

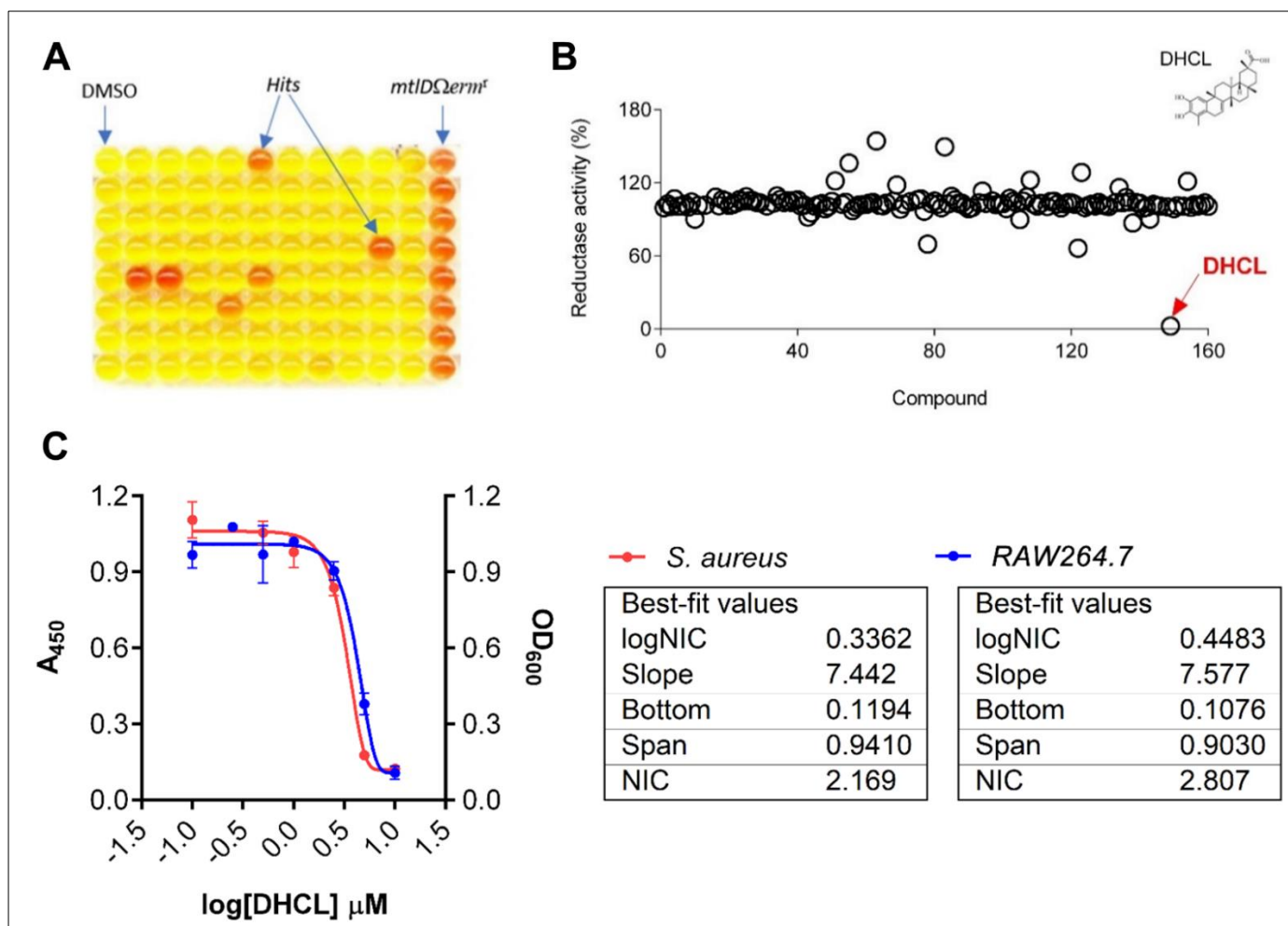
Fig. S5

Fig. S5. Functional assays for screening and identifying dihydrocelastrol (DHCL) as a potent inhibitor of SaM1PDH; and its effects on proliferation of *S. aureus* USA300 and RAW264.7 cells. (A) Primary screening assay to identify hits. Inhibiting mannitol-metabolizing capacity of WT *S. aureus* USA300 strain was used to screen for SaM1PDH inhibitors. In a mannitol salt broth supplemented with phenol red as a pH indicator, mannitol fermentation of WT *S. aureus* cells generates acidic byproducts that turn broth from red to yellow. Chemicals inhibiting that color change were recognized as hits. WT *S. aureus* cells grown in broth treated with DMSO and SaM1PDH knockout (*mtID Δ erm^F*) *S. aureus* cells grown in broth alone were included in each screening plate as a negative and positive control, respectively. Image is representative of screening plates photographed after 6 h incubation at 37 °C with arrows indicating negative controls, positive controls, and hits. (B) Secondary screening assay to validate primary hits. Inhibiting SaM1PDH F6P reductase activity *in vitro* was used to validate hits found in (A) and identified DHCL as a potent inhibitor of SaM1PDH. Briefly, purified SaM1PDH was incubated with cofactor (NADH), substrate (F6P), and hits found in (A) for 5 min at room temperature. Relative reductase activity of SaM1PDH was determined by measuring the absorbance of NADH at 340 nm and calculated as percentages of SaM1PDH activity in treated reaction-mix with hits compared to that in control reaction-mix treated with DMSO. Graph was constructed by plotting reductase activities of SaM1PDH (open circles) against examined hits. Arrow indicates F6P reduction activity of SaM1PDH treated with DHCL, whose chemical structure is shown above graph. (C) Effect of DHCL on proliferation of *S. aureus* USA300 and RAW264.7 cells. WT *S. aureus* USA300 and RAW264.7 cells were cultured in appropriate media supplemented with various concentrations of DHCL (0–10 μM). For *S. aureus* cells, DHCL susceptibility was assessed by measuring optical density of bacterial culture at 600 nm (OD_{600}).

For RAW264.7 cells, after addition of a water-soluble tetrazolium salts solution (EZ-Cytox Cell Viability Assay kit, DoGen, Korea) into cultured cells, cell viability was assessed by measuring absorbance at 450 nm (A_{450}). Graph was constructed by plotting either OD_{600} (*S. aureus*) or A_{450} (RAW264.7 cells) values against logarithms of examined DHCL concentrations. Circles (*S. aureus*, red; RAW264.7, blue) represent mean values obtained from at least three independent measurements, whereas whiskers represent one standard deviations of the means. Solid lines (*S. aureus*, red; RAW264.7, blue) represent the best-fit curves determined by nonlinear regression analyses using GraphPad Prism 7 (<http://www.graphpad.com>) to determine non-inhibitory concentrations (NICs) (1) of DHCL. Nonlinear regression analysis results are summarized in tables.

Reference

1. Lambert RJ, Pearson J. 2000. Susceptibility testing: accurate and reproducible minimum inhibitory concentration (MIC) and non-inhibitory concentration (NIC) values. *J Appl Microbiol* 88:784-90.