

Structure of the super-elongation complex subunit AFF4 C-terminal homology domain reveals requirements for AFF homo- and heterodimerization

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Supplementary information

Supplementary Tables: Table S1-S2

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Supplementary Tables

Supplementary Table S1, Primer list and sequences

Nucleotide sequences used in fluorescence anisotropy

Name	Sequence (5'-3')
FBS-35mer G-quadruplex RNA	6-FAM-AAGAAGAGGACAAGGAGGAAGAGGACGGGAGGAGGCUUC
HIV1 LTR-III G-quadruplex DNA	6-FAM-GGGAGGCGTGGCCTGGGCGGGACTGGGG
Unstructured TAR RNA	6-FAM- UUAAGGAAUUAAGUCGUGCGUCUAAUAACCGGAGAGGGAACCCACU
Stem-loop- TAR RNA	6-FAM- ACCAGAUCUGAGCCUGGGAGCUCUCUGGCUAACUAGGGAACCCACU

Primers use to clone AFF4-CHD and AFF1-CHD to protein expression vector

Name	Sequence (5'-3')	Description
AFF4-899- LICv1-Fw	TACTTCCAATCCAATGCATCTAAGCCTCGGAGAACAAA GCTTG	His ₆ -tagged AFF4-CHD
AFF4-1163- LICv1-Rev	TTATCCACTTCCAATGTTATTAAGATATCAACTTGGCAT CCTGGC	
AFF1-938- LICv1-Fw	TACTTCCAATCCAATGCAAATCCTTTTCCAGTGCCTTCTT TGC	His ₆ -MBP- tagged AFF1- CHD
AFF1-1210- LICv1- Rev	TTATCCACTTCCAATGTTATTAAGGTGTTTTGGTTAATTC TTGTAGCTGCTG	

Primers used to generate mutants of AFF4-CHD

Name	Sequence (5'-3')	Description
AFF4- H1090A-Fw	TTCAGTGACCATTCCACAGAAGATCGCACAGATGGCAGC CAGC	His ₆ -tagged AFF4- CHD-5MA mutant
AFF4- H1090A-Rev	GCTGGCTGCCATCTGTGCGATCTTCTGTGGAATGGTCACT GAA	

Name	Sequence (5'-3')	Description
AFF4-YV1096,1607 AA-Fw	CAGATGGCAGCCAGCGCAGCACAGGTCACATCCAAC	
AFF4-YV1096,1607 AA-Rev	GTTGGATGTGACCTGTGCTGCGCTGGCTGCCATCTG	
AFF4-FL1103,1134 AA-Fw	CAGGTCACATCCAACGCAGCATATGCCACCGAAATTTGGGACCAAGC	
AFF4-FL1103,1134 AA-Rev	GCTTGGTCCCAAATTTTCGGTGGCATATGCTGCGTTGGATGTGACCTG	
AFF4-H1090D-Fw	AGTGACCATTCCACAGAAGATCGATCAGATGGCAGCCAGC	His ₆ -tagged AFF4-CHD-5MD mutant
AFF4-H1090D-Rev	GCTGGCTGCCATCTGATCGATCTTCTGTGGAATGGTCACT	
AFF4-YV1096,1097 DD-Fw	CAGATGGCAGCCAGCGATGATCAGGTCACATCCAACGATGATTAT	
AFF4-YV1096,1097 DD-Rev	ATAATCATCGTTGGATGTGACCTGATCATCGCTGGCTGCCATCTG	
AFF4-FL1103,1134 DD-Fw	CAGGTCACATCCAACGATGATTATGCCACCGAAATTTGGGACCAAG	
AFF4-FL1103,1134 DD-Rev	CTTGGTCCCAAATTTTCGGTGGCATAATCATCGTTGGATGTGACCTG	

Supplementary Table S2, Curve fitting data for AFF4-CHD RNA and DNA binding as determined by fluorescence anisotropy. Fluorescence anisotropy data on Figure 3 and 4 were fitted with the quadratic single site binding equation in GraphPad Prism, see Experimental Procedure.

Curve fitting data for Figure 3B, the binding of FBS-35mer G-quadruplex RNA to AFF4-CHD in buffer containing different KCl concentration.

KCl concentration	$K_{d, app}$ (μM)	R^2	B_{max}
50 mM	0.422 ± 0.016	0.9935	124.4 ± 1.15
100 mM	0.645 ± 0.036	0.9861	91.54 ± 1.39
200 mM	0.691 ± 0.081	0.9436	65.09 ± 2.11

Curve fitting data for Figure 3C, the binding of different nucleotides to AFF4-CHD in buffer containing 100 mM KCl.

Nucleotides	$K_{d, app}$ (μM)	R^2	B_{max}
FBS-35mer G-quadruplex RNA	0.270 ± 0.026	0.9784	90.88 ± 2.00
HIV1 LTR-III G-quadruplex DNA	1.647 ± 0.176	0.9792	51.05 ± 1.87
Unstructured TAR RNA	0.228 ± 0.044	0.9058	69.66 ± 3.05
Stem-loop-TAR RNA	0.251 ± 0.026	0.9724	67.85 ± 1.60

Curve fitting for Figure 4C, the binding of FBS-35mer G-quadruplex RNA to phosphorylated and Apo AFF4-CHD.

Phosphorylation state	$K_{d, app}$ (μM)	R^2	B_{max}
Phosphorylated AFF4-CHD (P-TEFb WT)	0.693 ± 0.068	0.9545	81.98 ± 2.24
Apo AFF4-CHD (P-TEFb N-mut)	0.275 ± 0.019	0.9751	74.90 ± 1.20

Legends to Supplementary Figures

Figure S1. Alignments of AFF proteins from human, mouse, chicken, zebra fish and drosophila. The sequence ID of the proteins in non-redundant protein sequences databank are as the following: *Homo sapiens* AFF4 (NP_055238.1), *Mus musculus* AFF4 (NP_291043.1), *Gallus gallus* AFF4 (XP_004945087.1), *Danio rerio* AFF4 (XP_005173957.1), *Homo sapiens* AFF1 (NP_001160165.1), *Mus musculus* AFF1 (NP_001074267.1), *Gallus gallus* AFF1 (XP_025006145.1), *Homo sapiens* AFF3 (NP_002276.2), *Mus musculus* AFF3 (NP_001277743.1), *Gallus gallus* AFF3 (XP_05133278.1), *Homo sapiens* AFF2 (NP_001162594.1), *Mus pahari* AFF2 (021043331.1), *Danio rerio* AFF2 (XP_002664429), *Gallus gallus* AFF2 (XP_015134139.2), *Danio rerio* AFF3 (XP_009302946), *Drosophila melanogaster* Lillipitian (NP_523464.1). The alignment was made using full-length AFF proteins and only the C-terminal sequences are shown in the figure. Stars indicate phosphorylation sites identified by mass spectrometry. Black stars mark the residues that are not highly conserved among AFF proteins. Brown stars indicate the conserved residues that are identified as phosphorylation sites. Residues on dimerization interface are marked with black and orange squares. Orange squares indicate the residues that are mutated in 5MA and 5MD constructs.

Figure S2. (A)-(C) Superposition of AFF4-CHD with top hits from DALI search. (A) Superposition of AFF4-CHD onto 5G05-O, the anaphase-promoting complex subunit 5 (APC5) (30). AFF4-CHD is colored in dark green and APC5 is colored in gray. AFF4-CHD superposes well with the last 5 helices in the C-terminal of APC5, with an RMSD of 2.7 over 452 atoms (30). (B) Superposition of AFF4-CHD onto 4JHR-B (32), the G-protein-signaling modulator 2. AFF4-CHD is colored in dark green and G-protein modulator 2 is colored in gray. AFF4-CHD superposes well with the 5 helices in the N-terminal of G-protein modulator 2, with an RMSD of 2.216 over 424 atoms. (C) Superposition of AFF4-CHD onto 6HEP-B (in gray), the 14-3-3 protein β , in the presence of CFTR R-domain peptide pS753-pS768 as the ligand (6HEP-E, in black) (33). (D-E) SDS-PAGE of the elution fractions of analytical gel filtration. (D) Shows the fractions of analytical gel filtrations of AFF4-CHD-WT, AFF4-CHD-5MA and AFF4-CHD-5MD, corresponding to the elution profile in Figure 2B. The same fractions that cover the peaks of all three runs are shown on the SDS-PAGE and stained with instant blue (Expendeon). (E) shows the fractions of Figure 2D. The same fractions from all four runs were loaded to SDS-PAGE and stained with instant blue (Expendeon).

Figure S3. (A)-(B) SDS-PAGE of the AFF4-CHD before and after phosphorylation by P-TEFb and P-TEFb N mutant. This figure shows the phosphorylation of the protein used in Figure 4C and 4D. (A) shows the protein involved in the phosphorylation of replication 1 (R1). (B) shows the protein involved in the phosphorylation of replication 2 and 3 on the same PAGE (R2 and R3). (C) Calibration of analytical gel filtration column Superdex 200 3.2/300 increase (GE Healthcare). The protein standards and their molecular weight are as indicated in the figure. The Log Mw (y) of each protein was plotted against the elution peak volume of each protein (x) in GraphPad Prism. Linear regression curve was fitted to the data points. The fitting equation and the R^2 of the fitting are indicated on the panel. The molecular weight of an unknown protein can be calculated using the given equation and the elution volume (x). (D)-(E) Static light scattering results of AFF4-CHD-WT, AFF4-CHD-5MD, MBP-AFF1-CHD and MBP-AFF1-CHD/AFF4-CHD-WT complex. In each panel, the elution profile of a SEC run (A280, curve in black) and the molecular weight measured from light scattering (Mw, curve in red) are plotted against elution volume. The values are indicated as left and right y-axis respectively. For panel G, to form the heterodimeric MBP-AFF1-CHD/AFF4-CHD-WT complex, MBP-AFF1-CHD and AFF4-CHD-WT were mixed in 1:3 molar ratio and incubated overnight before injecting to the column. The molecular weight of the heteromeric peak and the excess AFF4 are both analyzed and indicated on the figure.

Figure S1

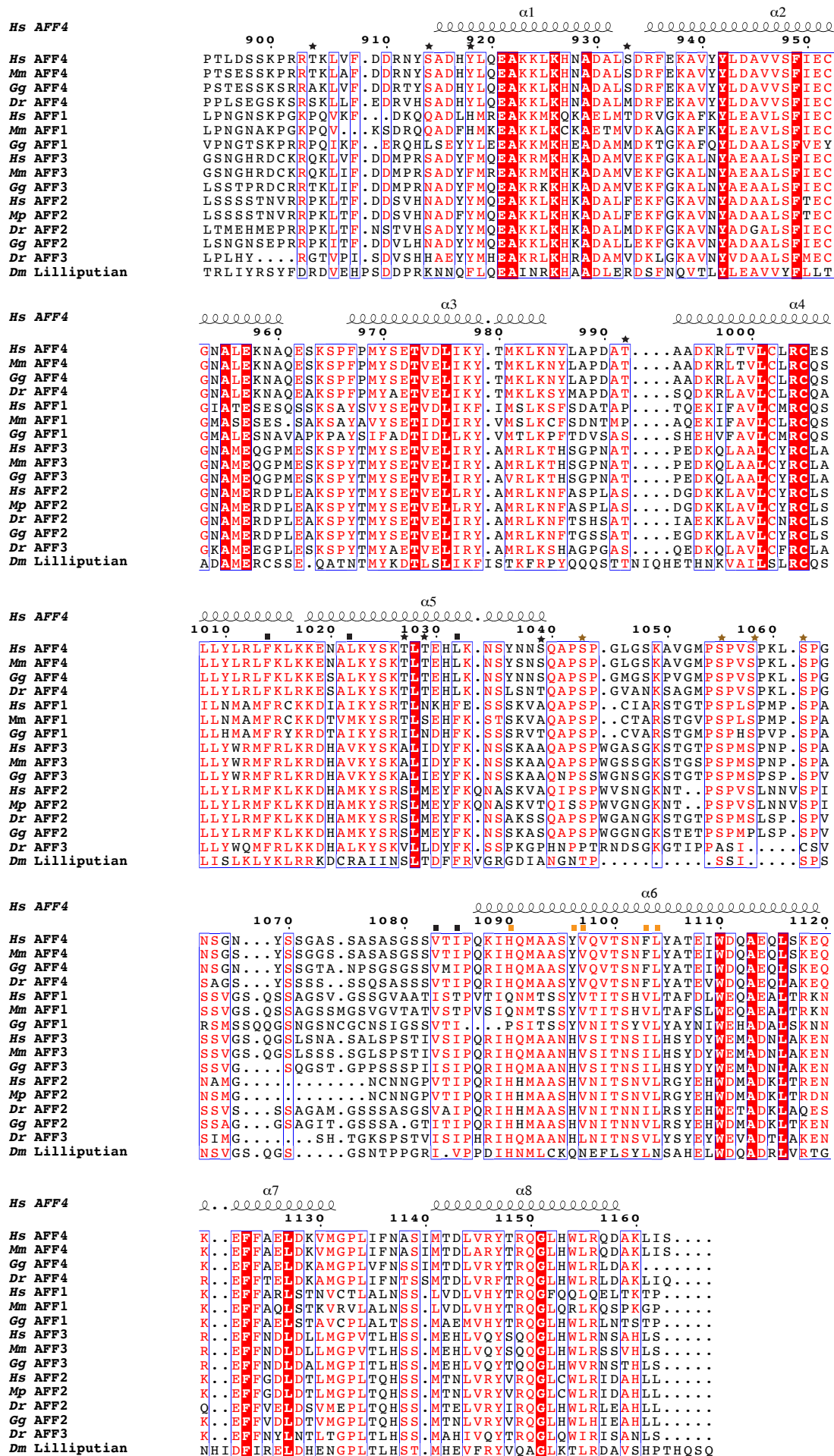


Figure S2

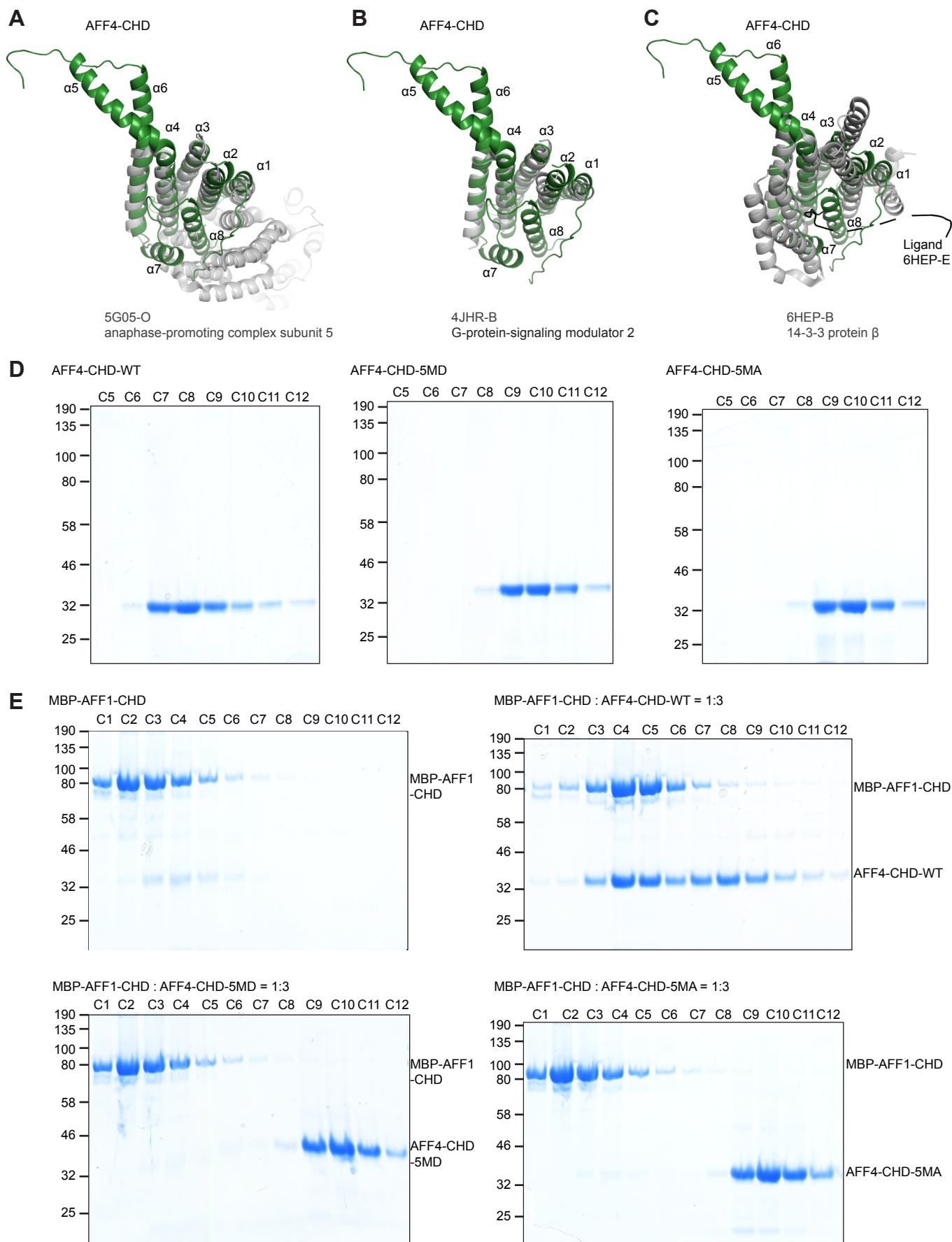
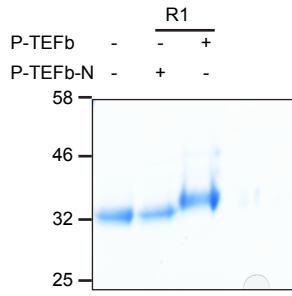
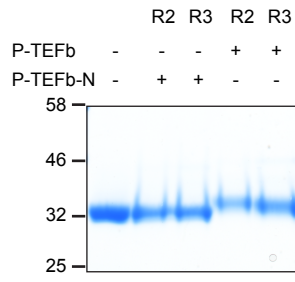


Figure S3

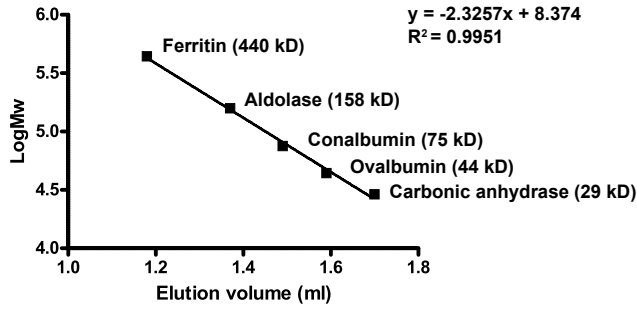
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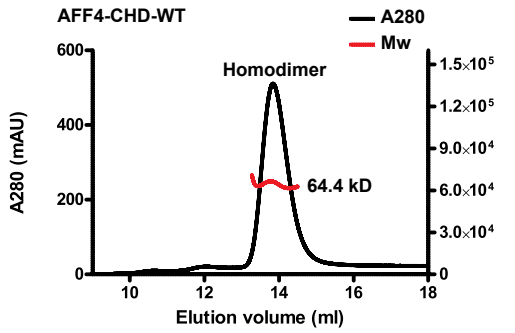
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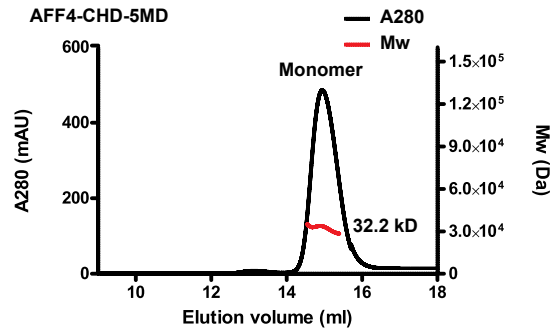
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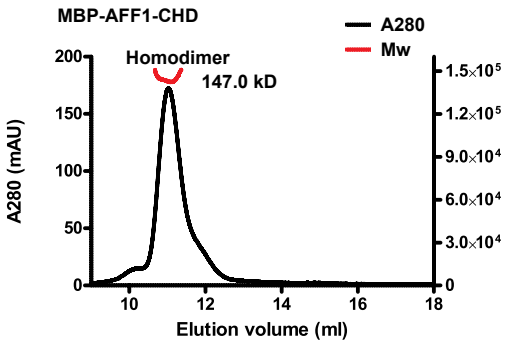
D



E



F



G

