## **Supplementary Data for:**

## Multiple Pleomorphic Tetramers of Thermostable Direct Hemolysin from *Grimontia hollisae* in Exerting Hemolysis and Membrane Binding

Yu-Kuo Wang<sup>a</sup>, Sheng-Cih Huang<sup>a</sup>, Chin-Yuan Chang<sup>a</sup>, Wan-Ting Huang<sup>a</sup>, Man-Jun Liao<sup>a</sup>, Bak-Sau Yip<sup>c</sup>,

Feng-Pai Chou<sup>a</sup>, Thomas Tien-Hsiung Li<sup>d\*</sup>, and Tung-Kung Wu<sup>a,b\*</sup>

<sup>a</sup>Department of Biological Science and Technology, National Chiao Tung University, Hsin-Chu 30010,

Taiwan, Republic of China

<sup>b</sup>Center for Emergent Functional Matter Science, National Chiao Tung University, 1001 Ta-Hsueh Rd.,

Hsinchu 30010, Taiwan, Republic of China

<sup>c</sup>Department of Neurology, National Taiwan University Hospital, Hsin-Chu, 30059, Taiwan, Republic of

China

<sup>d</sup>Graduate Institute of Biochemistry, National Chung Hsing University, Taichung, 40227 Taiwan, Republic

of China

\* To whom correspondence should be addressed:

E-mail: tkwmll@mail.nctu.edu.tw (T. K. Wu)

Fax: +886-3-5725700

Tel: +886-3-5712121-56917

E-mail: lithomas@dragon.nchc.edu.tw (Thomas T. H. Li)

Fax: +886-4-22853487

Tel: +886-4-22840468

**Fig. S1** Superposition of the overall structures of the TDH tetramers (Gh–TDH Oligomer–I) between *G*. *hollisae* (green, PDB entry 4WX3) and *V. parahaemolyticus* (red, PDB entry 3A57). The spheres colored in orange marked the position of the *N*-terminal  $Gly^{12}$  for each TDH monomer. The Vp–TDH tetramer were generated by applying crystallographic symmetries (Space group *I*<sub>4</sub>) on the coordinates of the published Vp–TDH monomeric structure.



**Fig. S2.** Illustration of residues of Gh–TDH involved in protomer–protomer interactions. (A) The crystal packing pattern corresponds to the Gh–TDH–I oligomer, and protomer–protomer interactions in (B) Oligomer–I, (C) Oligomer–II, and (D) Oligomer–III.





Fig. S3. Flow cytometry analysis of erythrocytes binding of wild-type and mutated Gh-TDHs.

**Fig. S4A.** Illustrations of the analysis of the topology of Gh–TDH Oligomer–I, –II , and –III using program CAVER <sup>50</sup>. The narrowest part of the side–channels were marked with measurements.



Fig. S4B. Top-view of Oligomer-I from the N-terminus direction and of Oligomer-II and -III.



Figure S4C. Bottom-view of Gh-TDH Oligomer-I from the C-terminus direction and of Oligomer-II and – III.



Fig. S5. Sequence alignment of Vp–TDH and Gh–TDH.

G.hollisae_TDH V.parahaemolyticus_TDH2	FELPSIPFPSPGSDEILFVVRDTTFNTKEPVNVKVSDFWTNRNVKRKPYK FELPSVPFPAPGSDEILFVVRDTTFNTNAPVNVEVSDFWTNRNVKRKPYK ****:***:****************************	50 50
G.hollisae_TDH V.parahaemolyticus_TDH2	DVYGQSVFTTSGSKWLTSYMTVSINNKDYTMAAVSGYKDGFSSVFVKSGQ DVYGQSVFTTSGTKWLTSYMTVNINDKDYTMAAVSGYKHGHSAVFVKSDQ ************************************	100 100
G.hollisae_TDH V.parahaemolyticus_TDH2	IQLQHYYNSVADFVGGDENSIPSKTYLDETPEYFVNVEAYESGSGNILVM VQLQHSYDSVANFVGEDEDSIPSKMYLDETPEYFVNVEAYESGSGNILVM :**** *:***:*** **:**** **************	150 150
G.hollisae_TDH V.parahaemolyticus_TDH2	CISNKESYFECESQQ 165 CISNKESFFECKHQQ 165 ******:**: **	

**Fig. S6.** Analytical ultracentrifugation analysis (AUC) results of Gh–TDH<sup>WT</sup> and various Gh–TDH<sup>mut</sup> proteins. (A) Gh–TDH<sup>WT</sup>; (B) Gh–TDH<sup>Y53D</sup>; (C) Gh–TDH<sup>F159D</sup>; (D) Gh–TDH<sup>R46E</sup>; (E) Gh–TDH<sup>Y53D/F159D</sup>; (F) Gh–TDH<sup>D78A</sup>; (G) Gh–TDH<sup>K97A</sup>; (H) Gh–TDH<sup>N108A</sup>.



(A) Gh-TDH<sup>WT</sup>





(E) Gh-TDH<sup>Y53D/F159D</sup>





0.000002

0.000000

-0.000002

Ò

100000

±30.42082 15997.06384 ±60.73138 0.26194 ±0.00086 Α

400000

500000

300000

500000

Molecular Mass (Da)

200000

## Table S1

MAD data <sup>a</sup>	(	Gh-TDH-I oligome	r	Gh-TDH-II oligomer					
	peak	inflection	remote	peak	inflection	remote			
space group		P21212			P2 <sub>1</sub> 2 <sub>1</sub> 2				
cell dimension (Å)		105.3, 112.6, 60.8	3		105.3, 112.7, 60.9				
		α, β, γ = 90º			α, β, γ = 909				
wavelength	0.9789	0.9791	0.9639	0.9790	0.9792	0.9638			
resolution (Å)	30-1.79	50-1.79	30-1.70	30-2.30	30-2.30	30-2.40			
	(1.73-1.79)	(1.73-1.79)	(1.70-1.76)	(2.38-2.30)	(2.38-2.30)	(2.38-2.40)			
total reflections	700,387	345,485	359,546	263,840	132,647	133,719			
unique reflection	71,759	70,365	73,925	32,500	32,652	32,462			
R <sub>merge</sub>	0.077(0.597)	0.073(0.643)	0.064(0.570)	0.083(0.264)	0.080(0.226)	0.068(0.222)			
redundancy	9.8 (8.7) <sup>b</sup>	4.9 (4.3)	4.9 (4.1)	8.1 (7.2)	4.1 (3.7)	4.1 (3.8)			
l/σ(l)	5.1 (4.8)	4.2 (4.1)	4.3 (4.5)	21.8 (6.7)	14.2 (5.9)	17.0 (6.3)			
data completeness (%)	94.4 (93.6)	92.4 (82.5)	92.2 (79.3)	99.6 (97.9)	99.5 (97.4)	99.5 (98.3)			
refinement statics									
resolution (Å)		23.85 - 1.70			26.77 – 2.30				
No. reflections		71,897		32,609					
R <sub>work</sub> /R <sub>free</sub> (%)		20.11 / 23.36		19.34 / 24.07					
protein atoms		4,817		4,854					
waters		240		184					
B <sub>factor</sub> (Ų)									
protein		26.935		31.334					
water		29.428		32.134					
R.M.S. deviation (Å)									
bond lengths (Å)		0.005		0.007					
bond angles (°)		0.847		1.008					
Ramanchandran favored (%)		96.6%		97.25%					
MolProbity score <sup>e</sup> (100 <sup>th</sup> percentile)		0.87			0.95				
All-atom clash score <sup>f</sup> (100 <sup>th</sup> percentile)	0.43 1.06								

a. The multiple anomalous dispersion (MAD) X-ray diffraction data were named based on the Gh-TDH tetrameric structure determined in the crystallographic asymmetric unit.

b. Values in parentheses are parameters for high-resolution shell.

c.  $R_{merge} = \sum_{hkl} (\sum_{i} (|I_{hkl,i}-\langle I_{hkl} \rangle|)) / \sum_{hkl,i} \langle I_{hkl} \rangle$ , where  $I_{hkl,i}$  is the intensity of the reflection with Miller indices h, k and l, and  $\langle I_{hkl} \rangle$  is the mean intensity of the reflection.

d.  $R_{work} = \sum_{hkl} (||Fobs_{hkl}| - |Fcalc_{hkl}||)/|Fobs_{hkl}|$ , where  $|Fobs_{hkl}|$  and  $|Fcalc_{hkl}|$  are the observed and calculated structure factor amplitudes.  $R_{free}$  is calculated with statistically selected ~2000 reflections omitted from the refinement process.

e. MolProbity score is defined as the following:

0.42574\*log(1+clashscore)+0.32996\*log(1+max(0,pctRotOut-1))+0.24979\*log(1+max(0,100-pctRamaFavored-2))+0.5 100<sup>th</sup> percentile is the best among structures of comparable resolution; 0<sup>th</sup> percentile is the worst.

f. The clashscore is the number of serious overlaps of non-donor-acceptor atoms by more than 0.4 Å per 1000 atoms.

**Table S2.** Molecular mass determination of Gh–TDH<sup>WT</sup> and various Gh–TDH<sup>mut</sup> proteins.

Mw. (kDa)	WT	Y53D	F159D	R46E	Y53D/ F159D	D78A	K97A	S98A	Q104A	N108A	Y126A	E129A
SDS PAGE	22	22	22	22	22	22	22	22	22	22	22	22
Native- PAGE	90~140	72	72	66	66	90~140	70~95	70~95	70~95	70~95	70~95	70~95
AUC	74.54 ± 0.007	33.09 ± 0.07	28.28 ± 0.15	18.86 ± 0.007	21.88 ± 0.05	72.18 ± 0.02	65.88 ± 0.003	N.D.	N.D.	71.30 ± 0.03	N.D.	N.D.

N. D. = not determined