

Table S2. Primers, plasmids and bacterial strains used in this study.**A. Primers used in this study**

Primer name	Sequence (5'→3') ^a	Purpose
<i>mtlD</i> _fwd	TAGGATCCATGAAAGCAGTTCACCTTTGGTG (<i>Bam</i> HI)	cloning of <i>S. aureus mtlD</i> gene into plasmid pVFT1S
<i>mtlD</i> _rev	GTCTCGAGTTATGAAAGTGAATTGTACGCTTCA (<i>Xho</i> I)	
<i>mtlD</i> _comp._fwd	ATTATAGGTACCAGAAA GGAGGT AAACATATGAAAGCAGTTC ACTTTG (<i>Kpn</i> I)	cloning of <i>mtlD</i> wild type and mutants into plasmid pRMC2
<i>mtlD</i> _comp._rev	AATAATGAATTCTTATGAAAGTGAATTGTAC (<i>Eco</i> RI)	
<i>R283S</i> _fwd	CACGTGTCGGAAGCGGTACATTACGTAAAATTGGCCC	introducing <i>R283S</i> mutation into <i>mtlD</i>
<i>R283S</i> _rev	GGGCAATTTTACGTAATGTACCGCTTCCGACACGTG	
<i>R287S</i> _fwd	GTCGGACGAGGTACATTAAGTAAAATTGGCCCTAAAG	introducing <i>R287S</i> mutation into <i>mtlD</i>
<i>R287S</i> _rev	CTTTAGGGCCAATTTTACTTAATGTACCTCGTCCGAC	
<i>R294F</i> _fwd	GCCCTAAAGATTTTCATTATAAAAAC	introducing <i>R294F</i> mutation into <i>mtlD</i>
<i>R294F</i> _rev	GGTTTTATAATGAAATCTTTAGGGC	
<i>mtlA</i> _fwd	AGCAATTCCTCATGGCACAG	qRT-PCR for <i>mtlA</i> gene
<i>mtlA</i> _rev	TTACCAGCAATTCCCACGAC	
<i>mtlR</i> _fwd	TCGTTTGCAGCAGTGGTATG	qRT-PCR for <i>mtlR</i> gene
<i>mtlR</i> _rev	ATCGCTGACTGATGCTTGTG	
<i>mtlF</i> _fwd	AAAGGCACTGCAAAGCAAC	qRT-PCR for <i>mtlF</i> gene
<i>mtlF</i> _rev	TGCAACACCAGTCATTCCAC	
<i>mtlD</i> _fwd	TGGCTTACGCCGTGTTTTAG	qRT-PCR for <i>mtlD</i> gene
<i>mtlD</i> _rev	AATGTACCTCGTCCGACACG	
<i>rho</i> _fwd	AACGTTTGACGAACCACCAG	qRT-PCR for <i>rho</i> gene
<i>rho</i> _rev	ATAAGCGCGTGCTAATCTCG	
<i>rpoB</i> _fwd	TGAAGAGAACGGCGTTGAG	qRT-PCR for <i>rpoB</i> gene
<i>rpoB</i> _rev	CCAACCTGCAACGATTGGAC	

^a The restriction sites are underlined, and the corresponding restriction endonucleases are shown in parentheses. The Shine-Dalgarno sequence of *gapA* promoter is in bold.

B. Plasmids used in this study.

Plasmids	Descriptions	References
pVFT1S	<i>km^r</i> ; expression vector tagging His ₆ to the N-terminus of proteins	Korean Patent 1020050051893
pVFT1S_ <i>mtlD</i>	<i>km^r</i> ; pVFT1S expressing <i>SaM1PDH</i> wild type	This study
pVFT1S_ <i>R283S</i>	<i>km^r</i> ; pVFT1S expressing <i>SaM1PDH R283S</i> mutant	This study
pVFT1S_ <i>R287S</i>	<i>km^r</i> ; pVFT1S expressing <i>SaM1PDH R287S</i> mutant	This study
pVFT1S_ <i>R294F</i>	<i>km^r</i> ; pVFT1S expressing <i>SaM1PDH R294</i> mutant	This study
pRMC2	<i>amp^r</i> , <i>cm^r</i> ; expression vector under control of tetracycline inducible P _{<i>xyll/tetO</i>}	Reference (1)
pRMC2_ <i>mtlD</i>	<i>amp^r</i> , <i>cm^r</i> ; pRMC2 expressing <i>SaM1PDH</i> wild type	This study
pRMC2_ <i>R283S</i>	<i>amp^r</i> , <i>cm^r</i> ; pRMC2 expressing <i>SaM1PDH R283S</i> mutant	This study
pRMC2_ <i>R287S</i>	<i>amp^r</i> , <i>cm^r</i> ; pRMC2 expressing <i>SaM1PDH R287S</i> mutant	This study
pRMC2_ <i>R294F</i>	<i>amp^r</i> , <i>cm^r</i> ; pRMC2 expressing <i>SaM1PDH R294F</i> mutant	This study

C. Bacterial strains used in this study.

Strains	Descriptions	Sources or References
<i>Escherichia coli</i>		
DH5 α	Strain for cloning and plasmid amplification	Invitrogen, USA
BL21(DE3)	Strain for protein overexpression	Novagen, USA
B834(DE3)	Strain for selenomethionine-substituted protein overexpression	Novagen, USA
<i>Staphylococcus aureus</i>		
RN4220	Restriction-deficient strain of NCTC8325	Reference (2)
RN_RMC2	<i>cm^r</i> , <i>amp^r</i> ; RN4220 bearing plasmid pRMC2	This study
RN_ <i>mtlD</i>	<i>amp^r</i> , <i>cm^r</i> ; RN4220 bearing plasmid pRMC2_ <i>mtlD</i>	This study
RNR283S	<i>amp^r</i> , <i>cm^r</i> ; RN4220 bearing plasmid pRMC2_ <i>R283S</i>	This study
RNR287S	<i>amp^r</i> , <i>cm^r</i> ; RN4220 bearing plasmid pRMC2_ <i>R287S</i>	This study
RNR294F	<i>amp^r</i> , <i>cm^r</i> ; RN4220 bearing plasmid pRMC2_ <i>R294F</i>	This study
WT SAUSA300	JE2, wild-type epidemic community-associated methicillin-resistant <i>S. aureus</i> isolate USA300 LAC	Nebraska library (3)
<i>mtlA</i> Ω <i>erm^r</i>	NE929, <i>erm^r</i> ; JE2 bearing <i>mtlA::Tn917</i>	Nebraska library (3)
<i>mtlR</i> Ω <i>erm^r</i>	NE837, <i>erm^r</i> ; JE2 bearing <i>mtlR::Tn917</i>	Nebraska library (3)
<i>mtlF</i> Ω <i>erm^r</i>	NE1737, <i>erm^r</i> ; JE2 bearing <i>mtlF::Tn917</i>	Nebraska library (3)
<i>mtlD</i> Ω <i>erm^r</i>	NE1263, <i>erm^r</i> ; JE2 bearing <i>mtlD::Tn917</i>	Nebraska library (3)
WT SAUSA300_pRMC2	<i>cm^r</i> , <i>amp^r</i> ; WT SAUSA300 bearing plasmid pRMC2	This study
<i>mtlD</i> Ω <i>erm^r</i> _pRMC2	<i>erm^r</i> , <i>amp^r</i> , <i>cm^r</i> ; NE1263 bearing plasmid pRMC2	This study
<i>mtlD</i> Ω <i>erm^r</i> _mtlD	<i>erm^r</i> , <i>amp^r</i> , <i>cm^r</i> ; NE1263 bearing plasmid pRMC2_ <i>mtlD</i>	This study

<i>mtlD</i> Ω <i>erm</i> ^r _{R283S}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE1263 bearing plasmid pRMC2_R283S	This study
<i>mtlD</i> Ω <i>erm</i> ^r _{R287S}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE1263 bearing plasmid pRMC2_R287S	This study
<i>mtlD</i> Ω <i>erm</i> ^r _{R294F}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE1263 bearing plasmid pRMC2_R294F	This study
<i>mtlA</i> Ω <i>erm</i> ^r _{<i>mtlD</i>}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE929 bearing plasmid pRMC2_ <i>mtlD</i>	This study
<i>mtlR</i> Ω <i>erm</i> ^r _{<i>mtlD</i>}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE837 bearing plasmid pRMC2_ <i>mtlD</i>	This study
<i>mtlF</i> Ω <i>erm</i> ^r _{<i>mtlD</i>}	<i>erm</i> ^r , <i>amp</i> ^r , <i>cm</i> ^r ; NE1737 bearing plasmid pRMC2_ <i>mtlD</i>	This study
WT_pRMC2	<i>cm</i> ^r ; WT SAUSA300 bearing plasmid pRMC2	This study

References

1. Corrigan RM, Foster TJ. 2009. An improved tetracycline-inducible expression vector for *Staphylococcus aureus*. *Plasmid* 61:126-9.
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3. Fey PD, Endres JL, Yajjala VK, Widhelm TJ, Boissy RJ, Bose JL, Bayles KW. 2013. A genetic resource for rapid and comprehensive phenotype screening of nonessential *Staphylococcus aureus* genes. *MBio* 4:e00537-12.