New Phytologist Supporting Information

Abscisic acid is a substrate of the ABC transporter encoded by the durable wheat disease resistance gene *Lr34*

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Fig. S1 Lr34res induces a multiple stress response in rice. (a) Venn diagram showing number of genes of the 'Lr34res-responsive core gene set' that were found to be differentially expressed by biotic and/or abiotic stress in a rice meta-analysis (Shaik & Ramakrishna, 2014). (b) Semi quantitative RT-PCR of known stress-responsive genes. Most of these genes are not or only very weakly expressed in the sib line in a stress-free environment but they are strongly induced in the Lr34res-containing rice line 19. 19sib-1, 19sib-2 and 19sib-3 represent three independent biological replicates of 19sib whereas 19-1, 19-2 and 19-3 are three independent biological replicates of line 19. OsLEA3-1 = late embryogenesis abundant 3-1; OsMIOX = myo-inositol oxygenase; OsEFA27 = caleosin; sa/T = salt stress-induced; Os/CL = isocitrate lyase; OsMS; malate synthase; OsRSs1 = sucrose synthase-1 (reference gene) (Wang et al., 1992). (c) Quantitative RT-PCR expression analysis of the stress-responsive genes OsCHIT8, OsRCI2-5, OsASR1, OsBBTI4 and OsRNS4. RT-qPCRs were done on plants of the Lr34res-expressing rice line 19 and its sib 19sib. Relative fold changes (19sib was set to 1) and P-values (Student's t-test on log₁₀ transformed expression values) are indicated. Three biological replicates were used, mean ± SEM. Details on the function of the genes used in (b) and (c) are given in Table S3.



Fig. S2 Characterization of the *Lr34sus* **rice line 131**. (a) *Lr34* expression levels in coleoptiles of *Lr34res* lines 8, 11 and 19 compared to the *Lr34sus* line 131. RNA was extracted from at least 10 pooled coleoptiles. n = 3 technical replicates, mean ± SEM. A primer pair amplifying both *Lr34* alleles was used in the upper panel and a *Lr34res*-specific primer pair was used in the lower panel. (b) RT-qPCR for *Lr34* and the *Lr34res*-responsive genes *OsRCl2-5*, *OsRNS4* and *CHIT8* on four-week-old rice seedlings of lines 19, 131 and 131sib. n = 4 biological replicates, mean ± SEM, letters indicate lines with similar expression levels (P > 0.05; Tukey's honest significance test). These genes were chosen because they have a known function in stress tolerance (see Table S3) (c) Full-length cDNA amplification of the *Lr34sus* allele in line 131. The expected fragment size was 4,321bp. The integrity of the *Lr34sus* full-length cDNA was verified by sequencing. (d) Southern blot showing copy number in line 131. A probe for the selectable marker gene *HPT* was used (Risk *et al.*, 2013).



Fig. S3 LR34 changes ABA fluxes. (a) ³H-JA accumulation (disintegrations per minute, d.p.m.) in rice seedlings of lines 11 and 19 compared to respective sib lines. N = 3, n = 6-17, mean \pm SEM. (b) ³H-ABA accumulation in non-germinated caryopses of rice lines 11 and 19 compared to sib lines. N = 3, n = 3-17, mean \pm SEM. (c) Relative ³H-ABA accumulation in yeast strain YMM12 after addition of cold phytohormones or hormone analogs [3 μ M] salicylic acid (SA), metyl jasmonate (MJ), coronatine (Cor). The control (Con) without inhibitor was set to 100%, mean \pm SEM. * P < 0.05, ** P < 0.01. (D) Relative ³H-ABA accumulation in yeast strain YMM12 after addition of the indole alkaloid camalexin (CamX) and the diterpene sclareol (ScL) [3 μ M]. Both components have been reported to contribute to disease resistance in Arabidopsis. The control (Con) without competitor was set to 100%, mean \pm SEM. * P < 0.01.



Fig. S4 ABA concentrations in whole leaves of three-week-old plants of *Lr34res*containing lines 19 and 16 compared to sib lines. n = 3 biological replicates, mean ± SEM, n.s. = non-significant difference (Student's t-test, P < 0.05).



Fig. S5 Cluster analysis of the leaf sheath infection assay shown in Fig. 5c. Numbers at the edges indicate the Approximately Unbiased (AU) p-values.

Table S1. Primer sequences used in this study

semi RT-qPCR					
gene name	forward primer	reverse primer	fragment size [bp]	Tm [°C]	cycle number
OsLEA3-1	GACCTCCAGCACGTCGCAGG	GTGGCAGAGGTGTCCTTGTTG	327	60	35
OsMIOX	GCCATGACGACTACATGTACC	CGATGAGTGACATGTAGTAGG	249	60	35
OsEFA27	GGTCTAAGCTACCCAACTCTG	CACGTAGAGCAATATCCACTC	282	60	30
salT	GACGCTGGTGAAGATTGGTC	CCAGTTTAATCTCTGTAGAGGTG	206	65	40
OsICL	GTGTGGCAGTTCATCACGCTC	CATGAGCCCTTGAACTGCTC	266	62	35
OsMS	GACCGCACCGTCGAGATCAC	GTGATCGTGCCGGCAACCGC	173	65	40
Lr34_full_lenght cDNA	GAGTACGGCTAGGCAATAGC	GGCAAGTAGCTATATCTGTAAC	4,321 (sus) 4,318(res)	Touch down	* 35
RT-gPCR					
· · · ·				primer	
gene name	forward primer	reverse primer	fragment size [bp]	efficiency	reference
Lr34sus and Lr34res rice	GACAGCGCCAGAATGGTGC	GACATCAACCCTGTCAATTC	179	105%	this study
Lr34sus and Lr34res barley	ACTGGATTCGCTTGTATGGATA	ACTGGCAGAAGAACCTTGAAACA	80	101.70%	this study
HvGAPDH	CCGGGTTCCCACTGTGGAT	TGACTAGCAACTCGGTGCGG	451	102%	this study
OsASR1	GAGCATACGCCATGCACGAG	GATCTCCTCCTTCACCCTGTG	80	105.7%	this study
OsRCI2-5	TACGGCTTGGGTATTGAGTTCTGG	CCGACGCTGGCTCTGCTC	125	95.7%	Li et al. (2014)
OsRNS4	CTCATCCTCATGTGGCCTGG	GTTGCACCTGACAACAGCTG	147	95.1%	this study
RC24/CHIT8	GTGATGACAGGGCAGTGGAC	CTCTGGTTGTAGCAGTCCAAG	209	114%	this study
BBTI4	GTGACAGCATCGTGCAGCTG	GCGGCGCACTTGTTCACCTC	277	100.80%	this study
cloning of HA-Lr34					
	forward primer	reverse primer	fragment size [bp]	Tm [°C]	cycle number
Lr34_fragment1	CCAATATATCCTGTCAAACACTG	CAGCATAATCTGGAACATCGTATGGATACATCTCAAGATGGGTGAGTTAAATCG	2,815	55	35
Lr34_fragment2	ATGTATCCATACGATGTTCCAGATTATGCTGAGGGCCTCGCAAGAGAGAC	GGCATATGAACCCACACTCCTCAA	4,691	55	35
Lr34_fragment3	TTGAGGAGTGTGGGTTCATATGCC	TTGACTGCACGAATAACAATGGCTG	2,987	55	35
Lr34_fragment4	GCAGCCATTGTTATTCGTGCAGTC	TTGACTGCACGAATAACAATGGCTG	2,754	55	35
Lr34_fragment5	CACCATAGGCGCTTATAATGATCAGAC	ATGCAAGCTGATCCACTAG	3,314	55	35
yeast Leu casette	CTCCACGAAAATATCCGAACGCAGCAAGATTGGGTCCTTTTCATCACGTGC	TGCCCAGGCAAGACCGAGATGCACCGCGATGCGGCCGCCACCGCGGT	3,792	60	40

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Locus ID Gene name Gene Log₂ **Reported function** Reference FC¹⁾ description LOC Os05g46480 11.9 OsLEA3-1 late embryo-Improved drought (Xiao et al., genesis tolerance in 2007) transgenic rice abundant protein LOC Os06g36560 **OsMIOX** *myo*-inositol 3.4 Improved drought (Duan et al., oxygenase tolerance in 2012) transgenic rice LOC_Os03g17790 putative low 4.1 Improved drought (Li et al., OsRCI2-5 temperature and tolerance in 2014) salt responsive transgenic rice protein 3.6 LOC Os09g36680 OsRNS4 S-like Improved salinity (Zheng et ribonuclease al., 2014) tolerance in transgenic rice LOC Os04g43200 caleosin 8.1 Responsive to (Frandsen et OsEFA27 osmotic stress al., 1996) LOC_Os07g34520 OsICL isocitrate lyase 12.2 Increased expression (Maruyama after dehydration et al., 2014) LOC Os04g40990 6.5 Increased expression (Maruyama **OsMS** malate synthase after dehydration et al., 2014) 7.0 LOC Os01g24710 sa/T jacalin-like lectin Responsive to (Claes et al., domain drought stress 1990) containing protein LOC Os01g72900 ABA-, stress-2.5 Increased expression (Perez-Diaz OsASR1 and ripeningafter drought stress et al., 2014) induced Enhanced stripe rust LOC Os06g51060 RC24/CHIT8 4.6 chitinase (Huang et resistance in al., 2013) transgenic wheat 6.1 LOC Os01g03340 BBTI4 Bowman-Birk Conferred partial, (Pang et al., type bran trypsin broad-spectrum 2013) inhibitor resistance against bacterial blight in transgenic rice

Table S3. *Lr34res*-responsive core genes with a reported function in abiotic or biotic stress tolerance.

¹⁾ Fold change (FC) is represented as the average of the two independent transgenic lines 5 and 16 at seedling and adult plant stages.

Table S4. Comparison of the 'Lr34res-responsive core gene set' to a microarray study of seven-day-old rice seedlings incubated in solutions of 100 µM abscisic acid (ABA), 100 µM salicylic acid (SA), 100 µM jasmonic acid (JA), 50 µM benzyl aminopurine (BAP; cytokinin), 50 µM indole-3-acetic-acid (IAA; auxin) or 100 µM 1-aminocyclopropane-1carboxylic acid (ACC; ethylene derivative) (Garg et al., 2012). In total 4,171 hormoneresponsive genes were identified. 80 (79 up-regulated and 1 down-regulated) of the 159 Lr34res-responsive core genes were present in this list of 4,171 hormone-responsive genes. The numbers in the table list the number of genes (out of 80) that were responsive to the respective hormone. A relative quality score was assigned as following: If a gene of the 80 Lr34res-responsive core genes was differentially expressed by the respective hormone in the hormone study with the same direction (up- or down-regulated) it was given the score +1. If a gene of the 80 *Lr34res*-responsive core genes was not differentially expressed in the hormone study or differentially expressed in the opposite direction it was given the score -1. The relative quality score was calculated as the sum of the individual ratings. A score of 80 indicates perfect correlation between the two data sets whereas a score of -80 indicates no correlation.

up-regulated in <i>Lr34res</i> core gene set	79	down-regulated in <i>Lr34res</i> core gene set	1	relative quality score	exclusively regulated by respective hormone	also ABA regulated
up-regulated by ABA	68	down-regulated by ABA	0	56	35	-
up-regulated by SA	26	down-regulated by SA	0	-28	2	23
up-regulated by JA	19	down-regulated by JA	0	-42	1	14
up-regulated by BAP	8	down-regulated by BAP	0	-64	2	6
up-regulated by IAA	4	down-regulated by IAA	0	-72	0	3
up-regulated by ACC	3	down-regulated by ACC	0	-74	0	3

Table S5. Comparison of the '*Lr34res*-responsive core gene set' to an RNAseq analysis performed in Brachypodium seedlings incubated in 10 μ M abscisic acid (ABA), 100 μ M salicylic acid (SA), 30 μ M methyl jasmonate (MJ), 1 μ M cytokinin (CK) or 10 μ M indole-3-acetic acid (IAA; auxin) (Kakei *et al.*, 2015). DEG = differentially expressed genes. 'DEG with rice orthologs' lists the Brachypodium genes that had a clear ortholog in rice.

Hormome	DEG in Brachypodium	DEG with rice orthologs	Found in <i>Lr34res</i> core gene set	Expected*	p-value
ABA	445	409	25	3.35	<0.0001
MJ	383	352	5	2.89	0.21
IAA	125	118	0	1	0.32
SA	81	75	0	0.5	0.48
СК	23	22	0	0	-

* Number of genes expected to be found in the '*Lr34res*-responsive core gene set' if there were no enrichment for a particular hormone. The values were calculated based on the number of 'DEG with rice orthologs' and the total number of genes (19,399) identified in the hormone study (Kakei *et al.*, 2015). For example, 409 DEG for ABA represent 2.1% of the total genes. 2.1% of 159 *Lr34res*-responsive genes correspond to 3.35 genes that would be expected in the '*Lr34res*-responsive core gene set' by chance. The p-value was calculated based on the expected and observed genes using a Chi-square test.

Table S6. P-values of the generalized linear analysis based on the leaf sheath assay shown in Fig. 5c. P-values < 0.05 are marked in grey.

	p-value				
Comparison	Level 1	Level 2	Level 3	Level 4	
19sib x 27-3 - 19 x wt	0.0005	0.9860	0.0015	0.0290	
19 x 27-3 - 19 x wt	< 0.001	< 0.001	< 0.001	0.42957	
19sib x wt - 19 x wt	< 0.001	< 0.001	< 0.001	< 0.001	
19 x 27-3 - 19sib x 27-3	< 0.001	< 0.001	< 0.001	0.00294	
19sib x wt - 19sib x 27-3	< 0.001	< 0.001	< 0.001	0.23143	
19sib x wt - 19 x 27-3	< 0.001	0.6780	< 0.001	< 0.001	

Level-wise generalized linear model

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