

Figure S1. Organization and gene expression patterns in the mouse embryo at E7.5. Lateral view of the mouse embryo at E7.5 with the anterior side to the left. The illustration on the left-hand side indicates the different tissue layers and structures of the embryo, and the one to the right depicts the main gene expression pattern in those regions. The sources for the outlines shown here can be found in Table S1 and (Edri et al., 2018).

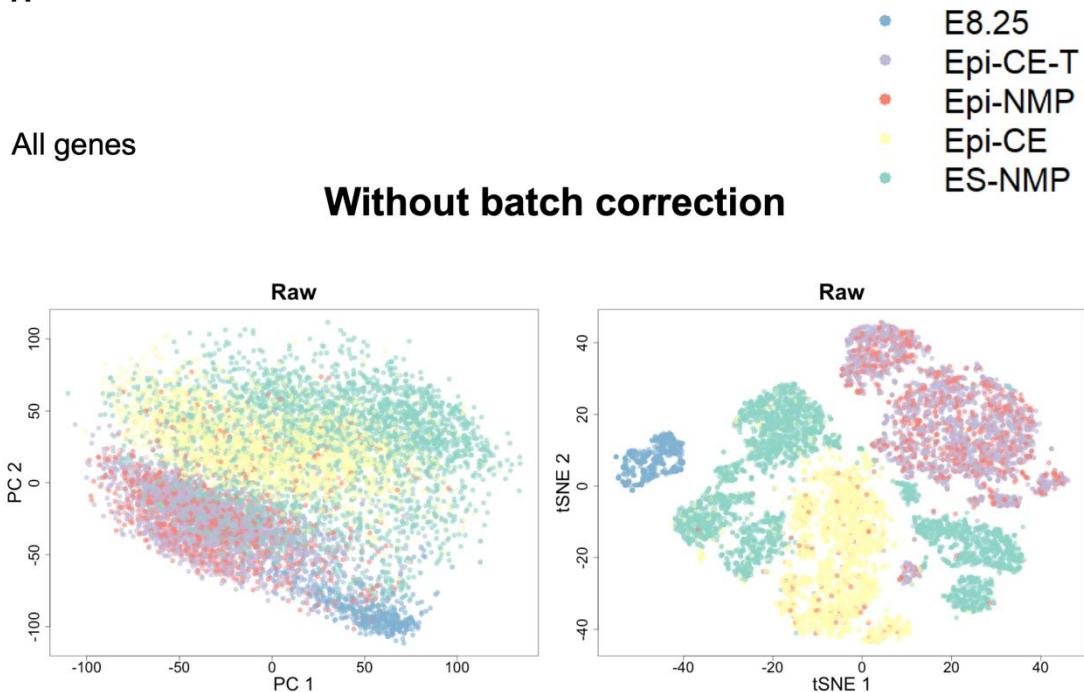
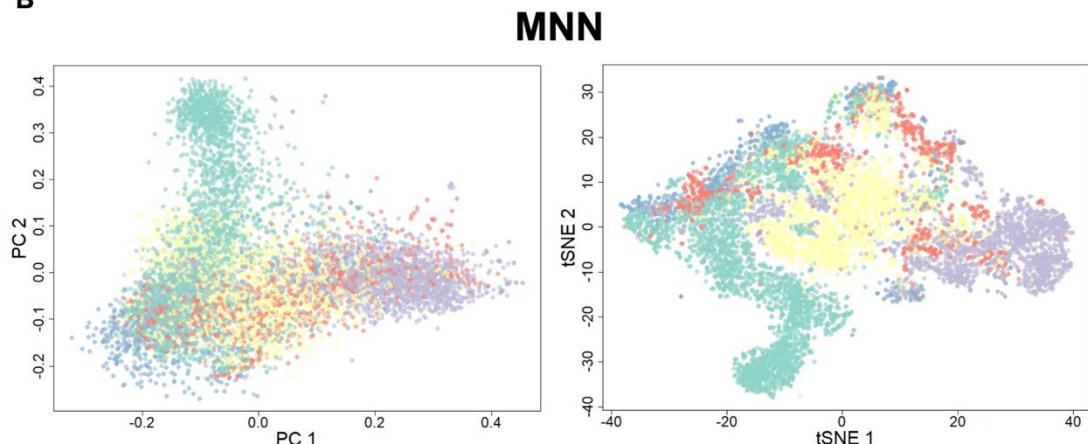
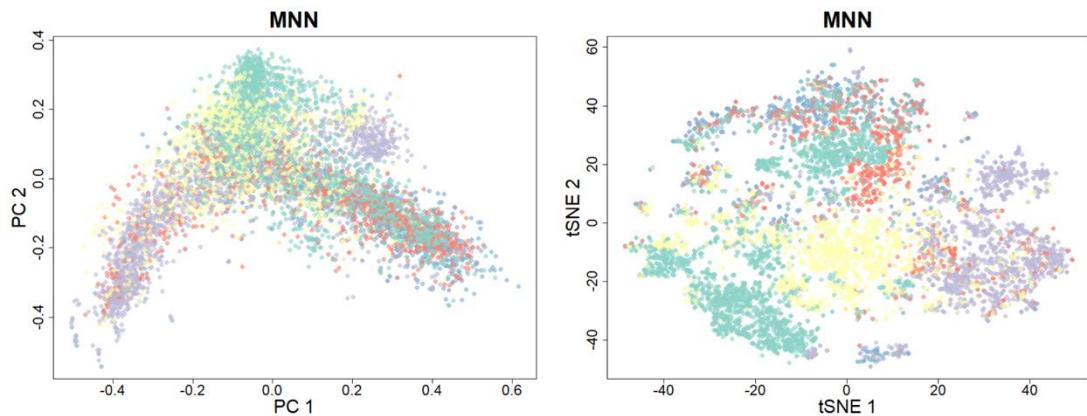
A**B**

Figure S2. Correction for RNA-seq single cells technical variation. **A.** PCA and tSNE plots of the raw data after completing the quality control for the cells and genes. The plots were performed using all genes (14,822 genes) that passed the quality control. **B.** Batch effects correction using mutual nearest neighbour (MNN, (Haghverdi et al., 2018)).

A

- E8.25
- Epi-CE-T
- Epi-NMP
- Epi-CE
- ES-NMP

Highly variable genes detected by MNN algorithm:

**B**

Informative genes:

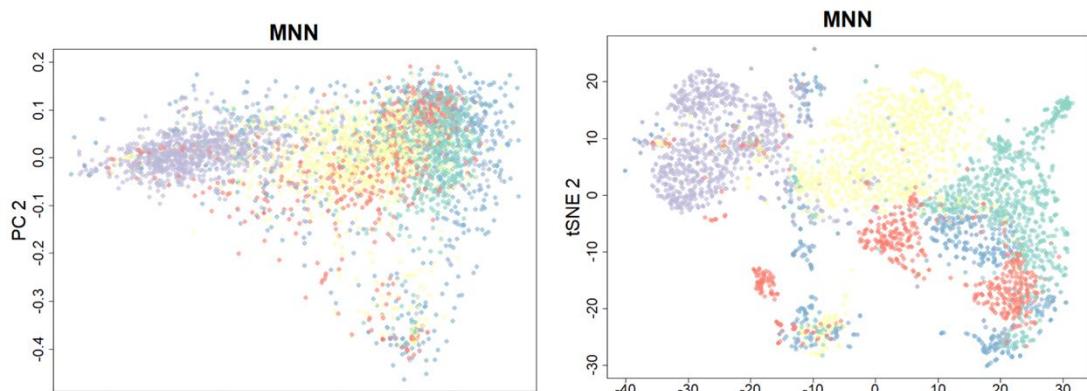


Figure S3. Correction for RNA-seq single cells technical variation for subset of genes. **A.** MNN batch effects correction using the highly variable genes detected by MNN. **B.** MNN batch effects correction using 1,342 genes of interest gathered from a literature review (see main text and Table S2).

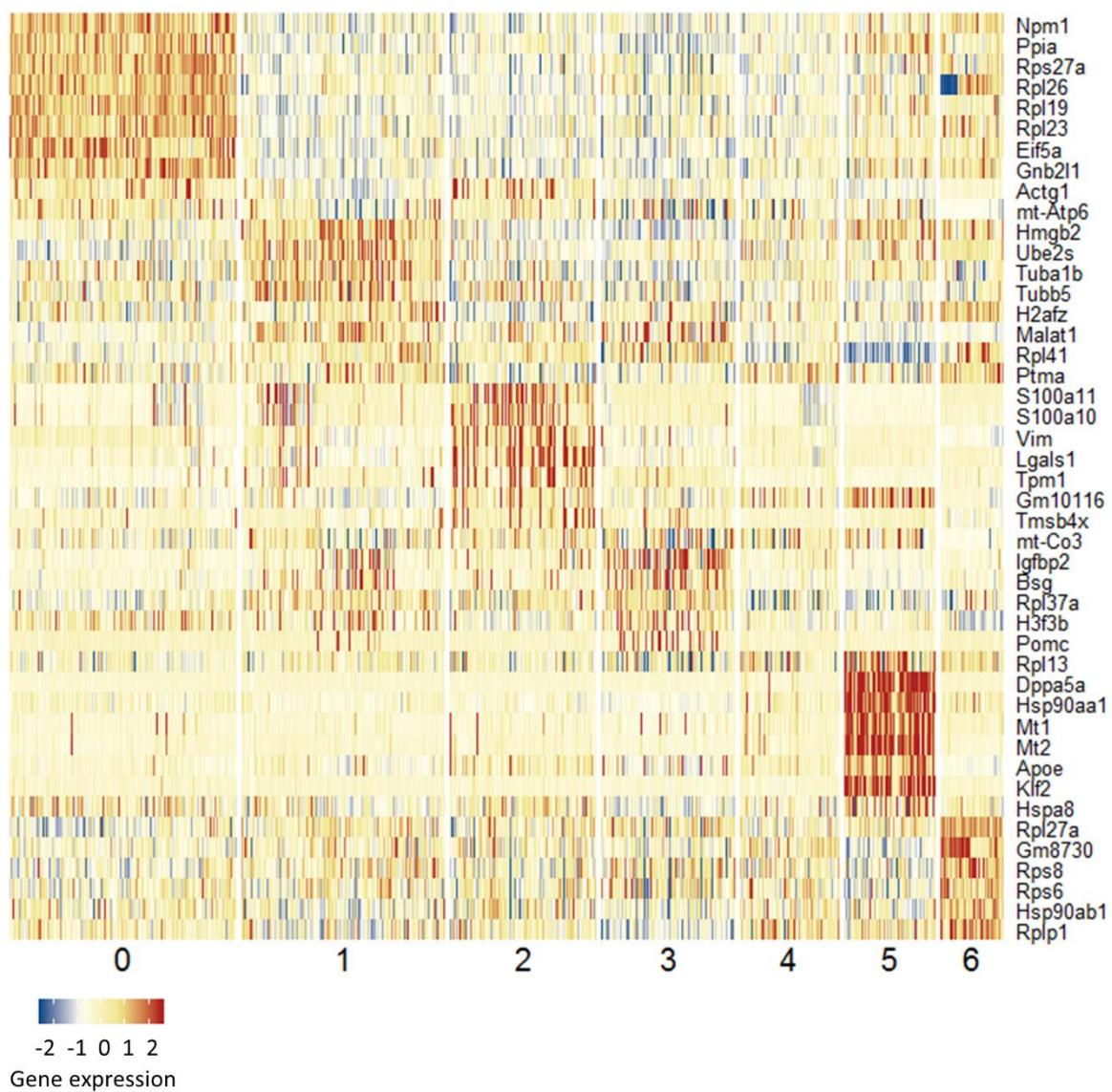


Figure S4. Clustering the embryo dataset and the *in vitro* populations after batch correcting process. Top 10 marker genes characterizing each of the 7 clusters obtained by Seurat package.

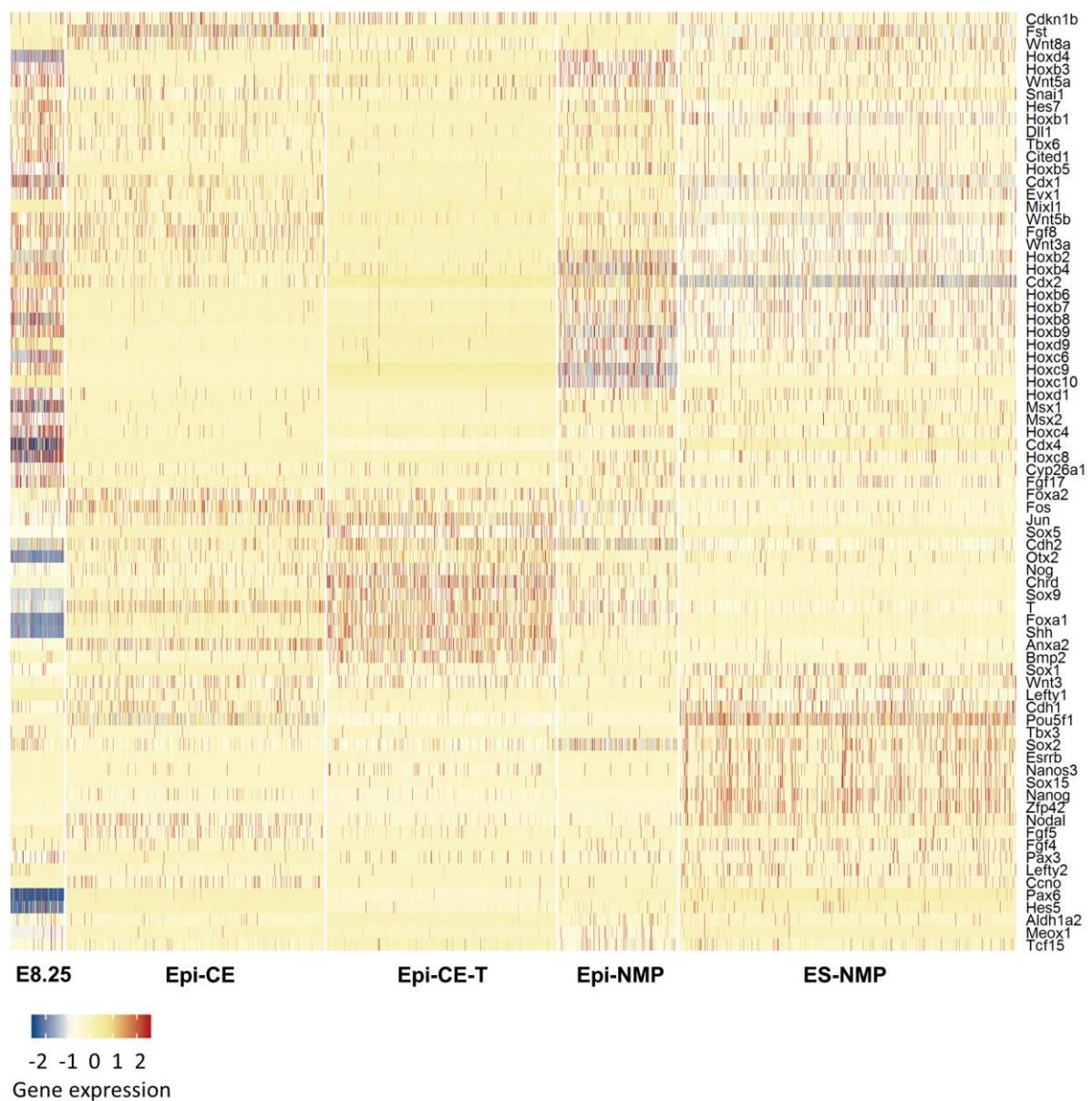


Figure S5. Visualization of gene expression in the *in vivo* and *in vitro* populations.

Expression of chosen marker genes of pluripotent state, CE (E7.5), CLE, neural, mesoderm and the node along the different samples.

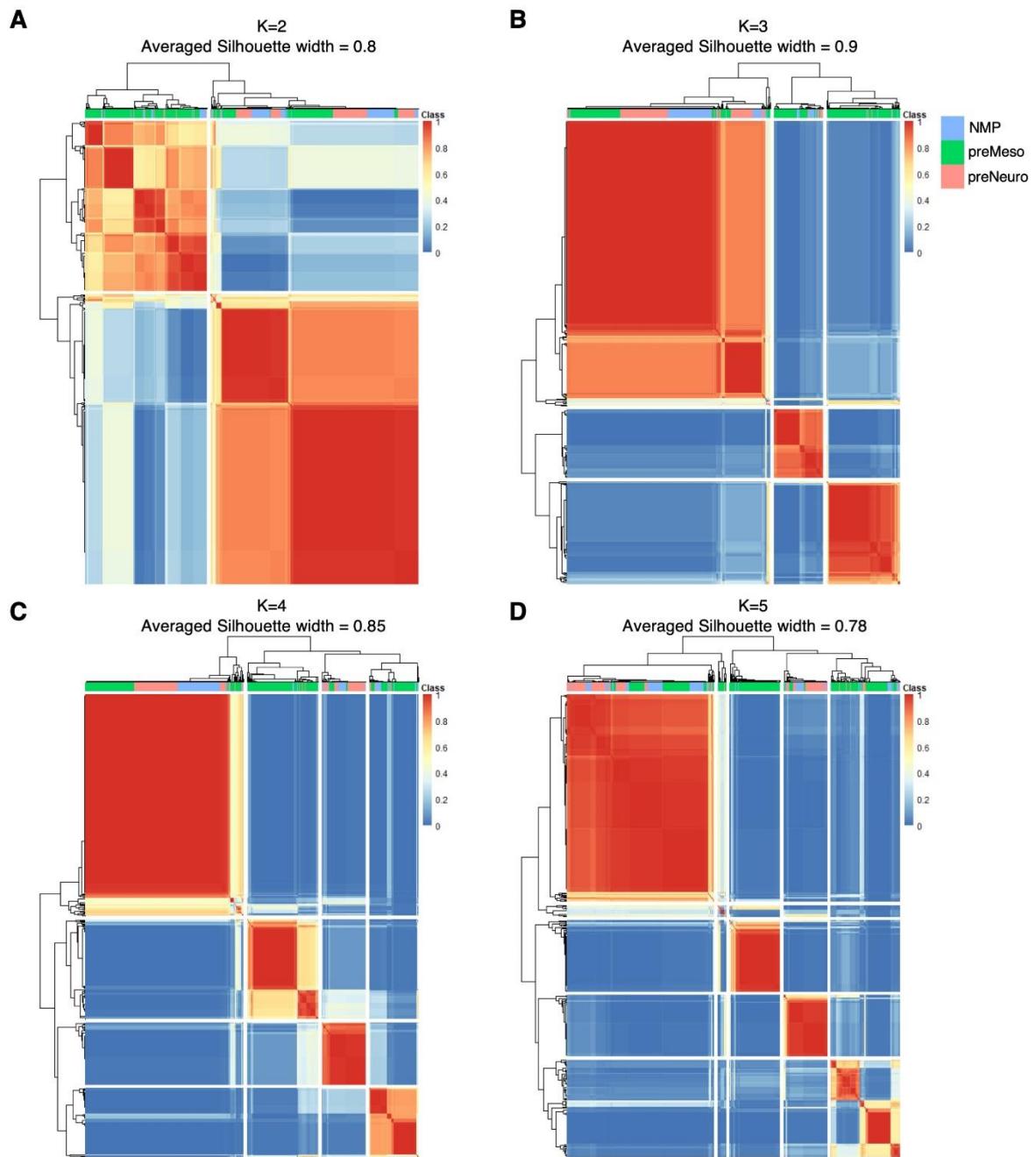


Figure S6. The averaged Silhouette width and the consensus matrices obtained for the embryo CLE for number of clusters: A. k = 2; B. k = 3; C. k = 4 and D. k = 5. Cells that were defined as NMP candidates, mesodermal progenitors (preMeso) and neural progenitors (preNeuro) are marked in blue, green and pink respectively.

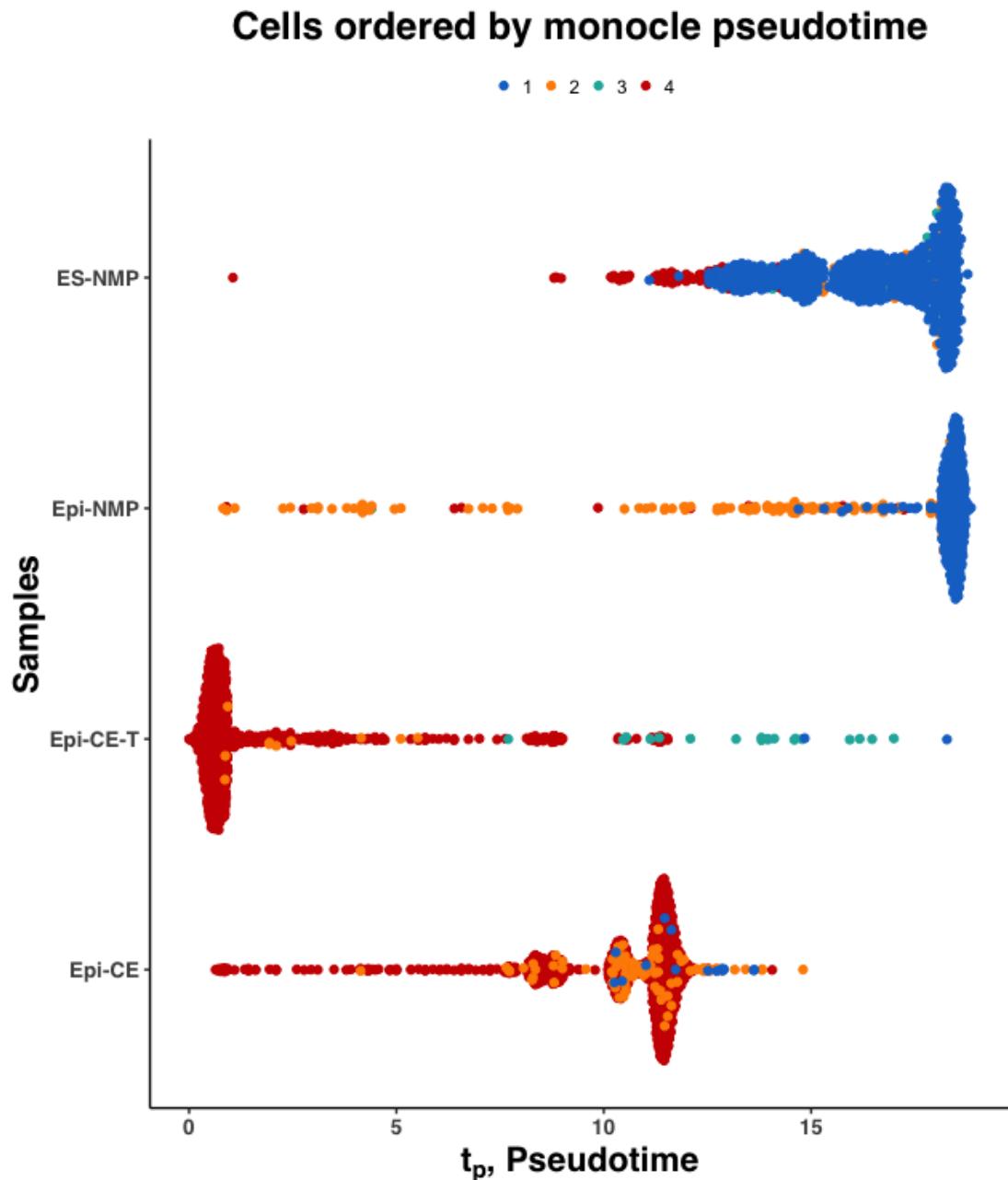


Figure S7. Monocle Pseudotemporal order of the *in vitro* cells. The cells are ordered along the pseudotime in the x-axis for every *in vitro* sample in the y-axis, where the cells that were classified to the 4 classes are coloured.

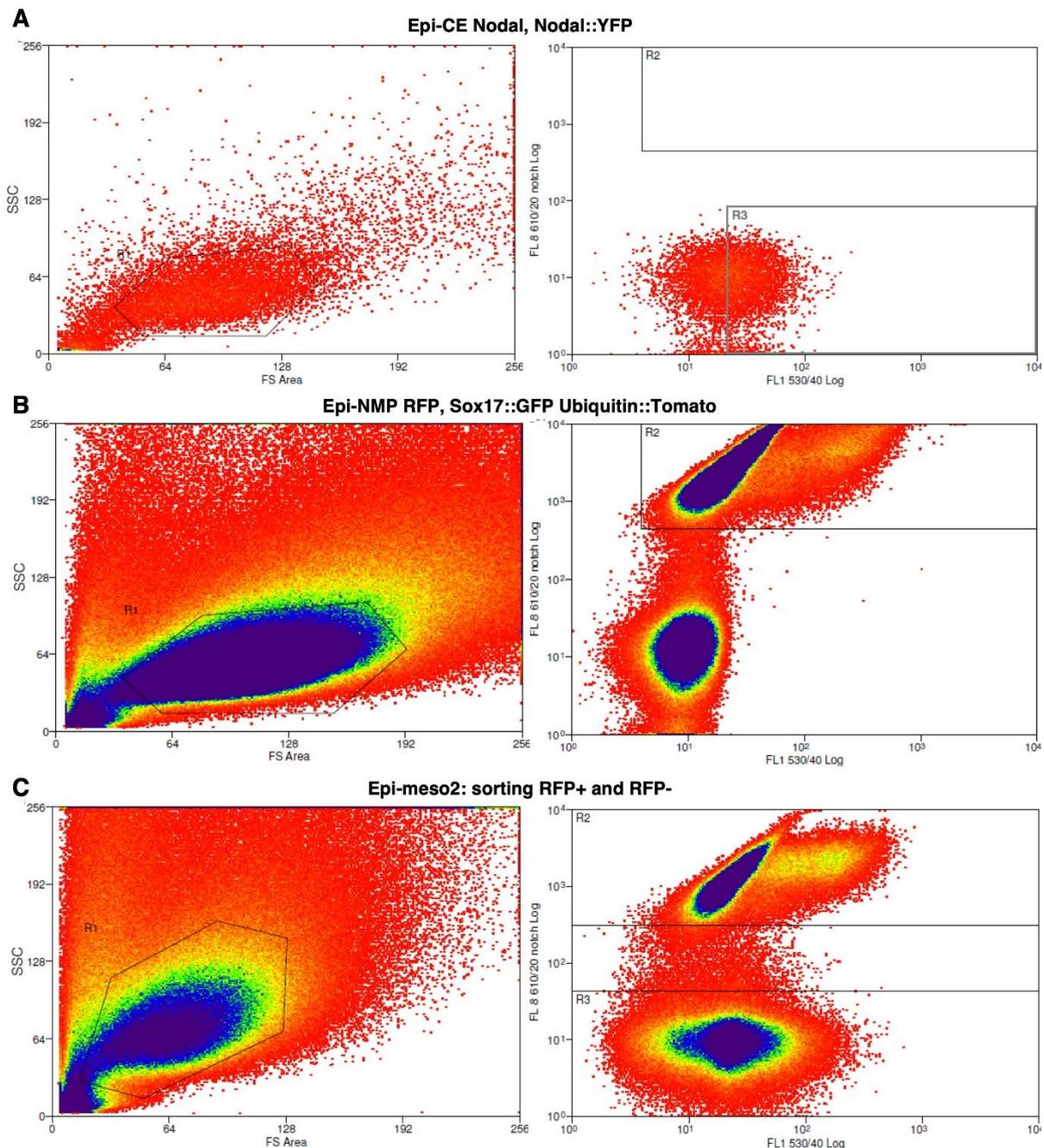


Figure S8. Node like cell with Epi-NMP mixture and sorting. Cells were sorted using a MoFlo sorter (Beckman Coulter). Single cells were collected according to the plot of SSC (side scattered light) versus FC (forward scattered light) Area (SSC versus FS area plots). The lasers that were used are 647nm laser with emission filter of 610/20 to collect the RFP positive cells and 488nm laser with emission filter of 530/40 to collect the YFP positive cells (FL8 610/20 versus FL1 530/40 log scale plots). **A.** Epi-CE Nodal sample composed of Nodal::YFP cells that were cultured under the Epi-CE protocol. Only the YFP positive cells (R3 rectangle) were used to make the mixture of Epi-meso2 (see Materials and Methods). **B.** Epi-NMP RFP sample composed of Sox17::GFP Ubiquitin::Tomato cells, were culture under the Epi-NMP protocol. Only the RFP positive cells (R2 rectangle) were used to make the Epi-meso2 mixture together with the Epi-CE Nodal YFP positive cells. **C.** After growing the mixture of Epi-meso2 for 4 days the cells were sorted based on their RFP fluorescence: RFP positive cells (R2 rectangle, sample named EM2-RFP+4d) and RFP negative cells (R3 rectangle, sample named EM2-RFP-4d). The sorted cells were quantified for their RNA of a chosen set of genes using RT-qPCR technique (Materials and Methods).

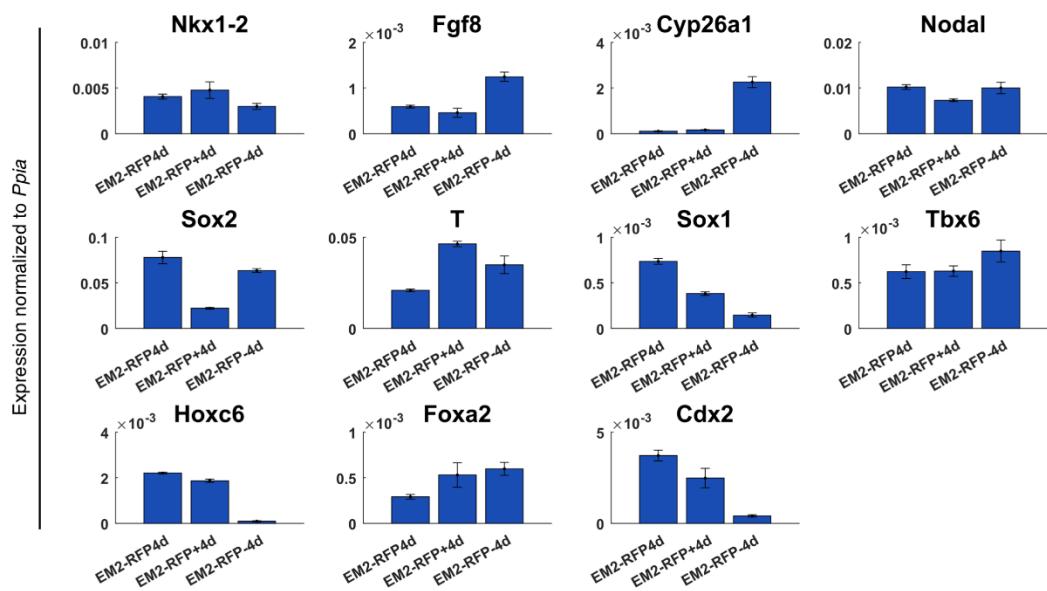


Table S1 - Genes expression in the mouse embryo

genes	Expression in the embryo E7.5 or E8.5	Category or pattern of expression	References
Nanog	-	Pluripotency and germ cells	(Chambers et al., 2003; Chambers et al., 2007)
Rex1	-	Pluripotency	(Toyooka et al., 2008)
Esrrb	-	Pluripotency	(Festuccia et al., 2012; Papp and Plath, 2012)
Fgf4	CE E7.5 and some expression in the CLE E8.5/9: the expression moves from the primitive streak to a region of the dorsal CE	Pluripotency and expression in the streak	(Niswander and Martin, 1992; Wright et al., 2003)
Sox15	-	Pluripotency	(Maruyama et al., 2005)
Nanos	-	Germ cells	(Tsuda et al., 2003)
Tbx3	-	Pluripotency and germ cells	(Pontecorvi et al., 2008; Russell et al., 2015)
Anxa2		Early epiblast	(Wang et al., 2015)
Fgf5	CE E7.5	Early epiblast	(Hebert et al., 1991; Khoa et al., 2016)
Otx2	CE E7.5	Early epiblast, primitive streak and node	(Acampora et al., 2009; Cajal et al., 2012)
Cdh1	CE E7.5	Pluripotency and early epiblast	(Basilicata et al., 2016; Cano et al., 2000; De Vries et al., 2004; Redmer et al., 2011)
Oct4 (Pou5f1)	CE E7.5	Pluripotency, early epiblast, caudal epiblast until E8.0	(Downs, 2008; Shi and Jin, 2010)
Pou3f1	Expression from E5.5 onward.	Early expression in the anterior epiblast, later expression in the nervous system (midbrain and forebrain) and in the peripheral nervous system. At	(Zhu et al., 2014)

		E8.0 <i>Pouu3f1</i> expression is restricted to the anterior neuroectoderm.	
Pou4f1	E9.5	Cardiac development and development of cochleovestibular ganglion neurons in mouse inner ear.	(Deng et al., 2014; Maskell et al., 2017)
Tcf15	-	Pre gastrulation, primed pluripotency and somites.	(Davies et al., 2013)
Cdh2	CLE E8.5/9	Primitive streak and neural progenitors	(Basilicata et al., 2016)
Cdx1, Cdx2, Cdx4	Both in CE E7.5 and CLE E8.5/9	Caudal epiblast and tail but (also posterior endoderm)	(Deschamps and van Nes, 2005)
Sox2	Both in CE E7.5 and CLE E8.5/9	Pluripotency, neural progenitors	(Henrique et al., 2015)
T	Both in CE E7.5 and CLE E8.5/9	Primitive Streak, caudal epiblast, tail bud and node.	(Henrique et al., 2015)
Cyp26a1	CLE E8.5/9	Caudal epiblast and tail bud from E8.0	(Sakai et al., 2001; Sirbu and Duester, 2006)
Nkx1-2	Both in CE E7.5 and CLE E8.5/9	Caudal epiblast and tail bud from E8.0; preneural spinal cord	(Henrique et al., 2015; Schubert et al., 1995)
Fgf8	Both in CE E7.5 and CLE E8.5/9	Caudal epiblast and tail bud	(Cunningham et al., 2015; Dunty et al., 2008; Sirbu and Duester, 2006)
Epha5	CLE E8.5/9	At E9.0 expression in the tail bud.	(Cooper et al., 2009)
Fos, Jun		Not known	
Fst1	CE E7.5	It is turned off in the CLE with the onset of somitogenesis.	(Cunningham et al., 2016)
Tbx6	Starts at CE E7.5	Paraxial mesoderm	(Chalamalasetty et al., 2011; Chalamalasetty et al., 2014; Dunty et

			al., 2008; White et al., 2003)
Meox1	CLE E8.5/9	Presomitic and somatic mesoderm	(Jukkola et al., 2005)
Aldh1a2	Both in CE E7.5 and CLE E8.5/9	Paraxial mesoderm at E7.5, presomitic and somatic mesoderm.	(Duester, 2008; Sirbu and Duester, 2006)
Cited1	CLE E8.5/9	Mesoderm. Early kidney development, progenitors of the heart, limb, axial skeleton and placenta.	(Boyle et al., 2007; Dunwoodie et al., 1998; Rodriguez et al., 2004)
Mesp1	From gastrulation E6.5 onwards.	Initially Mesp1 was expressed at the onset of gastrulation in the primitive streak and then in cardiac mesoderm. At E7.5 expression in the base of the allantois. At E8.0 Mesp1 is expressed on both sides of the node.	(Saga et al., 1996; Saga et al., 1997)
Mesp2	Initially detected at E8.0	At E8.0 expression was detected on both sides of the node at the same locations as for Mesp1. From E8.5 expressed in the PSM until E12.5.	(Saga et al., 1997)
Ifitm1	CLE E8.5/9	At E7.5 expression in the allantois. At E8.5 expression in the brain and in the neural tube. At E9.5 expression in the brain, primordial gut, the somites and the PSM. Expression in primordial germ cells (PGCs).	(Klymiuk et al., 2012; Lange et al., 2003)

Fn1		At E10 fibronectin matrix enriched at intersomitic borders.	(Bajanca et al., 2004)
Snai1	CE E7.5	EMT.	(Carver et al., 2001)
Bmp2	Both in CE E7.5 and CLE E8.5/9	IM, extra embryonic tissue (yolk sac and allantois), at E8.25 expression in the neural fold and LPM.	(Danesh et al., 2009)
Bmp4	Both in CE E7.5 and CLE E8.5/9	embryonic tissue (yolk sac and allantois), posterior part of the embryo, at E8.25 high expression in the posterior LPM, neural tube and in the caudal mesoderm of the tail.	(Danesh et al., 2009; Lawson et al., 1999; Zakin and De Robertis, 2004)
Evx1	Both in CE E7.5 and CLE E8.5/9	Posterior primitive streak. At E8.5 expression in the tail bud and in the allantois.	(Cambray and Wilson, 2007; Kojima et al., 2014; Schubert et al., 1995)
Mixl1	CE E7.5 and E8.5/9	Primitive streak, progenitors of endoderm. At E8.5 some expression in the tip of the tail bud and in the allantois, also expression in the crown cells of the node.	(Dunty et al., 2014; Hart et al., 2002; Kojima et al., 2014; Pearce and Evans, 1999; Robb et al., 2000; Wolfe and Downs, 2014)
Foxb1	E8.5	At E8.5 expression in the somites, presomitic mesoderm and in the neural plate	(Zhao et al., 2007).
Fgf3	Both in CE E7.5 and CLE E8.5/9	Fgf3 together with Fgf8 and Fgf17 are expressed in the primitive streak, PSM and tail bud. Later (E9.5) expressed in the ectoderm	(Bachler and Neubüser, 2001; Wahl et al., 2007)

		covering the midfacial region.	
Fgf9	Detected from E9.5.	At E9.5 together with Fgf3, Fgf8 and Fgf17, is expressed in the ectoderm covering the midfacial region. At E10.5 expressed the epithelium of the developing lung.	(Bachler and Neubüser, 2001; del Moral et al., 2006)
Fgf15	Expression detected at E9.5	Expressed in the developing CNS	(Fischer et al., 2011)
Fgf17	Low expression in the primitive streak at E7.5 and in the CLE E8.5	expressed in prestreak- and streak-stage embryos. Lower expression than Fgf8 in the posterior primitive streak and then in the presumptive mid/hindbrain junction, presomitic mesoderm and developing heart.	(Maruoka et al., 1998; Sun et al., 1999)
Msx1, Msx2	Starts at E7.5	At E7.5 expression in the embryonic and extraembryonic mesoderm (including the allantois), at E8.0 expression in the LPM and in the visceral endoderm together with the primordial germ cells, expression in the neural crest.	(Ishii et al., 2005; Sun et al., 2016)
Osr1	E8.5	At E8.5 expression in IM. Later expression in the developing heart, limb, lung, and craniofacial structures.	(Lan et al., 2011; Wang et al., 2005)
Phlda2	E8.0-8.5	yolk sac endoderm (expression in the	(Hou et al., 2007; Lefebvre, 2012)

		placenta), LPM and ventral definitive endoderm	
Krt18, Kr8		Extra embryonic endoderm	(Maurer et al., 2008)
Cldn6, Cldn7	At E6.5-E7.5 Cldn6 expressed in the epiblast excluding the primitive streak. At E8.5 expression begins to be restricted to the endoderm.	Definitive endoderm, kidney development	(Anderson et al., 2008; Khairallah et al., 2014)
Sparc	E9.0	At E9.0 present in extra embryonic tissues, later expressed in the parietal endoderm.	(Howe et al., 1988)
Pax6	E8.0 in the neuroepithelium	Spinal cord	(Bel-Vialar et al., 2007)
Pax3	First detected at E8.5	Expressed in the developing spinal cord, later (10-12 day) in the neural crest. Also, marker of limb muscle progenitor cells and expression in the dorsal neural tube.	(Goulding et al., 1991; Relaix et al., 2004)
Sox1	Detected at E7.5 and continue to be expressed at E9.5	Spinal cord: neural plate and neural tube	(Pevny et al., 1998; Wood and Episkopou, 1999)
Sox3	Sox2 and Sox3 expressed early in the embryo at E6.5 and at E8.5.	Both Sox2 and Sox3 are expressed throughout the epiblast. At E7.5 Sox3 is expressed in the posterior epiblast. At E8.5 Sox3 shows anteroposterior gradients of expression with bias to the posterior region.	(Wood and Episkopou, 1999)
Sox9	Expression starts at E9.5	Involved in chondrogenesis, formation of	(Barrionuevo et al., 2006; Wright et al., 1995)

		cardiac valves, and neural crest, testis and spinal cord development. Also expressed in the notochord.	
Sox5	Later expression than E9.0	Involved together with Sox6 in chondrogenesis.	(Akiyama et al., 2002)
Sox6	Later expression than E9.0	Expressed in wide range of tissues amongst them: the central nervous system (brain and spinal cord), somites, notochord, limb buds and liver.	(Akiyama et al., 2002; Hagiwara, 2011)
Sema3a	Expression detected at E9.5	Expression in the peripheral nervous system (PNS) and in the somites.	(Fujisawa and Kitsukawa, 1998; Schwarz et al., 2009)
Hes3	Both in CE E7.5 and CLE E8.5/9	At E7.5 expressed in the primitive streak, at E8.5 expression in the hindbrain, the neural plate and primitive streak.	(Lobe, 1997)
Hes5	First detected at E8-8.5	Neuroepithelium: midbrain, hindbrain and neural tube.	(Hitoshi et al., 2011)
Hes7	CLE E8.5	Expression in the PSM.	(Bessho et al., 2001a; Bessho et al., 2001b)
Ptn		Expressed in the central and peripheral nervous system	(Rosenfield et al., 2012)
Crabp2		At E10.5 can be detected in the spinal cord.	(Colbert et al., 1995; Duester, 2008)
Wnt3	First expressed at E5.5 and at E7.5 expression is evident in the epiblast.	Expression in the posterior visceral endoderm (E5.5) and later in the adjacent posterior epiblast.	(Tortelote et al., 2013)(Stuckey et al., 2011)
Wnt3a, Wnt8a	Both in CE E7.5 and CLE E8.5/9	Caudal epiblast and tail bud.	(Cambray and Wilson, 2007; Cunningham et al., 2015; Girós et al.,

			2011; Parr Brian A. et al., 1993)
Wnt5a	CLE E8.5/9	Caudal epiblast and tail bud.	(Yamaguchi et al., 1999)
Nodal	CE E7.5	Expression in the node.	(Lu and Robertson, 2004)
Foxa2	CE E7.5	Marker of the node and anterior mesendoderm. Also expressed in the extraembryonic tissue.	(Kojima et al., 2014; Lu and Robertson, 2004)
Ccno	CE E7.5-E8.0	Expression in the node and in the posterior tip of the embryo.	(Funk et al., 2015)
Zic1, Zic2, Zic3	From E8.0 to E16.5. Expressed in the CLE.	Expression in the CNS, somites, limb (not Zic1) and eye. Around E8.0, all the three expressed in the dorsal spinal cord and in the somites, Zic1 is evenly distributed along the AP axis but no expression was detected in the tail bud. Zic2 and Zic3 are highly expressed in the head and in the tail bud, but weakly in the trunk.	(Nagai et al., 1997)
Hox5-10	CLE E8.5 – E10.5	Anterior – posterior patterning, specify segment identity. Expression from the last cervical vertebrae (ribs) till the first sacral vertebrae (pelvis).	(Myers, 2008)
Hoxc10	Expression at E10.5		(Lee et al., 1999)

Table S2 – List of genes that were used to cluster the embryo cells (attached xls file).

[Click here to Download Table S2](#)

Table S3 – The marker genes of the 4 clusters of the CLE embryo data (attached xls file).

[Click here to Download Table S3](#)

Table S4 – The 82 informative genes that were used to build the SVM and classify the cells to the 4 clusters

[Click here to Download Table S4](#)

Table S5 – The highly expressed genes in the 2 pseudotime ranges of class 4 (attached xls file).

[Click here to Download Table S5](#)

Table S6 – Primer sequences used for qRT-PCR.

Gene	Forward primer sequence	Reverse primer sequence
1 <i>Cdx2</i>	TCCTGCTGACTGCTTCTGA	CCCTTCCTGATTGTGGAGA
2 <i>Cyp26a1</i>	TCTGGGACCTGTACTGTGTGA	AAGCCGTATTCCTGCGCTT
3 <i>Fgf8</i>	AGGACTGCGTATTACAGAGAT	CATGTACCAGCCCTCGTACT
4 <i>Foxa2</i>	CATTACGCCTCAACCACCC	GGTAGTGCATGACCTGTTCG
5 <i>Hoxc6</i>	CCCTCTTCTCCCTTGCTC	CCACGTCTGACTCCCTGTTT
6 <i>Nkx1-2</i>	ACAACCACACAAGCCACTGA	CCATCCTGGAAACCCTTATT
7 <i>Nodal</i>	AGCCACTGTCCAGTTCTCCAG	GTGTCTGCCAAGCATACATCTC
8 <i>Sox1</i>	AGACAGCGTGCCTTGATT	TGGGATAAGACCTGGGTGAG
9 <i>Sox2</i>	CATGAGAGCAAGTACTGGCAAG	CCAACGATATCAACCTGCATGG
10 <i>T</i>	CTGGGAGCTCAGTTCTTCG	GTCCACGAGGCTATGAGGAG
11 <i>Tbx6</i>	CCAGAACCTAGGATCACACA	CCCGAAGTTCTCTTCACA

Reference

- Acampora, D., Di Giovannantonio, L. G., Di Salvio, M., Mancuso, P. and Simeone, A.** (2009). Selective inactivation of Otx2 mRNA isoforms reveals isoform-specific requirement for visceral endoderm anteriorization and head morphogenesis and highlights cell diversity in the visceral endoderm. *Mech. Dev.* **126**, 882–897.
- Akiyama, H., Chaboissier, M.-C., Martin, J. F., Schedl, A. and de Crombrugghe, B.** (2002). The transcription factor Sox9 has essential roles in successive steps of the chondrocyte differentiation pathway and is required for expression of Sox5 and Sox6. *Genes Dev.* **16**, 2813–28.
- Anderson, W. J., Zhou, Q., Alcalde, V., Kaneko, O. F., Blank, L. J., Sherwood, R. I., Guseh, J. S., Rajagopal, J. and Melton, D. A.** (2008). Genetic targeting of the endoderm with claudin-6CreER. *Dev. Dyn.* **237**, 504–512.
- Bachler, M. and Neubüser, A.** (2001). Expression of members of the Fgf family and their receptors during midfacial development. *Mech. Dev.* **100**, 313–316.
- Bajanca, F., Luz, M., Duxson, M. J. and Thorsteinsdóttir, S.** (2004). Integrins in the mouse myotome: Developmental changes and differences between the epaxial and hypaxial lineage. *Dev. Dyn.* **231**, 402–415.
- Barrientos, F., Taketo, M. M., Scherer, G. and Kispert, A.** (2006). Sox9 is required for notochord maintenance in mice. *Dev. Biol.* **295**, 128–140.
- Basilicata, M. F., Frank, M., Solter, D., Brabletz, T. and Stemmler, M. P.** (2016). Inappropriate cadherin switching in the mouse epiblast compromises proper signaling between the epiblast and the extraembryonic ectoderm during gastrulation. *Sci. Rep.* **6**, 26562.
- Bel-Vialar, S., Medevielle, F. and Pituello, F.** (2007). The on/off of Pax6 controls the tempo of neuronal differentiation in the developing spinal cord. *Dev. Biol.* **305**, 659–673.
- Bessho, Y., Miyoshi, G., Sakata, R. and Kageyama, R.** (2001a). Hes7: a bHLH-type repressor gene regulated by Notch and expressed in the presomitic mesoderm. *Genes to Cells* **6**, 175–185.
- Bessho, Y., Sakata, R., Komatsu, S., Shiota, K., Yamada, S. and Kageyama, R.** (2001b). Dynamic expression and essential functions of Hes7 in somite segmentation. *Genes Dev.* **15**, 2642–7.
- Boyle, S., Shioda, T., Perantoni, A. O. and de Caestecker, M.** (2007). Cited1 and Cited2 are differentially expressed in the developing kidney but are not required for nephrogenesis. *Dev. Dyn.* **236**, 2321–2330.
- Cajal, M., Lawson, K. A., Hill, B., Moreau, A., Rao, J., Ross, A., Collignon, J., Camus, A., Simeone, A. and Levi, G.** (2012). Clonal and molecular analysis of the prospective anterior neural boundary in the mouse embryo. *Development* **139**, 423–36.
- Cambray, N. and Wilson, V.** (2007). Two distinct sources for a population of maturing axial progenitors. *Development* **134**, 2829–2840.
- Cano, A., Pérez-Moreno, M. A., Rodrigo, I., Locascio, A., Blanco, M. J., del Barrio, M. G., Portillo, F. and Nieto, M. A.** (2000). The transcription factor Snail controls epithelial–mesenchymal transitions by repressing E-cadherin expression. *Nat. Cell Biol.* **2**, 76–83.

- Carver, E. A., Jiang, R., Lan, Y., Oram, K. F. and Gridley, T.** (2001). The mouse snail gene encodes a key regulator of the epithelial-mesenchymal transition. *Mol. Cell. Biol.* **21**, 8184–8.
- Chalamalasetty, R. B., Dnty, W. C., Biris, K. K., Ajima, R., Iacovino, M., Beisaw, A., Feigenbaum, L., Chapman, D. L., Yoon, J. K., Kyba, M., et al.** (2011). The Wnt3a/β-catenin target gene Mesogenin1 controls the segmentation clock by activating a Notch signalling program. *Nat. Commun.* **2**, 390.
- Chalamalasetty, R. B., Garriock, R. J., Dnty, W. C., Kennedy, M. W., Jailwala, P., Si, H. and Yamaguchi, T. P.** (2014). Mesogenin 1 is a master regulator of paraxial presomitic mesoderm differentiation. *Development* **141**, 4285–97.
- Chambers, I., Colby, D., Robertson, M., Nichols, J., Lee, S., Tweedie, S. and Smith, A.** (2003). Functional Expression Cloning of Nanog, a Pluripotency Sustaining Factor in Embryonic Stem Cells. *Cell* **113**, 643–655.
- Chambers, I., Silva, J., Colby, D., Nichols, J., Nijmeijer, B., Robertson, M., Vrana, J., Jones, K., Grotewold, L. and Smith, A.** (2007). Nanog safeguards pluripotency and mediates germline development. *Nature* **450**, 1230–1234.
- Colbert, M. C., Rubin, W. W., Linney, E. and LaMantia, A.-S.** (1995). Retinoid signaling and the generation of regional and cellular diversity in the embryonic mouse spinal cord. *Dev. Dyn.* **204**, 1–12.
- Cooper, M. A., Crockett, D. P., Nowakowski, R. S., Gale, N. W. and Zhou, R.** (2009). Distribution of EphA5 receptor protein in the developing and adult mouse nervous system. *J. Comp. Neurol.* **514**, 310–28.
- Cunningham, T. J., Kumar, S., Yamaguchi, T. P. and Duester, G.** (2015). Wnt8a and Wnt3a cooperate in the axial stem cell niche to promote mammalian body axis extension. *Dev. Dyn.* **244**, 797–807.
- Cunningham, T. J., Colas, A. and Duester, G.** (2016). Early molecular events during retinoic acid induced differentiation of neuromesodermal progenitors. *Biol. Open* **5**, bio.020891.
- Danesh, S. M., Villasenor, A., Chong, D., Soukup, C. and Cleaver, O.** (2009). BMP and BMP receptor expression during murine organogenesis. *Gene Expr. Patterns* **9**, 255–65.
- Davies, O. R., Lin, C.-Y., Radzisheuskaya, A., Zhou, X., Taube, J., Blin, G., Waterhouse, A., Smith, A. J. H. and Lowell, S.** (2013). Tcf15 primes pluripotent cells for differentiation. *Cell Rep.* **3**, 472–84.
- De Vries, W. N., Evsikov, A. V., Haac, B. E., Fancher, K. S., Holbrook, A. E., Kemler, R., Solter, D. and Knowles, B. B.** (2004). Maternal beta-catenin and E-cadherin in mouse development. *Development* **131**, 4435–45.
- del Moral, P.-M., De Langhe, S. P., Sala, F. G., Veltmaat, J. M., Tefft, D., Wang, K., Warburton, D. and Bellusci, S.** (2006). Differential role of FGF9 on epithelium and mesenchyme in mouse embryonic lung. *Dev. Biol.* **293**, 77–89.
- Deng, M., Yang, H., Xie, X., Liang, G. and Gan, L.** (2014). Comparative expression analysis of POU4F1, POU4F2 and ISL1 in developing mouse cochleovestibular ganglion neurons. *Gene Expr. Patterns* **15**, 31–7.
- Deschamps, J. and van Nes, J.** (2005). Developmental regulation of the Hox genes during axial morphogenesis in the mouse. *Development* **132**, 2931–42.

- Downs, K. M.** (2008). Systematic localization of Oct-3/4 to the gastrulating mouse conceptus suggests manifold roles in mammalian development. *Dev. Dyn.* **237**, 464–475.
- Duester, G.** (2008). Retinoic Acid Synthesis and Signaling during Early Organogenesis. *Cell* **134**, 921–931.
- Dunty, W. C., Biris, K. K., Chalamalasetty, R. B., Taketo, M. M., Lewandoski, M. and Yamaguchi, T. P.** (2008). Wnt3a/beta-catenin signaling controls posterior body development by coordinating mesoderm formation and segmentation. *Development* **135**, 85–94.
- Dunty, W. C., Kennedy, M. W. L., Chalamalasetty, R. B., Campbell, K. and Yamaguchi, T. P.** (2014). Transcriptional profiling of Wnt3a mutants identifies Sp transcription factors as essential effectors of the Wnt/β-catenin pathway in neuromesodermal stem cells. *PLoS One* **9**, e87018.
- Dunwoodie, S. L., Rodriguez, T. A. and Beddington, R. S. P.** (1998). Msg1 and Mrg1, founding members of a gene family, show distinct patterns of gene expression during mouse embryogenesis. *Mech. Dev.* **72**, 27–40.
- Edri, S., Hayward, P., Baillie-Johnson, P., Steventon, B. and Martinez Arias, A.** (2018). An Epiblast Stem Cell derived multipotent progenitor population for axial extension. *bioRxiv* 242461.
- Festuccia, N., Osorno, R., Halbritter, F., Karwacki-Neisius, V., Navarro, P., Colby, D., Wong, F., Yates, A., Tomlinson, S. R. and Chambers, I.** (2012). Esrrb is a direct Nanog target gene that can substitute for Nanog function in pluripotent cells. *Cell Stem Cell* **11**, 477–90.
- Fischer, T., Faus-Kessler, T., Welzl, G., Simeone, A., Wurst, W. and Prakash, N.** (2011). Fgf15-mediated control of neurogenic and proneural gene expression regulates dorsal midbrain neurogenesis. *Dev. Biol.* **350**, 496–510.
- Fujisawa, H. and Kitsukawa, T.** (1998). Receptors for collapsin/semaphorins. *Curr. Opin. Neurobiol.* **8**, 587–592.
- Funk, M. C., Bera, A. N., Menchen, T., Kuales, G., Thriene, K., Lienkamp, S. S., Dengjel, J., Omran, H., Frank, M. and Arnold, S. J.** (2015). Cyclin O (Ccno) functions during deuterosome-mediated centriole amplification of multiciliated cells. *EMBO J.* **34**, 1078–89.
- Girós, A., Grgur, K., Gossler, A. and Costell, M.** (2011). α5β1 Integrin-Mediated Adhesion to Fibronectin Is Required for Axis Elongation and Somitogenesis in Mice. *PLoS One* **6**, e22002.
- Goulding, M. D., Chalepakis, G., Deutsch, U., Erselius, J. R. and Gruss, P.** (1991). Pax-3, a novel murine DNA binding protein expressed during early neurogenesis. *EMBO J.* **10**, 1135–47.
- Haghverdi, L., Lun, A. T. L., Morgan, M. D. and Marioni, J. C.** (2018). Batch effects in single-cell RNA-sequencing data are corrected by matching mutual nearest neighbors. *Nat. Biotechnol.* **36**, 421–427.
- Hagiwara, N.** (2011). Sox6, jack of all trades: a versatile regulatory protein in vertebrate development. *Dev. Dyn.* **240**, 1311–21.
- Hart, A. H., Hartley, L., Sourris, K., Stadler, E. S., Li, R., Stanley, E. G., Tam, P. P. L., Elefanty, A. G. and Robb, L.** (2002). Mixl1 is required for axial mesendoderm

- morphogenesis and patterning in the murine embryo. *Development* **129**, 3597–608.
- Hebert, J. M., Boyle, M. and Martin, G. R.** (1991). mRNA localization studies suggest that murine FGF-5 plays a role in gastrulation. *Development* **112**,
- Henrique, D., Abranches, E., Verrier, L. and Storey, K. G.** (2015). Neuromesodermal progenitors and the making of the spinal cord. *Development* **142**, 2864–2875.
- Hitoshi, S., Ishino, Y., Kumar, A., Jasmine, S., Tanaka, K. F., Kondo, T., Kato, S., Hosoya, T., Hotta, Y. and Ikenaka, K.** (2011). Mammalian Gcm genes induce Hes5 expression by active DNA demethylation and induce neural stem cells. *Nat. Neurosci.* **14**, 957–964.
- Hou, J., Charters, A. M., Lee, S. C., Zhao, Y., Wu, M. K., Jones, S. J., Marra, M. A. and Hoodless, P. A.** (2007). A systematic screen for genes expressed in definitive endoderm by Serial Analysis of Gene Expression (SAGE). *BMC Dev. Biol.* **7**, 92.
- Howe, C. C., Overton, G. C., Sawicki, J., Solter, D., Stein, P. and Strickland, S.** (1988). Expression of SPARC/osteonectin transcript in murine embryos and gonads. *Differentiation* **37**, 20–25.
- Ishii, M., Han, J., Yen, H.-Y., Sucov, H. M., Chai, Y. and Maxson, R. E.** (2005). Combined deficiencies of Msx1 and Msx2 cause impaired patterning and survival of the cranial neural crest. *Development* **132**, 4937–50.
- Jukkola, T., Trokovic, R., Maj, P., Lamberg, A., Mankoo, B., Pachnis, V., Savilahti, H. and Partanen, J.** (2005). Meox1Cre: A mouse line expressing Cre recombinase in somitic mesoderm. *genesis* **43**, 148–153.
- Khairallah, H., El Andalousi, J., Simard, A., Haddad, N., Chen, Y.-H., Hou, J., Ryan, A. K. and Gupta, I. R.** (2014). Claudin-7, -16, and -19 during mouse kidney development. *Tissue barriers* **2**, e964547.
- Khoa, L. T. P., Azami, T., Tsukiyama, T., Matsushita, J., Tsukiyama-Fujii, S., Takahashi, S. and Ema, M.** (2016). Visualization of the Epiblast and Visceral Endodermal Cells Using Fgf5-P2A-Venus BAC Transgenic Mice and Epiblast Stem Cells. *PLoS One* **11**, e0159246.
- Klymiuk, I., Kenner, L., Adler, T., Busch, D. H., Boersma, A., Irmler, M., Gailus-Durner, V., Fuchs, H., Leitner, N., Müller, M., et al.** (2012). In Vivo Functional Requirement of the Mouse Ifitm1 Gene for Germ Cell Development, Interferon Mediated Immune Response and Somitogenesis. *PLoS One* **7**, e44609.
- Kojima, Y., Kaufman-Francis, K., Studdert, J. B., Steiner, K. A., Power, M. D., Loebel, D. A. F., Jones, V., Hor, A., De Alencastro, G., Logan, G. J., et al.** (2014). The transcriptional and functional properties of mouse epiblast stem cells resemble the anterior primitive streak. *Cell Stem Cell* **14**, 107–120.
- Lan, Y., Liu, H., Ovitt, C. E. and Jiang, R.** (2011). Generation of Osr1 conditional mutant mice. *Genesis* **49**, 419–22.
- Lange, U. C., Saitou, M., Western, P. S., Barton, S. C. and Surani, M. A.** (2003). The fragilis interferon-inducible gene family of transmembrane proteins is associated with germ cell specification in mice. *BMC Dev. Biol.* **3**, 1.
- Lawson, K. A., Dunn, N. R., Roelen, B. A., Zeinstra, L. M., Davis, A. M., Wright, C. V., Korving, J. P. and Hogan, B. L.** (1999). Bmp4 is required for the generation of primordial germ cells in the mouse embryo. *Genes Dev.* **13**, 424–36.

- Lee, S.-J., McPherron, A. C. and Lawler, A. M.** (1999). Regulation of anterior/posterior patterning of the axial skeleton by growth/differentiation factor 11. *Nat. Genet.* **22**, 260–264.
- Lefebvre, L.** (2012). The placental imprintome and imprinted gene function in the trophoblast glycogen cell lineage. *Reprod. Biomed. Online* **25**, 44–57.
- Lobe, C. G.** (1997). Expression of the helix-loop-helix factor, Hes3, during embryo development suggests a role in early midbrain-hindbrain patterning. *Mech. Dev.* **62**, 227–237.
- Lu, C. C. and Robertson, E. J.** (2004). Multiple roles for Nodal in the epiblast of the mouse embryo in the establishment of anterior-posterior patterning. *Dev. Biol.* **273**, 149–159.
- Maruoka, Y., Ohbayashi, N., Hoshikawa, M., Itoh, N., Hogan, B. L. M. and Furuta, Y.** (1998). Comparison of the expression of three highly related genes, Fgf8, Fgf17 and Fgf18, in the mouse embryo. *Mech. Dev.* **74**, 175–177.
- Maruyama, M., Ichisaka, T., Nakagawa, M. and Yamanaka, S.** (2005). Differential roles for Sox15 and Sox2 in transcriptional control in mouse embryonic stem cells. *J. Biol. Chem.* **280**, 24371–9.
- Maskell, L. J., Qamar, K., Babakr, A. A., Hawkins, T. A., Heads, R. J. and Budhraam-Mahadeo, V. S.** (2017). Essential but partially redundant roles for POU4F1/Brn-3a and POU4F2/Brn-3b transcription factors in the developing heart. *Cell Death Dis.* **8**, e2861.
- Maurer, J., Nelson, B., Ceceña, G., Bajpai, R., Mercola, M., Terskikh, A. and Oshima, R. G.** (2008). Contrasting Expression of Keratins in Mouse and Human Embryonic Stem Cells. *PLoS One* **3**, e3451.
- Myers, P.** (2008). Hox Genes in Development: The Hox Code. *Nature*.
- Nagai, T., Aruga, J., Takada, S., Günther, T., Spörle, R., Schughart, K. and Mikoshiba, K.** (1997). The Expression of the Mouse Zic1, Zic2, and Zic3 Gene Suggests an Essential Role for Zic Genes in Body Pattern Formation. *Dev. Biol.* **182**, 299–313.
- Niswander, L. and Martin, G. R.** (1992). Fgf-4 expression during gastrulation, myogenesis, limb and tooth development in the mouse. *Development* **114**,
- Papp, B. and Plath, K.** (2012). Pluripotency re-centered around Esrrb. *EMBO J.* **31**, 4255–7.
- Parr Brian A., Shea Martin J., Vassileva Galya and McMahon Andrew P.** (1993). Mouse Wnt genes exhibit discrete domains of expression in the early embryonic CNS and limb buds. *Development* **119**, 247–261.
- Pearce, J. J. H. and Evans, M. J.** (1999). Mml, a mouse Mix-like gene expressed in the primitive streak. *Mech. Dev.* **87**, 189–192.
- Pevny, L. H., Sockanathan, S., Placzek, M. and Lovell-Badge, R.** (1998). A role for SOX1 in neural determination. *Development* **125**,
- Pontecorvi, M., Goding, C. R., Richardson, W. D. and Kessaris, N.** (2008). Expression of Tbx2 and Tbx3 in the developing hypothalamic–pituitary axis. *Gene Expr. Patterns* **8**, 411–417.
- Redmer, T., Diecke, S., Grigoryan, T., Quiroga-Negreira, A., Birchmeier, W. and Besser, D.** (2011). E-cadherin is crucial for embryonic stem cell pluripotency and can replace OCT4 during somatic cell reprogramming. *EMBO Rep.* **12**, 720–6.
- Relaix, F., Rocancourt, D., Mansouri, A. and Buckingham, M.** (2004). Divergent

- functions of murine Pax3 and Pax7 in limb muscle development. *Genes Dev.* **18**, 1088–105.
- Robb, L., Hartley, L., Begley, C. G., Brodnicki, T. C., Copeland, N. G., Gilbert, D. J., Jenkins, N. A. and Elefanty, A. G.** (2000). Cloning, expression analysis, and chromosomal localization of murine and human homologues of aXenopus Mix gene. *Dev. Dyn.* **219**, 497–504.
- Rodriguez, T. A., Sparrow, D. B., Scott, A. N., Withington, S. L., Preis, J. I., Michalicek, J., Clements, M., Tsang, T. E., Shioda, T., Beddington, R. S. P., et al.** (2004). Cited1 is required in trophoblasts for placental development and for embryo growth and survival. *Mol. Cell. Biol.* **24**, 228–44.
- Rosenfield, S. M., Bowden, E. T., Cohen-Missner, S., Gibby, K. A., Ory, V., Henke, R. T., Riegel, A. T. and Wellstein, A.** (2012). Pleiotrophin (PTN) expression and function and in the mouse mammary gland and mammary epithelial cells. *PLoS One* **7**, e47876.
- Russell, R., Ilg, M., Lin, Q., Wu, G., Lechel, A., Bergmann, W., Eiseler, T., Linta, L., Kumar P, P., Klingenstein, M., et al.** (2015). A Dynamic Role of TBX3 in the Pluripotency Circuitry. *Stem cell reports* **5**, 1155–1170.
- Saga, Y., Hata, N., Kobayashi, S., Magnuson, T., Seldin, M. F. and Taketo, M. M.** (1996). MesP1: a novel basic helix-loop-helix protein expressed in the nascent mesodermal cells during mouse gastrulation. *Development* **122**,
- Saga, Y., Hata, N., Koseki, H. and Taketo, M. M.** (1997). Mesp2: a novel mouse gene expressed in the presegmented mesoderm and essential for segmentation initiation. *Genes Dev.* **11**, 1827–39.
- Sakai, Y., Meno, C., Fujii, H., Nishino, J., Shiratori, H., Saijoh, Y., Rossant, J. and Hamada, H.** (2001). The retinoic acid-inactivating enzyme CYP26 is essential for establishing an uneven distribution of retinoic acid along the antero-posterior axis within the mouse embryo. *Genes Dev.* **15**, 213–25.
- Schubert, F. R., Fainsod, A., Gruenbaum, Y. and Gruss, P.** (1995). Expression of the novel murine homeobox gene Sax-1 in the developing nervous system. *Mech. Dev.* **51**, 99–114.
- Schwarz, Q., Maden, C. H., Vieira, J. M. and Ruhrberg, C.** (2009). Neuropilin 1 signaling guides neural crest cells to coordinate pathway choice with cell specification. *Proc. Natl. Acad. Sci. U. S. A.* **106**, 6164–9.
- Shi, G. and Jin, Y.** (2010). Role of Oct4 in maintaining and regaining stem cell pluripotency. *Stem Cell Res. Ther.* **1**, 39.
- Sirbu, I. O. and Duester, G.** (2006). Retinoic-acid signalling in node ectoderm and posterior neural plate directs left-right patterning of somitic mesoderm. *Nat. Cell Biol.* **8**, 271–277.
- Stuckey, D. W., Di Gregorio, A., Clements, M. and Rodriguez, T. A.** (2011). Correct Patterning of the Primitive Streak Requires the Anterior Visceral Endoderm. *PLoS One* **6**, e17620.
- Sun, X., Meyers, E. N., Lewandoski, M. and Martin, G. R.** (1999). Targeted disruption of Fgf8 causes failure of cell migration in the gastrulating mouse embryo. *Genes Dev.* **13**, 1834–46.
- Sun, J., Ting, M.-C., Ishii, M. and Maxson, R.** (2016). Msx1 and Msx2 function together in

- the regulation of primordial germ cell migration in the mouse. *Dev. Biol.* **417**, 11–24.
- Tortelote, G. G., Hernández-Hernández, J. M., Quaresma, A. J. C., Nickerson, J. A., Imbalzano, A. N. and Rivera-Pérez, J. A.** (2013). Wnt3 function in the epiblast is required for the maintenance but not the initiation of gastrulation in mice. *Dev. Biol.* **374**, 164–73.
- Toyooka, Y., Shimosato, D., Murakami, K., Takahashi, K. and Niwa, H.** (2008). Identification and characterization of subpopulations in undifferentiated ES cell culture. *Development* **135**, 909–18.
- Tsuda, M., Sasaoka, Y., Kiso, M., Abe, K., Haraguchi, S., Kobayashi, S. and Saga, Y.** (2003). Conserved Role of nanos Proteins in Germ Cell Development. *Science (80-.).* **301**, 1239–1241.
- Wahl, M. B., Deng, C., Lewandoski, M. and Pourquié, O.** (2007). FGF signaling acts upstream of the NOTCH and WNT signaling pathways to control segmentation clock oscillations in mouse somitogenesis. *Development* **134**, 4033–41.
- Wang, Q., Lan, Y., Cho, E.-S., Maltby, K. M. and Jiang, R.** (2005). Odd-skipped related 1 (Odd1) is an essential regulator of heart and urogenital development. *Dev. Biol.* **288**, 582–594.
- Wang, B., Ye, T.-M., Lee, K.-F., Chiu, P. C. N., Pang, R. T. K., Ng, E. H. Y. and Yeung, W. S. B.** (2015). Annexin A2 Acts as an Adhesion Molecule on the Endometrial Epithelium during Implantation in Mice. *PLoS One* **10**, e0139506.
- Weinreb, C., Wolock, S. and Klein, A. M.** (2017). SPRING: a kinetic interface for visualizing high dimensional single-cell expression data. *Bioinformatics*.
- White, P. H., Farkas, D. R., McFadden, E. E. and Chapman, D. L.** (2003). Defective somite patterning in mouse embryos with reduced levels of Tbx6. *Development* **130**, 1681–90.
- Wolfe, A. D. and Downs, K. M.** (2014). Mixl1 localizes to putative axial stem cell reservoirs and their posterior descendants in the mouse embryo. *Gene Expr. Patterns* **15**, 8–20.
- Wood, H. B. and Episkopou, V.** (1999). Comparative expression of the mouse Sox1, Sox2 and Sox3 genes from pre-gastrulation to early somite stages. *Mech. Dev.* **86**, 197–201.
- Wright, E., Hargrave, M. R., Christiansen, J., Cooper, L., Kun, J., Evans, T., Gangadharan, U., Greenfield, A. and Koopman, P.** (1995). The Sry-related gene Sox9 is expressed during chondrogenesis in mouse embryos. *Nat. Genet.* **9**, 15–20.
- Wright, T. J., Hatch, E. P., Karabagli, H., Karabagli, P., Schoenwolf, G. C. and Mansour, S. L.** (2003). Expression of mouse fibroblast growth factor and fibroblast growth factor receptor genes during early inner ear development. *Dev. Dyn.* **228**, 267–272.
- Yamaguchi, T. P., Bradley, A., McMahon, A. P. and Jones, S.** (1999). A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo. *Development* **126**, 1211–1223.
- Zakin, L. and De Robertis, E. M.** (2004). Inactivation of mouse Twisted gastrulation reveals its role in promoting Bmp4 activity during forebrain development. *Development* **131**, 413–24.
- Zhao, T., Zhou, X., Szabó, N., Leitges, M. and Alvarez-Bolado, G.** (2007). Foxb1-drivenCre expression in somites and the neuroepithelium of diencephalon, brainstem,

and spinal cord. *genesis* **45**, 781–787.

Zhu, Q., Song, L., Peng, G., Sun, N., Chen, J., Zhang, T., Sheng, N., Tang, W., Qian, C., Qiao, Y., et al. (2014). The transcription factor Pou3f1 promotes neural fate commitment via activation of neural lineage genes and inhibition of external signaling pathways. *Elife* **3**.