Fig. S3

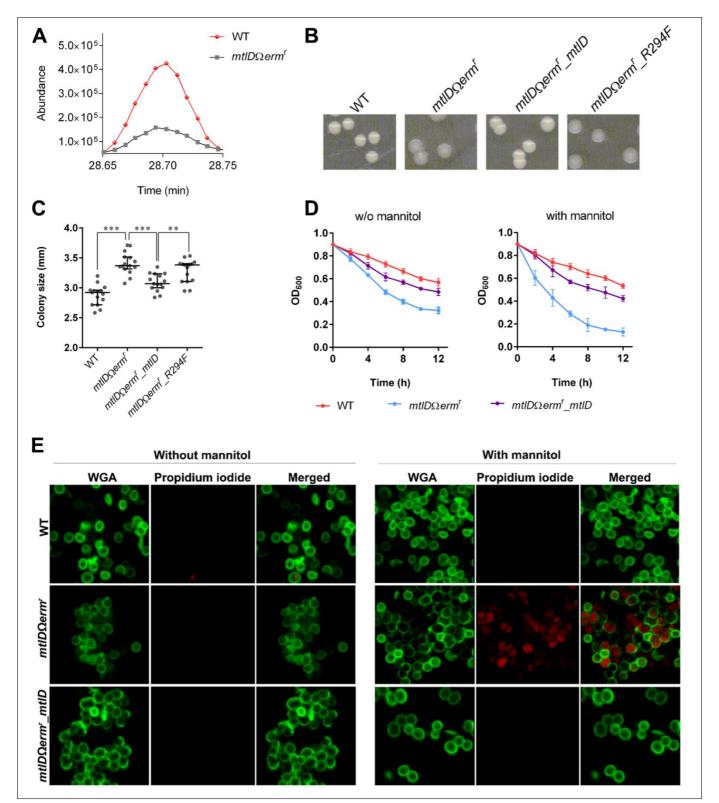


Fig. S3. Assessment of intracellular accumulations of mannitol in *S. aureus* strains and its impact on colony morphology of various *S. aureus* USA300 strains under control and stress conditions. (A) Gas chromatography-mass spectrometry (GC-MS) analysis of intracellular mannitol. Briefly, cytoplasmic contents were extracted from cell pellets of WT and *Sa*M1PDH knockout (*mtlD* Ω *erm*^r) *S. aureus* strains by flash-freezing cells in liquid nitrogen followed by lysostaphin treatment and sonication. After derivatization, samples were subjected to GC-MS analysis in which D-mannitol and adonitol were used as a reference and

an internal standard, respectively. After comparing spectra of chromatogram peaks with an electron impact mass spectrum library NIST08 (NIST, USA) followed by normalization, relative abundance of mannitol in each sample was determined by absorbance peak of mannitol in chromatography retention profiles. Data points (WT, diamond; knockout, square) represent absorbance of mannitol recorded at each chromatographic retention time. Solid lines (WT, red; knockout, gray) connect data points. Data are representative of at least three independent measurements. The SaM1PDH knockout ($mtlD\Omega erm^{r}$) was found to accumulate significantly higher amount of mannitol than that of WT S. aureus USA300. (B) Effect of mannitol on colony morphologies of S. aureus strains wherein colonies of WT, knockout (mtlDQerm^r), WT complemented $(mtlD\Omega erm^{r} mtlD)$, and R294F mutant complemented $(mtlD\Omega erm^{r} R294F)$ S. aureus strains on mannitolcontaining nutrient agar. Indicated *S. aureus* strains were plated onto BHI agar containing 27.5 mM mannitol. After incubation at 37 °C for 48 h, plates were photographed. Images are representative of at least two independent experiments for each strain. Colony sizes of WT, knockout, WT complemented, and R294F mutant complemented S. aureus strains on mannitol-containing nutrient agar. (C) From the photographs, 15– 25 colonies from each strain were randomly selected for diameter measurements. Horizontal lines represent means of colony sizes, whereas whiskers represent one standard deviation of the means. Statistical significance was calculated by Student's *t*-test (** p < 0.01, *** p < 0.001). (**D**) Optical density-based Triton X-100-induced autolysis assay depicting the effect of mannitol on S. aureus USA300 cell lysis. Briefly, after being cultured in either (left), BHI media or (right) BHI media containing 27.5 mM mannitol, bacterial cell densities were adjusted to an OD₆₀₀ of 0.9 in PBS containing 0.05% Triton X-100 (1, 2). The time-dependent OD_{600} was monitored at room temperature for 12 h. The reduction in OD_{600} of the *mtlDQerm^r* mutant (Clearance of cell turbidity) in Triton X-100-induced autolysis assay confirmed that the cell wall of the *mtlDQerm^r* mutant strain is much weaker than those of the WT and complemented strains. (E) Visualization of mannitol-induced cell damage of *mtlDQerm^r* knockout S. *aureus* USA300 strain using cell wall-specific wheat germ agglutinin (WTA) fluorescent dye. The indicated bacteria were spotted on BHI agar containing without or with 27.5 mM mannitol and incubated at 37 °C for 48 h. The S. aureus cells were harvested and subsequently washed with PBS for 3 times. Bacteria were then resuspended in PBS containing Alexa flour 488 conjugated WGA and incubated for 5 min at room temperature to allow staining of cell-wall of all S. aureus cells (3). The free WGA fluorescent dye was removed by 3 times PBS washing. The WGA stained, and washed S. aureus cells were resuspended in PBS containing propidium iodide (PI) to visualize membranedamaged S. aureus by assessing the entry and staining with PI. These results confirmed that $mt D\Omega erm^{r}$ knockout strain lacking SaM1PDH activity were cytolyzed under mannitol treated condition.

References

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