

Supplemental material

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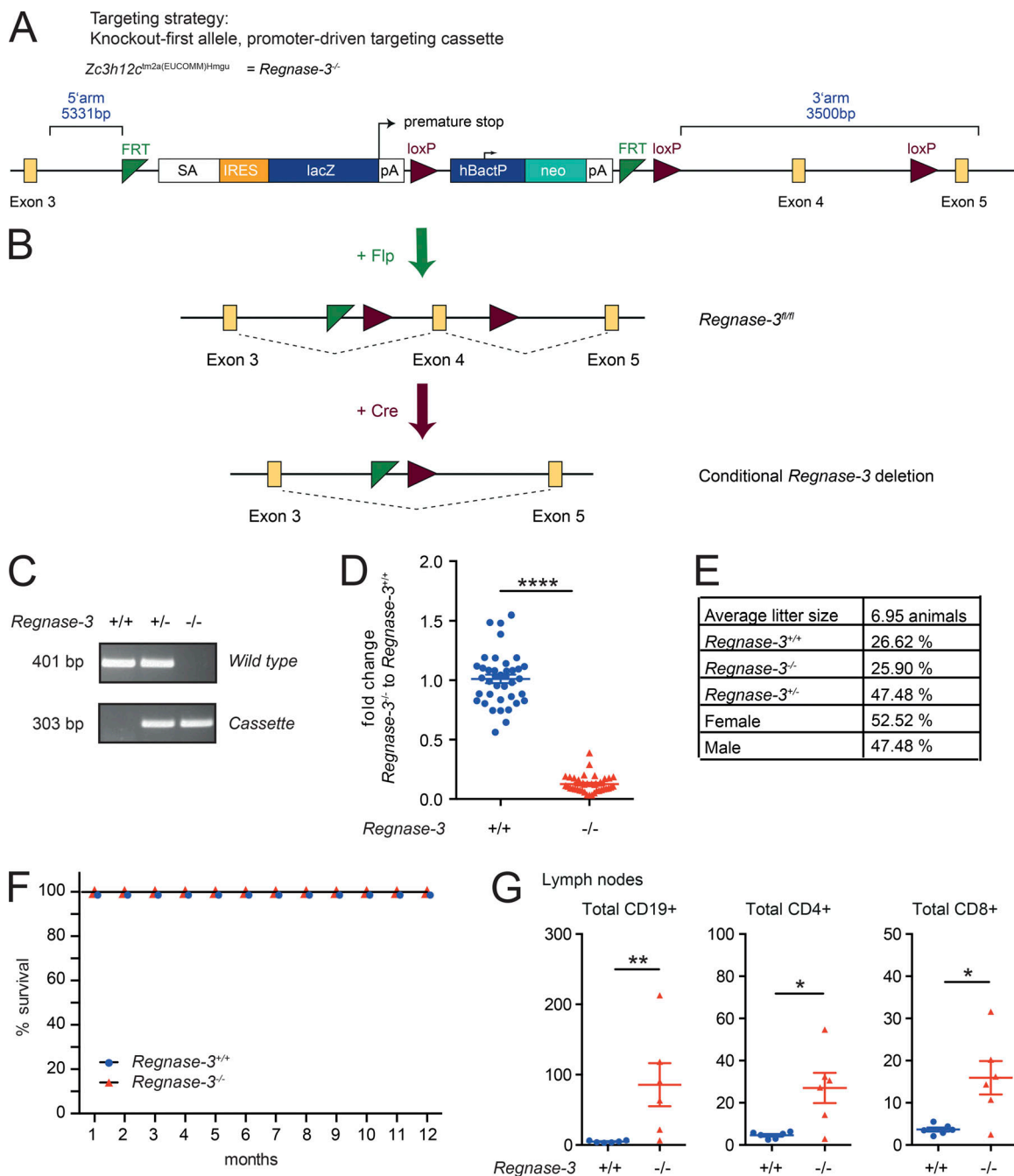


Figure S1. **Generation of *Regnase-3* (*Zc3h12c*)-deficient mice. (A)** Schematic representation of the targeting vector for generation of *Regnase-3* (*Zc3h12c*)-deficient mice: FRT, FLP recombinase target; SA, splice acceptor; IRES, internal ribosome entry site; lacZ, β -galactosidase; pA, polyA; hBactP, promoter; neo, neomycin resistance gene. **(B)** Mating strategies for generation of mice with conditional deletion of *Regnase-3* (*Zc3h12c*) with promoter-driven Cre recombinases. **(C)** PCR analysis of mouse ear tissue to identify the *Regnase-3* genotype using wild type primers and primers to identify the cassette. **(D)** Quantitative RT-PCR of the *Regnase-3* (*Zc3h12c*) gene transcript in liver tissue from *Regnase-3^{+/+}* and *Regnase-3^{-/-}* mice ($n = 39/39$), relative to expression in *Regnase-3^{+/+}* mice. **(E)** 20 representative litters from het \times het matings with a total of 139 mice were used to calculate average litter size and the distribution of the genotypes and sex. **(F)** Survival rate in a cohort of *Regnase-3^{+/+}* and *Regnase-3^{-/-}* mice ($n = 10/10$). **(G)** Number of calculated total B cells (CD19⁺) and CD4⁺ and CD8⁺ T cells in lymph nodes of *Regnase-3^{+/+}* and *Regnase-3^{-/-}* mice ($n = 6/6$). Data are represented as mean \pm SEM and were compared by Mann-Whitney *U* test (*, $P \leq 0.05$; **, $P \leq 0.01$; ****, $P \leq 0.0001$).

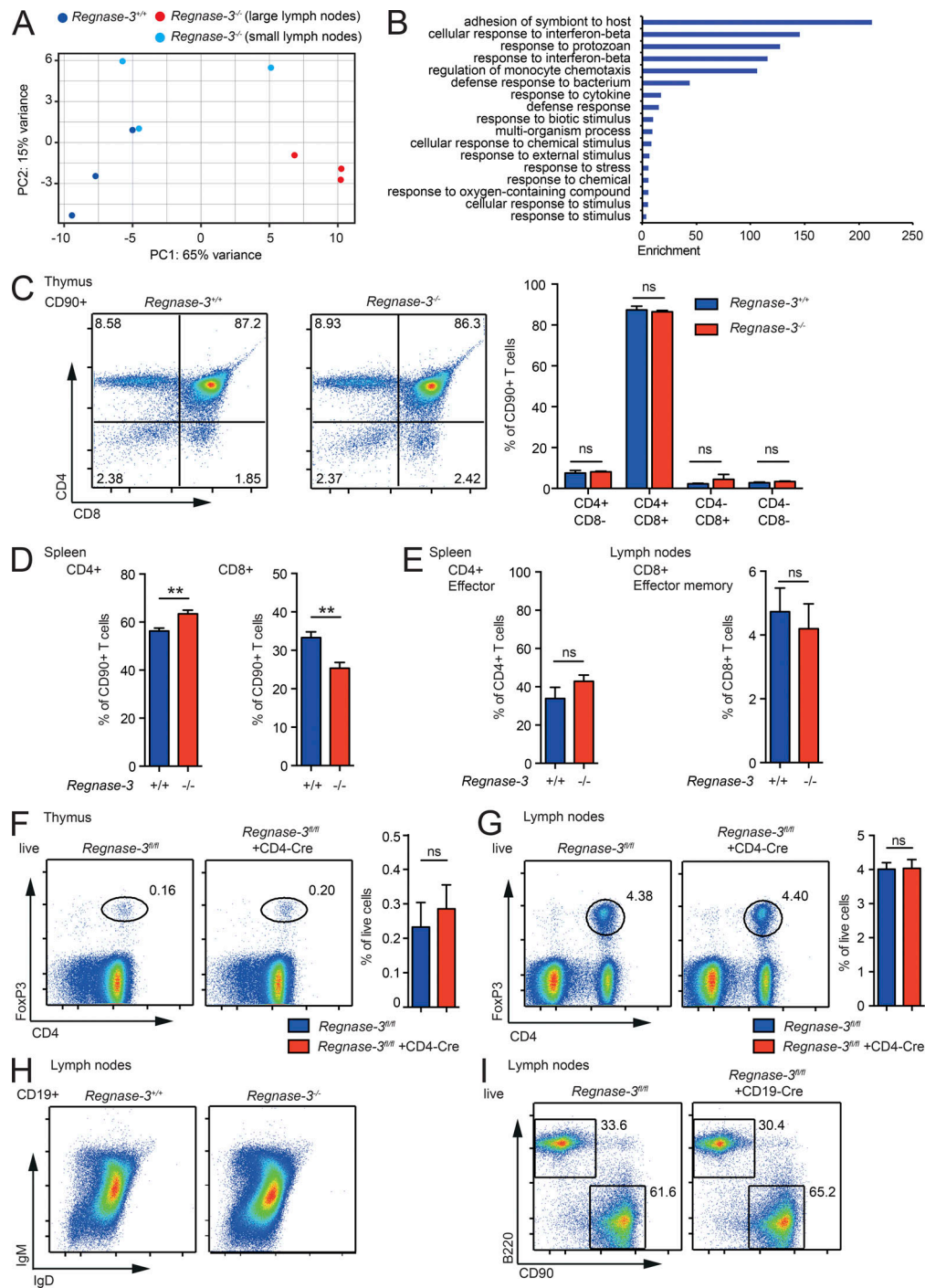


Figure S2. T and B cell analysis of *Regnase-3^{-/-}* mice. (A) B cells (CD19⁺) were isolated from enlarged lymph nodes and normal-sized lymph nodes from the same *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermates ($n = 3/3$), and RNA was isolated and subjected to RNA sequencing. The individual B cell samples are plotted based on PCA. (B) Enriched GO terms ($P < 0.05$) for all up-regulated ($\geq 2 \log_2$ fold) genes (as shown in Fig. 4) ordered according to their enrichment value. (C) Frequencies of double-negative, double-positive, and CD4 and CD8 single-positive T cells in the thymus of *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermate controls at 6 mo of age assessed by flow cytometry ($n = 8/8$). Left: Representative blots. Right: Statistics. (D) Frequencies of splenic CD4⁺ and CD8⁺ T cells within the T cell compartment (CD90⁺) of *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermate controls at 6 mo of age assessed by flow cytometry ($n = 9/9$). (E) Frequencies of splenic CD4⁺ effector T cells (CD19⁻, CD90⁺, CD4⁺, CD62L⁻/CD44^{hi}) in *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermate controls at 6 mo of age ($n = 6/6$), and frequencies of CD8⁺ effector memory T cells (CD19⁻, CD90⁺, CD8⁺, CD62L⁻/CD44^{hi}) in lymph nodes of *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermate controls at 6 mo of age ($n = 6/6$). (F and G) Frequencies of regulatory T cells (CD4⁺, FoxP3⁺) in *Regnase-3^{fl/fl}* +CD4Cre mice and *Regnase-3^{fl/fl}* controls at 5 mo of age in the thymus (F) and lymph nodes (G), assessed by flow cytometry ($n = 9/9$). Left: Representative blots. Right: Statistics. (H) IgD versus IgM within B cells (CD19⁺) in lymph nodes of *Regnase-3^{-/-}* mice and their *Regnase-3^{+/+}* littermate controls at 6 mo of age, assessed by flow cytometry (representative blot of $n = 6/6$). (I) Frequencies of B cells (B220⁺) and T cells (CD90⁺) in lymph nodes of *Regnase-3^{fl/fl}* +CD19-Cre mice and their *Regnase-3^{fl/fl}* littermates at 5 mo of age (representative flow cytometry blots of $n = 13/13$). Data are represented as mean \pm SEM and were compared by Mann-Whitney U test (**, $P \leq 0.01$; ns, not significant).

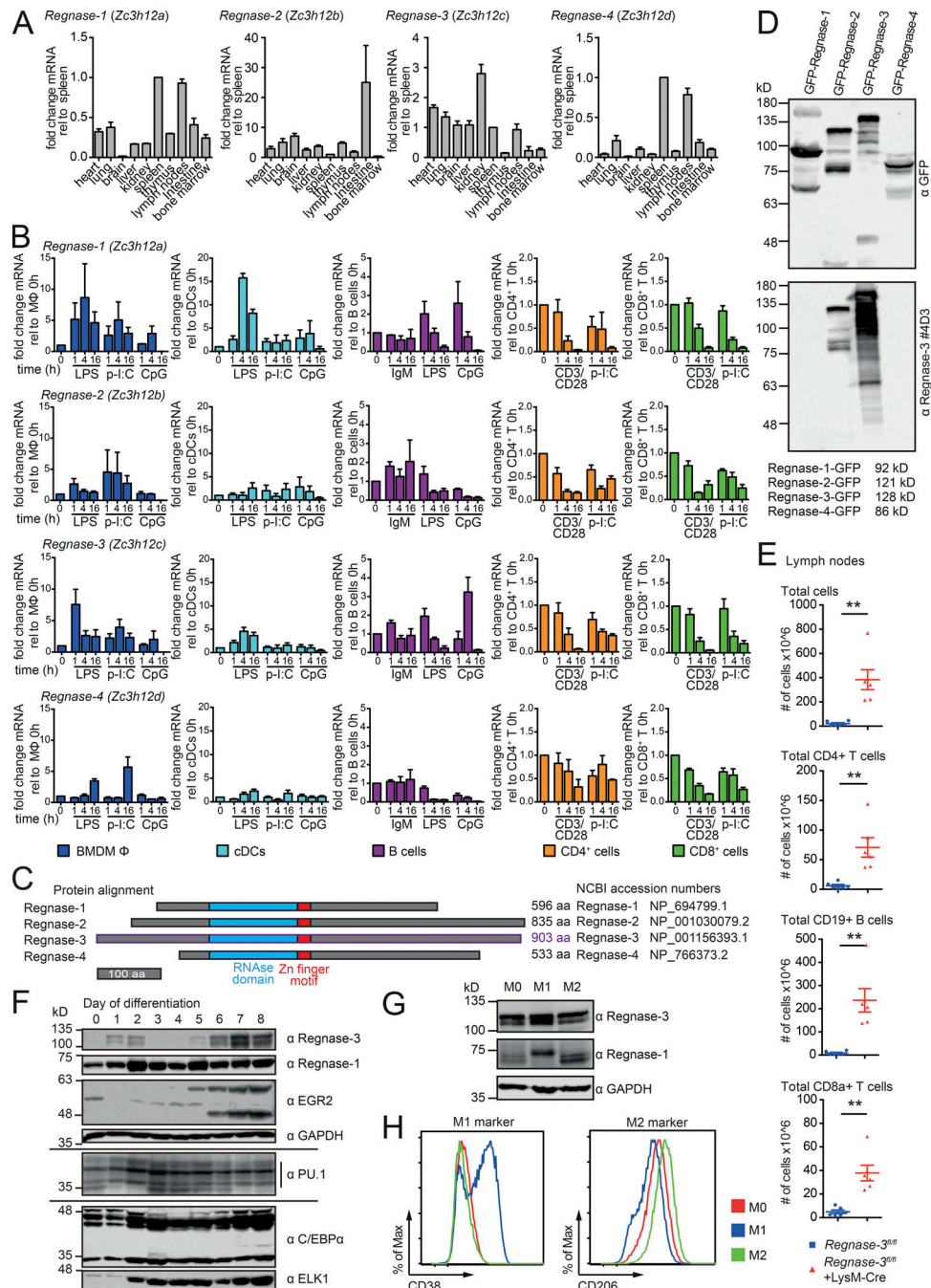


Figure S3. Regnase family member mRNA expression levels in tissues and immune cells. (A) mRNA expression in tissues for *Regnase-1 (Zc3h12a)*, *Regnase-2 (Zc3h12b)*, *Regnase-3 (Zc3h12c)*, and *Regnase-4 (Zc3h12d)* by quantitative RT-PCR, normalized to *Hprt* relative to (relto) their expression in spleen. Mean \pm SEM from three C57BL/6J mice. (B) mRNA expression in immune cells for *Regnase-1-4 (Zc3h12a-d)* by quantitative RT-PCR, normalized to *Hprt* and relative to untreated cells. Mean \pm SEM from three independent experiments: In vitro differentiated BMDMs (Φ) and in vitro differentiated cDCs were treated with LPS (100 ng/ml), low molecular weight poly-I:C (20 μ g/ml), or C-type CpG ODN 2395 (1 μ M) for indicated times. Splenic B cells (B220⁺) were stimulated with anti-IgM (10 μ g/ml), LPS (20 μ g/ml), or C-type CpG ODN 2395 (1 μ M) for indicated times. Splenic CD4⁺ and CD8⁺ T cells were stimulated with plate-bound anti-CD3 and anti-CD28 or low molecular weight poly-I:C (20 μ g/ml) for indicated times. (C) Protein alignment of Regnase family members (National Center for Biotechnology Information accession nos. as indicated). NCBI, National Center for Biotechnology Information. (D) Verification of anti-Regnase-3 antibody (clone 4D3): Regnase family members with N-terminal GFP tag were overexpressed and analyzed by immunoblot. Top: Anti-GFP as loading control. Bottom: Anti-Regnase-3 antibody (clone 4D3). (E) Number of total cells and calculated total B cells (CD19⁺) and CD4⁺ and CD8⁺ T cells in lymph nodes of *Regnase-3^{fl/fl}* +*LysM-Cre* mice and their *Regnase-3^{fl/fl}* littermate controls at 6 mo of age ($n = 6/6$). $**$, $P < 0.01$. (F) ER-Hoxb8 cells were differentiated into macrophages by estrogen withdrawal and addition of recombinant GM-CSF (20 ng/ml). Cells were harvested after each day of differentiation and analyzed by immunoblot (representative blot from $n = 2$ independent experiments). (G) BMDM from C57BL/6J mice were classically activated (M1 condition) with LPS (100 ng/ml) and IFN γ (20 ng/ml), alternatively activated (M2 condition) with IL-4 (20 ng/ml), or left untreated (M0 condition) for 24 h and analyzed by immunoblot (representative blot of three experiments). (H) Shift toward M1 and M2 condition in G was verified with surface markers reported to be up-regulated under M1 condition (CD38) or M2 condition (CD206) by flow cytometry.

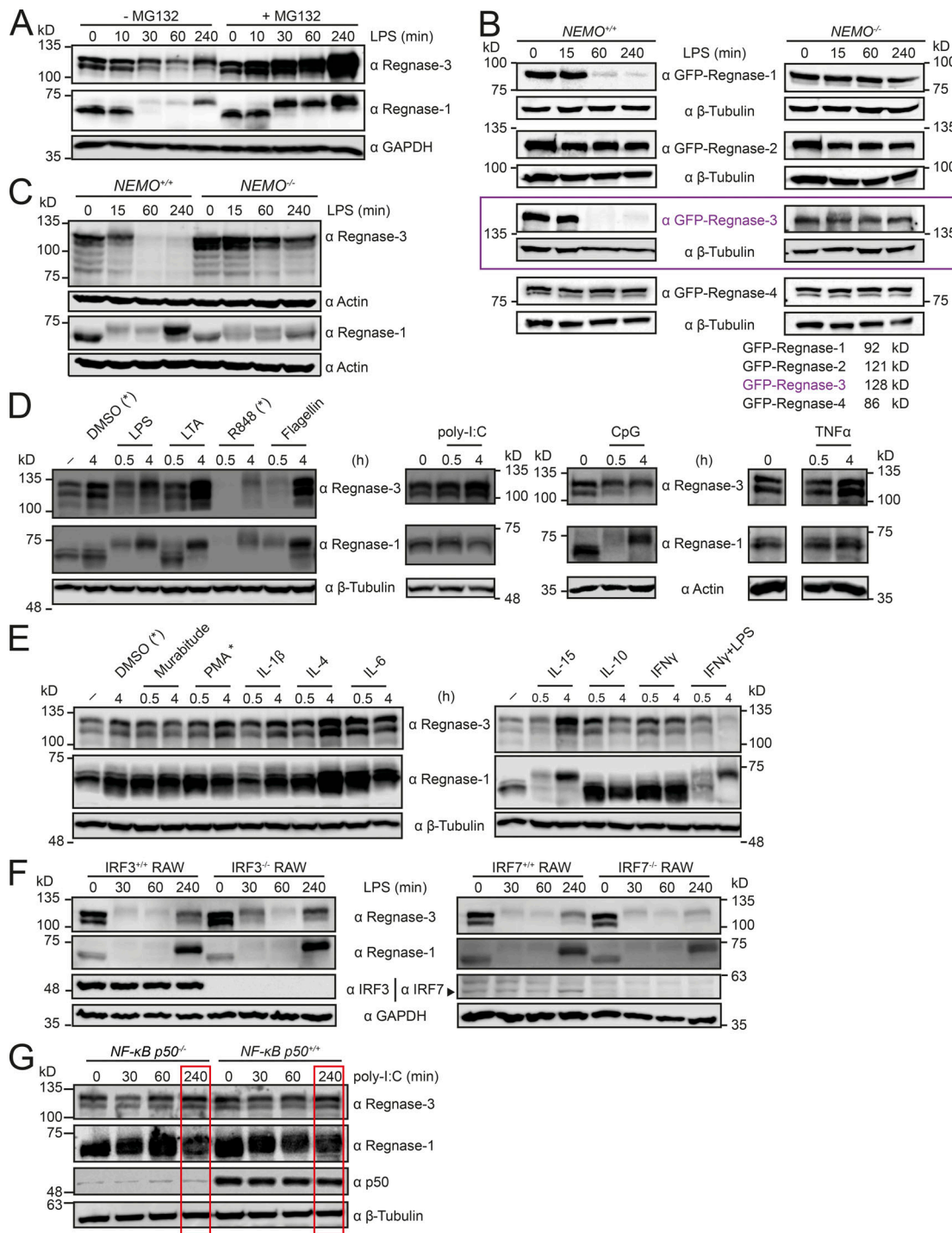


Figure S4. **Regulation of Regnase-1 and -3 protein levels in the presence of cytokines and TLR agonists.** (A) BMDMs from C57BL/6j mice were pre-incubated with MG132 (10 μM) or DMSO (vehicle) for 30 min and then stimulated with LPS (100 ng/ml) for indicated times and analyzed by immunoblot (representative blot of three independent experiments). (B) Constructs of Regnase family members Regnase-1-4 with N-terminal GFP tag were overexpressed in *NEMO*-deficient MEFs or *NEMO*-reconstituted cells, stimulated with LPS (1 μg/ml) for indicated times, and analyzed by immunoblot (representative blot of three independent experiments). (C) *NEMO*-deficient MEFs or *NEMO*-reconstituted cells were stimulated with LPS (1 μg/ml) for indicated timepoints and analyzed by immunoblot (representative blot of three independent experiments). (D and E) BMDMs from C57BL/6j mice were left untreated, were treated with vehicle (compounds dissolved in DMSO are marked with asterisk), or were stimulated with LPS (100 ng/ml), LTA (1 μg/ml), R848 (Resiquimod; 1 μg/ml), Flagellin (100 ng/ml), high molecular weight poly-I:C (10 μg/ml), C-type CpG ODN 2395 (1 μg/ml), TNFα (50 ng/ml), Murabutide (100 ng/ml), PMA (25 ng/ml), IL-1β (50 ng/ml), IL-4 (50 ng/ml), IL-6 (50 ng/ml), IL-15 (50 ng/ml), IL-10 (50 ng/ml), IFNγ (50 ng/ml), or IFNγ plus LPS (50 and 100 ng/ml) for 0.5 or 4 h and analyzed by immunoblot (representative blots of three experiments). (F) *IRF3*^{-/-} or *IRF7*^{-/-} deficient RAW cells and wild type RAW cells were stimulated with LPS (100 ng/ml) for indicated times and analyzed by immunoblot (representative blot of three independent experiments). (G) BMDMs from *NF-κB p50*^{+/+} and *NF-κB p50*^{-/-} mice were stimulated with high molecular weight poly-I:C (μg/ml) for indicated times and analyzed by immunoblot (representative blot from two mice).

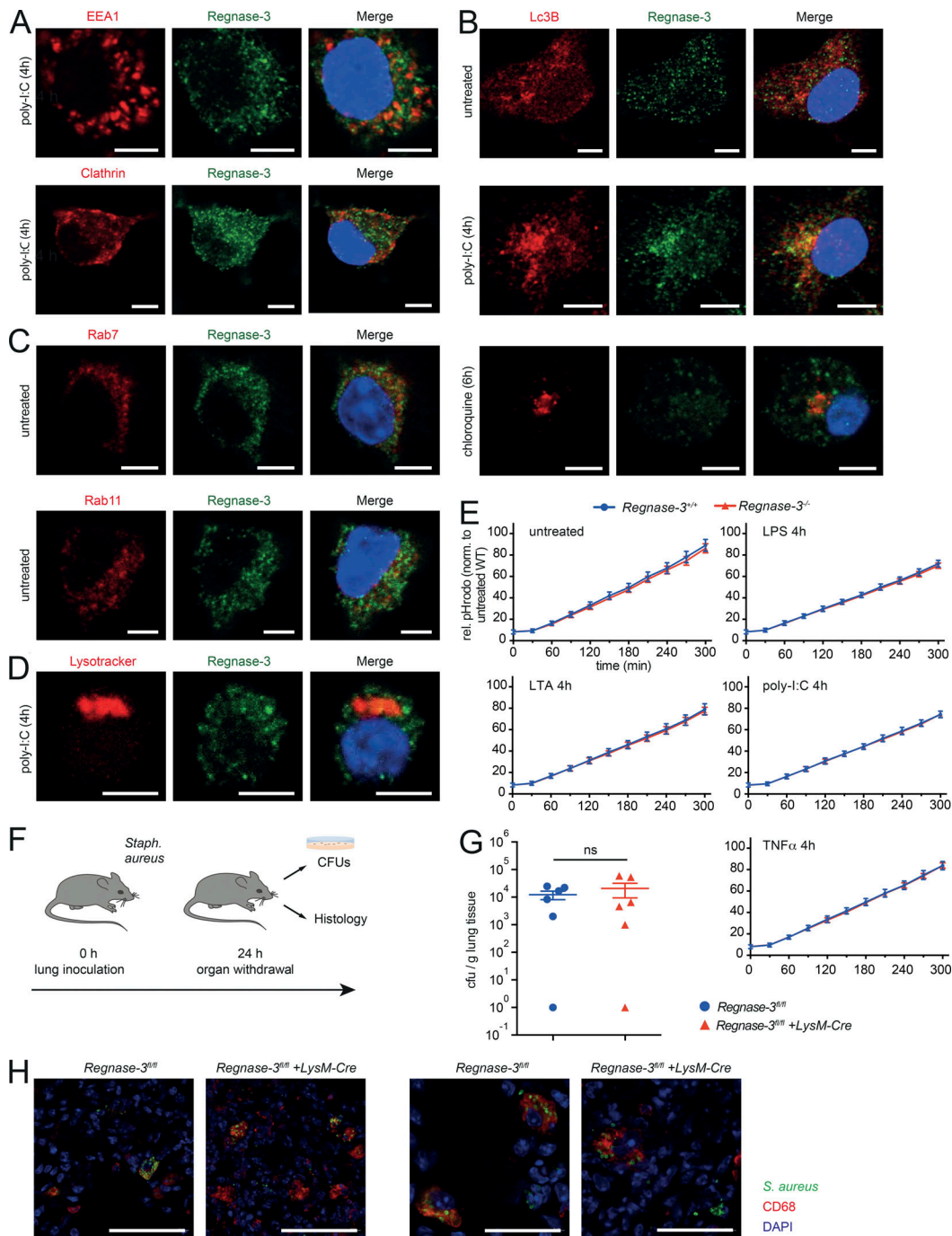


Figure S5. Subcellular localization and effects of Regnase-3 on phagocytosis. (A–D) Confocal microscopy images. BMDMs from C57BL/6J mice were pretreated with high molecular weight poly-I:C (10 μ g/ml) for 4 h, autophagy inhibitor chloroquine (25 μ M) for 6 h, or left untreated, as indicated. Red and green as indicated; counterstain for nuclei with DAPI in blue. Representative images of three or more independent experiments are shown. Bars, 5 μ m. **(A)** BMDMs from C57BL/6J mice were immunostained for Regnase-3 and EEA1 (early endosomes) or Clathrin (Clathrin-mediated endocytosis). **(B)** Immunostaining for autophagosome marker Lc3B and Regnase-3. **(C)** Immunostaining for Regnase-3 and endosomal markers Rab7 or Rab11. **(D)** Lysosomes in poly-I:C-treated BMDMs were labeled with LysoTracker dye for 30 min, fixed, and immunostained for Regnase-3. **(E)** BMDMs from *Regnase-3^{+/+}* and *Regnase-3^{-/-}* mice were left untreated or were preincubated for 4 h with LPS (100 ng/ml), LTA (1 μ g/ml), high molecular weight poly-I:C (10 μ g/ml), or TNF α (50 ng/ml). Subsequently, *S. aureus* bioparticles coupled to pH-sensitive pHrodo fluorophore were subjected to cells, and the increase in fluorescence was acquired over time with a microplate reader. Maximal fluorescence of unstimulated *Regnase-3^{+/+}* BMDMs in each assay was set to 100% (mean \pm SEM from four independent assays with BMDMs from two mice in each assay; total $n = 8/8$). Norm., normalized; rel., relative. **(F)** Schematic representation of in vivo pneumonia model. The lungs of *Regnase-3^{fl/fl}* + *LysM-Cre* and *Regnase-3^{fl/fl}* littermate controls were inoculated with MRSA. 24 h after inoculation, organs were withdrawn to determine CFUs and to perform immunohistochemical analysis. **(G)** In vivo pneumonia model. CFUs in *Regnase-3^{fl/fl}* + *LysM-Cre* and *Regnase-3^{fl/fl}* littermate controls. **(H)** Immunofluorescence stains of lungs from pneumonia model. Lung sections were stained for *S. aureus* (green) and macrophages with anti-CD68 (red) and counterstained for nuclei with DAPI (blue). Images were acquired by confocal microscopy. Representative images are shown. Bars: 50 μ m (left); 25 μ m (right). Data are represented as mean \pm SEM and were compared by Mann-Whitney *U* test. ns, not significant.