Myeloid hypoxia inducible factor- 1α is essential for skeletal muscle

regeneration in mice

Supplementary material

Supplementary figure 1

HIF-1α in skeletal muscle cells did not influence the acute severity of trauma and was

not required for adequate skeletal muscle regeneration. (A and B) Neither the extent

of traumatic lesion nor the serum creatine kinase levels differed significantly between

HIF- $1\alpha^{+f/+f}$ and MCKCre/HIF- $1\alpha^{+f/+f}$ mice. (**C** and **D**) There were neither differences in

the cross-sectional area of regenerating myofibres nor in the extent of necrotic cell

debris at 7d after injury between HIF- $1\alpha^{+f/+f}$ and MCKCre/HIF- $1\alpha^{+f/+f}$ mice. Each group

included four to five mice. Data are mean ± SEM. XSA=cross-sectional area.

Supplementary figure 2

Myeloid-specific knock-out of VEGF did neither impair regeneration of muscle tissue

nor of vasculature after skeletal muscle injury. (A) The extent of necrotic cell debris

was not changed in mice with a VEGF knock-out in myeloid cells. (B) Numbers of

CD34-positive blood-endothelial cells were not decreased at the injury site of

LysMCre/VEGF+f/+f mice. Each group included five to six mice. Data are mean ±

SEM.

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Supplementary figure 3

Myeloid cells, isolated from injured skeletal muscle, showed high purity and high deletion efficiency for HIF-1 α . **(A)** The purity of isolated myeloid cells accounted more than 80 % and was examined by FACS-analysis with a CD11b antibody. **(B)** The HIF-1 α deletion efficiency of myeloid cells, isolated from LysMCre/HIF-1 α ^{+f/+f} mice, was approximately 75 %. Each group included five to six mice. Data are mean \pm SEM.

Supplementary figure 4

Macrophages of injured skeletal muscles of HIF- $1\alpha^{+f/+f}$ and LysMCre/HIF- $1\alpha^{+f/+f}$ mice showed a M2-phenotype. (**A** and **B**) Macrophages, isolated from injured skeletal muscles, showed approximately 100 times higher expression of arginase-1 mRNA than iNOS mRNA. (**C**) Representative iNOS- and YM-1-immunostaining. Both wild-type and HIF- 1α -deficient macrophages showed low expression of the M1-marker iNOS, but high expression of the M2-marker YM-1. Each group included five to nine mice. Scale bars: $50\mu m$. Data are mean \pm SEM. YM-1=T-lymphocyte-derived eosinophil chemotactic factor.



















