*In vitro* seeding activity of glycoform-deficient prions from variably protease-sensitive prionopathy and familial CJD associated with PrP<sup>V180I</sup> mutation

Zerui Wang<sup>1,2</sup>, Jue Yuan<sup>2</sup>, Pingping Shen<sup>1</sup>, Romany Abskharon<sup>3</sup>, Yue Lang<sup>1,2</sup>, Johnny Dang<sup>2</sup>,

Alise Adornato<sup>2</sup>, Ling Xu<sup>2</sup>, Jiafeng Chen<sup>1</sup>, Jiachun Feng<sup>1</sup>, Mohammed Moudjou<sup>4</sup>, Tetsuyuki

Kitamoto<sup>5</sup>, Jan Langeveld<sup>6</sup>, Brian Appleby<sup>2,7,8</sup>, Jiyan Ma<sup>3</sup>, Qingzhong Kong<sup>2,7,8</sup>, Robert B.

Petersen<sup>2,9\*</sup>, Wen-Quan Zou<sup>1,2,7,8,10\*</sup>, Li Cui<sup>1\*</sup>

<sup>1</sup>Department of Neurology, The First Hospital of Jilin University, Changchun, Jilin Province, the People's Republic of China.

<sup>2</sup>Department of Pathology, Case Western Reserve University School of Medicine, Cleveland, Ohio, USA.

<sup>3</sup>Center for Neurodegenerative Science, Van Andel Research Institute, Grand Rapids, 49503 Michigan, USA.

<sup>4</sup>Virologie Immunologie Moléculaires, INRA, Jouy-en-Josas, France.

<sup>5</sup>Center for Prion Diseases, Tohoku University Graduate School of Medicine, 2-1 Seiryo-machi, Aoba-ku, Sendai 980-8575, Japan.

<sup>6</sup>Wageningen BioVeterinary Research, Houtribweg 39, Lelystad, the Netherlands.

<sup>7</sup>National Prion Disease Pathology Surveillance Center, Case Western Reserve University School of Medicine, Cleveland, Ohio, USA.

<sup>8</sup>Department of Neurology, Case Western Reserve University School of Medicine, Cleveland, Ohio, USA.

<sup>9</sup>Foundation Sciences, Central Michigan University College of Medicine, Mount Pleasant, Michigan, USA.

<sup>10</sup>State Key Laboratory for Infectious Disease Prevention and Control, National Institute for Viral Disease Control and Prevention, China Center for Disease Control and Prevention, Beijing, China.

\*Corresponding: WQZ at <u>wxz6@case.edu</u>; or LC at <u>chuili1967@126.com</u>; or RBP at <u>robert.petersen@cmich.edu</u>

## Table S1

Table S1 ANOVA analysis			
Groups	Comparison	p values	
Fig. 2b	Seeds	>0.05	
	Substrates	>0.05	
	Interaction	<0.0005	
Fig. 2e	Seeds	>0.05	
	Substrates	<0.0001	
	Interaction	<0.0001	
Fig. 3c	Seeds	<0.0005	
	Substrates	<0.0001	
	Interaction	<0.0001	
Fig. 3g	Seeds	<0.0001	
	Substrates	<0.0001	
	Interaction	<0.0001	

Fig. S1



Fig. S1 Schematic diagram of locations of the 1E4 and 3F4 epitopes on PrP and structure of human PrP. The mature human PrP contains 209 amino acids from residues 23 to 231 with an unstructured N-terminal domain from residues 23 to 127 and a structured C-terminal domain from residues 128 to 231. The 1E4 and 3F4 epitopes are next to each other, localizing in the unstructured domain between residues 97 and 105 for the 1E4 epitope and between residues 106 and 112 for the 3F4 epitope. In the structured C-terminal domain, there are two short  $\beta$ -sheets (PrP128-131 for  $\beta$ 1 and PrP161-164 for  $\beta$ 2) and three  $\alpha$ -helixes (PrP144-154 for  $\alpha$ 1, PrP173-194 for  $\alpha$ 2 and PrP200-228 for  $\alpha$ 3) [26]. There are two N-linked glycosylation sites at residues 181 and 197 (black quad arrow callouts) and a GPI anchor at the residue 231 (black circular arrow) [27, 28].



Fig. S2 Comparison of gel profile of the PrP molecule from brain homogenates of humanized TgMM, Tg180 and TgVV mice. Representative Western blotting of PrP from brain homogenates of humanized Tg mice expressing human wild-type PrP with 129MM polymorphism (TgMM), PrP<sup>V180I</sup> mutation (Tg180), or wild-type PrP with 129VV (TgVV) treated with or without digestion of PK at 50 µg/ml. The blot was probed with 3F4. Molecular weight markers are shown in kDa on the left side of the blots.

Fig	۱.	<b>S</b> 3



Fig. S3 Serial PMCA of PrP<sup>Sc</sup> from VPSPr and fCJD<sup>V180I</sup> in human or Tg mouse brain homogenate substrate. (a) Representative Western blotting of PrP<sup>Sc</sup> amplified with 4-6 rounds of sPMCA by seeding PrP<sup>Sc</sup> from VPSPrMM, VPSPrVV, VPSPrMV, fCJD<sup>V180I</sup>, and fCJD<sup>T183A</sup> in the hMM or hVV brain substrate. (b, c) Representative Western blotting of PrP<sup>Sc</sup> amplified with 4-6 rounds of sPMCA by seeding PrP<sup>Sc</sup> from VPSPrMM and VPSPrVV (b) as well as VPSPrMV and fCJD<sup>V180I</sup>MM (c) in humanized transgenic mouse brain homogenates from TgVV, TgMM, Tg180, or TgMM + Tg180 mouse lines probed with the

Tohoku 2 antibody. sPMCA-R: sPMCA rounds. Molecular weight markers are shown in kDa on the left side of the blots.

## Fig. S4



Fig. S4 Serial PMCA of PrP<sup>Sc</sup> from VPSPr and fCJD<sup>V1801</sup> in human brain homogenates or humanized Tg mouse brain homogenates. Representative Western blotting of PrP<sup>Sc</sup> from VPSPrMV, VPSPrMM, VPSPrVV, or fCJD<sup>V1801</sup> amplified with 4-6 rounds of sPMCA in human brain homogenates from non-CJD MM (hMM) or VV (hVV) or mouse brain homogenates from humanized TgVV mice probed with the Bar209 (**a**) or V14 (**b**) antibody. sPMCA-R: sPMCA rounds. Molecular weight markers are shown in kDa on the left side of the blots. PK: Proteinase K.