

Supplemental Table S1. The primer sequences (Liu et al., *J. Immunology*, 2016, 197:4736)

Primer	Sequence (5' to 3')
zebrafish <i>ifn1</i> -F	GAGCACATGAACTCGGTGAA
zebrafish <i>ifn1</i> -R	TGCGTATCTTGCCACACATT
zebrafish <i>ifn2</i> -F	CCTCTTTGCCAACGACAGTT
zebrafish <i>ifn2</i> -R	CGGTTCTTGAGCTCTCATC
zebrafish <i>rsad</i> -F	AGCAGATCACCGCTCTCAAT
zebrafish <i>rsad</i> -R	CCAGACACTGGATGCTCTGA
zebrafish <i>mxh</i> -F	AATGGTGATCCGCTATCTGC
zebrafish <i>mxh</i> -R	TCTGGCGGCTCAGTAAGTTT
zebrafish <i>mxr</i> -F	GAGGCTTCACTTGGCAACTC
zebrafish <i>mxr</i> -R	TTGTTCCAATAAGGCCAAGC
zebrafish <i>pkz</i> -F	GGAGCACCGTACAGGACATT
zebrafish <i>pkz</i> -R	CTCGGGCTTTATTTGCTCTG
zebrafish <i>mavs</i> -F	GTTCCCGGTCCAAGACACTA
zebrafish <i>mavs</i> -R	TTGTGCGCTGAGTTGTTCTG
zebrafish <i>rig1</i> -F	TTGAGGAGCTGCATGAACAC
zebrafish <i>rig1</i> -R	CCGCTTGAATCTCCTCAGAC
zebrafish <i>lta</i> -F	AAGCCAAACGAAGGTCA
zebrafish <i>lta</i> -R	AACCCATTTTCAGCGATTGTC
zebrafish β - <i>actin</i> -F	TACAATGAGCTCCGTGTTGC
zebrafish β - <i>actin</i> -R	ACATACAATGGCAGGGGTGTT
EPC- <i>ifn</i> -F	ATGAAAACTCAAATGTGGACGTA
EPC- <i>ifn</i> -R	GATAGTTTCCACCCATTTCTTAA
EPC- <i>isg15</i> -F	CAGCCTTGAGGATGATTCCAG
EPC- <i>isg15</i> -R	TGCCGTTGTAAATCAGTCG
EPC- <i>viperin</i> -F	AGCGAGGCTTACGACTTCTG
EPC- <i>viperin</i> -R	GCACCAACTCTCCAGAAAA
EPC- β 2M-RT-F	CTCCATTGAACTGCTGAAAGATG
EPC- β 2M-RT-R	CAAATAACTGTCTTCATTTGCTCAT
EPC- β - <i>actin</i> -F	CACTGTGCCCATCTACGAG
EPC- β - <i>actin</i> -R	CCATCTCCTGCTCGAAGTC
SVCV- <i>P</i> protein-F	TTGGACCTGGGATAGTGA
SVCV- <i>P</i> protein-R	CTTGCTTGTTTGTGGG
SVCV- <i>G</i> protein-F	CGACCTGGATTAGACTTG
SVCV- <i>G</i> protein-R	AATGTTCGGTTTCTCACT
SVCV- <i>N</i> protein-F	TGAGGTGAGTGCTGAGGATG
SVCV- <i>N</i> protein-R	CCATCAGCAAAGTCCGGTAT

Supplemental Figure Legends

Supplemental Figure S1. MLN4924 inhibits the expression of *ifn1* and *pkz* in zebrafish larvae upon SVCV infection.

(A-B) Zebrafish larvae (3 dpf) were pretreated with different dosage of MLN4924 for 24 h, and then were infected with SVCV (+SVCV) or PBS control (-SVCV). The expression of *ifn1* and *pkz* was examined by qRT-PCR.

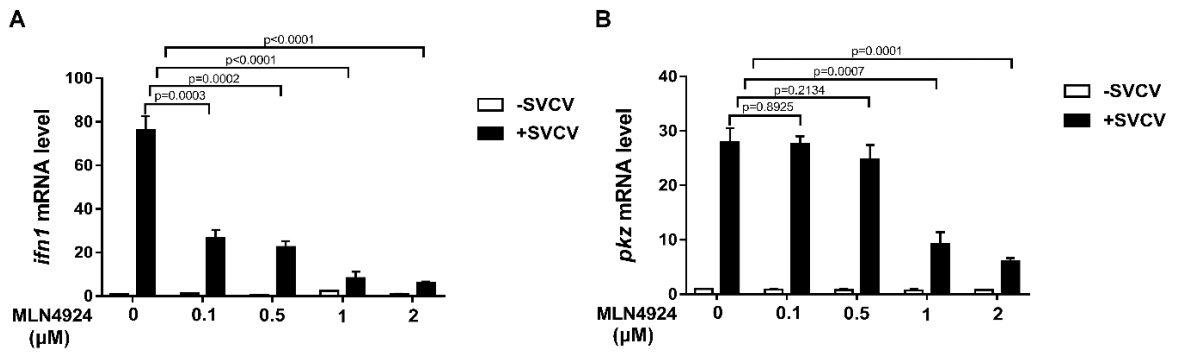
Supplemental Figure S2. Generation of *nedd8*-null zebrafish via CRISPR/Cas9 technology.

(A) Scheme of the genomic structure of zebrafish *nedd8* and the sequence information in mutant *nedd8* zebrafish (mutant1: *nedd8*^{ihb1227/ihb1227}, in which 4 bp was deleted in exon 3). (B) The sequence information of targeting sites (the blank line) in wild type (WT) and mutant *nedd8* allele. Upper panel: the sequence of the Cas9 target site is indicated with a black line and the *nedd8* mutant allele are shown. The red dashed line indicates the deleted nucleotide. (C) The predicted protein of *nedd8* in WT and the mutant. (D) The *nedd8* mRNA level in spleens from the *nedd8*^{+/+} or the *nedd8*-null mutant (*nedd8*^{ihb1227/1227}) (4 mpf ; n=3, respectively). (E) The Nedd8 protein in WT and the mutant larvae (3 dpf). Mpf, month post fertilization. Data are presented as means \pm SEM of three independent experiments, performed in triplicate.

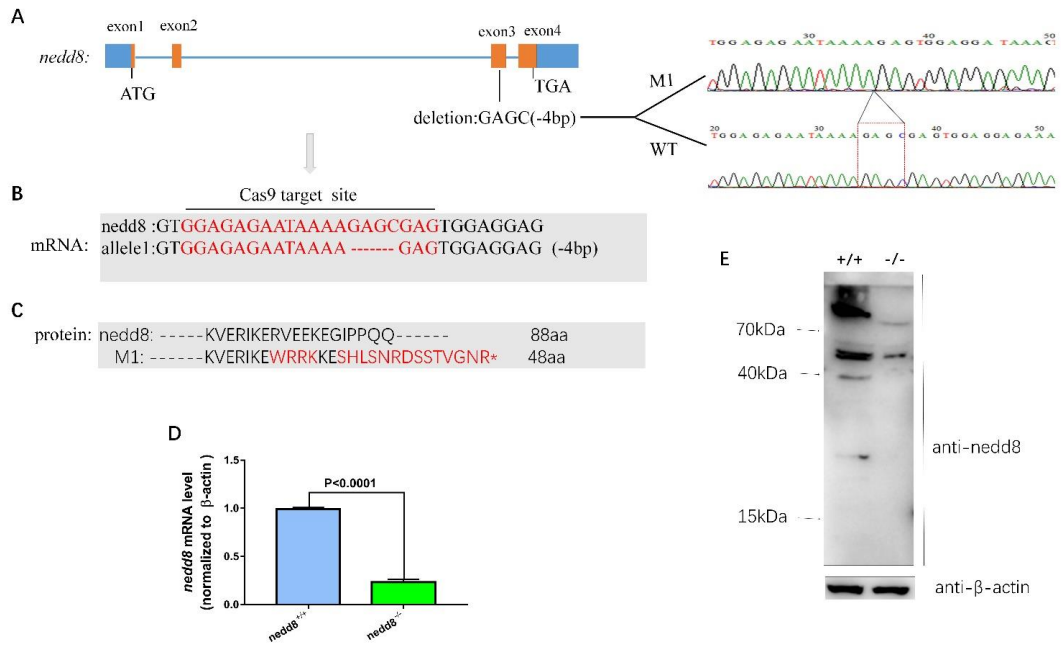
Supplemental Figure S3. Neddylation does not targets Mda5, Mavs and Tbk1.

(A-C) HEK 293T cells were transfected with the indicated plasmids together with His-*nedd8* respectively (5 μ g/each). After 36 hr, cells were lysed in guanidinium chloride, and His-*nedd8* was purified with Ni²⁺-NTA agarose. TCL, total cell lysates; IP, immunoprecipitation.

Supplemental Figure S1.



Supplemental Figure S2.



Supplemental Figure S3.

