

Versican is crucial for the initiation of cardiovascular lumen development in medaka (*Oryzias latipes*)

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Supplementary figures

Fig. S1: Wild-type and *lht* mutant embryos are phenotypically indistinguishable. Phase contrast microscopic image showing representative WT and *lht* mutant embryos at 40 hpf (n = 10). Wild-type and mutant embryos were phenotypically similar at this stage. Scale bar: 0.2 mm (A–D). Heartbeat rate of WT and *lht* mutant medaka embryo, at 2 dpf and 3 dpf (n = 13). The WT and *lht* mutant medaka exhibited a similar heartbeat rate (E,F). Each bar represents the mean ± SEM from 13 individual samples. Student's *t*-test, ***P < 0.0001, N.S., not significant.

Fig. S2: Amino acid sequence alignment of medaka, human, and mouse versican. Amino acid sequence of medaka, human, and mouse versican aligned by the ClustalW program shown with their UniProt accession codes. The black box indicates the region of high homology and corresponds to the G1 and G3 domains. The rest of the amino acid sequence belongs to the G2 domain.

Fig. S3: Analysis of versican mRNA expression in medaka. RT-PCR and qPCR data for versican expression in various organs of adult medaka (A, B). Versican expression was prominent in the heart, eyes, gills, muscle, ovary, and testis. Whole-mount *in situ* hybridization for versican at multiple stages of embryonic development (C). Versican expression was prominent in the eye, heart tube, and ICM until 48 hpf (C; left and middle panels). At 72 hpf, the embryo shows strong versican expression in the gill region (C; right panel). Versican expression in the heart at various stages of cardiac development (D). (n > 10) (B). ICM, intermediate cell mass. Scale bar: 500 µm (C), 100 µm (D). Each error bar represents the SEM from 3 individual samples. β-Actin was used as endogenous reference gene to calculate ΔCT value. To calculate ΔΔCT, testis was selected randomly as reference sample.

Fig. S4: The 3' UTR sequence of the versican region of interest is conserved in all medaka strains. We used the same set of primers we designed for genotyping, and analysed the sequence for QURT1, HN1, Hd-rR, Kaga, and Ok-cab. The region of interest did not show any polymorphism (A). Western blot results for the versican expression in WT embryo, *lht* embryo, WT cell line transfected with either ctrl MO or versican MO (B). This result was obtained when a new lot of MC-955 (versican) antibody was used. Cell lines derived from WT medaka embryos were transfected with either ctrl or versican MO, and immunoassayed with versican G2 domain specific (MC-955) antibody (C). From three independent experiments, the total number of cells expressing versican around the cell boundary were counted using GFP

expression and nuclear staining by DAPI (D). Morpholino (MO) injection assays and quantification data (E–F). Versican-a MO was injected at the one-cell stage into embryos obtained from WT zebrafish. A morphant injected with versican-a MO (0.5 mM) phenocopies the *lht* mutant (C). Graphical representation of versican-a MO (0.5 mM) showing all the *lht* mutant phenotypes (D) ctrl: control. The results are represented as mean percentages of three independent experiment, mean \pm SD.

Fig. S5: Comparative study of mRNA and protein levels in WT and *lht* mutant medaka. Structural representation of the V0, V1, V2, and V3 versican isoforms (A). Versican mRNA expression data for WT and mutant embryos using qPCR (B). RT-PCR results show the comparison of mRNA expression levels at the exon-exon boundary of WT and the *lht* mutant (C). Primers were designed for exon boundaries. Coloured regions indicate the specific isoform exon-exon boundaries. RT-PCR results show the comparison of mRNA expression levels at the exon-exon boundaries of WT and the *lht* mutant ($n = 3$). Cell lines derived from WT and *lht* mutant embryos were immunoassayed with a versican G1 domain-specific antibody. In the WT-derived cell line, versican was expressed at the cell membrane (D; left panel). However, versican expression was absent at the cell membrane of the *lht* mutant-derived cell line (D; right panel). Each error bar represents the SEM from three individual samples. β -Actin was used as endogenous reference gene to calculate ΔCT value. To calculate $\Delta\Delta CT$, WT was selected as reference sample. Student's *t*-test, *** $P < 0.0001$, N.S., not significant.

Fig. S6: The *lht* mutant displays constriction of the outflow tract. Stereomicroscope images of embryos injected with fluorescence beads into the sinus venosus at 4 dpf. Panels A and B show the bright field images and panels C and D show the fluorescent images. The fluorescent beads injected into the heart (C) flowed into the vascular network of WT embryos (D), but pooled in the heart tube of *lht* mutant embryos that lacked blood circulation. Scale bar: 200 μ m (A–D)

Fig. S7: Versican loss of function did not alter the chamber-specific differentiation of cardiomyocytes. *In situ* hybridization for the cardiac-specific genes *cmlc2* (A, B), *amhc* (C), and *vmhc* (D) ($n > 10$) in the heart at 3 dpf. Red circle marks the presence of *cmlc2*-positive cells localized near the outflow tract. Scale bar: 100 μ m (A–D).

Fig. S8: The heart of the *lht* mutant medaka embryo did not show any apoptosis at 50 hpf. Analysis of apoptosis in live embryos with acridine orange staining examined with a

KEYENCE fluorescence microscope. We did not find any apoptotic cells in the *lht* mutant heart. The white arrow indicates acridine orange positive apoptotic cells in the head region of WT and *lht* mutants.

Fig. S9: The heart of the *lht* mutant medaka embryo did not show any differences in cardiomyocyte proliferation compared to that of WT at 50 hpf. Analysis of cardiomyocyte proliferation by whole-mount immunohistochemistry at 50 hpf, using a phosphohistone H3 antibody. The z stack images were obtained by confocal microscopy (A, B). There was no significant difference in the rate of cardiomyocyte proliferation between WT and *lht* mutant embryos (C). White arrows indicate the phosphohistone H3 positive cells in the heart. Each bar represents the mean \pm SEM from nine individual samples. Student's *t*-test, ****P* < 0.0001, N.S., not significant.

Fig. S10: The *lht* mutant shows disrupted endothelial alignment. Time-lapse microscopic images of WT ($n > 15$) (A–D) and *lht* mutant embryos ($n = 5$) (E–H) at 38, 42, 46, and 50 hpf. Embryos obtained from a Tg(fli1:GFP):*lht* heterozygous cross were dechorionated and embedded in 1% low-melt agarose. These embryos were kept under the microscope for the duration of the time-lapse imaging from 32 to 50 hpf. Images were taken at 20 min intervals. The temperature was maintained at 28.5°C throughout the experiment. A, atrium; CCV, common cardinal vein; V, ventricle. Scale bar: 100 μ m.

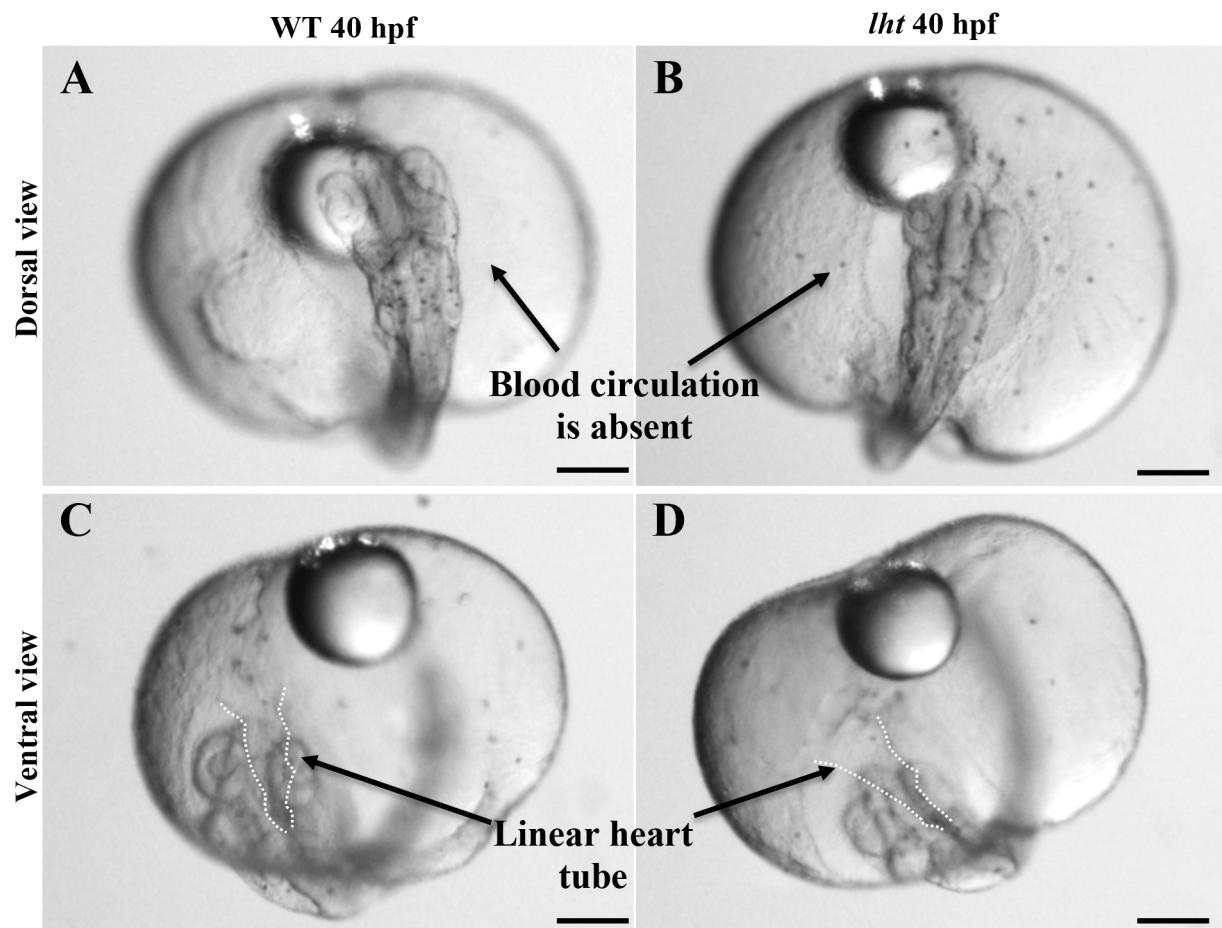
Fig. S11: The expression of etgase is specific to yolk sac blood vessels. Bright field image of etgase mRNA expression in WT embryos, after removal of the yolk, using *in-situ* hybridization. etgase was expressed exclusively in yolk sac blood vessels.

Fig. S12: Mutation in 3'UTR in the *lht* mutants creates putative binding sites for microRNA mmu-miR 871-3p.

Fig. S13: Full-length gels of Figure 2F. Note: WT: wild type, het: heterozygous, *lht*: *lht* homozygous mutant.

Fig. S14: Full-length gels of Figure S4B. Note: WT: wild type, *lht*: *lht* homozygous mutant, Ctrl MO: medaka fibroblasts nucleofected with control morpholino, versican MO: medaka fibroblasts nucleofected with versican morpholino.

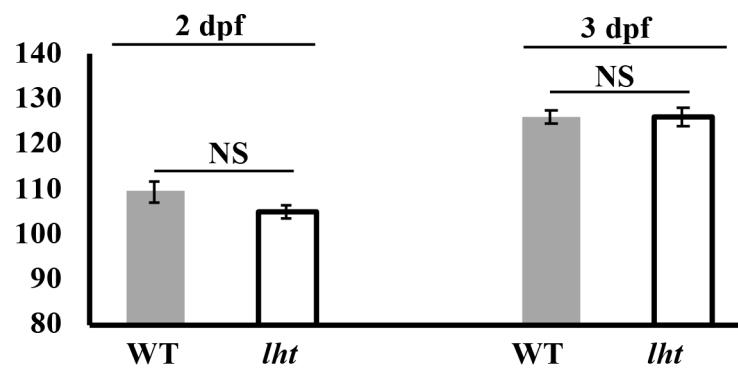
Fig. S1



E

Embryo stage	WT embryo (Mean \pm SEM)	<i>lht</i> mutant embryo (Mean \pm SEM)	P value
2 dpf	109 \pm 2.14	105 \pm 1.66	0.145
3 dpf	125.92 \pm 1.51	125.77 \pm 1.87	0.956

F



G1 domain

MEDAKA QHPVQLHCSAQRSLLHHCLSKMILLWLT*QCCQLLHLLTKQKIIILPLPQIVW*ITLIIP 1951
HUMAN ASTFE-VYSSTQRSDQLIL-----PFELE-----SPNVATSSDSGTRKSFMSLT 1777
MOUSE AHQGE-VRAATERSDHLLL-----TPELE-----SSNVDASSDLATWEGFILET 1747
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MEDAKA KKRSSQRITSAVQ*NPFPVLTVELHCPLK*LQCSAPQKVLGMAQMIHERLTHNSNCVXXX 2011
HUMAN TPTQSERE---MTDSTPVFTET-----NTLENLGAQTTEHSSIHQ-PG 1816
MOUSE TPTESEKE---MANSTPVFRET-----IGVANVEAQPFEHSSSSH-PR 1786
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MEDAKA XTDGTSFSSKIASMLSTTESSGDGTDDFT--KDSLITAT-----VSSTQGTVSPSVV 2062
HUMAN VQEGLTTLPRSPASAF-MEQGSGEAAADPETTVSSFSLNVEYAIQAEKEVAGTLSPHVE 1875
MOUSE VQEELTTLSGNPPSLF-TDLGSGDASTGMELITASLFTLDLESETKVKKELPSTPSPSVE 1845
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MEDAKA ASLTPEKSETLGTASTAATLLSSTDVKLSETSARIQQIFTSSKPLDESTKPEASSIKLST 2122
HUMAN TTFSTEP-----TGLVLSTVMDRVVAENITQTS----- 1903
MOUSE ISSSFEP-----TGLTPSTVLDIEIAGVMSQTS----- 1873
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MEDAKA DAQRLSVETGPSSKPTARPTGSSLFSTEKPTSLFKENDTVTVDLT-ILPVTPSGNQTE 2181
HUMAN REIVISERLGEPNYGAEIRGFSTGFPLEEDFSGDF-REYSTVSHPIAKEETVMMEGSGDA 1962
MOUSE QKTLISEISGKPTSQSGVRDLYTGFPMGEDFSGDF-SEYPTVSYPTMKEETVGMGGSDDE 1932
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MEDAKA DYPS-STSDSVVDHLDHTQETFIPKDYVSSTVKSFSTTDGKKSLPSVHMPSVKDTYTDM 2240
HUMAN AFRDTQTSPSTVP-----TSVHISHISDSEGPSSTM 1993
MOUSE RVRDTQTSSSIPT-----TSDNIYPVPDSKGPDSTV 1963
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MEDAKA ESSTDLPDEESSSN-DESGSGLTEFTTESL-----IEFIVTTDE-----AEINE 2283
HUMAN VSTSAFPWEEFTSSAEGSGEQLVTVSSSVVPVLSAVQKFSGTASSIIIDEGLGEVGTVNE 2053
MOUSE ASTTAFPWEEVMSSAEGSGEQLASVRSSVGPVPLAVDIFSGTESPYFDEEFEEVAATV
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MEDAKA TGSTLKIVST-----ASSTYSLTPSTQSS-HLAS----- 2312
HUMAN IDRRSTILPTAEVEGTKAPVEKEEVKVGTVSTNFPQTIPEAKLWSRQEVPVROEIESE 2113
MOUSE ANERPTVLPTAASGNTVDLTENGYIEVNSTMSDLFPQTMEPSKLWSPKEVNLDKQEIGRE 2083
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MEDAKA -----KSLPLE 2318
HUMAN TTSEEQIQEEKSFESPQNSPATEQTIFDSQTFTETELKTTDYSVLTTKTYSDDKEMKEE 2173
MOUSE TVTKEKAQGQKTFESLHSSFAPEQTILETQSLIETFQTSYDYSMLTTLKTYITNKEVEEE 2143
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MEDAKA -QSSVDFSVDISSIONSESFTFKPSIILERKPFLSTAHTVDPQATFLYSTEKAIASSLDI 2377
HUMAN DTSLVNMSTPDPDANGLESYTLPEATEKSHFFLATALVTESIPAEHV---VTDSPPIKK 2229
MOUSE GMSIAHMSTPGPGIJKDLESYTHPEAPGKSHFSATALVTEGARSV---LMDSTSQEE 2199
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MEDAKA HTSVNIFPTFHSTEKPSVPTAKVSPFH---TVRTQSPPEILNLSRVQLLTEDKTSGDPA 2434
HUMAN EESTKHFPGMRPTIQESDTELLFSGLGSGEELPTLPTESVNFTVEQINNNTLYPHT-- 2287
MOUSE EESIKLFQKGVKLTNKESNADLSFSGLGSGEA-LPPLPTTSVNLDMQIISTLYAET-- 2256
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MEDAKA NESFISKLLVLGNFTSSTPTGKTESLPALDNLSFS---LEGETAEYTKDS-LISQATDSS 2490
HUMAN ----SQVEST-----SSDKIEDFNRMENVAKEVGPLVSQTDIFEGSGSVTSTLIEI 2335
MOUSE ----SHMESLGT---SILGDKMEDHERMEDVSNEVRMLISKTSISQDS---TEALDTT 2305
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MEDAKA TGNDGANLPRNTLHPSHTASLLPESLKSDQTVPDFTSETVSSLKDKVSSIPTAFPTIY 2550
 HUMAN LSDTGAEGPTVA-----PLPFST--- 2353
 MOUSE LSHTGTEEPTTS-----TLPFVKLM- 2325
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MEDAKA RSIADQQVGIITPSTKQVETIKAGQTPTMVHLHEPKASISVYKIFTEEAKNELISGGTE 2610
 HUMAN -----DIGHPQ----NQTV---RWAEEIQ----- 2370
 MOUSE -----DLERSP----KQTL---RWEETQ----- 2342
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MEDAKA SLTPESITSEFITKDNAI---I---DTISTAQVPFFYPTVLTSGGIKADTKAQKLKIMEET 2665
 HUMAN TSRPQTITEQDSNKNSSAEINETTSSTDFLARAYGFEMAKEFVTSAKPBSDLYEPESG 2430
 MOUSE THRPQTMSGlisnenSSASEAEEAATSPTAFLPQTYSVEMLKFAPSESQPSDFNVNSG 2402
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MEDAKA EGSGTARSAILTPTPSILSAAPKSESVLTSHGIISTM----- 2703
 HUMAN EGSGEVDIVDSFHTSATTQATRQESSTTFVSDGSLEKHPEVPSAKAVTADGFPTVSVMLP 2490
 MOUSE EGSGEVDTLDLVYTSGTTQASSQG-DSMLASHGFLEKHPEVSKTEAGATDVSPTASAMFL 2461
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MEDAKA -----RTERD---SA---KT----- 2712
 HUMAN LHSEQNKSSPDPTSTLSNTVSYERSTDGSFQ--DRFREFEDSTLKPNRKKPTENIIDLD 2548
 MOUSE HHS-EYKSSLYPTSTLPSTEYKSPSEGIEDGLQDNIQFEGSTLKPSSRKTTESIIDLD
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MEDAKA SENYDDLSVKMITSSVYSMFS-----TQKPTLDKDIVPGDLDQSIFTLSTA 2759
 HUMAN KED-KDLILITESTILEILPELTSKNTIIDHTKPVYEDILGMQTDIDTEVPSEPHD 2607
 MOUSE KEDSKDLGLTITESAIVEILPELTSKNTIIDHTKPVYEYIPGIQTDLDPEIKLESHG 2580
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MEDAKA DS-NISENTLLSETLETETPVSPIEETFKTTEKD---ETQTLHSSGVSEQ-KKEF--L 2811
 HUMAN SNDESNDSTQVQEIEYAAVNLSLTEETFEGSADV-LASYTQATHDESMTYEDRSQLDHM 2666
 MOUSE SS---EESLQVQEKEYEGAVTLSPTESFEGSGDALLAGYTQAIYNESVTPNDGKQAEDI 2636
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MEDAKA PIGRITSAPTHEEITSSVEISPNASTVSFPQSTPKSKVTVQFVTTFALQPDTIQTVENTFO 2871
 HUMAN GFHFTTGIPAPSTETELDVLLPTATSLPIPRKSATV----- 2702
 MOUSE SFSFATGIPVSSTETELHTFFPTASTLHIPSCLTTA----- 2672
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MEDAKA HARSEPHQFRNDSSLESQVRSEIQSTQTPQANHNSQDASLTTILPISTSHQGSQITKF 2931
 HUMAN -----IPEIE-----GIKAEEKALDDMFESSTLSDGQAIADQ 2734
 MOUSE -----SPEID-----KPNIEAISLDDIFESSTLSDGQAIADQ 2704
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MEDAKA TAIHPGDHSVTEAGSVLEDGKTLKLKTPSSDTKYI-----DNIDYTAPD----- 2976
 HUMAN SEIIPTLGQFERTQEEYEDKK---HAGPSFQPEFSSGAAEALVDHTPYLSIATTHLMDQ 2790
 MOUSE SEVISTLGHLEKTQEEYEEKK---YGGPSFQPEFFSGVGVEVLTDPAYVSIGSTYLIAQ 2760
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MEDAKA --YDLVDPIRLESVPKYKNNSKEMEDSLAKPQTPISTSPISFYESGSESTSSEESMPLT 3034
 HUMAN SVTEVPDVMEGSNPPYYTDT-TLAVSTFAKLSSQTPSSPLTIYSGSEASGH---TEIPQP 2846
 MOUSE TLTELPNVVRPSDSTHYTEA-TPEVSSLAELSPQIPSSPFPVYDNGVSKF---PEVPHT 2816
 : : : : . *.. . .: *: . :**: .* .. * .:*

MEDAKA STAKV-----DSYVRGKIPSDLSSPTTMSSVLDPG 3065
 HUMAN SALPGIDVGSSVMSPQDSFKEIHVNIEATFKPSSEYLYHITEPPS-LSPDTKLEPSEDDG 2905
 MOUSE SAQPVSTVTSSQKSIESPFKEVHANIEETIKPLGG-NVRTEPPS-MSRDPALDVSEDES 2874
 *: : : * . :* :. * .

G3 domain

Fig. S3

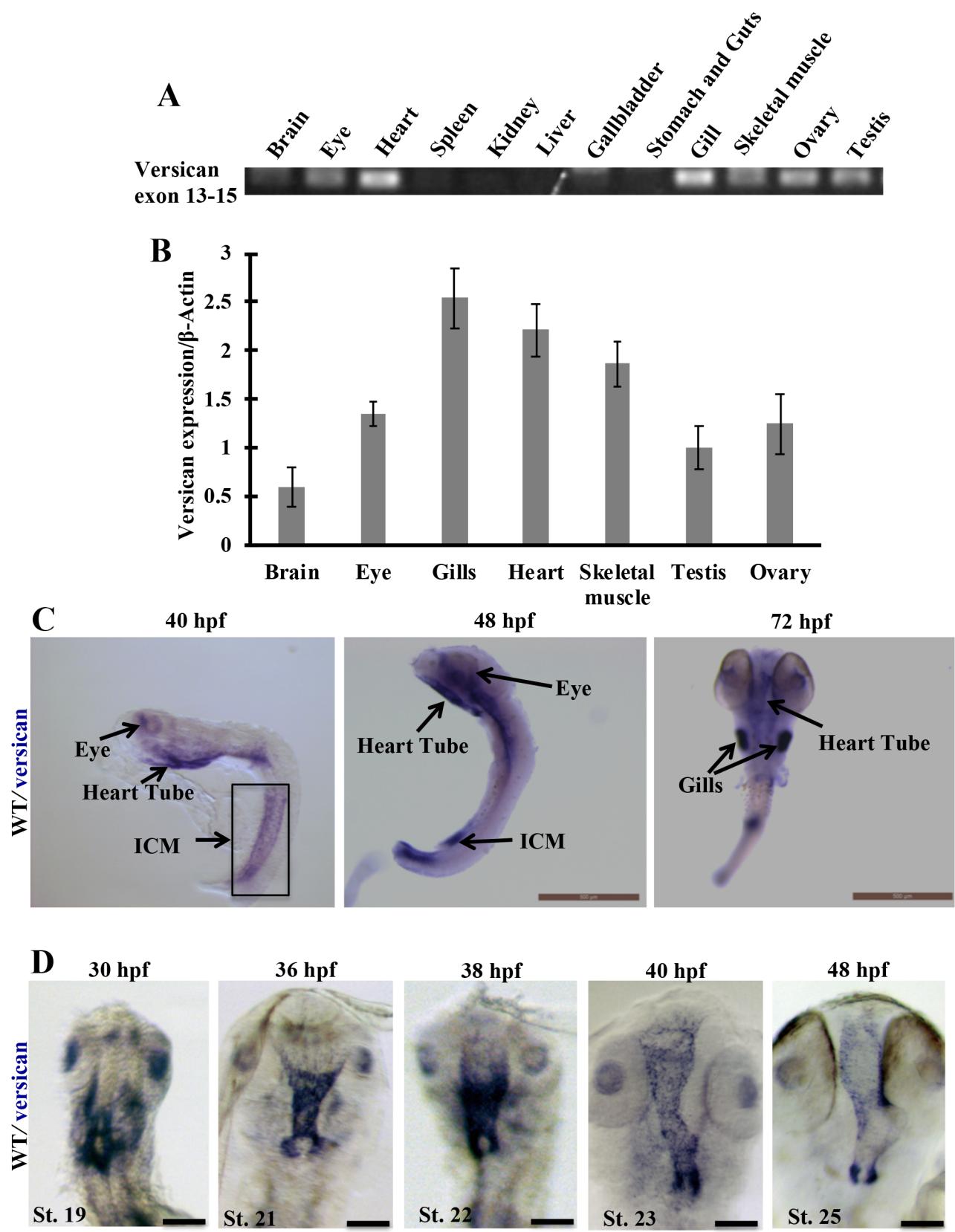


Fig. S4

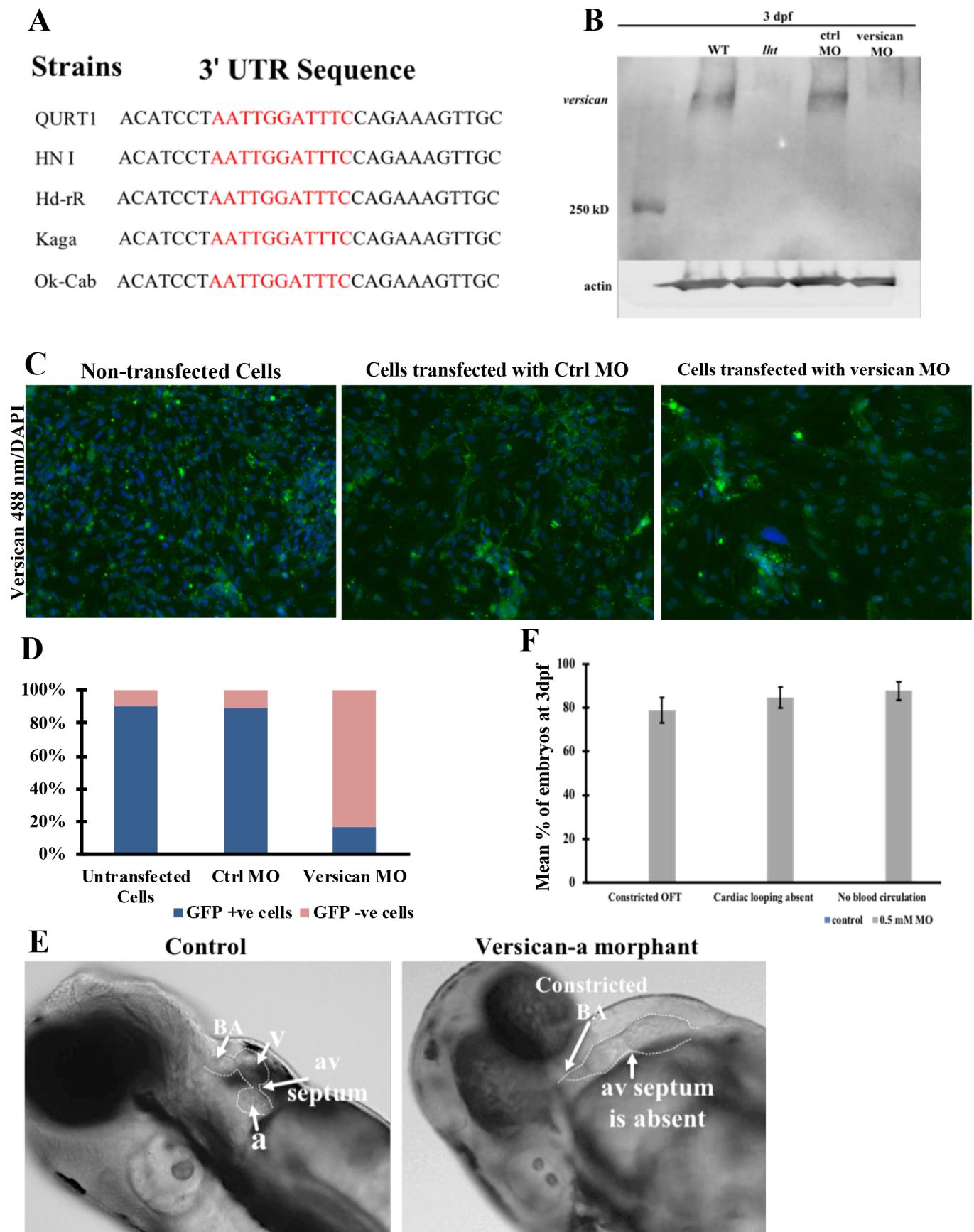


Fig. S5

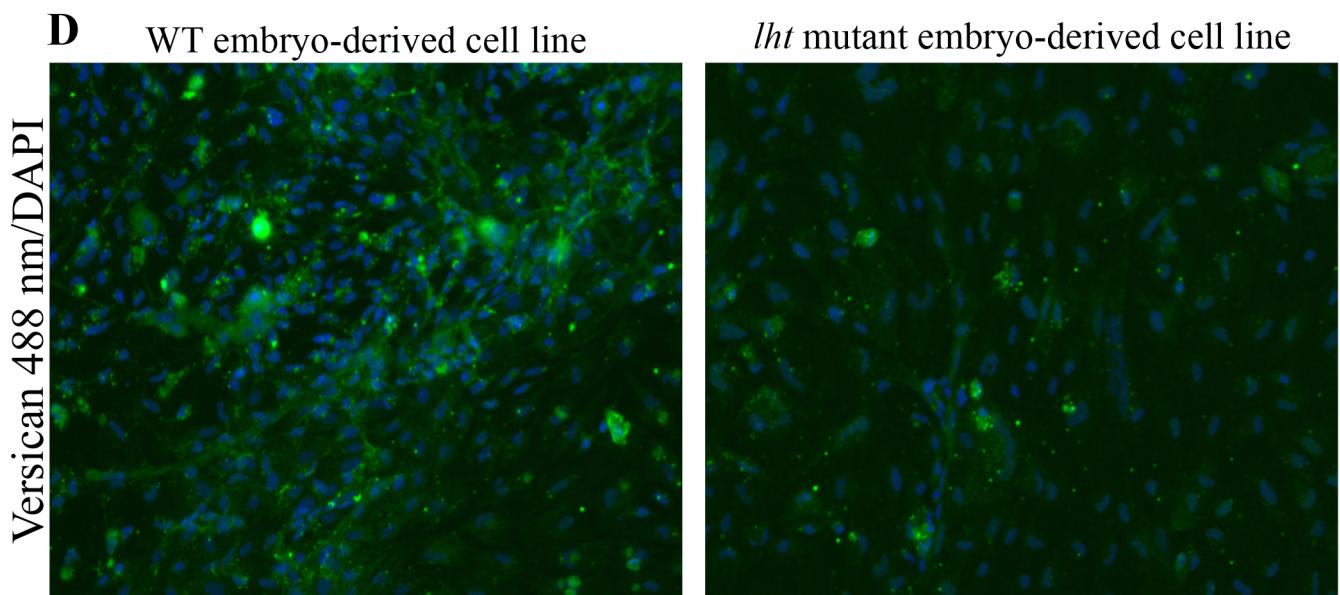
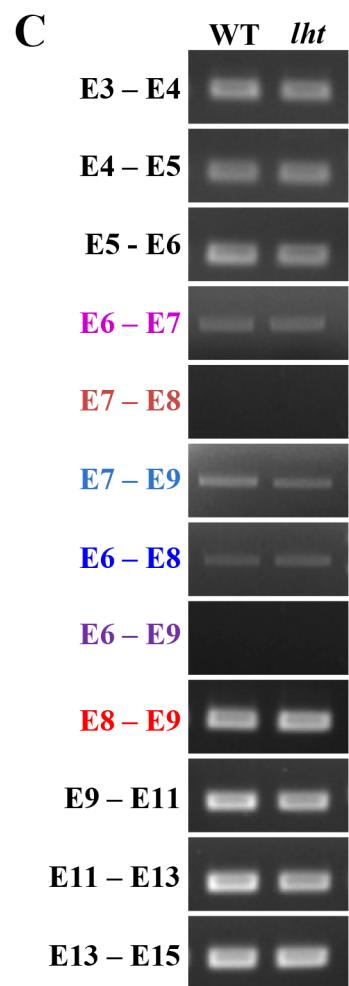
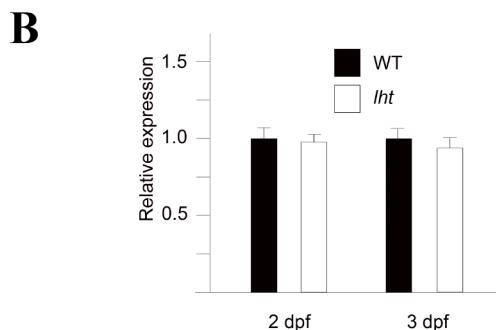
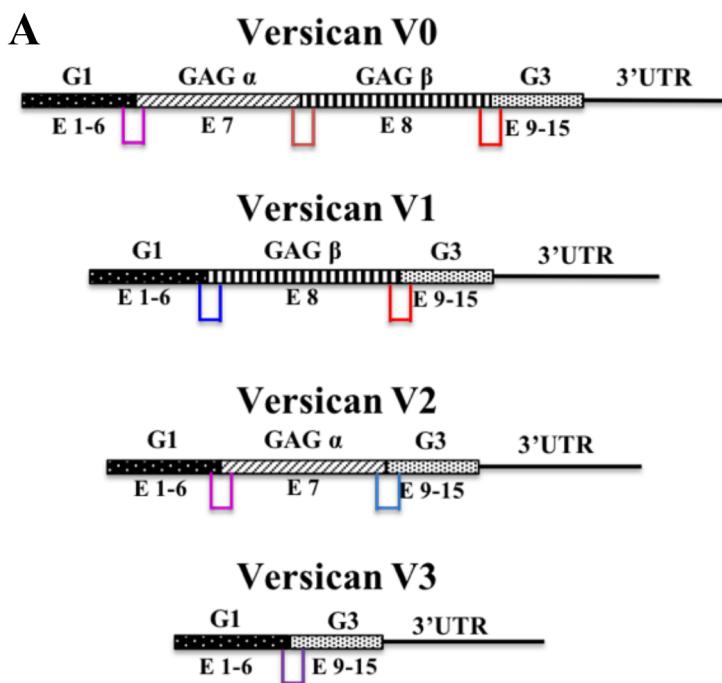


Fig. S6

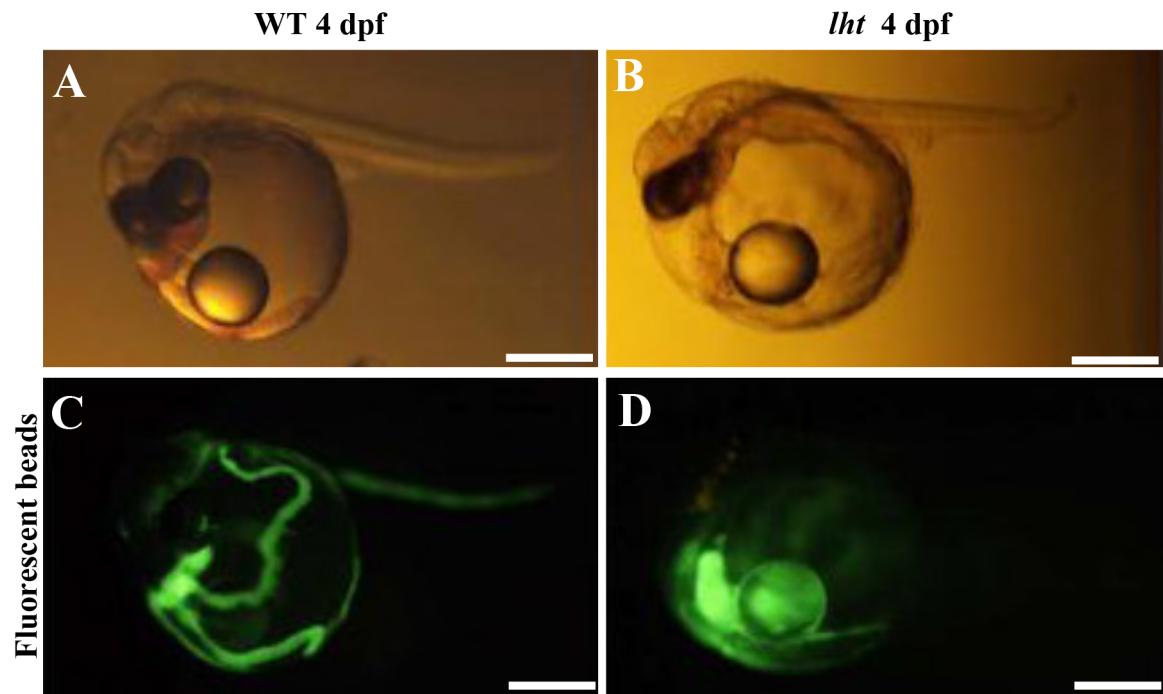


Fig. S7

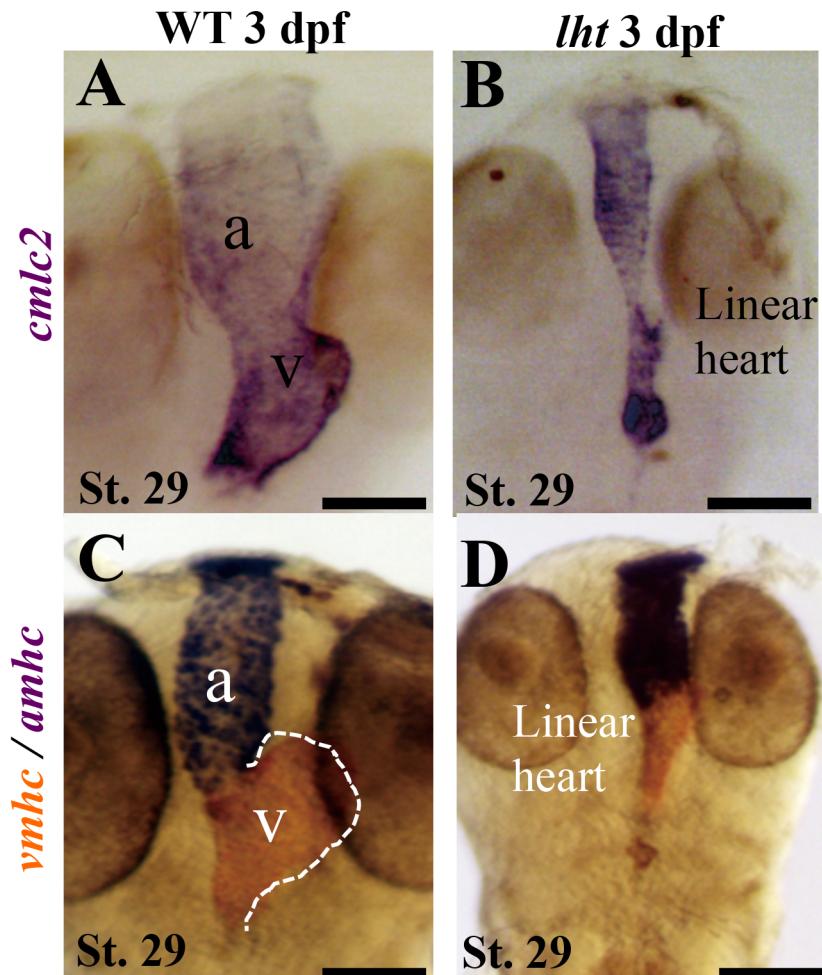


Fig. S8

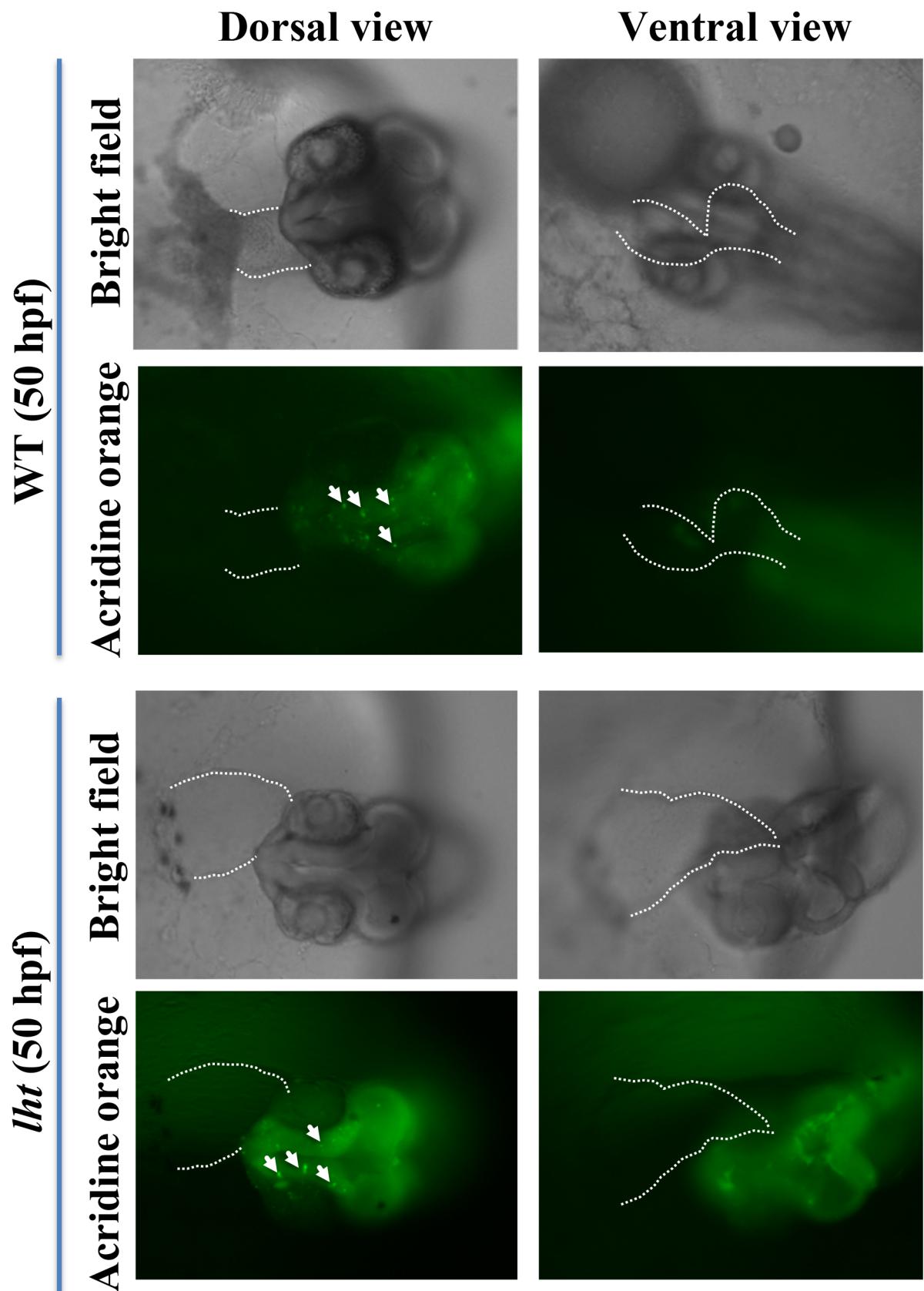


Fig. S9

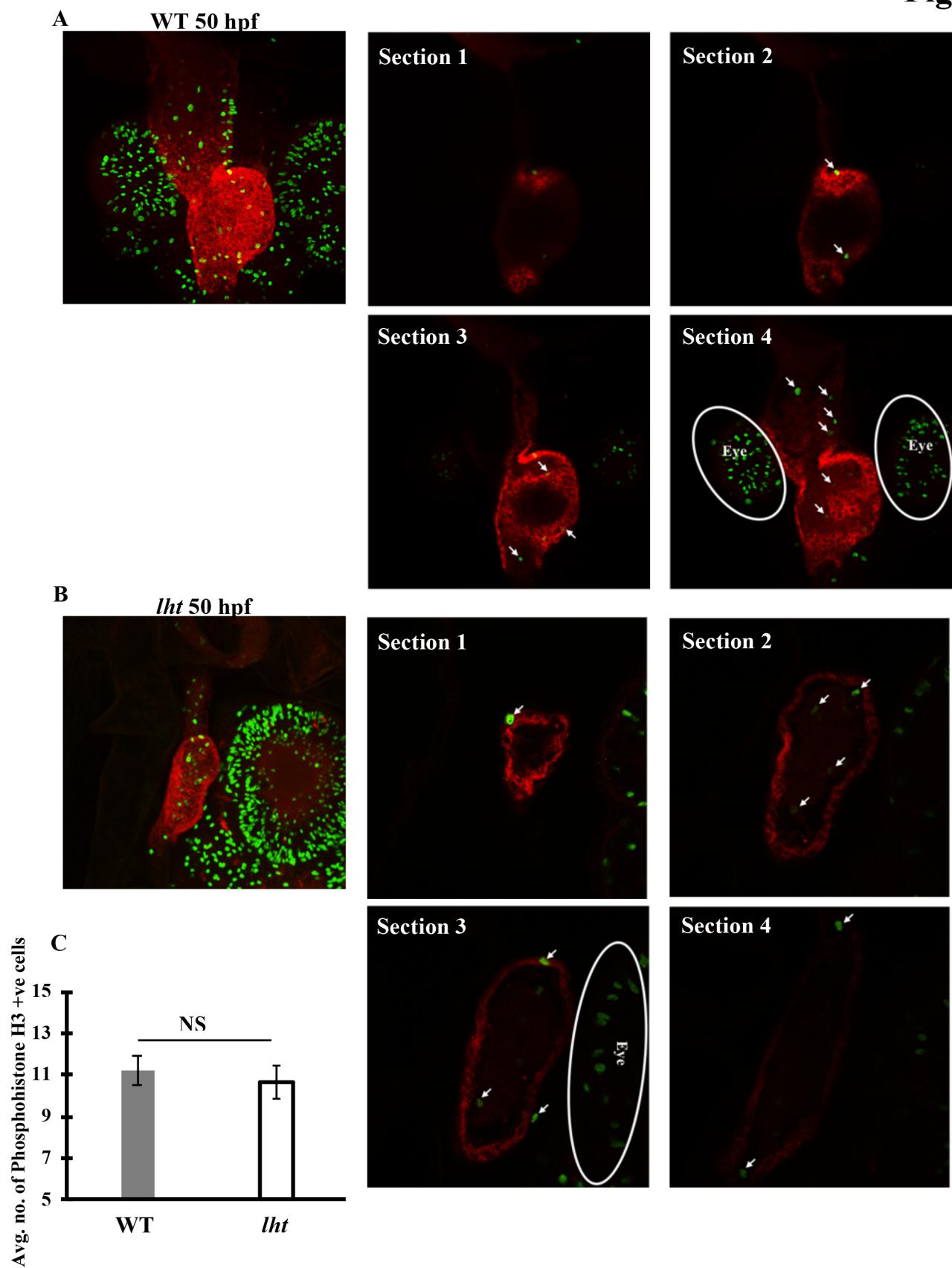


Fig S10

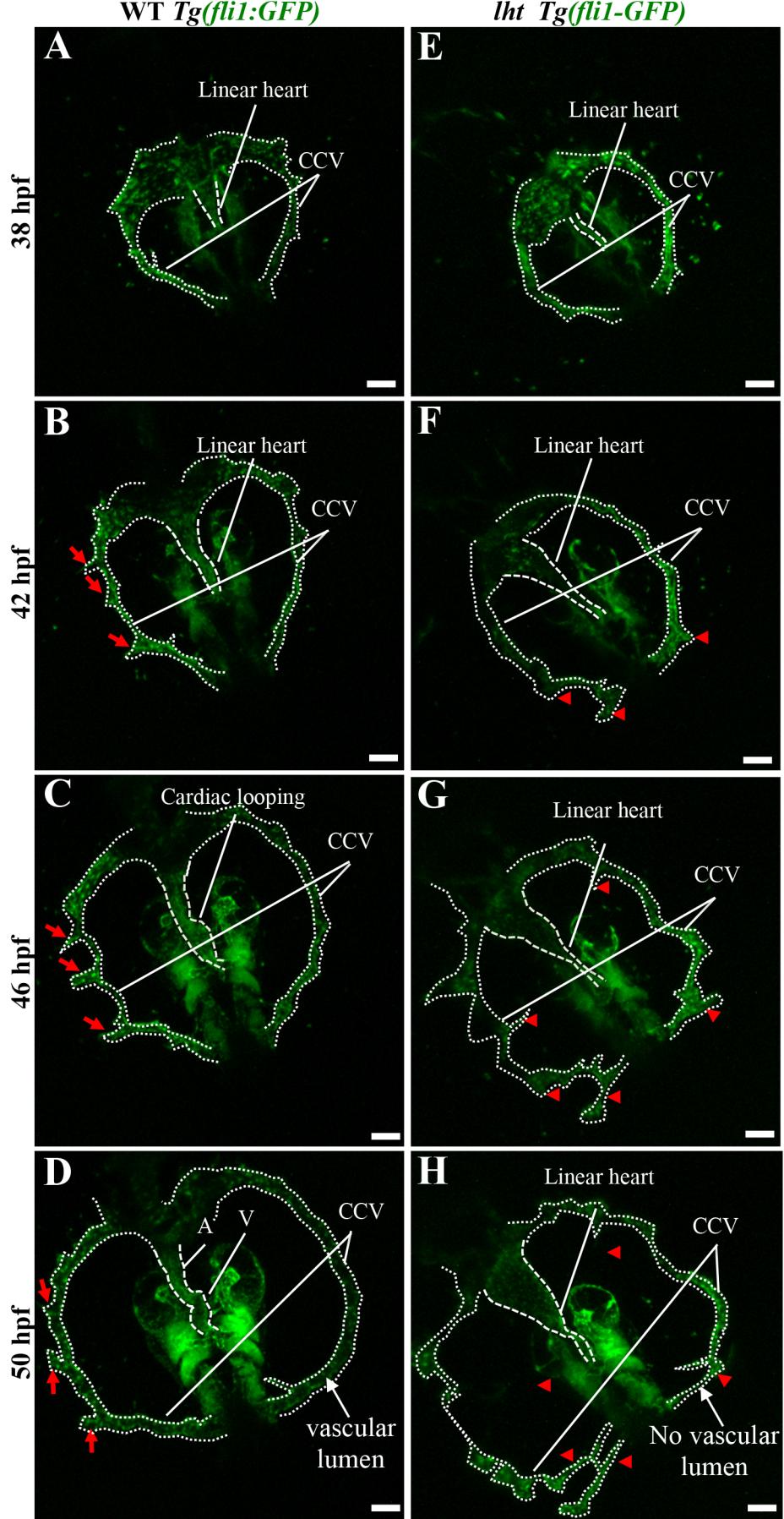


Fig. S11

WT 2.3 dpf (after yolk removal)

etgase



Fig. S12

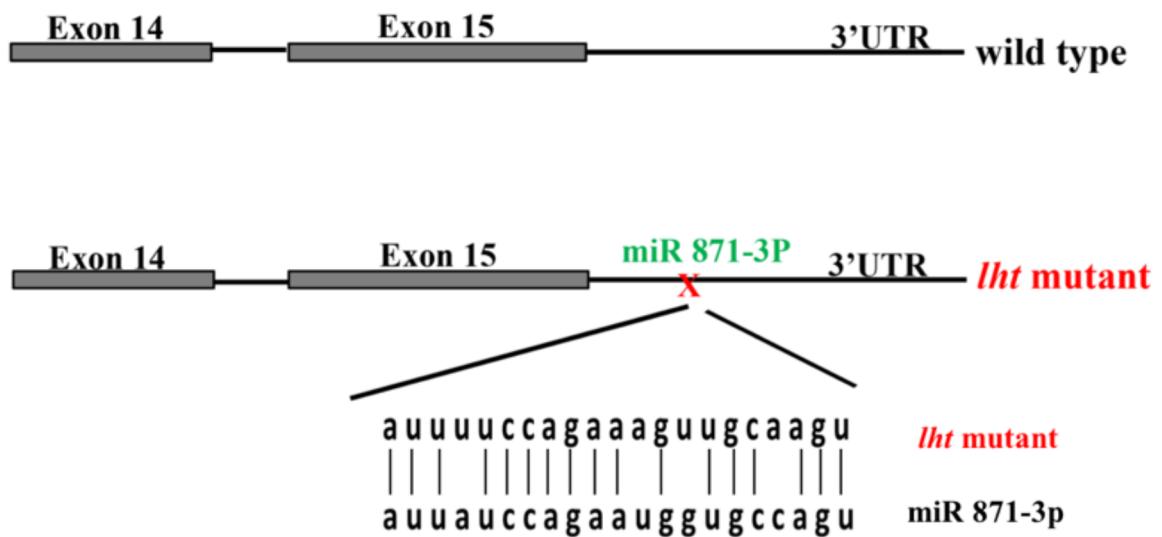


Fig. S13

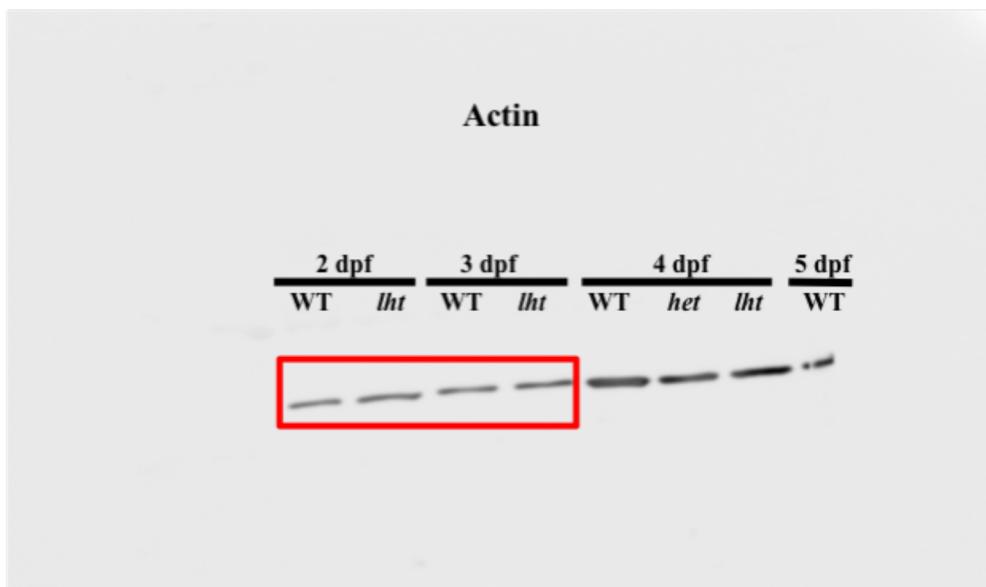
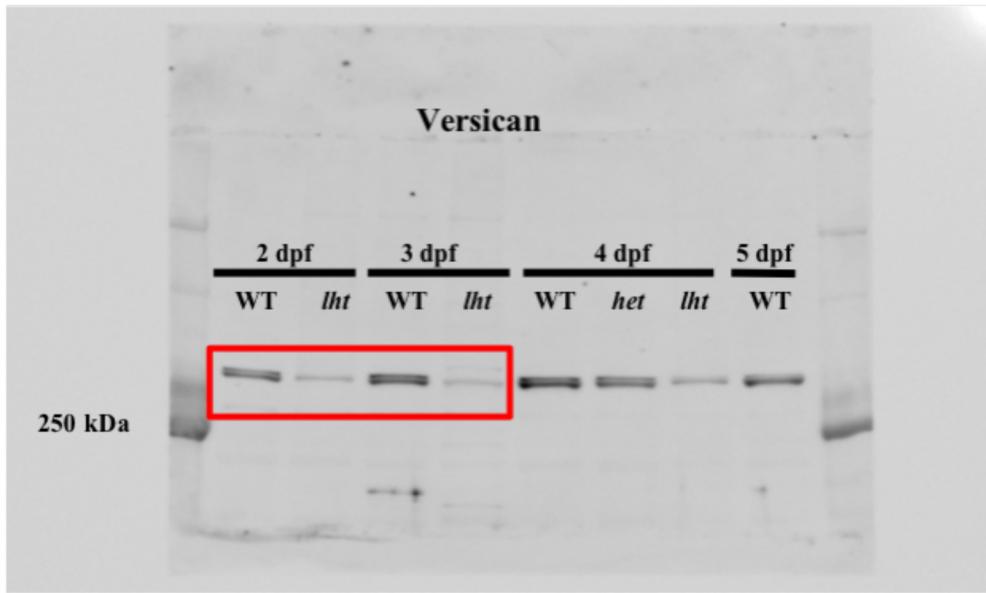


Fig. S14

Versican

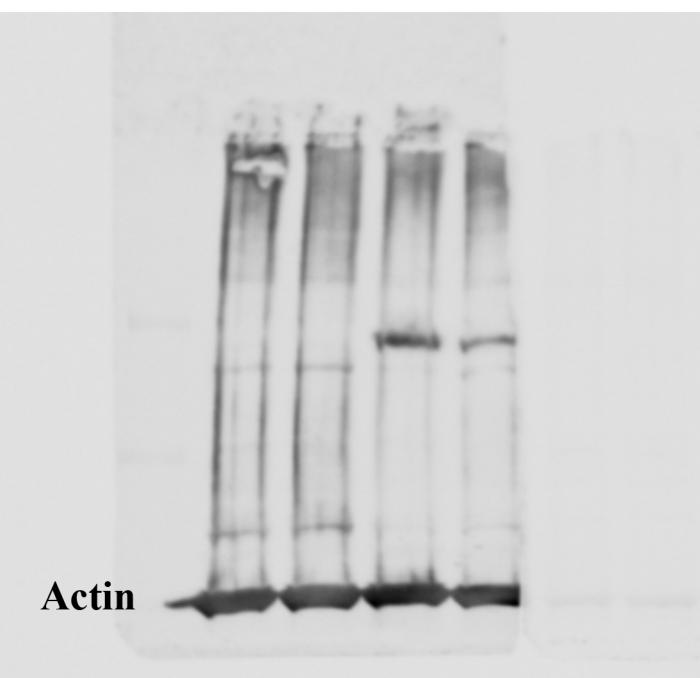
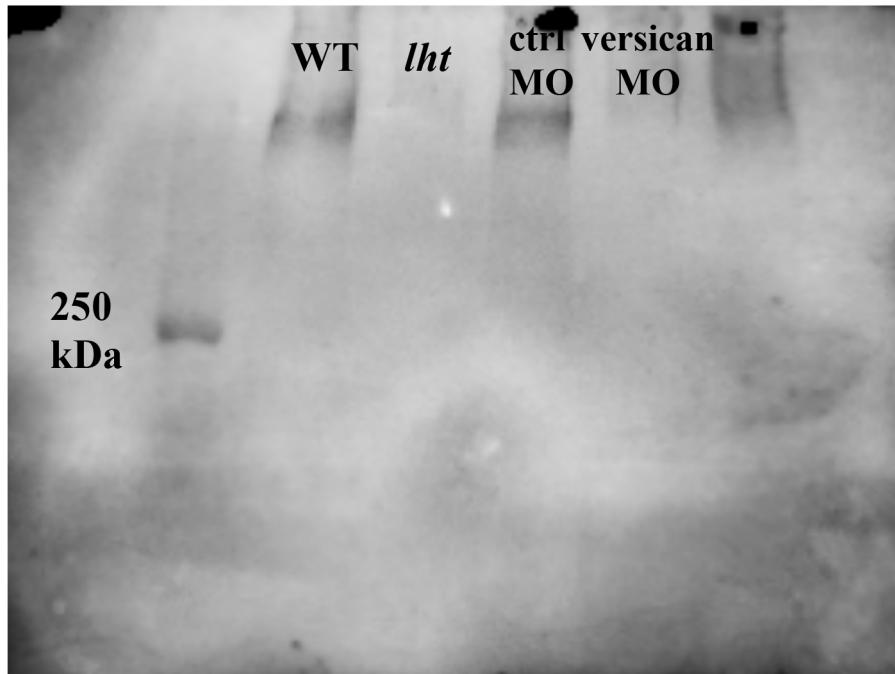


TABLE 1: Primers used for mapping

Contig. Name	Forward Primer (5'-----3')	Reverse Primer (5'-----3')	Restriction Enzyme
Olb0807j	TCGACACCTTCTGTTGTGAGAGCG	GGCCTTAGCAGTTGTTCTGGCA	HinfI
MFOISSA038C07	CCTGTCGTTGCTGAGTCCTTC	AGGTGTAATGCTGTTCGACCTC	RSaI
OI11523	ATTTAGTCCCCCAGGAGTGG	CGTTCTCGTTCTCCTGCAA	RSaI
OI12320	GCAGGTGGAAGTCGACAAA	CCAATGGACCATCACTTCC	RSaI
OI12338i5	TCCTTCAAAGGGAGTGAATGG	ACCCAGAGGTGAAGACATGC	HinfI
OI12419	AGTCAAGGGCAAGGATGATG	AGAGTTGGCTGATGGAGCAT	Hae3
OI1262li4	TCTGCCATCTCTACTTCTTATGG	TGAGGACAGGTCTGCACTGTT	HinfI
OI102119	CCAGCCAGAAGAACAAATGC	GCGAAAGCCCTGTTAAACT	RSaI
TiT8j07	TGAGCATTGGGTTACTGTG	ACTTGTGCGCTTGAGATG	RSaI

TABLE 2: Primers used to verify splicing of versican gene

Primer Name	Forward Primer (5'-----3')	Primer Name	Reverse Primer (5'-----3')
Vcan ex 3	CAGTGCCCAGTCATCCTGAA	Vcan ex 4	ATCTTGACAGGCCCGAACAG
Vcan ex 4	CTGTTCGGGCCTGTCAAGA	Vcan ex 5	TGCCAGGGTTAGTCATGAGG
Vcan ex 5	CCTCATGACTAACCCCTGGCA	Vcan ex 6	CCATAGTCGCATCTGTCTAGC
Vcan ex 6	CCAGAATTCAAGTGTGGGGGA	Vcan ex 7	TGGATGGAGGCCATGCTAGTG
Vcan ex 7	CTACCACAATAAGTTAGAGGAACAT	Vcan ex 8	AGACTTAGCAGGGATCCTCG
Vcan ex 7	CTACCACAATAAGTTAGAGGAACAT	VCan ex 9	AAGAGGACCAGTGTAAACCGG
Vcan ex 6	ATGCCCTTGACAAGCACTGC	Vcan ex 8	AGACTTAGCAGGGATCCTCG
Vcan ex 6	ATGCCCTTGACAAGCACTGC	Vcan ex 9	AAGAGGACCAGTGTAAACCGG
Vcan ex 8	GATGGCAAACAACCTGGCGA	Vcan ex 9	AAGAGGACCAGTGTAAACCGG
Vcan ex 9	CCGGTTACACTGGTCCTCTT	Vcan ex 11	CATTCTCGCTCTGCTGAGTC
Vcan ex 11	GACTCAGCAGAGCGAGAATG	Vcan ex 13	CGTCATTCCACTGTCCATCC
Vcan ex 13	GGATGGACAGTGGAATGACG	Vcan ex 15	GCTCCTCTGGTAGCTTGAAG

TABLE 3: Primers for real-time PCR

Primer Name	Forward Primer (5'-----3')	Reverse Primer (5'-----3')
BNP	CTTCATCCCTTTGGGGGC	TTCTCGCTCCTCCTGTG
Versican	GGATGGACAGTGGAAATGACG	GCTCCTCTGGTAGCTTGAAG

TABLE 4: Primers for in-situ Probe design

Primer Name	Forward Primer (5'-----3')	Reverse Primer (5'-----3')
Cmlc2	AATGTCTTTCCATGTTGAGC	CTCCTCTTCTCATCCCCATG
Amhc	TGCACTGATGGCTGAATTG	ACTTGATCTACA CCTTGGCC
Vmhc	GCTGAGATGTCCGTGTATGGTGC	GCTCCTCACGAGGCCTCTGCTTG
Bnp	GATCCATCCATCCATCATCC	TGATACTTAAAGACACAATGTCCAA
Versican	CACGACAATTCTCCCCATCT	TGCTCTTCCAGTCTCCTGGT
e-tgase	CCCACCCCTGACAGAGTTTC	GTCACAAACACGAGACGGGAT
TMEM 205	GCTCTACGTCTGAACTTGCG	ATCTCATGCCAGTTGTTCAA
Egfl7	GCACCTACAAGACCACCTAC	GGGACGAGGAGAATCACGAA