

Fig. S1. (A) Community similarity between samples and between DNA and RNA, comparing 18S cloning vs. high-throughput sequencing (HTS). Bray-Curtis tree of the 18S cloning (99% OTUs) and HTS (98% OTUs) diversity (i.e.: richness + evenness) by sample. DNA (black text) or RNA (grey text), as well as surface (surf; blue) or SCM samples (shades of green), are indicated. Note that no RNA remained for HTS analysis of PP-2. The scale bar equals 10% dissimilarity. **(B)** Proportions of shared HTS OTUs (lower triangle) and reads (upper triangle) between each pairwise sample combination.

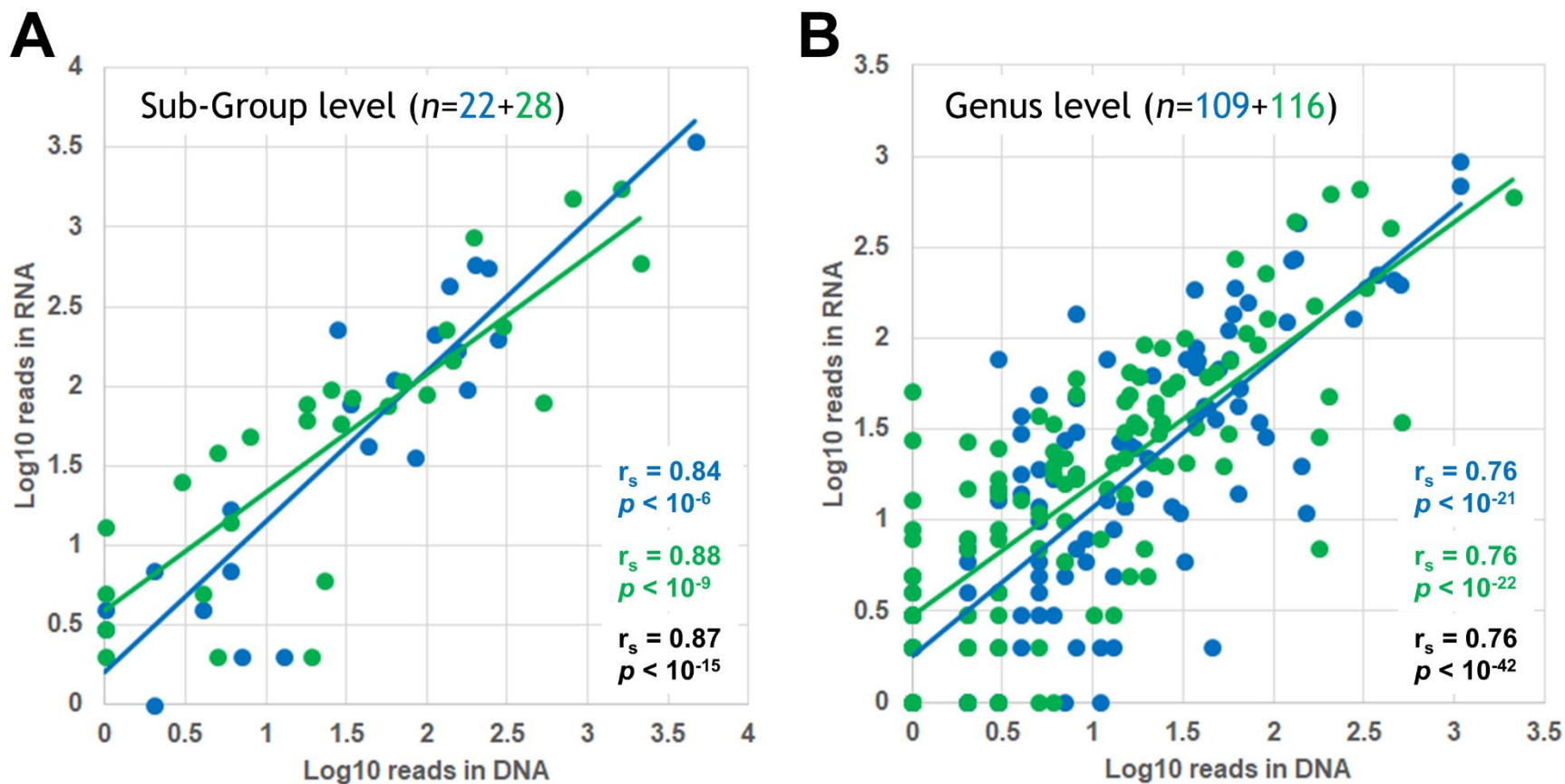


Fig. S2. Correlations of log-scale taxa high-throughput sequencing (HTS) reads between DNA and RNA samples at the coarser Sub-Group level (**A**) and at the finer genus level (**B**). Only CB-15 surface (blue) and CB-15 SCM (green) samples are presented as no RNA remained for HTS analysis of station PP-2. Listed are the Spearman r_s and p -values for the correlations of each individual sample (in matching colors) and for the overall trend with both samples combined (in black).

Table S1. Sample and station characteristics (“-” indicates missing data) of screened public Arctic metagenome studies.

	AGulf	JGI Sea of Change	JGI W. Arctic Ocean	UAF GEOTRACES
Region(s)	AGulf	NS+GS	CB	CB+AO
# of samples	1	6	8	17
# of stations	1	6	4	9
Dates of sampling	Dec.2007	Jun.-Jul.2012	Oct.2015	Aug.-Oct.2015
Range of latitudes (°N)	71.62	69.23 to 79.02	73.22 to 79.25	72.80 to 89.99
Range of longitudes (°E)	-126.07	-9.52 to 9.86	-150.23 to -150.06	-180.00 to -89.25
Depths of sampling (m)	10	10-20	5-79	1-100
Size fraction	0.2-50 µm	1.2-100 µm	0.2-3.0 µm	>0.22 µm
Accession #	PRJNA337945	JGI Proposal 532	JGI Proposal 2011	PRJEB14154
Reference	(1)	(2)	(3)	(4)
<i>Ranges of physicochemical characteristics</i>				
Salinity (psu)	30.56	31.03-35.15	25.70-31.48	27.41-36.68
Temperature (°C)	-1.60	-1.03 to +9.10	-1.46 to +0.02	-1.67 to +1.64
Nitrate (µM)	-	bd-6.6	bd-4.7	bd-17.5
Phosphate (µM)	-	0.46-0.88	0.52-1.11	-
Silicate (µM)	-	1.5-5.5	2.4-10.8	-
<i>Sequence characteristics</i>				
Average # reads screened/sample (M)	314	372	200	5.5
Range # reads screened/sample (M)	314	327-450	161-219	2.0-11.1
Total # reads screened (M)	314	2,234	1,599	93
Total # assembled GenSeed-HMM contigs	1	87	64	96
Total # of target plankton NR contigs	1	55	40	15
Total # of non-target eukaryote contigs	0	23	7	22
Total # of non-target prokaryote contigs	0	5	16	53
Average % identity of target NR matches	100%	82%	85%	79%
Range % identity of target NR matches	100%	45-100%	53-100%	51-99%

AGulf, Amundsen Gulf; AO, Arctic Ocean; bd, below detection; CB, Canada Basin; GS, Greenland Sea; JGI, Joint Genome Institute; NR, nitrate reductase; NS, Norwegian Sea; UAF, University of Alaska - Fairbanks.

1. Joli N, Monier A, Logares R, Lovejoy C. 2017. Seasonal patterns in Arctic prasinophytes and inferred ecology of Bathycoccus unveiled in an Arctic winter metagenome. *ISME J* 11:1372-1385.
2. Mock T. 2014. Sea of Change: Eukaryotic phytoplankton communities in the Arctic Ocean. JGI: <https://genome.jgi.doe.gov/portal/SeaofArcticOcean/SeaofArcticOcean.info.html>
3. Walsh D. 2016. Metagenomics of western Arctic Ocean microbial communities. JGI: <https://genome.jgi.doe.gov/portal/Metofommunities/Metofommunities.info.html>
4. Collins E. 2016. Arctic Ocean metagenomes sampled aboard CGC Healy during the 2015 GEOTRACES Arctic research cruise. NCBI: <https://www.ncbi.nlm.nih.gov/bioproject/PRJEB14154>

Table S3. Primers used for cloning and (pyro)sequencing of 18S rRNA and nitrate reductase (NR) genes

Assay/use	Primer ^a	Product size	Sequence (5'-3') ^a	Source
18S V4 pyrosequencing	A#-E572F B-E1009R	508 bp	(RocheA+MID#) CYGCGGTAATTCAGCTC (RocheB) AYGGTATCTRATCRTCCTTYG	(1)
18S cloning – PCR	NSF4/18 EukR	~1800 bp	CTG GTT GAT YCT GCC AGT TGA TCC TTC TGC AGG TTC ACC TAC	(2) (3)
18S cloning – sequencing	Euk528f	n/a	GCG GTA ATT CCA GCT CCA A	(4)
NR cloning – 1 st round	NRPt907F NRPt2325R	1419 bp	GGY GGN MGN ATG RTB AAG TGG CT GGG VGT GAT RCC HGT VCC DCC VGC	(5)
NR cloning – 2 nd round	NRPt1000F NRPt1389R	390 bp	GRD GGH TGG TGG TAC AAG CC GTT GTT YMY CAT BCC CAT	(5)

^a Pyrosequencing forward primers exist in 12 numbered versions (#), differing only by the 12 individual multiplex identifiers (MID = bar-codes). Also incorporated into the primers were the Roche A (forward) and B (reverse) adaptors.

1. Comeau AM, Li WKW, Tremblay J-E, Carmack EC, Lovejoy C. 2011. Arctic Ocean microbial community structure before and after the 2007 record sea ice minimum. *PLoS ONE* 6:e27492.
2. Hendriks L, Goris A, Neefs JM, Vandeppeer Y, Hennebert G, Dewachter R. 1989. The nucleotide sequence of the small ribosomal-subunit RNA of the yeast *Candida albicans* and the evolutionary position of the fungi among the Eukaryotes. *Syst Appl Microbiol* 12:223-229.
3. Medlin LK, Elwood HJ, Stickel S, Sogin ML. 1988. The characterization of enzymatically amplified eukaryotic 16S-like rRNA-coding regions. *Gene* 71:491-499.
4. Elwood HJ, Olsen GJ, Sogin ML 1985. The small-subunit ribosomal RNA gene sequences from the hypotrichous ciliates *Oxytricha nova* and *Stylonychia pustulata*. *Mol Biol Evol* 2:399-410.
5. Allen AE, Ward BB, Song B. 2005. Characterization of diatom (Bacillariophyceae) nitrate reductase genes and their detection in marine phytoplankton communities. *J Phycol* 41:95-104.