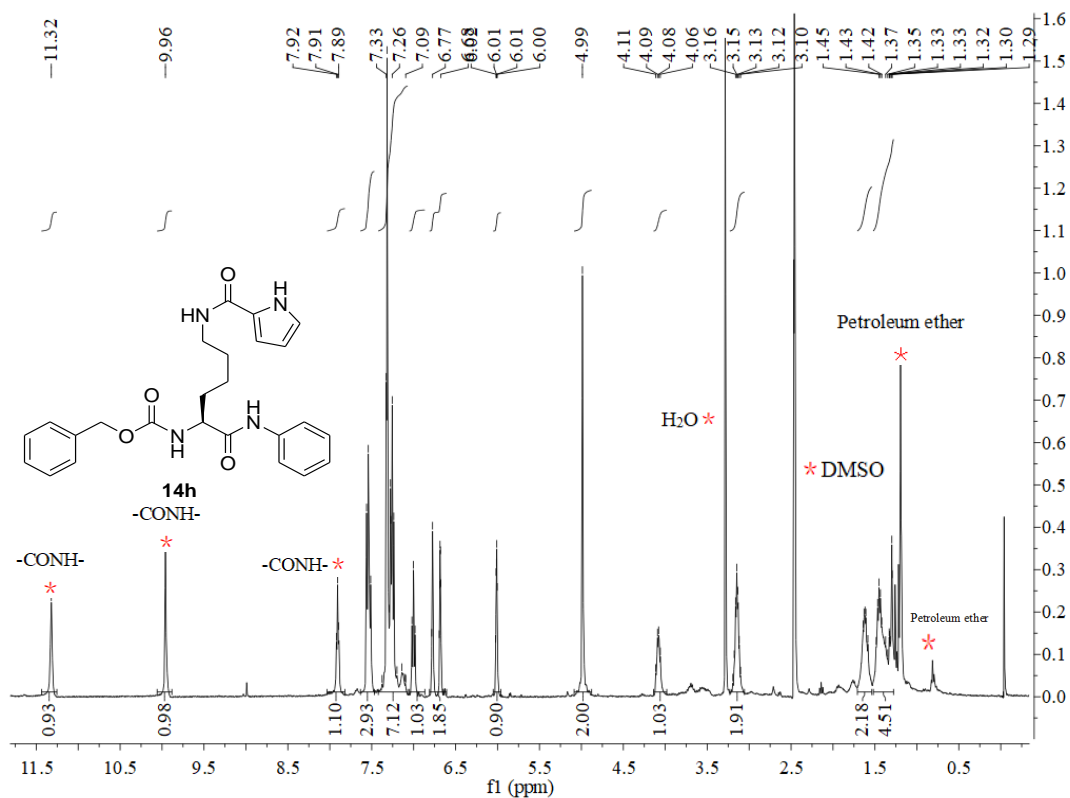
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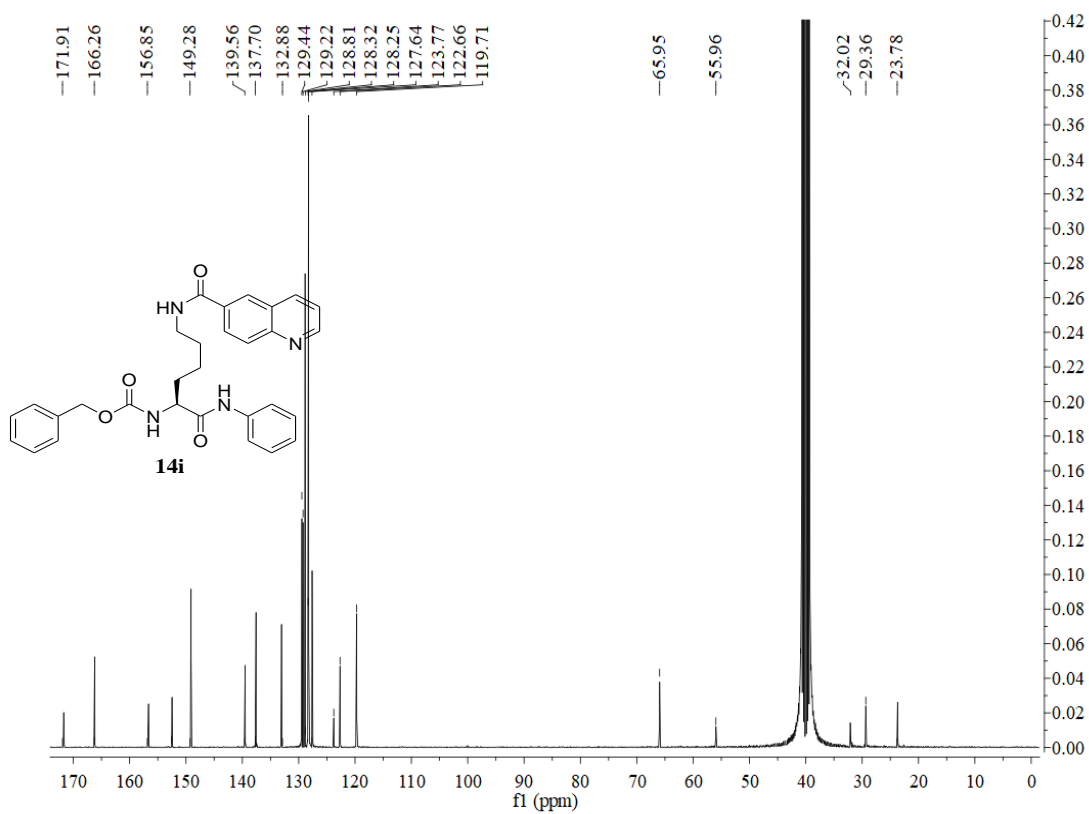
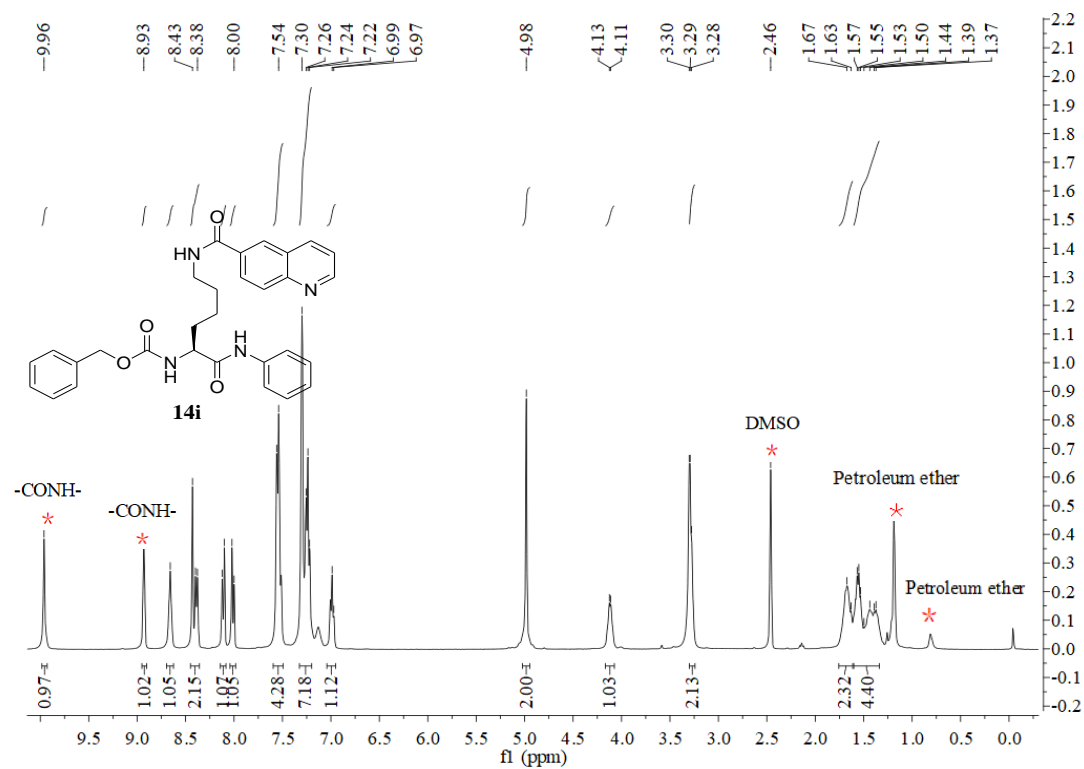
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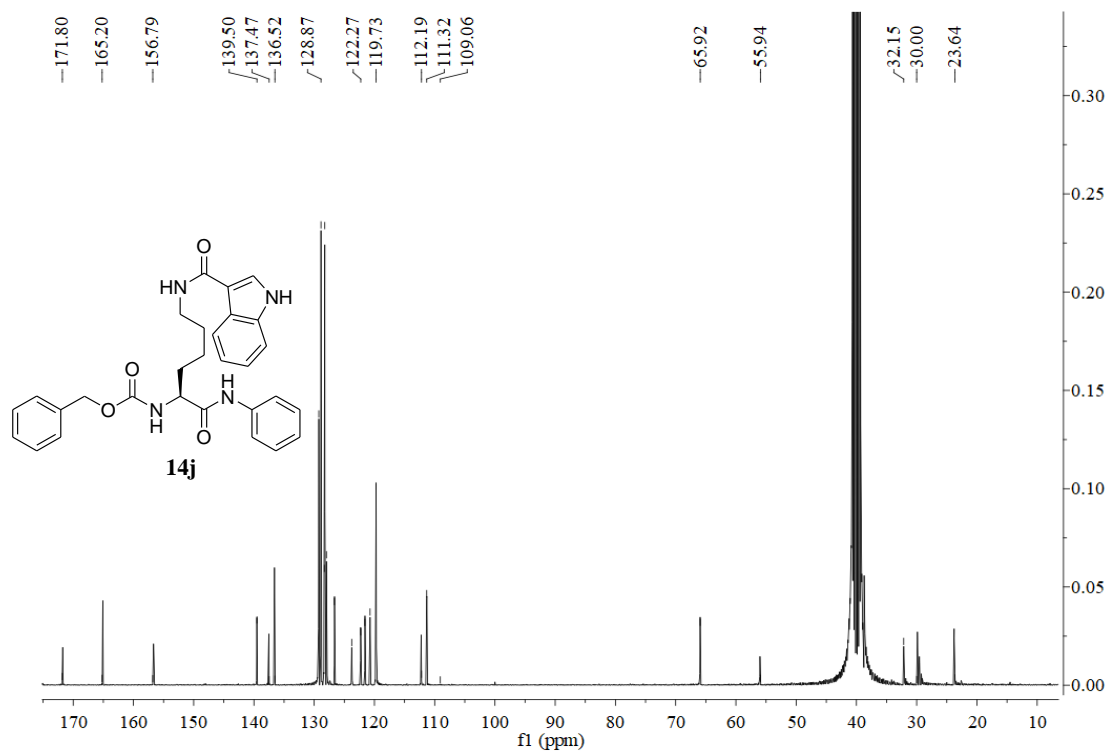
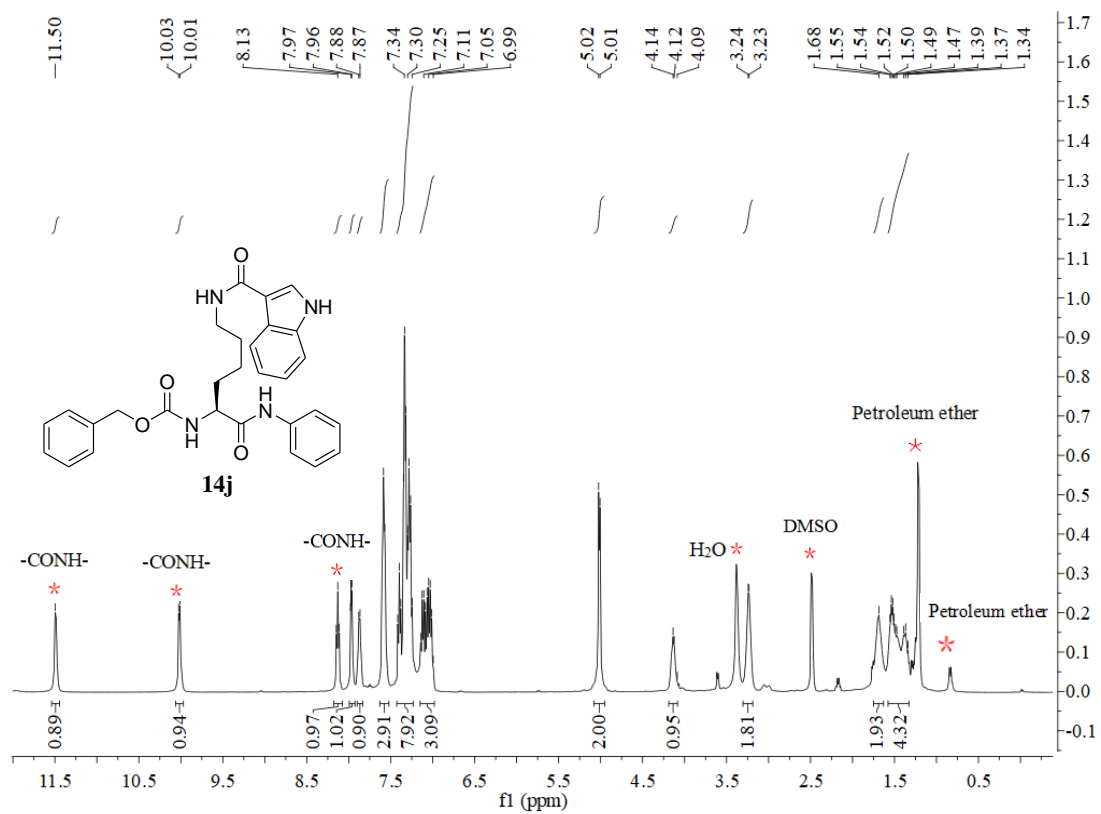
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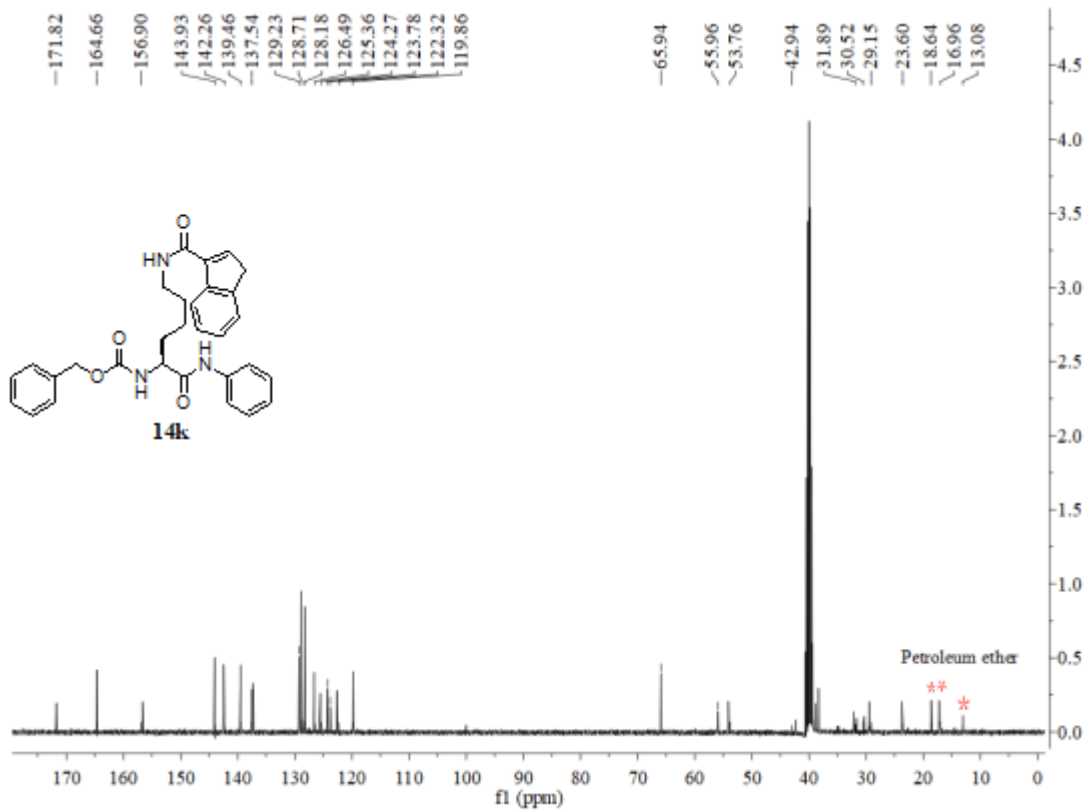
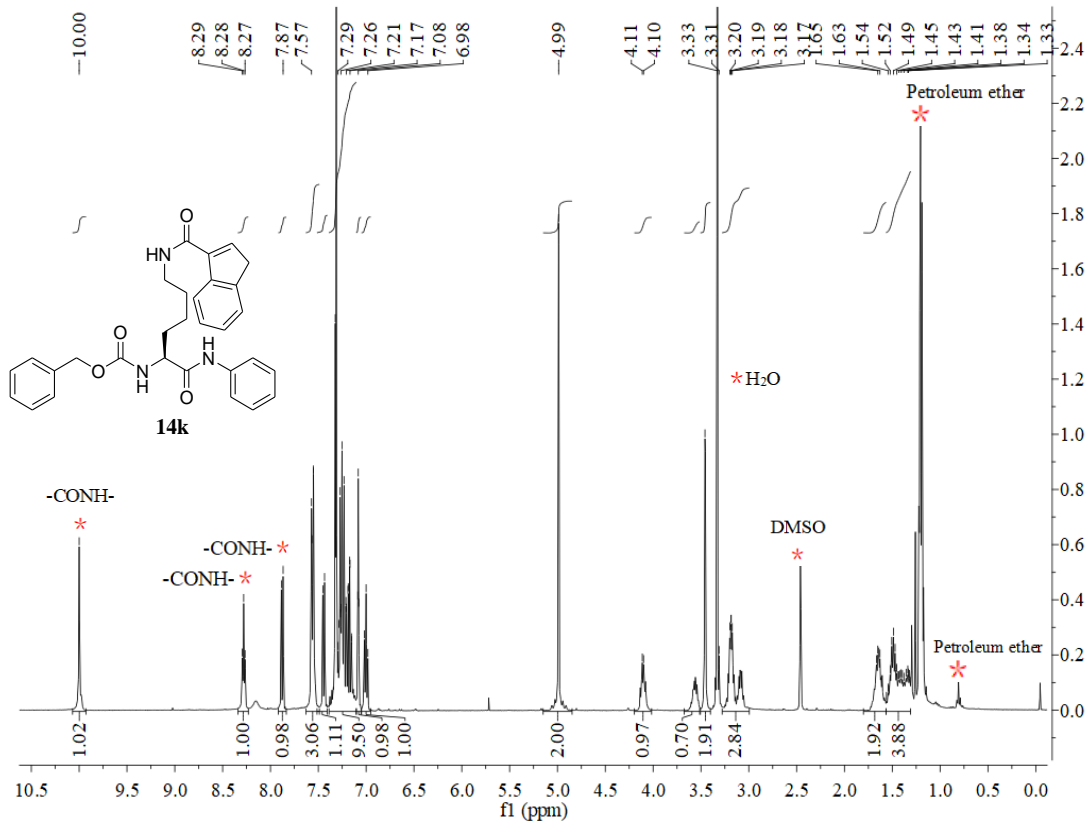
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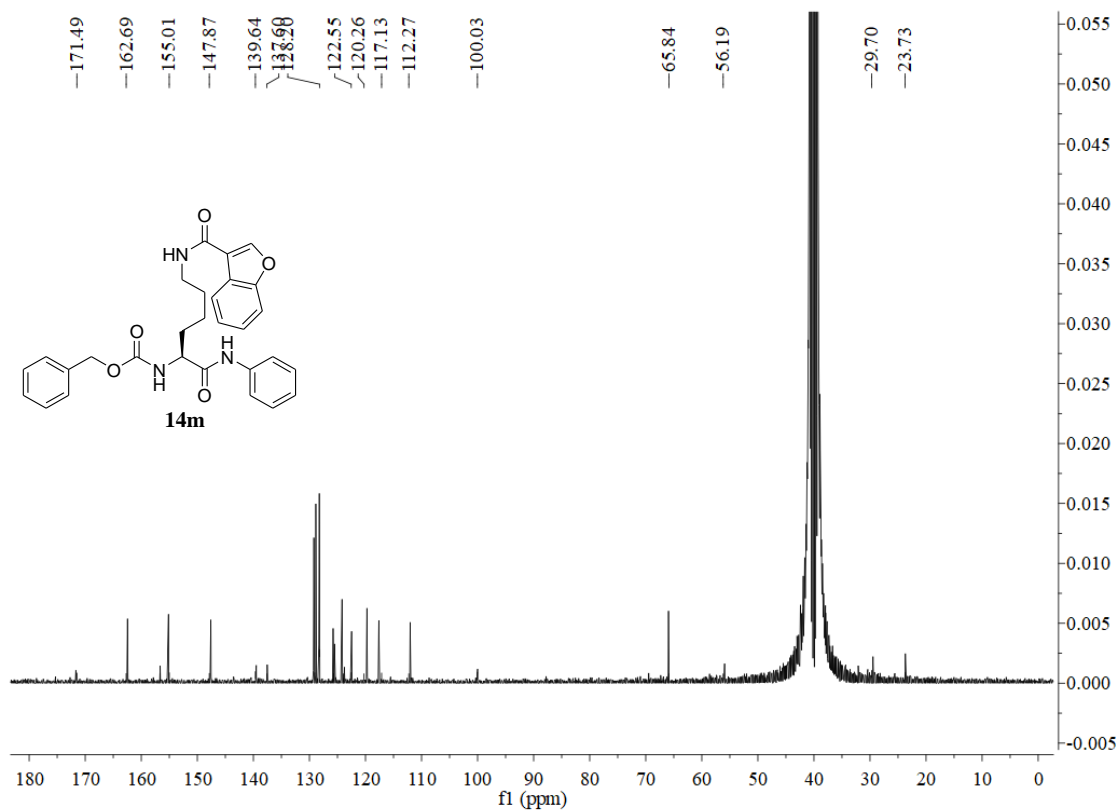
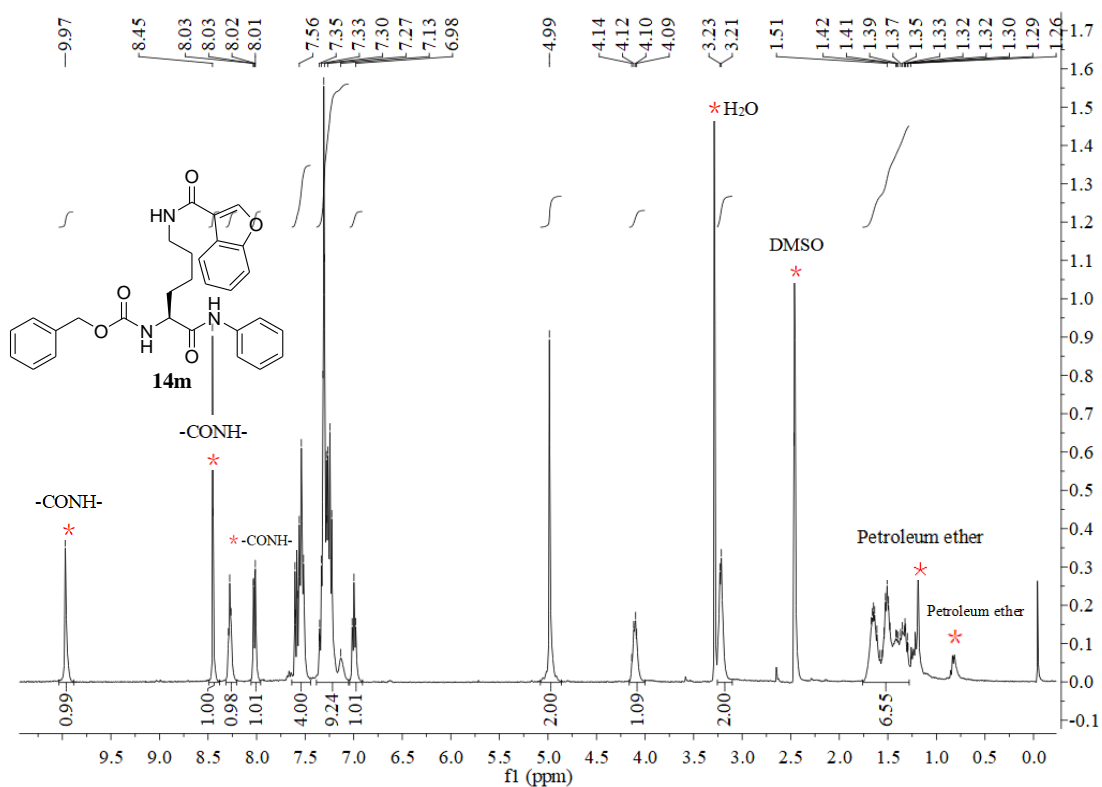
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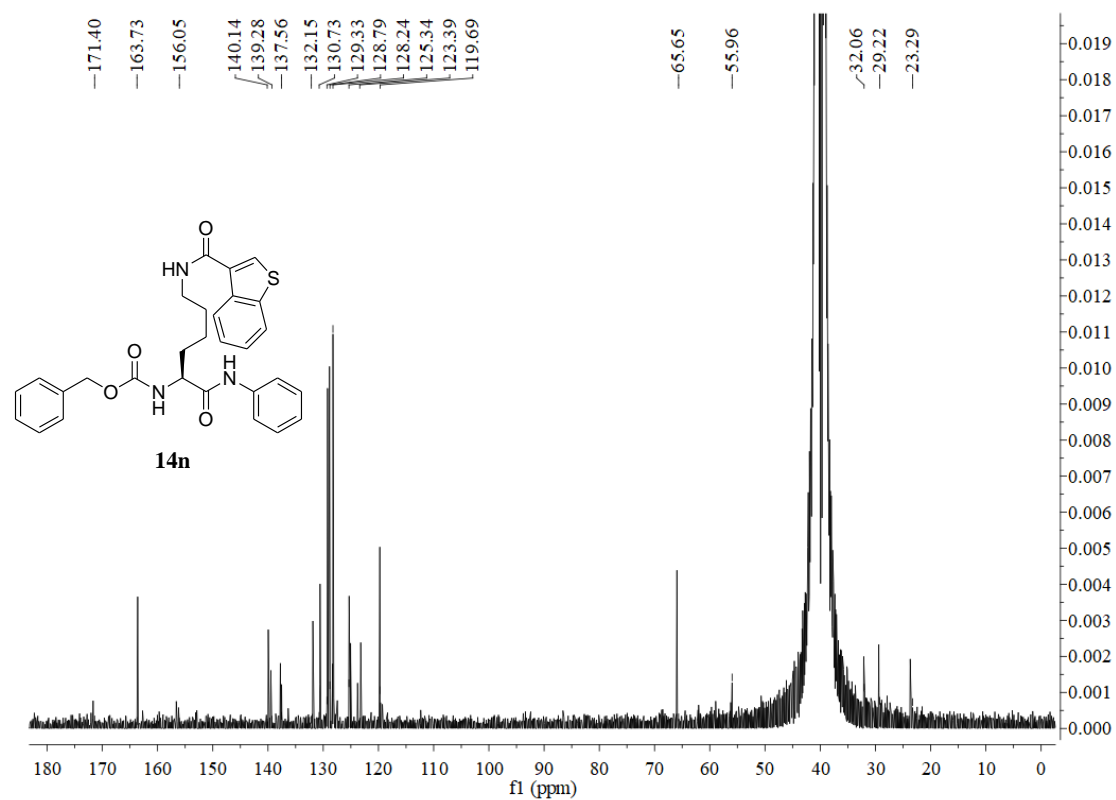
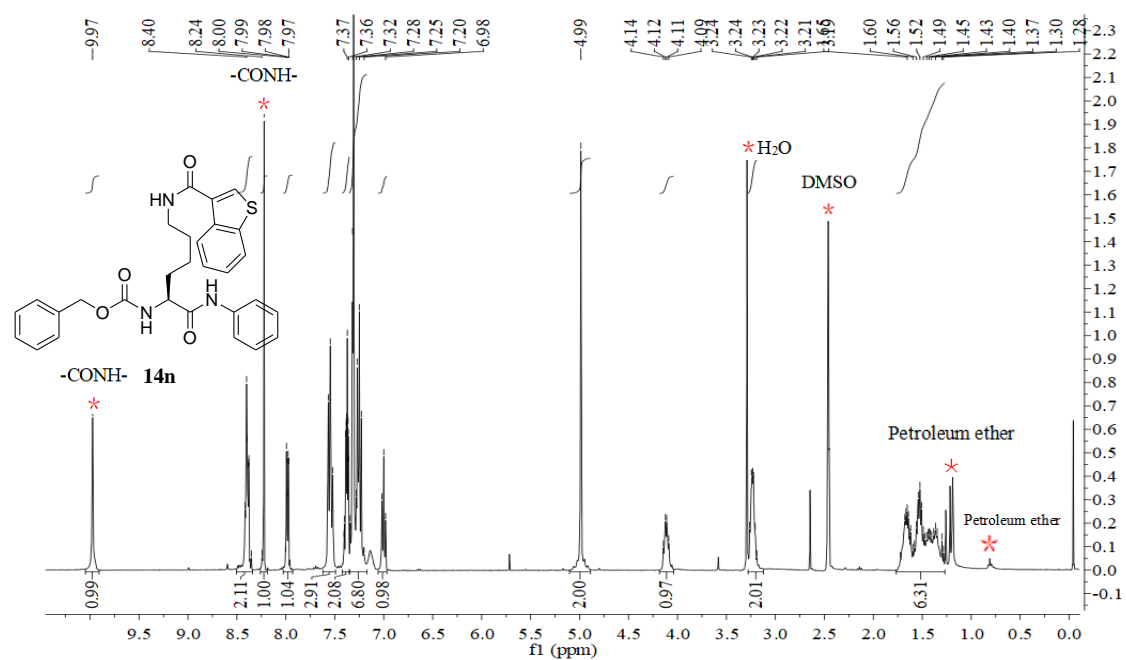
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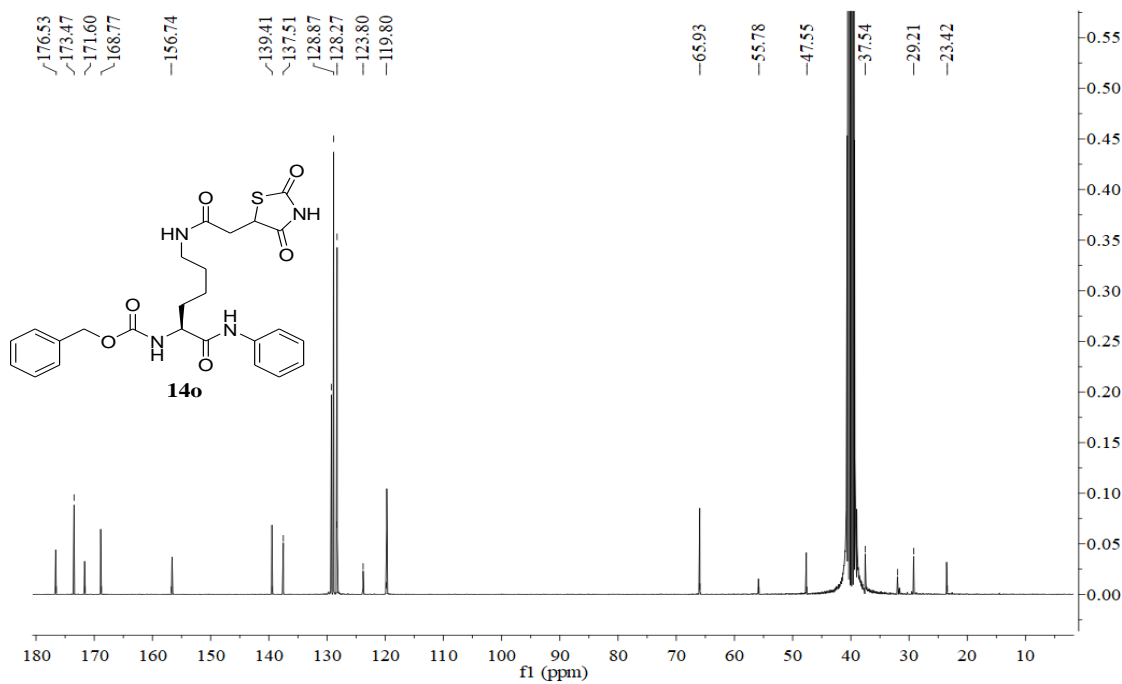
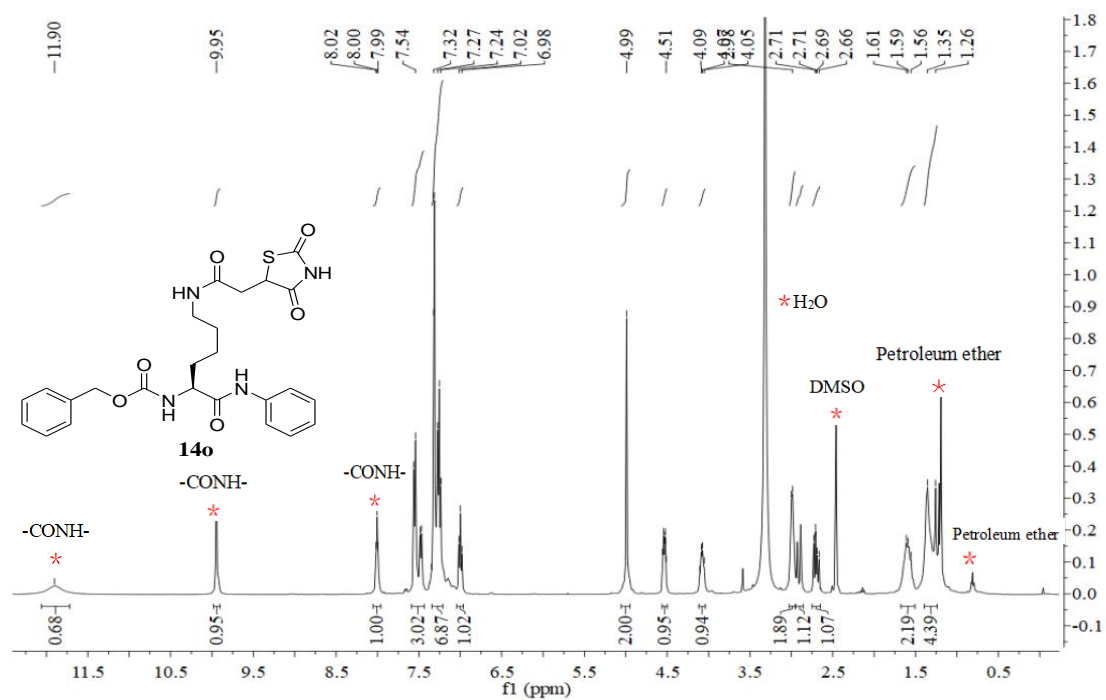
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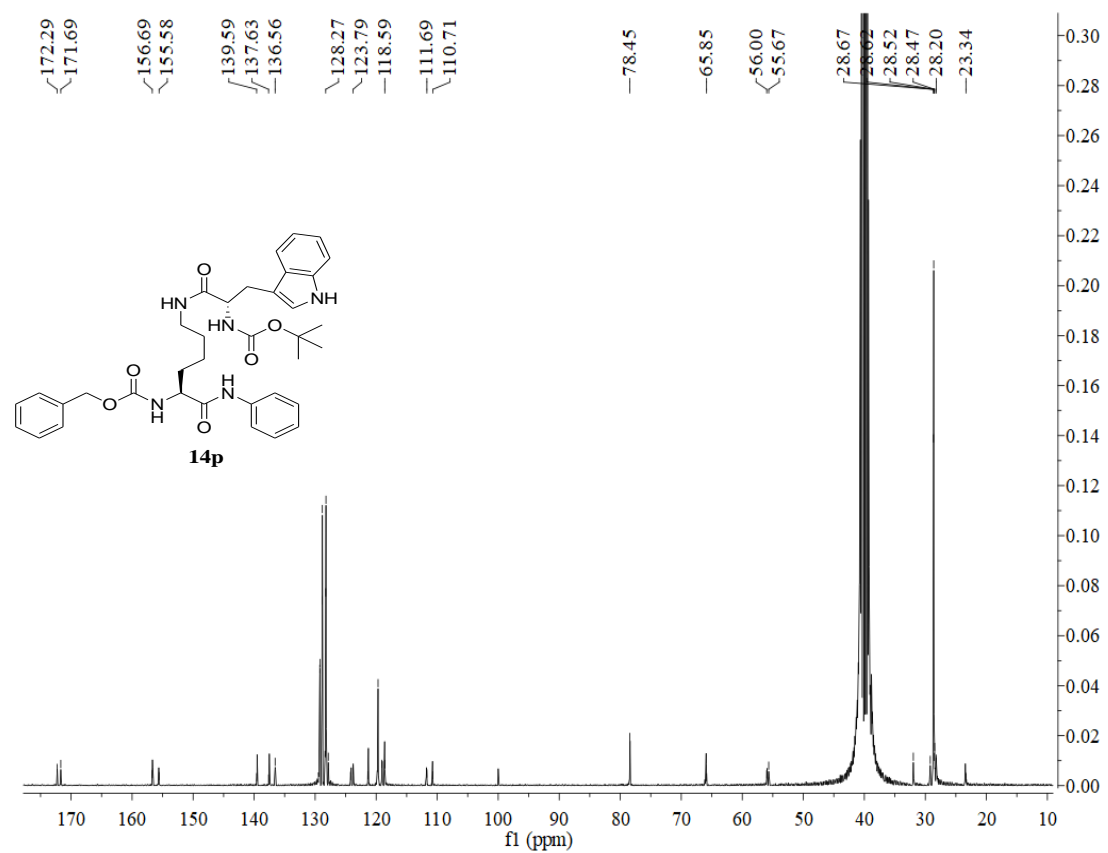
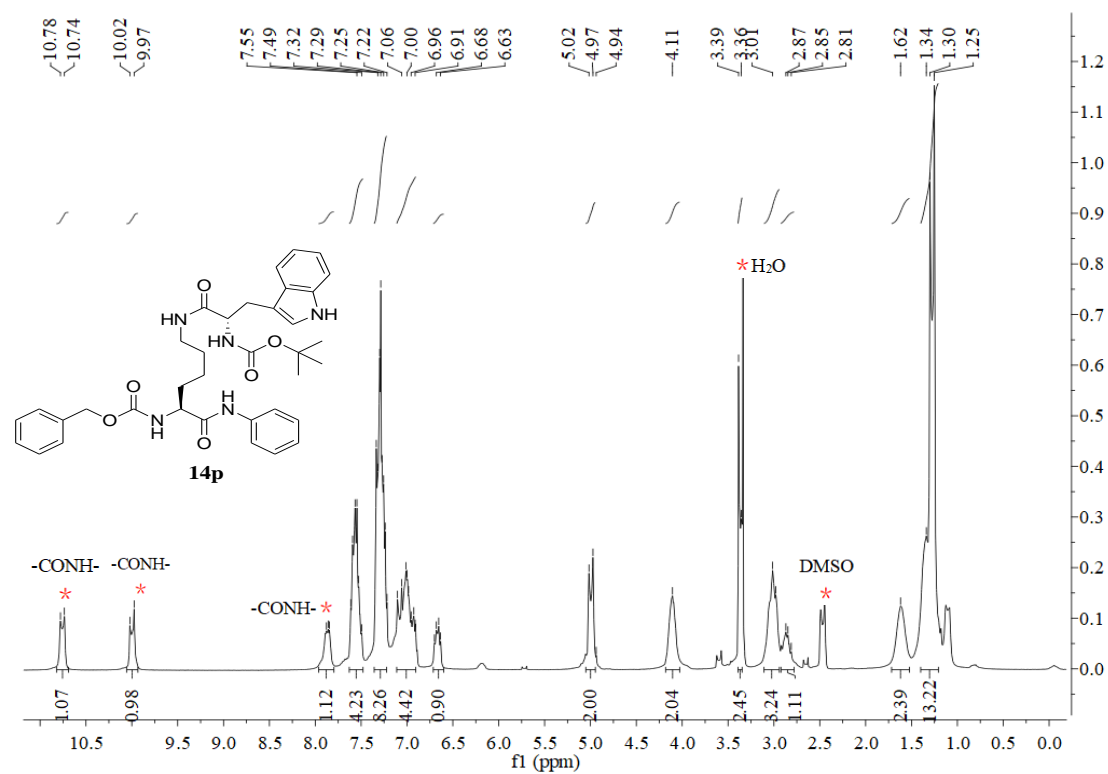
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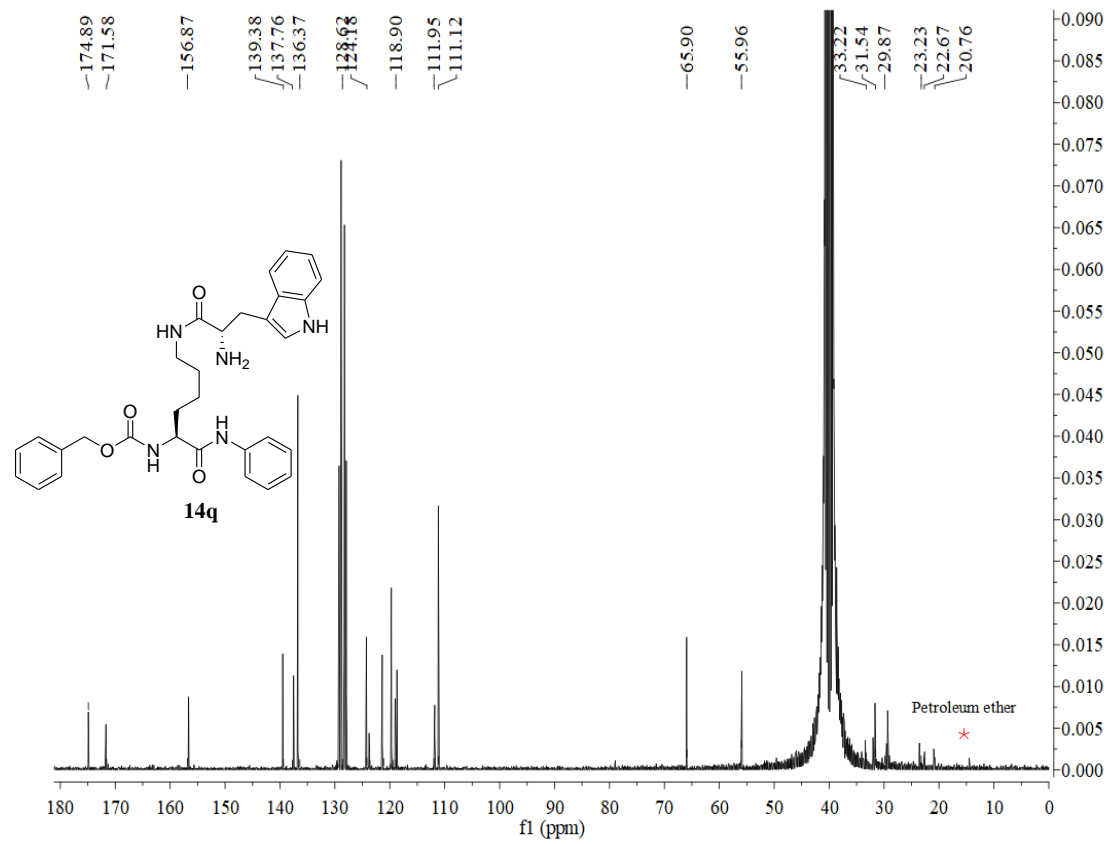
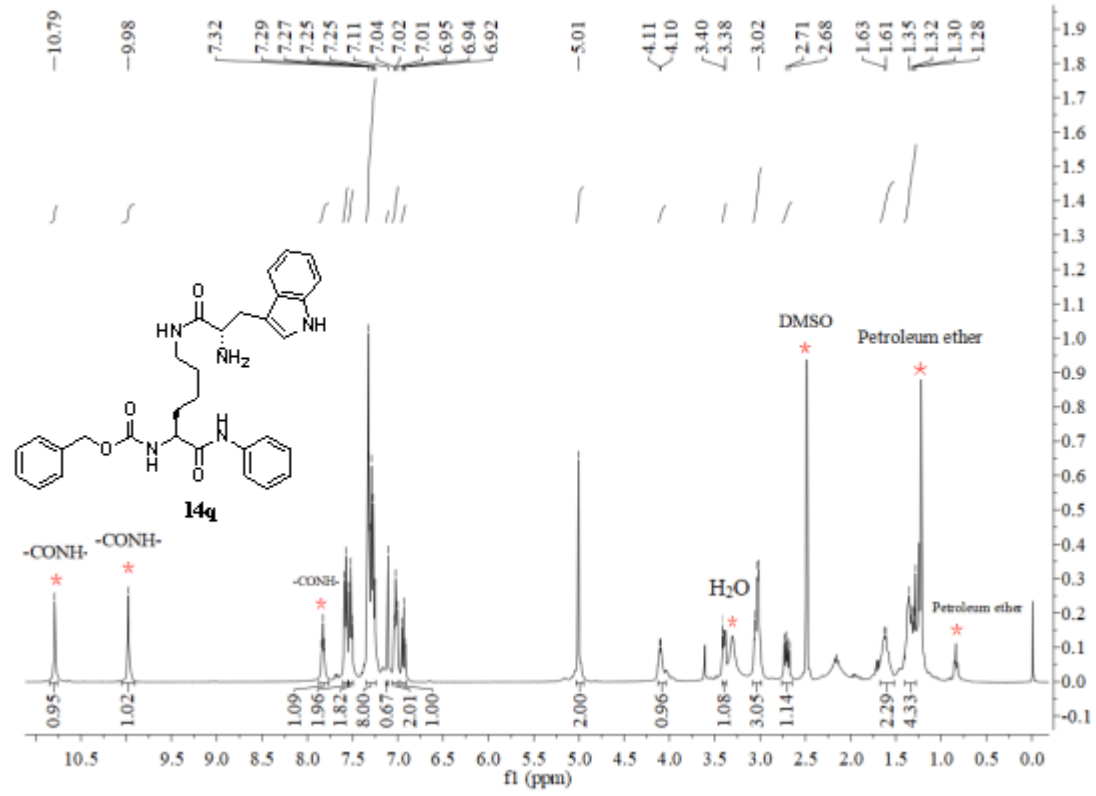
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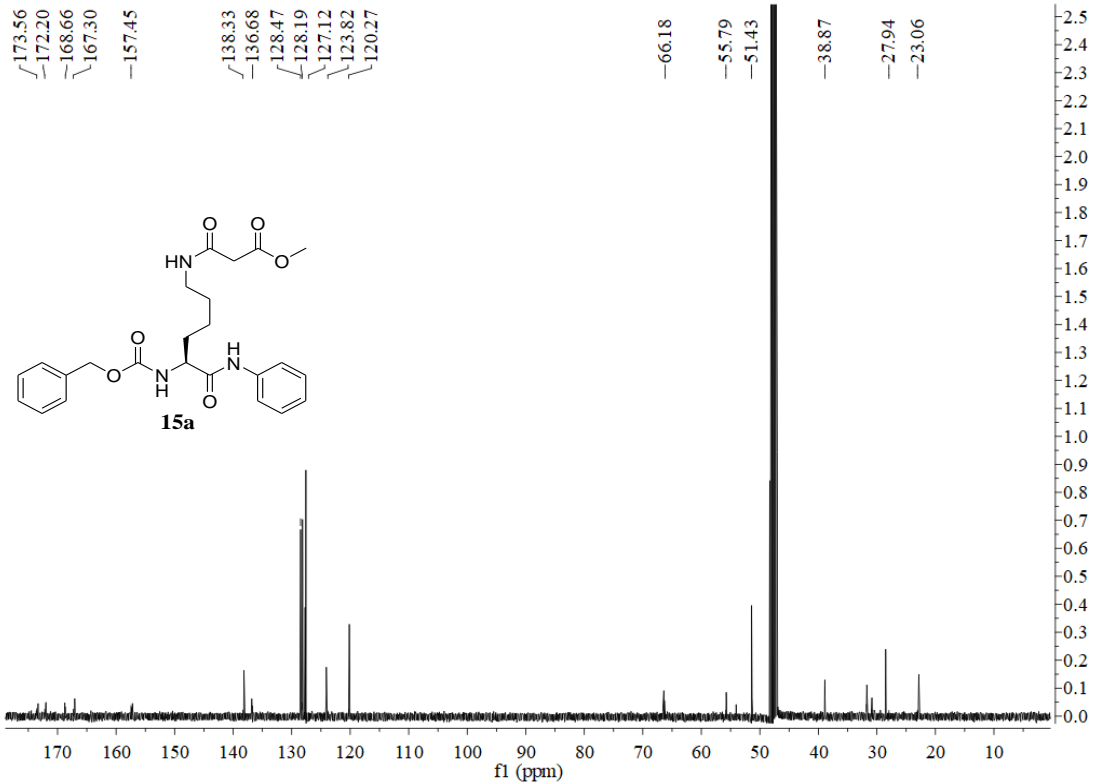
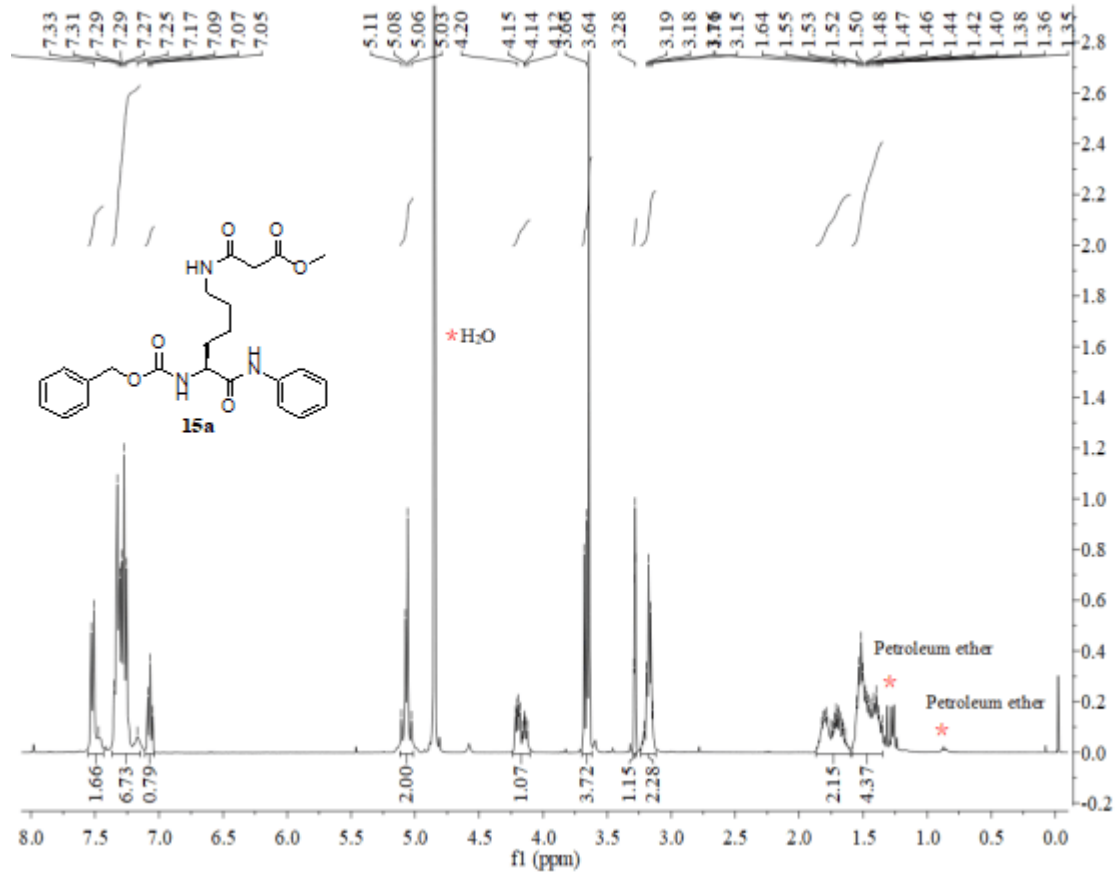
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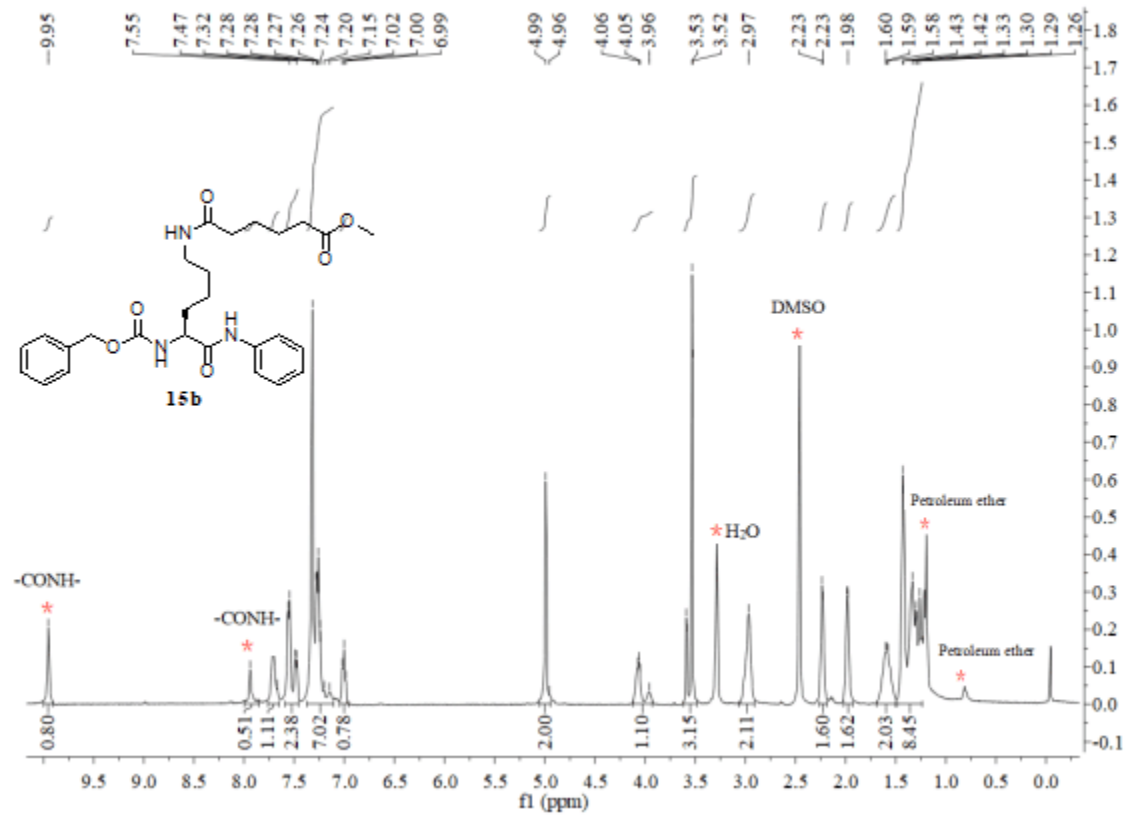
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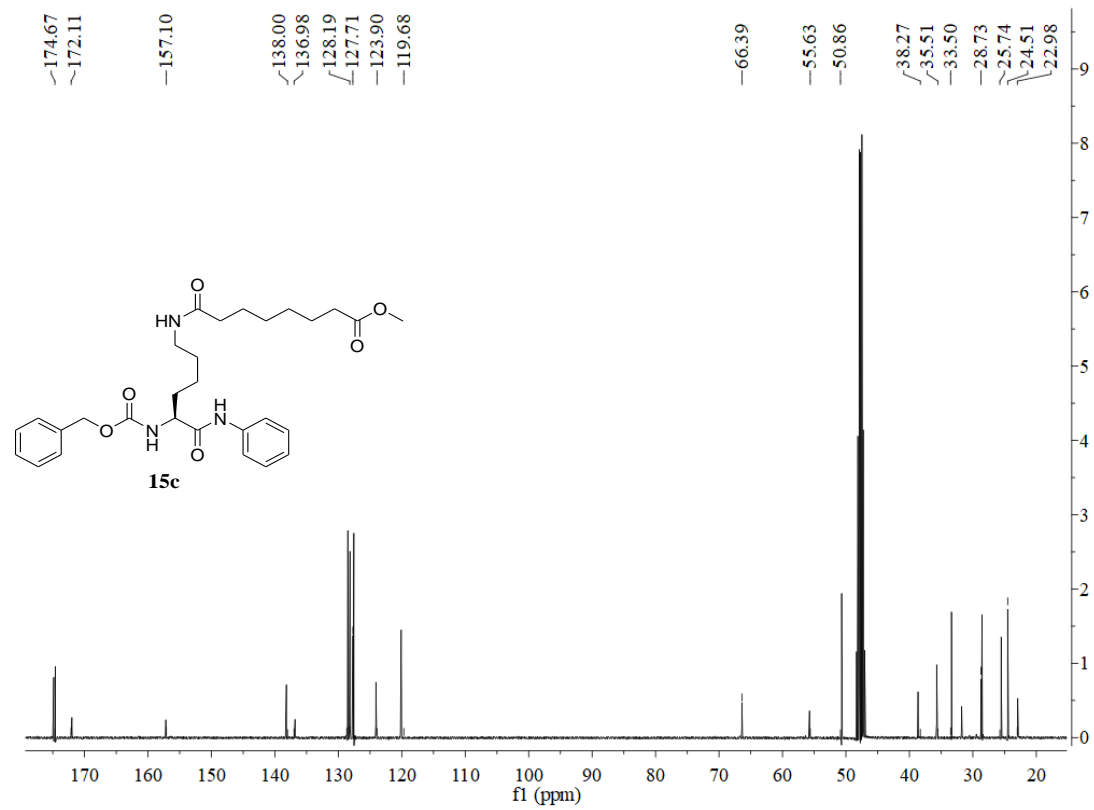
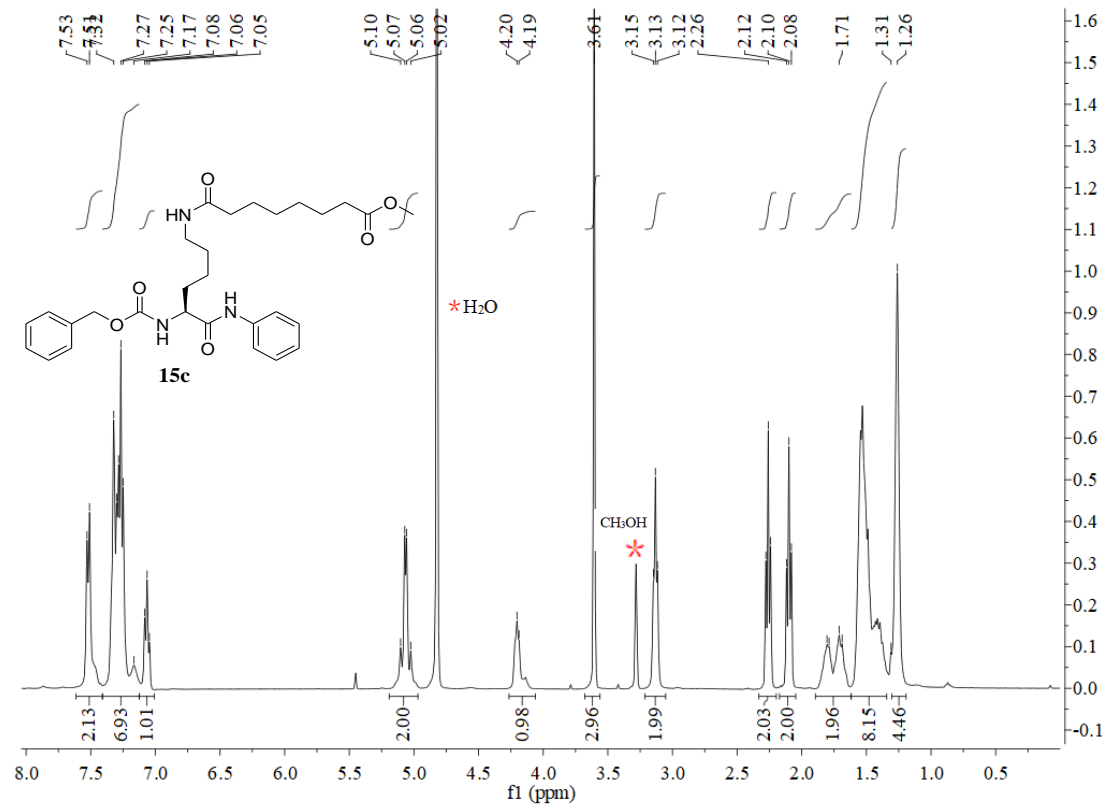
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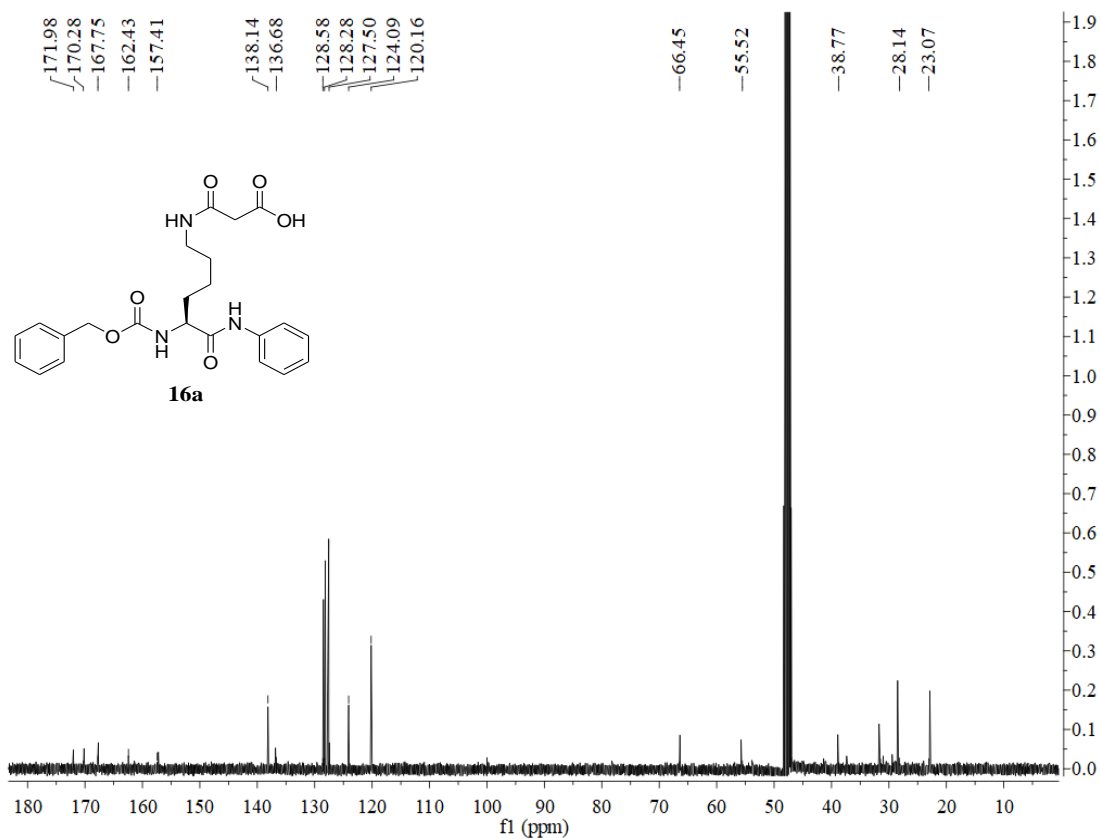
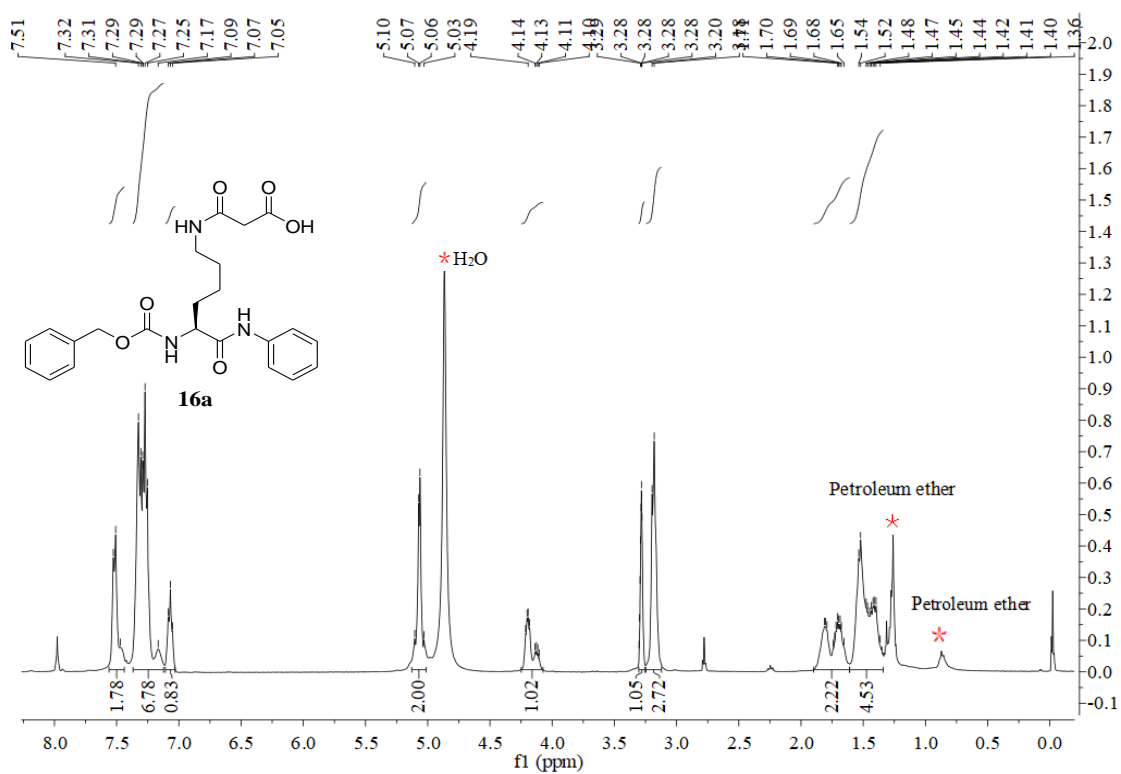
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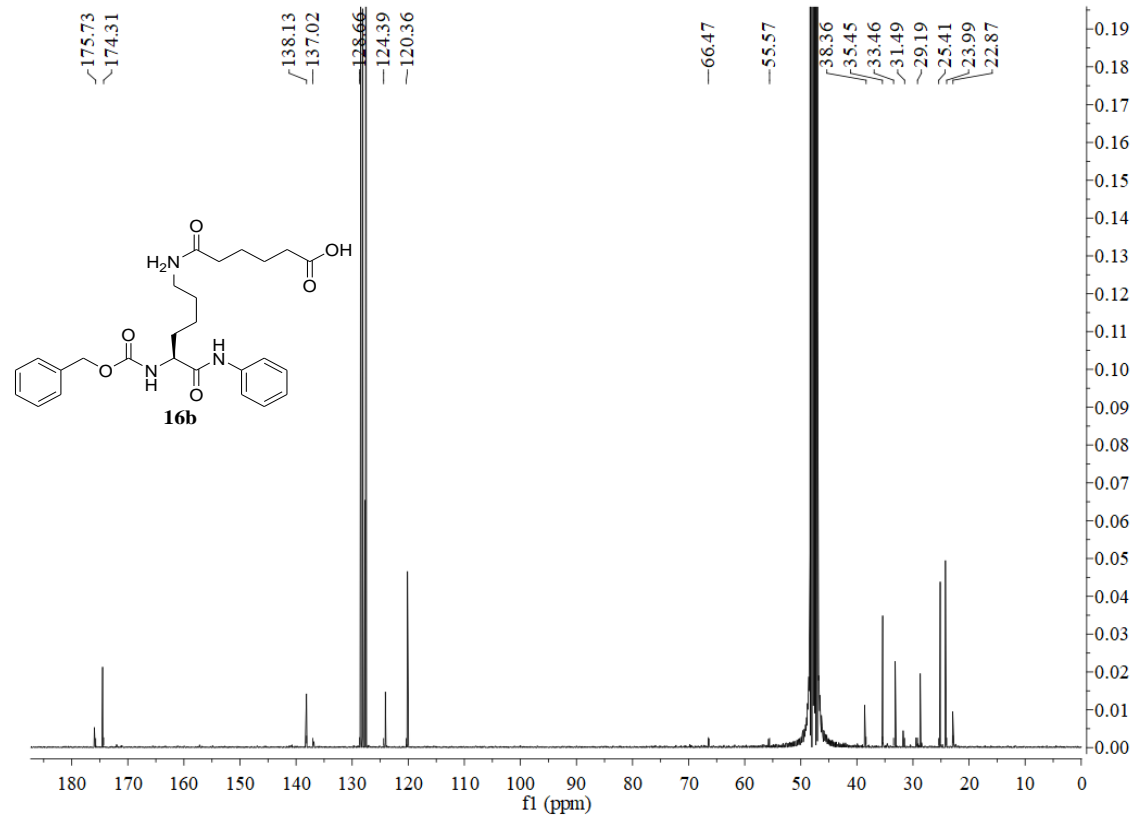
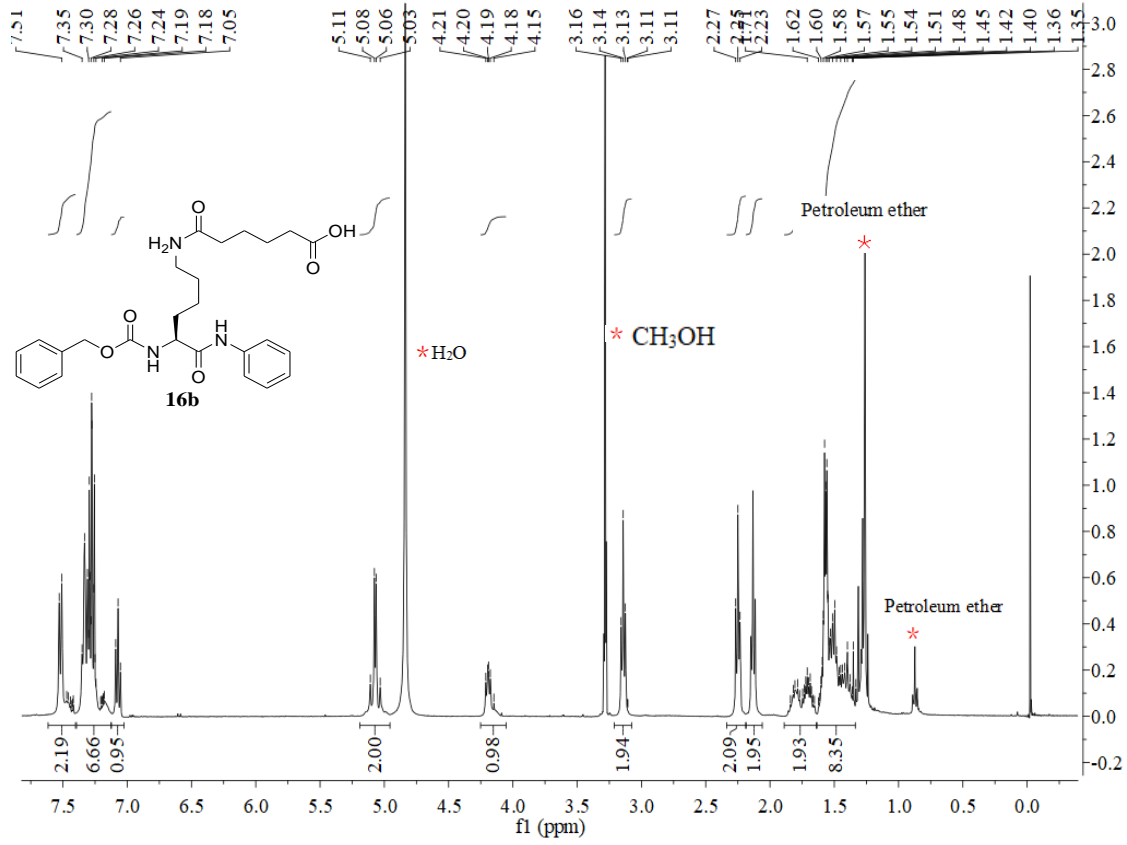
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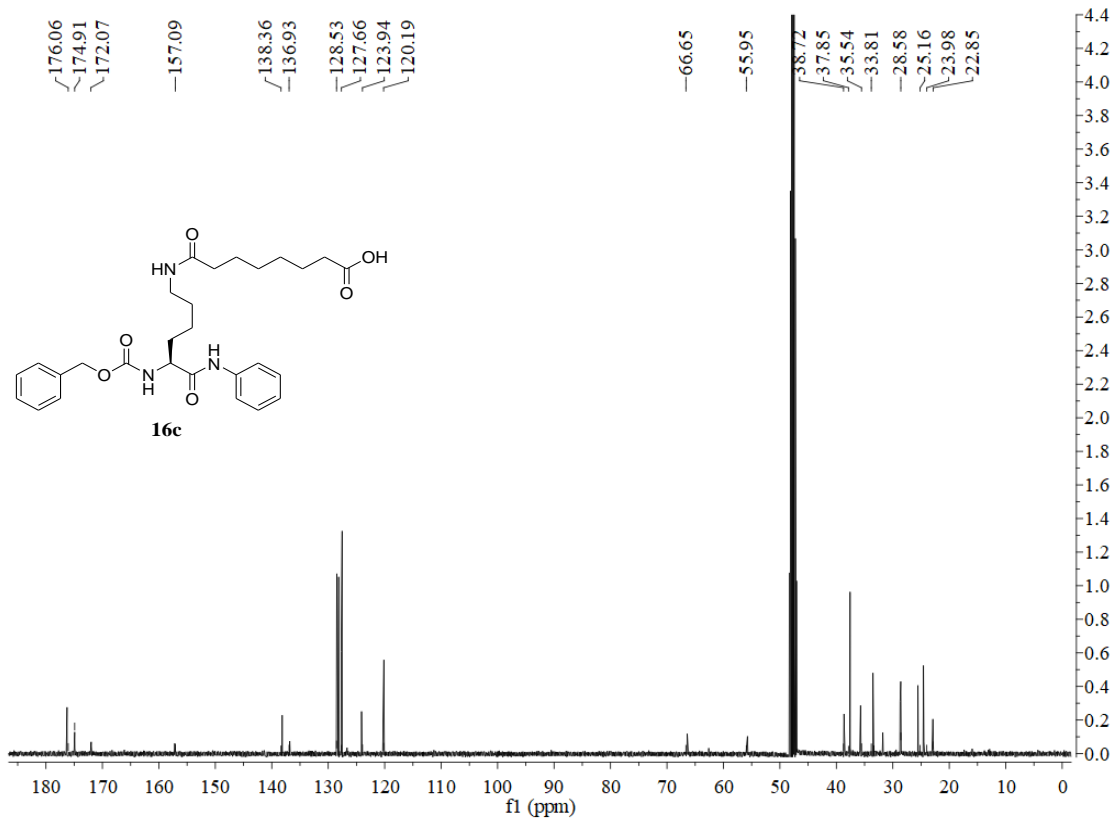
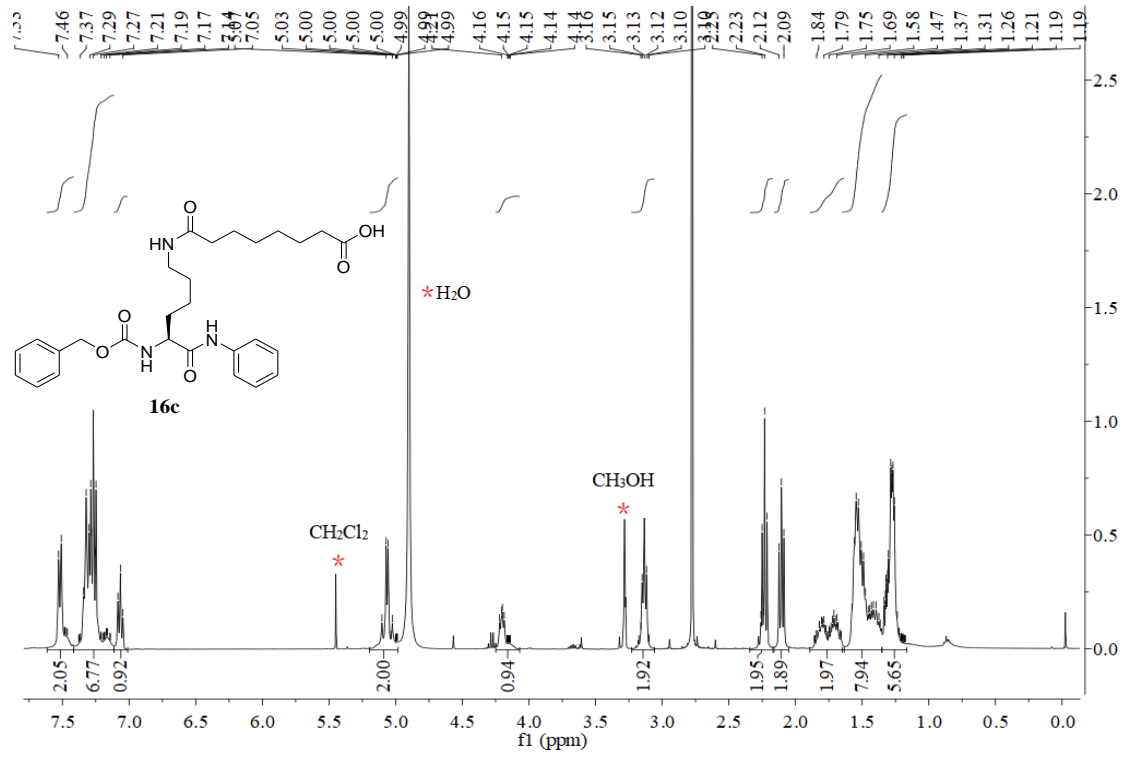
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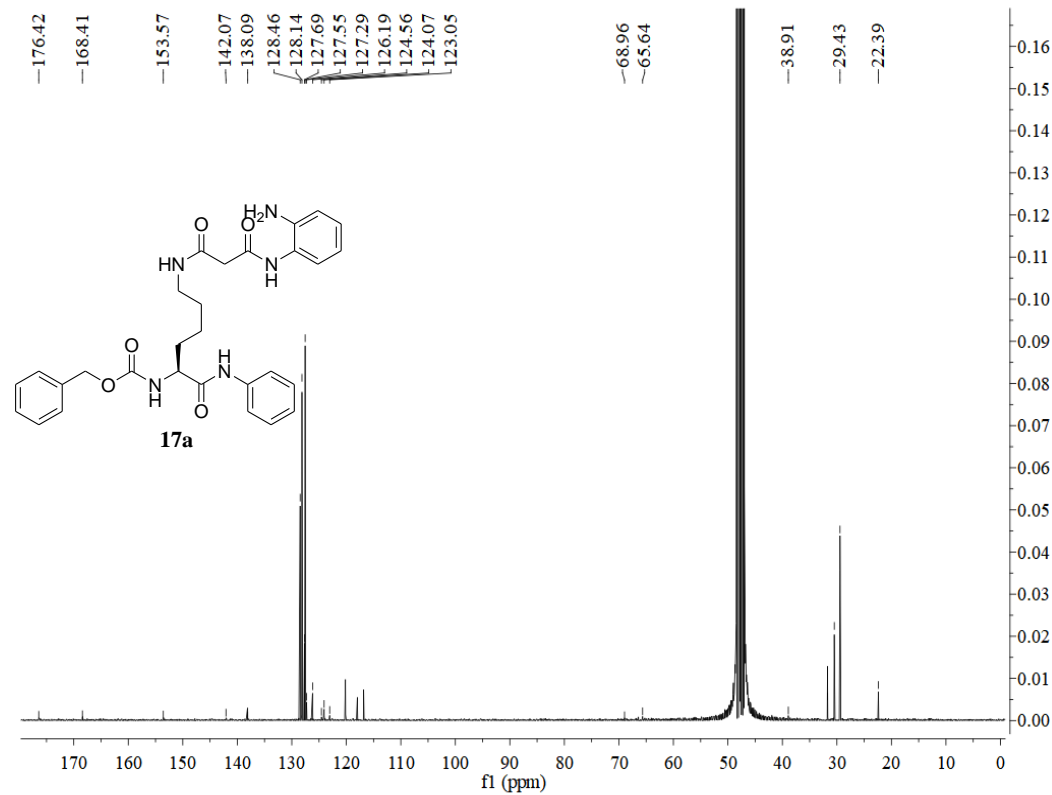
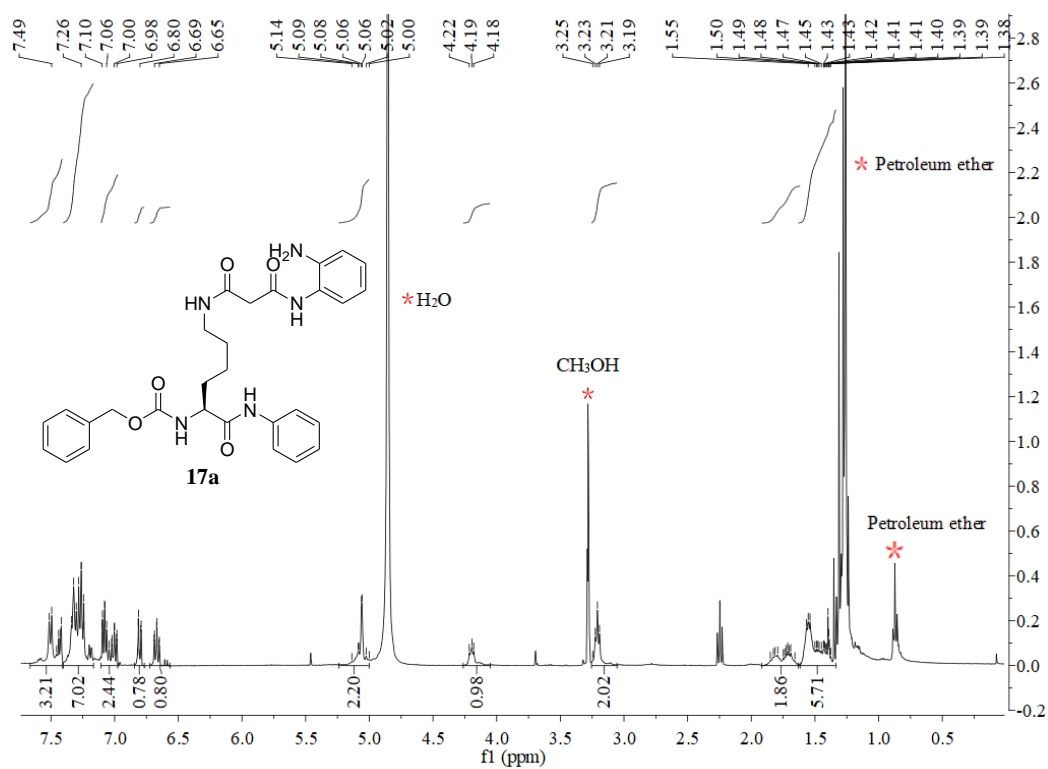
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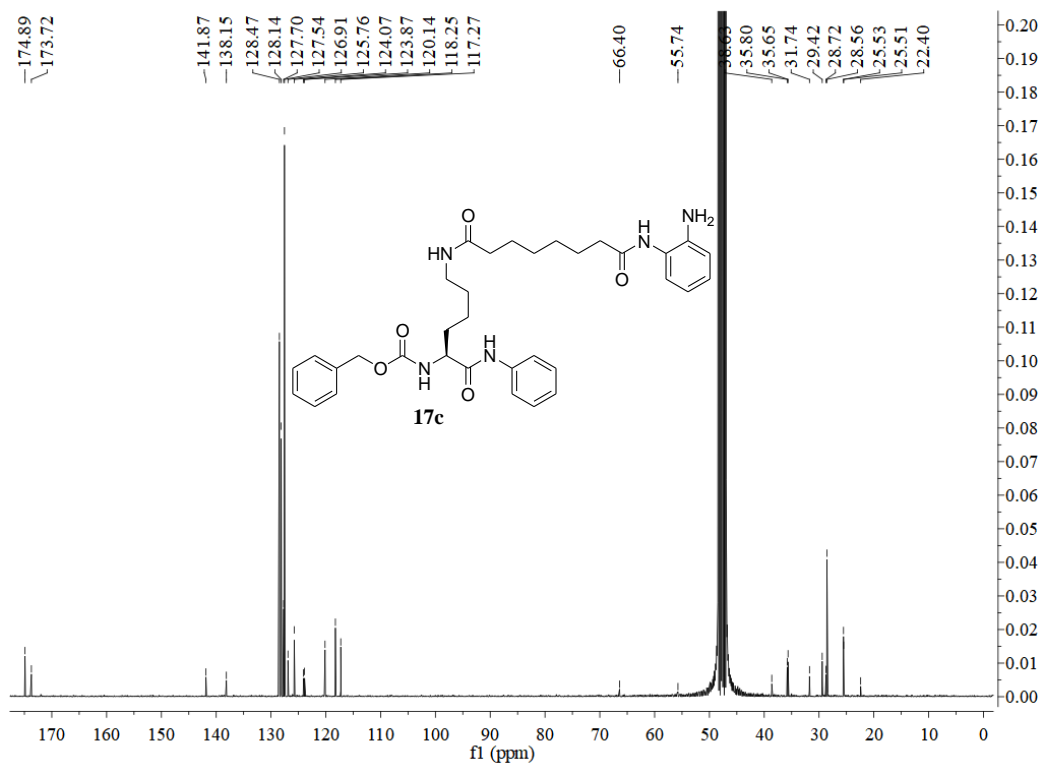
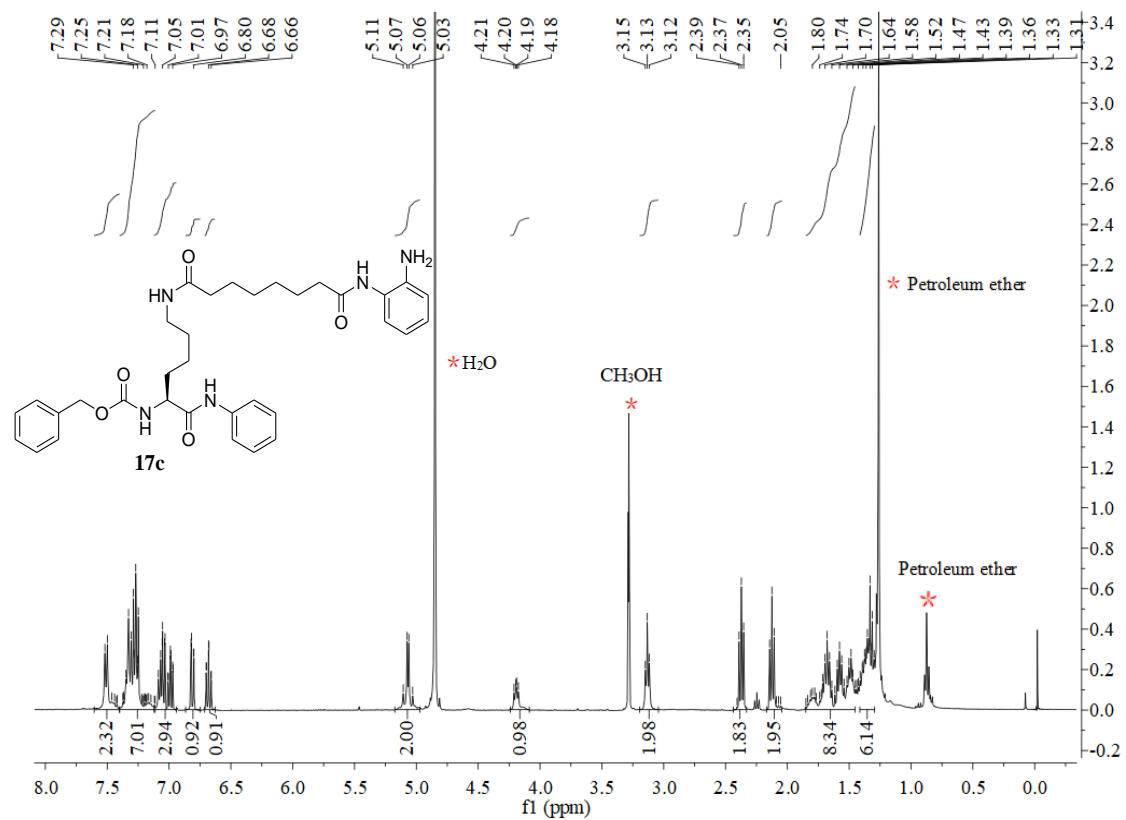
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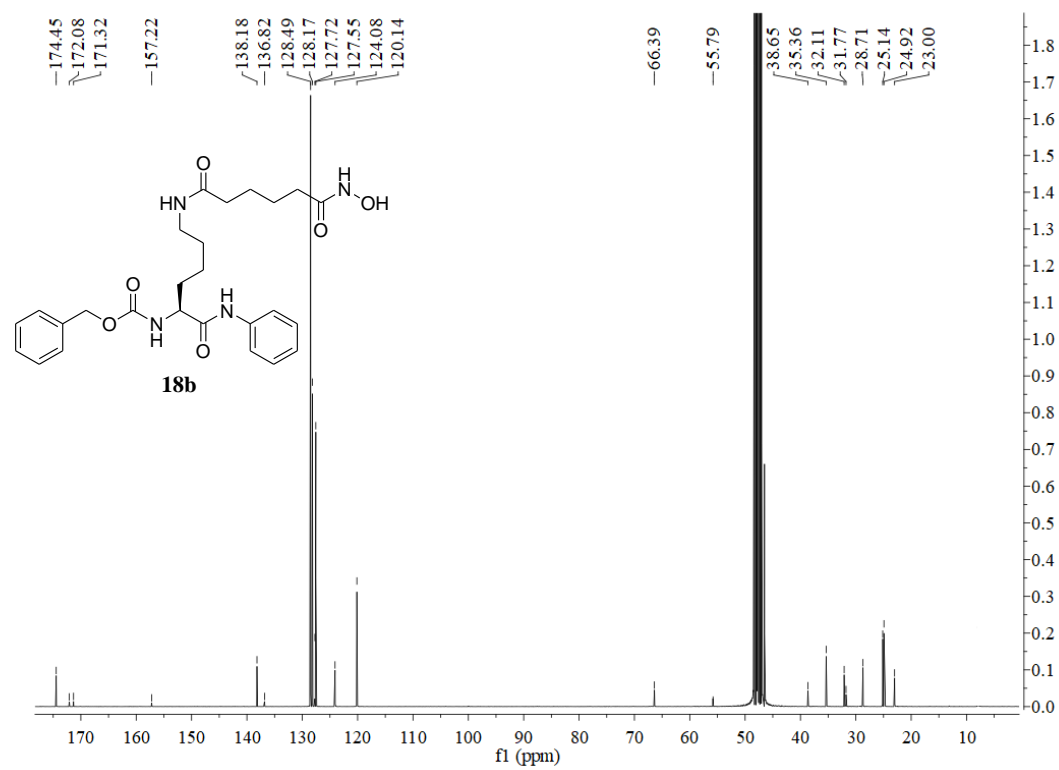
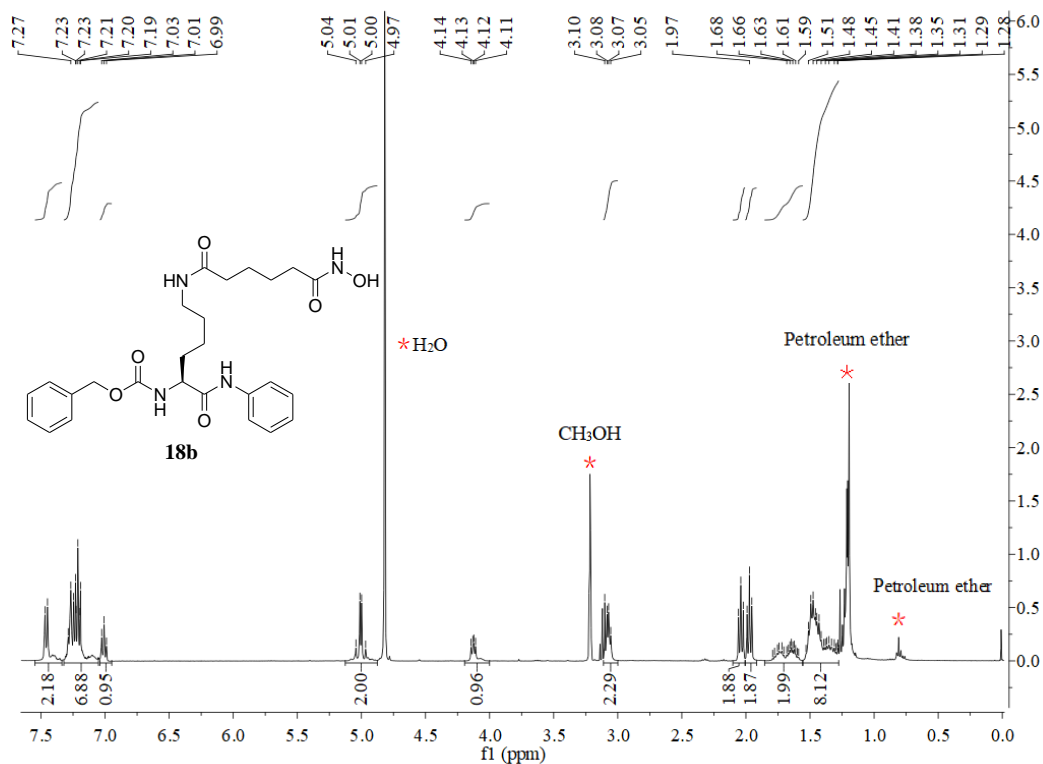
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