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Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their parents

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Manuscripts

1 **Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their**
2 **parents**

3
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24
25 **Keywords:** Metabolomics; lipids; inflammation; reference values; parents; children;
26 inheritance patterns; correlation studies; epidemiologic studies; cross-sectional studies.

27
28 **Word count: 4181**

29
30 **Abbreviations:** ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; BCAA: Branched
31 chain amino acid; CDC: Centers for Disease Prevention and Control; CVD: Cardiovascular
32 disease; CPS1: Carbamoyl-phosphate synthase 1; DHA: Docosahexaenoic acid; DOB: Date
33 of birth; EDTA: Ethylenediaminetetraacetic acid; GlycA: Glycoprotein acetyls; HbA1c:
34 Haemoglobin A1c; HDL: High-density lipoprotein; HOMA: Homeostatic model assessment;
35 IDL: Intermediate density lipoprotein; LA: Linoleic acid; LDL: Low-density lipoprotein;
36 LiH: Lithium Heparin; LSAC: Longitudinal Study of Australian Children; MUFA:

1 Monounsaturated fatty acid; NMR: Nuclear magnetic resonance; PCOS: Polycystic Ovary
2 Syndrome; PUFA: Polyunsaturated fatty acid; SFA: Saturated fatty acids; SST: serum
3 separating tubes; T2D: Type 2 diabetes; T2DM: Type 2 diabetes mellitus; VLDL: Very low
4 density lipoprotein; XL: Very large; XXL: Chylomicrons and extremely large; XS: Very
5 small.

6 7 **ABSTRACT**

8
9 **Objectives:** Nuclear Magnetic Resonance (NMR) metabolomics is high throughput and cost
10 effective, with the potential to improving the understanding of disease and risk. We examine
11 the circulating metabolic profile by quantitative NMR metabolomics of a sample of
12 Australian 11-12 year old children and their parents, describe differences by age and sex, and
13 explore correlation of metabolites in parent-child dyads.

14 **Design:** The population-based cross-sectional Child Health CheckPoint study nested within
15 the Longitudinal Study of Australian Children.

16 **Setting:** Blood samples collected from CheckPoint participants at assessment centres in six
17 Australian capital cities and eight selected regional centres between February 2015-March
18 2016.

19 **Participants:** 1180 children and 1325 parents provided a blood sample and had
20 metabolomics data available. This included 1133 parent-child dyads (518 mother-daughter,
21 469 mother-son, 68 father-daughter, and 78 father-son).

22 **Outcome measures:** 228 metabolic measures were obtained for each participant. We
23 focused on 70 biomarkers that captured variation in amino acid species, lipoprotein subclass
24 measures, lipid measures, fatty acids, measures related to fatty acid saturation, and composite
25 markers of inflammation and energy homeostasis.

26 **Results:** We identified sex-specific metabolic profiles in children and adults and differences
27 in the level of specific metabolites between childhood and adulthood. In general, metabolite
28 concentrations were higher in adults than children and sex differences were larger in adults
29 than in children. Positive correlations within parent-child dyads were observed for the
30 majority of metabolites. Correlations ranged from 0.03 (95% CI -0.05 to 0.12) for
31 acetoacetate in mother-daughter dyads, to 0.39 (95% CI 0.31 to 0.47) for isoleucine in
32 mother-son comparisons.

33 **Conclusions:** We report the serum metabolite profiles from mid-childhood and adulthood in
34 a population-based sample, together with parent-child concordance. Distinct age- and sex-
35 specific profiles were observed. These data will be informative for investigation of the

1 childhood origins of adult non-communicable diseases and for comparative studies in other
2 populations.

3 **Strengths and limitations of this study:**

- 4 • In a large population-based cohort, venous blood was collected for children and their
5 attending parent on the same day using the same methods
- 6 • Rapidly processed, high quality serum samples with standardised metabolomic data
7 generated as a single batch
- 8 • Cross-sectional design does not enable longitudinal analysis of specific metabolite
9 species over short term or longer periods of time
- 10 • Assessment of paternal influences on offspring metabolite measures is limited by a
11 relatively small sample size compared to mother-child pairs, reducing the precision of
12 estimates
- 13 • Factors known to influence metabolomic profile (such as body mass index) were not
14 considered

1 INTRODUCTION

2
3 Metabolomics involves the quantitative analysis of a large number of metabolites and lipids
4 involved in a diverse range of biochemical pathways.¹ Genetic/gene expression and
5 environmental exposures are associated with specific metabolic changes across many tissues
6 and body fluids.^{2 3} As such, metabolomics is recognised as a powerful top-down approach to
7 understand genetic and environmental influences on health and disease. Metabolomic
8 profiling also has considerable potential to identify clinically relevant biomarkers for risk
9 stratification and disease monitoring.

10 Recent advances in nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry
11 have enabled the simultaneous quantitative measurement of hundreds of metabolites. These
12 approaches are sufficiently cost effective and high throughput to be applicable to large cohort
13 studies. For example, NMR metabolomics of serum from the Cardiovascular Risk in Young
14 Finns Study identified many biomarkers from multiple metabolic pathways reflective of fatty
15 liver disease.⁴ These were also predictive of risk 10 years prior to diagnosis, indicating that
16 metabolic disruptions precede overt phenotype. Similar population and disease-specific
17 studies have identified metabolomic profiles associated with a range of exposures and health
18 outcomes with potential to reveal clinically important biomarkers and information on disease
19 mechanisms.⁵ In addition, specific serum metabolites can also be considered ‘intermediate
20 phenotypes’ linking genetic risk with disease outcomes.^{6 7}

21 Previous research indicates that some blood metabolites change with age, particularly from
22 mid to late adulthood.^{8 9} However, in adults sex appears to be a major driver of variation in
23 metabolite profile, potentially interacting with age. For example, the effects of sex appeared
24 to be greater in younger (age 25-35) than older Japanese adults.¹⁰ A study of 26,000 Northern
25 European adults identified many sex-specific metabolic species at the population level.⁹ In
26 men, several lipid measures begin to rise at early middle age whereas a similar increase is
27 only observed in females post menopause. This pattern is consistent for all cholesterol
28 measures – very low density lipoprotein (VLDL), intermediate density lipoprotein (IDL) and
29 low density lipoprotein (LDL) subclass particle concentrations - as well as for triglycerides.⁹
30 Physiological states such as pregnancy also have consistent and measurable influence on
31 serum metabolome.¹¹ However, it remains unclear how the serum metabolome responds to
32 age, sex and hormonal-specific factors in childhood.

33 Moreover, factors regulating the metabolic trajectory from early life to adulthood, the role of
34 metabolomic profile in health at the population level and the extent to which blood
35 metabolomic profiles are concordant for parents and children have not been fully explored.
36 One small study has reported correlations between parents (n=179) and their offspring

(n=255) for a range of cardiometabolic risk factors including standard lipid profile measured using conventional methods; this proved stronger for total cholesterol and LDL cholesterol than for high density lipoprotein (HDL) cholesterol or triglycerides.¹² Parent-child correlations of NMR metabolites have not been reported previously.

Here, we present (1) NMR-based metabolomics analysis of a population-based cohort of 11-12 year old children and their parents, (2) identify age and sex-specific metabolomic profiles and (3) report sex-specific parent-child concordance.

METHODS

Study Design: The Child Health CheckPoint comprised a detailed cross-sectional assessment of physical health and biomarkers in a population-based national sample of children (age 11-12 years) and their parents between February 2015 to March 2016. The CheckPoint was nested between waves 6 (2014) and 7 (2016) of the Longitudinal Study of Australian Children.¹³ The Longitudinal Study of Australian Children commenced in 2004, when two cohorts (the 'B' and 'K' cohorts, of which the B cohort only was included in the present study) were recruited who have since been followed biennially. Further details regarding the CheckPoint study design and methods are available elsewhere.¹⁴

Participants: Of the 8,921 families contacted to be part of the LSAC B cohort 5,107 families (57%) agreed to take part in the first wave of data collection in 2004; 4,484 families were retained for Wave 6 in 2014. During the Wave 6 LSAC home visit, B cohort families were introduced to the upcoming Child Health CheckPoint and asked to consent to their contact details being shared with the CheckPoint team. A total of 3,513 families provided permission to receive an information pack by mail and an information and recruitment phone call regarding the CheckPoint study (78% of Wave 6 cohort, 69% of original cohort). Of the families agreeing to receive information about the CheckPoint study, 1,874 families took part (53% of eligible participants, 42% of Wave 6 cohort and 37% of original cohort).

Ethics and consent: The CheckPoint data collection protocol was approved by The Royal Children's Hospital (Melbourne, Australia) Human Research Ethics Committee (33225D) and the Australian Institute of Family Studies Ethics Committee (14-26). The attending parent/caregiver provided written informed consent for themselves and their child to participate in the study, and asked to provide optional consent for the collection and use of biological samples.

Procedure: The specialised CheckPoint assessment centre sequentially visited six Australian capital cities and nine smaller regional centres between February 2015 and March 2016.¹⁴ Each participating child attended the centre with one parent or caregiver (usually the

1 biological mother) at which both participated in a wide range of measures relevant to non-
2 communicable disease. Those families who could not attend a centre were offered a home
3 visit. Participants were included in the current analyses if metabolomic data from CheckPoint
4 were available (figure 1). Venous blood was not available for home-visit participants, but was
5 collected at all capital city and most regional assessment centres. Participant pairs were
6 excluded from the concordance analyses in this study if the attending parent was not the
7 biological parent.

8 An experienced phlebotomist collected approximately 28mL of blood from the brachial vein
9 of the non-dominant arm of semi-reclining, semi-fasted participants (at the time of collection,
10 participants reported when they last ate or drank). Blood was collected sequentially into four
11 vacutainer tubes using a butterfly needle so only a single venepuncture was required. Order
12 of collection was (i) 2.7mL EDTA, (ii) 9mL EDTA, (iii) 9mL serum, (iv) 7.5mL Lithium
13 Heparin. The latter two tubes were immediately inverted 6 times to ensure mixing with
14 anticoagulant, and all tubes were transferred to the on-site laboratory. Time of collection was
15 scheduled earlier in the visit for parents than for children.

16 Collection tube barcodes were linked to the participant and samples were immediately
17 transported to an on-site laboratory where they were processed within two hours into 0.5mL
18 aliquots of plasma, serum, buffy coat (lymphocytes), whole blood and/or an aliquot tube
19 containing a blood clot (1.0mL FluidX screwcap tubes, Cheshire, UK) and stored
20 immediately at -809°C (Thermo Fisher Scientific, Waltham, USA). Each FluidX tube
21 contained a unique 2D barcode linked to the original collection tube and participant. As each
22 assessment centre closed, samples were shipped on dry ice to the Melbourne Children's
23 Bioresource Centre for long term storage at -80°C (serum, whole blood, plasma, blood clot)
24 or vapour phase liquid nitrogen (lymphocytes). At a later date, single 0.5ml serum aliquot
25 was removed for every CheckPoint participant and the combined aliquots were shipped in a
26 single batch to Nightingale Health (Helsinki, Finland) on dry ice for NMR metabolomics.

27 **Measures**

28 29 Metabolomic profiling

30 The Nightingale[®] NMR metabolomics platform (Helsinki, Finland) was used to obtain
31 metabolomics for children and parents using the 2016-version quantification algorithm.
32 Details of this platform and methodology have been extensively described elsewhere,^{6 15} and
33 epidemiological applications were recently reviewed.¹⁶ Briefly, metabolites were measured
34 from 0.35mL of serum using a single high-throughput experimental setup for the
35 simultaneous quantification of routine lipids, lipoprotein subclass distributions, particle size
36 and composition, fatty acids, and other low-molecular weight metabolites such as amino

1 acids and glycolysis-related metabolites. This generated data on 228 serum metabolite
 2 measures in absolute concentration units (eg millimoles per liter) and ratios (summarised in
 3 Table 1). Whilst widely used for epidemiological research, the NMR-based quantification has
 4 not been certified for clinical diagnostics. Further analytical validation of the quantification
 5 protocols for the biomarker subset routinely used in clinical settings (eg established
 6 cholesterol measures and creatinine) is expected to lead to recalibration of certain metabolite
 7 concentrations to better match clinical gold standards.¹⁶

8 **Table 1. Summary of biomarkers and derived variables obtained via high-throughput NMR**

Metabolic group	Species and derived measures
Amino acids	Alanine, Glutamine, Glycine, Histidine
	Branched chain: Isoleucine, Leucine, Valine
	Aromatic: Phenylalanine, Tyrosine
Cholesterol	VLDL, LDL, HDL, HDL2, HDL3, Total, Free, Esterified, Remnant
Triglycerides and phospholipids	Triglycerides (VLDL, LDL, HDL, total)
	Phosphoglycerides
	Ratio of triglycerides to phosphoglycerides*
	Phosphatidylcholine
	Sphingomyelins
	Total cholines
Apolipoproteins	Apolipoprotein A-1 (ApoA-1)
	Apolipoprotein B (Apo B)
	Ratio of Apolipoprotein B to Apolipoprotein A-1 (ApoB/Apo A-1)*
Fatty acids (FA)	Total, Omega-3, Omega-6, Polyunsaturated (PUFA), Saturated (SFA)
	Monounsaturated (MUFA), Docosahexaenoic acid (DHA), Linoleic (LA)
	Estimated degree of unsaturation
Fatty acid ratios	Omega-3/total FA*, Omega-6/total FA*, PUFA/total FA* (all %)
	SFA/total FA*, MUFA/total FA*, DHA/total FA*, LA/total FA* (all %)
Lipoprotein subclasses*	12 lipids in each of 14 subclasses: VLDL (XXL, XL, L, M, S, XS), IDL, LDL (L,M,S), HDL (XL, L, M,S)
Lipoprotein size*	Mean diameter of VLDL, LDL and HDL particles
Ketone bodies	Acetate, Acetoacetate, 3-hydroxybutyrate
Glycolysis related	Glucose, Lactate, Pyruvate, Citrate, Glycerol
Fluid balance	Creatinine, Albumin
Inflammation	Glycoprotein acetyls (GlycA)

9 Information obtained from <https://nightingalehealth.com/science/biomarkers>

10 * ratio; ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; DHA: Docosahexaenoic acid; GlycA: Glycoprotein acetyls;
 11 HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; L: Large; LDL: Low-density lipoprotein; LA:
 12 Linoleic acid; M: Medium; MUFA: Monounsaturated fatty acid; PUFA: Polyunsaturated fatty acid; S: Small; SFA:
 13 Saturated fatty acids; VLDL: Very low density lipoprotein; XL: Very large; XXL: Chylomicrons and extremely large; XS:
 14 Very small.

15
 16 Many of the 228 metabolomics measures correlate substantially both in adults
 17 (supplementary figure 1) and children (supplementary figure 2). For clarity, we therefore
 18 focussed on an informative subset of 70 lipid and metabolites in analyses that capture the
 19 majority of variation within the dataset. Most metabolites were reported as absolute

1 concentrations (ie not including ratios). We excluded glucose and lactate given the known
2 sensitivity of these measures to variable time of fasting and specific processing variables.
3 Some derived ratios, such as fatty acids and apolipoprotein B to apolipoprotein A-1, were
4 included because of previous data indicating clinical utility.¹⁷

5 Other measures and sample characteristics

6 *Age and sex:* Child date of birth and sex were obtained as detailed elsewhere.¹⁴ The attending
7 parent reported their own date of birth and sex by questionnaire. Age at assessment was
8 calculated to nearest week.

9 *Body mass index:* Using a portable stadiometer (Invicta IP0955, Leicester, UK), participant
10 height was measured without shoes, in light clothing, and in duplicate, to the nearest 0.1 cm.
11 A third measurement was taken if the difference of the first two height measurements was
12 greater than 0.5 cm; final height was the mean of all measurements made. Weight, to the
13 nearest 0.1 kg, was measured with an InBody230 bio-electrical impedance analysis scale
14 (Biospace Co. Ltd. Seoul, South Korea). Body mass index (BMI; kg/m²) was calculated, and
15 for children was converted to age- and sex-adjusted z-score using the 1970 US Centers for
16 Disease Control (CDC) growth reference charts.¹⁸

17 *Socioeconomic Disadvantage:* Neighbourhood socioeconomic position data were obtained
18 from the 2011 Australian Census Socio-Economic Indexes for Areas scores of the postcode
19 region where the participating family lived. This paper used the Socio-Economic Indexes for
20 Areas Index of Relative Socio-economic Disadvantage (Disadvantage Index), a standardised
21 score that summarises the social and economic conditions of Australian neighbourhoods
22 (national mean of 1000 and a standard deviation (SD) of 100, where higher values represent
23 less disadvantage).¹⁹

24 *Time of blood collection, processing and fasting time:* Time of blood collection and start of
25 laboratory processing were recorded. When missing, collection time was estimated using the
26 midpoint between the time the CheckPoint visit began and time that processing of the sample
27 commenced. Processing lag time was calculated as the minutes between blood collection and
28 the processing commencement. Most samples were processed within two hours.

29 Fasting time was calculated as the hours between last eating/drinking to time of blood
30 collection. The last time of eating/drinking was cross-checked against when the participant
31 was taking part in other CheckPoint stations (and known not to be eating) as well as sleep and
32 wake times from accelerometry data (to identify usual activity, and therefore likely eating
33 patterns) when available. Further details of cleaning processes for the time of last eat/drink
34 can be found elsewhere.²⁰

1 **Statistical analysis**

2 Continuous descriptive variables were summarised using weighted means and standard
3 deviations (SD) for children and adults separately, by sex and overall. Summaries of parent
4 and child metabolite measures we focussed on were reported using weighted means and SDs,
5 or weighted geometric means and relative SD for skewed metabolites, for children and adults
6 separately, by sex and overall. Population summary statistics were estimated by applying
7 survey weights and survey procedures that corrected for sampling, participation and non-
8 response biases, and took into account clustering in the sampling frame. Standard errors were
9 calculated taking into account the complex design and weights.²¹ More detail on the
10 calculation of weights is provided elsewhere.²²

11 To compare the distribution of metabolites by age and sex, density plots were examined.
12 Skewed metabolites (skewness greater or equal to 2) were log-transformed. We used two-
13 sided paired t-tests to compare mean metabolite concentrations between children and adults
14 in parent-child dyads, and t-tests to compare mean metabolite concentrations between
15 males and females for children and adults separately. The analyses were repeated using
16 weighted multi-level survey analyses and compared to unweighted analyses. As there
17 appeared to be no major effect of response patterns on results we reported results from
18 unweighted analyses.

19 Concordance between parents and children was assessed by 1) Pearson's correlation
20 coefficients (CC) with 95% confidence intervals, and 2) partial correlation coefficients
21 (PCC), controlling for age, socioeconomic status, fasting time and processing lag time. In
22 addition, the Pearson's correlation coefficients described above were repeated using weighted
23 multi-level survey analyses and compared to unweighted analyses. As there appeared to be no
24 major effect of response patterns on results we reported results from unweighted analyses.
25 Scatterplots of parent versus child metabolites (log-transformed where needed as above) were
26 examined to check for outliers and to ensure assumptions were met.

27 Analyses were undertaken using Stata version 14.2 and R version 3.3.2.

28

1 RESULTS

3 Sample characteristics

5 The recruitment and retention of participants in the Child Health CheckPoint are described
6 elsewhere.¹⁴ Of the 1874 families who participated in CheckPoint assessment centres, blood
7 serum samples of analysable quality from 1180 children and 1325 parents (figure 1) were
8 sent for NMR quantification of metabolites. The majority of excluded families undertook
9 home visits or attended a regional centre, where blood samples could not be collected (n=385,
10 20.5%), while some participants declined a blood sample (children, n=150, 8.0%; adults,
11 n=108, 5.8%). Few data were lost due to insufficient volume or poor quality samples at the
12 assessment centre (figure 1). The sample characteristics of parents and children are outlined
13 in table 2. Summary statistics for our main child and parent metabolite measures are
14 presented in supplementary table 1.

Table 2: Sample characteristics; values are weighted mean (standard deviation)

Characteristic	All	Male	Female
Child			
n	1152-1180	558-575	594-605
Age, years	12.0 (0.4)	12.0 (0.4)	12.0 (0.4)
BMI, (kg/m ²)	19.4 (3.5)	19.2 (3.4)	19.6 (3.7)
BMI z-score	0.38 (1.0)	0.40 (1.0)	0.37 (1.0)
Disadvantage Index	1012 (63)	1011 (65)	1014 (61)
Fasting time (hours)	4.2 (1.2)	4.3 (1.3)	4.2 (1.1)
Time of day - blood collection	14.16 (2.0)	14.12 (2.0)	14.20 (2.1)
Parent			
n	1272 - 1325	174-177	1098-1148
Age, years	43.9 (5.6)	46.9 (6.9)	43.4 (5.2)
BMI, (kg/m ²)	28.4 (6.4)	28.9 (4.7)	28.4 (6.6)
Fasting time (hours)	3.3 (1.6)	3.6 (2.0)	3.2 (1.5)
Time of day - blood collection	13.10 (2.0)	13.18 (2.1)	13.09 (2.0)

BMI, body mass index; Disadvantage Index: Index of Relative Socioeconomic Disadvantage; n: number of participants in cohort with this measure.

Differences in metabolite levels in children and adults

Figure 2 shows differences in mean metabolite levels for adults relative to children in standard deviation (SD) units. Most concentrations were higher in adults than children. Values that were similar in adults and children included total lipids in very large HDL lipoprotein subclass particles, acetoacetate, tyrosine and VLDL particle size. Levels in children were higher than those of adults for the glycolysis related measures (pyruvate, citrate and glycerol), the ketone body 3-hydroxybutrate, the amino acid glutamine, many fatty acid ratios and LDL and HDL particle sizes.

Supplementary table 2 lists the corresponding estimates in absolute concentration units, while supplementary figures 3-7 show density plots comparing the distributions of metabolites for boys, girls and adults.

Sex-specific differences in metabolite levels in children and adults

Figure 3 shows differences in mean metabolite levels by sex for children and adults separately in SD units, with estimates in absolute concentration units listed in supplementary table 3 and 4.

1 In general, sex differences appeared more pronounced in adulthood, resulting in distinct
2 overall patterns for children and adults. Children generally showed smaller differences by sex
3 than adults. Of note, sex differences for apolipoproteins, fatty acids and inflammation
4 measures showed different patterns in children compared to adults.

5 Girls had lower levels of apolipoprotein-A-1 (ApoA-1) and higher ApoB than boys. In adults,
6 the opposite pattern was observed with mothers having higher ApoA-1 and lower ApoB than
7 fathers. In children, most fatty acid concentrations were similar in girls and boys. In contrast,
8 many adult fatty acid measures were higher in fathers. There was no evidence of a difference
9 in the level of inflammation (GlycA) by sex in children. In adults, GlycA levels were higher
10 in fathers than mothers.

11 For some metabolites, sex differences in children mirrored (but were smaller than) those of
12 adults, particularly for the ketone bodies acetate and acetoacetate and some key amino acids.
13 At both ages, the amino acid glycine was higher in females but the branched-chain amino
14 acids leucine and valine were higher in males.

15 **Parent-child concordance**

16 Figure 4 shows the correlations between metabolite measures for all children with all parents,
17 and for boys and girls with mothers (but not with the 177 fathers, given the small numbers).
18 The corresponding correlation coefficients and partial correlation coefficients are listed in
19 supplementary tables 5 and 6.

20 A positive correlation was found for many metabolite measures irrespective of child sex, with
21 Correlation Coefficients (CC) ranging from 0.03 (95% CI -0.05 to 0.12) for acetoacetate in
22 mother-daughter pairs to 0.39 (95% CI 0.31 to 0.47) for isoleucine in mother-son
23 comparisons. Additional adjustment for factors that potentially influence metabolite levels
24 (age, socioeconomic status, fasting time and processing lag time) had little effect on the
25 degree of correlation in any comparison (supplementary tables 5 and 6). Correlations for all
26 parents and all children showed similar patterns to that observed for mother and child by sex.

27 Confidence intervals (95%) for all mother-son and mother-daughter correlations overlapped.
28 However, some metabolites showed differences by child sex. For example, fatty acids and 3-
29 hydroxybutyrate were more highly correlated in mother-daughter than mother-son pairs.
30 Other metabolites were more highly correlated in mother-son than mother-daughter
31 comparisons, including acetoacetate and the branched-chain amino acids (isoleucine, leucine
32 and valine).

1 **DISCUSSION**

2 **Principal findings**

3 Here we present age and sex differences on the detailed/NMR-based metabolic profiles from
4 1133 Australian parent-child dyads, and demonstrate that many metabolite measures have
5 high parent-child concordance. In accord with previous studies, we identified major
6 differences in metabolite levels between childhood and adulthood and also sex-specific
7 profiles in both childhood and adulthood. We also observed variability in the level of sex-
8 specific differences for several metabolites in childhood compared to adulthood and
9 identified a complex interplay of correlations of specific metabolites between parents and
10 their children according to parent-child sex relationships.

11 **Strengths and weaknesses**

12 This is the first major cohort study to report both sex- and cross-generational differences in
13 metabolomic concentrations in mid-childhood to adulthood utilising the NMR platform.
14 Further strengths include the large number of parent-child dyads representing a wide range of
15 parent ages, the national population-based sample and the state-of-the-art measurements.
16 Replication studies exploring sex differences at earlier and later stages of childhood and
17 adolescence would strengthen findings.

18 An important limitation is that paternal factors were not fully represented, as most parental
19 samples were from mothers (a well-documented problem in longitudinal cohort studies). This
20 also limited sex-specific parental contribution analysis; further studies including more fathers
21 are warranted. Additional limitations are that, without samples from both parents for each
22 child, we could not estimate heritability, and our results might not apply to mid-life adults
23 who are not parents (although we see no good reason why these would differ greatly). The
24 original uptake of just over 50% and subsequent attrition within LSAC and then the
25 CheckPoint has led to a relatively advantaged sample, but nonetheless participants varied
26 widely on key potential confounders (eg disadvantage, age) and this was at least partly offset
27 by application or consideration of survey weights. Given the large number of metabolites and
28 modest sample size, considerable uncertainty remains in any ranking of the various effects
29 across metabolites.

30 **Meaning and implications for clinicians and policymakers**

31 Overall, we found a clear difference in metabolite profile between children and their parents.
32 This was apparent for specific metabolite measures (such as some amino acids) as well as the

1 distribution of metabolites (such as lipid composition of lipoproteins of different density).
2 Some measures were higher in adults, some similar, while a minority were lower. Previous
3 studies, largely in adults, have identified a range of specific metabolite changes with age,
4 particularly from mid to late adulthood.²³ This includes a general decrease in several amino
5 acid species, which contrasts with our findings from childhood to mid adulthood.⁸ Only the
6 amino acid glutamine showed this pattern in our dataset.

7 Sex-specific differences in children (± 0.2 SD) were generally much smaller than in adults
8 (± 0.8 SD). Large metabolomic studies using alternative platforms have previously reported
9 reproducible, sex-specific signatures in circulating metabolite profile in adults.^{24 25} This
10 includes differences in amino acid and lipid serum concentrations, potentially influenced by
11 sex-specific effects of genetic polymorphisms on metabolite levels.^{25 26} As in our study,
12 most amino acids have usually been reported to be higher in men than women.^{25 27} For
13 example, in a recent study of 507 metabolic markers in 1756 individuals (903 female and 853
14 male aged ~ 60 years), one third of metabolites showed significant sexual dimorphism. These
15 were predominantly related to pathways of steroid metabolism, fatty acids, other lipids, and a
16 large proportion of amino acids.²⁷ Of particular note, branched chain amino acids (BCAAs)
17 and their related metabolic products were amongst the most differentially represented, with
18 much higher isoleucine, leucine and valine in males. A similar finding of higher leucine and
19 valine was also noted in the Cooperative Health Research in the Region of Augsburg
20 (KORA) follow-ups 3 (F3) and 4 (F4) analysis of >3000 adults,²⁵ consistent with our
21 observations in adulthood.

22 In children, we found sex-specific differences for leucine and valine were smaller but in the
23 same direction as adults. Several lines of evidence implicate BCAA metabolism with
24 metabolic risk in humans. For example, three candidate genes for obesity and/or type 2
25 diabetes mellitus (T2DM) are involved in the BCAA metabolic pathway.²⁸ In a recent large
26 meta-analysis of metabolomics in diabetes, a $>30\%$ higher risk of type 2 diabetes was found
27 per SD increase in isoleucine, leucine, valine or tyrosine, whereas glycine and glutamine
28 were inversely associated with risk.²⁸ Several clinical studies have also reported that BCAAs
29 positively correlate with insulin resistance, homeostatic model assessment (HOMA) index
30 and levels of haemoglobin A1c (HbA1c), while longitudinal studies have reported that
31 increased blood BCAAs are predictive of future insulin resistance and type 2 diabetes
32 (T2D).²⁹ It is intriguing to speculate that the higher BCAA in males from early life could
33 contribute to the well-described increasing prevalence of T2D in men. Levels of BCAA are

1 elevated in females with Polycystic Ovary Syndrome (PCOS), potentially contributing to the
2 associated insulin resistance.³⁰ However, it remains unclear whether BCAA are on the causal
3 pathway to T2D or result from adverse metabolic health. Our demonstration that the sex-
4 specific differences in BCAA arise early in life offers potential to track their association with
5 sex-specific measures of metabolic health from an early age to help clarify where they lie on
6 the causal pathway.

7 In accord with previous adult studies²⁵, we found higher levels of glycine in mothers than
8 fathers, and (less markedly) in girls than boys. Interestingly, recent metabolomics and genetic
9 analyses of ~10,000 adults with cardiovascular disease (CVD), with replication in >53,000
10 subjects, identified a genetic variant in carbamoyl-phosphate synthase 1 (*CPS1*) (linked to
11 plasma glycine levels) to be strongly associated with a reduced risk of CVD in women
12 ($p=6.3 \times 10^{-5}$) but not men ($p=0.95$), suggesting a direct link between glycine levels and CVD
13 risk, although whether this is a causal association remains unclear.³¹ It will be interesting in
14 the future to explore the link between variants in *CPS1* and circulating glycine levels from
15 early life to adulthood in relation to markers of cardiovascular health in females.

16 The small sex-differences of HDL cholesterol and ApoA-1 in children compared to adults is
17 consistent with modest differences in children, whereas substantial differences in adulthood
18 have previously been reported.³² ApoA-1 was more abundant in boys, while ApoB was
19 higher in girls, leading to a higher ApoB/ApoA-1 ratio in girls. The opposite pattern was
20 found in our limited sample of fathers relative to mothers. These data are surprising and
21 differ from a similarly sized study of slightly older European adolescent children (mean age
22 15 years) that found higher ApoA-1 and ApoB in girls relative to boys.³³ Interestingly, a
23 higher ApoB/ApoA-1 ratio has been strongly linked to increased coronary risk in adults,³⁴⁻³⁶
24 suggesting that the sex-specific differences may alter with increasing age, in keeping with the
25 increased CVD risk in adult males. ApoA-1 is the main protein component of HDL
26 cholesterol³⁷ thus the differences in trajectories in lipids and HDL cholesterol for boys and
27 girls across childhood that have been reported^{38,39} could partially explain this observation.

28 These are the first data on the mother-child or parent-child correlations of NMR metabolites.
29 Smaller studies have reported positive correlations between parents and children for a limited
30 range of cardiometabolic risk factors including total cholesterol, LDL cholesterol, HDL
31 cholesterol and triglycerides measured using conventional methods. We found positive
32 correlations between parents and children for the same lipid measures (although measured
33 using NMR) consistent with previously reported findings. One study reported a positive

1 association between the serum lipid levels of 4 year old children (n=127) and their parents
2 (122 mothers and 118 fathers)⁴⁰ while another study of children aged 6-18 (n=255) and their
3 parents (n=179) found that the age of the child influenced the degree of correlation of several
4 lipid measures, with older (10-18 years) children more similar to their parents in terms of
5 triglyceride levels than younger individuals (6-9 years).¹²

6 **Unanswered questions and future research**

7 The temporal and sex specific dynamism of the metabolomics data we describe here offer
8 considerable opportunities for identification of biomarkers of risk for a range of non-
9 communicable diseases early in life, to inform targeted interventions and monitor their
10 efficacy. Combining metabolomics with other 'omics data (such as genetics), as is
11 increasingly reported from large adult studies, offers considerable promise in understanding
12 the causal pathways that link early life exposures, genetics and intermediate phenotypes with
13 later onset chronic disease, and in identifying clinically relevant biomarkers.

14 In conclusion, we report detailed circulating metabolite profile from mid-childhood and
15 adulthood in a population-based sample, together with parent-child concordance and sex-
16 specific profiles in children and adults. Distinct age- and sex-specific profiles were observed,
17 as well as considerable evidence of parent-child concordance. These data will be informative
18 for investigation of the childhood origins of adult non-communicable diseases and for
19 comparative studies across populations.

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2 This paper uses unit record data from *Growing Up in Australia*, the Longitudinal Study of
3 Australian Children. The study is conducted in partnership between the Department of Social
4 Services (DSS), the Australian Institute of Family Studies (AIFS) and the Australian Bureau
5 of Statistics (ABS). The findings and views reported in this paper are those of the author and
6 should not be attributed to DSS, AIFS or the ABS.

7
8 REDCap (Research Electronic Data Capture) tools⁴¹ were used in this study. More
9 information about this software can be found at: www.project-redcap.org.

10 We thank the LSAC and CheckPoint study participants, staff and students for their
11 contributions.

12 COMPETING INTERESTS

13
14 All authors have completed the ICMJE uniform disclosure form at
15 www.icmje.org/coi_disclosure.pdf and declare financial support for the submitted work from
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27
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12 **CONTRIBUTIONS**

14 DB, RS and JC conceptualised and developed the Metabolomics Checkpoint study. SE and
15 JC undertook all aspects of data analysis. SAC coordinated the acquisition of metabolomics
16 data and provided critical review of this manuscript. MW, the Principal Investigator of the
17 Child Health CheckPoint, planned the analyses and provided critical review of this
18 manuscript. SE and RS drafted the manuscript. PW, MJ, TD, KL, JC, DB provided critical
19 expert advice and critical review of this manuscript.

20 **DATA SHARING STATEMENT**

22 Dataset and technical documents available from *Growing Up in Australia: The Longitudinal*
23 *Study of Australian Children* via low-cost license for bona fide researchers. More information
24 is available at www.growingupinaustralia.gov.au

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1 **FIGURE CAPTIONS AND FOOTNOTES**

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3 **Figure 1: Participant flow chart.**

4 n=number of families, c=number of children, p=number of attending adults,

5 MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,

6 LSAC=Longitudinal Study of Australian Children

7 *Unable to analyse due to insufficient volume or poor quality sample

8 ^Data from 6 non-biological child-parent pairs excluded from concordance analyses

9

10 **Figure 2: Differences in metabolite levels between children and adults.**

11 Association measures are SD difference in metabolite concentration for adults compared to
12 children. Error bars represent 95% confidence intervals. Association measures in absolute
13 concentration units, 95% confidence intervals and associated p-values are listed in
14 supplementary table 2. HDL: High-density lipoprotein; IDL: Intermediate density
15 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

16

17 **Figure 3: Sex-specific differences in metabolite levels in childhood and adulthood.**

18 Association measures are SD difference in metabolite concentration for females compared to
19 males in children (A) and adults (B). Error bars represent 95% confidence intervals.
20 Association measures in absolute concentration units, 95% confidence intervals and
21 associated p-values are listed in supplementary table 3 and 4. HDL: High-density lipoprotein;
22 IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low
23 density lipoprotein.

24

25 **Figure 4: Parent:child correlation for metabolite measures.**

26 Pearson's correlation coefficients for all children with all parents (A); and for boys (blue)
27 with mothers and for girls (red) with mothers (B). Error bars represent 95% confidence
28 intervals. Correlation coefficients with associated 95% confidence intervals are listed in
29 supplementary table 5 and 6. HDL: High-density lipoprotein; IDL: Intermediate density
30 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

31

SUPPLEMENTARY DOCUMENTS

Supplementary figure 1: Correlation of NMR metabolite measures in children.

Heatmap showing the correlation between metabolite measures in children. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 2: Correlation of NMR metabolite measures in parents.

Heatmap showing the correlation between metabolite measures in parents. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 3: Density plots for cholesterol and apolipoprotein measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each cholesterol and apolipoprotein measure.

Supplementary figure 4: Density plots for fatty acid and fatty acid ratio measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each fatty acid and fatty acid ratio measure.

Supplementary figure 5: Density plots for total lipids in the 14 lipoprotein subclass particles.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for total lipids within each of the 14 lipoprotein subclass particles.

Supplementary figure 6: Density plots for lipoprotein particle size and triglyceride measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for lipoprotein particle sizes and triglyceride measures.

Supplementary figure 7: Density plots for glycolysis related, amino acid, ketone body and inflammation measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for glycolysis related, amino acid, ketone body and inflammation measures.

Supplementary table 1: Weighted mean (SD) of metabolite measures in children and parents.

Supplementary table 2: Differences in mean metabolite levels in adults compared to children in absolute concentration units.

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1 **Supplementary table 3: Differences in mean metabolite levels in girls compared to boys**
2 **in absolute concentration units.**

4 **Supplementary table 4: Differences in mean metabolite levels in female compared to**
5 **male adults in absolute concentration units.**

7 **Supplementary table 5: Mother-child concordance; correlations and partial correlations**
8 **between mothers and their sons, daughters and all children.**

10 **Supplementary table 6: Parent-child concordance; correlation and partial correlations**
11 **between all parents and their sons, daughters and all children.**

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3 **1 REFERENCES**

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1. Nicholson JK, Lindon JC, Holmes E. 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica* 1999;**29**(11):1181-9 doi: 10.1080/004982599238047.
 2. Nath AP, Ritchie SC, Byars SG, et al. An interaction map of circulating metabolites, immune gene networks, and their genetic regulation. *Genome biology* 2017;**18**(1):146 doi: 10.1186/s13059-017-1279-y.
 3. Shah SH, Newgard CB. Integrated metabolomics and genomics: systems approaches to biomarkers and mechanisms of cardiovascular disease. *Circ Cardiovasc Genet* 2015;**8**(2):410-9 doi: 10.1161/CIRCGENETICS.114.000223.
 4. Kaikkonen JE, Wurtz P, Suomela E, et al. Metabolic profiling of fatty liver in young and middle-aged adults: Cross-sectional and prospective analyses of the Young Finns Study. *Hepatology* 2017;**65**(2):491-500 doi: 10.1002/hep.28899.
 5. Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol* 2016;**17**(7):451-9 doi: 10.1038/nrm.2016.25.
 6. Kettunen J, Tukiainen T, Sarin AP, et al. Genome-wide association study identifies multiple loci influencing human serum metabolite levels. *Nat Genet* 2012;**44**(3):269-76 doi: 10.1038/ng.1073.
 7. Suhre K, Gieger C. Genetic variation in metabolic phenotypes: study designs and applications. *Nat Rev Genet* 2012;**13**(11):759-69 doi: 10.1038/nrg3314.
 8. Yu Z, Zhai G, Singmann P, et al. Human serum metabolic profiles are age dependent. *Aging Cell* 2012;**11**(6):960-7 doi: 10.1111/j.1474-9726.2012.00865.x.
 9. Auro K, Joensuu A, Fischer K, et al. A metabolic view on menopause and ageing. *Nat Commun* 2014;**5**:4708 doi: 10.1038/ncomms5708.
 10. Saito K, Maekawa K, Kinchen JM, et al. Gender- and Age-Associated Differences in Serum Metabolite Profiles among Japanese Populations. *Biol Pharm Bull* 2016;**39**(7):1179-86 doi: 10.1248/bpb.b16-00226.
 11. Wang Q, Wurtz P, Auro K, et al. Metabolic profiling of pregnancy: cross-sectional and longitudinal evidence. *BMC Med* 2016;**14**(1):205 doi: 10.1186/s12916-016-0733-0.
 12. Halvorsen T, Moran A, Jacobs DR, Jr., et al. Relation of Cardiometabolic Risk Factors between Parents and Children. *J Pediatr* 2015;**167**(5):1049-56 e2 doi: 10.1016/j.jpeds.2015.07.053.
 13. Wake M, Clifford S, York E, et al. Introducing Growing Up in Australia's Child Health CheckPoint: A physical and biomarkers module for the Longitudinal Study of Australian Children. *Family Matters* 2014;**95**:15-23.
 14. Clifford S, Davies S, Wake M. Growing Up in Australia's Child Health CheckPoint cohort summary and methodology. Submitted to BMJ Open October 2017.
 15. Soininen P, Kangas AJ, Wurtz P, et al. High-throughput serum NMR metabonomics for cost-effective holistic studies on systemic metabolism. *Analyst* 2009;**134**(9):1781-5 doi: 10.1039/b910205a.
 16. Wurtz P, Kangas AJ, Soininen P, et al. Quantitative Serum NMR Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technology. *Am J Epidemiol* 2017.

17. Lu M, Lu Q, Zhang Y, et al. ApoB/apoA1 is an effective predictor of coronary heart disease risk in overweight and obesity. *J Biomed Res* 2011;**25**(4):266-73 doi: 10.1016/S1674-8301(11)60036-5.
18. Kuczmarski RJ, Ogden CL, Grummer-Strawn LM, et al. CDC growth charts: United States. *Adv Data* 2000(314):1-27.
19. Australian Bureau of S. Census of population and housing: Socio-Economic Indexes for Areas (SEIFA) 2011. Cat. no. 2033.0.55.001, 2011.
20. Davies S, Clifford S, Gillespie A, et al. LSAC's Child Health CheckPoint Data Issues Paper 2017. *Melbourne, Australia: Murdoch Children's Research Institute* 2017.
21. S.G. H, B.T. W, P.A. B. *Applied survey data analysis*. Boca Raton.: CRC press, 2010.
22. Ellul S, Hiscock R, Mensah F, et al. Longitudinal Study of Australian Children's Child Health CheckPoint: Weighting and Non-Response. 2017.
23. Menni C, Kastenmüller G, Petersen AK, et al. Metabolomic markers reveal novel pathways of ageing and early development in human populations. *International Journal of Epidemiology* 2013;**42**(4):1111-19 doi: 10.1093/ije/dyt094.
24. Dunn WB, Lin W, Broadhurst D, et al. Molecular phenotyping of a UK population: defining the human serum metabolome. *Metabolomics* 2015;**11**:9-26 doi: 10.1007/s11306-014-0707-1.
25. Mittelstrass K, Ried JS, Yu Z, et al. Discovery of sexual dimorphisms in metabolic and genetic biomarkers. *PLoS Genet* 2011;**7**(8):e1002215 doi: 10.1371/journal.pgen.1002215.
26. Kolz M, Johnson T, Sanna S, et al. Meta-analysis of 28,141 individuals identifies common variants within five new loci that influence uric acid concentrations. *PLoS Genet* 2009;**5**(6):e1000504 doi: 10.1371/journal.pgen.1000504.
27. Krumsiek J, Mittelstrass K, Do KT, et al. Gender-specific pathway differences in the human serum metabolome. *Metabolomics* 2015;**11**(6):1815-33 doi: 10.1007/s11306-015-0829-0.
28. Guasch-Ferré M, Hruby A, Toledo E, et al. Metabolomics in Prediabetes and Diabetes: A Systematic Review and Meta-analysis. *Diabetes Care* 2016;**39**(5):833-46 doi: 10.2337/dc15-2251.
29. Lynch CJ, Adams SH. Branched-chain amino acids in metabolic signalling and insulin resistance. *Nature reviews Endocrinology* 2014;**10**(12):723-36 doi: 10.1038/nrendo.2014.171.
30. Chang AY, Lalia AZ, Jenkins GD, et al. Combining a nontargeted and targeted metabolomics approach to identify metabolic pathways significantly altered in polycystic ovary syndrome. *Metabolism* 2017;**71**(Supplement C):52-63 doi: <https://doi.org/10.1016/j.metabol.2017.03.002>.
31. Hartiala JA, Wilson Tang WH, Wang Z, et al. Genome-wide association study and targeted metabolomics identifies sex-specific association of CPS1 with coronary artery disease. *Nature Communications* 2016;**7**:10558 doi: 10.1038/ncomms10558.
32. Davis CE, Williams DH, Oganov RG, et al. Sex Difference in High Density Lipoprotein Cholesterol in Six Countries. *American Journal of Epidemiology* 1996;**143**(11):1100-06 doi: 10.1093/oxfordjournals.aje.a008686.
33. Spinneker A, Egert S, Gonzalez-Gross M, et al. Lipid, lipoprotein and apolipoprotein profiles in European adolescents and its associations with gender, biological maturity and body fat--the HELENA Study. *Eur J Clin Nutr* 2012;**66**(6):727-35 doi: 10.1038/ejcn.2011.222.

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2
3 1 34. Walldius G, Jungner I, Aastveit AH, et al. The apoB/apoA-I ratio is better than the
4 2 cholesterol ratios to estimate the balance between plasma proatherogenic and
5 3 antiatherogenic lipoproteins and to predict coronary risk. *Clin Chem Lab Med*
6 4 2004;**42**(12):1355-63 doi: 10.1515/CCLM.2004.254.
7 5
8 5 35. Walldius G, Jungner I. The apoB/apoA-I ratio: a strong, new risk factor for cardiovascular
9 6 disease and a target for lipid-lowering therapy--a review of the evidence. *J Intern*
10 7 *Med* 2006;**259**(5):493-519 doi: 10.1111/j.1365-2796.2006.01643.x.
11 8
12 8 36. Mozaffarian D, Benjamin EJ, Go AS, et al. Heart disease and stroke statistics--2015
13 9 update: a report from the American Heart Association. *Circulation* 2015;**131**(4):e29-
14 10 322 doi: 10.1161/CIR.000000000000152.
15 11
16 11 37. Upadhyay RK. Emerging Risk Biomarkers in Cardiovascular Diseases and Disorders.
17 12 *Journal of Lipids* 2015;**2015**:971453 doi: 10.1155/2015/971453.
18 13
19 13 38. Hardy R, Lawlor DA, Kuh D. A life course approach to cardiovascular aging. *Future*
20 14 *cardiology* 2015;**11**(1):101-13 doi: 10.2217/fca.14.67.
21 15
22 15 39. Jolliffe CJ, Janssen I. Distribution of Lipoproteins by Age and Gender in Adolescents.
23 16 *Circulation* 2006;**114**(10):1056.
24 17
25 17 40. Ohlund I, Hernell O, Hornell A, et al. Serum lipid and apolipoprotein levels in 4-year-old
26 18 children are associated with parental levels and track over time. *Eur J Clin Nutr*
27 19 2011;**65**(4):463-69.
28 20
29 20 41. Harris PA, Taylor R, Thielke R, et al. Research electronic data capture (REDCap)—A
30 21 metadata-driven methodology and workflow process for providing translational
31 22 research informatics support. *Journal of Biomedical Informatics* 2009;**42**(2):377-81
32 23 doi: <https://doi.org/10.1016/j.jbi.2008.08.010>.
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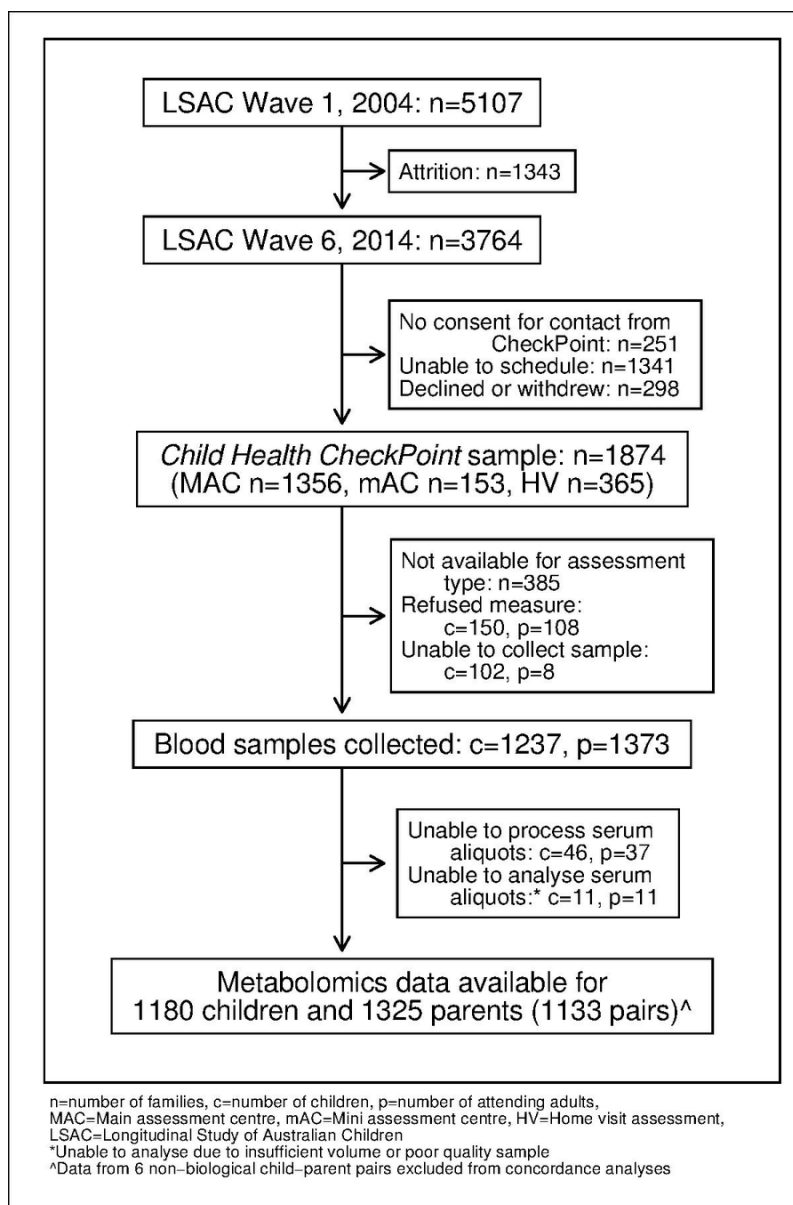


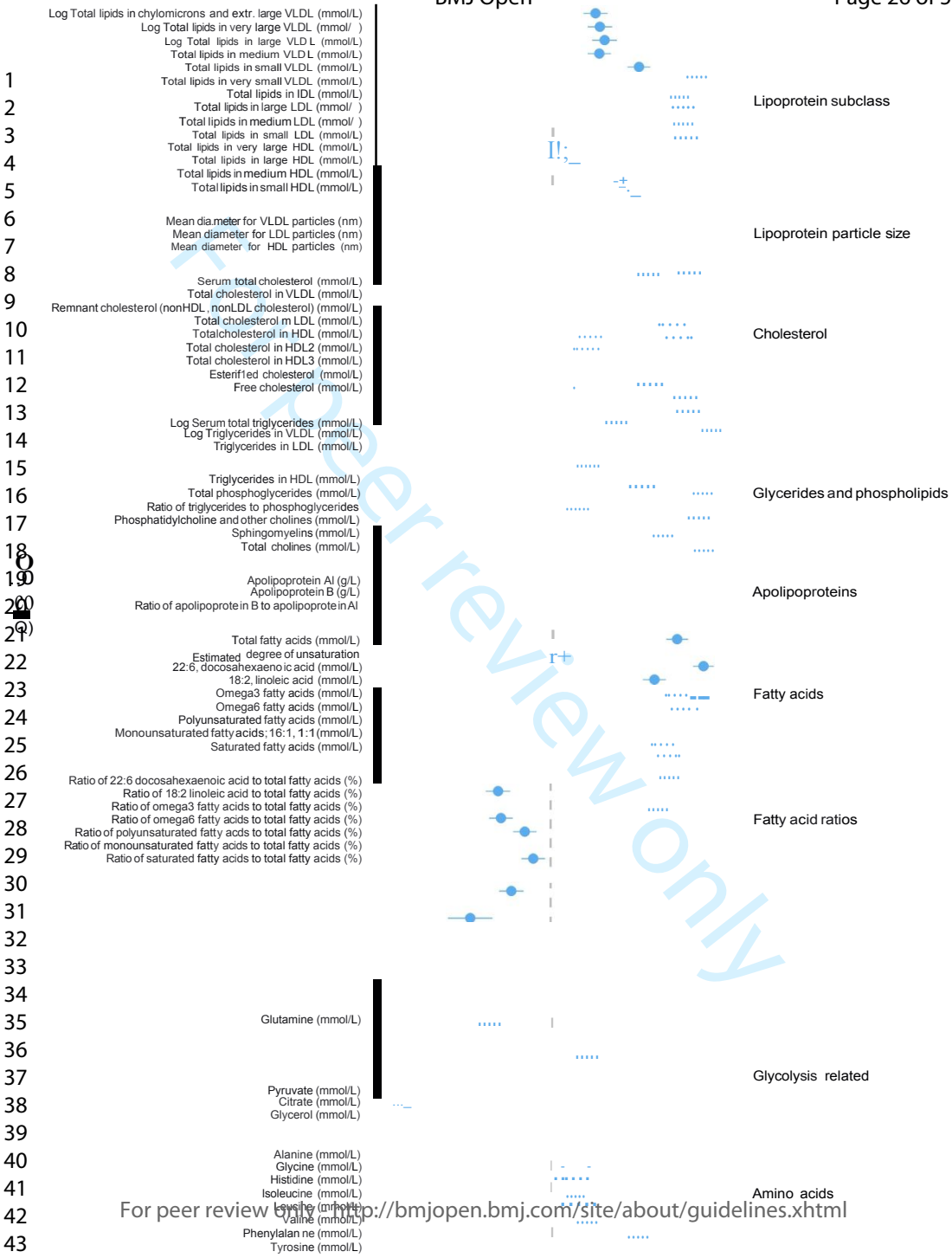
Figure 1: Participant flow chart.

n=number of families, c=number of children, p=number of attending adults,
MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,
LSAC=Longitudinal Study of Australian Children

*Unable to analyse due to insufficient volume or poor quality sample

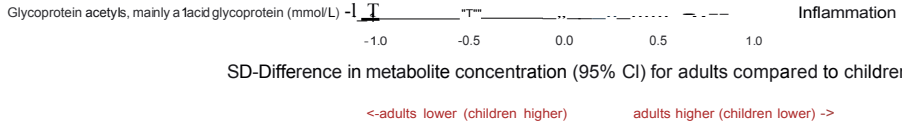
^Data from 6 non-biological child-parent pairs excluded from concordance analyses

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Log Acetate (mmol/L)
Log Acetoacetate (mmol/L)
Log 3hydroxybutyrate (mmol/L)

Ketone bodies



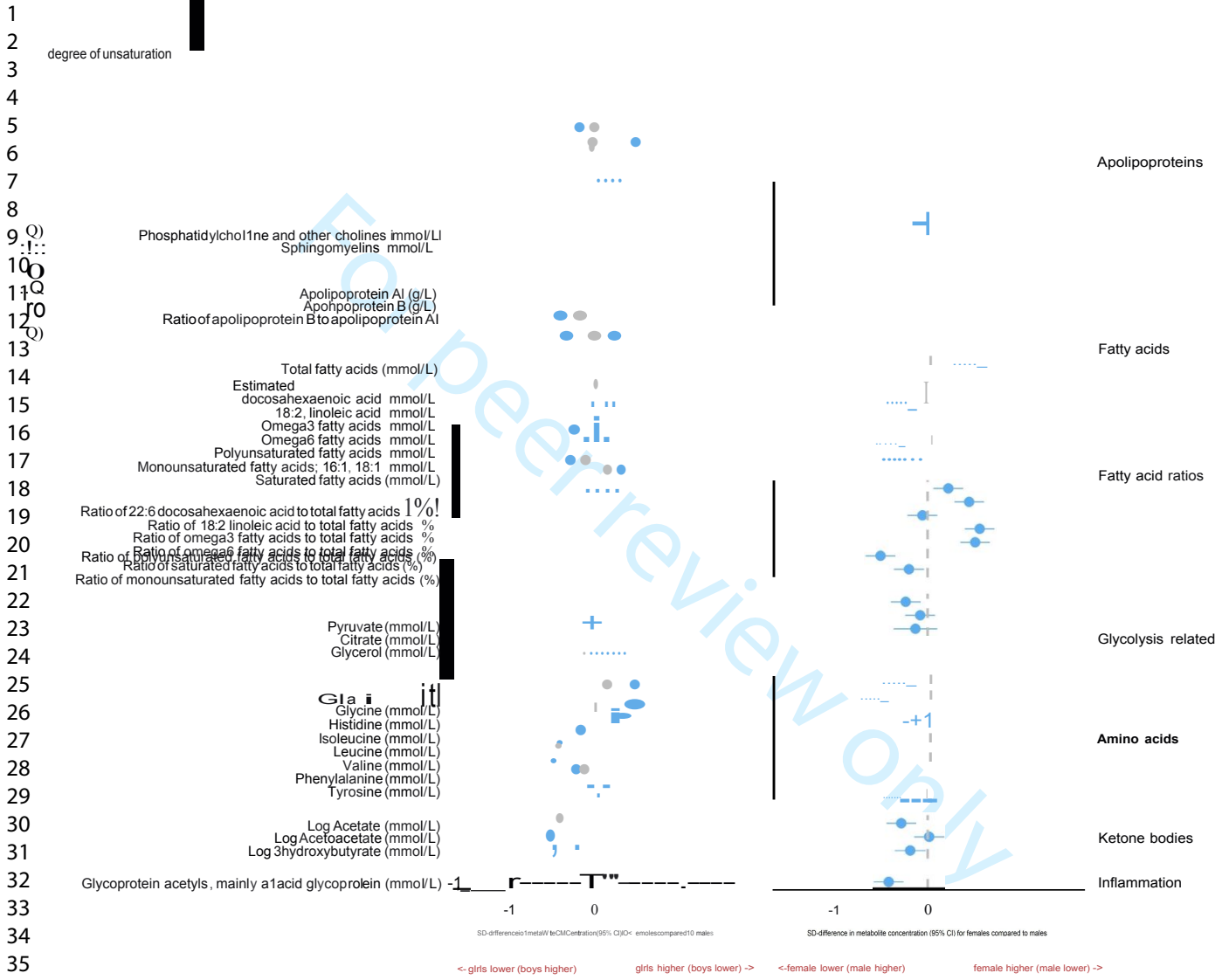
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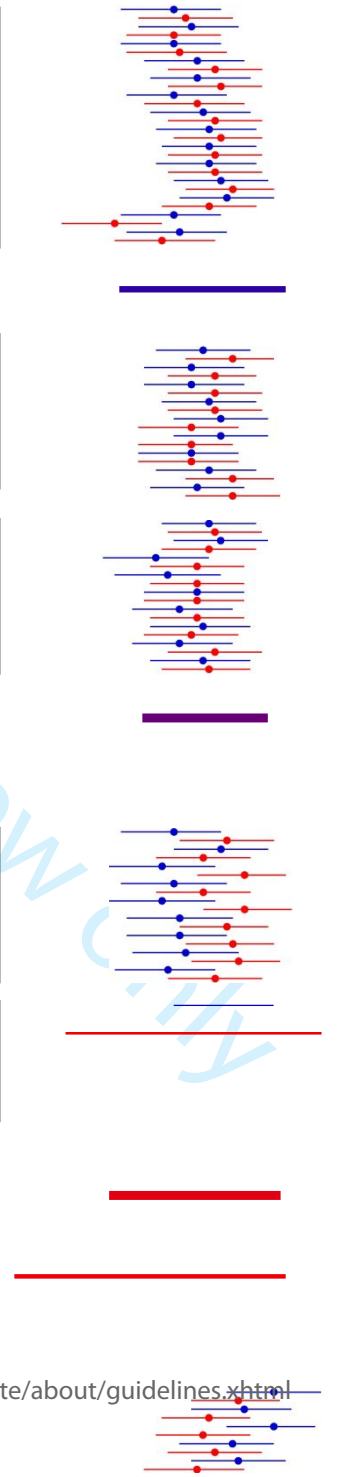
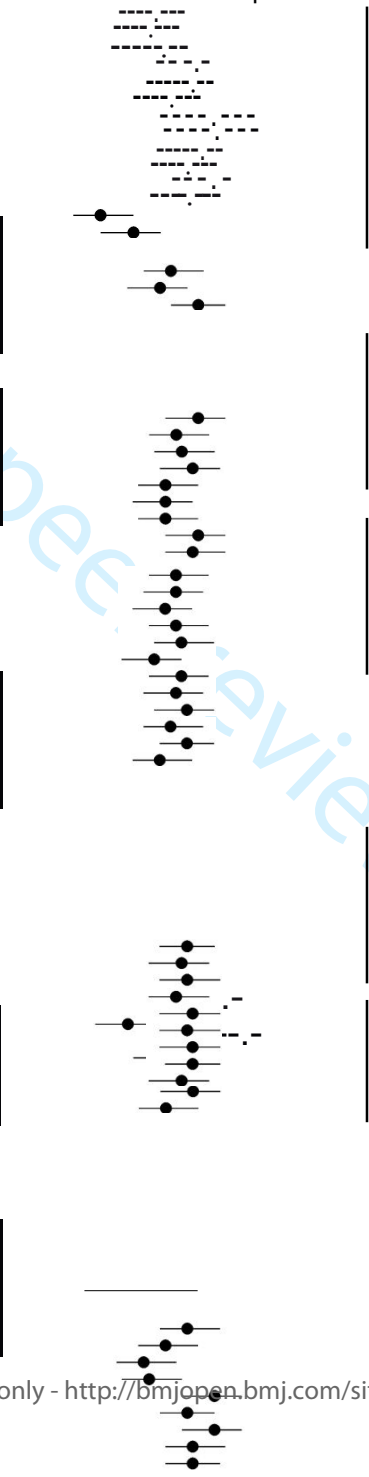


All children with all parents
BMJ Open

Boys and girls with mothers

Boys
Girls

- 1 Log Total lipids in chylomicrons & ex. large VLDL (mmol/L)
- 2 Log Total lipids in very large VLDL (mmol/L)
- 3 Log Total lipids in large VLDL (mmol/L)
- 4 Total lipids in medium VLDL (mmol/L)
- 5 Total lipids in small VLDL (mmol/L)
- 6 Total lipids in very small VLDL (mmol/L)
- 7 Total lipids in IDL (mmol/L)
- 8 Total lipids in large LDL (mmol/L)
- 9 Total lipids in medium LDL (mmol/L)
- 10 Total lipids in small LDL (mmol/L)
- 11 Total lipids in very large HDL (mmol/L)
- 12 Total lipids in large HDL (mmol/L)
- 13 Total lipids in medium HDL (mmol/L)
- 14 Total lipids in small HDL (mmol/L)
- 15 Mean diameter for VLDL particles (nm)
- 16 Mean diameter for LDL particles (nm)
- 17 Mean diameter for HDL particles (nm)
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- 19 cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)
- 20 Remnant
- 21 Serum total cholesterol (mmol/L)
- 22 Total cholesterol in VLDL (mmol/L)
- 23 Total cholesterol in LDL (mmol/L)
- 24 Total cholesterol in HDL (mmol/L)
- 25 Total cholesterol in HDL2 (mmol/L)
- 26 Total cholesterol in HDL3 (mmol/L)
- 27 Esterified cholesterol (mmol/L)
- 28 Free cholesterol (mmol/L)
- 29 Log Serum total triglycerides (mmol/L)
- 30 Log Triglycerides in VLDL (mmol/L)
- 31 Triglycerides in LDL (mmol/L)
- 32 Triglycerides in HDL (mmol/L)
- 33 Total phosphoglycerides (mmol/L)
- 34 Ratio of triglycerides to phosphoglycerides
- 35 Phosphatidylcholine & other cholines (mmol/L)
- 36 Sphingomyelins (mmol/L)
- 37 Total cholines (mmol/L)
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- 39 Apolipoprotein A1 (g/L)
- 40 Apolipoprotein B (g/L)
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Lipoprotein subclass lipids

Lipoprotein particle size

Cholesterol

Glycerides and

phospholipids Apolipoproteins

Fatty acids

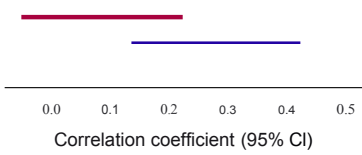
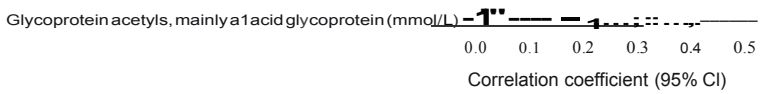
Fatty acid ratios

Glycolysis related

Amino acids

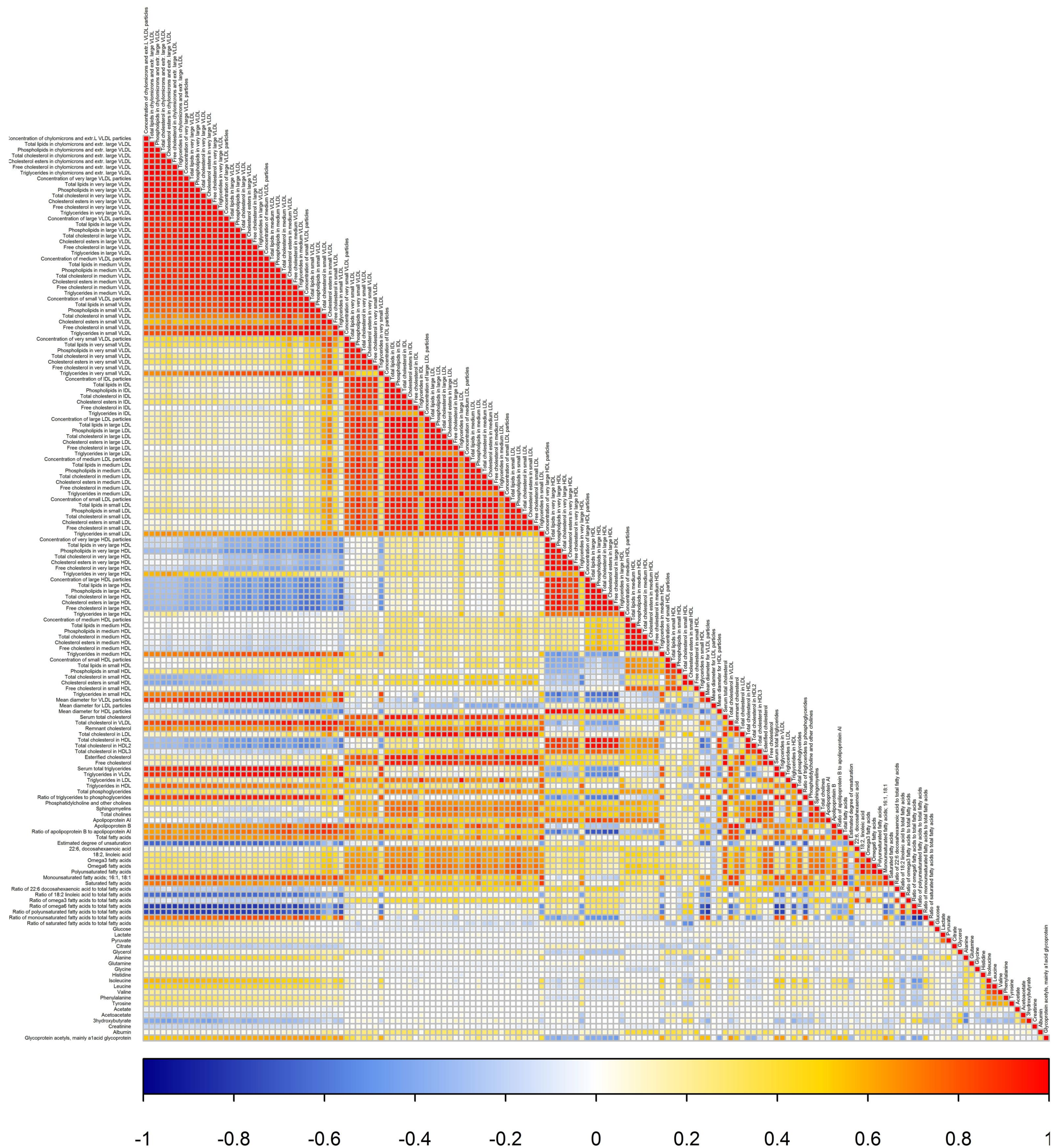
Log Acetate (mmol/L)
Log Acetoacetate (mmol/L)
Log 3hydroxybutyrate (mmol/L)

Ketone bodies



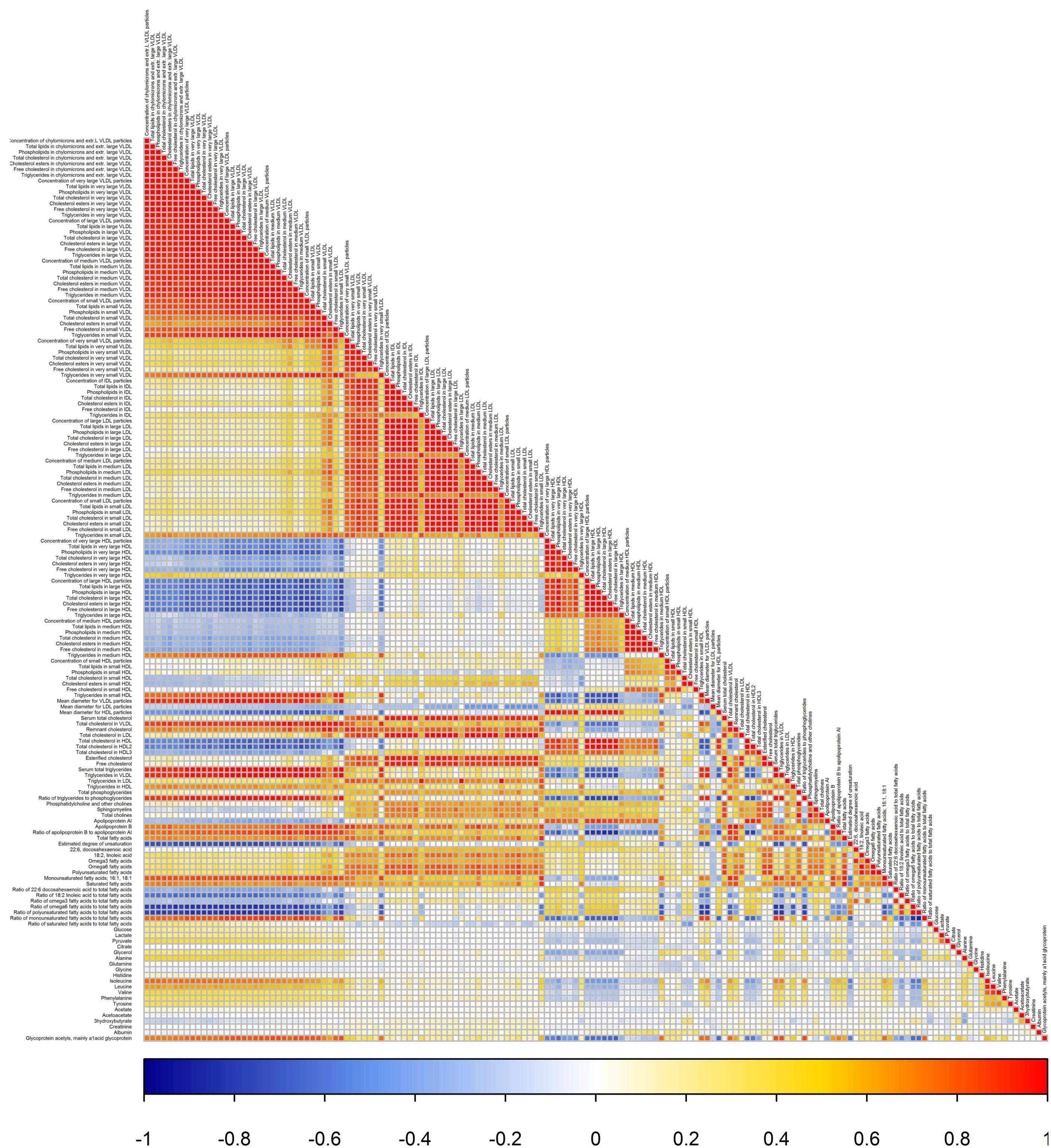
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Supplementary Figure 1: Correlations between Child Health CheckPoint metabolites - children



Note: Correlations (spearman) between metabolites for the CheckPoint child metabolomics data

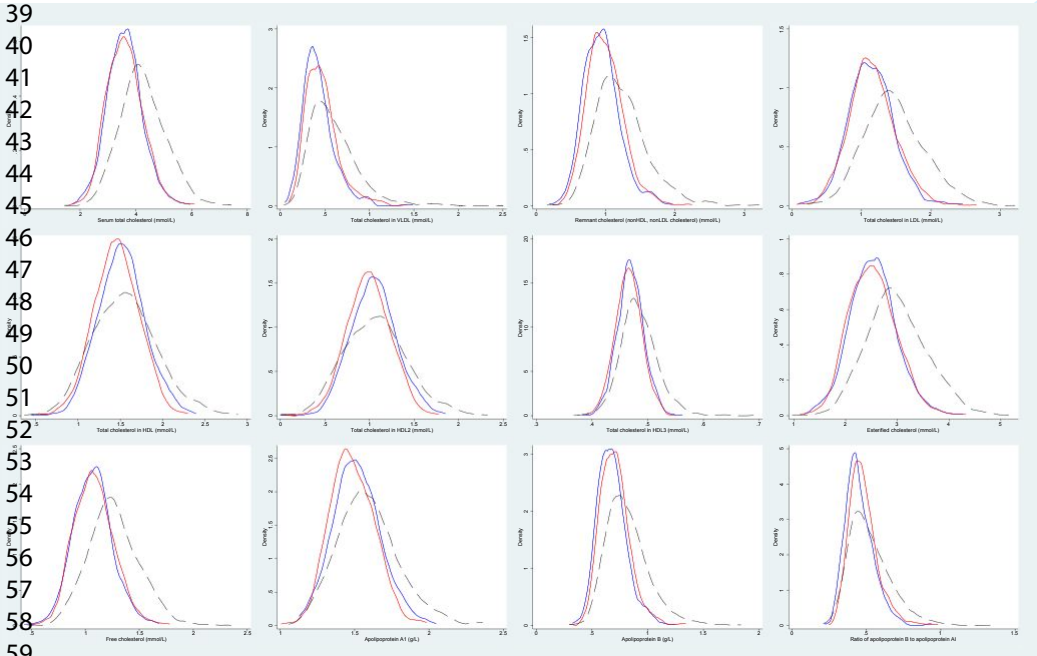
Supplementary Figure 2: Correlations between Child Health CheckPoint metabolites - parents



Note: Correlations (spearman) between metabolites for the CheckPoint parent metabolomics data

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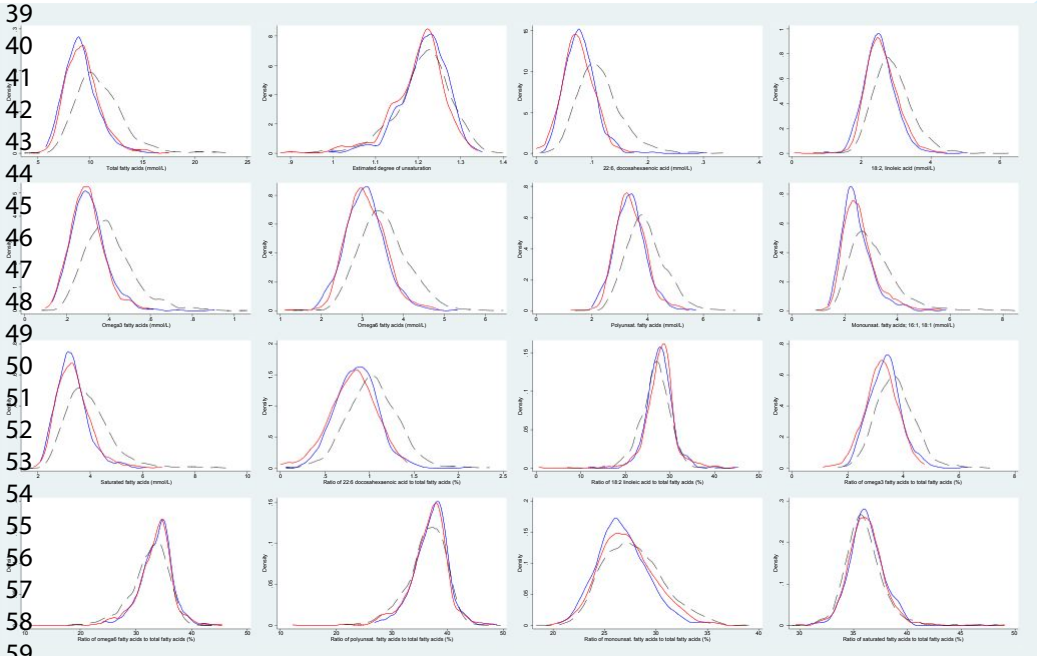
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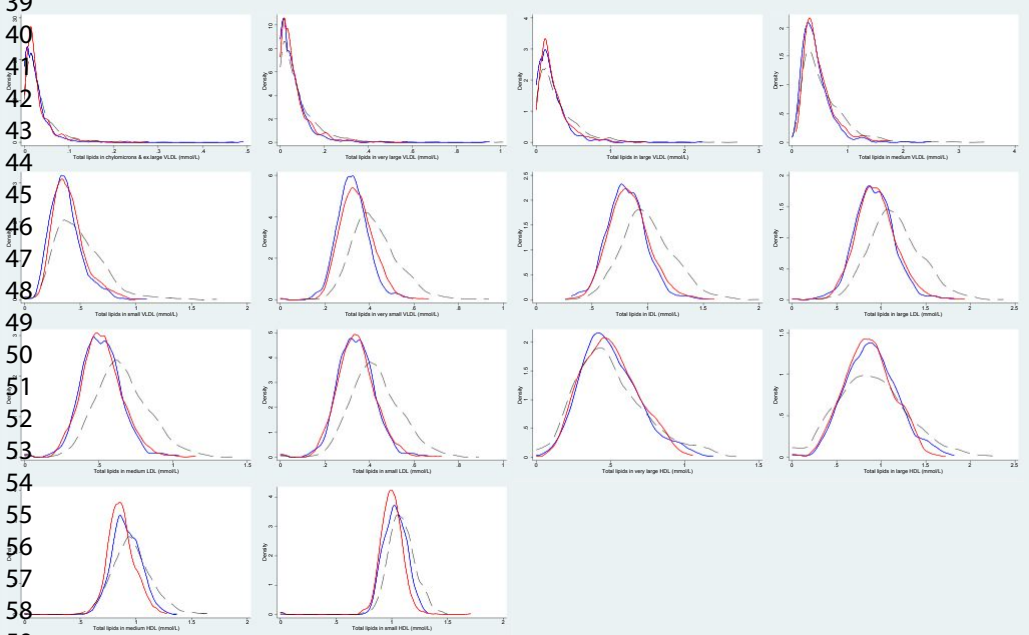
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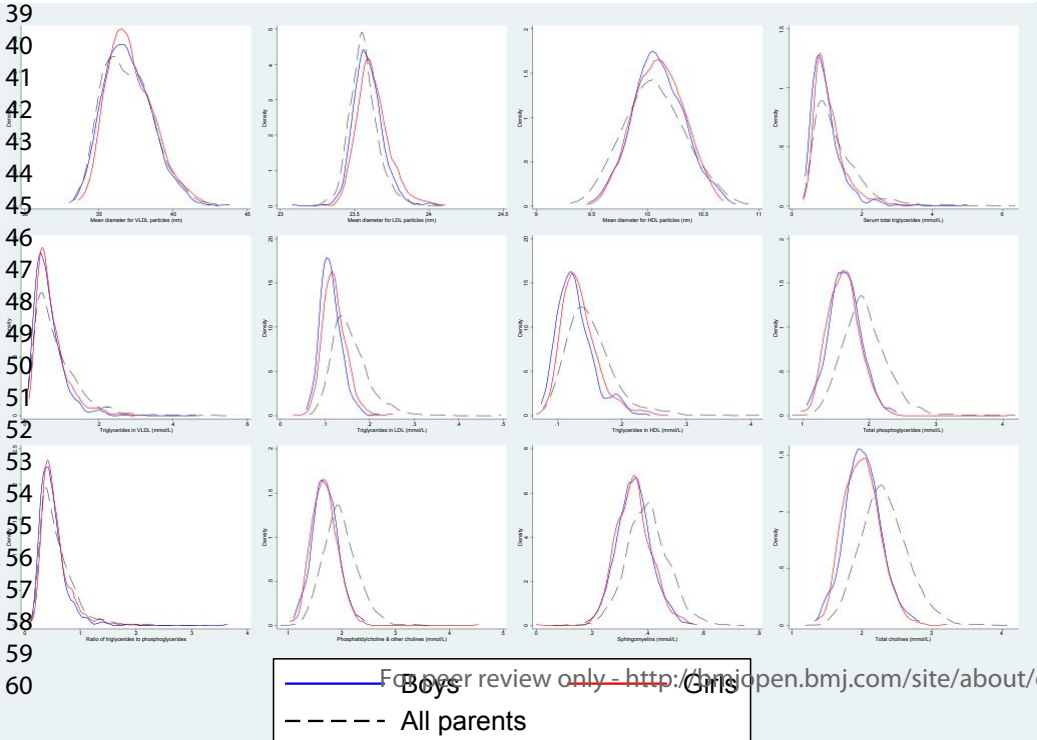
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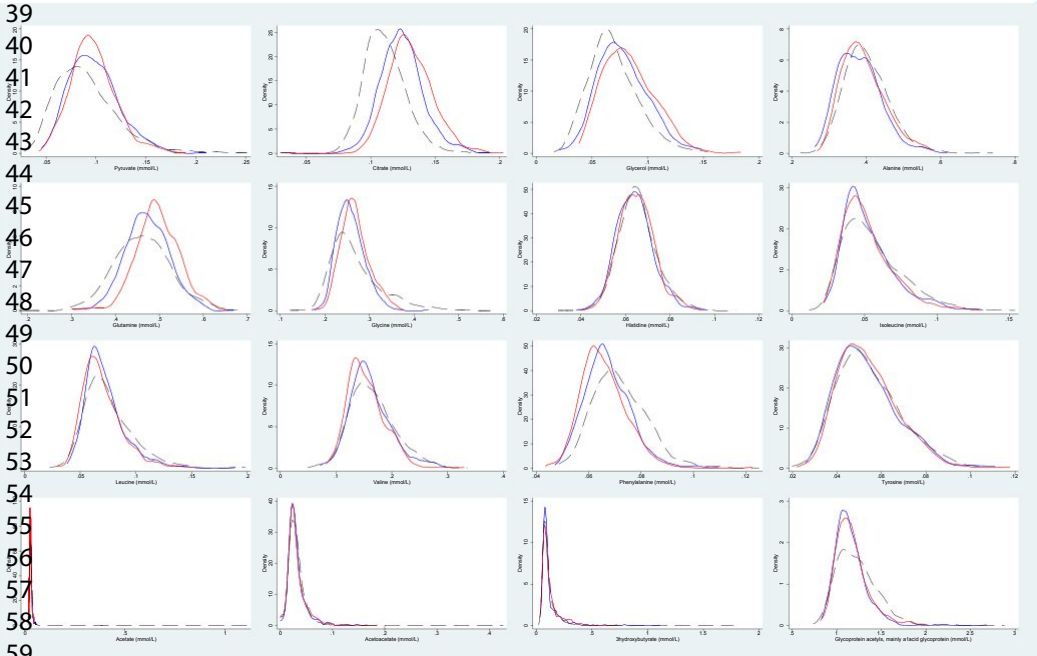
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— Boys
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Supplementary table 1: Weighted mean (SD)* of metabolite measures in children and parents.

Metabolic subgroup	Children									Adults								
	Boys			Girls			All			Male			Female			All		
	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	575	0.005	0.60	605	0.011	0.54	1180	0.007	0.58	177	0.040	0.53	1148	0.013	0.51	1325	0.015	0.52
Total lipids in very large VLDL (mmol/L)*	575	0.006	0.73	605	0.014	0.74	1180	0.010	0.74	177	0.087	0.79	1148	0.022	0.67	1325	0.027	0.70
Total lipids in large VLDL (mmol/L)*	575	0.059	0.99	605	0.121	0.93	1180	0.085	0.99	177	0.423	1.13	1148	0.161	0.73	1325	0.182	0.78
Total lipids in medium VLDL (mmol/L)	575	0.44	0.27	605	0.48	0.27	1180	0.46	0.27	177	0.96	0.63	1148	0.55	0.37	1325	0.60	0.43
Total lipids in small VLDL (mmol/L)	575	0.38	0.15	605	0.40	0.15	1180	0.39	0.15	177	0.69	0.28	1148	0.49	0.22	1325	0.52	0.23
Total lipids in very small VLDL (mmol/L)	575	0.32	0.07	605	0.34	0.08	1180	0.33	0.07	177	0.45	0.11	1148	0.43	0.11	1325	0.43	0.11
Total lipids in IDL (mmol/L)	575	0.80	0.18	605	0.83	0.18	1180	0.82	0.18	177	0.98	0.26	1148	1.00	0.24	1325	1.00	0.24
Total lipids in large LDL (mmol/L)	575	0.92	0.22	605	0.94	0.23	1180	0.93	0.22	177	1.16	0.33	1148	1.15	0.30	1325	1.16	0.30
Total lipids in medium LDL (mmol/L)	575	0.51	0.14	605	0.52	0.14	1180	0.52	0.14	177	0.68	0.21	1148	0.66	0.18	1325	0.66	0.19
Total lipids in small LDL (mmol/L)	575	0.34	0.08	605	0.34	0.09	1180	0.34	0.08	177	0.44	0.14	1148	0.42	0.11	1325	0.43	0.12
Total lipids in very large HDL (mmol/L)	575	0.48	0.20	605	0.49	0.18	1180	0.49	0.19	177	0.32	0.19	1148	0.50	0.23	1325	0.47	0.23
Total lipids in large HDL (mmol/L)	575	0.87	0.29	605	0.86	0.27	1180	0.87	0.28	177	0.51	0.34	1148	0.90	0.38	1325	0.85	0.40
Total lipids in medium HDL (mmol/L)	575	0.92	0.13	605	0.87	0.13	1180	0.89	0.13	177	0.83	0.24	1148	0.97	0.18	1325	0.95	0.19
Total lipids in small HDL (mmol/L)	575	1.04	0.10	605	1.00	0.11	1180	1.02	0.11	177	1.06	0.25	1148	1.08	0.14	1325	1.08	0.16
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	575	37.06	1.63	605	37.24	1.56	1180	37.15	1.59	177	38.53	1.74	1148	36.94	1.60	1325	37.15	1.70
Mean diameter for LDL particles (nm)	575	23.59	0.10	605	23.63	0.11	1180	23.61	0.11	177	23.49	0.09	1148	23.57	0.10	1325	23.56	0.10
Mean diameter for HDL particles (nm)	575	10.08	0.23	605	10.10	0.22	1180	10.09	0.23	177	9.80	0.24	1148	10.07	0.26	1325	10.03	0.28
Cholesterol																		
Serum total cholesterol (mmol/L)	575	3.58	0.62	605	3.60	0.64	1180	3.59	0.63	177	4.16	0.88	1148	4.23	0.83	1325	4.22	0.83
Total cholesterol in VLDL (mmol/L)	575	0.44	0.19	605	0.47	0.19	1180	0.46	0.19	177	0.83	0.40	1148	0.59	0.26	1325	0.62	0.30
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	575	0.94	0.27	605	0.99	0.27	1180	0.97	0.27	177	1.45	0.47	1148	1.22	0.38	1325	1.25	0.40
Total cholesterol in LDL (mmol/L)	575	1.13	0.33	605	1.15	0.34	1180	1.14	0.33	177	1.49	0.50	1148	1.46	0.44	1325	1.46	0.45
Total cholesterol in HDL (mmol/L)	575	1.50	0.27	605	1.45	0.27	1180	1.48	0.27	177	1.22	0.35	1148	1.56	0.36	1325	1.51	0.38
Total cholesterol in HDL2 (mmol/L)	575	1.03	0.25	605	0.99	0.25	1180	1.01	0.25	177	0.75	0.33	1148	1.07	0.34	1325	1.03	0.35
Total cholesterol in HDL3 (mmol/L)	575	0.47	0.02	605	0.47	0.02	1180	0.47	0.02	177	0.47	0.03	1148	0.48	0.03	1325	0.48	0.03
Esterified cholesterol (mmol/L)	572	2.52	0.45	604	2.52	0.46	1176	2.52	0.45	176	2.94	0.64	1147	2.98	0.59	1323	2.97	0.60
Free cholesterol (mmol/L)	572	1.06	0.18	604	1.08	0.19	1176	1.07	0.18	176	1.21	0.27	1147	1.26	0.24	1323	1.25	0.24
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	575	0.918	4.77	605	1.005	74.68	1180	0.962	10.07	177	1.686	0.96	1148	1.129	3.73	1325	1.19	2.75
Triglycerides in VLDL (mmol/L)*	575	0.582	1.07	605	0.648	1.21	1180	0.615	1.13	177	1.249	2.80	1148	0.694	1.75	1325	0.75	2.31
Triglycerides in LDL (mmol/L)	575	0.11	0.02	605	0.12	0.03	1180	0.12	0.03	177	0.15	0.04	1148	0.16	0.04	1325	0.16	0.04
Triglycerides in HDL (mmol/L)	575	0.13	0.03	605	0.14	0.03	1180	0.13	0.03	177	0.16	0.05	1148	0.15	0.04	1325	0.15	0.04
Total phosphoglycerides (mmol/L)	572	1.63	0.24	604	1.62	0.26	1176	1.63	0.25	176	1.86	0.35	1147	1.93	0.34	1323	1.92	0.34
Ratio of triglycerides to phosphoglycerides	572	0.53	0.25	604	0.57	0.28	1176	0.55	0.27	176	0.95	0.59	1147	0.58	0.28	1323	0.63	0.36
Phosphatidylcholine & other cholines (mmol/L)	572	1.69	0.24	604	1.69	0.27	1176	1.69	0.25	175	1.88	0.32	1147	1.98	0.34	1322	1.97	0.33
Sphingomyelins (mmol/L)	572	0.35	0.06	604	0.35	0.06	1176	0.35	0.06	175	0.37	0.07	1147	0.40	0.08	1322	0.39	0.08
Total cholines (mmol/L)	572	2.01	0.26	604	2.00	0.26	1176	2.00	0.26	175	2.19	0.33	1147	2.32	0.35	1322	2.30	0.35
Apolipoproteins																		
Apolipoprotein A1 (g/L)	575	1.51	0.16	605	1.48	0.15	1180	1.50	0.15	177	1.46	0.18	1148	1.59	0.20	1325	1.57	0.21
Apolipoprotein B (g/L)	575	0.68	0.13	604	0.71	0.14	1179	0.69	0.14	177	0.96	0.25	1148	0.81	0.20	1325	0.83	0.21
Ratio of apolipoprotein B to apolipoprotein AI	575	0.46	0.10	604	0.48	0.10	1179	0.47	0.10	177	0.66	0.18	1148	0.52	0.14	1325	0.54	0.15
Fatty acids																		
Total fatty acids (mmol/L)	570	9.21	1.70	604	9.37	1.73	1174	9.29	1.71	173	11.85	2.72	1145	10.92	2.39	1318	11.03	2.45
Estimated degree of unsaturation	570	1.21	0.06	604	1.20	0.06	1174	1.20	0.06	173	1.18	0.07	1145	1.21	0.07	1318	1.21	0.07
22:6, docosahexaenoic acid (mmol/L)	570	0.08	0.03	604	0.07	0.03	1174	0.08	0.03	173	0.12	0.05	1145	0.11	0.04	1318	0.11	0.04

1	18:2, linoleic acid (mmol/L)	570	2.54	0.46	604	2.59	0.46	1174	2.57	0.46	173	2.92	0.57	1145	2.88	0.58	1318	2.89	0.58	
2	Omega3 fatty acids (mmol/L)	570	0.31	0.09	604	0.30	0.08	1174	0.30	0.08	173	0.45	0.16	1145	0.40	0.12	1318	0.41	0.12	
3	Omega6 fatty acids (mmol/L)	570	3.08	0.49	604	3.09	0.49	1174	3.09	0.49	173	3.55	0.65	1145	3.51	0.63	1318	3.51	0.63	
4	Polyunsat. fatty acids (mmol/L)	570	3.39	0.56	604	3.39	0.56	1174	3.39	0.56	173	4.00	0.77	1145	3.91	0.72	1318	3.92	0.73	
5	Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	570	2.50	0.63	604	2.59	0.66	1174	2.54	0.65	173	3.52	1.06	1145	3.08	0.92	1318	3.14	0.94	
5	Saturated fatty acids (mmol/L)	570	3.33	0.64	604	3.39	0.68	1174	3.36	0.66	173	4.33	1.09	1145	3.93	0.93	1318	3.98	0.96	
6	Fatty acid ratios																			
7	Ratio of 22:6 docosaheptaenoic acid to total fatty acids (%)	570	0.84	0.24	604	0.79	0.27	1174	0.81	0.25	173	0.98	0.31	1145	1.02	0.28	1318	1.02	0.29	
8	Ratio of 18:2 linoleic acid to total fatty acids (%)	570	27.74	3.13	604	27.87	3.27	1174	27.81	3.19	173	25.04	3.57	1145	26.66	3.42	1318	26.45	3.47	
9	Ratio of omega3 fatty acids to total fatty acids (%)	570	3.33	0.58	604	3.15	0.60	1174	3.24	0.59	173	3.76	0.80	1145	3.66	0.70	1318	3.67	0.71	
9	Ratio of omega6 fatty acids to total fatty acids (%)	570	33.65	3.05	604	33.31	3.23	1174	33.48	3.13	173	30.47	3.76	1145	32.50	3.31	1318	32.25	3.42	
10	Ratio of polyunsat. fatty acids to total fatty acids (%)	570	36.98	3.24	604	36.46	3.51	1174	36.71	3.38	173	34.23	3.88	1145	36.16	3.59	1318	35.92	3.67	
11	Ratio of monounsat. fatty acids to total fatty acids (%)	570	26.91	2.57	604	27.38	2.68	1174	27.15	2.63	173	29.37	2.97	1145	27.89	2.84	1318	28.07	2.89	
11	Ratio of saturated fatty acids to total fatty acids (%)	570	36.11	1.68	604	36.17	1.80	1174	36.14	1.73	173	36.40	2.03	1145	35.95	2.03	1318	36.01	2.03	
12	Amino acids																			
13	Pyruvate (mmol/L)	574	0.10	0.02	605	0.10	0.02	1179	0.10	0.02	177	0.10	0.03	1147	0.09	0.03	1324	0.09	0.03	
14	Citrate (mmol/L)	575	0.12	0.02	604	0.13	0.02	1179	0.13	0.02	177	0.11	0.02	1148	0.11	0.02	1325	0.11	0.02	
15	Glycerol (mmol/L)	240	0.08	0.02	283	0.08	0.02	523	0.08	0.02	84	0.07	0.02	470	0.07	0.02	554	0.07	0.02	
16	Ketone bodies																			
17	Alanine (mmol/L)	575	0.39	0.06	605	0.40	0.06	1180	0.39	0.06	176	0.42	0.06	1147	0.40	0.06	1323	0.40	0.06	
18	Glutamine (mmol/L)	575	0.47	0.05	605	0.50	0.05	1180	0.49	0.05	177	0.49	0.06	1148	0.46	0.07	1325	0.46	0.07	
18	Glycine (mmol/L)	574	0.26	0.03	604	0.27	0.03	1178	0.27	0.03	176	0.24	0.03	1148	0.27	0.06	1324	0.27	0.06	
19	Histidine (mmol/L)	574	0.06	0.01	605	0.07	0.01	1179	0.07	0.01	176	0.07	0.01	1148	0.07	0.01	1324	0.07	0.01	
20	Isoleucine (mmol/L)	574	0.05	0.02	605	0.05	0.02	1179	0.05	0.02	174	0.07	0.02	1146	0.05	0.02	1320	0.06	0.02	
20	Leucine (mmol/L)	575	0.07	0.02	605	0.07	0.02	1180	0.07	0.02	177	0.10	0.03	1148	0.07	0.02	1325	0.08	0.02	
21	Valine (mmol/L)	575	0.16	0.04	604	0.16	0.04	1179	0.16	0.04	177	0.19	0.04	1147	0.16	0.04	1324	0.17	0.04	
22	Phenylalanine (mmol/L)	575	0.07	0.01	605	0.07	0.01	1180	0.07	0.01	177	0.07	0.01	1148	0.07	0.01	1325	0.07	0.01	
22	Tyrosine (mmol/L)	574	0.05	0.01	605	0.05	0.01	1179	0.05	0.01	176	0.06	0.01	1148	0.05	0.01	1324	0.05	0.01	
23	Glycolysis related																			
24	Acetate (mmol/L)*	575	0.031	0.05	605	0.03	0.04	1180	0.03	0.05	177	0.037	0.11	1146	0.033	0.09	1323	0.034	0.09	
25	Acetoacetate (mmol/L)*	575	0.025	0.27	605	0.023	0.30	1180	0.024	0.28	177	0.023	0.45	1147	0.024	0.26	1324	0.024	0.29	
26	3hydroxybutyrate (mmol/L)*	555	0.10	0.21	580	0.103	0.23	1135	0.101	0.22	170	0.104	0.16	1098	0.096	0.20	1268	0.097	0.20	
27	Inflammation																			
28	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	575	1.17	0.19	605	1.17	0.19	1180	1.17	0.19	177	1.38	0.37	1148	1.24	0.23	1325	1.26	0.26	

* geometric mean [relative SD] when skewed variable

Supplementary table 2: Differences in mean metabolite levels in adults compared to children in absolute concentration units.

Metabolic subgroup	Differences by age (Parents - Child)			
	Estimate	95% CI	p-value	Conversion factor (SD) #
Lipoprotein subclass lipids				
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.704	(0.519, 0.890)	<0.001	2.53
Total lipids in very large VLDL (mmol/L)*	0.922	(0.700, 1.145)	<0.001	3.03
Total lipids in large VLDL (mmol/L)*	0.648	(0.502, 0.795)	<0.001	1.95
Total lipids in medium VLDL (mmol/L)	0.105	(0.080, 0.129)	<0.001	0.35
Total lipids in small VLDL (mmol/L)	0.107	(0.094, 0.121)	<0.001	0.20
Total lipids in very small VLDL (mmol/L)	0.093	(0.086, 0.099)	<0.001	0.10
Total lipids in IDL (mmol/L)	0.181	(0.166, 0.196)	<0.001	0.23
Total lipids in large LDL (mmol/L)	0.229	(0.211, 0.247)	<0.001	0.29
Total lipids in medium LDL (mmol/L)	0.144	(0.132, 0.155)	<0.001	0.18
Total lipids in small LDL (mmol/L)	0.089	(0.082, 0.096)	<0.001	0.11
Total lipids in very large HDL (mmol/L)	0.012	(-0.003, 0.027)	0.128	0.22
Total lipids in large HDL (mmol/L)	0.032	(0.007, 0.057)	0.011	0.35
Total lipids in medium HDL (mmol/L)	0.076	(0.064, 0.089)	<0.001	0.17
Total lipids in small HDL (mmol/L)	0.068	(0.058, 0.078)	<0.001	0.14
Lipoprotein particle size				
Mean diameter for VLDL particles (nm)	-0.147	(-0.263, -0.031)	0.013	1.63
Mean diameter for LDL particles (nm)	-0.044	(-0.052, -0.037)	<0.001	0.11
Mean diameter for HDL particles (nm)	-0.027	(-0.045, -0.010)	0.002	0.26
Cholesterol				
Serum total cholesterol (mmol/L)	0.670	(0.619, 0.721)	<0.001	0.80
Total cholesterol in VLDL (mmol/L)	0.146	(0.129, 0.163)	<0.001	0.25
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.261	(0.237, 0.284)	<0.001	0.36
Total cholesterol in LDL (mmol/L)	0.327	(0.300, 0.354)	<0.001	0.42
Total cholesterol in HDL (mmol/L)	0.082	(0.058, 0.106)	<0.001	0.34
Total cholesterol in HDL2 (mmol/L)	0.064	(0.042, 0.086)	<0.001	0.31
Total cholesterol in HDL3 (mmol/L)	0.018	(0.016, 0.020)	<0.001	0.03
Esterified cholesterol (mmol/L)	0.474	(0.437, 0.511)	<0.001	0.58
Free cholesterol (mmol/L)	0.195	(0.180, 0.210)	<0.001	0.23
Glycerides and phospholipids				
Serum total triglycerides (mmol/L)*	0.176	(0.145, 0.206)	<0.001	0.44
Triglycerides in VLDL (mmol/L)*	0.140	(0.096, 0.183)	<0.001	0.62
Triglycerides in LDL (mmol/L)	0.040	(0.038, 0.043)	<0.001	0.04
Triglycerides in HDL (mmol/L)	0.020	(0.017, 0.022)	<0.001	0.04
Total phosphoglycerides (mmol/L)	0.311	(0.290, 0.332)	<0.001	0.34
Ratio of triglycerides to phosphoglycerides	0.049	(0.027, 0.071)	<0.001	0.30
Phosphatidylcholine & other cholines (mmol/L)	0.295	(0.274, 0.316)	<0.001	0.33
Sphingomyelins (mmol/L)	0.052	(0.047, 0.057)	<0.001	0.08
Total cholines (mmol/L)	0.323	(0.302, 0.345)	<0.001	0.35
Apolipoproteins				
Apolipoprotein A1 (g/L)	0.099	(0.086, 0.112)	<0.001	0.19
Apolipoprotein B (g/L)	0.125	(0.113, 0.137)	<0.001	0.18
Ratio of apolipoprotein B to apolipoprotein A1	0.055	(0.046, 0.064)	<0.001	0.13
Fatty acids				
Total fatty acids (mmol/L)	1.738	(1.592, 1.885)	<0.001	2.24
Estimated degree of unsaturation	0.005	(0.000, 0.009)	0.030	0.06
22:6, docosahexaenoic acid (mmol/L)	0.037	(0.035, 0.040)	<0.001	0.04
18:2, linoleic acid (mmol/L)	0.347	(0.310, 0.384)	<0.001	0.54
Omega3 fatty acids (mmol/L)	0.105	(0.098, 0.113)	<0.001	0.12
Omega6 fatty acids (mmol/L)	0.453	(0.414, 0.492)	<0.001	0.60
Polyunsat. fatty acids (mmol/L)	0.558	(0.513, 0.603)	<0.001	0.70
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.568	(0.512, 0.625)	<0.001	0.85
Saturated fatty acids (mmol/L)	0.612	(0.554, 0.669)	<0.001	0.86
Fatty acid ratios				
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.209	(0.190, 0.228)	<0.001	0.29
Ratio of 18:2 linoleic acid to total fatty acids (%)	-1.079	(-1.331, -0.827)	<0.001	3.39
Ratio of omega3 fatty acids to total fatty acids (%)	0.446	(0.403, 0.490)	<0.001	0.69
Ratio of omega6 fatty acids to total fatty acids (%)	-0.992	(-1.227, -0.757)	<0.001	3.30
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.546	(-0.794, -0.298)	<0.001	3.51
Ratio of monounsat. fatty acids to total fatty acids (%)	0.741	(0.547, 0.934)	<0.001	2.80

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2	Ratio of saturated fatty acids to total fatty acids (%)	-0.195	(-0.328, -0.062)	0.004	1.86
3	Glycolysis related				
4	Pyruvate (mmol/L)	-0.007	(-0.009, -0.005)	<0.001	0.03
5	Citrate (mmol/L)	-0.017	(-0.018, -0.016)	<0.001	0.02
6	Glycerol (mmol/L)	-0.011	(-0.015, -0.008)	<0.001	0.02
7	Amino acids				
8	Alanine (mmol/L)	0.013	(0.009, 0.017)	<0.001	0.06
9	Glutamine (mmol/L)	-0.023	(-0.027, -0.019)	<0.001	0.06
10	Glycine (mmol/L)	0.007	(0.003, 0.010)	<0.001	0.05
11	Histidine (mmol/L)	0.001	(0.000, 0.002)	0.005	0.01
12	Isoleucine (mmol/L)	0.003	(0.002, 0.004)	<0.001	0.02
13	Leucine (mmol/L)	0.004	(0.003, 0.006)	<0.001	0.02
14	Valine (mmol/L)	0.009	(0.006, 0.011)	<0.001	0.04
15	Phenylalanine (mmol/L)	0.005	(0.005, 0.006)	<0.001	0.01
16	Tyrosine (mmol/L)	0.001	(-0.000, 0.002)	0.100	0.01
17	Ketone bodies				
18	Acetate (mmol/L)*	0.101	(0.084, 0.117)	<0.001	0.23
19	Acetoacetate (mmol/L)*	-0.004	(-0.086, 0.078)	0.922	1.02
20	3hydroxybutyrate (mmol/L)*	-0.064	(-0.100, -0.028)	0.001	0.49
21	Inflammation				
22	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.062	(0.047, 0.078)	<0.001	0.22

*metabolite has been log transformed

Associations in Figure 2 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 3: Differences in mean metabolite levels in girls compared to boys in absolute concentration units.

Metabolic subgroup	Differences for children (Girls - Boys)			
	Estimate	95% CI	p-value	Conversion factor (SD) #
Lipoprotein subclass lipids				
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.737	(0.414, 1.059)	<0.001	2.84
Total lipids in very large VLDL (mmol/L)*	0.744	(0.355, 1.134)	<0.001	3.43
Total lipids in large VLDL (mmol/L)*	0.663	(0.390, 0.936)	<0.001	2.41
Total lipids in medium VLDL (mmol/L)	0.049	(0.018, 0.080)	0.002	0.27
Total lipids in small VLDL (mmol/L)	0.032	(0.015, 0.048)	<0.001	0.15
Total lipids in very small VLDL (mmol/L)	0.018	(0.010, 0.027)	<0.001	0.07
Total lipids in IDL (mmol/L)	0.025	(0.004, 0.046)	0.017	0.18
Total lipids in large LDL (mmol/L)	0.020	(-0.006, 0.046)	0.132	0.23
Total lipids in medium LDL (mmol/L)	0.008	(-0.008, 0.024)	0.338	0.14
Total lipids in small LDL (mmol/L)	0.001	(-0.008, 0.011)	0.788	0.09
Total lipids in very large HDL (mmol/L)	-0.002	(-0.023, 0.020)	0.882	0.19
Total lipids in large HDL (mmol/L)	-0.033	(-0.066, -0.001)	0.044	0.28
Total lipids in medium HDL (mmol/L)	-0.045	(-0.059, -0.030)	<0.001	0.13
Total lipids in small HDL (mmol/L)	-0.035	(-0.048, -0.022)	<0.001	0.12
Lipoprotein particle size				
Mean diameter for VLDL particles (nm)	0.215	(0.035, 0.395)	0.019	1.58
Mean diameter for LDL particles (nm)	0.035	(0.023, 0.047)	<0.001	0.11
Mean diameter for HDL particles (nm)	0.003	(-0.023, 0.028)	0.847	0.23
Cholesterol				
Serum total cholesterol (mmol/L)	0.007	(-0.066, 0.079)	0.857	0.63
Total cholesterol in VLDL (mmol/L)	0.040	(0.019, 0.061)	<0.001	0.18
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.053	(0.023, 0.083)	0.001	0.27
Total cholesterol in LDL (mmol/L)	0.016	(-0.023, 0.054)	0.427	0.34
Total cholesterol in HDL (mmol/L)	-0.062	(-0.093, -0.031)	<0.001	0.27
Total cholesterol in HDL2 (mmol/L)	-0.059	(-0.087, -0.030)	<0.001	0.25
Total cholesterol in HDL3 (mmol/L)	-0.003	(-0.006, -0.001)	0.013	0.02
Esterified cholesterol (mmol/L)	-0.008	(-0.060, 0.044)	0.755	0.45
Free cholesterol (mmol/L)	0.013	(-0.008, 0.034)	0.211	0.18
Glycerides and phospholipids				
Serum total triglycerides (mmol/L)*	0.101	(0.056, 0.145)	<0.001	0.39
Triglycerides in VLDL (mmol/L)*	0.125	(0.062, 0.187)	<0.001	0.55
Triglycerides in LDL (mmol/L)	0.009	(0.006, 0.012)	<0.001	0.03
Triglycerides in HDL (mmol/L)	0.007	(0.004, 0.011)	<0.001	0.03
Total phosphoglycerides (mmol/L)	-0.018	(-0.047, 0.010)	0.206	0.25
Ratio of triglycerides to phosphoglycerides	0.051	(0.020, 0.083)	0.001	0.27
Phosphatidylcholine & other cholines (mmol/L)	-0.011	(-0.040, 0.018)	0.447	0.25
Sphingomyelins (mmol/L)	-0.001	(-0.009, 0.006)	0.706	0.06
Total cholines (mmol/L)	-0.017	(-0.046, 0.013)	0.268	0.26
Apolipoproteins				
Apolipoprotein A1 (g/L)	-0.030	(-0.048, -0.013)	0.001	0.16
Apolipoprotein B (g/L)	0.027	(0.012, 0.042)	0.001	0.13
Ratio of apolipoprotein B to apolipoprotein A1	0.027	(0.016, 0.038)	<0.001	0.10
Fatty acids				
Total fatty acids (mmol/L)	0.200	(0.011, 0.389)	0.038	1.65
Estimated degree of unsaturation	-0.016	(-0.022, -0.009)	<0.001	0.06
22:6, docosahexaenoic acid (mmol/L)	-0.004	(-0.007, -0.000)	0.033	0.03
18:2, linoleic acid (mmol/L)	0.068	(0.015, 0.120)	0.011	0.46
Omega3 fatty acids (mmol/L)	-0.008	(-0.018, 0.001)	0.083	0.08
Omega6 fatty acids (mmol/L)	0.031	(-0.024, 0.087)	0.271	0.48
Polyunsat. fatty acids (mmol/L)	0.023	(-0.041, 0.086)	0.483	0.55
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.105	(0.034, 0.176)	0.004	0.62
Saturated fatty acids (mmol/L)	0.073	(-0.000, 0.146)	0.052	0.64
Fatty acid ratios				
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	-0.049	(-0.078, -0.019)	0.001	0.25
Ratio of 18:2 linoleic acid to total fatty acids (%)	0.197	(-0.173, 0.567)	0.297	3.23
Ratio of omega3 fatty acids to total fatty acids (%)	-0.145	(-0.213, -0.078)	<0.001	0.59
Ratio of omega6 fatty acids to total fatty acids (%)	-0.302	(-0.658, 0.054)	0.096	3.11
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.447	(-0.830, -0.065)	0.022	3.35
Ratio of monounsat. fatty acids to total fatty acids (%)	0.499	(0.202, 0.796)	0.001	2.61
Ratio of saturated fatty acids to total fatty acids (%)	-0.051	(-0.251, 0.148)	0.614	1.74
Glycolysis related				
Pyruvate (mmol/L)	-0.001	(-0.004, 0.002)	0.524	0.02
Citrate (mmol/L)	0.007	(0.005, 0.009)	<0.001	0.02
Glycerol (mmol/L)	0.006	(0.002, 0.010)	0.004	0.02
Amino acids				
Alanine (mmol/L)	0.011	(0.004, 0.017)	0.002	0.06
Glutamine (mmol/L)	0.023	(0.018, 0.029)	<0.001	0.05
Glycine (mmol/L)	0.010	(0.006, 0.014)	<0.001	0.03
Histidine (mmol/L)	0.001	(-0.000, 0.002)	0.075	0.01
Isoleucine (mmol/L)	-0.000	(-0.003, 0.002)	0.637	0.02
Leucine (mmol/L)	-0.002	(-0.005, -0.000)	0.022	0.02
Valine (mmol/L)	-0.007	(-0.011, -0.003)	0.001	0.04
Phenylalanine (mmol/L)	-0.002	(-0.003, -0.001)	0.003	0.01
Tyrosine (mmol/L)	0.000	(-0.001, 0.002)	0.582	0.01
Ketone bodies				
Acetate (mmol/L)*	-0.030	(-0.048, -0.011)	0.002	0.17
Acetoacetate (mmol/L)*	-0.058	(-0.172, 0.055)	0.313	0.99

1	3hydroxybutyrate (mmol/L)*	0.041	(-0.019, 0.101)	0.178	0.51
2					
3	Inflammation				
4	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.017	(-0.004, 0.038)	0.104	0.18

5 *metabolite has been log transformed

6 # Associations for children in Figure 3 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure in children
7 can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

8 Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

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Supplementary table 4: Differences in mean metabolite levels in female compared to male adults in absolute concentration units.

Metabolic subgroup	Differences for adults (Female - Male)			
	Estimate	95% CI	p-value	Conversion factor (SD) #
Lipoprotein subclass lipids				
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	-0.930	(-1.271, -0.589)	<0.001	2.17
Total lipids in very large VLDL (mmol/L)*	-1.217	(-1.617, -0.818)	<0.001	2.55
Total lipids in large VLDL (mmol/L)*	-0.900	(-1.107, -0.693)	<0.001	1.34
Total lipids in medium VLDL (mmol/L)	-0.325	(-0.385, -0.264)	<0.001	0.40
Total lipids in small VLDL (mmol/L)	-0.167	(-0.201, -0.133)	<0.001	0.22
Total lipids in very small VLDL (mmol/L)	-0.022	(-0.039, -0.005)	0.013	0.11
Total lipids in IDL (mmol/L)	0.014	(-0.023, 0.051)	0.465	0.24
Total lipids in large LDL (mmol/L)	-0.011	(-0.058, 0.035)	0.634	0.29
Total lipids in medium LDL (mmol/L)	-0.028	(-0.057, 0.001)	0.058	0.18
Total lipids in small LDL (mmol/L)	-0.017	(-0.035, 0.001)	0.061	0.11
Total lipids in very large HDL (mmol/L)	0.195	(0.158, 0.231)	<0.001	0.24
Total lipids in large HDL (mmol/L)	0.395	(0.334, 0.455)	<0.001	0.41
Total lipids in medium HDL (mmol/L)	0.129	(0.101, 0.158)	<0.001	0.18
Total lipids in small HDL (mmol/L)	0.002	(-0.021, 0.025)	0.850	0.14
Lipoprotein particle size				
Mean diameter for VLDL particles (nm)	-1.414	(-1.669, -1.159)	<0.001	1.68
Mean diameter for LDL particles (nm)	0.081	(0.066, 0.096)	<0.001	0.10
Mean diameter for HDL particles (nm)	0.278	(0.236, 0.320)	<0.001	0.28
Cholesterol				
Serum total cholesterol (mmol/L)	0.112	(-0.018, 0.241)	0.091	0.82
Total cholesterol in VLDL (mmol/L)	-0.187	(-0.230, -0.145)	<0.001	0.28
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	-0.184	(-0.244, -0.124)	<0.001	0.38
Total cholesterol in LDL (mmol/L)	-0.048	(-0.117, 0.021)	0.175	0.44
Total cholesterol in HDL (mmol/L)	0.344	(0.286, 0.401)	<0.001	0.38
Total cholesterol in HDL2 (mmol/L)	0.324	(0.271, 0.378)	<0.001	0.35
Total cholesterol in HDL3 (mmol/L)	0.019	(0.014, 0.025)	<0.001	0.03
Esterified cholesterol (mmol/L)	0.070	(-0.023, 0.163)	0.139	0.59
Free cholesterol (mmol/L)	0.046	(0.009, 0.084)	0.016	0.24
Glycerides and phospholipids				
Serum total triglycerides (mmol/L)*	-0.344	(-0.416, -0.273)	<0.001	0.47
Triglycerides in VLDL (mmol/L)*	-0.530	(-0.630, -0.429)	<0.001	0.66
Triglycerides in LDL (mmol/L)	0.008	(0.001, 0.014)	0.033	0.04
Triglycerides in HDL (mmol/L)	-0.004	(-0.010, 0.002)	0.228	0.04
Total phosphoglycerides (mmol/L)	0.106	(0.052, 0.159)	<0.001	0.34
Ratio of triglycerides to phosphoglycerides	-0.289	(-0.337, -0.241)	<0.001	0.32
Phosphatidylcholine & other cholines (mmol/L)	0.138	(0.086, 0.190)	<0.001	0.33
Sphingomyelins (mmol/L)	0.032	(0.020, 0.045)	<0.001	0.08
Total cholines (mmol/L)	0.170	(0.115, 0.224)	<0.001	0.35
Apolipoproteins				
Apolipoprotein A1 (g/L)	0.146	(0.114, 0.178)	<0.001	0.21
Apolipoprotein B (g/L)	-0.115	(-0.146, -0.084)	<0.001	0.20
Ratio of apolipoprotein B to apolipoprotein AI	-0.126	(-0.148, -0.105)	<0.001	0.14
Fatty acids				
Total fatty acids (mmol/L)	-0.711	(-1.091, -0.330)	<0.001	2.39
Estimated degree of unsaturation	0.031	(0.021, 0.042)	<0.001	0.07
22:6, docosahexaenoic acid (mmol/L)	-0.002	(-0.008, 0.005)	0.622	0.04
18:2, linoleic acid (mmol/L)	0.004	(-0.087, 0.094)	0.934	0.57
Omega3 fatty acids (mmol/L)	-0.035	(-0.054, -0.016)	<0.001	0.12
Omega6 fatty acids (mmol/L)	0.004	(-0.094, 0.102)	0.936	0.61
Polyunsat. fatty acids (mmol/L)	-0.031	(-0.144, 0.082)	0.592	0.71
Monounsatur. fatty acids; 16:1, 18:1 (mmol/L)	-0.372	(-0.520, -0.225)	<0.001	0.93
Saturated fatty acids (mmol/L)	-0.307	(-0.455, -0.159)	<0.001	0.93
Fatty acid ratios				
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.064	(0.018, 0.110)	0.006	0.29
Ratio of 18:2 linoleic acid to total fatty acids (%)	1.527	(0.984, 2.070)	<0.001	3.43
Ratio of omega3 fatty acids to total fatty acids (%)	-0.038	(-0.152, 0.075)	0.508	0.71
Ratio of omega6 fatty acids to total fatty acids (%)	1.882	(1.351, 2.412)	<0.001	3.38
Ratio of polyunsatur. fatty acids to total fatty acids (%)	1.843	(1.272, 2.414)	<0.001	3.62
Ratio of monounsatur. fatty acids to total fatty acids (%)	-1.456	(-1.914, -0.998)	<0.001	2.90
Ratio of saturated fatty acids to total fatty acids (%)	-0.387	(-0.700, -0.074)	0.015	1.96

Glycolysis related

Pyruvate (mmol/L)	-0.007	(-0.013, -0.002)	0.004	0.03
Citrate (mmol/L)	-0.001	(-0.004, 0.001)	0.335	0.02
Glycerol (mmol/L)	-0.003	(-0.008, 0.002)	0.279	0.02

Amino acids

Alanine (mmol/L)	-0.020	(-0.030, -0.011)	<0.001	0.06
Glutamine (mmol/L)	-0.038	(-0.048, -0.028)	<0.001	0.07
Glycine (mmol/L)	0.029	(0.020, 0.038)	<0.001	0.06
Histidine (mmol/L)	-0.001	(-0.003, 0.000)	0.116	0.01
Isoleucine (mmol/L)	-0.016	(-0.019, -0.013)	<0.001	0.02
Leucine (mmol/L)	-0.019	(-0.022, -0.016)	<0.001	0.02
Valine (mmol/L)	-0.029	(-0.035, -0.022)	<0.001	0.04
Phenylalanine (mmol/L)	-0.000	(-0.002, 0.001)	0.576	0.01
Tyrosine (mmol/L)	-0.005	(-0.007, -0.003)	<0.001	0.01

Ketone bodies

Acetate (mmol/L)*	-0.076	(-0.119, -0.033)	0.001	0.27
Acetoacetate (mmol/L)*	0.018	(-0.148, 0.184)	0.828	1.05
3hydroxybutyrate (mmol/L)*	-0.087	(-0.163, -0.011)	0.025	0.47

Inflammation

Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	-0.098	(-0.136, -0.061)	<0.001	0.24
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*metabolite has been log transformed

Associations for parents in Figure 3 are presented in SD-units.

The conversion factor provided (unweighted standard deviation of each metabolite measure in adults/parents)

can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 5: Mother-child concordance; correlations and partial correlations between mothers and their sons, daughters and all children.

Metabolic subgroup	Mother																	
	Boys				Girls				All Children									
	n	CC	95% CI	n	PCC#	95% CI	n	CC	95% CI	n	CC	95% CI	n	PCC#	95% CI			
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	469	0.22	0.13 - 0.30	468	0.22	0.13 - 0.31	518	0.24	0.16 - 0.32	517	0.23	0.14 - 0.31	987	0.23	0.17 - 0.29	985	0.23	0.17 - 0.29
Total lipids in very large VLDL (mmol/L)*	469	0.25	0.16 - 0.33	468	0.25	0.16 - 0.33	518	0.22	0.14 - 0.30	517	0.21	0.13 - 0.29	987	0.24	0.18 - 0.29	985	0.23	0.17 - 0.29
Total lipids in large VLDL (mmol/L)*	469	0.22	0.13 - 0.30	468	0.22	0.13 - 0.30	518	0.23	0.14 - 0.31	517	0.21	0.13 - 0.29	987	0.22	0.16 - 0.28	985	0.21	0.15 - 0.27
Total lipids in medium VLDL (mmol/L)	469	0.26	0.17 - 0.34	468	0.25	0.17 - 0.34	518	0.29	0.21 - 0.37	517	0.28	0.20 - 0.36	987	0.28	0.22 - 0.34	985	0.27	0.22 - 0.33
Total lipids in small VLDL (mmol/L)	469	0.26	0.18 - 0.35	468	0.26	0.17 - 0.34	518	0.30	0.21 - 0.37	517	0.29	0.21 - 0.36	987	0.29	0.23 - 0.34	985	0.28	0.22 - 0.33
Total lipids in very small VLDL (mmol/L)	469	0.22	0.14 - 0.31	468	0.22	0.13 - 0.30	518	0.26	0.17 - 0.34	517	0.26	0.18 - 0.34	987	0.25	0.19 - 0.30	985	0.24	0.18 - 0.30
Total lipids in IDL (mmol/L)	469	0.27	0.18 - 0.35	468	0.26	0.17 - 0.34	518	0.29	0.21 - 0.37	517	0.30	0.22 - 0.37	987	0.29	0.23 - 0.34	985	0.28	0.22 - 0.34
Total lipids in large LDL (mmol/L)	469	0.28	0.19 - 0.36	468	0.27	0.18 - 0.35	518	0.30	0.22 - 0.37	517	0.30	0.22 - 0.38	987	0.29	0.23 - 0.35	985	0.29	0.23 - 0.34
Total lipids in medium LDL (mmol/L)	469	0.28	0.20 - 0.36	468	0.27	0.19 - 0.35	518	0.29	0.21 - 0.37	517	0.29	0.21 - 0.37	987	0.29	0.23 - 0.35	985	0.29	0.23 - 0.34
Total lipids in small LDL (mmol/L)	469	0.28	0.19 - 0.36	468	0.27	0.18 - 0.35	518	0.29	0.21 - 0.37	517	0.29	0.21 - 0.37	987	0.28	0.22 - 0.34	985	0.28	0.22 - 0.34
Total lipids in very large HDL (mmol/L)	469	0.30	0.22 - 0.38	468	0.29	0.21 - 0.38	518	0.32	0.24 - 0.39	517	0.32	0.24 - 0.39	987	0.31	0.25 - 0.36	985	0.30	0.25 - 0.36
Total lipids in large HDL (mmol/L)	469	0.31	0.23 - 0.39	468	0.31	0.22 - 0.39	518	0.28	0.20 - 0.36	517	0.28	0.20 - 0.36	987	0.30	0.24 - 0.35	985	0.29	0.23 - 0.35
Total lipids in medium HDL (mmol/L)	469	0.22	0.13 - 0.30	468	0.21	0.12 - 0.30	518	0.12	0.03 - 0.20	517	0.13	0.04 - 0.21	987	0.17	0.11 - 0.23	985	0.17	0.11 - 0.23
Total lipids in small HDL (mmol/L)	469	0.23	0.14 - 0.31	468	0.22	0.13 - 0.31	518	0.20	0.12 - 0.29	517	0.20	0.12 - 0.29	987	0.21	0.15 - 0.27	985	0.21	0.15 - 0.27
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	469	0.30	0.22 - 0.38	468	0.30	0.22 - 0.38	518	0.27	0.19 - 0.35	517	0.26	0.18 - 0.34	987	0.29	0.23 - 0.35	985	0.28	0.22 - 0.34
Mean diameter for LDL particles (nm)	469	0.22	0.13 - 0.31	468	0.22	0.13 - 0.31	518	0.30	0.22 - 0.38	517	0.31	0.23 - 0.38	987	0.26	0.20 - 0.31	985	0.26	0.20 - 0.32
Mean diameter for HDL particles (nm)	469	0.32	0.23 - 0.40	468	0.31	0.23 - 0.39	518	0.33	0.26 - 0.41	517	0.33	0.25 - 0.41	987	0.33	0.27 - 0.38	985	0.32	0.26 - 0.38
Cholesterol																		
Serum total cholesterol (mmol/L)	469	0.27	0.19 - 0.35	468	0.27	0.18 - 0.35	518	0.32	0.24 - 0.39	517	0.32	0.24 - 0.40	987	0.30	0.24 - 0.35	985	0.29	0.24 - 0.35
Total cholesterol in VLDL (mmol/L)	469	0.25	0.17 - 0.34	468	0.25	0.16 - 0.33	518	0.29	0.21 - 0.36	517	0.28	0.20 - 0.36	987	0.28	0.22 - 0.33	985	0.27	0.21 - 0.33
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	469	0.25	0.17 - 0.34	468	0.25	0.16 - 0.33	518	0.29	0.21 - 0.37	517	0.29	0.21 - 0.37	987	0.28	0.22 - 0.34	985	0.28	0.22 - 0.33
Total cholesterol in LDL (mmol/L)	469	0.28	0.20 - 0.36	468	0.27	0.19 - 0.35	518	0.29	0.21 - 0.37	517	0.29	0.21 - 0.37	987	0.29	0.23 - 0.34	985	0.29	0.23 - 0.34
Total cholesterol in HDL (mmol/L)	469	0.30	0.22 - 0.38	468	0.30	0.21 - 0.38	518	0.25	0.16 - 0.33	517	0.24	0.16 - 0.32	987	0.28	0.22 - 0.33	985	0.27	0.21 - 0.32
Total cholesterol in HDL2 (mmol/L)	469	0.30	0.22 - 0.38	468	0.30	0.21 - 0.38	518	0.25	0.16 - 0.32	517	0.24	0.16 - 0.32	987	0.28	0.22 - 0.33	985	0.27	0.21 - 0.33
Total cholesterol in HDL3 (mmol/L)	469	0.25	0.16 - 0.33	468	0.24	0.16 - 0.33	518	0.25	0.16 - 0.33	517	0.24	0.16 - 0.32	987	0.25	0.19 - 0.31	985	0.24	0.18 - 0.30
Esterified cholesterol (mmol/L)	465	0.28	0.19 - 0.36	464	0.27	0.19 - 0.35	518	0.32	0.24 - 0.39	517	0.32	0.24 - 0.39	983	0.30	0.24 - 0.35	981	0.30	0.24 - 0.35
Free cholesterol (mmol/L)	465	0.26	0.18 - 0.34	464	0.26	0.17 - 0.34	518	0.32	0.24 - 0.40	517	0.33	0.25 - 0.40	983	0.30	0.24 - 0.35	981	0.29	0.24 - 0.35
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	469	0.28	0.20 - 0.36	468	0.28	0.19 - 0.36	518	0.29	0.21 - 0.37	517	0.28	0.20 - 0.36	987	0.29	0.23 - 0.35	985	0.28	0.23 - 0.34
Triglycerides in VLDL (mmol/L)*	469	0.30	0.22 - 0.38	468	0.29	0.21 - 0.38	518	0.28	0.20 - 0.36	517	0.27	0.19 - 0.35	987	0.29	0.23 - 0.35	985	0.28	0.22 - 0.34
Triglycerides in LDL (mmol/L)	469	0.19	0.10 - 0.28	468	0.19	0.10 - 0.28	518	0.26	0.18 - 0.34	517	0.26	0.18 - 0.34	987	0.23	0.17 - 0.29	985	0.23	0.17 - 0.29
Triglycerides in HDL (mmol/L)	469	0.21	0.12 - 0.30	468	0.21	0.13 - 0.30	518	0.26	0.18 - 0.34	517	0.25	0.17 - 0.33	987	0.24	0.18 - 0.30	985	0.24	0.18 - 0.29
Total phosphoglycerides (mmol/L)	465	0.26	0.17 - 0.34	464	0.26	0.17 - 0.34	518	0.26	0.17 - 0.34	517	0.26	0.18 - 0.34	983	0.26	0.20 - 0.32	981	0.26	0.20 - 0.32
Ratio of triglycerides to phosphoglycerides	465	0.23	0.15 - 0.32	464	0.23	0.14 - 0.32	518	0.26	0.18 - 0.34	517	0.25	0.17 - 0.33	983	0.25	0.20 - 0.31	981	0.25	0.19 - 0.30
Phosphatidylcholine & other cholines (mmol/L)	465	0.27	0.18 - 0.35	464	0.27	0.18 - 0.35	518	0.25	0.17 - 0.33	517	0.25	0.17 - 0.33	983	0.26	0.20 - 0.32	981	0.26	0.20 - 0.32
Sphingomyelin (mmol/L)	465	0.23	0.15 - 0.32	464	0.23	0.15 - 0.32	518	0.29	0.21 - 0.37	517	0.29	0.21 - 0.37	983	0.27	0.21 - 0.32	981	0.26	0.20 - 0.32
Total cholines (mmol/L)	465	0.27	0.18 - 0.35	464	0.27	0.18 - 0.35	518	0.28	0.20 - 0.35	517	0.28	0.20 - 0.36	983	0.27	0.21 - 0.33	981	0.27	0.21 - 0.33
Apolipoproteins																		
Apolipoprotein A1 (g/L)	469	0.28	0.20 - 0.36	468	0.28	0.19 - 0.36	518	0.26	0.18 - 0.34	517	0.26	0.18 - 0.34	987	0.27	0.21 - 0.33	985	0.27	0.21 - 0.32
Apolipoprotein B (g/L)	469	0.26	0.18 - 0.35	468	0.26	0.17 - 0.34	517	0.30	0.22 - 0.38	516	0.30	0.22 - 0.38	986	0.29	0.23 - 0.35	984	0.29	0.23 - 0.35
Ratio of apolipoprotein B to apolipoprotein AI	469	0.28	0.20 - 0.36	468	0.27	0.19 - 0.35	517	0.25	0.17 - 0.33	516	0.24	0.16 - 0.32	986	0.27	0.21 - 0.33	984	0.26	0.20 - 0.32
Fatty acids																		
Total fatty acids (mmol/L)	462	0.22	0.13 - 0.30	461	0.23	0.14 - 0.31	517	0.31	0.23 - 0.39	516	0.31	0.23 - 0.39	979	0.27	0.22 - 0.33	977	0.28	0.22 - 0.33
Estimated degree of unsaturation	462	0.30	0.22 - 0.38	461	0.30	0.21 - 0.38	517	0.27	0.19 - 0.35	516	0.25	0.17 - 0.33	979	0.29	0.23 - 0.34	977	0.28	0.22 - 0.33
22:6, docosahexaenoic acid (mmol/L)	462	0.19	0.10 - 0.28	461	0.19	0.10 - 0.28	517	0.26	0.18 - 0.34	516	0.27	0.19 - 0.35	979	0.23	0.17 - 0.29	977	0.23	0.17 - 0.29
18:2, linoleic acid (mmol/L)	462	0.22	0.13 - 0.31	461	0.23	0.15 - 0.32	517	0.27	0.19 - 0.35	516	0.27	0.19 - 0.35	979	0.25	0.19 - 0.31	977	0.26	0.20 - 0.32
Omega3 fatty acids (mmol/L)	462	0.20	0.11 - 0.29	461	0.21	0.12 - 0.30	517	0.34	0.27 - 0.42	516	0.34	0.26 - 0.41	979	0.27	0.21 - 0.33	977	0.27	0.21 - 0.33
Omega6 fatty acids (mmol/L)	462	0.23	0.14 - 0.31	461	0.24	0.16 - 0.33	517	0.31	0.23 - 0.38	516	0.31	0.23 - 0.38	979	0.27	0.21 - 0.33	977	0.28	0.22 - 0.33
Polysat. fatty acids (mmol/L)	462	0.23	0.14 - 0.31	461	0.24	0.15 - 0.33	517	0.32	0.24 - 0.39	516	0.32	0.24 - 0.39	979	0.28	0.22 - 0.33	977	0.28	0.22 - 0.34
Monosat. fatty acids; 16:1, 18:1 (mmol/L)	462	0.24	0.15 - 0.33	461	0.24	0.16 - 0.33	517	0.33	0.25 - 0.40	516	0.32	0.24 - 0.39	979	0.29	0.24 - 0.35	977	0.29	0.23 - 0.35
Saturated fatty acids (mmol/L)	462	0.21	0.12 - 0.29	461	0.21	0.12 - 0.30	517	0.29	0.21 - 0.37	516	0.29	0.21 - 0.37	979	0.26	0.20 - 0.32	977	0.26	0.20 - 0.32
Fatty acid ratios																		
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	462	0.31	0.22 - 0.39	461	0.29	0.21 - 0.38	517	0.35	0.27 - 0.42	516	0.33	0.25 - 0.41	979	0.33	0.27 - 0.39	977	0.31	0.25 - 0.37
Ratio of 18:2 linoleic acid to total fatty acids (%)	462	0.13	0.04 - 0.22	461	0.13	0.03 - 0.21	517	0.20	0.11 - 0.28	516	0.19	0.10 - 0.27	979	0.17	0.11 - 0.23	977	0.16	0.10 - 0.22
Ratio of omega3 fatty acids to total fatty acids (%)	462	0.32	0.24 - 0.40	461	0.33	0.24 - 0.40	517	0.40	0.33 - 0.47	516	0.38	0.30 - 0.45	979	0.36	0.31 - 0.41	977	0.35	0.29 - 0.40
Ratio of omega6 fatty acids to total fatty acids (%)	462	0.23	0.15 - 0.32	461	0.23	0.14 - 0.32	517	0.25	0.17 - 0.33	516	0.24	0.15 - 0.32	979	0.24	0.18 - 0.30	977	0.24	0.18 - 0.30
Ratio of polysat. fatty acids to total fatty acids (%)	462	0.27	0.19 - 0.36	461	0.27	0.18 - 0.35	517	0.28	0.20 - 0.36	516								

Supplementary table 6: Parent-child concordance; correlation and partial correlations between all parents and their sons, daughters and all children.

Metabolic subgroup	All Parents																	
	Boys					Girls					All Children							
	n	CC	95% CI	n	PCC#	95% CI	n	CC	95% CI	n	PCC#	95% CI	n	CC	95% CI			
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	547	0.22	0.14 - 0.30	544	0.22	0.14 - 0.30	586	0.20	0.12 - 0.28	585	0.19	0.11 - 0.27	1133	0.22	0.16 - 0.27	1129	0.21	0.16 - 0.27
Total lipids in very large VLDL (mmol/L)*	547	0.23	0.15 - 0.31	544	0.23	0.15 - 0.31	586	0.17	0.10 - 0.25	585	0.17	0.09 - 0.24	1133	0.21	0.15 - 0.26	1129	0.20	0.15 - 0.26
Total lipids in large VLDL (mmol/L)*	547	0.20	0.12 - 0.28	544	0.20	0.12 - 0.28	586	0.20	0.12 - 0.28	585	0.18	0.10 - 0.26	1133	0.20	0.15 - 0.26	1129	0.19	0.13 - 0.25
Total lipids in medium VLDL (mmol/L)	547	0.27	0.19 - 0.35	544	0.26	0.18 - 0.34	586	0.25	0.17 - 0.32	585	0.23	0.16 - 0.31	1133	0.26	0.21 - 0.31	1129	0.25	0.19 - 0.30
Total lipids in small VLDL (mmol/L)	547	0.27	0.19 - 0.34	544	0.26	0.18 - 0.33	586	0.26	0.18 - 0.33	585	0.25	0.17 - 0.32	1133	0.26	0.21 - 0.32	1129	0.25	0.20 - 0.31
Total lipids in very small VLDL (mmol/L)	547	0.23	0.15 - 0.31	544	0.23	0.14 - 0.30	586	0.25	0.17 - 0.33	585	0.26	0.18 - 0.33	1133	0.25	0.19 - 0.30	1129	0.24	0.19 - 0.30
Total lipids in LDL (mmol/L)	547	0.28	0.20 - 0.35	544	0.26	0.19 - 0.34	586	0.29	0.21 - 0.36	585	0.29	0.22 - 0.36	1133	0.29	0.23 - 0.34	1129	0.28	0.23 - 0.33
Total lipids in large LDL (mmol/L)	547	0.28	0.20 - 0.36	544	0.27	0.19 - 0.35	586	0.29	0.21 - 0.36	585	0.29	0.21 - 0.36	1133	0.29	0.23 - 0.34	1129	0.28	0.23 - 0.34
Total lipids in medium LDL (mmol/L)	547	0.29	0.21 - 0.36	544	0.28	0.20 - 0.35	586	0.28	0.21 - 0.36	585	0.28	0.21 - 0.36	1133	0.28	0.23 - 0.34	1129	0.28	0.23 - 0.33
Total lipids in small LDL (mmol/L)	547	0.28	0.20 - 0.35	544	0.27	0.19 - 0.35	586	0.28	0.20 - 0.35	585	0.28	0.20 - 0.35	1133	0.28	0.22 - 0.33	1129	0.27	0.22 - 0.33
Total lipids in very large HDL (mmol/L)	547	0.29	0.21 - 0.37	544	0.28	0.20 - 0.36	586	0.29	0.21 - 0.36	585	0.28	0.21 - 0.36	1133	0.29	0.24 - 0.34	1129	0.28	0.23 - 0.33
Total lipids in large HDL (mmol/L)	547	0.29	0.21 - 0.36	544	0.27	0.20 - 0.35	586	0.24	0.17 - 0.32	585	0.24	0.16 - 0.31	1133	0.27	0.21 - 0.32	1129	0.25	0.20 - 0.31
Total lipids in medium HDL (mmol/L)	547	0.15	0.07 - 0.24	544	0.14	0.06 - 0.23	586	0.10	0.02 - 0.18	585	0.10	0.02 - 0.18	1133	0.12	0.07 - 0.18	1129	0.12	0.06 - 0.18
Total lipids in small HDL (mmol/L)	547	0.19	0.10 - 0.27	544	0.18	0.10 - 0.26	586	0.18	0.10 - 0.26	585	0.18	0.10 - 0.26	1133	0.18	0.12 - 0.23	1129	0.18	0.12 - 0.23
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	547	0.29	0.21 - 0.36	544	0.28	0.20 - 0.36	586	0.22	0.14 - 0.29	585	0.21	0.13 - 0.28	1133	0.25	0.20 - 0.31	1129	0.24	0.19 - 0.30
Mean diameter for LDL particles (nm)	547	0.19	0.11 - 0.27	544	0.19	0.11 - 0.27	586	0.27	0.19 - 0.34	585	0.27	0.20 - 0.35	1133	0.23	0.17 - 0.28	1129	0.23	0.17 - 0.28
Mean diameter for HDL particles (nm)	547	0.31	0.23 - 0.38	544	0.30	0.22 - 0.37	586	0.30	0.22 - 0.37	585	0.29	0.22 - 0.36	1133	0.30	0.25 - 0.35	1129	0.29	0.24 - 0.35
Cholesterol																		
Serum total cholesterol (mmol/L)	547	0.28	0.20 - 0.36	544	0.27	0.19 - 0.35	586	0.31	0.24 - 0.38	585	0.31	0.24 - 0.39	1133	0.30	0.24 - 0.35	1129	0.29	0.24 - 0.34
Total cholesterol in VLDL (mmol/L)	547	0.26	0.18 - 0.34	544	0.26	0.18 - 0.33	586	0.26	0.18 - 0.33	585	0.25	0.17 - 0.32	1133	0.26	0.21 - 0.32	1129	0.25	0.20 - 0.31
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	547	0.27	0.19 - 0.34	544	0.26	0.18 - 0.34	586	0.28	0.20 - 0.35	585	0.28	0.20 - 0.35	1133	0.27	0.22 - 0.33	1129	0.27	0.21 - 0.32
Total cholesterol in LDL (mmol/L)	547	0.29	0.21 - 0.36	544	0.28	0.20 - 0.36	586	0.28	0.21 - 0.35	585	0.28	0.21 - 0.35	1133	0.29	0.23 - 0.34	1129	0.28	0.23 - 0.34
Total cholesterol in HDL (mmol/L)	547	0.27	0.19 - 0.35	544	0.25	0.17 - 0.33	586	0.22	0.14 - 0.29	585	0.21	0.13 - 0.28	1133	0.24	0.19 - 0.30	1129	0.23	0.17 - 0.28
Total cholesterol in LDL3 (mmol/L)	547	0.27	0.19 - 0.34	544	0.25	0.17 - 0.33	586	0.21	0.14 - 0.29	585	0.20	0.13 - 0.28	1133	0.24	0.18 - 0.29	1129	0.23	0.17 - 0.28
Total cholesterol in HDL3 (mmol/L)	547	0.25	0.17 - 0.33	544	0.23	0.15 - 0.31	586	0.23	0.15 - 0.31	585	0.22	0.15 - 0.30	1133	0.24	0.19 - 0.30	1129	0.23	0.17 - 0.28
Esterified cholesterol (mmol/L)	543	0.28	0.20 - 0.36	540	0.27	0.19 - 0.35	584	0.31	0.24 - 0.38	583	0.31	0.24 - 0.38	1127	0.30	0.24 - 0.35	1123	0.29	0.24 - 0.34
Free cholesterol (mmol/L)	543	0.27	0.19 - 0.34	540	0.26	0.18 - 0.33	584	0.32	0.24 - 0.39	583	0.32	0.24 - 0.39	1127	0.29	0.24 - 0.35	1123	0.29	0.23 - 0.34
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	547	0.28	0.20 - 0.35	544	0.27	0.19 - 0.34	586	0.25	0.17 - 0.32	585	0.24	0.16 - 0.31	1133	0.26	0.21 - 0.32	1129	0.25	0.20 - 0.31
Triglycerides in VLDL (mmol/L)*	547	0.28	0.20 - 0.36	544	0.28	0.20 - 0.35	586	0.23	0.15 - 0.30	585	0.21	0.14 - 0.29	1133	0.26	0.20 - 0.31	1129	0.25	0.19 - 0.30
Triglycerides in LDL (mmol/L)	547	0.20	0.12 - 0.28	544	0.19	0.11 - 0.27	586	0.27	0.19 - 0.34	585	0.27	0.19 - 0.34	1133	0.24	0.18 - 0.29	1129	0.23	0.18 - 0.29
Triglycerides in HDL (mmol/L)	547	0.25	0.17 - 0.33	544	0.25	0.17 - 0.33	586	0.27	0.20 - 0.35	585	0.27	0.19 - 0.34	1133	0.26	0.21 - 0.32	1129	0.26	0.20 - 0.31
Total phosphoglycerides (mmol/L)	543	0.28	0.20 - 0.36	540	0.27	0.19 - 0.35	584	0.27	0.19 - 0.34	583	0.27	0.20 - 0.35	1127	0.27	0.22 - 0.33	1123	0.27	0.22 - 0.32
Ratio of triglycerides to phosphoglycerides	543	0.23	0.15 - 0.31	540	0.22	0.14 - 0.30	584	0.21	0.13 - 0.28	583	0.19	0.11 - 0.27	1127	0.22	0.16 - 0.27	1123	0.21	0.15 - 0.26
Phosphatidylcholine & other choline (mmol/L)	542	0.28	0.20 - 0.36	539	0.27	0.19 - 0.35	584	0.26	0.19 - 0.34	583	0.26	0.19 - 0.34	1126	0.27	0.21 - 0.32	1122	0.27	0.21 - 0.32
Sphingomyelin (mmol/L)	542	0.23	0.15 - 0.31	539	0.22	0.13 - 0.30	584	0.28	0.21 - 0.36	583	0.28	0.21 - 0.36	1126	0.26	0.20 - 0.31	1122	0.25	0.19 - 0.30
Total choline (mmol/L)	542	0.27	0.19 - 0.35	539	0.27	0.19 - 0.34	584	0.29	0.21 - 0.36	583	0.29	0.21 - 0.36	1126	0.28	0.22 - 0.33	1122	0.27	0.22 - 0.33
Apolipoproteins																		
Apolipoprotein A1 (g/L)	547	0.26	0.18 - 0.34	544	0.25	0.17 - 0.33	586	0.25	0.17 - 0.33	585	0.25	0.17 - 0.32	1133	0.25	0.20 - 0.31	1129	0.24	0.19 - 0.30
Apolipoprotein B (g/L)	547	0.27	0.19 - 0.35	544	0.27	0.19 - 0.34	585	0.28	0.20 - 0.35	584	0.28	0.20 - 0.35	1132	0.28	0.23 - 0.33	1128	0.28	0.22 - 0.33
Ratio of apolipoprotein B to apolipoprotein A1	547	0.26	0.18 - 0.33	544	0.25	0.17 - 0.33	585	0.21	0.13 - 0.29	584	0.20	0.12 - 0.28	1132	0.23	0.18 - 0.29	1128	0.23	0.17 - 0.28
Fatty acids																		
Total fatty acids (mmol/L)	537	0.26	0.18 - 0.33	534	0.26	0.18 - 0.34	583	0.30	0.22 - 0.37	582	0.29	0.22 - 0.37	1120	0.28	0.23 - 0.33	1116	0.28	0.22 - 0.33
Estimated degree of unsaturation	537	0.30	0.22 - 0.37	534	0.30	0.22 - 0.37	583	0.24	0.17 - 0.32	582	0.23	0.15 - 0.30	1120	0.27	0.21 - 0.32	1116	0.26	0.20 - 0.31
22:6, docosahexaenoic acid (mmol/L)	537	0.23	0.15 - 0.31	534	0.22	0.14 - 0.30	583	0.33	0.26 - 0.40	582	0.33	0.25 - 0.40	1120	0.28	0.23 - 0.34	1116	0.27	0.22 - 0.33
18:2, linoleic acid (mmol/L)	537	0.24	0.16 - 0.32	534	0.26	0.17 - 0.33	583	0.27	0.19 - 0.34	582	0.27	0.19 - 0.34	1120	0.26	0.21 - 0.32	1116	0.26	0.21 - 0.32
Omega3 fatty acids (mmol/L)	537	0.24	0.16 - 0.32	534	0.24	0.16 - 0.32	583	0.34	0.27 - 0.41	582	0.34	0.27 - 0.41	1120	0.29	0.23 - 0.34	1116	0.29	0.23 - 0.34
Omega6 fatty acids (mmol/L)	537	0.26	0.18 - 0.33	534	0.26	0.18 - 0.34	583	0.30	0.23 - 0.38	582	0.30	0.23 - 0.38	1120	0.28	0.23 - 0.34	1116	0.28	0.23 - 0.34
Polysat. fatty acids (mmol/L)	537	0.26	0.17 - 0.33	534	0.26	0.18 - 0.34	583	0.31	0.24 - 0.39	582	0.31	0.24 - 0.39	1120	0.29	0.23 - 0.34	1116	0.29	0.23 - 0.34
Monosat. fatty acids; 16:1, 18:1 (mmol/L)	537	0.27	0.19 - 0.35	534	0.27	0.19 - 0.35	583	0.30	0.22 - 0.37	582	0.29	0.22 - 0.37	1120	0.29	0.24 - 0.34	1116	0.28	0.23 - 0.34
Saturated fatty acids (mmol/L)	537	0.25	0.17 - 0.33	534	0.25	0.17 - 0.33	583	0.28	0.20 - 0.35	582	0.28	0.20 - 0.35	1120	0.27	0.21 - 0.32	1116	0.27	0.21 - 0.32
Fatty acid ratios																		
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	537	0.31	0.23 - 0.38	534	0.30	0.22 - 0.37	583	0.33	0.26 - 0.40	582	0.32	0.24 - 0.39	1120	0.32	0.27 - 0.37	1116	0.31	0.25 - 0.36
Ratio of 18:2 linoleic acid to total fatty acids (%)	537	0.15	0.07 - 0.23	534	0.15	0.06 - 0.23	583	0.18	0.10 - 0.26	582	0.17	0.09 - 0.25	1120	0.17	0.11 - 0.23	1116	0.16	0.11 - 0.22
Ratio of omega3 fatty acids to total fatty acids (%)	537	0.32	0.24 - 0.39	534	0.32	0.24 - 0.39	583	0.40	0.33 - 0.47	582	0.39	0.32 - 0.46	1120	0.36	0.31 - 0.41	1116	0.35	0.30 - 0.40
Ratio of omega6 fatty acids to total fatty acids (%)	537	0.24	0.16 - 0.32	534	0.24	0.16 - 0.32	583	0.23	0.15 - 0.30	582	0.22	0.14 - 0.29	1120	0.24	0.18 - 0.29	1116	0.23	0.17 - 0.28
Ratio of polysat. fatty acids to total fatty acids (%)	537																	

STROBE Statement—checklist of items that should be included in reports of observational studies

Paper title: Metabolomics: Population epidemiology and concordance in 11-12 year old

Australians and their parents

Person completing checklist: Susan Ellul

	Item No	Recommendation	Page number	Line number
Title and abstract	1	(a) Indicate the study's design with a commonly used term in the title or the abstract	2	14-15
		(b) Provide in the abstract an informative and balanced summary of what was done and what was found	2	9-13 26-32
Introduction				
Background/rationale	2	Explain the scientific background and rationale for the investigation being reported	4-5	Pg 4 (3-9, 21-22, 31-32, 33-35) Pg 5 (3-4)
Objectives	3	State specific objectives, including any prespecified hypotheses	5	5-7
Methods				
Study design	4	Present key elements of study design early in the paper	5	10-17
Setting	5	Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection	5-6	Pg 5 (33-35) Pg 6 (1-27)
Participants	6	(a) <i>Cohort study</i> —Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up <i>Case control study</i>—Give the eligibility criteria, and the sources and methods of case ascertainment and control selection. Give the rationale for the choice of cases and controls <i>Cross-sectional study</i> —Give the eligibility criteria, and the sources and methods of selection of participants	5	18-26
		(b) <i>Cohort study</i>—For matched studies, give matching criteria and number of exposed and unexposed <i>Case control study</i>—For matched studies, give matching criteria and the number of controls per case		
Variables	7	Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable	6-8	Pg 6 (29-36) Pg 7 (1-6, table 1, 15-19) Pg 8 (1-33)
Data sources/ measurement	8*	For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group	6-8	Pg 6 (29-36) Pg 7 (1-6, table 1, 15-19) Pg 8 (1-33)
Bias	9	Describe any efforts to address potential sources of bias	9	6-10
Study size	10	Explain how the study size was arrived at	10	5-14, figure 1

Quantitative variables	11	Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why	9	2-26
Statistical methods	12	(a) Describe all statistical methods, including those used to control for confounding	9	2-26
		(b) Describe any methods used to examine subgroups and interactions	9	2-26
		(c) Explain how missing data were addressed	9	6-10
		(d) <i>Cohort study</i> —If applicable, explain how loss to follow-up was addressed	9	6-10, 15-17, 20-23
		(e) <i>Case-control study</i>—If applicable, explain how matching of cases and controls was addressed		
		(e) Describe any sensitivity analyses	9	15-17, 20-23

Results			Page number	Line number
Participants	13*	(a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed	10 and figure 1	5-12
		(b) Give reasons for non-participation at each stage	10 6 figure 1	8-12 5-6
		(c) Consider use of a flow diagram	figure 1	
Descriptive data	14*	(a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders	11 (table 2)	2
		(b) Indicate number of participants with missing data for each variable of interest	figure 1 table 2 sup Table 1	2
		(c) <i>Cohort study</i> —Summarise follow-up time (eg, average and total amount)	NA	
Outcome data	15*	<i>Cohort study</i> —Report numbers of outcome events or summary measures over time	sup table 1	
		<i>Case-control study</i> —Report numbers in each exposure category, or summary measures of exposure	NA	
		<i>Cross-sectional study</i> —Report numbers of outcome events or summary measures	NA	
Main results	16	(a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included	12 supp tables 2-6	20-25
		(b) Report category boundaries when continuous variables were categorized	NA	
		(c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period	NA	

Other analyses	17	Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses	supp tables 2-6	
Discussion				
Key results	18	Summarise key results with reference to study objectives	13	3-10
Limitations	19	Discuss limitations of the study, taking into account sources of potential bias or imprecision. Discuss both direction and magnitude of any potential bias	13	18-28
Interpretation	20	Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence	13-16	Pg 13 (30-32) Pg 14 (all) Pg 15 (all) Pg 16 (1-2)
Generalisability	21	Discuss the generalisability (external validity) of the study results	13	19-27
Other information				
Funding	22	Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based	17-18	Pg 17 (28-33) Pg 18 (1-3)

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.

BMJ Open

Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their parents

Journal:	<i>BMJ Open</i>
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Keywords:	Metabolomics, Lipids, Reference values, Children, Inheritance patterns, Epidemiologic studies

SCHOLARONE™
Manuscripts

1 **Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their**
2 **parents**

3
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17
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24
25 **Keywords:** Metabolomics; lipids; inflammation; reference values; parents; children;
26 inheritance patterns; correlation studies; epidemiologic studies; cross-sectional studies.

27
28 **Word count: 4270**

29
30 **Abbreviations:** ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; BCAA: Branched
31 chain amino acid; BD: Becton Dickinson; CDC: Centers for Disease Prevention and Control;
32 CVD: Cardiovascular disease; CPS1: Carbamoyl-phosphate synthase 1; DHA:
33 Docosahexaenoic acid; DOB: Date of birth; EDTA: Ethylenediaminetetraacetic acid; FDR:
34 False discovery rate; GlycA: Glycoprotein acetyls; HbA1c: Haemoglobin A1c; HDL: High-
35 density lipoprotein; HOMA: Homeostatic model assessment; IDL: Intermediate density
36 lipoprotein; LA: Linoleic acid; LDL: Low-density lipoprotein; LiH: Lithium Heparin; LSAC:

1 Longitudinal Study of Australian Children; MUFA: Monounsaturated fatty acid; NMR:
2 Nuclear magnetic resonance; PCOS: Polycystic Ovary Syndrome; PUFA: Polyunsaturated
3 fatty acid; SFA: Saturated fatty acids; SST: serum separating tubes; T2D: Type 2 diabetes;
4 T2DM: Type 2 diabetes mellitus; VLDL: Very low density lipoprotein; XL: Very large;
5 XXL: Chylomicrons and extremely large; XS: Very small.

6 7 **ABSTRACT**

8
9 **Objectives:** Nuclear Magnetic Resonance (NMR) metabolomics is high throughput and cost
10 effective, with the potential to improve the understanding of disease and risk. We examine
11 the circulating metabolic profile by quantitative NMR metabolomics of a sample of
12 Australian 11-12 year old children and their parents, describe differences by age and sex, and
13 explore correlation of metabolites in parent-child dyads.

14 **Design:** The population-based cross-sectional Child Health CheckPoint study nested within
15 the Longitudinal Study of Australian Children.

16 **Setting:** Blood samples collected from CheckPoint participants at assessment centres in
17 seven Australian cities and eight regional towns; February 2015-March 2016.

18 **Participants:** 1180 children and 1325 parents provided a blood sample and had
19 metabolomics data available. This included 1133 parent-child dyads (518 mother-daughter,
20 469 mother-son, 68 father-daughter, and 78 father-son).

21 **Outcome measures:** 228 metabolic measures were obtained for each participant. We
22 focused on 74 biomarkers including amino acid species, lipoprotein subclass measures, lipid
23 measures, fatty acids, measures related to fatty acid saturation, and composite markers of
24 inflammation and energy homeostasis.

25 **Results:** We identified sex-specific metabolic profiles in children and adults and differences
26 in the concentration of specific metabolites between childhood and adulthood. In general,
27 metabolite concentrations were higher in adults than children and sex differences were larger
28 in adults than in children. Positive correlations were observed for the majority of metabolites
29 including for isoleucine (CC 0.33, 95% CI 0.27 to 0.38), total cholesterol (CC 0.30, 95% CI
30 0.24 to 0.35) and omega 6 fatty acids (CC 0.28, 95% CI 0.23 to 0.34) in parent-child
31 comparisons.

32 **Conclusions:** We describe the serum metabolite profiles from mid-childhood and adulthood
33 in a population-based sample, together with parent-child concordance. Distinct age- and sex-
34 specific profiles were observed. These data will be informative for investigation of the
35 childhood origins of adult non-communicable diseases and for comparative studies in other
36 populations.

Strengths and limitations of this study:

- In a large population-based cohort, venous blood was collected for children and their attending parent on the same day using the same methods
- Rapidly processed, high quality serum samples with standardised metabolomic data generated as a single batch
- Cross-sectional design does not enable longitudinal analysis of specific metabolite species over short term or longer periods of time
- Assessment of paternal associations with offspring metabolite measures is limited by a relatively small sample size compared to mother-child pairs, reducing the precision of estimates
- Factors known to influence metabolomic profile (such as body mass index) were not considered as the aim was to describe the distribution of metabolites in children and their parents.

1 INTRODUCTION

2
3 Metabolomics involves the quantitative analysis of a large number of metabolites and lipids
4 involved in a diverse range of biochemical pathways.¹ Genetic/gene expression and
5 environmental exposures are associated with specific metabolic changes across many tissues
6 and body fluids.^{2 3} As such, metabolomics is recognised as a powerful top-down approach to
7 understanding genetic and environmental influences on health and disease. Metabolomic
8 profiling also has considerable potential to identify clinically relevant biomarkers for risk
9 stratification and disease monitoring.

10 Recent advances in nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry
11 have enabled the simultaneous quantitative measurement of hundreds of metabolites. These
12 approaches are sufficiently cost effective and high throughput to be applicable to large cohort
13 studies. For example, NMR metabolomics of serum from the Cardiovascular Risk in Young
14 Finns Study identified many biomarkers from multiple metabolic pathways reflective of fatty
15 liver disease.⁴ These were also predictive of risk 10 years prior to diagnosis, indicating that
16 metabolic disruptions precede overt phenotype. Similar population and disease-specific
17 studies have identified metabolomic profiles associated with a range of exposures and health
18 outcomes with potential to reveal clinically important biomarkers and information on disease
19 mechanisms.⁵ In addition, specific serum metabolites can also be considered ‘intermediate
20 phenotypes’ linking genetic risk with disease outcomes.^{6 7}

21 Previous research indicates that some blood metabolites change with age, particularly from
22 mid to late adulthood.^{8 9} However, in adults sex appears to be a major driver of variation in
23 metabolite profile, potentially interacting with age. For example, the effects of sex appeared
24 to be greater in younger (age 25-35) than older Japanese adults.¹⁰ A study of 26,000 Northern
25 European adults identified many sex-specific metabolic species at the population level.⁹ In
26 men, several lipid measures begin to rise at early middle age whereas a similar increase is
27 only observed in females post menopause. This pattern is consistent for all non-HDL
28 cholesterol measures – very low density lipoprotein (VLDL), intermediate density lipoprotein
29 (IDL) and low density lipoprotein (LDL) subclass particle concentrations - as well as for
30 triglycerides.⁹ Physiological states such as pregnancy also have consistent and measurable
31 influence on serum metabolome.¹¹ However, it remains unclear how the serum metabolome
32 differs in adults compared to children and by sex particularly in childhood.

33 Moreover, factors regulating the metabolic trajectory from early life to adulthood, the role of
34 metabolomic profile in health at the population level and the extent to which blood
35 metabolomic profiles are concordant for parents and children have not been fully explored.
36 One small study has reported correlations between parents (n=179) and their offspring

(n=255) for a range of cardiometabolic risk factors including standard lipid profile measured using conventional methods; this proved stronger for total cholesterol and LDL cholesterol than for high density lipoprotein (HDL) cholesterol or triglycerides.¹² Considerable evidence exists that the metabolomic profile is regulated, at least in part, by genetic factors^{13 14} and is also influenced by dietary and lifestyle factors. Each of these influences is likely to be shared between parents and their offspring to varying degrees, however, parent-child correlations of metabolites from NMR-based platforms have not been reported previously.

Here, we describe (1) the distribution of NMR-based metabolite measures in a population-based cohort of 11-12 year old children and their parents, differences in metabolite concentrations (2) by age (adults compared to children) and (3) by sex in children and adults; and (4) report sex-specific parent-child concordance.

METHODS

Study Design: Details of the initial Longitudinal Study of Australian Children (LSAC) study design and recruitment are outlined elsewhere.^{15 16} The LSAC commenced in 2004, when two cohorts (the 'B' and 'K' cohorts, of which the B cohort only was included in the present study) were recruited who have since been followed biennially. The Child Health CheckPoint comprised a detailed cross-sectional assessment of physical health and biomarkers in a population-based national sample of children (age 11-12 years) and their parents between February 2015 to March 2016. The CheckPoint was nested between waves 6 (2014) and 7 (2016) of the LSAC. Further details regarding the CheckPoint study design and methods are available elsewhere.^{17 18}

Participants: Of the 8,921 families contacted to be part of the LSAC B cohort 5,107 families (57%) agreed to take part in the first wave of data collection in 2004; 4,484 families were retained for Wave 6 in 2014. During the Wave 6 LSAC home visit, B cohort families were introduced to the upcoming Child Health CheckPoint and asked to consent to their contact details being shared with the CheckPoint team. A total of 3,513 families provided permission to receive an information pack by mail and an information and recruitment phone call regarding the CheckPoint study (78% of Wave 6 cohort, 69% of original cohort). Of the families agreeing to receive information about the CheckPoint study, 1,874 families took part (53% of eligible participants, 42% of Wave 6 cohort and 37% of original cohort).

Ethics and consent: The CheckPoint data collection protocol was approved by The Royal Children's Hospital (Melbourne, Australia) Human Research Ethics Committee (33225D) and the Australian Institute of Family Studies Ethics Committee (14-26). The attending parent/caregiver provided written informed consent for themselves and their child to

1 participate in the study, and asked to provide optional consent for the collection and use of
2 biological samples.

3 **Procedure:** The specialised CheckPoint assessment centre sequentially visited seven
4 Australian cities and eight regional towns between February 2015 and March 2016.¹⁸ Each
5 participating child attended the centre with one parent or caregiver (usually the biological
6 mother) at which both participated in a wide range of measures relevant to non-
7 communicable disease. Those families who could not attend a centre were offered a home
8 visit. Participants were included in the current analyses if metabolomic data from CheckPoint
9 were available (figure 1). Venous blood was not available for home-visit participants, but was
10 collected at all city and most regional assessment centres. Participant pairs were excluded
11 from the concordance analyses in this study if the attending parent was not the biological
12 parent.

13 An experienced phlebotomist collected approximately 28mL of blood from the brachial vein
14 of the non-dominant arm of semi-reclining, semi-fasted participants (at the time of collection,
15 participants reported when they last ate or drank). Blood was collected sequentially into four
16 Becton Dickinson (BD) Vacutainer[®] tubes using a butterfly needle so only a single
17 venepuncture was required. Order of collection was (i) 2.7mL EDTA, (ii) 9mL EDTA, (iii)
18 9mL serum, (iv) 7.5mL Lithium Heparin. The latter two tubes were immediately inverted 6
19 times to ensure mixing with anticoagulant, and all tubes were transferred to the on-site
20 laboratory. Time of collection was scheduled earlier in the visit for parents than for children.

21 Collection tube barcodes were linked to the participant and samples were immediately
22 transported to an on-site laboratory where they were processed within two hours. Blood
23 clotting was allowed at room temperature for at least 30 minutes after collection. The sample
24 tubes were spun at 550g relative centrifugal force for 10 minutes at room temperature and
25 distributed into 0.5mL aliquots of plasma, serum, buffy coat (lymphocytes), whole blood
26 and/or an aliquot tube containing a blood clot (1.0mL FluidX screwcap tubes, Cheshire, UK)
27 and stored immediately at -80°C (Thermo Fisher Scientific, Waltham, USA). Each FluidX
28 tube contained a unique 2D barcode linked to the original collection tube and participant. As
29 each assessment centre closed, samples were shipped on dry ice to the Melbourne Children's
30 Bioresource Centre for long term storage at -80°C (serum, whole blood, plasma, blood clot)
31 or vapour phase liquid nitrogen (lymphocytes). At a later date, single 0.5ml serum aliquot
32 was removed for every CheckPoint participant and the combined aliquots were shipped in a
33 single batch to Nightingale Health (Helsinki, Finland) on dry ice for NMR metabolomics.

34
35

1 **Measures**

2 3 Metabolomic profiling

4 The Nightingale[®] NMR metabolomics platform (Helsinki, Finland) was used to obtain
5 metabolomics for children and parents using the 2016-version quantification algorithm.
6 Details of this platform and methodology have been extensively described elsewhere,^{6 19} and
7 epidemiological applications were recently reviewed.²⁰ Briefly, metabolites were measured
8 from 0.35mL of serum using a single high-throughput experimental setup for the
9 simultaneous quantification of routine lipids, lipoprotein subclass distributions, particle size
10 and composition, fatty acids, and other low-molecular weight metabolites such as amino
11 acids and glycolysis-related metabolites. This generated data on 228 serum metabolite
12 measures in absolute concentration units (eg millimoles per liter) and ratios (summarised in
13 Table 1). Whilst widely used for epidemiological research, the NMR-based quantification has
14 not been certified for clinical diagnostics. Further analytical validation of the quantification
15 protocols for the biomarker subset routinely used in clinical settings (eg established
16 cholesterol measures and creatinine) is expected to lead to recalibration of certain metabolite
17 concentrations to better match clinical gold standards.²⁰

18

1 **Table 1. Summary of biomarkers and derived variables obtained via high-throughput NMR**

Metabolic group	Species and derived measures
Amino acids	Alanine, Glutamine, Glycine, Histidine
	Branched chain: Isoleucine, Leucine, Valine
	Aromatic: Phenylalanine, Tyrosine
Cholesterol	VLDL, LDL, HDL, HDL2, HDL3, Total, Free, Esterified, Remnant
Triglycerides and phospholipids	Triglycerides (VLDL, LDL, HDL, total)
	Phosphoglycerides
	Ratio of triglycerides to phosphoglycerides*
	Phosphatidylcholine
	Sphingomyelins
Total cholines	
Apolipoproteins	Apolipoprotein A-1 (ApoA-1)
	Apolipoprotein B (Apo B)
	Ratio of Apolipoprotein B to Apolipoprotein A-1 (ApoB/Apo A-1)*
Fatty acids (FA)	Total, Omega-3, Omega-6, Polyunsaturated (PUFA), Saturated (SFA)
	Monounsaturated (MUFA), Docosahexaenoic acid (DHA), Linoleic (LA)
	Estimated degree of unsaturation
Fatty acid ratios	Omega-3/total FA*, Omega-6/total FA*, PUFA/total FA* (all %)
	SFA/total FA*, MUFA/total FA*, DHA/total FA*, LA/total FA* (all %)
Lipoprotein subclasses*	12 lipid measures in each of 14 subclasses VLDL (XXL, XL, L, M, S, XS), IDL, LDL (L,M,S), HDL (XL, L, M,S): Particle concentration, Total lipids, Esterified cholesterol, Total cholesterol, Phospholipids, Free cholesterol, Triglycerides and Esterified cholesterol/Total lipids (%), Free cholesterol/Total lipids (%), Total cholesterol/Total lipids (%), Triglycerides/Total lipids (%) and Phospholipids/Total lipids (%).
Lipoprotein size*	Mean diameter of VLDL, LDL and HDL particles
Ketone bodies	Acetate, Acetoacetate, 3-hydroxybutyrate
Glycolysis related	Glucose, Lactate, Pyruvate, Citrate, Glycerol
Fluid balance	Creatinine, Albumin
Inflammation	Glycoprotein acetyls (GlycA)

2 Information obtained from <https://nightingalehealth.com/science/biomarkers>

3 * ratio; ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; DHA: Docosahexaenoic acid; GlycA: Glycoprotein acetyls;
4 HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; L: Large; LDL: Low-density lipoprotein; LA:
5 Linoleic acid; M: Medium; MUFA: Monounsaturated fatty acid; PUFA: Polyunsaturated fatty acid; S: Small; SFA:
6 Saturated fatty acids; VLDL: Very low density lipoprotein; XL: Very large; XXL: Chylomicrons and extremely large; XS:
7 Very small.

8
9 Many of the 228 metabolomics measures correlate substantially both in adults
10 (supplementary figure 1) and children (supplementary figure 2) and the pattern of correlations
11 were similar for adults and children. For clarity, we therefore focused on a subset of 74
12 metabolites in analyses. We eliminated the 5 ratio measures for each of the 14 lipoprotein
13 subclass particles. In addition, the 7 other measures within each of the lipoproteins (esterified
14 cholesterol, free cholesterol, total cholesterol, triglycerides, phospholipids, total lipids and
15 particle concentration) are all highly correlated and therefore we only reported total lipids for
16 each of the lipoprotein subclass particles.

1 Other measures and sample characteristics

2 *Age and sex:* For children, LSAC provided date of birth (DOB) and sex, which was originally
3 exported from the Medicare Australia database. In parents, DOB and sex was self-reported in
4 the CheckPoint questionnaire. Age in years was calculated as the difference between date of
5 the CheckPoint assessment and DOB divided by 365.

6 *Disadvantage index:* LSAC provided contact details of families consenting to be contacted
7 by CheckPoint. The family's residential postcode was confirmed during the CheckPoint
8 recruitment phone call and updated, if required. The disadvantage index score of postcode
9 was used to summarise neighbourhood socioeconomic position. Generated by the ABS
10 from the 2011 national Census, the index numerically summarises the social and economic
11 conditions of Australian neighbourhoods; national mean 1000, standard deviation 100;
12 higher scores indicate less disadvantage.²¹

13
14 *Time of blood collection, processing and fasting time:* Time of blood collection and start of
15 laboratory processing were recorded. When missing, collection time was estimated using the
16 midpoint between the time the CheckPoint visit began and time that processing of the sample
17 commenced. Processing lag time was calculated as the minutes between blood collection and
18 the processing commencement. Most samples were processed within two hours.

19 Fasting time was calculated as the hours between last eating/drinking to time of blood
20 collection. The last time of eating/drinking was cross-checked against when the participant
21 was taking part in other CheckPoint stations (and known not to be eating) as well as sleep and
22 wake times from accelerometry data (to identify usual activity, and therefore likely eating
23 patterns) when available. Further details of cleaning processes for the time of last eat/drink
24 can be found elsewhere.²²

25 **Statistical analysis**

26 Sample Characteristics

27
28 Continuous descriptive variables and metabolite measures were summarised using means and
29 standard deviations (SD) for children and adults separately, by sex and overall. For skewed
30 metabolites, geometric means and relative SD were reported. To provide visual comparisons
31 of distributions of metabolites by age and sex, density plots were used. Population summary
32 statistics were estimated by applying survey weights and survey procedures that corrected for
33 sampling, participation and non-response biases, and took into account clustering in the
34 sampling frame. Standard errors were calculated taking into account the complex design and
35 weights.²³ More detail on the calculation of weights is provided elsewhere.²⁴

1 Differences in metabolite concentration by age (adults compared to children) and by sex
2 (adults, children)

3 Skewed metabolites (skewness greater or equal to 2) were log-transformed. We used two-
4 sided paired and unpaired t-tests (as appropriate) to assess differences in mean metabolite
5 concentrations between adults and children in parent-child dyads, and between males and
6 females for adults and children separately. P-values were adjusted using Benjamini-
7 Hochberg (B-H) with a false discovery rate (FDR) of 10% to account for multiple
8 comparisons.

9 Parent-Child concordance

10 Concordance between parents and children was assessed by 1) Pearson's correlation
11 coefficients (CC) with 95% confidence intervals, and 2) partial correlation coefficients
12 (PCC), adjusting for child and parent age, disadvantage index, fasting time, processing lag
13 time (and for child and parent sex where appropriate). Scatterplots of parent versus child
14 metabolites (log-transformed where needed as above) were examined to check for outliers
15 and to ensure assumptions were met.

16 The analyses were repeated using weighted multi-level survey analyses and compared to
17 unweighted analyses. As there appeared to be no major effect of response patterns on
18 results we reported results from unweighted analyses. Analyses were undertaken using
19 Stata version 14.2 (StataCorp, College Station, TX) and R version 3.3.2.²⁵

20 **Patient and Public Involvement:** Because LSAC is a population-based longitudinal study,
21 no patient groups were involved in its design or conduct. To our knowledge, the public was
22 not involved in the study design, recruitment or conduct of LSAC study or its CheckPoint
23 module. Parents received a summary health report for their child and themselves at or soon
24 after the assessment visit. They consented to take part knowing that they would not otherwise
25 receive individual results about themselves or their child.

1 RESULTS

3 Sample characteristics

5 The recruitment and retention of participants in the Child Health CheckPoint are described
6 elsewhere.¹⁸ Of the 1874 families who participated in CheckPoint assessment centres, blood
7 serum samples of analysable quality from 1180 children and 1325 parents (figure 1) were
8 sent for NMR quantification of metabolites. The majority of excluded families undertook
9 home visits or attended a regional centre, where blood samples could not be collected (n=385,
10 20.5%), while some participants declined a blood sample (children, n=150, 8.0%; adults,
11 n=108, 5.8%). Few data were lost due to insufficient volume or poor quality samples at the
12 assessment centre (figure 1). The sample characteristics of parents and children are outlined
13 in table 2. Summary statistics for our main child and parent metabolite measures are
14 presented in supplementary table 1. Supplementary figures 3-7 show density plots comparing
15 the distributions of metabolites for boys, girls and adults.

Table 2: Sample characteristics; values are weighted mean (standard deviation)

Characteristic	All	Male	Female
Child			
n	1152-1180	558-575	594-605
Age, years	12.0 (0.4)	12.0 (0.4)	12.0 (0.4)
Disadvantage Index	1012 (63)	1011 (65)	1014 (61)
Fasting time (hours)	4.2 (1.2)	4.3 (1.3)	4.2 (1.1)
Time of day - blood collection	14.16 (2.0)	14.12 (2.0)	14.20 (2.1)
Processing lag time (hours)	1.16 (0.5)	1.18 (0.5)	1.14 (0.5)
Parent			
n	1272-1325	174-177	1098-1148
Age, years	43.9 (5.6)	46.9 (6.9)	43.4 (5.2)
Fasting time (hours)	3.3 (1.6)	3.6 (2.0)	3.2 (1.5)
Time of day - blood collection	13.10 (2.0)	13.18 (2.1)	13.09 (2.0)
Processing lag time (hours)	1.26 (0.5)	1.31 (0.5)	1.26 (0.5)

Disadvantage Index: Index of Relative Socioeconomic Disadvantage; n: number of participants in cohort with this measure.

Differences in metabolite levels by age (adults compared to children)

Figure 2 shows mean differences in metabolite levels for adults relative to children in standard deviation (SD) units. Most concentrations were higher in adults than children. Values that were similar in adults and children included total lipids in very large HDL lipoprotein subclass particles, acetoacetate, tyrosine and glucose. Levels in children were higher than those of adults for the majority of glycolysis related measures (lactate, pyruvate, citrate and glycerol), the ketone body 3-hydroxybutrate, the amino acid glutamine, many fatty acid ratios and all lipoprotein particle sizes. Supplementary table 2 lists the corresponding estimates in absolute concentration units.

Sex-specific differences in metabolite levels in children and adults

Figure 3 shows differences in mean metabolite levels by sex for children and adults separately in SD units, with estimates in absolute concentration units listed in supplementary table 3 and 4.

In general, sex differences were more pronounced in adulthood, resulting in distinct overall patterns for children and adults. Children generally showed smaller differences by sex than

1 adults. Of note, sex differences for apolipoproteins and fatty acid measures showed different
2 patterns in children compared to adults.

3 Girls had lower levels of apolipoprotein-A-1 (ApoA-1) and higher ApoB than boys. In adults,
4 the opposite pattern was observed with females having higher ApoA-1 and lower ApoB than
5 males. In children, some fatty acid concentrations were higher in girls than boys. In contrast,
6 many adult fatty acid measures were higher in males. There was no evidence of a difference
7 in the level of inflammation (GlycA) by sex in children, while in adults, GlycA levels tended
8 to be higher in males than females.

9 For some metabolites, sex differences in children mirrored (but were smaller in magnitude
10 than) those of adults, particularly for the ketone body acetate and some key amino acids. At
11 both ages, the amino acid glycine was higher in females but the branched-chain amino acids
12 leucine and valine tended to be higher in males.

13 **Parent-child concordance**

14 Figure 4 shows the correlations between metabolite measures for all children with all parents,
15 and for boys and girls with mothers (but not with the 177 fathers, given the small numbers).
16 The corresponding correlation coefficients and partial correlation coefficients are listed in
17 supplementary tables 5 and 6.

18 Correlations for all parents and all children showed similar patterns to that observed for
19 mother and child by sex. While there was little suggestion of substantial correlation within
20 parent-child dyads for some metabolites (eg glucose, acetate) a positive correlation was found
21 for many metabolite measures irrespective of child sex. For example, positive correlations
22 were observed for isoleucine (CC 0.33, 95% CI 0.27 to 0.38), total serum cholesterol (CC
23 0.30, 95% CI 0.24 to 0.35) and omega 6 fatty acids (CC 0.28, 95% CI 0.23 to 0.34) in parent-
24 child comparisons. Additional adjustment for factors that potentially influence metabolite
25 levels (age, socioeconomic status, fasting time and processing lag time) had little effect on
26 the degree of correlation in any comparison (supplementary tables 5 and 6).

27

1 DISCUSSION

2 Principal findings

3 Here we present age and sex differences, describing the distribution of detailed/NMR-based
4 metabolite measures in Australian 11-12 year old children and their parents, and demonstrate
5 that many metabolite measures have high parent-child concordance. In accord with previous
6 studies, we observed major differences in metabolite levels between childhood and adulthood
7 and also differences by sex in both childhood and adulthood. We also observed variability in
8 the magnitude of differences by sex for several metabolites in childhood compared to
9 adulthood and identified a complex interplay of correlations of specific metabolites between
10 parents and their children according to parent-child sex relationships.

11 Strengths and weaknesses

12 This is the first major cohort study to report both sex- and cross-generational differences in
13 metabolomic concentrations in mid-childhood to adulthood utilising the NMR platform.
14 Further strengths include the large number of parent-child dyads representing a wide range of
15 parent ages, the national population-based sample and the state-of-the-art measurements.
16 Replication studies exploring sex differences at earlier and later stages of childhood and
17 adolescence would strengthen findings.

18 An important limitation is that paternal factors were not fully represented, as most parental
19 samples were from mothers (a well-documented problem in longitudinal cohort studies). This
20 also limited sex-specific parental contribution analysis; further studies including more fathers
21 are warranted. Additional limitations are that, without samples from both parents for each
22 child, we could not estimate heritability, and our results might not apply to mid-life adults
23 who are not parents (although we see no good reason why these would differ greatly). The
24 original uptake of just over 50% and subsequent attrition within LSAC and then the
25 CheckPoint has led to a relatively advantaged sample, but nonetheless participants varied
26 widely on key potential confounders (eg disadvantage, age) and this was at least partly offset
27 by application or consideration of survey weights. Given the large number of metabolites and
28 modest sample size, considerable uncertainty remains in any ranking of the various effects
29 across metabolites.

30 Meaning and implications for clinicians and policymakers

31 Overall, we found a clear difference in metabolite profile between children and their parents.
32 This was apparent for specific metabolite measures (such as some amino acids) as well as the

1 distribution of metabolites (such as lipid composition of lipoproteins of different density).
2 Some measures were higher in adults, some similar, while a minority were lower. Previous
3 studies, largely in adults, have identified a range of specific metabolite changes with age,
4 particularly from mid to late adulthood.²⁶ This includes a general decrease in several amino
5 acid species, which contrasts with our findings from childhood to mid adulthood.⁸ Only the
6 amino acid glutamine showed this pattern in our dataset.

7 Sex-specific differences in children (± 0.2 SD) were generally much smaller than in adults
8 (± 0.8 SD). Large metabolomic studies using alternative platforms have previously reported
9 reproducible, sex-specific signatures in circulating metabolite profile in adults.²⁷⁻²⁸ This
10 includes differences in amino acid and lipid serum concentrations, potentially influenced by
11 sex-specific effects of genetic polymorphisms on metabolite levels.²⁸⁻²⁹ As in our study, most
12 amino acids have usually been reported to be higher in men than women.²⁸⁻³⁰ For example, in a
13 recent study of 507 metabolic markers in 1756 individuals (903 female and 853 male aged
14 ~60 years), one third of metabolites showed significant sexual dimorphism. These were
15 predominantly related to pathways of steroid metabolism, fatty acids, other lipids, and a large
16 proportion of amino acids.³⁰ Of particular note, branched chain amino acids (BCAAs) and
17 their related metabolic products were amongst the most differentially represented, with much
18 higher isoleucine, leucine and valine in males. A similar finding of higher leucine and valine
19 was also noted in the Cooperative Health Research in the Region of Augsburg (KORA)
20 follow-ups 3 (F3) and 4 (F4) analysis of >3000 adults,²⁸ consistent with our observations in
21 adulthood.

22 In children, we found sex-specific differences for leucine and valine were smaller but in the
23 same direction as adults. Several lines of evidence implicate BCAA metabolism with
24 metabolic risk in humans. For example, three candidate genes for obesity and/or type 2
25 diabetes mellitus (T2DM) are involved in the BCAA metabolic pathway.³¹ In a recent large
26 meta-analysis of metabolomics in diabetes, a >30% higher risk of type 2 diabetes was found
27 per SD increase in isoleucine, leucine, valine or tyrosine, whereas glycine and glutamine
28 were inversely associated with risk.³¹ Several clinical studies have also reported that BCAAs
29 positively correlate with insulin resistance, homeostatic model assessment (HOMA) index
30 and levels of haemoglobin A1c (HbA1c), while longitudinal studies have reported that
31 increased blood BCAAs are predictive of future insulin resistance and type 2 diabetes
32 (T2D).³² It is intriguing to speculate that the higher BCAA in males from early life could
33 contribute to the well-described increasing prevalence of T2D in men. Levels of BCAA are

1 elevated in females with Polycystic Ovary Syndrome (PCOS), potentially contributing to the
2 associated insulin resistance.³³ However, it remains unclear whether BCAA are on the causal
3 pathway to T2D or result from adverse metabolic health. Our demonstration that the sex-
4 specific differences in BCAA arise early in life offers potential to track their association with
5 sex-specific measures of metabolic health from an early age to help clarify where they lie on
6 the causal pathway.

7 In accord with previous adult studies²⁸, we found higher levels of glycine in mothers than
8 fathers, and (less markedly) in girls than boys. Interestingly, recent metabolomics and genetic
9 analyses of ~10,000 adults with cardiovascular disease (CVD), with replication in >53,000
10 subjects, identified a genetic variant in carbamoyl-phosphate synthase 1 (*CPS1*) (linked to
11 plasma glycine levels) to be strongly associated with a reduced risk of CVD in women
12 ($p=6.3 \times 10^{-5}$) but not men ($p=0.95$), suggesting a direct link between glycine levels and CVD
13 risk, although whether this is a causal association remains unclear.³⁴ It will be interesting in
14 the future to explore the link between variants in *CPS1* and circulating glycine levels from
15 early life to adulthood in relation to markers of cardiovascular health in females.

16 The small sex-differences of HDL cholesterol and ApoA-1 in children compared to adults is
17 consistent with modest differences in children, whereas substantial differences in adulthood
18 have previously been reported.³⁵ ApoA-1 was more abundant in boys, while ApoB was
19 higher in girls, leading to a higher ApoB/ApoA-1 ratio in girls. The opposite pattern was
20 found in our limited sample of fathers relative to mothers. These data are surprising and
21 differ from a similarly sized study of slightly older European adolescent children (mean age
22 15 years) that found higher ApoA-1 and ApoB in girls relative to boys.³⁶ Interestingly, a
23 higher ApoB/ApoA-1 ratio has been strongly linked to increased coronary risk in adults,³⁷⁻³⁹
24 suggesting that the sex-specific differences may alter with increasing age, in keeping with the
25 increased CVD risk in adult males. ApoA-1 is the main protein component of HDL
26 cholesterol⁴⁰ thus the differences in trajectories in lipids and HDL cholesterol for boys and
27 girls across childhood that have been reported^{41 42} could partially explain this observation.

28 These are the first data on the mother-child or parent-child correlations of NMR metabolites.
29 Smaller studies have reported positive correlations between parents and children for a limited
30 range of cardiometabolic risk factors including total cholesterol, LDL cholesterol, HDL
31 cholesterol and triglycerides measured using conventional methods. We found positive
32 correlations between parents and children for the same lipid measures (although measured
33 using NMR) consistent with previously reported findings. One study reported a positive

1 association between the serum lipid levels of 4 year old children (n=127) and their parents
2 (122 mothers and 118 fathers)⁴³ while another study of children aged 6-18 (n=255) and their
3 parents (n=179) found that the age of the child influenced the degree of correlation of several
4 lipid measures, with older (10-18 years) children more similar to their parents in terms of
5 triglyceride levels than younger individuals (6-9 years).¹²

6 **Unanswered questions and future research**

7 The temporal and sex specific dynamism of the metabolomics data we describe here offer
8 considerable opportunities for identification of biomarkers of risk for a range of non-
9 communicable diseases early in life, to inform targeted interventions and monitor their
10 efficacy. Combining metabolomics with other 'omics data (such as genetics), as is
11 increasingly reported from large adult studies, offers considerable promise in understanding
12 the causal pathways that link early life exposures, genetics and intermediate phenotypes with
13 later onset chronic disease, and in identifying clinically relevant biomarkers.

14 In conclusion, we describe the metabolite profile from mid-childhood and adulthood in a
15 population-based sample, together with parent-child concordance and sex-specific differences
16 in children and adults. In this descriptive paper, distinct age- and sex-specific profiles were
17 observed, as well as considerable evidence of correlation between parent and child measures.
18 These data will be informative for investigation of the childhood origins of adult non-
19 communicable diseases and for comparative studies across populations.

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6 should not be attributed to DSS, AIFS or the ABS.

7
8 REDCap (Research Electronic Data Capture) tools [ENREF 43](#)⁴⁴ were used in this study. More
9 information about this software can be found at: www.project-redcap.org.

10 We thank the LSAC and CheckPoint study participants, staff and students for their
11 contributions.

12 COMPETING INTERESTS

13
14 All authors have completed the ICMJE uniform disclosure form at
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3

4 **CONTRIBUTIONS**

5
6 DB, RS and JC conceptualised and developed the Metabolomics Checkpoint study. SE and
7 JC undertook all aspects of data analysis. SAC coordinated the acquisition of metabolomics
8 data and provided critical review of this manuscript. MW, the Principal Investigator of the
9 Child Health CheckPoint, planned the analyses and provided critical review of this
10 manuscript. SE and RS drafted the manuscript. PW, MJ, TD, KL, JC, DB provided critical
11 expert advice and critical review of this manuscript.

12 **DATA SHARING STATEMENT**

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14 Dataset and technical documents available from *Growing Up in Australia: The Longitudinal*
15 *Study of Australian Children* via low-cost license for bone fide researchers. More information
16 is available at www.growingupinaustralia.gov.au

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1 **FIGURE CAPTIONS AND FOOTNOTES**

2 **Figure 1: Participant flow chart.**

3 n=number of families, c=number of children, p=number of attending adults,

4 MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,

5 LSAC=Longitudinal Study of Australian Children

6 *Unable to analyse due to insufficient volume or poor quality sample

7 ^Data from 6 non-biological child-parent pairs excluded from concordance analyses

8 **Figure 2: Differences in metabolite levels between children and adults.**

9 Association measures are SD difference in metabolite concentration for adults compared to
10 children. Error bars represent 95% confidence intervals. Significant associations after p-
11 values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold
12 (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals
13 and associated p-values are listed in supplementary table 2. HDL: High-density lipoprotein;
14 IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low
15 density lipoprotein.

16 **Figure 3: Sex-specific differences in metabolite levels in childhood and adulthood.**

17 Association measures are SD difference in metabolite concentration for females compared to
18 males in children (A) and adults (B). Error bars represent 95% confidence intervals.
19 Significant associations after p-values adjusted for multiple testing using Benjamini-
20 Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute
21 concentration units, 95% confidence intervals and associated p-values are listed in
22 supplementary table 3 and 4. HDL: High-density lipoprotein; IDL: Intermediate density
23 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

24 **Figure 4: Parent:child correlation for metabolite measures.**

25 Pearson's correlation coefficients for all children with all parents (A); and for boys (blue)
26 with mothers and for girls (red) with mothers (B). Error bars represent 95% confidence
27 intervals. Correlation coefficients with associated 95% confidence intervals are listed in

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1 supplementary table 5 and 6. HDL: High-density lipoprotein; IDL: Intermediate density
2 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.
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SUPPLEMENTARY DOCUMENTS

Supplementary figure 1: Correlation of NMR measures in children.

Heatmap showing the correlation between metabolite measures in children. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 2: Correlation of NMR metabolite measures in parents.

Heatmap showing the correlation between metabolite measures in parents. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 3: Density plots for cholesterol and apolipoprotein measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each cholesterol and apolipoprotein measure.

Supplementary figure 4: Density plots for fatty acid and fatty acid ratio measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each fatty acid and fatty acid ratio measure.

Supplementary figure 5: Density plots for total lipids in the 14 lipoprotein subclass particles.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for total lipids within each of the 14 lipoprotein subclass particles.

Supplementary figure 6: Density plots for lipoprotein particle size and triglyceride measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for lipoprotein particle sizes and triglyceride measures.

Supplementary figure 7: Density plots for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

Supplementary table 1: Weighted mean (SD) of metabolite measures in children and parents.

Supplementary table 2: Mean difference in metabolite levels in adults compared to children in absolute concentration units.

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1 **Supplementary table 3: Differences in mean metabolite levels in girls compared to boys**
2 **in absolute concentration units.**

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4 **Supplementary table 4: Differences in mean metabolite levels in female compared to**
5 **male adults in absolute concentration units.**

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8 **Supplementary table 5: Mother-child concordance; correlations and partial correlations**
9 **between mothers and their sons, daughters and all children.**

10 **Supplementary table 6: Parent-child concordance; correlation and partial correlations**
11 **between all parents and their sons, daughters and all children.**
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1 REFERENCES

1. Nicholson JK, Lindon JC, Holmes E. 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica* 1999;29(11):1181-9. doi: 10.1080/004982599238047
2. Nath AP, Ritchie SC, Byars SG, et al. An interaction map of circulating metabolites, immune gene networks, and their genetic regulation. *Genome biology* 2017;18(1):146. doi: 10.1186/s13059-017-1279-y
3. Shah SH, Newgard CB. Integrated metabolomics and genomics: systems approaches to biomarkers and mechanisms of cardiovascular disease. *Circ Cardiovasc Genet* 2015;8(2):410-9. doi: 10.1161/CIRCGENETICS.114.000223
4. Kaikkonen JE, Wurtz P, Suomela E, et al. Metabolic profiling of fatty liver in young and middle-aged adults: Cross-sectional and prospective analyses of the Young Finns Study. *Hepatology* 2017;65(2):491-500. doi: 10.1002/hep.28899
5. Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol* 2016;17(7):451-9. doi: 10.1038/nrm.2016.25
6. Kettunen J, Tukiainen T, Sarin AP, et al. Genome-wide association study identifies multiple loci influencing human serum metabolite levels. *Nat Genet* 2012;44(3):269-76. doi: 10.1038/ng.1073
7. Suhre K, Gieger C. Genetic variation in metabolic phenotypes: study designs and applications. *Nat Rev Genet* 2012;13(11):759-69. doi: 10.1038/nrg3314
8. Yu Z, Zhai G, Singmann P, et al. Human serum metabolic profiles are age dependent. *Aging Cell* 2012;11(6):960-7. doi: 10.1111/j.1474-9726.2012.00865.x
9. Auro K, Joensuu A, Fischer K, et al. A metabolic view on menopause and ageing. *Nat Commun* 2014;5:4708. doi: 10.1038/ncomms5708
10. Saito K, Maekawa K, Kinchen JM, et al. Gender- and Age-Associated Differences in Serum Metabolite Profiles among Japanese Populations. *Biol Pharm Bull* 2016;39(7):1179-86. doi: 10.1248/bpb.b16-00226
11. Wang Q, Wurtz P, Auro K, et al. Metabolic profiling of pregnancy: cross-sectional and longitudinal evidence. *BMC Med* 2016;14(1):205. doi: 10.1186/s12916-016-0733-0
12. Halvorsen T, Moran A, Jacobs DR, Jr., et al. Relation of Cardiometabolic Risk Factors between Parents and Children. *J Pediatr* 2015;167(5):1049-56 e2. doi: 10.1016/j.jpeds.2015.07.053
13. Rueedi R, Ledda M, Nicholls AW, et al. Genome-Wide Association Study of Metabolic Traits Reveals Novel Gene-Metabolite-Disease Links. *PLOS Genetics* 2014;10(2):e1004132. doi: 10.1371/journal.pgen.1004132
14. Kettunen J, Demirkan A, Würtz P, et al. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nature Communications* 2016;7:11122. doi: 10.1038/ncomms11122
<https://www.nature.com/articles/ncomms11122#supplementary-information>
15. Edwards B. Growing Up in Australia: The Longitudinal Study of Australian Children: Entering adolescence and becoming a young adult. *Family Matters* 2014(95):5-14.
16. Sanson A, Johnstone R. The LSAC Research Consortium & FaCS LSAC Project Team. Growing Up in Australia takes its first steps. *Family Matters* 2004;67:46-53.

17. Wake M, Clifford SA, York E, et al. Introducing Growing Up in Australia's Child Health CheckPoint. *Family Matters* 2014;94:15-23.
18. Clifford SA, Davies S, Wake M. Child Health CheckPoint: Cohort summary and methodology of a physical health and biospecimen module for the Longitudinal Study of Australian Children. Submitted to BMJ Open October 2017.
19. Soininen P, Kangas AJ, Wurtz P, et al. High-throughput serum NMR metabolomics for cost-effective holistic studies on systemic metabolism. *Analyst* 2009;134(9):1781-5. doi: 10.1039/b910205a
20. Wurtz P, Kangas AJ, Soininen P, et al. Quantitative Serum NMR Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technology. *Am J Epidemiol* 2017 [published Online First: 10 May 2017]
21. Australian Bureau of S. Census of population and housing: Socio-Economic Indexes for Areas (SEIFA) 2011. Cat. no. 2033.0.55.001, 2011.
22. Davies S, Clifford S, Gillespie A, et al. LSAC's Child Health CheckPoint Data Issues Paper 2018. *Melbourne: Murdoch Children's Research Institute* 2018 doi: 10.25374/MCRI.5821230.
23. Heeringa SG, West BT, Berglund PA. Applied survey data analysis. Boca Raton.: CRC press 2010.
24. Ellul S, Hiscock R, Mensah FK, et al. Longitudinal Study of Australian Children's Child Health CheckPoint Technical Paper 1: Weighting and Non-Response. *Melbourne: Murdoch Children's Research Institute* 2018 doi: <https://doi.org/10.25374/MCRI.5687593>
25. R: A language and environment for statistical computing [program]. Vienna, Austria: R Foundation for Statistical Computing, 2018.
26. Menni C, Kastenmüller G, Petersen AK, et al. Metabolomic markers reveal novel pathways of ageing and early development in human populations. *International Journal of Epidemiology* 2013;42(4):1111-19. doi: 10.1093/ije/dyt094
27. Dunn WB, Lin W, Broadhurst D, et al. Molecular phenotyping of a UK population: defining the human serum metabolome. *Metabolomics* 2015;11:9-26. doi: 10.1007/s11306-014-0707-1
28. Mittelstrass K, Ried JS, Yu Z, et al. Discovery of sexual dimorphisms in metabolic and genetic biomarkers. *PLoS Genet* 2011;7(8):e1002215. doi: 10.1371/journal.pgen.1002215
29. Kolz M, Johnson T, Sanna S, et al. Meta-analysis of 28,141 individuals identifies common variants within five new loci that influence uric acid concentrations. *PLoS Genet* 2009;5(6):e1000504. doi: 10.1371/journal.pgen.1000504
30. Krumsiek J, Mittelstrass K, Do KT, et al. Gender-specific pathway differences in the human serum metabolome. *Metabolomics* 2015;11(6):1815-33. doi: 10.1007/s11306-015-0829-0
31. Guasch-Ferré M, Hruby A, Toledo E, et al. Metabolomics in Prediabetes and Diabetes: A Systematic Review and Meta-analysis. *Diabetes Care* 2016;39(5):833-46. doi: 10.2337/dc15-2251
32. Lynch CJ, Adams SH. Branched-chain amino acids in metabolic signalling and insulin resistance. *Nature reviews Endocrinology* 2014;10(12):723-36. doi: 10.1038/nrendo.2014.171
33. Chang AY, Lalia AZ, Jenkins GD, et al. Combining a nontargeted and targeted metabolomics approach to identify metabolic pathways significantly altered in

- 1 polycystic ovary syndrome. *Metabolism* 2017;71(Supplement C):52-63. doi:
2 <https://doi.org/10.1016/j.metabol.2017.03.002>
- 3 34. Hartiala JA, Wilson Tang WH, Wang Z, et al. Genome-wide association study and
4 targeted metabolomics identifies sex-specific association of CPS1 with coronary
5 artery disease. *Nature Communications* 2016;7:10558. doi: 10.1038/ncomms10558
- 6 35. Davis CE, Williams DH, Oganov RG, et al. Sex Difference in High Density Lipoprotein
7 Cholesterol in Six Countries. *American Journal of Epidemiology* 1996;143(11):1100-
8 06. doi: 10.1093/oxfordjournals.aje.a008686
- 9 36. Spinneker A, Egert S, Gonzalez-Gross M, et al. Lipid, lipoprotein and apolipoprotein
10 profiles in European adolescents and its associations with gender, biological maturity
11 and body fat--the HELENA Study. *Eur J Clin Nutr* 2012;66(6):727-35. doi:
12 10.1038/ejcn.2011.222
- 13 37. Walldius G, Jungner I, Aastveit AH, et al. The apoB/apoA-I ratio is better than the
14 cholesterol ratios to estimate the balance between plasma proatherogenic and
15 antiatherogenic lipoproteins and to predict coronary risk. *Clin Chem Lab Med*
16 2004;42(12):1355-63. doi: 10.1515/CCLM.2004.254
- 17 38. Walldius G, Jungner I. The apoB/apoA-I ratio: a strong, new risk factor for cardiovascular
18 disease and a target for lipid-lowering therapy--a review of the evidence. *J Intern*
19 *Med* 2006;259(5):493-519. doi: 10.1111/j.1365-2796.2006.01643.x
- 20 39. Mozaffarian D, Benjamin EJ, Go AS, et al. Heart disease and stroke statistics--2015
21 update: a report from the American Heart Association. *Circulation* 2015;131(4):e29-
22 322. doi: 10.1161/CIR.0000000000000152
- 23 40. Upadhyay RK. Emerging Risk Biomarkers in Cardiovascular Diseases and Disorders.
24 *Journal of Lipids* 2015;2015:971453. doi: 10.1155/2015/971453
- 25 41. Hardy R, Lawlor DA, Kuh D. A life course approach to cardiovascular aging. *Future*
26 *cardiology* 2015;11(1):101-13. doi: 10.2217/fca.14.67
- 27 42. Jolliffe CJ, Janssen I. Distribution of Lipoproteins by Age and Gender in Adolescents.
28 *Circulation* 2006;114(10):1056.
- 29 43. Ohlund I, Hernell O, Hornell A, et al. Serum lipid and apolipoprotein levels in 4-year-old
30 children are associated with parental levels and track over time. *Eur J Clin Nutr*
31 2011;65(4):463-69.
- 32 44. Harris PA, Taylor R, Thielke R, et al. Research electronic data capture (REDCap)—A
33 metadata-driven methodology and workflow process for providing translational
34 research informatics support. *Journal of Biomedical Informatics* 2009;42(2):377-81.
35 doi: <https://doi.org/10.1016/j.jbi.2008.08.010>
- 36

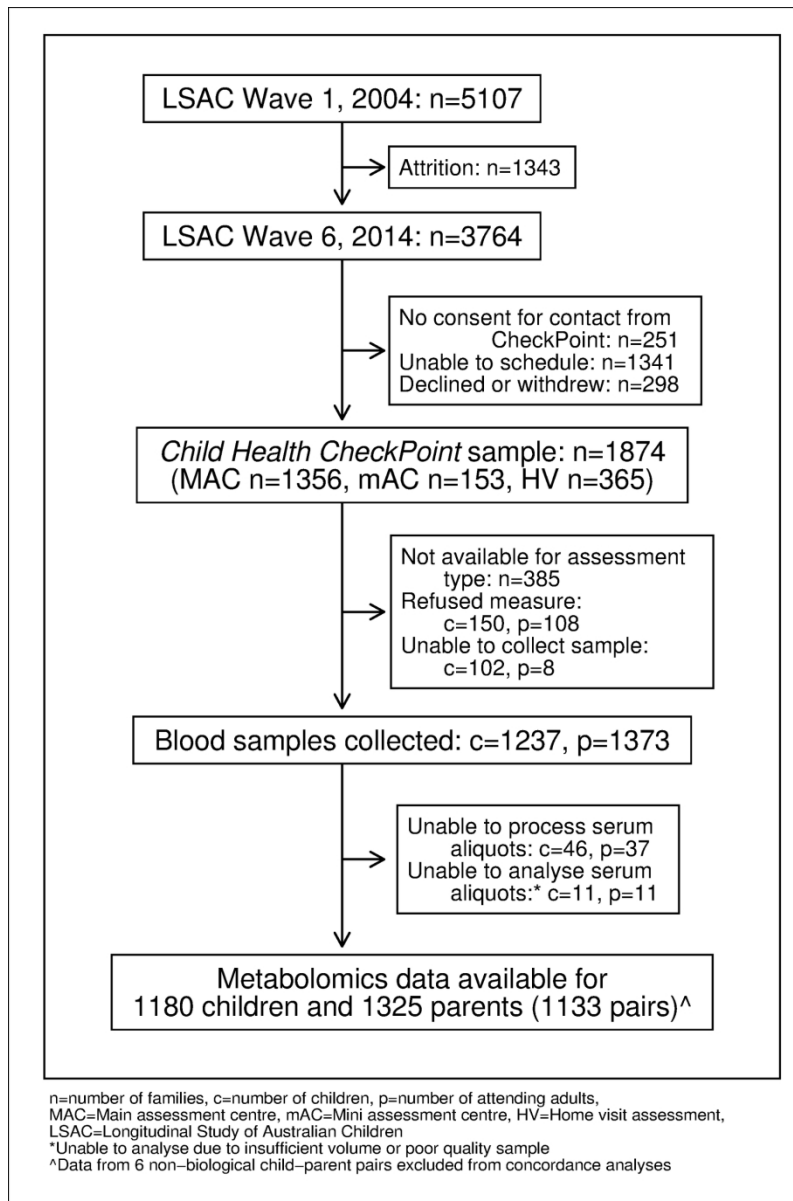


Figure 1: Participant flow chart.

n=number of families, c=number of children, p=number of attending adults,
MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,
LSAC=Longitudinal Study of Australian Children

*Unable to analyse due to insufficient volume or poor quality sample

^Data from 6 non-biological child-parent pairs excluded from concordance analyses

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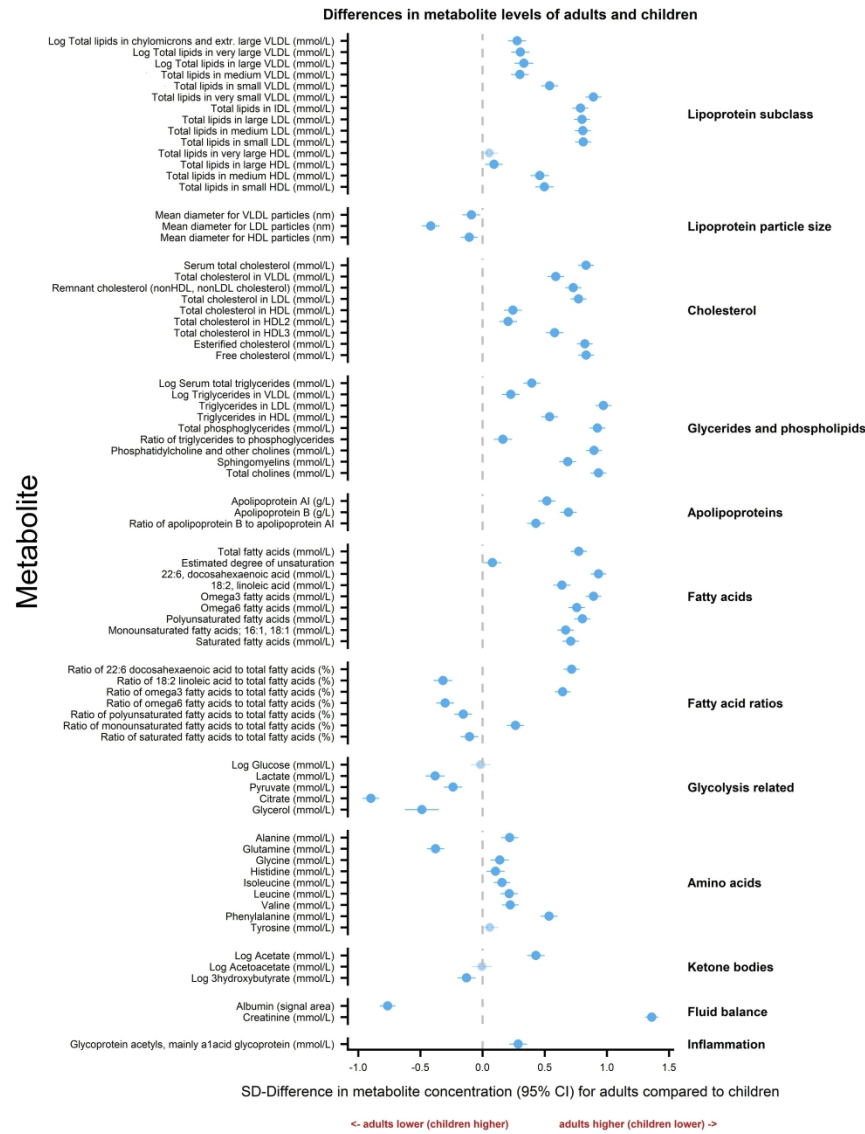


Figure 2: Differences in metabolite levels between children and adults.

Association measures are SD difference in metabolite concentration for adults compared to children. Error bars represent 95% confidence intervals. Significant associations after p-values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals and associated p-values are listed in supplementary table 2. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

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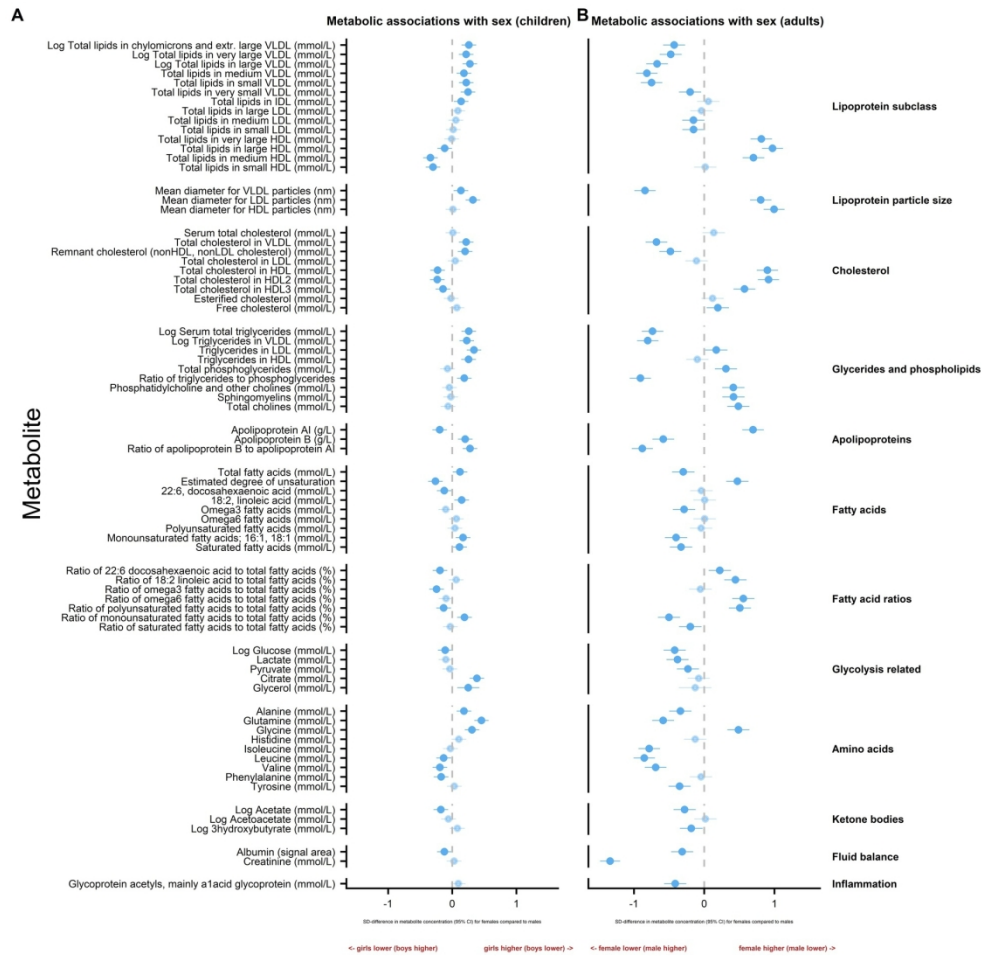


Figure 3: Sex-specific differences in metabolite levels in childhood and adulthood.

Association measures are SD difference in metabolite concentration for females compared to males in children (A) and adults (B). Error bars represent 95% confidence intervals. Significant associations after p-values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals and associated p-values are listed in supplementary table 3 and 4. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

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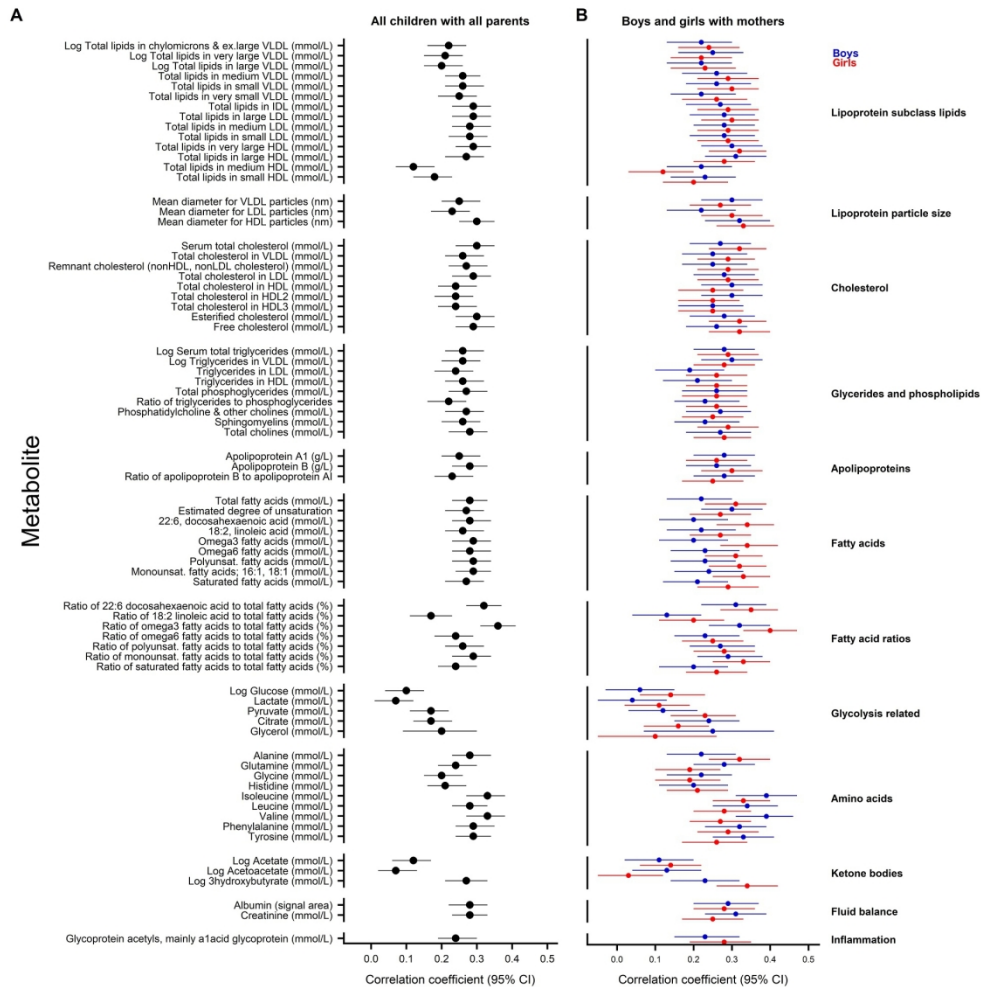


Figure 4: Parent:child correlation for metabolite measures. Pearson’s correlation coefficients for all children with all parents (A); and for boys (blue) with mothers and for girls (red) with mothers (B). Error bars represent 95% confidence intervals. Correlation coefficients with associated 95% confidence intervals are listed in supplementary table 5 and 6. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

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Supplementary table 1: Weighted mean (SD)* of metabolite measures in children and parents.

Metabolic subgroup	Children									Adults								
	Male			Female			All			Male			Female			All		
	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	575	0.005	4.826	605	0.011	3.260	1180	0.007	4.054	177	0.040	2.101	1148	0.013	2.822	1325	0.015	2.777
Total lipids in very large VLDL (mmol/L)*	575	0.006	6.246	605	0.014	4.800	1180	0.010	5.521	177	0.087	2.419	1148	0.022	3.458	1325	0.027	3.398
Total lipids in large VLDL (mmol/L)*	575	0.059	3.945	605	0.121	2.488	1180	0.085	3.234	177	0.423	1.287	1148	0.161	1.666	1325	0.182	1.665
Total lipids in medium VLDL (mmol/L)	575	0.441	0.270	605	0.478	0.272	1180	0.460	0.271	177	0.959	0.630	1148	0.548	0.366	1325	0.602	0.432
Total lipids in small VLDL (mmol/L)	575	0.381	0.153	605	0.405	0.146	1180	0.393	0.149	177	0.690	0.279	1148	0.494	0.216	1325	0.520	0.234
Total lipids in very small VLDL (mmol/L)	575	0.325	0.070	605	0.342	0.076	1180	0.334	0.073	177	0.451	0.111	1148	0.426	0.110	1325	0.429	0.110
Total lipids in IDL (mmol/L)	575	0.804	0.176	605	0.834	0.183	1180	0.819	0.180	177	0.985	0.261	1148	0.999	0.240	1325	0.997	0.242
Total lipids in large LDL (mmol/L)	575	0.917	0.220	605	0.941	0.228	1180	0.929	0.224	177	1.162	0.327	1148	1.155	0.298	1325	1.156	0.301
Total lipids in medium LDL (mmol/L)	575	0.511	0.136	605	0.519	0.140	1180	0.515	0.138	177	0.676	0.214	1148	0.655	0.185	1325	0.658	0.189
Total lipids in small LDL (mmol/L)	575	0.338	0.083	605	0.340	0.087	1180	0.339	0.085	177	0.439	0.135	1148	0.425	0.114	1325	0.427	0.117
Total lipids in very large HDL (mmol/L)	575	0.482	0.196	605	0.495	0.184	1180	0.488	0.189	177	0.320	0.189	1148	0.497	0.229	1325	0.474	0.232
Total lipids in large HDL (mmol/L)	575	0.874	0.291	605	0.859	0.275	1180	0.866	0.282	177	0.509	0.335	1148	0.900	0.382	1325	0.849	0.399
Total lipids in medium HDL (mmol/L)	575	0.917	0.127	605	0.871	0.126	1180	0.894	0.128	177	0.828	0.241	1148	0.971	0.175	1325	0.952	0.191
Total lipids in small HDL (mmol/L)	575	1.039	0.103	605	0.997	0.115	1180	1.018	0.111	177	1.055	0.254	1148	1.085	0.138	1325	1.081	0.157
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	575	37.063	1.633	605	37.238	1.557	1180	37.152	1.591	177	38.527	1.737	1148	36.943	1.599	1325	37.152	1.701
Mean diameter for LDL particles (nm)	575	23.587	0.103	605	23.628	0.109	1180	23.608	0.107	177	23.487	0.093	1148	23.573	0.100	1325	23.562	0.104
Mean diameter for HDL particles (nm)	575	10.081	0.233	605	10.102	0.221	1180	10.092	0.226	177	9.798	0.244	1148	10.068	0.262	1325	10.032	0.275
Cholesterol																		
Serum total cholesterol (mmol/L)	575	3.576	0.620	605	3.596	0.643	1180	3.586	0.629	177	4.161	0.885	1148	4.234	0.828	1325	4.225	0.835
Total cholesterol in VLDL (mmol/L)	575	0.438	0.188	605	0.472	0.189	1180	0.455	0.189	177	0.826	0.395	1148	0.592	0.265	1325	0.623	0.295
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	575	0.944	0.269	605	0.994	0.271	1180	0.970	0.270	177	1.452	0.472	1148	1.219	0.383	1325	1.250	0.402
Total cholesterol in LDL (mmol/L)	575	1.130	0.330	605	1.149	0.341	1180	1.139	0.334	177	1.492	0.498	1148	1.460	0.442	1325	1.464	0.449
Total cholesterol in HDL (mmol/L)	575	1.503	0.274	605	1.453	0.266	1180	1.477	0.270	177	1.217	0.350	1148	1.556	0.363	1325	1.511	0.379
Total cholesterol in HDL2 (mmol/L)	575	1.035	0.254	605	0.988	0.246	1180	1.011	0.250	177	0.751	0.327	1148	1.072	0.335	1325	1.030	0.351
Total cholesterol in HDL3 (mmol/L)	575	0.468	0.024	605	0.466	0.024	1180	0.467	0.024	177	0.466	0.035	1148	0.483	0.033	1325	0.481	0.034
Esterified cholesterol (mmol/L)	572	2.516	0.447	604	2.517	0.460	1176	2.517	0.452	176	2.941	0.636	1147	2.975	0.593	1323	2.971	0.597
Free cholesterol (mmol/L)	572	1.062	0.179	604	1.079	0.186	1176	1.070	0.182	176	1.211	0.273	1147	1.260	0.239	1323	1.253	0.244
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	575	0.918	0.709	605	1.005	0.681	1180	0.962	0.696	177	1.686	0.809	1148	1.129	0.755	1325	1.190	0.782
Triglycerides in VLDL (mmol/L)*	575	0.582	0.885	605	0.648	0.830	1180	0.615	0.858	177	1.249	0.927	1148	0.694	0.945	1325	0.750	0.972
Triglycerides in LDL (mmol/L)	575	0.113	0.024	605	0.123	0.027	1180	0.118	0.026	177	0.153	0.040	1148	0.158	0.044	1325	0.157	0.043
Triglycerides in HDL (mmol/L)	575	0.129	0.030	605	0.136	0.030	1180	0.133	0.030	177	0.161	0.047	1148	0.151	0.040	1325	0.153	0.041
Total phosphoglycerides (mmol/L)	572	1.632	0.240	604	1.620	0.261	1176	1.626	0.250	176	1.865	0.354	1147	1.926	0.340	1323	1.918	0.342
Ratio of triglycerides to phosphoglycerides	572	0.526	0.253	604	0.569	0.277	1176	0.548	0.265	176	0.946	0.587	1147	0.580	0.279	1323	0.628	0.355
Phosphatidylcholine & other cholines (mmol/L)	572	1.691	0.240	604	1.687	0.267	1176	1.689	0.253	175	1.877	0.320	1147	1.978	0.336	1322	1.965	0.335
Sphingomyelins (mmol/L)	572	0.348	0.061	604	0.349	0.064	1176	0.348	0.062	175	0.370	0.070	1147	0.397	0.078	1322	0.394	0.077
Total cholines (mmol/L)	572	2.005	0.256	604	1.997	0.264	1176	2.001	0.259	175	2.185	0.334	1147	2.317	0.351	1322	2.299	0.351
Apolipoproteins																		
Apolipoprotein A1 (g/L)	575	1.509	0.159	605	1.484	0.151	1180	1.497	0.155	177	1.461	0.178	1148	1.589	0.205	1325	1.572	0.206
Apolipoprotein B (g/L)	575	0.682	0.135	604	0.706	0.136	1179	0.694	0.135	177	0.955	0.245	1148	0.812	0.196	1325	0.831	0.208
Ratio of apolipoprotein B to apolipoprotein A	575	0.455	0.097	604	0.479	0.098	1179	0.467	0.098	177	0.660	0.178	1148	0.518	0.136	1325	0.537	0.150
Fatty acids																		
Total fatty acids (mmol/L)	570	9.215	1.697	604	9.370	1.730	1174	9.294	1.709	173	11.850	2.723	1145	10.917	2.392	1318	11.034	2.446
Estimated degree of unsaturation	570	1.212	0.056	604	1.196	0.065	1174	1.204	0.061	173	1.179	0.070	1145	1.212	0.066	1318	1.208	0.068
22:6, docosahexaenoic acid (mmol/L)	570	0.078	0.028	604	0.074	0.029	1174	0.076	0.028	173	0.118	0.051	1145	0.111	0.039	1318	0.112	0.041
18:2, linoleic acid (mmol/L)	570	2.539	0.456	604	2.592	0.464	1174	2.566	0.459	173	2.919	0.567	1145	2.880	0.584	1318	2.885	0.580
Omega3 fatty acids (mmol/L)	570	0.309	0.086	604	0.296	0.083	1174	0.302	0.085	173	0.451	0.160	1145	0.400	0.117	1318	0.406	0.124
Omega6 fatty acids (mmol/L)	570	3.077	0.493	604	3.094	0.492	1174	3.086	0.491	173	3.553	0.648	1145	3.507	0.627	1318	3.513	0.628
Polyunsat. fatty acids (mmol/L)	570	3.386	0.565	604	3.390	0.561	1174	3.388	0.560	173	4.004	0.775	1145	3.907	0.721	1318	3.919	0.726
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	570	2.500	0.632	604	2.587	0.661	1174	2.544	0.646	173	3.520	1.058	1145	3.080	0.917	1318	3.135	0.943
Saturated fatty acids (mmol/L)	570	3.328	0.642	604	3.393	0.682	1174	3.362	0.661	173	4.325	1.088	1145	3.930	0.932	1318	3.979	0.958

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Fatty acid ratios

Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	570	0.841	0.239	604	0.790	0.269	1174	0.815	0.255	173	0.984	0.315	1145	1.023	0.284	1318	1.018	0.287
Ratio of 18:2 linoleic acid to total fatty acids (%)	570	27.741	3.127	604	27.867	3.274	1174	27.805	3.191	173	25.038	3.570	1145	26.656	3.417	1318	26.453	3.467
Ratio of omega3 fatty acids to total fatty acids (%)	570	3.329	0.579	604	3.150	0.600	1174	3.238	0.594	173	3.763	0.799	1145	3.659	0.697	1318	3.672	0.709
Ratio of omega6 fatty acids to total fatty acids (%)	570	33.650	3.047	604	33.308	3.228	1174	33.475	3.133	173	30.468	3.757	1145	32.502	3.311	1318	32.247	3.424
Ratio of polyunsat. fatty acids to total fatty acids (%)	570	36.979	3.235	604	36.459	3.514	1174	36.713	3.377	173	34.231	3.878	1145	36.161	3.588	1318	35.918	3.669
Ratio of monounsat. fatty acids to total fatty acids (%)	570	26.911	2.569	604	27.375	2.678	1174	27.148	2.625	173	29.366	2.968	1145	27.889	2.842	1318	28.075	2.891
Ratio of saturated fatty acids to total fatty acids (%)	570	36.110	1.675	604	36.167	1.802	1174	36.139	1.734	173	36.402	2.027	1145	35.950	2.032	1318	36.007	2.031

Glycolysis related

Glucose (mmol/L)*	574	1.350	0.115	605	1.336	0.105	1179	1.342	0.11	176	1.415	0.205	1148	1.334	0.176	1324	1.344	0.182
Lactate (mmol/L)	575	1.770	0.459	605	1.718	0.434	1180	1.743	0.446	177	1.696	0.472	1148	1.562	0.480	1325	1.580	0.480
Pyruvate (mmol/L)	574	0.100	0.024	605	0.098	0.023	1179	0.099	0.023	177	0.101	0.031	1147	0.093	0.034	1324	0.094	0.033
Citrate (mmol/L)	575	0.125	0.017	604	0.131	0.018	1179	0.128	0.018	177	0.110	0.016	1148	0.111	0.016	1325	0.111	0.016
Glycerol (mmol/L)#	240	0.078	0.021	283	0.083	0.022	523	0.081	0.021	84	0.073	0.021	470	0.071	0.023	554	0.071	0.023

Amino acids

Alanine (mmol/L)	575	0.387	0.061	605	0.396	0.060	1180	0.391	0.060	176	0.423	0.065	1147	0.399	0.060	1323	0.402	0.061
Glutamine (mmol/L)	575	0.474	0.050	605	0.497	0.051	1180	0.485	0.051	177	0.490	0.063	1148	0.456	0.066	1325	0.461	0.066
Glycine (mmol/L)	574	0.261	0.032	604	0.270	0.034	1178	0.265	0.033	176	0.243	0.029	1148	0.274	0.061	1324	0.270	0.059
Histidine (mmol/L)	574	0.065	0.009	605	0.065	0.008	1179	0.065	0.008	176	0.066	0.008	1148	0.065	0.009	1324	0.065	0.009
Isoleucine (mmol/L)	574	0.054	0.019	605	0.053	0.019	1179	0.054	0.019	174	0.072	0.021	1146	0.055	0.020	1320	0.057	0.021
Leucine (mmol/L)	575	0.073	0.019	605	0.071	0.019	1180	0.072	0.019	177	0.097	0.029	1148	0.074	0.021	1325	0.077	0.023
Valine (mmol/L)	575	0.162	0.037	604	0.156	0.035	1179	0.159	0.036	177	0.192	0.036	1147	0.162	0.042	1324	0.166	0.042
Phenylalanine (mmol/L)	575	0.068	0.009	605	0.066	0.009	1180	0.067	0.009	177	0.073	0.011	1148	0.072	0.011	1325	0.073	0.011
Tyrosine (mmol/L)	574	0.054	0.014	605	0.055	0.014	1179	0.055	0.014	176	0.060	0.013	1148	0.054	0.015	1324	0.055	0.015

Ketone bodies

Acetate (mmol/L)*	575	0.031	0.423	605	0.030	0.404	1180	0.030	0.413	177	0.037	0.655	1146	0.033	0.600	1323	0.034	0.609
Acetoacetate (mmol/L)*	575	0.025	1.310	605	0.023	1.429	1180	0.024	1.367	177	0.023	2.116	1147	0.024	1.278	1324	0.024	1.403
3hydroxybutyrate (mmol/L)*#	555	0.100	0.786	580	0.103	0.826	1135	0.101	0.805	170	0.104	0.669	1098	0.096	0.781	1268	0.097	0.769

Fluid balance

Albumin (signal area)	574	0.093	0.005	605	0.092	0.005	1179	0.093	0.005	177	0.090	0.005	1148	0.088	0.005	1325	0.089	0.005
Creatinine (mmol/L)	570	0.040	0.006	600	0.040	0.006	1170	0.040	0.006	173	0.066	0.015	1139	0.054	0.009	1312	0.055	0.010

Inflammation

Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	575	1.170	0.191	605	1.173	0.186	1180	1.172	0.188	177	1.375	0.366	1148	1.242	0.233	1325	1.260	0.258
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* geometric mean [relative SD] when skewed variable

Note: The presence of ethanol in a sample can affect quantification of glycerol and on some occasions 3hydroxybutyrate. Ethanol can be introduced in to a sample from disinfectants used during blood collection/processing of sample.

Supplementary table 2: Mean difference in metabolite levels in adults compared to children in absolute concentration unit

Metabolic subgroup	Differences by age (Adults - Child)				Conversion factor (SD) #
	Estimate	95% CI	P-value	Adj_p-value^	
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.704	(0.519, 0.890)	<0.001	<0.001	2.534
Total lipids in very large VLDL (mmol/L)*	0.922	(0.700, 1.145)	<0.001	<0.001	3.031
Total lipids in large VLDL (mmol/L)*	0.648	(0.502, 0.795)	<0.001	<0.001	1.950
Total lipids in medium VLDL (mmol/L)	0.105	(0.080, 0.129)	<0.001	<0.001	0.348
Total lipids in small VLDL (mmol/L)	0.107	(0.094, 0.121)	<0.001	<0.001	0.199
Total lipids in very small VLDL (mmol/L)	0.093	(0.086, 0.099)	<0.001	<0.001	0.104
Total lipids in IDL (mmol/L)	0.181	(0.166, 0.196)	<0.001	<0.001	0.230
Total lipids in large LDL (mmol/L)	0.229	(0.211, 0.247)	<0.001	<0.001	0.286
Total lipids in medium LDL (mmol/L)	0.144	(0.132, 0.155)	<0.001	<0.001	0.178
Total lipids in small LDL (mmol/L)	0.089	(0.082, 0.096)	<0.001	<0.001	0.110
Total lipids in very large HDL (mmol/L)	0.012	(-0.003, 0.027)	0.128	0.132	0.217
Total lipids in large HDL (mmol/L)	0.032	(0.007, 0.057)	0.011	0.012	0.353
Total lipids in medium HDL (mmol/L)	0.076	(0.064, 0.089)	<0.001	<0.001	0.166
Total lipids in small HDL (mmol/L)	0.068	(0.058, 0.078)	<0.001	<0.001	0.137
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	-0.147	(-0.263, -0.031)	0.013	0.014	1.633
Mean diameter for LDL particles (nm)	-0.044	(-0.052, -0.037)	<0.001	<0.001	0.106
Mean diameter for HDL particles (nm)	-0.027	(-0.045, -0.010)	0.002	0.003	0.256
Cholesterol					
Serum total cholesterol (mmol/L)	0.670	(0.619, 0.721)	<0.001	<0.001	0.805
Total cholesterol in VLDL (mmol/L)	0.146	(0.129, 0.163)	<0.001	<0.001	0.249
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.261	(0.237, 0.284)	<0.001	<0.001	0.357
Total cholesterol in LDL (mmol/L)	0.327	(0.300, 0.354)	<0.001	<0.001	0.424
Total cholesterol in HDL (mmol/L)	0.082	(0.058, 0.106)	<0.001	<0.001	0.337
Total cholesterol in HDL2 (mmol/L)	0.064	(0.042, 0.086)	<0.001	<0.001	0.311
Total cholesterol in HDL3 (mmol/L)	0.018	(0.016, 0.020)	<0.001	<0.001	0.031
Esterified cholesterol (mmol/L)	0.474	(0.437, 0.511)	<0.001	<0.001	0.576
Free cholesterol (mmol/L)	0.195	(0.180, 0.210)	<0.001	<0.001	0.234
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	0.176	(0.145, 0.206)	<0.001	<0.001	0.443
Triglycerides in VLDL (mmol/L)*	0.140	(0.096, 0.183)	<0.001	<0.001	0.615
Triglycerides in LDL (mmol/L)	0.040	(0.038, 0.043)	<0.001	<0.001	0.042
Triglycerides in HDL (mmol/L)	0.020	(0.017, 0.022)	<0.001	<0.001	0.037
Total phosphoglycerides (mmol/L)	0.311	(0.290, 0.332)	<0.001	<0.001	0.337
Ratio of triglycerides to phosphoglycerides	0.049	(0.027, 0.071)	<0.001	<0.001	0.299
Phosphatidylcholine & other cholines (mmol/L)	0.295	(0.274, 0.316)	<0.001	<0.001	0.329
Sphingomyelins (mmol/L)	0.052	(0.047, 0.057)	<0.001	<0.001	0.075
Total cholines (mmol/L)	0.323	(0.302, 0.345)	<0.001	<0.001	0.347
Apolipoproteins					
Apolipoprotein A1 (g/L)	0.099	(0.086, 0.112)	<0.001	<0.001	0.191
Apolipoprotein B (g/L)	0.125	(0.113, 0.137)	<0.001	<0.001	0.182
Ratio of apolipoprotein B to apolipoprotein A1	0.055	(0.046, 0.064)	<0.001	<0.001	0.127
Fatty acids					
Total fatty acids (mmol/L)	1.738	(1.592, 1.885)	<0.001	<0.001	2.245
Estimated degree of unsaturation	0.005	(0.000, 0.009)	0.030	0.031	0.063
22:6, docosahexaenoic acid (mmol/L)	0.037	(0.035, 0.040)	<0.001	<0.001	0.040
18:2, linoleic acid (mmol/L)	0.347	(0.310, 0.384)	<0.001	<0.001	0.545
Omega3 fatty acids (mmol/L)	0.105	(0.098, 0.113)	<0.001	<0.001	0.118
Omega6 fatty acids (mmol/L)	0.453	(0.414, 0.492)	<0.001	<0.001	0.597
Polyunsat. fatty acids (mmol/L)	0.558	(0.513, 0.603)	<0.001	<0.001	0.695
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.568	(0.512, 0.625)	<0.001	<0.001	0.850
Saturated fatty acids (mmol/L)	0.612	(0.554, 0.669)	<0.001	<0.001	0.862

Fatty acid ratios

Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.209	(0.190, 0.228)	<0.001	<0.001	0.292
Ratio of 18:2 linoleic acid to total fatty acids (%)	-1.079	(-1.331, -0.827)	<0.001	<0.001	3.387
Ratio of omega3 fatty acids to total fatty acids (%)	0.446	(0.403, 0.490)	<0.001	<0.001	0.693
Ratio of omega6 fatty acids to total fatty acids (%)	-0.992	(-1.227, -0.757)	<0.001	<0.001	3.296
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.546	(-0.794, -0.298)	<0.001	<0.001	3.507
Ratio of monounsat. fatty acids to total fatty acids (%)	0.741	(0.547, 0.934)	<0.001	<0.001	2.797
Ratio of saturated fatty acids to total fatty acids (%)	-0.195	(-0.328, -0.062)	0.004	0.005	1.863

Glycolysis related

Glucose (mmol/L)*	-0.002	(-0.014, 0.009)	0.700	0.709	0.147
Lactate (mmol/L)	-0.180	(-0.215, -0.144)	<0.001	<0.001	0.471
Pyruvate (mmol/L)	-0.007	(-0.009, -0.005)	<0.001	<0.001	0.029
Citrate (mmol/L)	-0.017	(-0.018, -0.016)	<0.001	<0.001	0.019
Glycerol (mmol/L)	-0.011	(-0.015, -0.008)	<0.001	<0.001	0.023

Amino acids

Alanine (mmol/L)	0.013	(0.009, 0.017)	<0.001	<0.001	0.060
Glutamine (mmol/L)	-0.023	(-0.027, -0.019)	<0.001	<0.001	0.060
Glycine (mmol/L)	0.007	(0.003, 0.010)	<0.001	<0.001	0.049
Histidine (mmol/L)	0.001	(0.000, 0.002)	0.005	0.006	0.009
Isoleucine (mmol/L)	0.003	(0.002, 0.004)	<0.001	<0.001	0.019
Leucine (mmol/L)	0.004	(0.003, 0.006)	<0.001	<0.001	0.021
Valine (mmol/L)	0.009	(0.006, 0.011)	<0.001	<0.001	0.039
Phenylalanine (mmol/L)	0.005	(0.005, 0.006)	<0.001	<0.001	0.010
Tyrosine (mmol/L)	0.001	(-0.000, 0.002)	0.100	0.105	0.014

Ketone bodies

Acetate (mmol/L)*	0.101	(0.084, 0.117)	<0.001	<0.001	0.235
Acetoacetate (mmol/L)*	-0.004	(-0.086, 0.078)	0.922	0.922	1.022
3hydroxybutyrate (mmol/L)*	-0.064	(-0.100, -0.028)	0.001	0.001	0.493

Fluid balance

Albumin (signal area)	-0.004	(-0.004, -0.004)	<0.001	<0.001	0.005
Creatinine (mmol/L)	0.016	(0.015, 0.017)	<0.001	0.001	0.012

Inflammation

Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.062	(0.047, 0.078)	<0.001	<0.001	0.217
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* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations in Figure 2 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor. Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 3: Differences in mean metabolite levels in girls compared to boys in absolute concentration units.

Metabolic subgroup	Differences for children (Female - Male)				
	Estimate	95% CI	pvalue	Adj_p-value [^]	Conversion factor (SD) #
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.737	(0.414, 1.059)	<0.001	<0.001	2.845
Total lipids in very large VLDL (mmol/L)*	0.744	(0.355, 1.134)	<0.001	0.001	3.428
Total lipids in large VLDL (mmol/L)*	0.663	(0.390, 0.936)	<0.001	<0.001	2.411
Total lipids in medium VLDL (mmol/L)	0.049	(0.018, 0.080)	0.002	0.004	0.269
Total lipids in small VLDL (mmol/L)	0.032	(0.015, 0.048)	<0.001	0.001	0.146
Total lipids in very small VLDL (mmol/L)	0.018	(0.010, 0.027)	<0.001	<0.001	0.074
Total lipids in IDL (mmol/L)	0.025	(0.004, 0.046)	0.017	0.035	0.182
Total lipids in large LDL (mmol/L)	0.020	(-0.006, 0.046)	0.132	0.187	0.227
Total lipids in medium LDL (mmol/L)	0.008	(-0.008, 0.024)	0.338	0.416	0.139
Total lipids in small LDL (mmol/L)	0.001	(-0.008, 0.011)	0.788	0.822	0.086
Total lipids in very large HDL (mmol/L)	-0.002	(-0.023, 0.020)	0.882	0.882	0.190
Total lipids in large HDL (mmol/L)	-0.033	(-0.066, -0.001)	0.044	0.074	0.283
Total lipids in medium HDL (mmol/L)	-0.045	(-0.059, -0.030)	<0.001	<0.001	0.131
Total lipids in small HDL (mmol/L)	-0.035	(-0.048, -0.022)	<0.001	<0.001	0.116
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	0.215	(0.035, 0.395)	0.019	0.038	1.580
Mean diameter for LDL particles (nm)	0.035	(0.023, 0.047)	<0.001	0.000	0.108
Mean diameter for HDL particles (nm)	0.003	(-0.023, 0.028)	0.847	0.870	0.226
Cholesterol					
Serum total cholesterol (mmol/L)	0.007	(-0.066, 0.079)	0.857	0.869	0.634
Total cholesterol in VLDL (mmol/L)	0.040	(0.019, 0.061)	<0.001	0.001	0.184
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.053	(0.023, 0.083)	0.001	0.002	0.265
Total cholesterol in LDL (mmol/L)	0.016	(-0.023, 0.054)	0.427	0.518	0.336
Total cholesterol in HDL (mmol/L)	-0.062	(-0.093, -0.031)	<0.001	<0.001	0.273
Total cholesterol in HDL2 (mmol/L)	-0.059	(-0.087, -0.030)	<0.001	<0.001	0.253
Total cholesterol in HDL3 (mmol/L)	-0.003	(-0.006, -0.001)	0.013	0.027	0.024
Esterified cholesterol (mmol/L)	-0.008	(-0.060, 0.044)	0.755	0.798	0.455
Free cholesterol (mmol/L)	0.013	(-0.008, 0.034)	0.211	0.284	0.184
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	0.101	(0.056, 0.145)	<0.001	<0.001	0.390
Triglycerides in VLDL (mmol/L)*	0.125	(0.062, 0.187)	<0.001	<0.001	0.551
Triglycerides in LDL (mmol/L)	0.009	(0.006, 0.012)	<0.001	<0.001	0.026
Triglycerides in HDL (mmol/L)	0.007	(0.004, 0.011)	<0.001	<0.001	0.029
Total phosphoglycerides (mmol/L)	-0.018	(-0.047, 0.010)	0.206	0.282	0.249
Ratio of triglycerides to phosphoglycerides	0.051	(0.020, 0.083)	0.001	0.003	0.274
Phosphatidylcholine & other cholines (mmol/L)	-0.011	(-0.040, 0.018)	0.447	0.534	0.251
Sphingomyelins (mmol/L)	-0.001	(-0.009, 0.006)	0.706	0.757	0.063
Total cholines (mmol/L)	-0.017	(-0.046, 0.013)	0.268	0.354	0.257
Apolipoproteins					
Apolipoprotein A1 (g/L)	-0.030	(-0.048, -0.013)	0.001	0.002	0.155
Apolipoprotein B (g/L)	0.027	(0.012, 0.042)	0.001	0.002	0.133
Ratio of apolipoprotein B to apolipoprotein A1	0.027	(0.016, 0.038)	<0.001	<0.001	0.098
Fatty acids					
Total fatty acids (mmol/L)	0.200	(0.011, 0.389)	0.038	0.065	1.650
Estimated degree of unsaturation	-0.016	(-0.022, -0.009)	<0.001	<0.001	0.060
22:6, docosahexaenoic acid (mmol/L)	-0.004	(-0.007, -0.000)	0.033	0.059	0.028
18:2, linoleic acid (mmol/L)	0.068	(0.015, 0.120)	0.011	0.024	0.458
Omega3 fatty acids (mmol/L)	-0.008	(-0.018, 0.001)	0.083	0.128	0.084
Omega6 fatty acids (mmol/L)	0.031	(-0.024, 0.087)	0.271	0.352	0.485
Polyunsat. fatty acids (mmol/L)	0.023	(-0.041, 0.086)	0.483	0.567	0.553
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.105	(0.034, 0.176)	0.004	0.009	0.623
Saturated fatty acids (mmol/L)	0.073	(-0.000, 0.146)	0.052	0.085	0.638
Fatty acid ratios					
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	-0.049	(-0.078, -0.019)	0.001	0.003	0.255
Ratio of 18:2 linoleic acid to total fatty acids (%)	0.197	(-0.173, 0.567)	0.297	0.379	3.233
Ratio of omega3 fatty acids to total fatty acids (%)	-0.145	(-0.213, -0.078)	<0.001	0.000	0.593
Ratio of omega6 fatty acids to total fatty acids (%)	-0.302	(-0.658, 0.054)	0.096	0.142	3.109
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.447	(-0.830, -0.065)	0.022	0.042	3.346
Ratio of monounsat. fatty acids to total fatty acids (%)	0.499	(0.202, 0.796)	0.001	0.003	2.606
Ratio of saturated fatty acids to total fatty acids (%)	-0.051	(-0.251, 0.148)	0.614	0.689	1.743
Glycolysis related					
Glucose (mmol/L)*	-0.013	(-0.026, 0.001)	0.061	0.098	0.118
Lactate (mmol/L)	-0.045	(-0.097, 0.007)	0.088	0.133	0.456
Pyruvate (mmol/L)	-0.001	(-0.004, 0.002)	0.524	0.606	0.024
Citrate (mmol/L)	0.007	(0.005, 0.009)	<0.001	<0.001	0.018
Glycerol (mmol/L)	0.006	(0.002, 0.010)	0.004	0.009	0.023
Amino acids					
Alanine (mmol/L)	0.011	(0.004, 0.017)	0.002	0.004	0.058
Glutamine (mmol/L)	0.023	(0.018, 0.029)	<0.001	<0.001	0.051
Glycine (mmol/L)	0.010	(0.006, 0.014)	<0.001	<0.001	0.032
Histidine (mmol/L)	0.001	(-0.000, 0.002)	0.075	0.118	0.008
Isoleucine (mmol/L)	0.000	(-0.003, 0.002)	0.637	0.693	0.018

1	Leucine (mmol/L)	-0.002	(-0.005, -0.000)	0.022	0.041	0.018
2	Valine (mmol/L)	-0.007	(-0.011, -0.003)	0.001	0.003	0.036
3	Phenylalanine (mmol/L)	-0.002	(-0.003, -0.001)	0.003	0.007	0.009
4	Tyrosine (mmol/L)	0.000	(-0.001, 0.002)	0.582	0.663	0.014
5	Ketone bodies					
6	Acetate (mmol/L)*	-0.030	(-0.048, -0.011)	0.002	0.005	0.166
7	Acetoacetate (mmol/L)*	-0.058	(-0.172, 0.055)	0.313	0.393	0.992
8	3hydroxybutyrate (mmol/L)*	0.041	(-0.019, 0.101)	0.178	0.248	0.513
9	Fluid balance					
10	Albumin (signal area)	-0.001	(-0.001, -0.000)	0.037	0.064	0.005
11	Creatinine (mmol/L)	0.000	(-0.001, 0.001)	0.624	0.690	0.007
12	Inflammation					
13	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.017	(-0.004, 0.038)	0.104	0.151	0.183

* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations for children in Figure 3 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure in children) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 4: Differences in mean metabolite levels in female compared to male adults in absolute concentration units.

Metabolic subgroup	Differences for adults (Female - Male)				
	Estimate	95% CI	pvalue	Adj_p-value^	Conversion factor (SD) #
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	-0.930	(-1.271, -0.589)	<0.001	<0.001	2.173
Total lipids in very large VLDL (mmol/L)*	-1.217	(-1.617, -0.818)	<0.001	<0.001	2.555
Total lipids in large VLDL (mmol/L)*	-0.900	(-1.107, -0.693)	<0.001	<0.001	1.343
Total lipids in medium VLDL (mmol/L)	-0.325	(-0.385, -0.264)	<0.001	<0.001	0.398
Total lipids in small VLDL (mmol/L)	-0.167	(-0.201, -0.133)	<0.001	<0.001	0.223
Total lipids in very small VLDL (mmol/L)	-0.022	(-0.039, -0.005)	0.013	0.018	0.107
Total lipids in IDL (mmol/L)	0.014	(-0.023, 0.051)	0.465	0.530	0.236
Total lipids in large LDL (mmol/L)	-0.011	(-0.058, 0.035)	0.634	0.671	0.293
Total lipids in medium LDL (mmol/L)	-0.028	(-0.057, 0.001)	0.058	0.076	0.182
Total lipids in small LDL (mmol/L)	-0.017	(-0.035, 0.001)	0.061	0.079	0.113
Total lipids in very large HDL (mmol/L)	0.195	(0.158, 0.231)	<0.001	<0.001	0.239
Total lipids in large HDL (mmol/L)	0.395	(0.334, 0.455)	<0.001	<0.001	0.405
Total lipids in medium HDL (mmol/L)	0.129	(0.101, 0.158)	<0.001	<0.001	0.184
Total lipids in small HDL (mmol/L)	0.002	(-0.021, 0.025)	0.850	0.874	0.145
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	-1.414	(-1.669, -1.159)	<0.001	<0.001	1.678
Mean diameter for LDL particles (nm)	0.081	(0.066, 0.096)	<0.001	<0.001	0.100
Mean diameter for HDL particles (nm)	0.278	(0.236, 0.320)	<0.001	<0.001	0.279
Cholesterol					
Serum total cholesterol (mmol/L)	0.112	(-0.018, 0.241)	0.091	0.116	0.817
Total cholesterol in VLDL (mmol/L)	-0.187	(-0.230, -0.145)	<0.001	<0.001	0.275
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	-0.184	(-0.244, -0.124)	<0.001	<0.001	0.383
Total cholesterol in LDL (mmol/L)	-0.048	(-0.117, 0.021)	0.175	0.213	0.437
Total cholesterol in HDL (mmol/L)	0.344	(0.286, 0.401)	<0.001	<0.001	0.382
Total cholesterol in HDL2 (mmol/L)	0.324	(0.271, 0.378)	<0.001	<0.001	0.354
Total cholesterol in HDL3 (mmol/L)	0.019	(0.014, 0.025)	<0.001	<0.001	0.034
Esterified cholesterol (mmol/L)	0.070	(-0.023, 0.163)	0.139	0.172	0.586
Free cholesterol (mmol/L)	0.046	(0.009, 0.084)	0.016	0.023	0.238
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	-0.344	(-0.416, -0.273)	<0.001	<0.001	0.468
Triglycerides in VLDL (mmol/L)*	-0.530	(-0.630, -0.429)	<0.001	<0.001	0.659
Triglycerides in LDL (mmol/L)	0.008	(0.001, 0.014)	0.033	0.044	0.044
Triglycerides in HDL (mmol/L)	-0.004	(-0.010, 0.002)	0.228	0.272	0.040
Total phosphoglycerides (mmol/L)	0.106	(0.052, 0.159)	<0.001	<0.001	0.340
Ratio of triglycerides to phosphoglycerides	-0.289	(-0.337, -0.241)	<0.001	<0.001	0.318
Phosphatidylcholine & other cholines (mmol/L)	0.138	(0.086, 0.190)	<0.001	<0.001	0.331
Sphingomyelins (mmol/L)	0.032	(0.020, 0.045)	<0.001	<0.001	0.078
Total cholines (mmol/L)	0.170	(0.115, 0.224)	<0.001	<0.001	0.349
Apolipoproteins					
Apolipoprotein A1 (g/L)	0.146	(0.114, 0.178)	<0.001	<0.001	0.209
Apolipoprotein B (g/L)	-0.115	(-0.146, -0.084)	<0.001	<0.001	0.198
Ratio of apolipoprotein B to apolipoprotein A1	-0.126	(-0.148, -0.105)	<0.001	<0.001	0.144
Fatty acids					
Total fatty acids (mmol/L)	-0.711	(-1.091, -0.330)	<0.001	<0.001	2.388
Estimated degree of unsaturation	0.031	(0.021, 0.042)	<0.001	<0.001	0.066
22:6, docosahexaenoic acid (mmol/L)	-0.002	(-0.008, 0.005)	0.622	0.667	0.041
18:2, linoleic acid (mmol/L)	0.004	(-0.087, 0.094)	0.934	0.947	0.566
Omega3 fatty acids (mmol/L)	-0.035	(-0.054, -0.016)	<0.001	0.001	0.122
Omega6 fatty acids (mmol/L)	0.004	(-0.094, 0.102)	0.936	0.936	0.612
Polyunsat. fatty acids (mmol/L)	-0.031	(-0.144, 0.082)	0.592	0.644	0.708
Monounsatur. fatty acids; 16:1, 18:1 (mmol/L)	-0.372	(-0.520, -0.225)	<0.001	<0.001	0.930
Saturated fatty acids (mmol/L)	-0.307	(-0.455, -0.159)	<0.001	<0.001	0.932
Fatty acid ratios					
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.064	(0.018, 0.110)	0.006	0.009	0.288
Ratio of 18:2 linoleic acid to total fatty acids (%)	1.527	(0.984, 2.070)	<0.001	<0.001	3.430
Ratio of omega3 fatty acids to total fatty acids (%)	-0.038	(-0.152, 0.075)	0.508	0.570	0.710
Ratio of omega6 fatty acids to total fatty acids (%)	1.882	(1.351, 2.412)	<0.001	<0.001	3.376
Ratio of polyunsatur. fatty acids to total fatty acids (%)	1.843	(1.272, 2.414)	<0.001	<0.001	3.621
Ratio of monounsatur. fatty acids to total fatty acids (%)	-1.456	(-1.914, -0.998)	<0.001	<0.001	2.904
Ratio of saturated fatty acids to total fatty acids (%)	-0.387	(-0.700, -0.074)	0.015	0.022	1.959
Glycolysis related					
Glucose (mmol/L)*	-0.071	(-0.097, -0.044)	<0.001	<0.001	0.168
Lactate (mmol/L)	-0.178	(-0.252, -0.105)	<0.001	<0.001	0.469
Pyruvate (mmol/L)	-0.007	(-0.013, -0.002)	0.004	0.006	0.032

1	Citrate (mmol/L)	-0.001	(-0.004, 0.001)	0.335	0.387	0.016
2	Glycerol (mmol/L)	-0.003	(-0.008, 0.002)	0.279	0.328	0.022
3						
4	Amino acids					
5	Alanine (mmol/L)	-0.020	(-0.030, -0.011)	<0.001	<0.001	0.060
6	Glutamine (mmol/L)	-0.038	(-0.048, -0.028)	<0.001	<0.001	0.065
7	Glycine (mmol/L)	0.029	(0.020, 0.038)	<0.001	<0.001	0.059
8	Histidine (mmol/L)	-0.001	(-0.003, 0.000)	0.116	0.146	0.009
9	Isoleucine (mmol/L)	-0.016	(-0.019, -0.013)	<0.001	<0.001	0.021
10	Leucine (mmol/L)	-0.019	(-0.022, -0.016)	<0.001	<0.001	0.022
11	Valine (mmol/L)	-0.029	(-0.035, -0.022)	<0.001	<0.001	0.042
12	Phenylalanine (mmol/L)	0.000	(-0.002, 0.001)	0.576	0.637	0.010
13	Tyrosine (mmol/L)	-0.005	(-0.007, -0.003)	<0.001	<0.001	0.014
14						
15	Ketone bodies					
16	Acetate (mmol/L)*	-0.076	(-0.119, -0.033)	0.001	0.001	0.273
17	Acetoacetate (mmol/L)*	0.018	(-0.148, 0.184)	0.828	0.863	1.048
18	3hydroxybutyrate (mmol/L)*	-0.087	(-0.163, -0.011)	0.025	0.035	0.472
19						
20	Fluid balance					
21	Albumin (signal area)	-0.002	(-0.002, -0.001)	<0.001	<0.001	0.005
22	Creatinine (mmol/L)	-0.013	(-0.015, -0.012)	<0.001	<0.001	0.010
23						
24	Inflammation					
25	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	-0.098	(-0.136, -0.061)	<0.001	<0.001	0.239

* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations for parents in Figure 3 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure in adults/parents) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

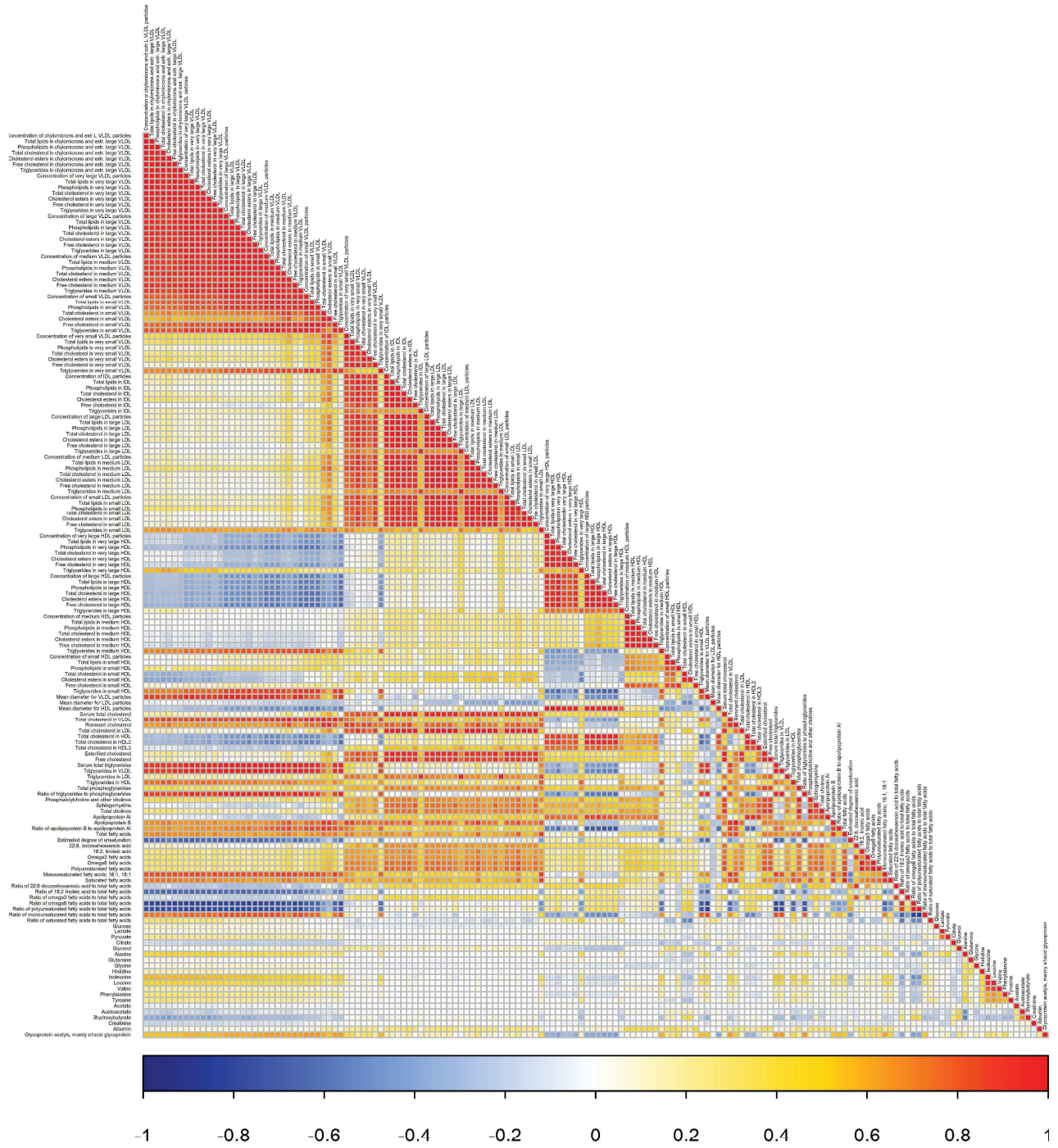
Supplementary table 5: Mother-child concordance; correlations and partial correlations between mothers and their sons, daughters and all children.

Metabolic subgroup	Mother																	
	Boys						Girls						All Children					
	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	PCC*	95% CI
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.Large VLDL (mmol/L)*	469	0.22	0.13-0.30	433	0.23	0.14-0.31	518	0.24	0.16-0.32	476	0.21	0.12-0.29	987	0.23	0.17-0.29	909	0.22	0.16-0.28
Total lipids in very large VLDL (mmol/L)*	469	0.25	0.16-0.33	433	0.25	0.16-0.34	518	0.22	0.14-0.30	476	0.20	0.12-0.29	987	0.24	0.18-0.29	909	0.23	0.17-0.29
Total lipids in large VLDL (mmol/L)*	469	0.22	0.13-0.30	433	0.24	0.15-0.33	518	0.23	0.14-0.31	476	0.22	0.14-0.31	987	0.22	0.16-0.28	909	0.23	0.17-0.29
Total lipids in medium VLDL (mmol/L)	469	0.26	0.17-0.34	433	0.28	0.19-0.36	518	0.29	0.21-0.37	476	0.30	0.21-0.38	987	0.28	0.22-0.34	909	0.29	0.23-0.35
Total lipids in small VLDL (mmol/L)	469	0.26	0.18-0.35	433	0.28	0.19-0.36	518	0.30	0.21-0.37	476	0.30	0.21-0.38	987	0.29	0.23-0.34	909	0.29	0.23-0.35
Total lipids in very small VLDL (mmol/L)	469	0.22	0.14-0.31	433	0.21	0.12-0.30	518	0.26	0.17-0.34	476	0.26	0.18-0.35	987	0.25	0.19-0.30	909	0.25	0.18-0.31
Total lipids in IDL (mmol/L)	469	0.27	0.18-0.35	433	0.23	0.14-0.32	518	0.29	0.21-0.37	476	0.31	0.23-0.39	987	0.29	0.23-0.34	909	0.28	0.22-0.34
Total lipids in large LDL (mmol/L)	469	0.28	0.19-0.36	433	0.24	0.15-0.33	518	0.30	0.22-0.37	476	0.32	0.24-0.40	987	0.29	0.23-0.35	909	0.29	0.23-0.35
Total lipids in medium LDL (mmol/L)	469	0.28	0.20-0.36	433	0.24	0.15-0.33	518	0.29	0.21-0.37	476	0.32	0.24-0.40	987	0.29	0.23-0.35	909	0.29	0.23-0.35
Total lipids in small LDL (mmol/L)	469	0.28	0.19-0.36	433	0.24	0.15-0.32	518	0.29	0.21-0.37	476	0.32	0.23-0.40	987	0.28	0.23-0.34	909	0.28	0.22-0.34
Total lipids in very large HDL (mmol/L)	469	0.30	0.22-0.38	433	0.30	0.21-0.39	518	0.32	0.24-0.39	476	0.30	0.21-0.38	987	0.31	0.25-0.36	909	0.30	0.24-0.36
Total lipids in large HDL (mmol/L)	469	0.31	0.23-0.39	433	0.31	0.23-0.40	518	0.28	0.20-0.36	476	0.26	0.18-0.34	987	0.30	0.24-0.35	909	0.29	0.23-0.35
Total lipids in medium HDL (mmol/L)	469	0.22	0.13-0.30	433	0.20	0.11-0.29	518	0.12	0.03-0.20	476	0.13	0.04-0.22	987	0.17	0.11-0.23	909	0.17	0.10-0.23
Total lipids in small HDL (mmol/L)	469	0.23	0.14-0.31	433	0.22	0.13-0.31	518	0.20	0.12-0.29	476	0.20	0.11-0.28	987	0.21	0.15-0.27	909	0.21	0.15-0.27
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	469	0.30	0.22-0.38	433	0.32	0.23-0.40	518	0.27	0.19-0.35	476	0.25	0.16-0.33	987	0.29	0.23-0.35	909	0.28	0.22-0.34
Mean diameter for LDL particles (nm)	469	0.22	0.13-0.31	433	0.20	0.11-0.29	518	0.30	0.22-0.38	476	0.32	0.24-0.40	987	0.26	0.20-0.31	909	0.26	0.20-0.32
Mean diameter for HDL particles (nm)	469	0.32	0.23-0.40	433	0.32	0.23-0.40	518	0.33	0.26-0.41	476	0.31	0.23-0.39	987	0.33	0.27-0.38	909	0.31	0.25-0.37
Cholesterol																		
Serum total cholesterol (mmol/L)	469	0.27	0.19-0.35	433	0.23	0.14-0.32	518	0.32	0.24-0.39	476	0.34	0.26-0.42	987	0.30	0.24-0.35	909	0.30	0.24-0.35
Total cholesterol in VLDL (mmol/L)	469	0.25	0.17-0.34	433	0.27	0.18-0.36	518	0.29	0.21-0.36	476	0.29	0.21-0.37	987	0.28	0.22-0.33	909	0.29	0.23-0.35
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	469	0.25	0.17-0.34	433	0.26	0.17-0.34	518	0.29	0.21-0.37	476	0.31	0.22-0.39	987	0.28	0.22-0.34	909	0.29	0.23-0.35
Total cholesterol in LDL (mmol/L)	469	0.28	0.20-0.36	433	0.24	0.15-0.33	518	0.29	0.21-0.37	476	0.32	0.23-0.40	987	0.29	0.23-0.34	909	0.29	0.23-0.35
Total cholesterol in HDL (mmol/L)	469	0.30	0.22-0.38	433	0.30	0.21-0.39	518	0.32	0.24-0.39	476	0.34	0.26-0.42	987	0.30	0.24-0.35	909	0.27	0.21-0.33
Total cholesterol in very large HDL (mmol/L)	469	0.30	0.22-0.38	433	0.31	0.22-0.39	518	0.32	0.24-0.40	476	0.33	0.25-0.41	987	0.28	0.22-0.33	909	0.27	0.21-0.33
Total cholesterol in large HDL (mmol/L)	469	0.25	0.16-0.33	433	0.23	0.14-0.32	518	0.25	0.16-0.33	476	0.24	0.16-0.33	987	0.25	0.19-0.31	909	0.24	0.17-0.30
Esterified cholesterol (mmol/L)	465	0.28	0.19-0.36	430	0.23	0.14-0.32	518	0.32	0.24-0.39	476	0.34	0.26-0.42	983	0.30	0.24-0.35	906	0.29	0.23-0.35
Free cholesterol (mmol/L)	465	0.26	0.18-0.34	430	0.22	0.13-0.31	518	0.32	0.24-0.40	476	0.35	0.27-0.43	983	0.30	0.24-0.35	906	0.30	0.24-0.35
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	469	0.28	0.20-0.36	433	0.30	0.21-0.38	518	0.29	0.21-0.37	476	0.29	0.20-0.37	987	0.29	0.23-0.35	909	0.30	0.24-0.36
Triglycerides in VLDL (mmol/L)*	469	0.30	0.22-0.38	433	0.31	0.23-0.40	518	0.28	0.20-0.36	476	0.27	0.18-0.35	987	0.29	0.23-0.35	909	0.29	0.23-0.35
Triglycerides in LDL (mmol/L)	469	0.19	0.10-0.28	433	0.18	0.09-0.27	518	0.26	0.18-0.34	476	0.27	0.18-0.35	987	0.23	0.17-0.29	909	0.23	0.17-0.29
Triglycerides in HDL (mmol/L)	469	0.21	0.12-0.30	433	0.23	0.14-0.32	518	0.26	0.18-0.34	476	0.26	0.18-0.34	987	0.24	0.18-0.30	909	0.25	0.19-0.31
Total phosphoglycerides (mmol/L)	465	0.26	0.17-0.34	430	0.23	0.14-0.32	518	0.26	0.17-0.34	476	0.27	0.18-0.35	983	0.26	0.20-0.32	906	0.25	0.18-0.31
Ratio of triglycerides to phosphoglycerides	465	0.23	0.15-0.32	430	0.26	0.16-0.34	518	0.26	0.18-0.34	476	0.27	0.19-0.35	983	0.25	0.20-0.31	906	0.27	0.20-0.33
Phosphatidylcholine & other cholinols (mmol/L)	465	0.27	0.18-0.35	430	0.24	0.14-0.32	518	0.25	0.17-0.33	476	0.27	0.18-0.35	983	0.26	0.20-0.32	906	0.25	0.19-0.31
Sphingomyelins (mmol/L)	465	0.23	0.15-0.32	430	0.22	0.12-0.30	518	0.29	0.21-0.37	476	0.31	0.23-0.39	983	0.27	0.21-0.32	906	0.27	0.21-0.33
Total cholinols (mmol/L)	465	0.27	0.18-0.35	430	0.23	0.14-0.32	518	0.28	0.20-0.35	476	0.28	0.20-0.37	983	0.27	0.21-0.33	906	0.26	0.20-0.32
Apolipoproteins																		
Apolipoprotein A1 (g/L)	469	0.28	0.20-0.36	433	0.26	0.17-0.35	518	0.26	0.18-0.34	476	0.26	0.17-0.34	987	0.27	0.21-0.33	909	0.26	0.20-0.32
Apolipoprotein B (g/L)	469	0.26	0.18-0.35	433	0.27	0.18-0.35	517	0.30	0.22-0.38	475	0.32	0.24-0.40	986	0.29	0.23-0.35	908	0.31	0.25-0.36
Ratio of apolipoprotein B to apolipoprotein A1	469	0.28	0.20-0.36	433	0.30	0.21-0.38	517	0.25	0.17-0.33	475	0.25	0.16-0.33	986	0.27	0.21-0.33	908	0.28	0.22-0.34
Fatty acids																		
Total fatty acids (mmol/L)	462	0.22	0.13-0.30	427	0.22	0.13-0.31	517	0.31	0.23-0.39	475	0.33	0.25-0.41	979	0.27	0.22-0.33	902	0.29	0.22-0.34
Estimated degree of unsaturation	462	0.30	0.22-0.38	427	0.32	0.23-0.41	517	0.27	0.19-0.35	475	0.24	0.16-0.33	979	0.29	0.23-0.34	902	0.28	0.22-0.34
22:6, docosahexaenoic acid (mmol/L)	462	0.20	0.11-0.29	427	0.18	0.09-0.27	517	0.34	0.26-0.41	475	0.32	0.24-0.40	979	0.27	0.21-0.32	902	0.25	0.19-0.31
18:2, linoleic acid (mmol/L)	462	0.22	0.13-0.31	427	0.22	0.13-0.31	517	0.27	0.19-0.35	475	0.30	0.21-0.38	979	0.25	0.19-0.31	902	0.27	0.21-0.33
Omega3 fatty acids (mmol/L)	462	0.20	0.11-0.29	427	0.19	0.09-0.28	517	0.34	0.27-0.42	475	0.34	0.26-0.42	979	0.27	0.21-0.33	902	0.26	0.20-0.32
Omega6 fatty acids (mmol/L)	462	0.23	0.14-0.32	427	0.22	0.13-0.31	517	0.31	0.23-0.38	475	0.33	0.25-0.41	979	0.27	0.21-0.33	902	0.28	0.22-0.34
Polysat. fatty acids (mmol/L)	462	0.23	0.14-0.31	427	0.22	0.12-0.30	517	0.32	0.24-0.39	475	0.34	0.26-0.42	979	0.28	0.22-0.33	902	0.28	0.22-0.34
Monounsat. fatty acids: 16:1, 18:1 (mmol/L)	462	0.24	0.15-0.33	427	0.25	0.16-0.34	517	0.33	0.25-0.40	475	0.33	0.25-0.41	979	0.29	0.24-0.35	902	0.30	0.24-0.36
Saturated fatty acids (mmol/L)	462	0.21	0.12-0.29	427	0.21	0.12-0.30	517	0.29	0.21-0.37	475	0.30	0.21-0.38	979	0.26	0.20-0.32	902	0.26	0.20-0.32
Fatty acid ratios																		
Ratio of 22:6:docosahexaenoic acid to total fatty acids (%)	462	0.31	0.22-0.39	427	0.29	0.20-0.38	517	0.35	0.27-0.42	475	0.32	0.24-0.40	979	0.33	0.27-0.39	902	0.31	0.25-0.36
Ratio of 18:2 linoleic acid to total fatty acids (%)	462	0.13	0.04-0.22	427	0.15	0.06-0.24	517	0.20	0.11-0.28	475	0.16	0.08-0.25	979	0.17	0.11-0.23	902	0.16	0.09-0.22
Ratio of omega3 fatty acids to total fatty acids (%)	462	0.32	0.24-0.40	427	0.32	0.23-0.40	517	0.40	0.33-0.47	475	0.37	0.29-0.44	979	0.36	0.31-0.41	902	0.34	0.28-0.39
Ratio of omega6 fatty acids to total fatty acids (%)	462	0.23	0.15-0.32	427	0.26	0.17-0.35	517	0.25	0.17-0.33	475	0.22	0.13-0.30	979	0.24	0.18-0.30	902	0.24	0.17-0.30
Ratio of polysat. fatty acids to total fatty acids (%)	462	0.27	0.19-0.36	427	0.30	0.21-0.38	517	0.28	0.20-0.36	475	0.24	0.16-0.33	979	0.28	0.22-0.33	902	0.27	0.

Supplementary table 6: Parent-child concordance, correlation and partial correlations between all parents and their sons, daughters and all children.

Metabolic subgroup	All Parents																	
	Male Child					Female Child					All Children							
	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI			
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	547	0.22	0.14 - 0.30	505	0.23	0.14 - 0.31	586	0.20	0.12 - 0.28	544	0.17	0.08 - 0.25	1133	0.22	0.16 - 0.27	1049	0.20	0.15 - 0.26
Total lipids in very large VLDL (mmol/L)*	547	0.23	0.15 - 0.31	505	0.23	0.15 - 0.31	586	0.17	0.10 - 0.25	544	0.16	0.08 - 0.24	1133	0.21	0.15 - 0.26	1049	0.20	0.14 - 0.26
Total lipids in large VLDL (mmol/L)*	547	0.20	0.12 - 0.28	505	0.22	0.13 - 0.30	586	0.20	0.12 - 0.28	544	0.19	0.11 - 0.27	1133	0.20	0.15 - 0.26	1049	0.21	0.15 - 0.26
Total lipids in medium VLDL (mmol/L)	547	0.27	0.19 - 0.35	505	0.28	0.20 - 0.36	586	0.25	0.17 - 0.32	544	0.24	0.16 - 0.32	1133	0.26	0.21 - 0.31	1049	0.26	0.20 - 0.32
Total lipids in small VLDL (mmol/L)	547	0.27	0.19 - 0.34	505	0.27	0.19 - 0.35	586	0.26	0.18 - 0.33	544	0.26	0.17 - 0.33	1133	0.26	0.21 - 0.32	1049	0.26	0.21 - 0.32
Total lipids in very small VLDL (mmol/L)	547	0.23	0.15 - 0.31	505	0.22	0.13 - 0.30	586	0.25	0.17 - 0.33	544	0.26	0.18 - 0.34	1133	0.25	0.19 - 0.30	1049	0.24	0.19 - 0.30
Total lipids in IDL (mmol/L)	547	0.28	0.20 - 0.35	505	0.25	0.16 - 0.33	586	0.29	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.28	0.23 - 0.34
Total lipids in large LDL (mmol/L)	547	0.28	0.20 - 0.36	505	0.25	0.17 - 0.33	586	0.29	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.29	0.23 - 0.34
Total lipids in medium LDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.25	0.17 - 0.33	586	0.28	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.28	0.23 - 0.34	1049	0.29	0.23 - 0.34
Total lipids in small LDL (mmol/L)	547	0.28	0.20 - 0.35	505	0.24	0.16 - 0.32	586	0.28	0.20 - 0.35	544	0.31	0.23 - 0.38	1133	0.28	0.22 - 0.33	1049	0.28	0.22 - 0.33
Total lipids in very large HDL (mmol/L)	547	0.29	0.21 - 0.37	505	0.28	0.20 - 0.36	586	0.29	0.21 - 0.36	544	0.27	0.19 - 0.35	1133	0.29	0.24 - 0.34	1049	0.28	0.22 - 0.33
Total lipids in large HDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.28	0.19 - 0.36	586	0.24	0.17 - 0.32	544	0.22	0.14 - 0.30	1133	0.27	0.21 - 0.32	1049	0.25	0.19 - 0.30
Total lipids in medium HDL (mmol/L)	547	0.15	0.07 - 0.24	505	0.13	0.04 - 0.21	586	0.10	0.02 - 0.18	544	0.10	0.02 - 0.19	1133	0.12	0.07 - 0.18	1049	0.11	0.05 - 0.17
Total lipids in small HDL (mmol/L)	547	0.19	0.10 - 0.27	505	0.16	0.08 - 0.25	586	0.18	0.10 - 0.26	544	0.17	0.09 - 0.25	1133	0.18	0.12 - 0.23	1049	0.17	0.11 - 0.22
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	547	0.29	0.21 - 0.36	505	0.30	0.22 - 0.38	586	0.22	0.14 - 0.29	544	0.19	0.11 - 0.27	1133	0.25	0.20 - 0.31	1049	0.24	0.19 - 0.30
Mean diameter for LDL particles (nm)	547	0.19	0.11 - 0.27	505	0.17	0.08 - 0.25	586	0.27	0.19 - 0.34	544	0.29	0.21 - 0.36	1133	0.23	0.17 - 0.28	1049	0.23	0.17 - 0.28
Mean diameter for HDL particles (nm)	547	0.31	0.23 - 0.38	505	0.30	0.22 - 0.38	586	0.30	0.22 - 0.37	544	0.27	0.19 - 0.35	1133	0.30	0.25 - 0.35	1049	0.29	0.23 - 0.34
Cholesterol																		
Serum total cholesterol (mmol/L)	547	0.28	0.20 - 0.36	505	0.24	0.16 - 0.32	586	0.31	0.24 - 0.38	544	0.33	0.26 - 0.41	1133	0.30	0.24 - 0.35	1049	0.29	0.24 - 0.35
Total cholesterol in VLDL (mmol/L)	547	0.26	0.18 - 0.34	505	0.27	0.19 - 0.35	586	0.26	0.18 - 0.33	544	0.26	0.18 - 0.34	1133	0.26	0.21 - 0.32	1049	0.27	0.21 - 0.32
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	547	0.27	0.19 - 0.34	505	0.26	0.18 - 0.34	586	0.28	0.20 - 0.35	544	0.29	0.21 - 0.37	1133	0.27	0.22 - 0.33	1049	0.28	0.22 - 0.34
Total cholesterol in LDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.26	0.17 - 0.34	586	0.28	0.21 - 0.35	544	0.30	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.28	0.23 - 0.34
Total cholesterol in HDL (mmol/L)	547	0.27	0.19 - 0.35	505	0.25	0.17 - 0.33	586	0.22	0.14 - 0.29	544	0.20	0.12 - 0.28	1133	0.24	0.19 - 0.30	1049	0.22	0.17 - 0.28
Total cholesterol in HDL2 (mmol/L)	547	0.27	0.19 - 0.34	505	0.25	0.17 - 0.33	586	0.21	0.14 - 0.29	544	0.20	0.11 - 0.28	1133	0.24	0.18 - 0.29	1049	0.22	0.16 - 0.28
Total cholesterol in HDL3 (mmol/L)	547	0.25	0.17 - 0.33	505	0.22	0.14 - 0.30	586	0.23	0.15 - 0.31	544	0.23	0.14 - 0.30	1133	0.24	0.19 - 0.30	1049	0.22	0.16 - 0.28
Esterified cholesterol (mmol/L)	543	0.28	0.20 - 0.36	502	0.24	0.15 - 0.32	584	0.31	0.24 - 0.38	542	0.33	0.25 - 0.40	1127	0.30	0.24 - 0.35	1044	0.29	0.23 - 0.34
Free cholesterol (mmol/L)	543	0.27	0.19 - 0.34	502	0.23	0.14 - 0.31	584	0.32	0.24 - 0.39	542	0.34	0.26 - 0.41	1127	0.29	0.24 - 0.35	1044	0.29	0.23 - 0.34
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	547	0.28	0.20 - 0.35	505	0.29	0.20 - 0.36	586	0.25	0.17 - 0.32	544	0.24	0.16 - 0.32	1133	0.26	0.21 - 0.32	1049	0.26	0.21 - 0.32
Triglycerides in VLDL (mmol/L)*	547	0.28	0.20 - 0.36	505	0.29	0.21 - 0.37	586	0.23	0.15 - 0.30	544	0.21	0.13 - 0.29	1133	0.26	0.20 - 0.31	1049	0.25	0.20 - 0.31
Triglycerides in LDL (mmol/L)	547	0.20	0.12 - 0.28	505	0.19	0.10 - 0.27	586	0.27	0.19 - 0.34	544	0.28	0.20 - 0.35	1133	0.24	0.18 - 0.29	1049	0.24	0.18 - 0.29
Triglycerides in HDL (mmol/L)	547	0.25	0.17 - 0.33	505	0.26	0.18 - 0.34	586	0.27	0.20 - 0.35	544	0.28	0.20 - 0.35	1133	0.26	0.21 - 0.32	1049	0.27	0.21 - 0.32
Total phosphoglycerides (mmol/L)	543	0.28	0.20 - 0.36	502	0.24	0.16 - 0.32	584	0.27	0.19 - 0.34	542	0.29	0.21 - 0.36	1127	0.27	0.22 - 0.33	1044	0.26	0.20 - 0.32
Ratio of triglycerides to phosphoglycerides	543	0.23	0.15 - 0.31	502	0.24	0.16 - 0.32	584	0.21	0.13 - 0.28	542	0.20	0.12 - 0.28	1127	0.22	0.16 - 0.27	1044	0.22	0.16 - 0.28
Phosphatidylcholine & other cholines (mmol/L)	542	0.28	0.20 - 0.36	501	0.23	0.15 - 0.32	584	0.26	0.19 - 0.34	542	0.28	0.20 - 0.36	1126	0.27	0.21 - 0.32	1043	0.26	0.20 - 0.31
Sphingomyelins (mmol/L)	542	0.23	0.15 - 0.31	501	0.20	0.12 - 0.29	584	0.28	0.21 - 0.36	542	0.30	0.22 - 0.37	1126	0.26	0.20 - 0.31	1043	0.26	0.20 - 0.31
Total cholines (mmol/L)	542	0.27	0.19 - 0.35	501	0.23	0.14 - 0.31	584	0.29	0.21 - 0.36	542	0.30	0.22 - 0.37	1126	0.28	0.22 - 0.33	1043	0.26	0.20 - 0.32
Apolipoproteins																		
Apolipoprotein A1 (g/L)	547	0.26	0.18 - 0.34	505	0.23	0.14 - 0.31	586	0.25	0.17 - 0.33	544	0.25	0.17 - 0.32	1133	0.25	0.20 - 0.31	1049	0.23	0.18 - 0.29
Apolipoprotein B (g/L)	547	0.27	0.19 - 0.35	505	0.27	0.19 - 0.35	585	0.28	0.20 - 0.35	543	0.30	0.22 - 0.37	1132	0.28	0.23 - 0.33	1048	0.29	0.23 - 0.34
Ratio of apolipoprotein B to apolipoprotein AI	547	0.26	0.18 - 0.33	505	0.27	0.18 - 0.34	585	0.21	0.13 - 0.29	543	0.20	0.12 - 0.28	1132	0.23	0.18 - 0.29	1048	0.24	0.18 - 0.29
Fatty acids																		
Total fatty acids (mmol/L)	537	0.26	0.18 - 0.33	496	0.25	0.17 - 0.33	583	0.30	0.22 - 0.37	541	0.31	0.23 - 0.39	1120	0.28	0.23 - 0.33	1037	0.29	0.23 - 0.34
Estimated degree of unsaturation	537	0.30	0.22 - 0.37	496	0.32	0.23 - 0.39	583	0.24	0.17 - 0.32	541	0.22	0.14 - 0.30	1120	0.27	0.21 - 0.32	1037	0.26	0.20 - 0.32
22:6, docosahexaenoic acid (mmol/L)	537	0.23	0.15 - 0.31	496	0.21	0.13 - 0.30	583	0.33	0.26 - 0.40	541	0.33	0.25 - 0.40	1120	0.28	0.23 - 0.34	1037	0.27	0.21 - 0.32
18:2, linoleic acid (mmol/L)	537	0.24	0.16 - 0.32	496	0.25	0.16 - 0.33	583	0.27	0.19 - 0.34	541	0.29	0.21 - 0.37	1120	0.26	0.21 - 0.32	1037	0.28	0.22 - 0.33
Omega3 fatty acids (mmol/L)	537	0.24	0.16 - 0.32	496	0.22	0.14 - 0.31	583	0.34	0.27 - 0.41	541	0.34	0.27 - 0.41	1120	0.29	0.23 - 0.34	1037	0.28	0.22 - 0.33
Omega6 fatty acids (mmol/L)	537	0.26	0.18 - 0.33	496	0.25	0.16 - 0.33	583	0.30	0.23 - 0.38	541	0.33	0.25 - 0.40	1120	0.28	0.23 - 0.34	1037	0.29	0.24 - 0.35
Polynsat. fatty acids (mmol/L)	537	0.26	0.17 - 0.33	496	0.24	0.16 - 0.33	583	0.31	0.24 - 0.39	541	0.34	0.26 - 0.41	1120	0.29	0.23 - 0.34	1037	0.29	0.24 - 0.35
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	537	0.27	0.19 - 0.35	496	0.28	0.19 - 0.36	583	0.30	0.22 - 0.37	541	0.30	0.23 - 0.38	1120	0.29	0.24 - 0.34	1037	0.29	0.24 - 0.35
Saturated fatty acids (mmol/L)	537	0.25	0.17 - 0.33	496	0.25	0.16 - 0.33	583	0.28	0.20 - 0.35	541	0.28	0.20 - 0.36	1120	0.27	0.21 - 0.32	1037	0.27	0.21 - 0.32
Fatty acid ratios																		
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	537	0.31	0.23 - 0.38	496	0.30	0.21 - 0.37	583	0.33	0.26 - 0.40	541	0.31	0.23 - 0.38	1120	0.32	0.27 - 0.37	1037	0.30	0.24 - 0.36
Ratio of 18:2 linoleic acid to total fatty acids (%)	537	0.15	0.07 - 0.23	496	0.17	0.08 - 0.25	583	0.18	0.10 - 0.26	541	0.16	0.08 - 0.24	1120	0.17	0.11 - 0.23	1037	0.16	0.10 - 0.22
Ratio of omega3 fatty acids to total fatty acids (%)	537	0.32	0.24 - 0.39	496	0.32	0.23 - 0.39	583	0.40	0.33 - 0.47	541	0.38	0.31 - 0.45	1120	0.36	0.31 - 0.41	1037	0.35	0.29 - 0.40
Ratio of omega6 fatty acids to total fatty acids (%)	537	0.24	0.16 -															

Supplementary Figure 1: Correlations between Child Health CheckPoint metabolites - children



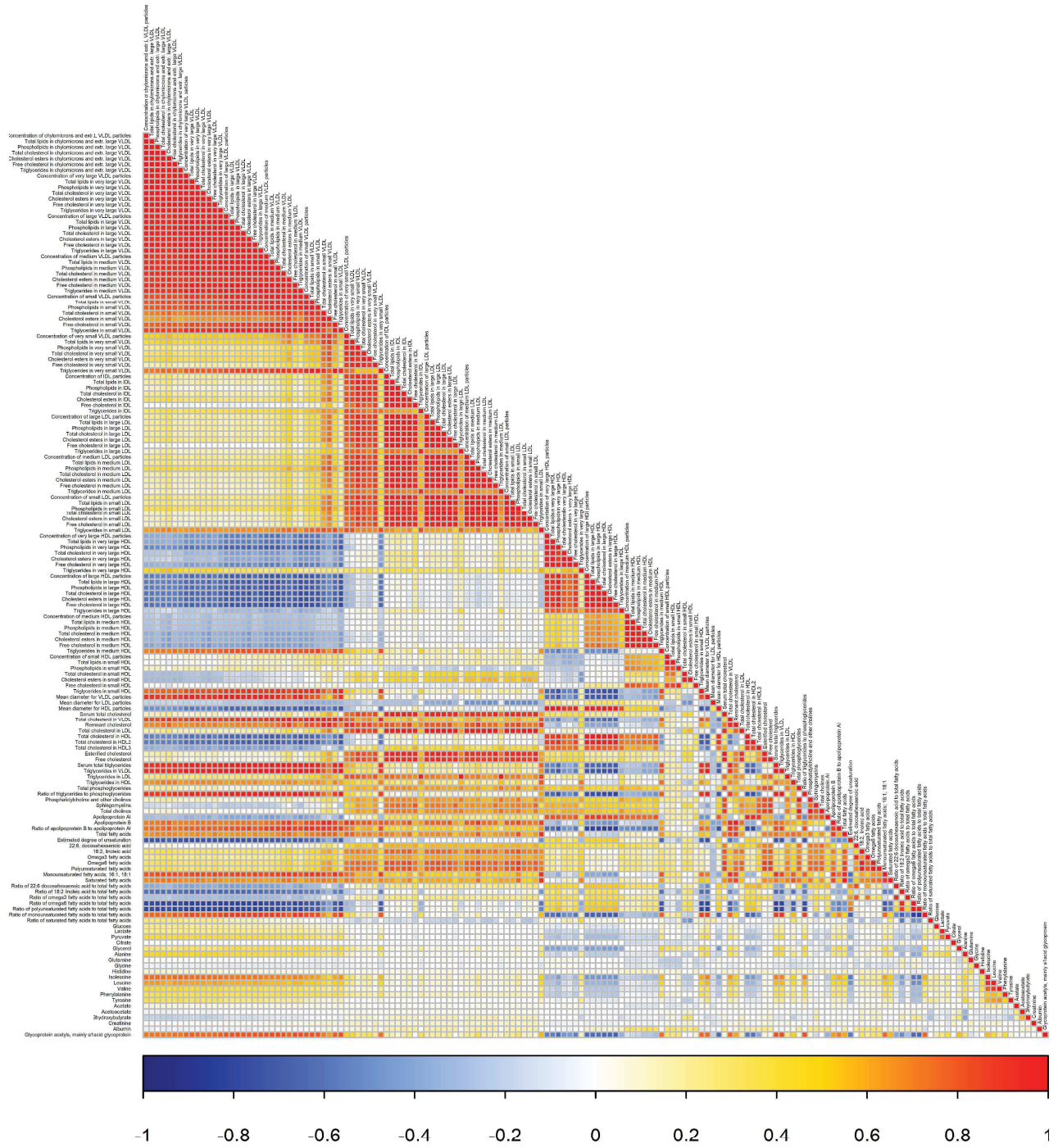
Note: Correlations (spearman) between metabolites for the CheckPoint child metabolomics data

Supplementary figure 1: Correlation of NMR measures in children.

Heatmap showing the correlation between metabolite measures in children. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

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Supplementary Figure 2: Correlations between Child Health CheckPoint metabolites - parents

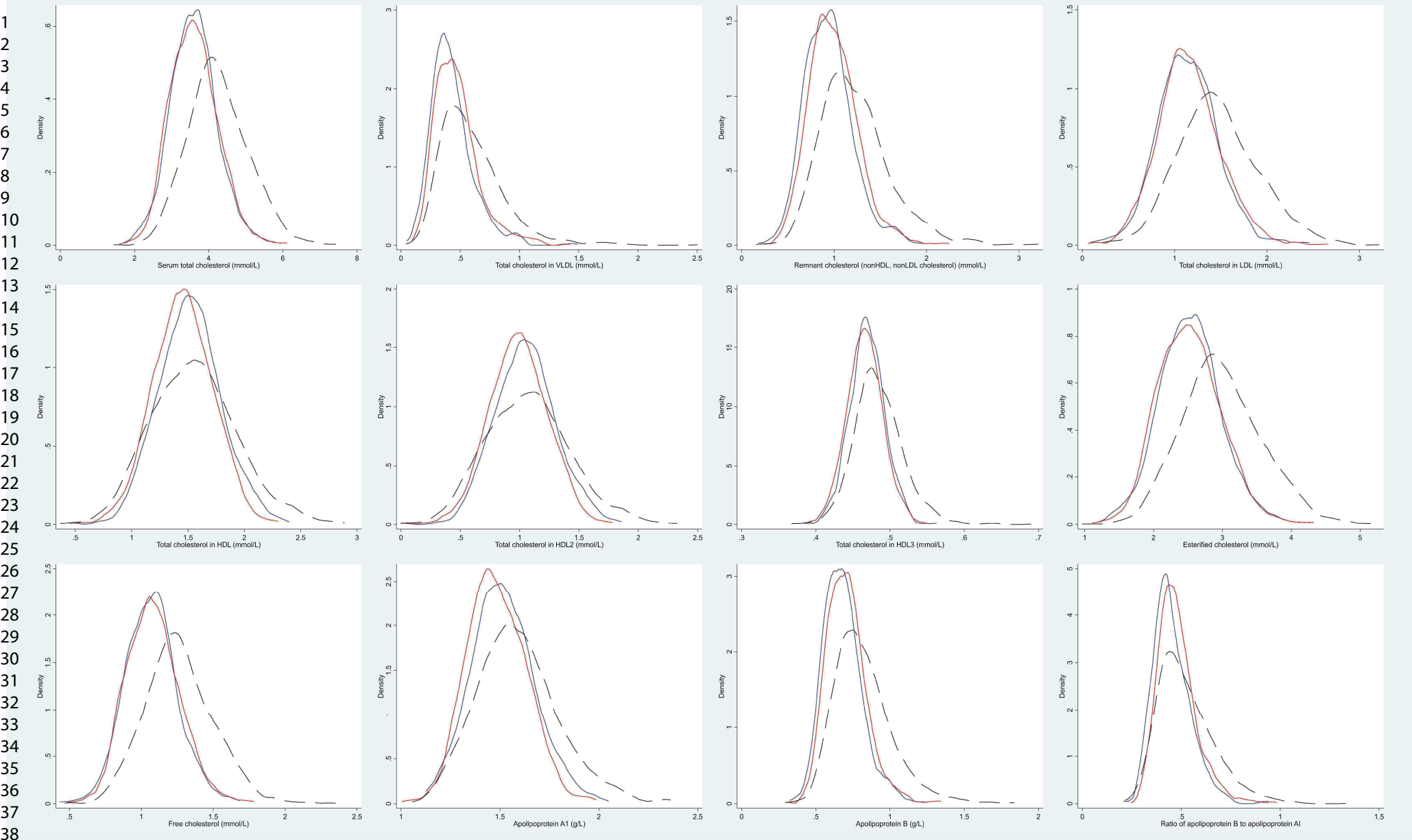


Note: Correlations (spearman) between metabolites for the CheckPoint parent metabolomics data

Supplementary figure 2: Correlation of NMR metabolite measures in parents.

Heatmap showing the correlation between metabolite measures in parents. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

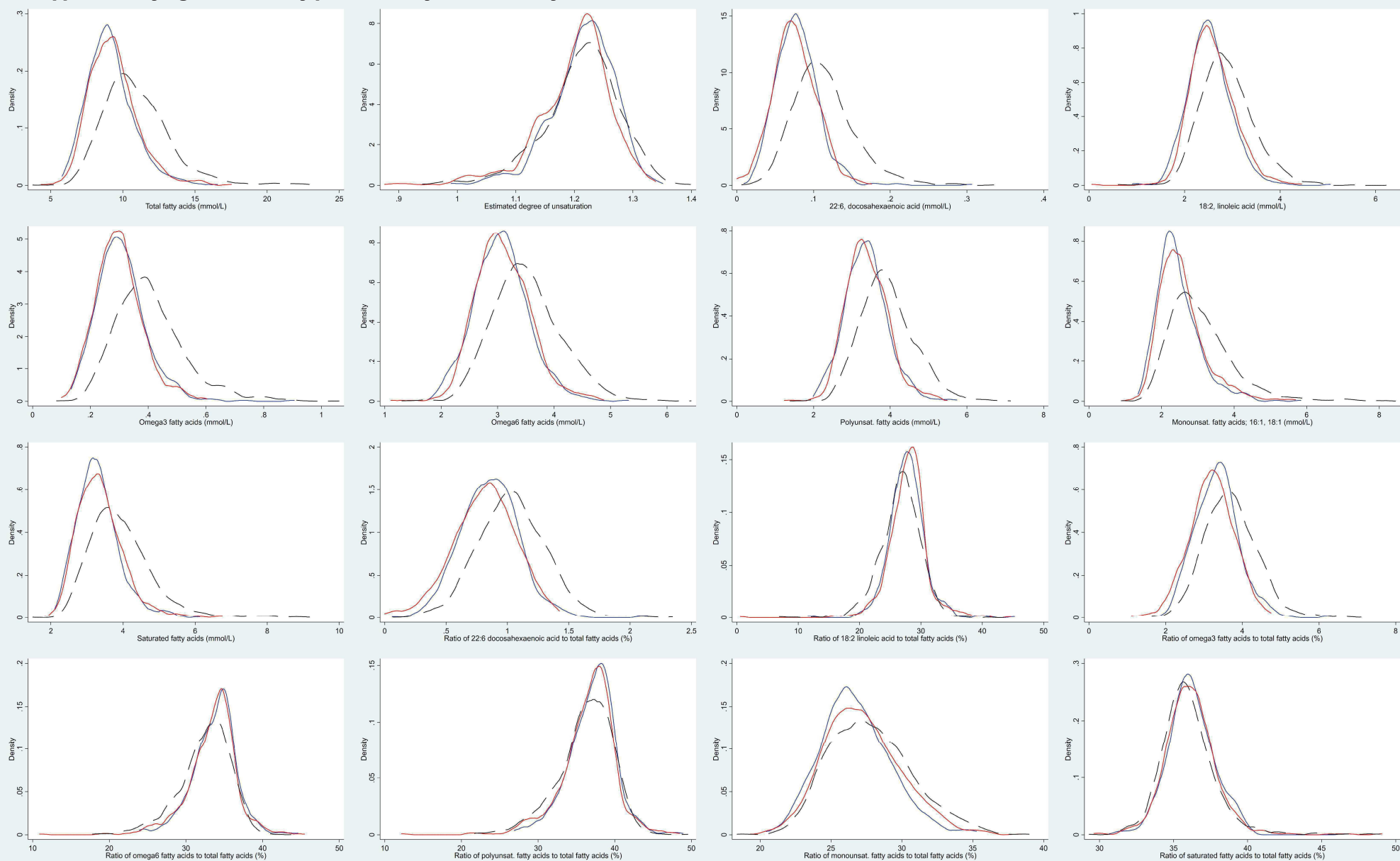
Supplementary figure 3: Density plots for cholesterol and apolipoprotein measures.



— Boys
 — Girls
 - - - All parents

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each cholesterol and apolipoprotein measure.

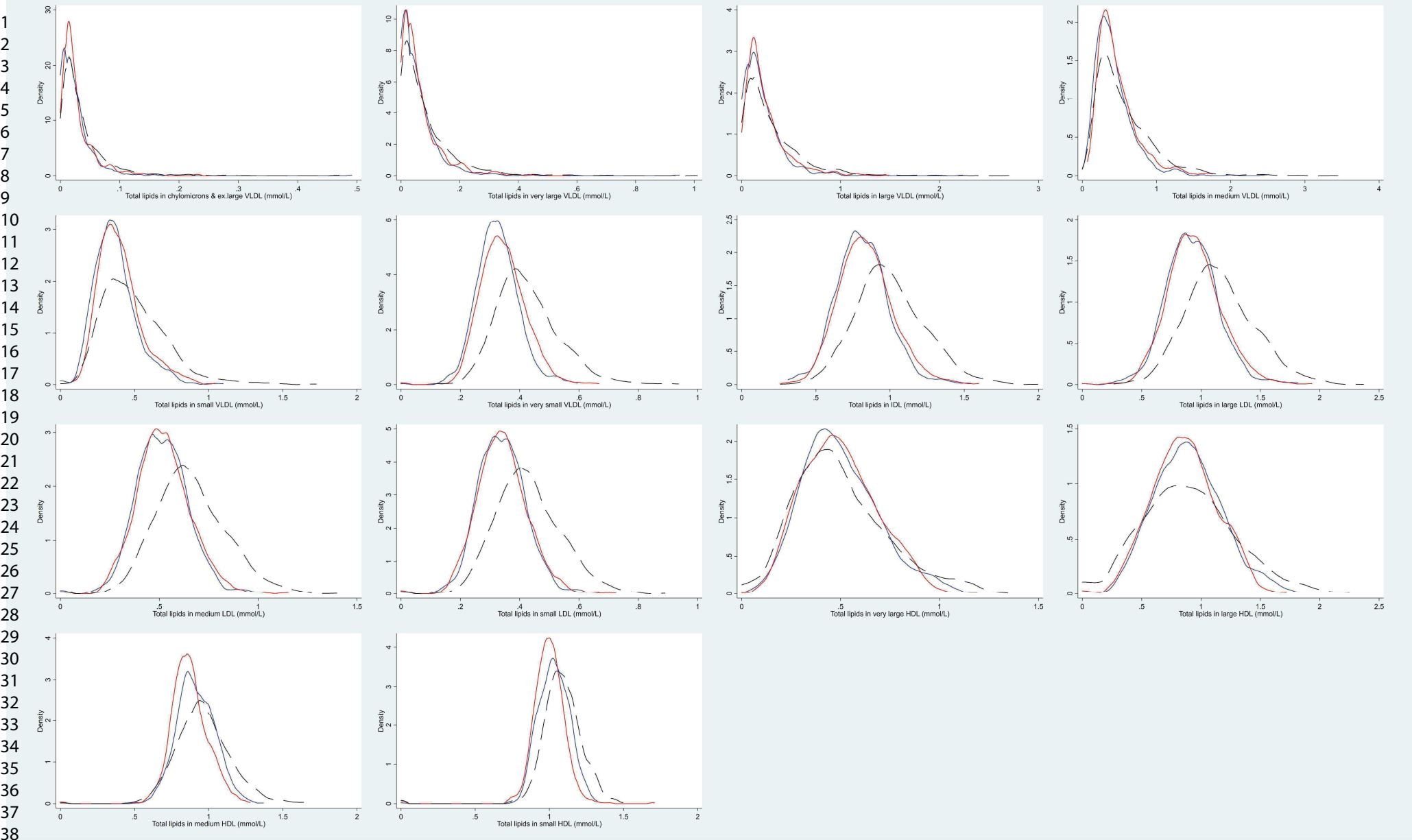
Supplementary figure 4: Density plots for fatty acid and fatty acid ratio measures.



Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each fatty acid and fatty acid ratio measure.

Preprint BMJ Open 2019;19:e024444. <https://doi.org/10.1136/bmjopen-2019-024444>

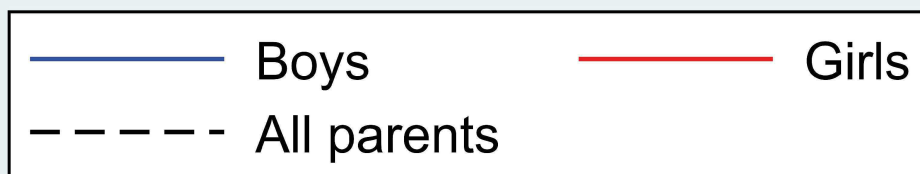
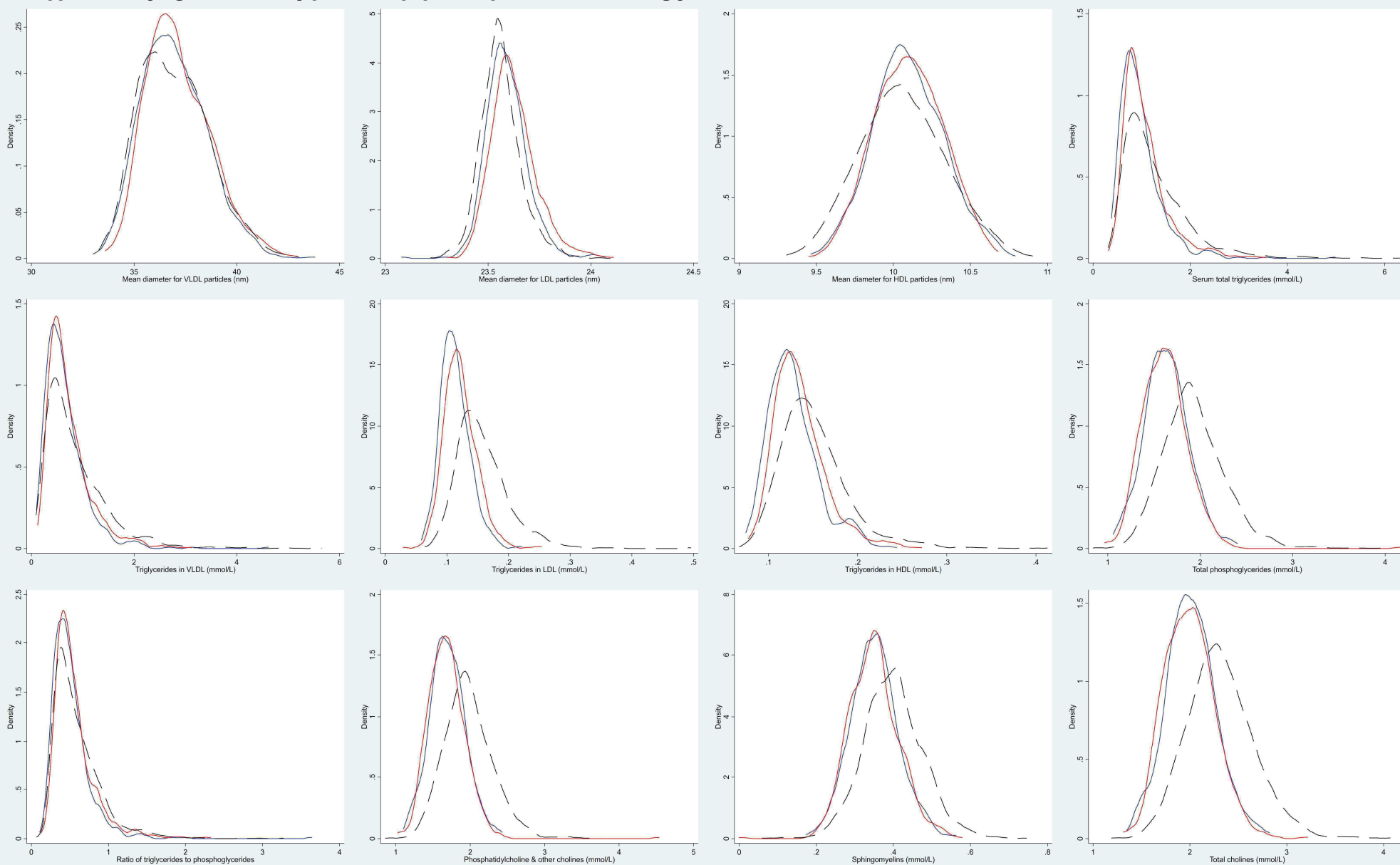
Supplementary figure 5: Density plots for total lipids in the 14 lipoprotein subclass particles.



Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for total lipids within each of the 14 lipoprotein subclass particles.

For peer review only: <http://bmjopen.bmj.com/site/about/guidelines.xhtml>

Supplementary figure 6: Density plots for lipoprotein particle size and triglyceride measures.

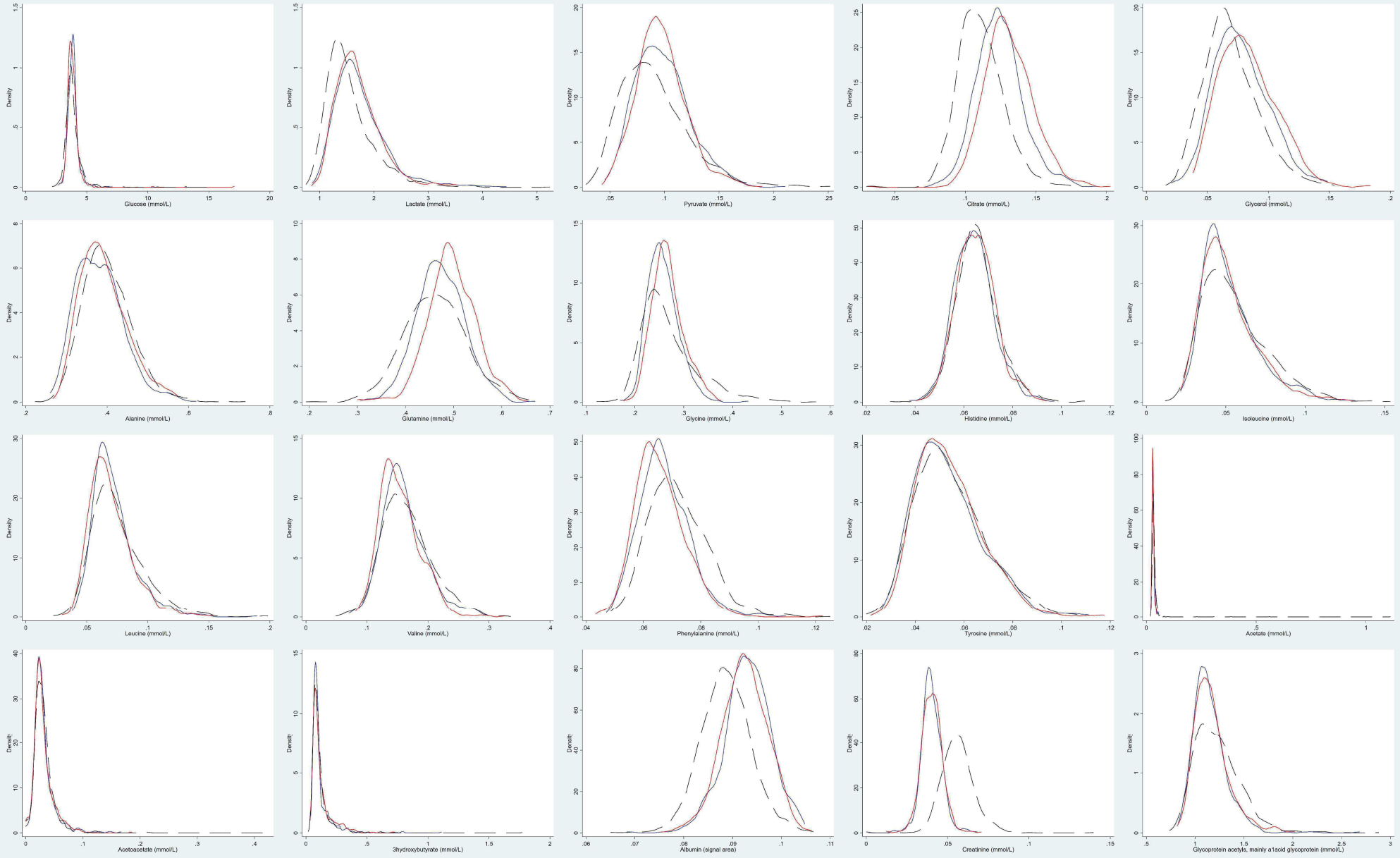


Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for lipoprotein particle sizes and triglyceride measures.

For peer review only: <http://bmjopen.bmj.com/site/about/guidelines.xhtml>

Supplementary figure 7: Density plots for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

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Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

STROBE Statement—checklist of items that should be included in reports of observational studies

Paper title: Metabolomics: Population epidemiology and concordance in 11-12 year old

Australians and their parents

Person completing checklist: Susan Ellul

	Item No	Recommendation	Page number	Line number
Title and abstract	1	(a) Indicate the study's design with a commonly used term in the title or the abstract	2	14-15
		(b) Provide in the abstract an informative and balanced summary of what was done and what was found	2	9-13 25-32
Introduction				
Background/rationale	2	Explain the scientific background and rationale for the investigation being reported	4-5	Pg 4 (3-9 ,21-22, 31-32, 33-35) Pg 5 (5-7)
Objectives	3	State specific objectives, including any prespecified hypotheses	5	8-11
Methods				
Study design	4	Present key elements of study design early in the paper	5	14-22
Setting	5	Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection	5-6	Pg 5 (17-20) Pg 6 (3-32)
Participants	6	(a) <i>Cohort study</i> —Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up <i>Case control study</i>—Give the eligibility criteria, and the sources and methods of case ascertainment and control selection. Give the rationale for the choice of cases and controls <i>Cross-sectional study</i> —Give the eligibility criteria, and the sources and methods of selection of participants	5	23-31
		(b) <i>Cohort study</i>—For matched studies, give matching criteria and number of exposed and unexposed <i>Case control study</i>—For matched studies, give matching criteria and the number of controls per case		
Variables	7	Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable	7-9	Pg 7 (4-17) Pg 8 (table 1, 9-16) Pg 9 (1-24)
Data sources/ measurement	8*	For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group	7-9	Pg 7 (4-17) Pg 8 (table 1, 9-16) Pg 9 (1-24)
Bias	9	Describe any efforts to address potential sources of bias	9 -10	Pg 9 (31-35) Pg 10 (16-18)

Study size	10	Explain how the study size was arrived at	11	5-14, figure 1
Quantitative variables	11	Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why	9-10	Pg 9 (28-35) Pg 10 (3-15)
Statistical methods	12	(a) Describe all statistical methods, including those used to control for confounding	9-10	Pg 9 (28-35) Pg 10 (3-15)
		(b) Describe any methods used to examine subgroups and interactions	9-10	Pg 9 (28-35) Pg 10 (3-15)
		(c) Explain how missing data were addressed	9 -10	Pg 9 (31-35) Pg 10 (16-18)
		(d) <i>Cohort study</i> —If applicable, explain how loss to follow-up was addressed	9	31-35
		(e) <i>Case-control study</i>—If applicable, explain how matching of cases and controls was addressed		
		(e) Describe any sensitivity analyses	10	16-18

Results			Page number	Line number
Participants	13*	(a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed	11 and figure 1	5-12
		(b) Give reasons for non-participation at each stage	11 6 figure 1	8-12 7-11
		(c) Consider use of a flow diagram	figure 1	
Descriptive data	14*	(a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders	12 (table 2)	2
		(b) Indicate number of participants with missing data for each variable of interest	figure 1 table 2 sup Table 1	2
		(c) <i>Cohort study</i> —Summarise follow-up time (eg, average and total amount)	NA	
Outcome data	15*	<i>Cohort study</i> —Report numbers of outcome events or summary measures over time	sup table 1	
		<i>Case-control study</i> —Report numbers in each exposure category, or summary measures of exposure	NA	
		<i>Cross-sectional study</i> —Report numbers of outcome events or summary measures	NA	
Main results	16	(a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included	13 supp tables 2-6	21-24
		(b) Report category boundaries when continuous variables were categorized	NA	

		(c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period	NA	
Other analyses	17	Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses	supp tables 2-6	
Discussion				
Key results	18	Summarise key results with reference to study objectives	14	3-10
Limitations	19	Discuss limitations of the study, taking into account sources of potential bias or imprecision. Discuss both direction and magnitude of any potential bias	14	18-28
Interpretation	20	Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence	14-17	Pg 14 (30-32) Pg 15 (all) Pg 16 (all) Pg 17 (1-2)
Generalisability	21	Discuss the generalisability (external validity) of the study results	14	18-28
Other information				
Funding	22	Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based	18-19	Pg 18 (21-33) Pg 19 (1-2)

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.

BMJ Open

Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their parents

Journal:	<i>BMJ Open</i>
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Keywords:	Metabolomics, Lipids, Reference values, Children, Inheritance patterns, Epidemiologic studies

SCHOLARONE™
Manuscripts

1 **Metabolomics: Population epidemiology and concordance in 11-12 year old Australians and their**
2 **parents**

3
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24
25 **Keywords:** Metabolomics; lipids; inflammation; reference values; parents; children;
26 inheritance patterns; correlation studies; epidemiologic studies; cross-sectional studies.

27
28 **Word count: 4328**

29
30 **Abbreviations:** ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; BCAA: Branched
31 chain amino acid; BD: Becton Dickinson; CDC: Centers for Disease Prevention and Control;
32 CVD: Cardiovascular disease; CPS1: Carbamoyl-phosphate synthase 1; DHA:
33 Docosahexaenoic acid; DOB: Date of birth; EDTA: Ethylenediaminetetraacetic acid; FDR:
34 False discovery rate; GlycA: Glycoprotein acetyls; HbA1c: Haemoglobin A1c; HDL: High-
35 density lipoprotein; HOMA: Homeostatic model assessment; IDL: Intermediate density
36 lipoprotein; LA: Linoleic acid; LDL: Low-density lipoprotein; LiH: Lithium Heparin; LSAC:

1 Longitudinal Study of Australian Children; MUFA: Monounsaturated fatty acid; NMR:
2 Nuclear magnetic resonance; PCOS: Polycystic Ovary Syndrome; PUFA: Polyunsaturated
3 fatty acid; REDCap: Research Electronic Data Capture; SFA: Saturated fatty acids; SST:
4 serum separating tubes; T2D: Type 2 diabetes; T2DM: Type 2 diabetes mellitus; VLDL:
5 Very low density lipoprotein; XL: Very large; XXL: Chylomicrons and extremely large; XS:
6 Very small.

7 8 **ABSTRACT**

9
10 **Objectives:** Nuclear Magnetic Resonance (NMR) metabolomics is high throughput and cost
11 effective, with the potential to improve the understanding of disease and risk. We examine
12 the circulating metabolic profile by quantitative NMR metabolomics of a sample of
13 Australian 11-12 year old children and their parents, describe differences by age and sex, and
14 explore correlation of metabolites in parent-child dyads.

15 **Design:** The population-based cross-sectional Child Health CheckPoint study nested within
16 the Longitudinal Study of Australian Children.

17 **Setting:** Blood samples collected from CheckPoint participants at assessment centres in
18 seven Australian cities and eight regional towns; February 2015-March 2016.

19 **Participants:** 1180 children and 1325 parents provided a blood sample and had
20 metabolomics data available. This included 1133 parent-child dyads (518 mother-daughter,
21 469 mother-son, 68 father-daughter, and 78 father-son).

22 **Outcome measures:** 228 metabolic measures were obtained for each participant. We
23 focused on 74 biomarkers including amino acid species, lipoprotein subclass measures, lipids,
24 fatty acids, measures related to fatty acid saturation, and composite markers of inflammation
25 and energy homeostasis.

26 **Results:** We identified differences in the concentration of specific metabolites between
27 childhood and adulthood and in metabolic profiles in children and adults by sex. In general,
28 metabolite concentrations were higher in adults than children and sex differences were larger
29 in adults than in children. Positive correlations were observed for the majority of metabolites
30 including isoleucine (CC 0.33, 95% CI 0.27 to 0.38), total cholesterol (CC 0.30, 95% CI 0.24
31 to 0.35) and omega 6 fatty acids (CC 0.28, 95% CI 0.23 to 0.34) in parent-child comparisons.

32 **Conclusions:** We describe the serum metabolite profiles from mid-childhood and adulthood
33 in a population-based sample, together with parent-child concordance. Differences in profiles
34 by age and sex were observed. These data will be informative for investigation of the
35 childhood origins of adult non-communicable diseases and for comparative studies in other
36 populations.

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2 **Strengths and limitations of this study:**

- 3 • In a large population-based cohort, venous blood was collected for children and their
4 attending parent on the same day using the same methods
- 5 • Rapidly processed, high quality serum samples with standardised metabolomic data
6 generated as a single batch
- 7 • Cross-sectional design does not enable longitudinal analysis of specific metabolite
8 species over short term or longer periods of time
- 9 • Assessment of paternal associations with offspring metabolite measures is limited by
10 a relatively small sample size compared to mother-child pairs, reducing the precision
11 of estimates
- 12 • Factors known to influence metabolomic profile (such as body mass index) were not
13 considered as the aim was to describe the distribution of metabolites in children and
14 their parents.

1 INTRODUCTION

2
3 Metabolomics involves the quantitative analysis of a large number of metabolites and lipids
4 involved in a diverse range of biochemical pathways.¹ Genetic/gene expression and
5 environmental exposures are associated with specific metabolic changes across many tissues
6 and body fluids.^{2 3} As such, metabolomics is recognised as a powerful top-down approach to
7 understanding genetic and environmental influences on health and disease. Metabolomic
8 profiling also has considerable potential to identify clinically relevant biomarkers for risk
9 stratification and disease monitoring.

10 Recent advances in nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry
11 have enabled the simultaneous quantitative measurement of hundreds of metabolites. These
12 approaches are sufficiently cost effective and high throughput to be applicable to large cohort
13 studies. For example, NMR metabolomics of serum from the Cardiovascular Risk in Young
14 Finns Study identified many biomarkers from multiple metabolic pathways reflective of fatty
15 liver disease.⁴ These were also predictive of risk 10 years prior to diagnosis, indicating that
16 metabolic disruptions precede overt phenotype. Similar population and disease-specific
17 studies have identified metabolomic profiles associated with a range of exposures and health
18 outcomes with potential to reveal clinically important biomarkers and information on disease
19 mechanisms.⁵ In addition, specific serum metabolites can also be considered ‘intermediate
20 phenotypes’ linking genetic risk with disease outcomes.^{6 7}

21 Previous research indicates that some blood metabolites change with age, particularly from
22 mid to late adulthood.^{8 9} However, in adults sex appears to be a major driver of variation in
23 metabolite profile, potentially interacting with age. For example, the effects of sex appeared
24 to be greater in younger (age 25-35) than older Japanese adults.¹⁰ A study of 26,000 Northern
25 European adults identified many sex-specific metabolic species at the population level.⁹ In
26 men, several lipid measures begin to rise at early middle age whereas a similar increase is
27 only observed in females post menopause. This pattern is consistent for all non-HDL
28 cholesterol measures – very low density lipoprotein (VLDL), intermediate density lipoprotein
29 (IDL) and low density lipoprotein (LDL) subclass particle concentrations - as well as for
30 triglycerides.⁹ Physiological states such as pregnancy also have consistent and measurable
31 influence on serum metabolome.¹¹ However, it remains unclear how the serum metabolome
32 differs in adults compared to children and by sex particularly in childhood.

33 Moreover, factors regulating the metabolic trajectory from early life to adulthood, the role of
34 metabolomic profile in health at the population level and the extent to which blood
35 metabolomic profiles are concordant for parents and children have not been fully explored.
36 One small study has reported correlations between parents (n=179) and their offspring

(n=255) for a range of cardiometabolic risk factors including standard lipid profile measured using conventional methods; this proved stronger for total cholesterol and LDL cholesterol than for high density lipoprotein (HDL) cholesterol or triglycerides.¹² Considerable evidence exists that the metabolomic profile is regulated, at least in part, by genetic factors^{13 14} and is also influenced by dietary and lifestyle factors. Each of these influences is likely to be shared between parents and their offspring to varying degrees, however, parent-child correlations of metabolites from NMR-based platforms have not been reported previously.

Here, we describe (1) the distribution of NMR-based metabolite measures in a population-based cohort of 11-12 year old children and their parents, differences in metabolite concentrations (2) by age (adults compared to children) and (3) by sex in children and adults; and (4) report sex-specific parent-child concordance.

METHODS

Study Design: Details of the initial Longitudinal Study of Australian Children (LSAC) study design and recruitment are outlined elsewhere.^{15 16} The LSAC commenced in 2004, when two cohorts (the 'B' and 'K' cohorts, of which the B cohort only was included in the present study) were recruited who have since been followed biennially. The Child Health CheckPoint comprised a detailed cross-sectional assessment of physical health and biomarkers in a population-based national sample of children (age 11-12 years) and their parents between February 2015 to March 2016. The CheckPoint was nested between waves 6 (2014) and 7 (2016) of the LSAC. Further details regarding the CheckPoint study design and methods are available elsewhere.^{17 18}

Participants: Of the 8,921 families contacted to be part of the LSAC B cohort 5,107 families (57%) agreed to take part in the first wave of data collection in 2004; 4,484 families were retained for Wave 6 in 2014. During the Wave 6 LSAC home visit, B cohort families were introduced to the upcoming Child Health CheckPoint and asked to consent to their contact details being shared with the CheckPoint team. A total of 3,513 families provided permission to receive an information pack by mail and an information and recruitment phone call regarding the CheckPoint study (78% of Wave 6 cohort, 69% of original cohort). Of the families agreeing to receive information about the CheckPoint study, 1,874 families took part (53% of eligible participants, 42% of Wave 6 cohort and 37% of original cohort).

Ethics and consent: The CheckPoint data collection protocol was approved by The Royal Children's Hospital (Melbourne, Australia) Human Research Ethics Committee (33225D) and the Australian Institute of Family Studies Ethics Committee (14-26). The attending parent/caregiver provided written informed consent for themselves and their child to

1 participate in the study, and asked to provide optional consent for the collection and use of
2 biological samples.

3 **Procedure:** The specialised CheckPoint assessment centre sequentially visited seven
4 Australian cities and eight regional towns between February 2015 and March 2016.¹⁸ Each
5 participating child attended the centre with one parent or caregiver (usually the biological
6 mother) at which both participated in a wide range of measures relevant to non-
7 communicable disease. Those families who could not attend a centre were offered a home
8 visit. Participants were included in the current analyses if metabolomic data from CheckPoint
9 were available (figure 1). Venous blood was not available for home-visit participants, but was
10 collected at all city and most regional assessment centres. Participant pairs were excluded
11 from the concordance analyses in this study if the attending parent was not the biological
12 parent.

13 An experienced phlebotomist collected approximately 28mL of blood from the brachial vein
14 of the non-dominant arm of semi-reclining, semi-fasted participants (at the time of collection,
15 participants reported when they last ate or drank). Blood was collected sequentially into four
16 Becton Dickinson (BD) Vacutainer[®] tubes using a butterfly needle so only a single
17 venepuncture was required. Order of collection was (i) 2.7mL EDTA, (ii) 9mL EDTA, (iii)
18 9mL serum, (iv) 7.5mL Lithium Heparin. The latter two tubes were immediately inverted 6
19 times to ensure mixing with anticoagulant, and all tubes were transferred to the on-site
20 laboratory. Time of collection was scheduled earlier in the visit for parents than for children.

21 Collection tube barcodes were linked to the participant and samples were immediately
22 transported to an on-site laboratory where they were processed within two hours. Blood
23 clotting was allowed at room temperature for at least 30 minutes after collection. The sample
24 tubes were spun at 550g relative centrifugal force for 10 minutes at room temperature and
25 distributed into 0.5mL aliquots of plasma, serum, buffy coat (lymphocytes), whole blood
26 and/or an aliquot tube containing a blood clot (1.0mL FluidX screwcap tubes, Cheshire, UK)
27 and stored immediately at -80°C (Thermo Fisher Scientific, Waltham, USA). Each FluidX
28 tube contained a unique 2D barcode linked to the original collection tube and participant. As
29 each assessment centre closed, samples were shipped on dry ice to the Melbourne Children's
30 Bioresource Centre for long term storage at -80°C (serum, whole blood, plasma, blood clot)
31 or vapour phase liquid nitrogen (lymphocytes). At a later date, single 0.5ml serum aliquot
32 was removed for every CheckPoint participant and the combined aliquots were shipped in a
33 single batch to Nightingale Health (Helsinki, Finland) on dry ice for NMR metabolomics.

34
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1 **Measures**

2 3 Metabolomic profiling

4 The Nightingale[®] NMR metabolomics platform (Helsinki, Finland) was used to obtain
5 metabolomics for children and parents using the 2016-version quantification algorithm.
6 Details of this platform and methodology have been extensively described elsewhere,^{6 19} and
7 epidemiological applications were recently reviewed.²⁰ Briefly, metabolites were measured
8 from 0.35mL of serum using a single high-throughput experimental setup for the
9 simultaneous quantification of routine lipids, lipoprotein subclass distributions, particle size
10 and composition, fatty acids, and other low-molecular weight metabolites such as amino
11 acids and glycolysis-related metabolites. This generated data on 228 serum metabolite
12 measures in absolute concentration units (eg millimoles per liter) and ratios (summarised in
13 Table 1). Whilst widely used for epidemiological research, the NMR-based quantification has
14 not been certified for clinical diagnostics. Further analytical validation of the quantification
15 protocols for the biomarker subset routinely used in clinical settings (eg established
16 cholesterol measures and creatinine) is expected to lead to recalibration of certain metabolite
17 concentrations to better match clinical gold standards.²⁰

18

1 **Table 1. Summary of biomarkers and derived variables obtained via high-throughput NMR**

Metabolic group	Species and derived measures
Amino acids	Alanine, Glutamine, Glycine, Histidine
	Branched chain: Isoleucine, Leucine, Valine
	Aromatic: Phenylalanine, Tyrosine
Cholesterol	VLDL, LDL, HDL, HDL2, HDL3, Total, Free, Esterified, Remnant
Triglycerides and phospholipids	Triglycerides (VLDL, LDL, HDL, total)
	Phosphoglycerides
	Ratio of triglycerides to phosphoglycerides*
	Phosphatidylcholine
	Sphingomyelins
Total cholines	
Apolipoproteins	Apolipoprotein A-1 (ApoA-1)
	Apolipoprotein B (Apo B)
	Ratio of Apolipoprotein B to Apolipoprotein A-1 (ApoB/Apo A-1)*
Fatty acids (FA)	Total, Omega-3, Omega-6, Polyunsaturated (PUFA), Saturated (SFA)
	Monounsaturated (MUFA), Docosahexaenoic acid (DHA), Linoleic (LA)
	Estimated degree of unsaturation
Fatty acid ratios	Omega-3/total FA*, Omega-6/total FA*, PUFA/total FA* (all %)
	SFA/total FA*, MUFA/total FA*, DHA/total FA*, LA/total FA* (all %)
Lipoprotein subclasses*	12 lipid measures in each of 14 subclasses VLDL (XXL, XL, L, M, S, XS), IDL, LDL (L,M,S), HDL (XL, L, M,S): Particle concentration, Total lipids, Esterified cholesterol, Total cholesterol, Phospholipids, Free cholesterol, Triglycerides and Esterified cholesterol/Total lipids (%), Free cholesterol/Total lipids (%), Total cholesterol/Total lipids (%), Triglycerides/Total lipids (%) and Phospholipids/Total lipids (%).
Lipoprotein size*	Mean diameter of VLDL, LDL and HDL particles
Ketone bodies	Acetate, Acetoacetate, 3-hydroxybutyrate
Glycolysis related	Glucose, Lactate, Pyruvate, Citrate, Glycerol
Fluid balance	Creatinine, Albumin
Inflammation	Glycoprotein acetyls (GlycA)

2 Information obtained from <https://nightingalehealth.com/science/biomarkers>

3 * ratio; ApoA-1: Apolipoprotein A-1; Apo B: Apolipoprotein B; DHA: Docosahexaenoic acid; GlycA: Glycoprotein acetyls;
4 HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; L: Large; LDL: Low-density lipoprotein; LA:
5 Linoleic acid; M: Medium; MUFA: Monounsaturated fatty acid; PUFA: Polyunsaturated fatty acid; S: Small; SFA:
6 Saturated fatty acids; VLDL: Very low density lipoprotein; XL: Very large; XXL: Chylomicrons and extremely large; XS:
7 Very small.

8
9 Many of the 228 metabolomics measures correlate substantially both in adults
10 (supplementary figure 1) and children (supplementary figure 2) and the pattern of correlations
11 were similar for adults and children. For clarity, we therefore focused on a subset of 74
12 metabolites in analyses. We eliminated the 5 ratio measures for each of the 14 lipoprotein
13 subclass particles. In addition, the 7 other measures within each of the lipoproteins (esterified
14 cholesterol, free cholesterol, total cholesterol, triglycerides, phospholipids, total lipids and
15 particle concentration) are all highly correlated and therefore we only reported total lipids for
16 each of the lipoprotein subclass particles.

1 Other measures and sample characteristics

2 *Age and sex:* For children, LSAC provided date of birth (DOB) and sex, which was originally
3 exported from the Medicare Australia database. In parents, DOB and sex was self-reported in
4 the CheckPoint questionnaire, which was administered on an iPad using the Research
5 Electronic Data Capture (REDCap) tool.²¹ Age in years was calculated as the difference
6 between date of the CheckPoint assessment and DOB divided by 365.

7 *Disadvantage index:* LSAC provided contact details of families consenting to be contacted
8 by CheckPoint. The family's residential postcode was confirmed during the CheckPoint
9 recruitment phone call and updated, if required. The disadvantage index score of postcode
10 was used to summarise neighbourhood socioeconomic position. Generated by the ABS
11 from the 2011 national Census, the index numerically summarises the social and economic
12 conditions of Australian neighbourhoods; national mean 1000, standard deviation 100;
13 higher scores indicate less disadvantage.²²

14
15 *Time of blood collection, processing and fasting time:* Time of blood collection and start of
16 laboratory processing were recorded. When missing, collection time was estimated using the
17 midpoint between the time the CheckPoint visit began and time that processing of the sample
18 commenced. Processing lag time was calculated as the minutes between blood collection and
19 the processing commencement. Most samples were processed within two hours.

20 Fasting time was calculated as the hours between last eating/drinking to time of blood
21 collection. The last time of eating/drinking was cross-checked against when the participant
22 was taking part in other CheckPoint stations (and known not to be eating) as well as sleep and
23 wake times from accelerometry data (to identify usual activity, and therefore likely eating
24 patterns) when available. Further details of cleaning processes for the time of last eat/drink
25 can be found elsewhere.²³

26 **Statistical analysis**

27 Sample Characteristics

28
29 Continuous descriptive variables and metabolite measures were summarised using means and
30 standard deviations (SD) for children and adults separately, by sex and overall. For skewed
31 metabolites, geometric means and relative SD were reported. To provide visual comparisons
32 of distributions of metabolites by age and sex, density plots were used. Population summary
33 statistics were estimated by applying survey weights and survey procedures that corrected for
34 sampling, participation and non-response biases, and took into account clustering in the
35 sampling frame. Standard errors were calculated taking into account the complex design and
36 weights.²⁴ More detail on the calculation of weights is provided elsewhere.²⁵

1 Differences in metabolite concentration by age (adults compared to children) and by sex
2 (adults, children)

3 Skewed metabolites (skewness greater or equal to 2) were log-transformed. We used two-
4 sided paired and unpaired t-tests (as appropriate) to assess differences in mean metabolite
5 concentrations between adults and children in parent-child dyads, and between males and
6 females for adults and children separately. P-values were adjusted using Benjamini-
7 Hochberg (B-H) with a false discovery rate (FDR) of 10% to account for multiple
8 comparisons.

9 Parent-Child concordance

10 Concordance between parents and children was assessed by 1) Pearson's correlation
11 coefficients (CC) with 95% confidence intervals, and 2) partial correlation coefficients
12 (PCC), adjusting for child and parent age, disadvantage index, fasting time, processing lag
13 time (and for child and parent sex where appropriate). Scatterplots of parent versus child
14 metabolites (log-transformed where needed as above) were examined to check for outliers
15 and to ensure assumptions were met.

16 The analyses were repeated using weighted multi-level survey analyses and compared to
17 unweighted analyses. As there appeared to be no major effect of response patterns on
18 results we reported results from unweighted analyses. Analyses were undertaken using
19 Stata version 14.2 (StataCorp, College Station, TX) and R version 3.3.2.²⁶

20 **Patient and Public Involvement:** Because LSAC is a population-based longitudinal study,
21 no patient groups were involved in its design or conduct. To our knowledge, the public was
22 not involved in the study design, recruitment or conduct of LSAC study or its CheckPoint
23 module. Parents received a summary health report for their child and themselves at or soon
24 after the assessment visit. They consented to take part knowing that they would not otherwise
25 receive individual results about themselves or their child.

1 RESULTS

3 Sample characteristics

5 The recruitment and retention of participants in the Child Health CheckPoint are described
6 elsewhere.¹⁸ Of the 1874 families who participated in CheckPoint assessment centres, blood
7 serum samples of analysable quality from 1180 children and 1325 parents (figure 1) were
8 sent for NMR quantification of metabolites. The majority of excluded families undertook
9 home visits or attended a regional centre, where blood samples could not be collected (n=385,
10 20.5%), while some participants declined a blood sample (children, n=150, 8.0%; adults,
11 n=108, 5.8%). Few data were lost due to insufficient volume or poor quality samples at the
12 assessment centre (figure 1). The sample characteristics of parents and children are outlined
13 in table 2. Summary statistics for our main child and parent metabolite measures are
14 presented in supplementary table 1. Supplementary figures 3-7 show density plots comparing
15 the distributions of metabolites for boys, girls and adults.

Table 2: Sample characteristics; values are weighted mean (standard deviation)

Characteristic	All	Male	Female
Child			
n	1152-1180	558-575	594-605
Age, years	12.0 (0.4)	12.0 (0.4)	12.0 (0.4)
BMI, (kg/m ²)	19.4 (3.5)	19.2 (3.4)	19.6 (3.7)
BMI z-score	0.38 (1.0)	0.40 (1.0)	0.37 (1.0)
Disadvantage Index	1012 (63)	1011 (65)	1014 (61)
Fasting time (hours)	4.2 (1.2)	4.3 (1.3)	4.2 (1.1)
Time of day - blood collection	14.16 (2.0)	14.12 (2.0)	14.20 (2.1)
Processing lag time (hours)	1.16 (0.5)	1.18 (0.5)	1.14 (0.5)
Parent			
n	1272-1325	174-177	1098-1148
Age, years	43.9 (5.6)	46.9 (6.9)	43.4 (5.2)
BMI, (kg/m ²)	28.4 (6.4)	28.9 (4.7)	28.4 (6.6)
Fasting time (hours)	3.3 (1.6)	3.6 (2.0)	3.2 (1.5)
Time of day - blood collection	13.10 (2.0)	13.18 (2.1)	13.09 (2.0)
Processing lag time (hours)	1.26 (0.5)	1.31 (0.5)	1.26 (0.5)

Disadvantage Index: Index of Relative Socioeconomic Disadvantage; n: number of participants in cohort with this measure.

Differences in metabolite levels - adults compared to children

Figure 2 shows mean differences in metabolite levels for adults relative to children in standard deviation (SD) units. Most concentrations were higher in adults than children. Values that were similar in adults and children included total lipids in very large HDL lipoprotein subclass particles, acetoacetate, tyrosine and glucose. Levels in children were higher than those of adults for the majority of glycolysis related measures (lactate, pyruvate, citrate and glycerol), the ketone body 3-hydroxybutrate, the amino acid glutamine, many fatty acid ratios and all lipoprotein particle sizes. Supplementary table 2 lists the corresponding estimates in absolute concentration units.

Sex differences in metabolite levels in children and adults

Figure 3 shows differences in mean metabolite levels by sex for children and adults separately in SD units, with estimates in absolute concentration units listed in supplementary table 3 and 4.

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3 1 In general, sex differences were more pronounced in adulthood, resulting in distinct overall
4 2 patterns for children and adults. Children generally showed smaller differences by sex than
5 3 adults. Of note, sex differences for apolipoproteins and fatty acid measures showed different
6 4 patterns in children compared to adults.

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10 5 Girls had lower levels of apolipoprotein-A-1 (ApoA-1) and higher ApoB than boys. In adults,
11 6 the opposite pattern was observed with females having higher ApoA-1 and lower ApoB than
12 7 males. In children, some fatty acid concentrations were higher in girls than boys. In contrast,
13 8 many adult fatty acid measures were higher in males. There was no evidence of a difference
14 9 in the level of inflammation (GlycA) by sex in children, while in adults, GlycA levels tended
15 10 to be higher in males than females.

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20 11 For some metabolites, sex differences in children mirrored (but were smaller in magnitude
21 12 than) those of adults, particularly for the ketone body acetate and some key amino acids. At
22 13 both ages, the amino acid glycine was higher in females but the branched-chain amino acids
23 14 leucine and valine tended to be higher in males.

24 15 **Parent-child concordance**

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29 16 Figure 4 shows the correlations between metabolite measures for all children with all parents,
30 17 and for boys and girls with mothers (but not with the 177 fathers, given the small numbers).
31 18 The corresponding correlation coefficients and partial correlation coefficients are listed in
32 19 supplementary tables 5 and 6.

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36 20 Correlations for all parents and all children showed similar patterns to that observed for
37 21 mother and child by sex. While there was little suggestion of substantial correlation within
38 22 parent-child dyads for some metabolites (eg glucose, acetate) a positive correlation was found
39 23 for many metabolite measures irrespective of child sex. For example, positive correlations
40 24 were observed for isoleucine (CC 0.33, 95% CI 0.27 to 0.38), total serum cholesterol (CC
41 25 0.30, 95% CI 0.24 to 0.35) and omega 6 fatty acids (CC 0.28, 95% CI 0.23 to 0.34) in parent-
42 26 child comparisons. Additional adjustment for factors that potentially influence metabolite
43 27 levels (age, socioeconomic status, fasting time and processing lag time) had little effect on
44 28 the degree of correlation in any comparison (supplementary tables 5 and 6).

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1 DISCUSSION

2 Principal findings

3 Here we present age and sex differences, describing the distribution of detailed/NMR-based
4 metabolite measures in Australian 11-12 year old children and their parents, and demonstrate
5 that many metabolite measures have moderate parent-child concordance and in general there
6 is a high level of agreement in the magnitude of concordance across metabolites. In accord
7 with previous studies, we observed major differences in metabolite levels between childhood
8 and adulthood and also differences by sex in both childhood and adulthood. We also
9 observed variability in the magnitude of differences by sex for several metabolites in
10 childhood compared to adulthood and identified a complex interplay of correlations of
11 specific metabolites between parents and their children according to parent-child sex
12 relationships.

13 Strengths and weaknesses

14 This is the first major cohort study to report both sex- and cross-generational differences in
15 metabolomic concentrations in mid-childhood to adulthood utilising the NMR platform.
16 Further strengths include the large number of parent-child dyads representing a wide range of
17 parent ages, the national population-based sample and the state-of-the-art measurements.
18 Replication studies exploring sex differences at earlier and later stages of childhood and
19 adolescence would strengthen findings.

20 An important limitation is that paternal factors were not fully represented, as most parental
21 samples were from mothers (a well-documented problem in longitudinal cohort studies). This
22 also limited sex-specific parental contribution analysis; further studies including more fathers
23 are warranted. Additional limitations are that, without samples from both parents for each
24 child, we could not estimate heritability, and our results might not apply to mid-life adults
25 who are not parents (although we see no good reason why these would differ greatly). The
26 original uptake of just over 50% and subsequent attrition within LSAC and then the
27 CheckPoint has led to a relatively advantaged sample, but nonetheless participants varied
28 widely on key potential confounders (eg disadvantage, age) and this was at least partly offset
29 by application or consideration of survey weights. Given the large number of metabolites and
30 modest sample size, considerable uncertainty remains in any ranking of the various effects
31 across metabolites. In addition, given the descriptive aims of the paper, additional factors and
32 potential confounders not considered could explain some of the results observed.

1 **Meaning and implications for clinicians and policymakers**

2 Overall, we found a difference in metabolite profile between children and their parents. This
3 was apparent for specific metabolite measures (such as some amino acids) as well as the
4 distribution of metabolites (such as lipid composition of lipoproteins of different density).
5 Some measures were higher in adults, some similar, while a minority were lower. Previous
6 studies, largely in adults, have identified a range of specific metabolite changes with age,
7 particularly from mid to late adulthood.²⁷ This includes a general decrease in several amino
8 acid species, which contrasts with our findings from childhood to mid adulthood.⁸ Only the
9 amino acid glutamine showed this pattern in our dataset.

10 Differences in children by sex (± 0.2 SD) were generally much smaller than in adults (± 0.8
11 SD). Large metabolomic studies using alternative platforms have previously reported
12 reproducible, sex-specific signatures in circulating metabolite profile in adults.²⁸⁻²⁹ This
13 includes differences in amino acid and lipid serum concentrations, potentially influenced by
14 sex-specific effects of genetic polymorphisms on metabolite levels.²⁹⁻³⁰ As in our study, most
15 amino acids have usually been reported to be higher in men than women.²⁹⁻³¹ For example, in a
16 recent study of 507 metabolic markers in 1756 individuals (903 female and 853 male aged
17 ~ 60 years), one third of metabolites showed significant sexual dimorphism. These were
18 predominantly related to pathways of steroid metabolism, fatty acids, other lipids, and a large
19 proportion of amino acids.³¹ Of particular note, branched chain amino acids (BCAAs) and
20 their related metabolic products were amongst the most differentially represented, with much
21 higher isoleucine, leucine and valine in males. A similar finding of higher leucine and valine
22 was also noted in the Cooperative Health Research in the Region of Augsburg (KORA)
23 follow-ups 3 (F3) and 4 (F4) analysis of >3000 adults,²⁹ consistent with our observations in
24 adulthood.

25 In children, we found that sex differences for leucine and valine were smaller but in the same
26 direction as adults. Several lines of evidence implicate BCAA metabolism with metabolic
27 risk in humans. For example, three candidate genes for obesity and/or type 2 diabetes
28 mellitus (T2DM) are involved in the BCAA metabolic pathway.³² In a recent large meta-
29 analysis of metabolomics in diabetes, a $>30\%$ higher risk of type 2 diabetes was found per
30 SD increase in isoleucine, leucine, valine or tyrosine, whereas glycine and glutamine were
31 inversely associated with risk.³² Several clinical studies have also reported that BCAAs
32 positively correlate with insulin resistance, homeostatic model assessment (HOMA) index
33 and levels of haemoglobin A1c (HbA1c), while longitudinal studies have reported that

1 increased blood BCAAs are predictive of future insulin resistance and type 2 diabetes
2 (T2D).³³ It is intriguing to speculate that the higher BCAA in males from early life could
3 contribute to the well-described increasing prevalence of T2D in men. Levels of BCAA are
4 elevated in females with Polycystic Ovary Syndrome (PCOS), potentially contributing to the
5 associated insulin resistance.³⁴ However, it remains unclear whether BCAA are on the causal
6 pathway to T2D or result from adverse metabolic health. Our demonstration that the sex
7 differences in BCAA possibly arise early in life offers potential to track their association with
8 sex-specific measures of metabolic health from an early age to help clarify where they lie on
9 the causal pathway.

10 In accord with previous adult studies²⁹, we found higher levels of glycine in mothers than
11 fathers, and (less markedly) in girls than boys. Interestingly, recent metabolomics and genetic
12 analyses of ~10,000 adults with cardiovascular disease (CVD), with replication in >53,000
13 subjects, identified a genetic variant in carbamoyl-phosphate synthase 1 (*CPS1*) (linked to
14 plasma glycine levels) to be strongly associated with a reduced risk of CVD in women
15 ($p=6.3 \times 10^{-5}$) but not men ($p=0.95$), suggesting a direct link between glycine levels and CVD
16 risk, although whether this is a causal association remains unclear.³⁵ It will be interesting in
17 the future to explore the link between variants in *CPS1* and circulating glycine levels from
18 early life to adulthood in relation to markers of cardiovascular health in females.

19 The small sex-differences of HDL cholesterol and ApoA-1 in children compared to adults is
20 consistent with modest differences in children, whereas substantial differences in adulthood
21 have previously been reported.³⁶ ApoA-1 was more abundant in boys, while ApoB was
22 higher in girls, leading to a higher ApoB/ApoA-1 ratio in girls. The opposite pattern was
23 found in our limited sample of fathers relative to mothers. These data are surprising and
24 differ from a similarly sized study of slightly older European adolescent children (mean age
25 15 years) that found higher ApoA-1 and ApoB in girls relative to boys.³⁷ Interestingly, a
26 higher ApoB/ApoA-1 ratio has been strongly linked to increased coronary risk in adults,³⁸⁻⁴⁰
27 suggesting that sex differences may alter with increasing age, in keeping with the increased
28 CVD risk in adult males. ApoA-1 is the main protein component of HDL cholesterol⁴¹ thus
29 the differences in trajectories in lipids and HDL cholesterol for boys and girls across
30 childhood that have been reported^{42 43} could partially explain this observation.

31 These are the first data on the mother-child or parent-child correlations of NMR metabolites.
32 Smaller studies have reported positive correlations between parents and children for a limited
33 range of cardiometabolic risk factors including total cholesterol, LDL cholesterol, HDL

1 cholesterol and triglycerides measured using conventional methods. We found positive
2 correlations between parents and children for the same lipid measures (although measured
3 using NMR) consistent with previously reported findings. One study reported a positive
4 association between the serum lipid levels of 4 year old children (n=127) and their parents
5 (122 mothers and 118 fathers)⁴⁴ while another study of children aged 6-18 (n=255) and their
6 parents (n=179) found that the age of the child influenced the degree of correlation of several
7 lipid measures, with older (10-18 years) children more similar to their parents in terms of
8 triglyceride levels than younger individuals (6-9 years).¹²

9 **Unanswered questions and future research**

10 The temporal and sex specific dynamism of the metabolomics data we describe here offer
11 considerable opportunities for identification of biomarkers of risk for a range of non-
12 communicable diseases early in life, to inform targeted interventions and monitor their
13 efficacy. Combining metabolomics with other 'omics data (such as genetics), as is
14 increasingly reported from large adult studies, offers considerable promise in understanding
15 the causal pathways that link early life exposures, genetics and intermediate phenotypes with
16 later onset chronic disease, and in identifying clinically relevant biomarkers.

17 In conclusion, we describe the metabolite profile from mid-childhood and adulthood in a
18 population-based sample, together with parent-child concordance and differences by sex in
19 children and adults. In this descriptive paper, distinct differences in profiles were observed by
20 age and sex, as well as considerable evidence of correlation between parent and child
21 measures. These data will be informative for investigation of the childhood origins of adult
22 non-communicable diseases and for comparative studies across populations.

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3 Australian Children. The study is conducted in partnership between the Department of Social
4 Services (DSS), the Australian Institute of Family Studies (AIFS) and the Australian Bureau
5 of Statistics (ABS). The findings and views reported in this paper are those of the author and
6 should not be attributed to DSS, AIFS or the ABS.

7
8 REDCap (Research Electronic Data Capture) tools were used in this study. More
9 information about this software can be found at: www.project-redcap.org.

10 We thank the LSAC and CheckPoint study participants, staff and students for their
11 contributions.

12 **COMPETING INTERESTS**

13
14 All authors have completed the ICMJE uniform disclosure form at
15 www.icmje.org/doi_disclosure.pdf and declare financial support as described in the funding
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30 played no role in the conduct or analysis of the trial. DSS played a role in study design;
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32 management, analysis, and interpretation; preparation, review, or approval of the manuscript;

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2 the Victorian Government's Operational Infrastructure Support Program.

3

4 **CONTRIBUTIONS**

5
6 DB, RS and JC conceptualised and developed the Metabolomics Checkpoint study. SE and
7 JC undertook all aspects of data analysis. SAC coordinated the acquisition of metabolomics
8 data and provided critical review of this manuscript. MW, the Principal Investigator of the
9 Child Health CheckPoint, planned the analyses and provided critical review of this
10 manuscript. SE and RS drafted the manuscript. PW, MJ, TD, KL, JC, DB provided critical
11 expert advice and critical review of this manuscript.

12 **DATA SHARING STATEMENT**

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14 Dataset and technical documents available from *Growing Up in Australia: The Longitudinal*
15 *Study of Australian Children* via low-cost license for bona fide researchers. More information
16 is available at www.growingupinaustralia.gov.au

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1 **FIGURE CAPTIONS AND FOOTNOTES**

2 **Figure 1: Participant flow chart.**

3 n=number of families, c=number of children, p=number of attending adults,

4 MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,

5 LSAC=Longitudinal Study of Australian Children

6 *Unable to analyse due to insufficient volume or poor quality sample

7 ^Data from 6 non-biological child-parent pairs excluded from concordance analyses

8 **Figure 2: Differences in metabolite levels between children and adults.**

9 Association measures are SD difference in metabolite concentration for adults compared to
10 children. Error bars represent 95% confidence intervals. Significant associations after p-
11 values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold
12 (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals
13 and associated p-values are listed in supplementary table 2. HDL: High-density lipoprotein;
14 IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low
15 density lipoprotein.

16 **Figure 3: Sex differences in metabolite levels in childhood and adulthood.**

17 Association measures are SD difference in metabolite concentration for females compared to
18 males in children (A) and adults (B). Error bars represent 95% confidence intervals.
19 Significant associations after p-values adjusted for multiple testing using Benjamini-
20 Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute
21 concentration units, 95% confidence intervals and associated p-values are listed in
22 supplementary table 3 and 4. HDL: High-density lipoprotein; IDL: Intermediate density
23 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

24 **Figure 4: Parent:child correlation for metabolite measures.**

25 Pearson's correlation coefficients for all children with all parents (A); and for boys (blue)
26 with mothers and for girls (red) with mothers (B). Error bars represent 95% confidence
27 intervals. Correlation coefficients with associated 95% confidence intervals are listed in

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1 supplementary table 5 and 6. HDL: High-density lipoprotein; IDL: Intermediate density
2 lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.
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SUPPLEMENTARY DOCUMENTS

Supplementary figure 1: Correlation of NMR measures in children.

Heatmap showing the correlation between metabolite measures in children. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 2: Correlation of NMR metabolite measures in parents.

Heatmap showing the correlation between metabolite measures in parents. The correlations shown are Spearman's correlation coefficients with blue cells representing negative correlations and red cells representing positive correlations.

Supplementary figure 3: Density plots for cholesterol and apolipoprotein measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each cholesterol and apolipoprotein measure.

Supplementary figure 4: Density plots for fatty acid and fatty acid ratio measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for each fatty acid and fatty acid ratio measure.

Supplementary figure 5: Density plots for total lipids in the 14 lipoprotein subclass particles.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for total lipids within each of the 14 lipoprotein subclass particles.

Supplementary figure 6: Density plots for lipoprotein particle size and triglyceride measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for lipoprotein particle sizes and triglyceride measures.

Supplementary figure 7: Density plots for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

Boys (blue), girls (red), and all parents (thin dotted black line) plotted on the same graph for glycolysis related, amino acid, ketone body, fluid balance and inflammation measures.

Supplementary table 1: Weighted mean (SD) of metabolite measures in children and parents.

Supplementary table 2: Mean difference in metabolite levels in adults compared to children in absolute concentration units.

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1 **Supplementary table 3: Differences in mean metabolite levels in girls compared to boys**
2 **in absolute concentration units.**

4 **Supplementary table 4: Differences in mean metabolite levels in female compared to**
5 **male adults in absolute concentration units.**

7 **Supplementary table 5: Mother-child concordance; correlations and partial correlations**
8 **between mothers and their sons, daughters and all children.**

10 **Supplementary table 6: Parent-child concordance; correlation and partial correlations**
11 **between all parents and their sons, daughters and all children.**

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1 REFERENCES

1. Nicholson JK, Lindon JC, Holmes E. 'Metabonomics': understanding the metabolic responses of living systems to pathophysiological stimuli via multivariate statistical analysis of biological NMR spectroscopic data. *Xenobiotica* 1999;29(11):1181-9. doi: 10.1080/004982599238047
2. Nath AP, Ritchie SC, Byars SG, et al. An interaction map of circulating metabolites, immune gene networks, and their genetic regulation. *Genome biology* 2017;18(1):146. doi: 10.1186/s13059-017-1279-y
3. Shah SH, Newgard CB. Integrated metabolomics and genomics: systems approaches to biomarkers and mechanisms of cardiovascular disease. *Circ Cardiovasc Genet* 2015;8(2):410-9. doi: 10.1161/CIRCGENETICS.114.000223
4. Kaikkonen JE, Wurtz P, Suomela E, et al. Metabolic profiling of fatty liver in young and middle-aged adults: Cross-sectional and prospective analyses of the Young Finns Study. *Hepatology* 2017;65(2):491-500. doi: 10.1002/hep.28899
5. Johnson CH, Ivanisevic J, Siuzdak G. Metabolomics: beyond biomarkers and towards mechanisms. *Nat Rev Mol Cell Biol* 2016;17(7):451-9. doi: 10.1038/nrm.2016.25
6. Kettunen J, Tukiainen T, Sarin AP, et al. Genome-wide association study identifies multiple loci influencing human serum metabolite levels. *Nat Genet* 2012;44(3):269-76. doi: 10.1038/ng.1073
7. Suhre K, Gieger C. Genetic variation in metabolic phenotypes: study designs and applications. *Nat Rev Genet* 2012;13(11):759-69. doi: 10.1038/nrg3314
8. Yu Z, Zhai G, Singmann P, et al. Human serum metabolic profiles are age dependent. *Aging Cell* 2012;11(6):960-7. doi: 10.1111/j.1474-9726.2012.00865.x
9. Auro K, Joensuu A, Fischer K, et al. A metabolic view on menopause and ageing. *Nat Commun* 2014;5:4708. doi: 10.1038/ncomms5708
10. Saito K, Maekawa K, Kinchen JM, et al. Gender- and Age-Associated Differences in Serum Metabolite Profiles among Japanese Populations. *Biol Pharm Bull* 2016;39(7):1179-86. doi: 10.1248/bpb.b16-00226
11. Wang Q, Wurtz P, Auro K, et al. Metabolic profiling of pregnancy: cross-sectional and longitudinal evidence. *BMC Med* 2016;14(1):205. doi: 10.1186/s12916-016-0733-0
12. Halvorsen T, Moran A, Jacobs DR, Jr., et al. Relation of Cardiometabolic Risk Factors between Parents and Children. *J Pediatr* 2015;167(5):1049-56 e2. doi: 10.1016/j.jpeds.2015.07.053
13. Rueedi R, Ledda M, Nicholls AW, et al. Genome-Wide Association Study of Metabolic Traits Reveals Novel Gene-Metabolite-Disease Links. *PLOS Genetics* 2014;10(2):e1004132. doi: 10.1371/journal.pgen.1004132
14. Kettunen J, Demirkan A, Würtz P, et al. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nature Communications* 2016;7:11122. doi: 10.1038/ncomms11122
<https://www.nature.com/articles/ncomms11122#supplementary-information>
15. Edwards B. Growing Up in Australia: The Longitudinal Study of Australian Children: Entering adolescence and becoming a young adult. *Family Matters* 2014(95):5-14.
16. Sanson A, Johnstone R. The LSAC Research Consortium & FaCS LSAC Project Team. Growing Up in Australia takes its first steps. *Family Matters* 2004;67:46-53.
17. Wake M, Clifford SA, York E, et al. Introducing Growing Up in Australia's Child Health CheckPoint. *Family Matters* 2014;94:15-23.

18. Clifford SA, Davies S, Wake M. Child Health CheckPoint: Cohort summary and methodology of a physical health and biospecimen module for the Longitudinal Study of Australian Children. Submitted to BMJ Open October 2017.
19. Soininen P, Kangas AJ, Wurtz P, et al. High-throughput serum NMR metabolomics for cost-effective holistic studies on systemic metabolism. *Analyst* 2009;134(9):1781-5. doi: 10.1039/b910205a
20. Wurtz P, Kangas AJ, Soininen P, et al. Quantitative Serum NMR Metabolomics in Large-Scale Epidemiology: A Primer on -Omic Technology. *Am J Epidemiol* 2017 [published Online First: 10 May 2017]
21. Harris PA, Taylor R, Thielke R, et al. Research electronic data capture (REDCap)--a metadata-driven methodology and workflow process for providing translational research informatics support. *J Biomed Inform* 2009;42(2):377-81. doi: 10.1016/j.jbi.2008.08.010 [published Online First: 2008/10/22]
22. Australian Bureau of S. Census of population and housing: Socio-Economic Indexes for Areas (SEIFA) 2011. Cat. no. 2033.0.55.001, 2011.
23. Davies S, Clifford S, Gillespie A, et al. LSAC's Child Health CheckPoint Data Issues Paper 2018. *Melbourne: Murdoch Children's Research Institute* 2018 doi: 10.25374/MCRI.5821230.
24. Heeringa SG, West BT, Berglund PA. Applied survey data analysis. Boca Raton.: CRC press 2010.
25. Ellul S, Hiscock R, Mensah FK, et al. Longitudinal Study of Australian Children's Child Health CheckPoint Technical Paper 1: Weighting and Non-Response. *Melbourne: Murdoch Children's Research Institute* 2018 doi: <https://doi.org/10.25374/MCRI.5687593>
26. R: A language and environment for statistical computing [program]. Vienna, Austria: R Foundation for Statistical Computing, 2018.
27. Menni C, Kastenmüller G, Petersen AK, et al. Metabolomic markers reveal novel pathways of ageing and early development in human populations. *International Journal of Epidemiology* 2013;42(4):1111-19. doi: 10.1093/ije/dyt094
28. Dunn WB, Lin W, Broadhurst D, et al. Molecular phenotyping of a UK population: defining the human serum metabolome. *Metabolomics* 2015;11:9-26. doi: 10.1007/s11306-014-0707-1
29. Mittelstrass K, Ried JS, Yu Z, et al. Discovery of sexual dimorphisms in metabolic and genetic biomarkers. *PLoS Genet* 2011;7(8):e1002215. doi: 10.1371/journal.pgen.1002215
30. Kolz M, Johnson T, Sanna S, et al. Meta-analysis of 28,141 individuals identifies common variants within five new loci that influence uric acid concentrations. *PLoS Genet* 2009;5(6):e1000504. doi: 10.1371/journal.pgen.1000504
31. Krumsiek J, Mittelstrass K, Do KT, et al. Gender-specific pathway differences in the human serum metabolome. *Metabolomics* 2015;11(6):1815-33. doi: 10.1007/s11306-015-0829-0
32. Guasch-Ferré M, Hruby A, Toledo E, et al. Metabolomics in Prediabetes and Diabetes: A Systematic Review and Meta-analysis. *Diabetes Care* 2016;39(5):833-46. doi: 10.2337/dc15-2251
33. Lynch CJ, Adams SH. Branched-chain amino acids in metabolic signalling and insulin resistance. *Nature reviews Endocrinology* 2014;10(12):723-36. doi: 10.1038/nrendo.2014.171
34. Chang AY, Lalia AZ, Jenkins GD, et al. Combining a nontargeted and targeted metabolomics approach to identify metabolic pathways significantly altered in

- 1 polycystic ovary syndrome. *Metabolism* 2017;71(Supplement C):52-63. doi:
2 <https://doi.org/10.1016/j.metabol.2017.03.002>
- 3 35. Hartiala JA, Wilson Tang WH, Wang Z, et al. Genome-wide association study and
4 targeted metabolomics identifies sex-specific association of CPS1 with coronary
5 artery disease. *Nature Communications* 2016;7:10558. doi: 10.1038/ncomms10558
- 6 36. Davis CE, Williams DH, Oganov RG, et al. Sex Difference in High Density Lipoprotein
7 Cholesterol in Six Countries. *American Journal of Epidemiology* 1996;143(11):1100-
8 06. doi: 10.1093/oxfordjournals.aje.a008686
- 9 37. Spinneker A, Egert S, Gonzalez-Gross M, et al. Lipid, lipoprotein and apolipoprotein
10 profiles in European adolescents and its associations with gender, biological maturity
11 and body fat--the HELENA Study. *Eur J Clin Nutr* 2012;66(6):727-35. doi:
12 10.1038/ejcn.2011.222
- 13 38. Walldius G, Jungner I, Aastveit AH, et al. The apoB/apoA-I ratio is better than the
14 cholesterol ratios to estimate the balance between plasma proatherogenic and
15 antiatherogenic lipoproteins and to predict coronary risk. *Clin Chem Lab Med*
16 2004;42(12):1355-63. doi: 10.1515/CCLM.2004.254
- 17 39. Walldius G, Jungner I. The apoB/apoA-I ratio: a strong, new risk factor for
18 cardiovascular disease and a target for lipid-lowering therapy--a review of the
19 evidence. *J Intern Med* 2006;259(5):493-519. doi: 10.1111/j.1365-2796.2006.01643.x
- 20 40. Mozaffarian D, Benjamin EJ, Go AS, et al. Heart disease and stroke statistics--2015
21 update: a report from the American Heart Association. *Circulation* 2015;131(4):e29-
22 322. doi: 10.1161/CIR.0000000000000152
- 23 41. Upadhyay RK. Emerging Risk Biomarkers in Cardiovascular Diseases and Disorders.
24 *Journal of Lipids* 2015;2015:971453. doi: 10.1155/2015/971453
- 25 42. Hardy R, Lawlor DA, Kuh D. A life course approach to cardiovascular aging. *Future*
26 *cardiology* 2015;11(1):101-13. doi: 10.2217/fca.14.67
- 27 43. Jolliffe CJ, Janssen I. Distribution of Lipoproteins by Age and Gender in Adolescents.
28 *Circulation* 2006;114(10):1056.
- 29 44. Ohlund I, Hernell O, Hornell A, et al. Serum lipid and apolipoprotein levels in 4-year-old
30 children are associated with parental levels and track over time. *Eur J Clin Nutr*
31 2011;65(4):463-69.
32

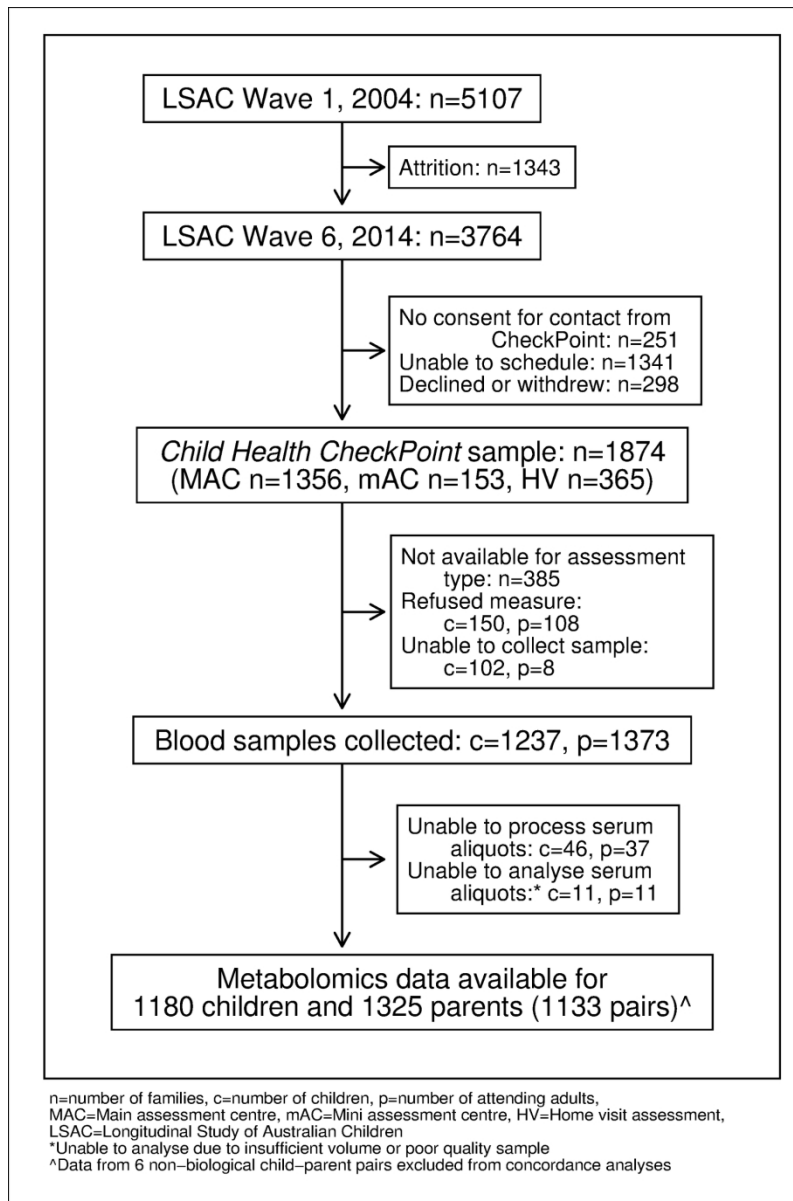


Figure 1: Participant flow chart.

n=number of families, c=number of children, p=number of attending adults,
MAC=Main assessment centre, mAC=Mini assessment centre, HV=Home visit assessment,
LSAC=Longitudinal Study of Australian Children

*Unable to analyse due to insufficient volume or poor quality sample

^Data from 6 non-biological child-parent pairs excluded from concordance analyses

76x114mm (600 x 600 DPI)

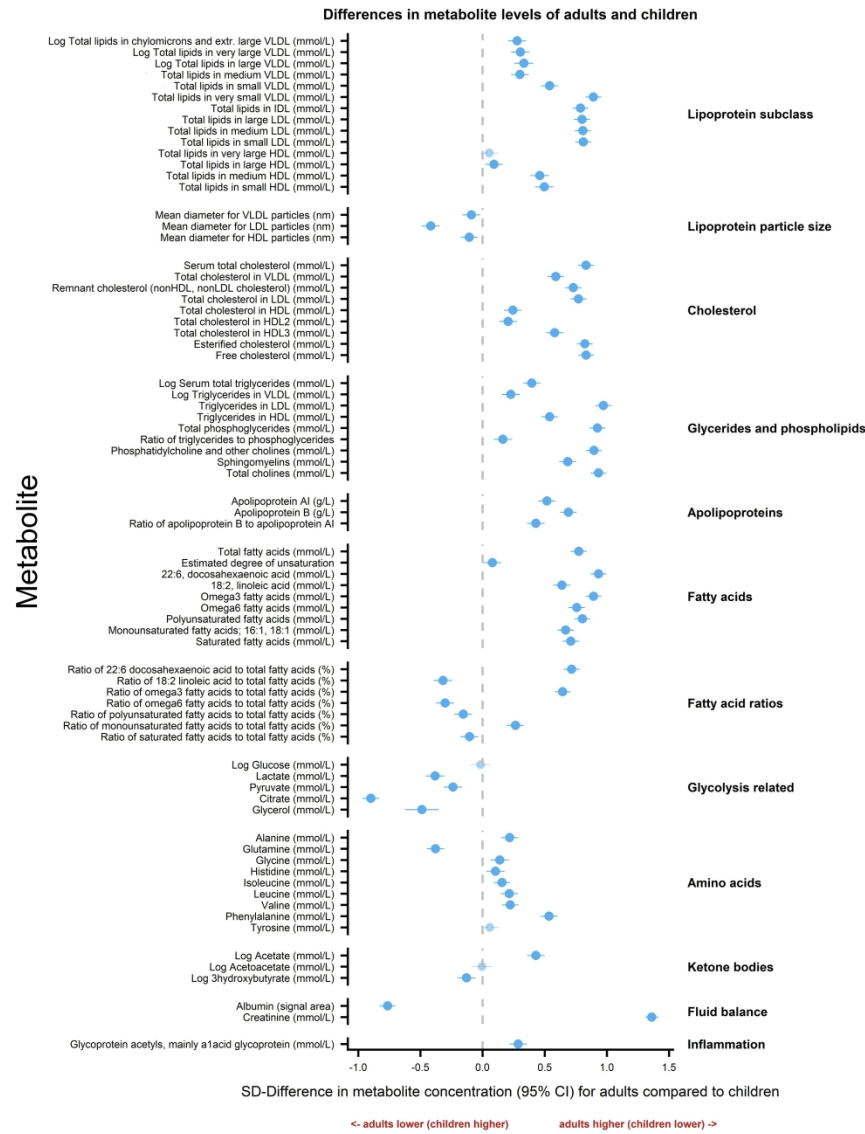


Figure 2: Differences in metabolite levels between children and adults.

Association measures are SD difference in metabolite concentration for adults compared to children. Error bars represent 95% confidence intervals. Significant associations after p-values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals and associated p-values are listed in supplementary table 2. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

254x338mm (300 x 300 DPI)

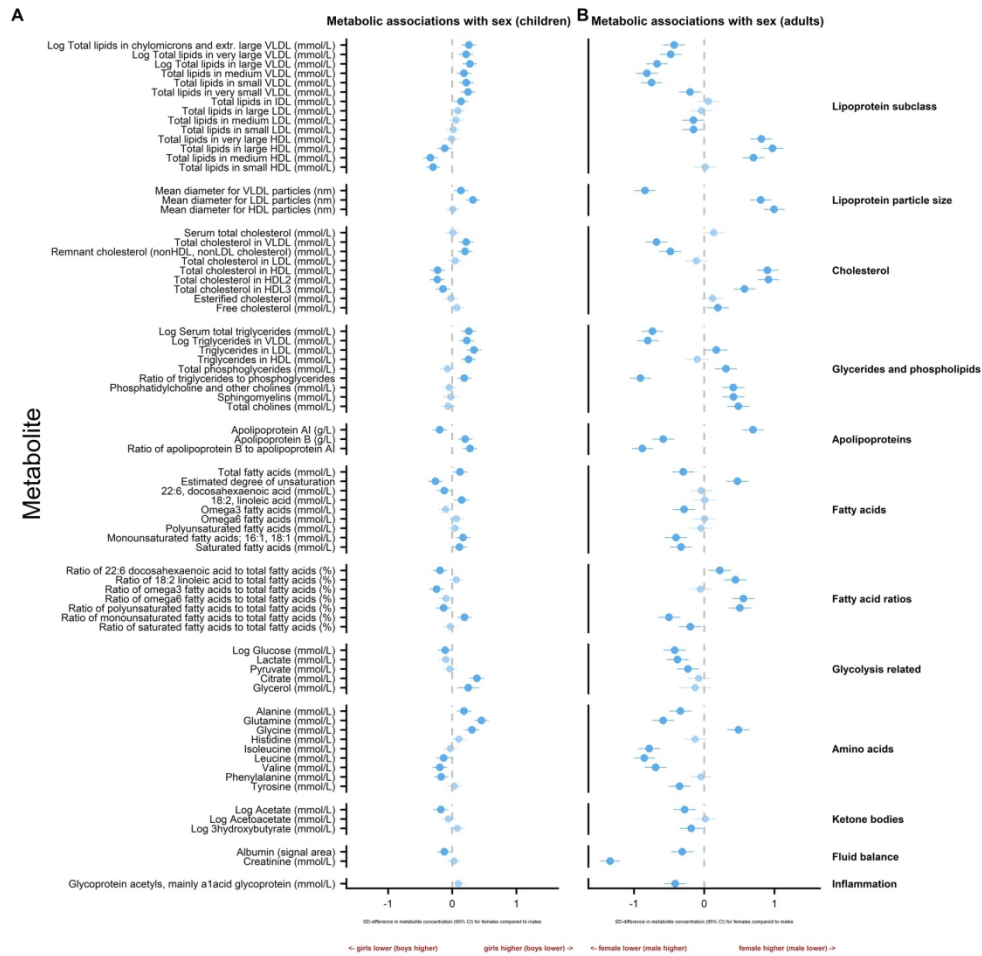


Figure 3: Sex-specific differences in metabolite levels in childhood and adulthood.

Association measures are SD difference in metabolite concentration for females compared to males in children (A) and adults (B). Error bars represent 95% confidence intervals. Significant associations after p-values adjusted for multiple testing using Benjamini-Hochberg procedure are shown in bold (FDR=0.10). Association measures in absolute concentration units, 95% confidence intervals and associated p-values are listed in supplementary table 3 and 4. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

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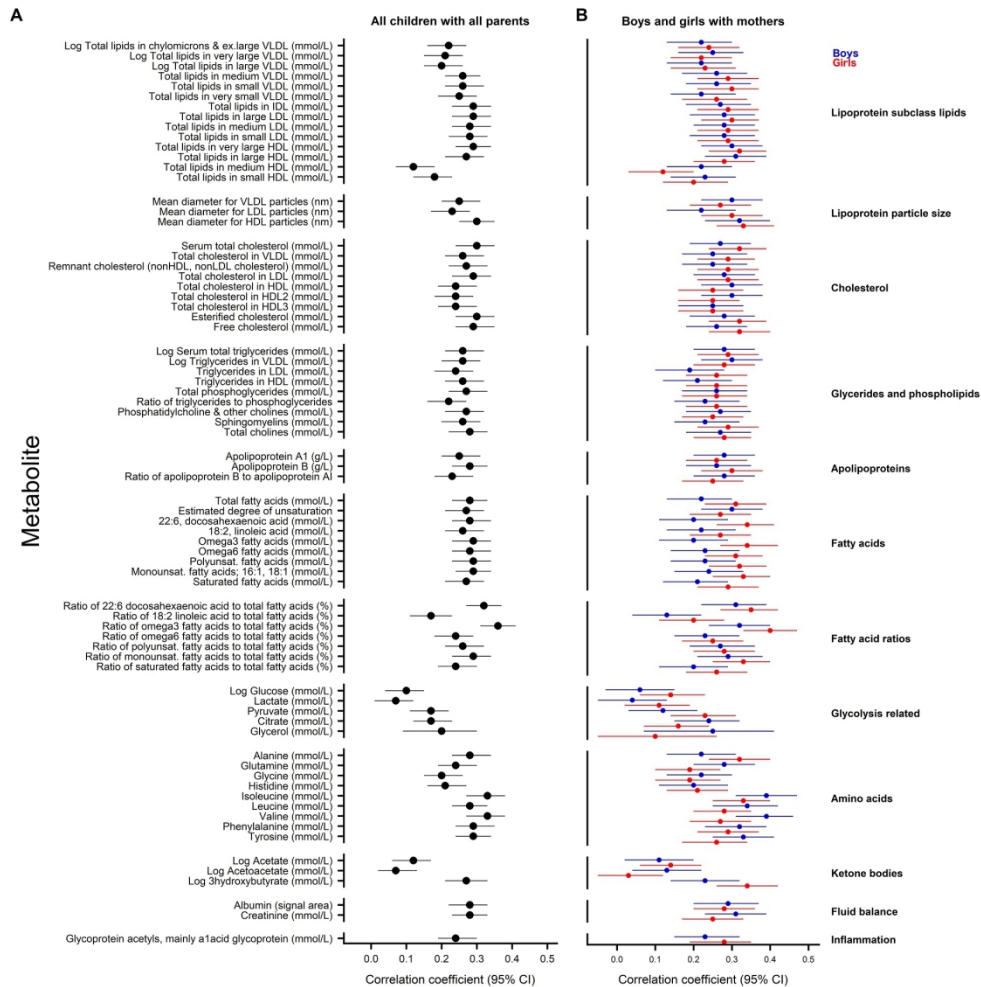


Figure 4: Parent:child correlation for metabolite measures. Pearson's correlation coefficients for all children with all parents (A); and for boys (blue) with mothers and for girls (red) with mothers (B). Error bars represent 95% confidence intervals. Correlation coefficients with associated 95% confidence intervals are listed in supplementary table 5 and 6. HDL: High-density lipoprotein; IDL: Intermediate density lipoprotein; LDL: Low-density lipoprotein; VLDL: Very low density lipoprotein.

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Supplementary table 1: Weighted mean (SD)* of metabolite measures in children and parents.

Metabolic subgroup	Children									Adults								
	Male			Female			All			Male			Female			All		
	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD	n	Mean	SD
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	575	0.005	4.826	605	0.011	3.260	1180	0.007	4.054	177	0.040	2.101	1148	0.013	2.822	1325	0.015	2.777
Total lipids in very large VLDL (mmol/L)*	575	0.006	6.246	605	0.014	4.800	1180	0.010	5.521	177	0.087	2.419	1148	0.022	3.458	1325	0.027	3.398
Total lipids in large VLDL (mmol/L)*	575	0.059	3.945	605	0.121	2.488	1180	0.085	3.234	177	0.423	1.287	1148	0.161	1.666	1325	0.182	1.665
Total lipids in medium VLDL (mmol/L)	575	0.441	0.270	605	0.478	0.272	1180	0.460	0.271	177	0.959	0.630	1148	0.548	0.366	1325	0.602	0.432
Total lipids in small VLDL (mmol/L)	575	0.381	0.153	605	0.405	0.146	1180	0.393	0.149	177	0.690	0.279	1148	0.494	0.216	1325	0.520	0.234
Total lipids in very small VLDL (mmol/L)	575	0.325	0.070	605	0.342	0.076	1180	0.334	0.073	177	0.451	0.111	1148	0.426	0.110	1325	0.429	0.110
Total lipids in IDL (mmol/L)	575	0.804	0.176	605	0.834	0.183	1180	0.819	0.180	177	0.985	0.261	1148	0.999	0.240	1325	0.997	0.242
Total lipids in large LDL (mmol/L)	575	0.917	0.220	605	0.941	0.228	1180	0.929	0.224	177	1.162	0.327	1148	1.155	0.298	1325	1.156	0.301
Total lipids in medium LDL (mmol/L)	575	0.511	0.136	605	0.519	0.140	1180	0.515	0.138	177	0.676	0.214	1148	0.655	0.185	1325	0.658	0.189
Total lipids in small LDL (mmol/L)	575	0.338	0.083	605	0.340	0.087	1180	0.339	0.085	177	0.439	0.135	1148	0.425	0.114	1325	0.427	0.117
Total lipids in very large HDL (mmol/L)	575	0.482	0.196	605	0.495	0.184	1180	0.488	0.189	177	0.320	0.189	1148	0.497	0.229	1325	0.474	0.232
Total lipids in large HDL (mmol/L)	575	0.874	0.291	605	0.859	0.275	1180	0.866	0.282	177	0.509	0.335	1148	0.900	0.382	1325	0.849	0.399
Total lipids in medium HDL (mmol/L)	575	0.917	0.127	605	0.871	0.126	1180	0.894	0.128	177	0.828	0.241	1148	0.971	0.175	1325	0.952	0.191
Total lipids in small HDL (mmol/L)	575	1.039	0.103	605	0.997	0.115	1180	1.018	0.111	177	1.055	0.254	1148	1.085	0.138	1325	1.081	0.157
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	575	37.063	1.633	605	37.238	1.557	1180	37.152	1.591	177	38.527	1.737	1148	36.943	1.599	1325	37.152	1.701
Mean diameter for LDL particles (nm)	575	23.587	0.103	605	23.628	0.109	1180	23.608	0.107	177	23.487	0.093	1148	23.573	0.100	1325	23.562	0.104
Mean diameter for HDL particles (nm)	575	10.081	0.233	605	10.102	0.221	1180	10.092	0.226	177	9.798	0.244	1148	10.068	0.262	1325	10.032	0.275
Cholesterol																		
Serum total cholesterol (mmol/L)	575	3.576	0.620	605	3.596	0.643	1180	3.586	0.629	177	4.161	0.885	1148	4.234	0.828	1325	4.225	0.835
Total cholesterol in VLDL (mmol/L)	575	0.438	0.188	605	0.472	0.189	1180	0.455	0.189	177	0.826	0.395	1148	0.592	0.265	1325	0.623	0.295
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	575	0.944	0.269	605	0.994	0.271	1180	0.970	0.270	177	1.452	0.472	1148	1.219	0.383	1325	1.250	0.402
Total cholesterol in LDL (mmol/L)	575	1.130	0.330	605	1.149	0.341	1180	1.139	0.334	177	1.492	0.498	1148	1.460	0.442	1325	1.464	0.449
Total cholesterol in HDL (mmol/L)	575	1.503	0.274	605	1.453	0.266	1180	1.477	0.270	177	1.217	0.350	1148	1.556	0.363	1325	1.511	0.379
Total cholesterol in HDL2 (mmol/L)	575	1.035	0.254	605	0.988	0.246	1180	1.011	0.250	177	0.751	0.327	1148	1.072	0.335	1325	1.030	0.351
Total cholesterol in HDL3 (mmol/L)	575	0.468	0.024	605	0.466	0.024	1180	0.467	0.024	177	0.466	0.035	1148	0.483	0.033	1325	0.481	0.034
Esterified cholesterol (mmol/L)	572	2.516	0.447	604	2.517	0.460	1176	2.517	0.452	176	2.941	0.636	1147	2.975	0.593	1323	2.971	0.597
Free cholesterol (mmol/L)	572	1.062	0.179	604	1.079	0.186	1176	1.070	0.182	176	1.211	0.273	1147	1.260	0.239	1323	1.253	0.244
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	575	0.918	0.709	605	1.005	0.681	1180	0.962	0.696	177	1.686	0.809	1148	1.129	0.755	1325	1.190	0.782
Triglycerides in VLDL (mmol/L)*	575	0.582	0.885	605	0.648	0.830	1180	0.615	0.858	177	1.249	0.927	1148	0.694	0.945	1325	0.750	0.972
Triglycerides in LDL (mmol/L)	575	0.113	0.024	605	0.123	0.027	1180	0.118	0.026	177	0.153	0.040	1148	0.158	0.044	1325	0.157	0.043
Triglycerides in HDL (mmol/L)	575	0.129	0.030	605	0.136	0.030	1180	0.133	0.030	177	0.161	0.047	1148	0.151	0.040	1325	0.153	0.041
Total phosphoglycerides (mmol/L)	572	1.632	0.240	604	1.620	0.261	1176	1.626	0.250	176	1.865	0.354	1147	1.926	0.340	1323	1.918	0.342
Ratio of triglycerides to phosphoglycerides	572	0.526	0.253	604	0.569	0.277	1176	0.548	0.265	176	0.946	0.587	1147	0.580	0.279	1323	0.628	0.355
Phosphatidylcholine & other cholines (mmol/L)	572	1.691	0.240	604	1.687	0.267	1176	1.689	0.253	175	1.877	0.320	1147	1.978	0.336	1322	1.965	0.335
Sphingomyelins (mmol/L)	572	0.348	0.061	604	0.349	0.064	1176	0.348	0.062	175	0.370	0.070	1147	0.397	0.078	1322	0.394	0.077
Total cholines (mmol/L)	572	2.005	0.256	604	1.997	0.264	1176	2.001	0.259	175	2.185	0.334	1147	2.317	0.351	1322	2.299	0.351
Apolipoproteins																		
Apolipoprotein A1 (g/L)	575	1.509	0.159	605	1.484	0.151	1180	1.497	0.155	177	1.461	0.178	1148	1.589	0.205	1325	1.572	0.206
Apolipoprotein B (g/L)	575	0.682	0.135	604	0.706	0.136	1179	0.694	0.135	177	0.955	0.245	1148	0.812	0.196	1325	0.831	0.208
Ratio of apolipoprotein B to apolipoprotein A	575	0.455	0.097	604	0.479	0.098	1179	0.467	0.098	177	0.660	0.178	1148	0.518	0.136	1325	0.537	0.150
Fatty acids																		
Total fatty acids (mmol/L)	570	9.215	1.697	604	9.370	1.730	1174	9.294	1.709	173	11.850	2.723	1145	10.917	2.392	1318	11.034	2.446
Estimated degree of unsaturation	570	1.212	0.056	604	1.196	0.065	1174	1.204	0.061	173	1.179	0.070	1145	1.212	0.066	1318	1.208	0.068
22:6, docosahexaenoic acid (mmol/L)	570	0.078	0.028	604	0.074	0.029	1174	0.076	0.028	173	0.118	0.051	1145	0.111	0.039	1318	0.112	0.041
18:2, linoleic acid (mmol/L)	570	2.539	0.456	604	2.592	0.464	1174	2.566	0.459	173	2.919	0.567	1145	2.880	0.584	1318	2.885	0.580
Omega3 fatty acids (mmol/L)	570	0.309	0.086	604	0.296	0.083	1174	0.302	0.085	173	0.451	0.160	1145	0.400	0.117	1318	0.406	0.124
Omega6 fatty acids (mmol/L)	570	3.077	0.493	604	3.094	0.492	1174	3.086	0.491	173	3.553	0.648	1145	3.507	0.627	1318	3.513	0.628
Polyunsat. fatty acids (mmol/L)	570	3.386	0.565	604	3.390	0.561	1174	3.388	0.560	173	4.004	0.775	1145	3.907	0.721	1318	3.919	0.726
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	570	2.500	0.632	604	2.587	0.661	1174	2.544	0.646	173	3.520	1.058	1145	3.080	0.917	1318	3.135	0.943
Saturated fatty acids (mmol/L)	570	3.328	0.642	604	3.393	0.682	1174	3.362	0.661	173	4.325	1.088	1145	3.930	0.932	1318	3.979	0.958

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Fatty acid ratios

Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	570	0.841	0.239	604	0.790	0.269	1174	0.815	0.255	173	0.984	0.315	1145	1.023	0.284	1318	1.018	0.287
Ratio of 18:2 linoleic acid to total fatty acids (%)	570	27.741	3.127	604	27.867	3.274	1174	27.805	3.191	173	25.038	3.570	1145	26.656	3.417	1318	26.453	3.467
Ratio of omega3 fatty acids to total fatty acids (%)	570	3.329	0.579	604	3.150	0.600	1174	3.238	0.594	173	3.763	0.799	1145	3.659	0.697	1318	3.672	0.709
Ratio of omega6 fatty acids to total fatty acids (%)	570	33.650	3.047	604	33.308	3.228	1174	33.475	3.133	173	30.468	3.757	1145	32.502	3.311	1318	32.247	3.424
Ratio of polyunsat. fatty acids to total fatty acids (%)	570	36.979	3.235	604	36.459	3.514	1174	36.713	3.377	173	34.231	3.878	1145	36.161	3.588	1318	35.918	3.669
Ratio of monounsat. fatty acids to total fatty acids (%)	570	26.911	2.569	604	27.375	2.678	1174	27.148	2.625	173	29.366	2.968	1145	27.889	2.842	1318	28.075	2.891
Ratio of saturated fatty acids to total fatty acids (%)	570	36.110	1.675	604	36.167	1.802	1174	36.139	1.734	173	36.402	2.027	1145	35.950	2.032	1318	36.007	2.031

Glycolysis related

Glucose (mmol/L)*	574	1.350	0.115	605	1.336	0.105	1179	1.342	0.11	176	1.415	0.205	1148	1.334	0.176	1324	1.344	0.182
Lactate (mmol/L)	575	1.770	0.459	605	1.718	0.434	1180	1.743	0.446	177	1.696	0.472	1148	1.562	0.480	1325	1.580	0.480
Pyruvate (mmol/L)	574	0.100	0.024	605	0.098	0.023	1179	0.099	0.023	177	0.101	0.031	1147	0.093	0.034	1324	0.094	0.033
Citrate (mmol/L)	575	0.125	0.017	604	0.131	0.018	1179	0.128	0.018	177	0.110	0.016	1148	0.111	0.016	1325	0.111	0.016
Glycerol (mmol/L)#	240	0.078	0.021	283	0.083	0.022	523	0.081	0.021	84	0.073	0.021	470	0.071	0.023	554	0.071	0.023

Amino acids

Alanine (mmol/L)	575	0.387	0.061	605	0.396	0.060	1180	0.391	0.060	176	0.423	0.065	1147	0.399	0.060	1323	0.402	0.061
Glutamine (mmol/L)	575	0.474	0.050	605	0.497	0.051	1180	0.485	0.051	177	0.490	0.063	1148	0.456	0.066	1325	0.461	0.066
Glycine (mmol/L)	574	0.261	0.032	604	0.270	0.034	1178	0.265	0.033	176	0.243	0.029	1148	0.274	0.061	1324	0.270	0.059
Histidine (mmol/L)	574	0.065	0.009	605	0.065	0.008	1179	0.065	0.008	176	0.066	0.008	1148	0.065	0.009	1324	0.065	0.009
Isoleucine (mmol/L)	574	0.054	0.019	605	0.053	0.019	1179	0.054	0.019	174	0.072	0.021	1146	0.055	0.020	1320	0.057	0.021
Leucine (mmol/L)	575	0.073	0.019	605	0.071	0.019	1180	0.072	0.019	177	0.097	0.029	1148	0.074	0.021	1325	0.077	0.023
Valine (mmol/L)	575	0.162	0.037	604	0.156	0.035	1179	0.159	0.036	177	0.192	0.036	1147	0.162	0.042	1324	0.166	0.042
Phenylalanine (mmol/L)	575	0.068	0.009	605	0.066	0.009	1180	0.067	0.009	177	0.073	0.011	1148	0.072	0.011	1325	0.073	0.011
Tyrosine (mmol/L)	574	0.054	0.014	605	0.055	0.014	1179	0.055	0.014	176	0.060	0.013	1148	0.054	0.015	1324	0.055	0.015

Ketone bodies

Acetate (mmol/L)*	575	0.031	0.423	605	0.030	0.404	1180	0.030	0.413	177	0.037	0.655	1146	0.033	0.600	1323	0.034	0.609
Acetoacetate (mmol/L)*	575	0.025	1.310	605	0.023	1.429	1180	0.024	1.367	177	0.023	2.116	1147	0.024	1.278	1324	0.024	1.403
3hydroxybutyrate (mmol/L)*#	555	0.100	0.786	580	0.103	0.826	1135	0.101	0.805	170	0.104	0.669	1098	0.096	0.781	1268	0.097	0.769

Fluid balance

Albumin (signal area)	574	0.093	0.005	605	0.092	0.005	1179	0.093	0.005	177	0.090	0.005	1148	0.088	0.005	1325	0.089	0.005
Creatinine (mmol/L)	570	0.040	0.006	600	0.040	0.006	1170	0.040	0.006	173	0.066	0.015	1139	0.054	0.009	1312	0.055	0.010

Inflammation

Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	575	1.170	0.191	605	1.173	0.186	1180	1.172	0.188	177	1.375	0.366	1148	1.242	0.233	1325	1.260	0.258
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* geometric mean [relative SD] when skewed variable

Note: The presence of ethanol in a sample can affect quantification of glycerol and on some occasions 3hydroxybutyrate.

Ethanol can be introduced in to a sample from disinfectants used during blood collection/processing of sample.

Supplementary table 2: Mean difference in metabolite levels in adults compared to children in absolute concentration unit

Metabolic subgroup	Differences by age (Adults - Child)				Conversion factor (SD) #
	Estimate	95% CI	P-value	Adj_p-value^	
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.704	(0.519, 0.890)	<0.001	<0.001	2.534
Total lipids in very large VLDL (mmol/L)*	0.922	(0.700, 1.145)	<0.001	<0.001	3.031
Total lipids in large VLDL (mmol/L)*	0.648	(0.502, 0.795)	<0.001	<0.001	1.950
Total lipids in medium VLDL (mmol/L)	0.105	(0.080, 0.129)	<0.001	<0.001	0.348
Total lipids in small VLDL (mmol/L)	0.107	(0.094, 0.121)	<0.001	<0.001	0.199
Total lipids in very small VLDL (mmol/L)	0.093	(0.086, 0.099)	<0.001	<0.001	0.104
Total lipids in IDL (mmol/L)	0.181	(0.166, 0.196)	<0.001	<0.001	0.230
Total lipids in large LDL (mmol/L)	0.229	(0.211, 0.247)	<0.001	<0.001	0.286
Total lipids in medium LDL (mmol/L)	0.144	(0.132, 0.155)	<0.001	<0.001	0.178
Total lipids in small LDL (mmol/L)	0.089	(0.082, 0.096)	<0.001	<0.001	0.110
Total lipids in very large HDL (mmol/L)	0.012	(-0.003, 0.027)	0.128	0.132	0.217
Total lipids in large HDL (mmol/L)	0.032	(0.007, 0.057)	0.011	0.012	0.353
Total lipids in medium HDL (mmol/L)	0.076	(0.064, 0.089)	<0.001	<0.001	0.166
Total lipids in small HDL (mmol/L)	0.068	(0.058, 0.078)	<0.001	<0.001	0.137
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	-0.147	(-0.263, -0.031)	0.013	0.014	1.633
Mean diameter for LDL particles (nm)	-0.044	(-0.052, -0.037)	<0.001	<0.001	0.106
Mean diameter for HDL particles (nm)	-0.027	(-0.045, -0.010)	0.002	0.003	0.256
Cholesterol					
Serum total cholesterol (mmol/L)	0.670	(0.619, 0.721)	<0.001	<0.001	0.805
Total cholesterol in VLDL (mmol/L)	0.146	(0.129, 0.163)	<0.001	<0.001	0.249
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.261	(0.237, 0.284)	<0.001	<0.001	0.357
Total cholesterol in LDL (mmol/L)	0.327	(0.300, 0.354)	<0.001	<0.001	0.424
Total cholesterol in HDL (mmol/L)	0.082	(0.058, 0.106)	<0.001	<0.001	0.337
Total cholesterol in HDL2 (mmol/L)	0.064	(0.042, 0.086)	<0.001	<0.001	0.311
Total cholesterol in HDL3 (mmol/L)	0.018	(0.016, 0.020)	<0.001	<0.001	0.031
Esterified cholesterol (mmol/L)	0.474	(0.437, 0.511)	<0.001	<0.001	0.576
Free cholesterol (mmol/L)	0.195	(0.180, 0.210)	<0.001	<0.001	0.234
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	0.176	(0.145, 0.206)	<0.001	<0.001	0.443
Triglycerides in VLDL (mmol/L)*	0.140	(0.096, 0.183)	<0.001	<0.001	0.615
Triglycerides in LDL (mmol/L)	0.040	(0.038, 0.043)	<0.001	<0.001	0.042
Triglycerides in HDL (mmol/L)	0.020	(0.017, 0.022)	<0.001	<0.001	0.037
Total phosphoglycerides (mmol/L)	0.311	(0.290, 0.332)	<0.001	<0.001	0.337
Ratio of triglycerides to phosphoglycerides	0.049	(0.027, 0.071)	<0.001	<0.001	0.299
Phosphatidylcholine & other cholines (mmol/L)	0.295	(0.274, 0.316)	<0.001	<0.001	0.329
Sphingomyelins (mmol/L)	0.052	(0.047, 0.057)	<0.001	<0.001	0.075
Total cholines (mmol/L)	0.323	(0.302, 0.345)	<0.001	<0.001	0.347
Apolipoproteins					
Apolipoprotein A1 (g/L)	0.099	(0.086, 0.112)	<0.001	<0.001	0.191
Apolipoprotein B (g/L)	0.125	(0.113, 0.137)	<0.001	<0.001	0.182
Ratio of apolipoprotein B to apolipoprotein A1	0.055	(0.046, 0.064)	<0.001	<0.001	0.127
Fatty acids					
Total fatty acids (mmol/L)	1.738	(1.592, 1.885)	<0.001	<0.001	2.245
Estimated degree of unsaturation	0.005	(0.000, 0.009)	0.030	0.031	0.063
22:6, docosahexaenoic acid (mmol/L)	0.037	(0.035, 0.040)	<0.001	<0.001	0.040
18:2, linoleic acid (mmol/L)	0.347	(0.310, 0.384)	<0.001	<0.001	0.545
Omega3 fatty acids (mmol/L)	0.105	(0.098, 0.113)	<0.001	<0.001	0.118
Omega6 fatty acids (mmol/L)	0.453	(0.414, 0.492)	<0.001	<0.001	0.597
Polyunsat. fatty acids (mmol/L)	0.558	(0.513, 0.603)	<0.001	<0.001	0.695
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.568	(0.512, 0.625)	<0.001	<0.001	0.850
Saturated fatty acids (mmol/L)	0.612	(0.554, 0.669)	<0.001	<0.001	0.862

Fatty acid ratios

Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.209	(0.190, 0.228)	<0.001	<0.001	0.292
Ratio of 18:2 linoleic acid to total fatty acids (%)	-1.079	(-1.331, -0.827)	<0.001	<0.001	3.387
Ratio of omega3 fatty acids to total fatty acids (%)	0.446	(0.403, 0.490)	<0.001	<0.001	0.693
Ratio of omega6 fatty acids to total fatty acids (%)	-0.992	(-1.227, -0.757)	<0.001	<0.001	3.296
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.546	(-0.794, -0.298)	<0.001	<0.001	3.507
Ratio of monounsat. fatty acids to total fatty acids (%)	0.741	(0.547, 0.934)	<0.001	<0.001	2.797
Ratio of saturated fatty acids to total fatty acids (%)	-0.195	(-0.328, -0.062)	0.004	0.005	1.863

Glycolysis related

Glucose (mmol/L)*	-0.002	(-0.014, 0.009)	0.700	0.709	0.147
Lactate (mmol/L)	-0.180	(-0.215, -0.144)	<0.001	<0.001	0.471
Pyruvate (mmol/L)	-0.007	(-0.009, -0.005)	<0.001	<0.001	0.029
Citrate (mmol/L)	-0.017	(-0.018, -0.016)	<0.001	<0.001	0.019
Glycerol (mmol/L)	-0.011	(-0.015, -0.008)	<0.001	<0.001	0.023

Amino acids

Alanine (mmol/L)	0.013	(0.009, 0.017)	<0.001	<0.001	0.060
Glutamine (mmol/L)	-0.023	(-0.027, -0.019)	<0.001	<0.001	0.060
Glycine (mmol/L)	0.007	(0.003, 0.010)	<0.001	<0.001	0.049
Histidine (mmol/L)	0.001	(0.000, 0.002)	0.005	0.006	0.009
Isoleucine (mmol/L)	0.003	(0.002, 0.004)	<0.001	<0.001	0.019
Leucine (mmol/L)	0.004	(0.003, 0.006)	<0.001	<0.001	0.021
Valine (mmol/L)	0.009	(0.006, 0.011)	<0.001	<0.001	0.039
Phenylalanine (mmol/L)	0.005	(0.005, 0.006)	<0.001	<0.001	0.010
Tyrosine (mmol/L)	0.001	(-0.000, 0.002)	0.100	0.105	0.014

Ketone bodies

Acetate (mmol/L)*	0.101	(0.084, 0.117)	<0.001	<0.001	0.235
Acetoacetate (mmol/L)*	-0.004	(-0.086, 0.078)	0.922	0.922	1.022
3hydroxybutyrate (mmol/L)*	-0.064	(-0.100, -0.028)	0.001	0.001	0.493

Fluid balance

Albumin (signal area)	-0.004	(-0.004, -0.004)	<0.001	<0.001	0.005
Creatinine (mmol/L)	0.016	(0.015, 0.017)	<0.001	0.001	0.012

Inflammation

Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.062	(0.047, 0.078)	<0.001	<0.001	0.217
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* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations in Figure 2 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor. Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 3: Differences in mean metabolite levels in girls compared to boys in absolute concentration units.

Metabolic subgroup	Differences for children (Female - Male)				
	Estimate	95% CI	pvalue	Adj_p-value [^]	Conversion factor (SD) #
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	0.737	(0.414, 1.059)	<0.001	<0.001	2.845
Total lipids in very large VLDL (mmol/L)*	0.744	(0.355, 1.134)	<0.001	0.001	3.428
Total lipids in large VLDL (mmol/L)*	0.663	(0.390, 0.936)	<0.001	<0.001	2.411
Total lipids in medium VLDL (mmol/L)	0.049	(0.018, 0.080)	0.002	0.004	0.269
Total lipids in small VLDL (mmol/L)	0.032	(0.015, 0.048)	<0.001	0.001	0.146
Total lipids in very small VLDL (mmol/L)	0.018	(0.010, 0.027)	<0.001	<0.001	0.074
Total lipids in IDL (mmol/L)	0.025	(0.004, 0.046)	0.017	0.035	0.182
Total lipids in large LDL (mmol/L)	0.020	(-0.006, 0.046)	0.132	0.187	0.227
Total lipids in medium LDL (mmol/L)	0.008	(-0.008, 0.024)	0.338	0.416	0.139
Total lipids in small LDL (mmol/L)	0.001	(-0.008, 0.011)	0.788	0.822	0.086
Total lipids in very large HDL (mmol/L)	-0.002	(-0.023, 0.020)	0.882	0.882	0.190
Total lipids in large HDL (mmol/L)	-0.033	(-0.066, -0.001)	0.044	0.074	0.283
Total lipids in medium HDL (mmol/L)	-0.045	(-0.059, -0.030)	<0.001	<0.001	0.131
Total lipids in small HDL (mmol/L)	-0.035	(-0.048, -0.022)	<0.001	<0.001	0.116
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	0.215	(0.035, 0.395)	0.019	0.038	1.580
Mean diameter for LDL particles (nm)	0.035	(0.023, 0.047)	<0.001	0.000	0.108
Mean diameter for HDL particles (nm)	0.003	(-0.023, 0.028)	0.847	0.870	0.226
Cholesterol					
Serum total cholesterol (mmol/L)	0.007	(-0.066, 0.079)	0.857	0.869	0.634
Total cholesterol in VLDL (mmol/L)	0.040	(0.019, 0.061)	<0.001	0.001	0.184
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	0.053	(0.023, 0.083)	0.001	0.002	0.265
Total cholesterol in LDL (mmol/L)	0.016	(-0.023, 0.054)	0.427	0.518	0.336
Total cholesterol in HDL (mmol/L)	-0.062	(-0.093, -0.031)	<0.001	<0.001	0.273
Total cholesterol in HDL2 (mmol/L)	-0.059	(-0.087, -0.030)	<0.001	<0.001	0.253
Total cholesterol in HDL3 (mmol/L)	-0.003	(-0.006, -0.001)	0.013	0.027	0.024
Esterified cholesterol (mmol/L)	-0.008	(-0.060, 0.044)	0.755	0.798	0.455
Free cholesterol (mmol/L)	0.013	(-0.008, 0.034)	0.211	0.284	0.184
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	0.101	(0.056, 0.145)	<0.001	<0.001	0.390
Triglycerides in VLDL (mmol/L)*	0.125	(0.062, 0.187)	<0.001	<0.001	0.551
Triglycerides in LDL (mmol/L)	0.009	(0.006, 0.012)	<0.001	<0.001	0.026
Triglycerides in HDL (mmol/L)	0.007	(0.004, 0.011)	<0.001	<0.001	0.029
Total phosphoglycerides (mmol/L)	-0.018	(-0.047, 0.010)	0.206	0.282	0.249
Ratio of triglycerides to phosphoglycerides	0.051	(0.020, 0.083)	0.001	0.003	0.274
Phosphatidylcholine & other cholines (mmol/L)	-0.011	(-0.040, 0.018)	0.447	0.534	0.251
Sphingomyelins (mmol/L)	-0.001	(-0.009, 0.006)	0.706	0.757	0.063
Total cholines (mmol/L)	-0.017	(-0.046, 0.013)	0.268	0.354	0.257
Apolipoproteins					
Apolipoprotein A1 (g/L)	-0.030	(-0.048, -0.013)	0.001	0.002	0.155
Apolipoprotein B (g/L)	0.027	(0.012, 0.042)	0.001	0.002	0.133
Ratio of apolipoprotein B to apolipoprotein A1	0.027	(0.016, 0.038)	<0.001	<0.001	0.098
Fatty acids					
Total fatty acids (mmol/L)	0.200	(0.011, 0.389)	0.038	0.065	1.650
Estimated degree of unsaturation	-0.016	(-0.022, -0.009)	<0.001	<0.001	0.060
22:6, docosahexaenoic acid (mmol/L)	-0.004	(-0.007, -0.000)	0.033	0.059	0.028
18:2, linoleic acid (mmol/L)	0.068	(0.015, 0.120)	0.011	0.024	0.458
Omega3 fatty acids (mmol/L)	-0.008	(-0.018, 0.001)	0.083	0.128	0.084
Omega6 fatty acids (mmol/L)	0.031	(-0.024, 0.087)	0.271	0.352	0.485
Polyunsat. fatty acids (mmol/L)	0.023	(-0.041, 0.086)	0.483	0.567	0.553
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	0.105	(0.034, 0.176)	0.004	0.009	0.623
Saturated fatty acids (mmol/L)	0.073	(-0.000, 0.146)	0.052	0.085	0.638
Fatty acid ratios					
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	-0.049	(-0.078, -0.019)	0.001	0.003	0.255
Ratio of 18:2 linoleic acid to total fatty acids (%)	0.197	(-0.173, 0.567)	0.297	0.379	3.233
Ratio of omega3 fatty acids to total fatty acids (%)	-0.145	(-0.213, -0.078)	<0.001	0.000	0.593
Ratio of omega6 fatty acids to total fatty acids (%)	-0.302	(-0.658, 0.054)	0.096	0.142	3.109
Ratio of polyunsat. fatty acids to total fatty acids (%)	-0.447	(-0.830, -0.065)	0.022	0.042	3.346
Ratio of monounsat. fatty acids to total fatty acids (%)	0.499	(0.202, 0.796)	0.001	0.003	2.606
Ratio of saturated fatty acids to total fatty acids (%)	-0.051	(-0.251, 0.148)	0.614	0.689	1.743
Glycolysis related					
Glucose (mmol/L)*	-0.013	(-0.026, 0.001)	0.061	0.098	0.118
Lactate (mmol/L)	-0.045	(-0.097, 0.007)	0.088	0.133	0.456
Pyruvate (mmol/L)	-0.001	(-0.004, 0.002)	0.524	0.606	0.024
Citrate (mmol/L)	0.007	(0.005, 0.009)	<0.001	<0.001	0.018
Glycerol (mmol/L)	0.006	(0.002, 0.010)	0.004	0.009	0.023
Amino acids					
Alanine (mmol/L)	0.011	(0.004, 0.017)	0.002	0.004	0.058
Glutamine (mmol/L)	0.023	(0.018, 0.029)	<0.001	<0.001	0.051
Glycine (mmol/L)	0.010	(0.006, 0.014)	<0.001	<0.001	0.032
Histidine (mmol/L)	0.001	(-0.000, 0.002)	0.075	0.118	0.008
Isoleucine (mmol/L)	0.000	(-0.003, 0.002)	0.637	0.693	0.018

1	Leucine (mmol/L)	-0.002	(-0.005, -0.000)	0.022	0.041	0.018
2	Valine (mmol/L)	-0.007	(-0.011, -0.003)	0.001	0.003	0.036
3	Phenylalanine (mmol/L)	-0.002	(-0.003, -0.001)	0.003	0.007	0.009
4	Tyrosine (mmol/L)	0.000	(-0.001, 0.002)	0.582	0.663	0.014
5	Ketone bodies					
6	Acetate (mmol/L)*	-0.030	(-0.048, -0.011)	0.002	0.005	0.166
7	Acetoacetate (mmol/L)*	-0.058	(-0.172, 0.055)	0.313	0.393	0.992
8	3hydroxybutyrate (mmol/L)*	0.041	(-0.019, 0.101)	0.178	0.248	0.513
9	Fluid balance					
10	Albumin (signal area)	-0.001	(-0.001, -0.000)	0.037	0.064	0.005
11	Creatinine (mmol/L)	0.000	(-0.001, 0.001)	0.624	0.690	0.007
12	Inflammation					
13	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	0.017	(-0.004, 0.038)	0.104	0.151	0.183

* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations for children in Figure 3 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure in children) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 4: Differences in mean metabolite levels in female compared to male adults in absolute concentration units.

Metabolic subgroup	Differences for adults (Female - Male)				
	Estimate	95% CI	pvalue	Adj_p-value^	Conversion factor (SD) #
Lipoprotein subclass lipids					
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	-0.930	(-1.271, -0.589)	<0.001	<0.001	2.173
Total lipids in very large VLDL (mmol/L)*	-1.217	(-1.617, -0.818)	<0.001	<0.001	2.555
Total lipids in large VLDL (mmol/L)*	-0.900	(-1.107, -0.693)	<0.001	<0.001	1.343
Total lipids in medium VLDL (mmol/L)	-0.325	(-0.385, -0.264)	<0.001	<0.001	0.398
Total lipids in small VLDL (mmol/L)	-0.167	(-0.201, -0.133)	<0.001	<0.001	0.223
Total lipids in very small VLDL (mmol/L)	-0.022	(-0.039, -0.005)	0.013	0.018	0.107
Total lipids in IDL (mmol/L)	0.014	(-0.023, 0.051)	0.465	0.530	0.236
Total lipids in large LDL (mmol/L)	-0.011	(-0.058, 0.035)	0.634	0.671	0.293
Total lipids in medium LDL (mmol/L)	-0.028	(-0.057, 0.001)	0.058	0.076	0.182
Total lipids in small LDL (mmol/L)	-0.017	(-0.035, 0.001)	0.061	0.079	0.113
Total lipids in very large HDL (mmol/L)	0.195	(0.158, 0.231)	<0.001	<0.001	0.239
Total lipids in large HDL (mmol/L)	0.395	(0.334, 0.455)	<0.001	<0.001	0.405
Total lipids in medium HDL (mmol/L)	0.129	(0.101, 0.158)	<0.001	<0.001	0.184
Total lipids in small HDL (mmol/L)	0.002	(-0.021, 0.025)	0.850	0.874	0.145
Lipoprotein particle size					
Mean diameter for VLDL particles (nm)	-1.414	(-1.669, -1.159)	<0.001	<0.001	1.678
Mean diameter for LDL particles (nm)	0.081	(0.066, 0.096)	<0.001	<0.001	0.100
Mean diameter for HDL particles (nm)	0.278	(0.236, 0.320)	<0.001	<0.001	0.279
Cholesterol					
Serum total cholesterol (mmol/L)	0.112	(-0.018, 0.241)	0.091	0.116	0.817
Total cholesterol in VLDL (mmol/L)	-0.187	(-0.230, -0.145)	<0.001	<0.001	0.275
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	-0.184	(-0.244, -0.124)	<0.001	<0.001	0.383
Total cholesterol in LDL (mmol/L)	-0.048	(-0.117, 0.021)	0.175	0.213	0.437
Total cholesterol in HDL (mmol/L)	0.344	(0.286, 0.401)	<0.001	<0.001	0.382
Total cholesterol in HDL2 (mmol/L)	0.324	(0.271, 0.378)	<0.001	<0.001	0.354
Total cholesterol in HDL3 (mmol/L)	0.019	(0.014, 0.025)	<0.001	<0.001	0.034
Esterified cholesterol (mmol/L)	0.070	(-0.023, 0.163)	0.139	0.172	0.586
Free cholesterol (mmol/L)	0.046	(0.009, 0.084)	0.016	0.023	0.238
Glycerides and phospholipids					
Serum total triglycerides (mmol/L)*	-0.344	(-0.416, -0.273)	<0.001	<0.001	0.468
Triglycerides in VLDL (mmol/L)*	-0.530	(-0.630, -0.429)	<0.001	<0.001	0.659
Triglycerides in LDL (mmol/L)	0.008	(0.001, 0.014)	0.033	0.044	0.044
Triglycerides in HDL (mmol/L)	-0.004	(-0.010, 0.002)	0.228	0.272	0.040
Total phosphoglycerides (mmol/L)	0.106	(0.052, 0.159)	<0.001	<0.001	0.340
Ratio of triglycerides to phosphoglycerides	-0.289	(-0.337, -0.241)	<0.001	<0.001	0.318
Phosphatidylcholine & other cholines (mmol/L)	0.138	(0.086, 0.190)	<0.001	<0.001	0.331
Sphingomyelins (mmol/L)	0.032	(0.020, 0.045)	<0.001	<0.001	0.078
Total cholines (mmol/L)	0.170	(0.115, 0.224)	<0.001	<0.001	0.349
Apolipoproteins					
Apolipoprotein A1 (g/L)	0.146	(0.114, 0.178)	<0.001	<0.001	0.209
Apolipoprotein B (g/L)	-0.115	(-0.146, -0.084)	<0.001	<0.001	0.198
Ratio of apolipoprotein B to apolipoprotein A1	-0.126	(-0.148, -0.105)	<0.001	<0.001	0.144
Fatty acids					
Total fatty acids (mmol/L)	-0.711	(-1.091, -0.330)	<0.001	<0.001	2.388
Estimated degree of unsaturation	0.031	(0.021, 0.042)	<0.001	<0.001	0.066
22:6, docosahexaenoic acid (mmol/L)	-0.002	(-0.008, 0.005)	0.622	0.667	0.041
18:2, linoleic acid (mmol/L)	0.004	(-0.087, 0.094)	0.934	0.947	0.566
Omega3 fatty acids (mmol/L)	-0.035	(-0.054, -0.016)	<0.001	0.001	0.122
Omega6 fatty acids (mmol/L)	0.004	(-0.094, 0.102)	0.936	0.936	0.612
Polyunsat. fatty acids (mmol/L)	-0.031	(-0.144, 0.082)	0.592	0.644	0.708
Monounsatur. fatty acids; 16:1, 18:1 (mmol/L)	-0.372	(-0.520, -0.225)	<0.001	<0.001	0.930
Saturated fatty acids (mmol/L)	-0.307	(-0.455, -0.159)	<0.001	<0.001	0.932
Fatty acid ratios					
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	0.064	(0.018, 0.110)	0.006	0.009	0.288
Ratio of 18:2 linoleic acid to total fatty acids (%)	1.527	(0.984, 2.070)	<0.001	<0.001	3.430
Ratio of omega3 fatty acids to total fatty acids (%)	-0.038	(-0.152, 0.075)	0.508	0.570	0.710
Ratio of omega6 fatty acids to total fatty acids (%)	1.882	(1.351, 2.412)	<0.001	<0.001	3.376
Ratio of polyunsatur. fatty acids to total fatty acids (%)	1.843	(1.272, 2.414)	<0.001	<0.001	3.621
Ratio of monounsatur. fatty acids to total fatty acids (%)	-1.456	(-1.914, -0.998)	<0.001	<0.001	2.904
Ratio of saturated fatty acids to total fatty acids (%)	-0.387	(-0.700, -0.074)	0.015	0.022	1.959
Glycolysis related					
Glucose (mmol/L)*	-0.071	(-0.097, -0.044)	<0.001	<0.001	0.168
Lactate (mmol/L)	-0.178	(-0.252, -0.105)	<0.001	<0.001	0.469
Pyruvate (mmol/L)	-0.007	(-0.013, -0.002)	0.004	0.006	0.032

1	Citrate (mmol/L)	-0.001	(-0.004, 0.001)	0.335	0.387	0.016
2	Glycerol (mmol/L)	-0.003	(-0.008, 0.002)	0.279	0.328	0.022
3						
4	Amino acids					
5	Alanine (mmol/L)	-0.020	(-0.030, -0.011)	<0.001	<0.001	0.060
6	Glutamine (mmol/L)	-0.038	(-0.048, -0.028)	<0.001	<0.001	0.065
7	Glycine (mmol/L)	0.029	(0.020, 0.038)	<0.001	<0.001	0.059
8	Histidine (mmol/L)	-0.001	(-0.003, 0.000)	0.116	0.146	0.009
9	Isoleucine (mmol/L)	-0.016	(-0.019, -0.013)	<0.001	<0.001	0.021
10	Leucine (mmol/L)	-0.019	(-0.022, -0.016)	<0.001	<0.001	0.022
11	Valine (mmol/L)	-0.029	(-0.035, -0.022)	<0.001	<0.001	0.042
12	Phenylalanine (mmol/L)	0.000	(-0.002, 0.001)	0.576	0.637	0.010
13	Tyrosine (mmol/L)	-0.005	(-0.007, -0.003)	<0.001	<0.001	0.014
14						
15	Ketone bodies					
16	Acetate (mmol/L)*	-0.076	(-0.119, -0.033)	0.001	0.001	0.273
17	Acetoacetate (mmol/L)*	0.018	(-0.148, 0.184)	0.828	0.863	1.048
18	3hydroxybutyrate (mmol/L)*	-0.087	(-0.163, -0.011)	0.025	0.035	0.472
19						
20	Fluid balance					
21	Albumin (signal area)	-0.002	(-0.002, -0.001)	<0.001	<0.001	0.005
22	Creatinine (mmol/L)	-0.013	(-0.015, -0.012)	<0.001	<0.001	0.010
23						
24	Inflammation					
25	Glycoprotein acetyls, mainly a1acid glycoprotein (mmol/L)	-0.098	(-0.136, -0.061)	<0.001	<0.001	0.239

* Metabolite has been log transformed

^ Benjamini-Hochberg adjusted p-value

Associations for parents in Figure 3 are presented in SD-units. The conversion factor provided (unweighted standard deviation of each metabolite measure in adults/parents) can be used to convert the association in absolute concentration to SD units by dividing by the conversion factor.

Where metabolite has been log transformed conversion factor is standard deviation of log transformed metabolite

Supplementary table 5: Mother-child concordance; correlations and partial correlations between mothers and their sons, daughters and all children.

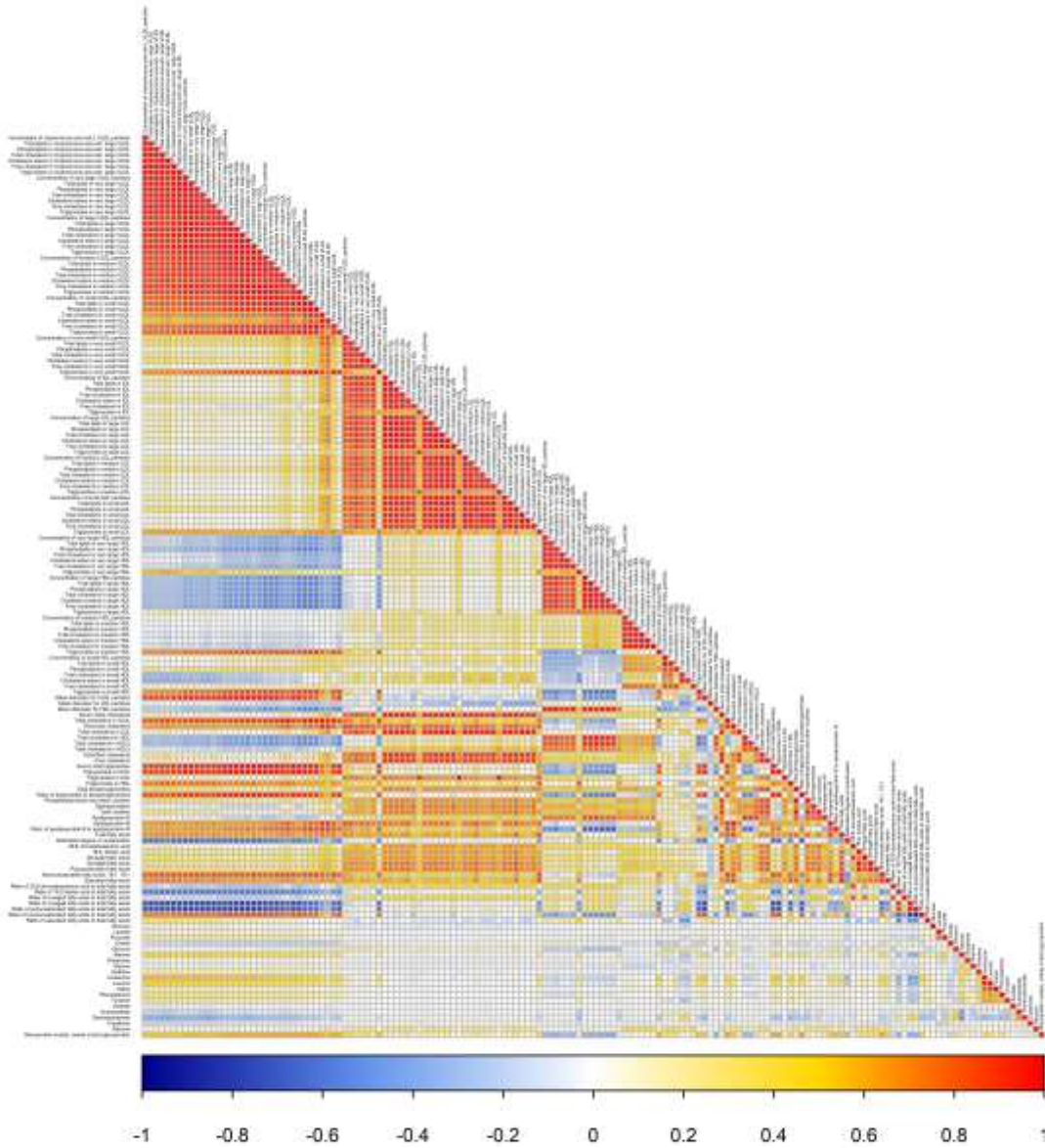
Metabolic subgroup	Mother																		
	Boys						Girls						All Children						
	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	PCC*	95% CI	
Lipoprotein subclass lipids																			
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	469	0.22	0.13-0.30	433	0.23	0.14-0.31	518	0.24	0.16-0.32	476	0.21	0.12-0.29	987	0.23	0.17-0.29	909	0.22	0.16-0.28	
Total lipids in very large VLDL (mmol/L)*	469	0.25	0.16-0.33	433	0.25	0.16-0.34	518	0.22	0.14-0.30	476	0.20	0.12-0.29	987	0.24	0.18-0.29	909	0.23	0.17-0.29	
Total lipids in large VLDL (mmol/L)*	469	0.22	0.13-0.30	433	0.24	0.15-0.33	518	0.23	0.14-0.31	476	0.22	0.14-0.31	987	0.22	0.16-0.28	909	0.23	0.17-0.29	
Total lipids in medium VLDL (mmol/L)	469	0.26	0.17-0.34	433	0.28	0.19-0.36	518	0.29	0.21-0.37	476	0.30	0.21-0.38	987	0.28	0.22-0.34	909	0.29	0.23-0.35	
Total lipids in small VLDL (mmol/L)	469	0.26	0.18-0.35	433	0.28	0.19-0.36	518	0.30	0.21-0.37	476	0.30	0.21-0.38	987	0.29	0.23-0.34	909	0.29	0.23-0.35	
Total lipids in very small VLDL (mmol/L)	469	0.22	0.14-0.31	433	0.21	0.12-0.30	518	0.26	0.17-0.34	476	0.26	0.18-0.35	987	0.25	0.19-0.30	909	0.25	0.18-0.31	
Total lipids in IDL (mmol/L)	469	0.27	0.18-0.35	433	0.23	0.14-0.32	518	0.29	0.21-0.37	476	0.31	0.23-0.39	987	0.29	0.23-0.34	909	0.28	0.22-0.34	
Total lipids in large LDL (mmol/L)	469	0.28	0.19-0.36	433	0.24	0.15-0.33	518	0.30	0.22-0.37	476	0.32	0.24-0.40	987	0.29	0.23-0.35	909	0.29	0.23-0.35	
Total lipids in medium LDL (mmol/L)	469	0.28	0.20-0.36	433	0.24	0.15-0.33	518	0.29	0.21-0.37	476	0.32	0.24-0.40	987	0.29	0.23-0.35	909	0.29	0.23-0.35	
Total lipids in small LDL (mmol/L)	469	0.28	0.19-0.36	433	0.24	0.15-0.32	518	0.29	0.21-0.37	476	0.32	0.23-0.40	987	0.28	0.23-0.34	909	0.28	0.22-0.34	
Total lipids in very large HDL (mmol/L)	469	0.30	0.22-0.38	433	0.30	0.22-0.39	518	0.32	0.24-0.39	476	0.30	0.21-0.38	987	0.31	0.25-0.36	909	0.30	0.24-0.36	
Total lipids in large HDL (mmol/L)	469	0.31	0.23-0.39	433	0.31	0.23-0.40	518	0.28	0.20-0.36	476	0.26	0.18-0.34	987	0.30	0.24-0.35	909	0.29	0.23-0.35	
Total lipids in medium HDL (mmol/L)	469	0.22	0.13-0.30	433	0.20	0.11-0.29	518	0.12	0.03-0.20	476	0.13	0.04-0.22	987	0.17	0.11-0.23	909	0.17	0.10-0.23	
Total lipids in small HDL (mmol/L)	469	0.23	0.14-0.31	433	0.22	0.13-0.31	518	0.20	0.12-0.29	476	0.20	0.11-0.28	987	0.21	0.15-0.27	909	0.21	0.15-0.27	
Lipoprotein particle size																			
Mean diameter for VLDL particles (nm)	469	0.30	0.22-0.38	433	0.32	0.23-0.40	518	0.27	0.19-0.35	476	0.25	0.16-0.33	987	0.29	0.23-0.35	909	0.28	0.22-0.34	
Mean diameter for LDL particles (nm)	469	0.22	0.13-0.31	433	0.20	0.11-0.29	518	0.30	0.22-0.38	476	0.32	0.24-0.40	987	0.26	0.20-0.31	909	0.26	0.20-0.32	
Mean diameter for HDL particles (nm)	469	0.32	0.23-0.40	433	0.32	0.23-0.40	518	0.33	0.26-0.41	476	0.31	0.23-0.39	987	0.33	0.27-0.38	909	0.31	0.25-0.37	
Cholesterol																			
Serum total cholesterol (mmol/L)	469	0.27	0.19-0.35	433	0.23	0.14-0.32	518	0.32	0.24-0.39	476	0.34	0.26-0.42	987	0.30	0.24-0.35	909	0.30	0.24-0.35	
Total cholesterol in VLDL (mmol/L)	469	0.25	0.17-0.34	433	0.27	0.18-0.36	518	0.29	0.21-0.36	476	0.29	0.21-0.37	987	0.28	0.22-0.33	909	0.29	0.23-0.35	
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	469	0.25	0.17-0.34	433	0.26	0.17-0.34	518	0.29	0.21-0.37	476	0.31	0.22-0.39	987	0.28	0.22-0.34	909	0.29	0.23-0.35	
Total cholesterol in LDL (mmol/L)	469	0.28	0.20-0.36	433	0.24	0.15-0.33	518	0.29	0.21-0.37	476	0.32	0.23-0.40	987	0.29	0.23-0.34	909	0.29	0.23-0.35	
Total cholesterol in HDL (mmol/L)	469	0.30	0.22-0.38	433	0.30	0.22-0.39	518	0.25	0.16-0.33	476	0.23	0.15-0.32	987	0.28	0.22-0.33	909	0.27	0.20-0.33	
Total cholesterol in very large HDL (mmol/L)	469	0.30	0.22-0.38	433	0.31	0.22-0.39	518	0.25	0.16-0.32	476	0.23	0.15-0.32	987	0.28	0.22-0.33	909	0.27	0.21-0.33	
Total cholesterol in HDL3 (mmol/L)	469	0.25	0.16-0.33	433	0.23	0.14-0.32	518	0.25	0.16-0.33	476	0.24	0.16-0.33	987	0.25	0.19-0.31	909	0.24	0.17-0.30	
Esterified cholesterol (mmol/L)	465	0.28	0.19-0.36	430	0.23	0.14-0.32	518	0.32	0.24-0.39	476	0.34	0.26-0.42	983	0.30	0.24-0.35	906	0.29	0.23-0.35	
Free cholesterol (mmol/L)	465	0.26	0.18-0.34	430	0.22	0.13-0.31	518	0.32	0.24-0.40	476	0.35	0.27-0.43	983	0.30	0.24-0.35	906	0.30	0.24-0.35	
Glycerides and phospholipids																			
Serum total triglycerides (mmol/L)*	469	0.28	0.20-0.36	433	0.30	0.21-0.38	518	0.29	0.21-0.37	476	0.29	0.20-0.37	987	0.29	0.23-0.35	909	0.30	0.24-0.36	
Total triglycerides in VLDL (mmol/L)*	469	0.30	0.22-0.38	433	0.31	0.23-0.40	518	0.28	0.20-0.36	476	0.27	0.18-0.35	987	0.29	0.23-0.35	909	0.29	0.23-0.35	
Triglycerides in LDL (mmol/L)	469	0.19	0.10-0.28	433	0.18	0.09-0.27	518	0.26	0.18-0.34	476	0.27	0.18-0.35	987	0.23	0.17-0.29	909	0.23	0.17-0.29	
Triglycerides in HDL (mmol/L)	469	0.21	0.12-0.30	433	0.23	0.14-0.32	518	0.26	0.18-0.34	476	0.26	0.18-0.34	987	0.24	0.18-0.30	909	0.25	0.19-0.31	
Total phosphoglycerides (mmol/L)	465	0.26	0.17-0.34	430	0.23	0.14-0.32	518	0.26	0.17-0.34	476	0.27	0.18-0.35	983	0.26	0.20-0.32	906	0.25	0.18-0.31	
Ratio of triglycerides to phosphoglycerides	465	0.23	0.15-0.32	430	0.26	0.16-0.34	518	0.26	0.18-0.34	476	0.27	0.19-0.35	983	0.25	0.20-0.31	906	0.27	0.20-0.33	
Phosphatidylcholine & other cholinols (mmol/L)	465	0.27	0.18-0.35	430	0.24	0.14-0.32	518	0.25	0.17-0.33	476	0.27	0.18-0.35	983	0.26	0.20-0.32	906	0.25	0.19-0.31	
Sphingomyelins (mmol/L)	465	0.23	0.15-0.32	430	0.22	0.12-0.30	518	0.29	0.21-0.37	476	0.31	0.23-0.39	983	0.27	0.21-0.32	906	0.27	0.21-0.33	
Total cholinols (mmol/L)	465	0.27	0.18-0.35	430	0.23	0.14-0.32	518	0.28	0.20-0.35	476	0.28	0.20-0.37	983	0.27	0.21-0.33	906	0.26	0.20-0.32	
Apolipoproteins																			
Apolipoprotein A1 (g/L)	469	0.28	0.20-0.36	433	0.26	0.17-0.35	518	0.26	0.18-0.34	476	0.26	0.17-0.34	987	0.27	0.21-0.33	909	0.26	0.20-0.32	
Apolipoprotein B (g/L)	469	0.26	0.18-0.35	433	0.27	0.18-0.35	517	0.30	0.22-0.38	475	0.32	0.24-0.40	986	0.29	0.23-0.35	908	0.31	0.25-0.36	
Ratio of apolipoprotein B to apolipoprotein A1	469	0.28	0.20-0.36	433	0.30	0.21-0.38	517	0.25	0.17-0.33	475	0.25	0.16-0.33	986	0.27	0.21-0.33	908	0.28	0.22-0.34	
Fatty acids																			
Total fatty acids (mmol/L)	462	0.22	0.13-0.30	427	0.22	0.13-0.31	517	0.31	0.23-0.39	475	0.33	0.25-0.41	979	0.27	0.22-0.33	902	0.29	0.22-0.34	
Estimated degree of unsaturation	462	0.30	0.22-0.38	427	0.32	0.23-0.41	517	0.27	0.19-0.35	475	0.24	0.16-0.33	979	0.29	0.23-0.34	902	0.28	0.22-0.34	
22:6, docosahexaenoic acid (mmol/L)	462	0.20	0.11-0.29	427	0.18	0.09-0.27	517	0.34	0.26-0.41	475	0.32	0.24-0.40	979	0.27	0.21-0.32	902	0.25	0.19-0.31	
18:2, linoleic acid (mmol/L)	462	0.22	0.13-0.31	427	0.22	0.13-0.31	517	0.27	0.19-0.35	475	0.30	0.21-0.38	979	0.25	0.19-0.31	902	0.27	0.21-0.33	
Omega3 fatty acids (mmol/L)	462	0.20	0.11-0.29	427	0.19	0.09-0.28	517	0.34	0.27-0.42	475	0.34	0.26-0.42	979	0.27	0.21-0.33	902	0.26	0.20-0.32	
Omega6 fatty acids (mmol/L)	462	0.23	0.14-0.32	427	0.22	0.13-0.31	517	0.31	0.23-0.38	475	0.33	0.25-0.41	979	0.27	0.21-0.33	902	0.28	0.22-0.34	
Polysat. fatty acids (mmol/L)	462	0.23	0.14-0.31	427	0.22	0.12-0.30	517	0.32	0.24-0.39	475	0.34	0.26-0.42	979	0.28	0.22-0.33	902	0.28	0.22-0.34	
Monounsat. fatty acids 16:1, 18:1 (mmol/L)	462	0.24	0.15-0.33	427	0.25	0.16-0.34	517	0.33	0.25-0.40	475	0.33	0.25-0.41	979	0.29	0.24-0.35	902	0.30	0.24-0.36	
Saturated fatty acids (mmol/L)	462	0.21	0.12-0.29	427	0.21	0.12-0.30	517	0.29	0.21-0.37	475	0.30	0.21-0.38	979	0.26	0.20-0.32	902	0.26	0.20-0.32	
Fatty acid ratios																			
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	462	0.31	0.22-0.39	427	0.29	0.20-0.38	517	0.35	0.27-0.42	475	0.32	0.24-0.40	979	0.33	0.27-0.39	902	0.31	0.25-0.36	
Ratio of 18:2 linoleic acid to total fatty acids (%)	462	0.13	0.04-0.22	427	0.15	0.06-0.24	517	0.20	0.11-0.28	475	0.16	0.08-0.25	979	0.17	0.11-0.23	902	0.16	0.09-0.22	
Ratio of omega3 fatty acids to total fatty acids (%)	462	0.32	0.24-0.40	427	0.32	0.23-0.40	517	0.40	0.33-0.47	475	0.37	0.29-0.44	979	0.36	0.31-0.41	902	0.34	0.28-0.39	
Ratio of omega6 fatty acids to total fatty acids (%)	462	0.23	0.15-0.32	427	0.26	0.17-0.35	517	0.25	0.17-0.33	475	0.22	0.13-0.30	979	0.24	0.18-0.30	902	0.24	0.17-0.30	
Ratio of polysat. fatty acids to total fatty acids (%)	462	0.27	0.19-0.36	427	0.30	0.21-0.38	517	0.28	0.20-0.36	475	0.24	0.16-0.33	979	0.28	0.22-0.33	902	0.27	0.20-	

Supplementary table 6: Parent-child concordance, correlation and partial correlations between all parents and their sons, daughters and all children.

Metabolic subgroup	All Parents																	
	Male Child					Female Child					All Children							
	n	CC	95% CI	n	PCC*	95% CI	n	CC	95% CI	n	CC	95% CI	n	PCC*	95% CI			
Lipoprotein subclass lipids																		
Total lipids in chylomicrons & ex.large VLDL (mmol/L)*	547	0.22	0.14 - 0.30	505	0.23	0.14 - 0.31	586	0.20	0.12 - 0.28	544	0.17	0.08 - 0.25	1133	0.22	0.16 - 0.27	1049	0.20	0.15 - 0.26
Total lipids in very large VLDL (mmol/L)*	547	0.23	0.15 - 0.31	505	0.23	0.15 - 0.31	586	0.17	0.10 - 0.25	544	0.16	0.08 - 0.24	1133	0.21	0.15 - 0.26	1049	0.20	0.14 - 0.26
Total lipids in large VLDL (mmol/L)*	547	0.20	0.12 - 0.28	505	0.22	0.13 - 0.30	586	0.20	0.12 - 0.28	544	0.19	0.11 - 0.27	1133	0.20	0.15 - 0.26	1049	0.21	0.15 - 0.26
Total lipids in medium VLDL (mmol/L)	547	0.27	0.19 - 0.35	505	0.28	0.20 - 0.36	586	0.25	0.17 - 0.32	544	0.24	0.16 - 0.32	1133	0.26	0.21 - 0.31	1049	0.26	0.20 - 0.32
Total lipids in small VLDL (mmol/L)	547	0.27	0.19 - 0.34	505	0.27	0.19 - 0.35	586	0.26	0.18 - 0.33	544	0.26	0.17 - 0.33	1133	0.26	0.21 - 0.32	1049	0.26	0.21 - 0.32
Total lipids in very small VLDL (mmol/L)	547	0.23	0.15 - 0.31	505	0.22	0.13 - 0.30	586	0.25	0.17 - 0.33	544	0.26	0.18 - 0.34	1133	0.25	0.19 - 0.30	1049	0.24	0.19 - 0.30
Total lipids in IDL (mmol/L)	547	0.28	0.20 - 0.35	505	0.25	0.16 - 0.33	586	0.29	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.28	0.23 - 0.34
Total lipids in large LDL (mmol/L)	547	0.28	0.20 - 0.36	505	0.25	0.17 - 0.33	586	0.29	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.29	0.23 - 0.34
Total lipids in medium LDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.25	0.17 - 0.33	586	0.28	0.21 - 0.36	544	0.31	0.23 - 0.38	1133	0.28	0.23 - 0.34	1049	0.29	0.23 - 0.34
Total lipids in small LDL (mmol/L)	547	0.28	0.20 - 0.35	505	0.24	0.16 - 0.32	586	0.28	0.20 - 0.35	544	0.31	0.23 - 0.38	1133	0.28	0.22 - 0.33	1049	0.28	0.22 - 0.33
Total lipids in very large HDL (mmol/L)	547	0.29	0.21 - 0.37	505	0.28	0.20 - 0.36	586	0.29	0.21 - 0.36	544	0.27	0.19 - 0.35	1133	0.29	0.24 - 0.34	1049	0.28	0.22 - 0.33
Total lipids in large HDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.28	0.19 - 0.36	586	0.24	0.17 - 0.32	544	0.22	0.14 - 0.30	1133	0.27	0.21 - 0.32	1049	0.25	0.19 - 0.30
Total lipids in medium HDL (mmol/L)	547	0.15	0.07 - 0.24	505	0.13	0.04 - 0.21	586	0.10	0.02 - 0.18	544	0.10	0.02 - 0.19	1133	0.12	0.07 - 0.18	1049	0.11	0.05 - 0.17
Total lipids in small HDL (mmol/L)	547	0.19	0.10 - 0.27	505	0.16	0.08 - 0.25	586	0.18	0.10 - 0.26	544	0.17	0.09 - 0.25	1133	0.18	0.12 - 0.23	1049	0.17	0.11 - 0.22
Lipoprotein particle size																		
Mean diameter for VLDL particles (nm)	547	0.29	0.21 - 0.36	505	0.30	0.22 - 0.38	586	0.22	0.14 - 0.29	544	0.19	0.11 - 0.27	1133	0.25	0.20 - 0.31	1049	0.24	0.19 - 0.30
Mean diameter for LDL particles (nm)	547	0.19	0.11 - 0.27	505	0.17	0.08 - 0.25	586	0.27	0.19 - 0.34	544	0.29	0.21 - 0.36	1133	0.23	0.17 - 0.28	1049	0.23	0.17 - 0.28
Mean diameter for HDL particles (nm)	547	0.31	0.23 - 0.38	505	0.30	0.22 - 0.38	586	0.30	0.22 - 0.37	544	0.27	0.19 - 0.35	1133	0.30	0.25 - 0.35	1049	0.29	0.23 - 0.34
Cholesterol																		
Serum total cholesterol (mmol/L)	547	0.28	0.20 - 0.36	505	0.24	0.16 - 0.32	586	0.31	0.24 - 0.38	544	0.33	0.26 - 0.41	1133	0.30	0.24 - 0.35	1049	0.29	0.24 - 0.35
Total cholesterol in VLDL (mmol/L)	547	0.26	0.18 - 0.34	505	0.27	0.19 - 0.35	586	0.26	0.18 - 0.33	544	0.26	0.18 - 0.34	1133	0.26	0.21 - 0.32	1049	0.27	0.21 - 0.32
Remnant cholesterol (nonHDL, nonLDL cholesterol) (mmol/L)	547	0.27	0.19 - 0.34	505	0.26	0.18 - 0.34	586	0.28	0.20 - 0.35	544	0.29	0.21 - 0.37	1133	0.27	0.22 - 0.33	1049	0.28	0.22 - 0.34
Total cholesterol in LDL (mmol/L)	547	0.29	0.21 - 0.36	505	0.26	0.17 - 0.34	586	0.28	0.21 - 0.35	544	0.30	0.23 - 0.38	1133	0.29	0.23 - 0.34	1049	0.28	0.23 - 0.34
Total cholesterol in HDL (mmol/L)	547	0.27	0.19 - 0.35	505	0.25	0.17 - 0.33	586	0.22	0.14 - 0.29	544	0.20	0.12 - 0.28	1133	0.24	0.19 - 0.30	1049	0.22	0.17 - 0.28
Total cholesterol in HDL2 (mmol/L)	547	0.27	0.19 - 0.34	505	0.25	0.17 - 0.33	586	0.21	0.14 - 0.29	544	0.20	0.11 - 0.28	1133	0.24	0.18 - 0.29	1049	0.22	0.16 - 0.28
Total cholesterol in HDL3 (mmol/L)	547	0.25	0.17 - 0.33	505	0.22	0.14 - 0.30	586	0.23	0.15 - 0.31	544	0.23	0.14 - 0.30	1133	0.24	0.19 - 0.30	1049	0.22	0.16 - 0.28
Esterified cholesterol (mmol/L)	543	0.28	0.20 - 0.36	502	0.24	0.15 - 0.32	584	0.31	0.24 - 0.38	542	0.33	0.25 - 0.40	1127	0.30	0.24 - 0.35	1044	0.29	0.23 - 0.34
Free cholesterol (mmol/L)	543	0.27	0.19 - 0.34	502	0.23	0.14 - 0.31	584	0.32	0.24 - 0.39	542	0.34	0.26 - 0.41	1127	0.29	0.24 - 0.35	1044	0.29	0.23 - 0.34
Glycerides and phospholipids																		
Serum total triglycerides (mmol/L)*	547	0.28	0.20 - 0.35	505	0.29	0.20 - 0.36	586	0.25	0.17 - 0.32	544	0.24	0.16 - 0.32	1133	0.26	0.21 - 0.32	1049	0.26	0.21 - 0.32
Triglycerides in VLDL (mmol/L)*	547	0.28	0.20 - 0.36	505	0.29	0.21 - 0.37	586	0.23	0.15 - 0.30	544	0.21	0.13 - 0.29	1133	0.26	0.20 - 0.31	1049	0.25	0.20 - 0.31
Triglycerides in LDL (mmol/L)	547	0.20	0.12 - 0.28	505	0.19	0.10 - 0.27	586	0.27	0.19 - 0.34	544	0.28	0.20 - 0.35	1133	0.24	0.18 - 0.29	1049	0.24	0.18 - 0.29
Triglycerides in HDL (mmol/L)	547	0.25	0.17 - 0.33	505	0.26	0.18 - 0.34	586	0.27	0.20 - 0.35	544	0.28	0.20 - 0.35	1133	0.26	0.21 - 0.32	1049	0.27	0.21 - 0.32
Total phosphoglycerides (mmol/L)	543	0.28	0.20 - 0.36	502	0.24	0.16 - 0.32	584	0.27	0.19 - 0.34	542	0.29	0.21 - 0.36	1127	0.27	0.22 - 0.33	1044	0.26	0.20 - 0.32
Ratio of triglycerides to phosphoglycerides	543	0.23	0.15 - 0.31	502	0.24	0.16 - 0.32	584	0.21	0.13 - 0.28	542	0.20	0.12 - 0.28	1127	0.22	0.16 - 0.27	1044	0.22	0.16 - 0.28
Phosphatidylcholine & other cholines (mmol/L)	542	0.28	0.20 - 0.36	501	0.23	0.15 - 0.32	584	0.26	0.19 - 0.34	542	0.28	0.20 - 0.36	1126	0.27	0.21 - 0.32	1043	0.26	0.20 - 0.31
Sphingomyelins (mmol/L)	542	0.23	0.15 - 0.31	501	0.20	0.12 - 0.29	584	0.28	0.21 - 0.36	542	0.30	0.22 - 0.37	1126	0.26	0.20 - 0.31	1043	0.26	0.20 - 0.31
Total cholines (mmol/L)	542	0.27	0.19 - 0.35	501	0.23	0.14 - 0.31	584	0.29	0.21 - 0.36	542	0.30	0.22 - 0.37	1126	0.28	0.22 - 0.33	1043	0.26	0.20 - 0.32
Apolipoproteins																		
Apolipoprotein A1 (g/L)	547	0.26	0.18 - 0.34	505	0.23	0.14 - 0.31	586	0.25	0.17 - 0.33	544	0.25	0.17 - 0.32	1133	0.25	0.20 - 0.31	1049	0.23	0.18 - 0.29
Apolipoprotein B (g/L)	547	0.27	0.19 - 0.35	505	0.27	0.19 - 0.35	585	0.28	0.20 - 0.35	543	0.30	0.22 - 0.37	1132	0.28	0.23 - 0.33	1048	0.29	0.23 - 0.34
Ratio of apolipoprotein B to apolipoprotein AI	547	0.26	0.18 - 0.33	505	0.27	0.18 - 0.34	585	0.21	0.13 - 0.29	543	0.20	0.12 - 0.28	1132	0.23	0.18 - 0.29	1048	0.24	0.18 - 0.29
Fatty acids																		
Total fatty acids (mmol/L)	537	0.26	0.18 - 0.33	496	0.25	0.17 - 0.33	583	0.30	0.22 - 0.37	541	0.31	0.23 - 0.39	1120	0.28	0.23 - 0.33	1037	0.29	0.23 - 0.34
Estimated degree of unsaturation	537	0.30	0.22 - 0.37	496	0.32	0.23 - 0.39	583	0.24	0.17 - 0.32	541	0.22	0.14 - 0.30	1120	0.27	0.21 - 0.32	1037	0.26	0.20 - 0.32
22:6, docosahexaenoic acid (mmol/L)	537	0.23	0.15 - 0.31	496	0.21	0.13 - 0.30	583	0.33	0.26 - 0.40	541	0.33	0.25 - 0.40	1120	0.28	0.23 - 0.34	1037	0.27	0.21 - 0.32
18:2, linoleic acid (mmol/L)	537	0.24	0.16 - 0.32	496	0.25	0.16 - 0.33	583	0.27	0.19 - 0.34	541	0.29	0.21 - 0.37	1120	0.26	0.21 - 0.32	1037	0.28	0.22 - 0.33
Omega3 fatty acids (mmol/L)	537	0.24	0.16 - 0.32	496	0.22	0.14 - 0.31	583	0.34	0.27 - 0.41	541	0.34	0.27 - 0.41	1120	0.29	0.23 - 0.34	1037	0.28	0.22 - 0.33
Omega6 fatty acids (mmol/L)	537	0.26	0.18 - 0.33	496	0.25	0.16 - 0.33	583	0.30	0.23 - 0.38	541	0.33	0.25 - 0.40	1120	0.28	0.23 - 0.34	1037	0.29	0.24 - 0.35
Polysat. fatty acids (mmol/L)	537	0.26	0.17 - 0.33	496	0.24	0.16 - 0.33	583	0.31	0.24 - 0.39	541	0.34	0.26 - 0.41	1120	0.29	0.23 - 0.34	1037	0.29	0.24 - 0.35
Monounsat. fatty acids; 16:1, 18:1 (mmol/L)	537	0.27	0.19 - 0.35	496	0.28	0.19 - 0.36	583	0.30	0.22 - 0.37	541	0.30	0.23 - 0.38	1120	0.29	0.24 - 0.34	1037	0.29	0.24 - 0.35
Saturated fatty acids (mmol/L)	537	0.25	0.17 - 0.33	496	0.25	0.16 - 0.33	583	0.28	0.20 - 0.35	541	0.28	0.20 - 0.36	1120	0.27	0.21 - 0.32	1037	0.27	0.21 - 0.32
Fatty acid ratios																		
Ratio of 22:6 docosahexaenoic acid to total fatty acids (%)	537	0.31	0.23 - 0.38	496	0.30	0.21 - 0.37	583	0.33	0.26 - 0.40	541	0.31	0.23 - 0.38	1120	0.32	0.27 - 0.37	1037	0.30	0.24 - 0.36
Ratio of 18:2 linoleic acid to total fatty acids (%)	537	0.15	0.07 - 0.23	496	0.17	0.08 - 0.25	583	0.18	0.10 - 0.26	541	0.16	0.08 - 0.24	1120	0.17	0.11 - 0.23	1037	0.16	0.10 - 0.22
Ratio of omega3 fatty acids to total fatty acids (%)	537	0.32	0.24 - 0.39	496	0.32	0.23 - 0.39	583	0.40	0.33 - 0.47	541	0.38	0.31 - 0.45	1120	0.36	0.31 - 0.41	1037	0.35	0.29 - 0.40
Ratio of omega6 fatty acids to total fatty acids (%)	537	0.24	0.16 -															

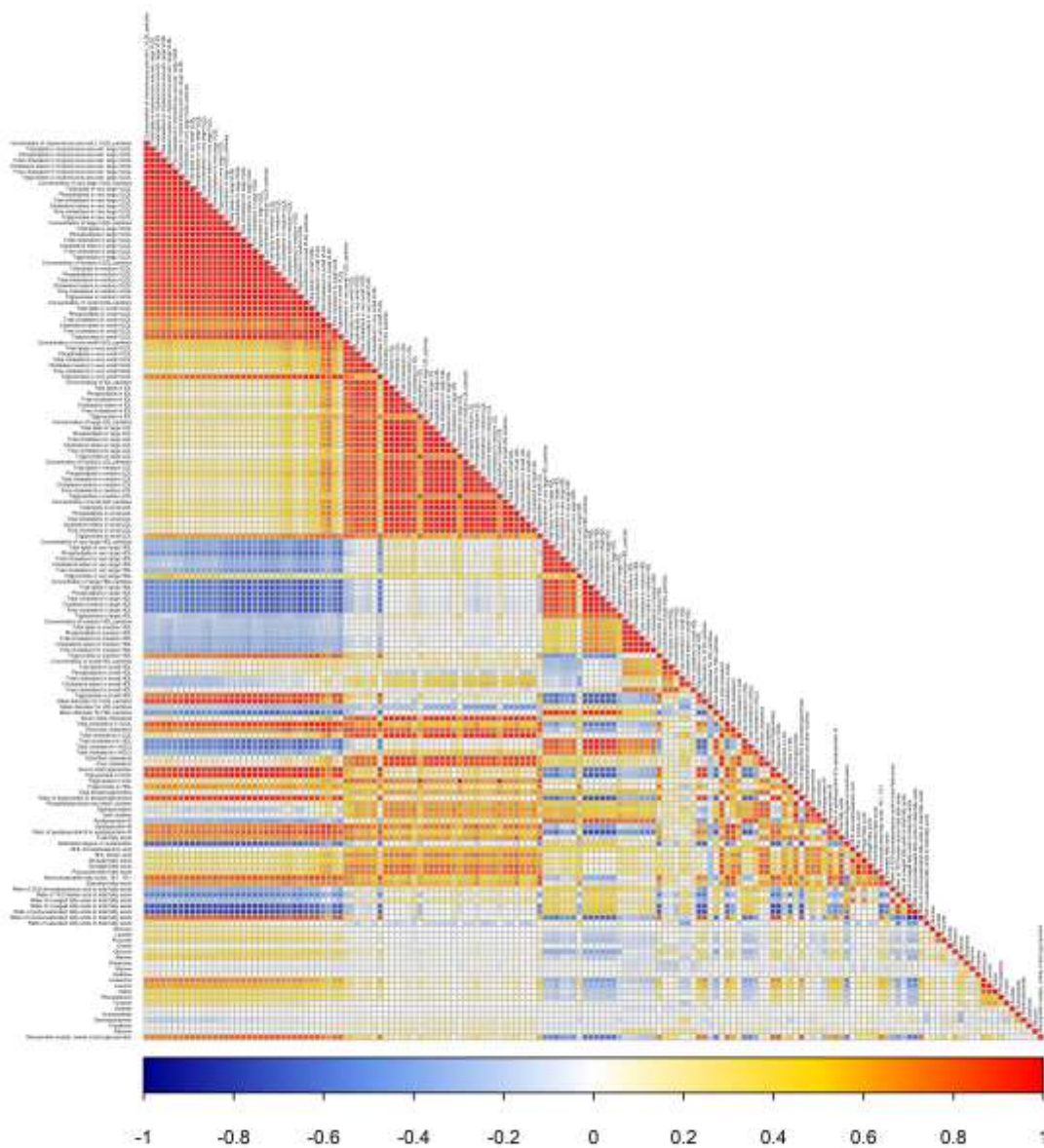
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Supplementary Figure 1: Correlations between Child Health CheckPoint metabolites - children



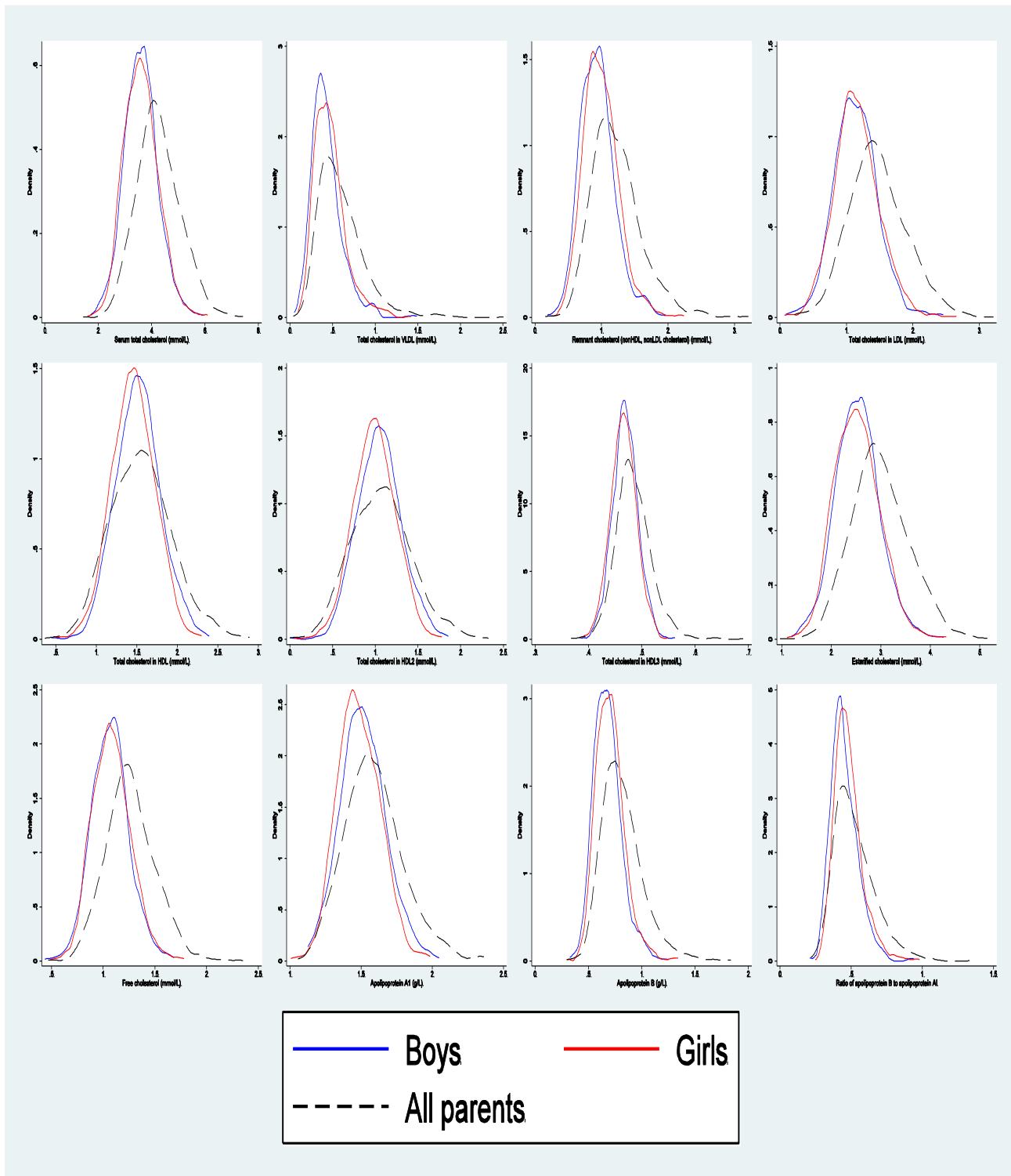
Note: Correlations (spearman) between metabolites for the CheckPoint child metabolomics data

Supplementary Figure 2: Correlations between Child Health CheckPoint metabolites - parents

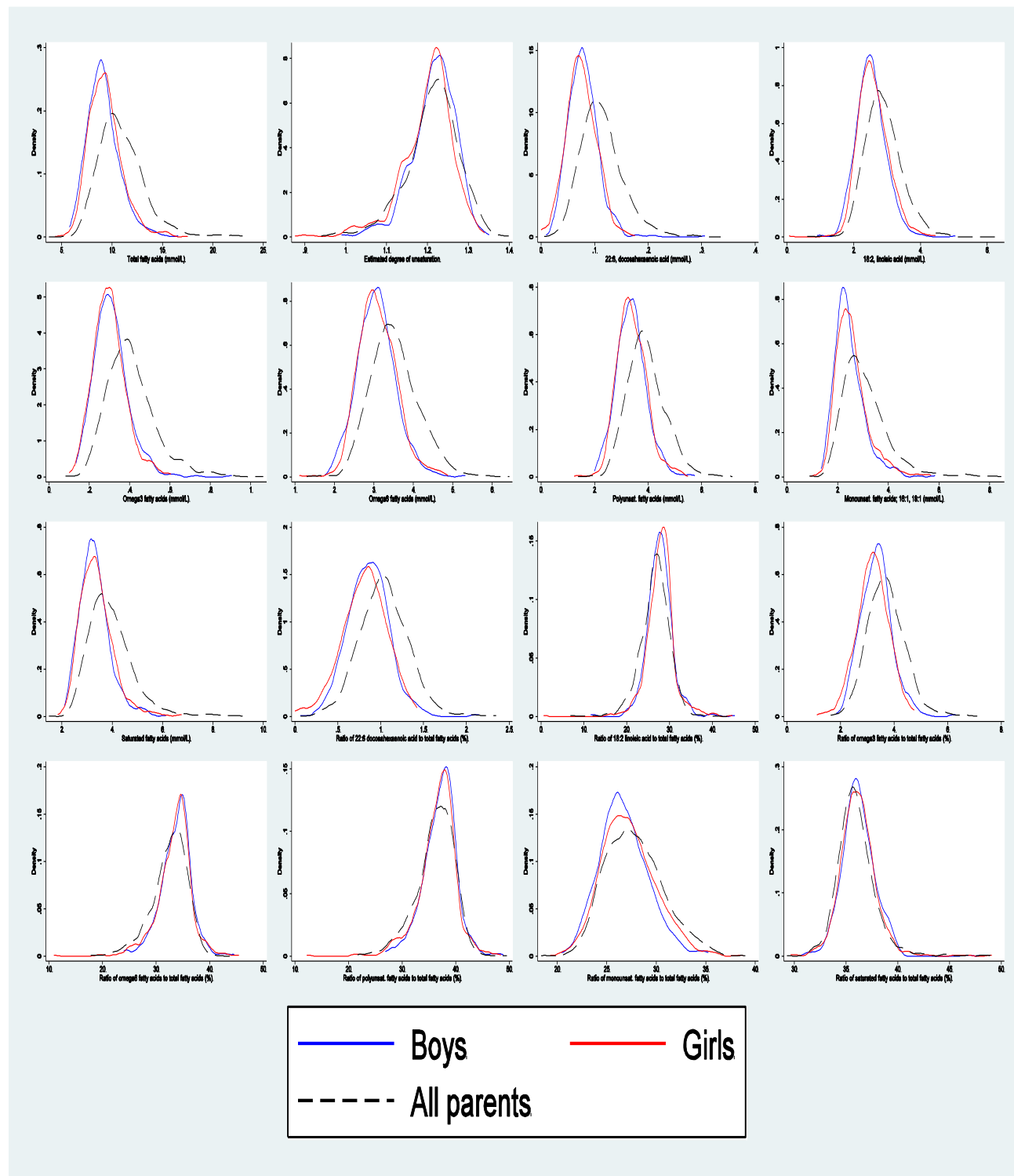


Note: Correlations (spearman) between metabolites for the CheckPoint parent metabolomics data

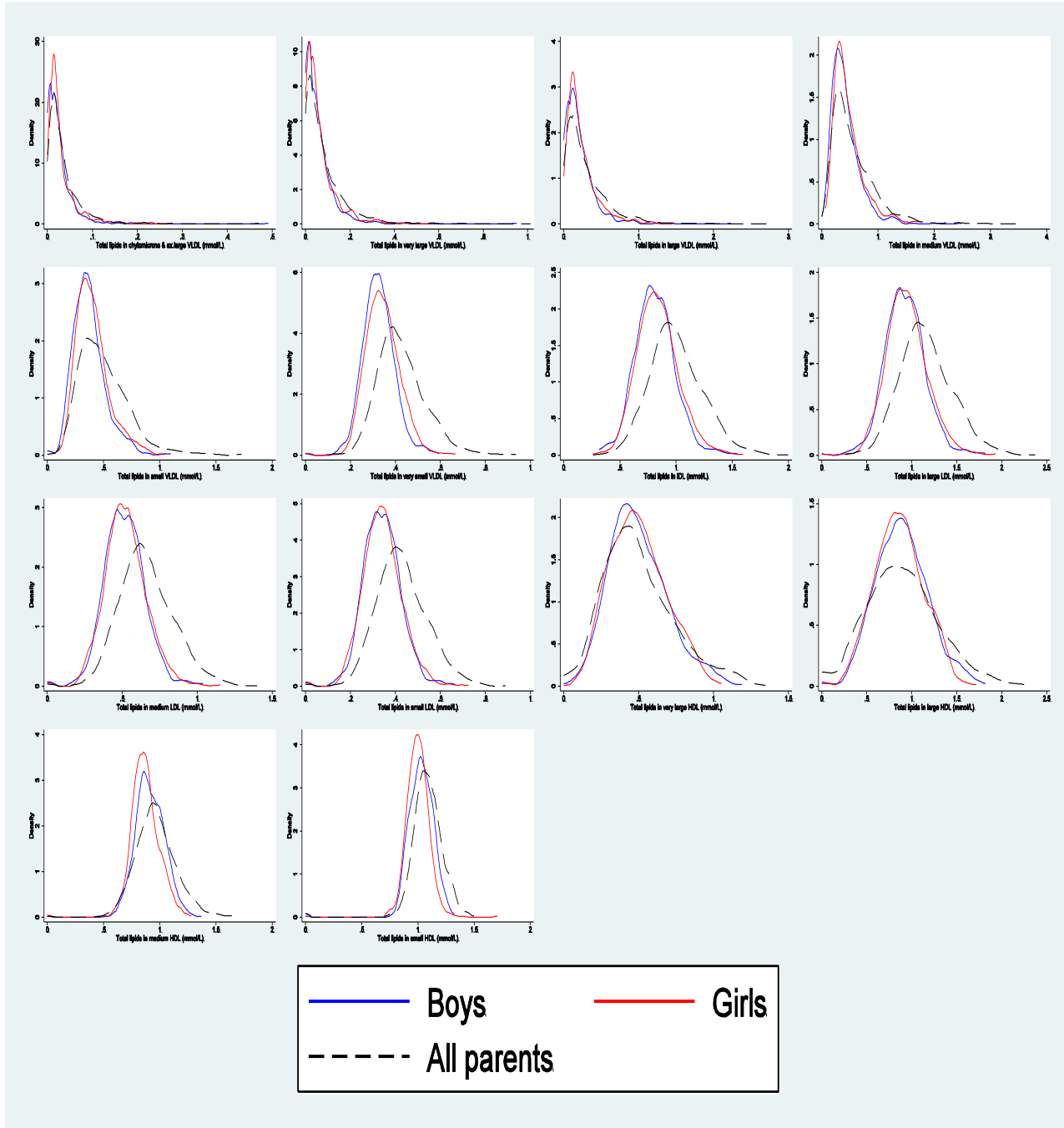
Supplementary Figure 3



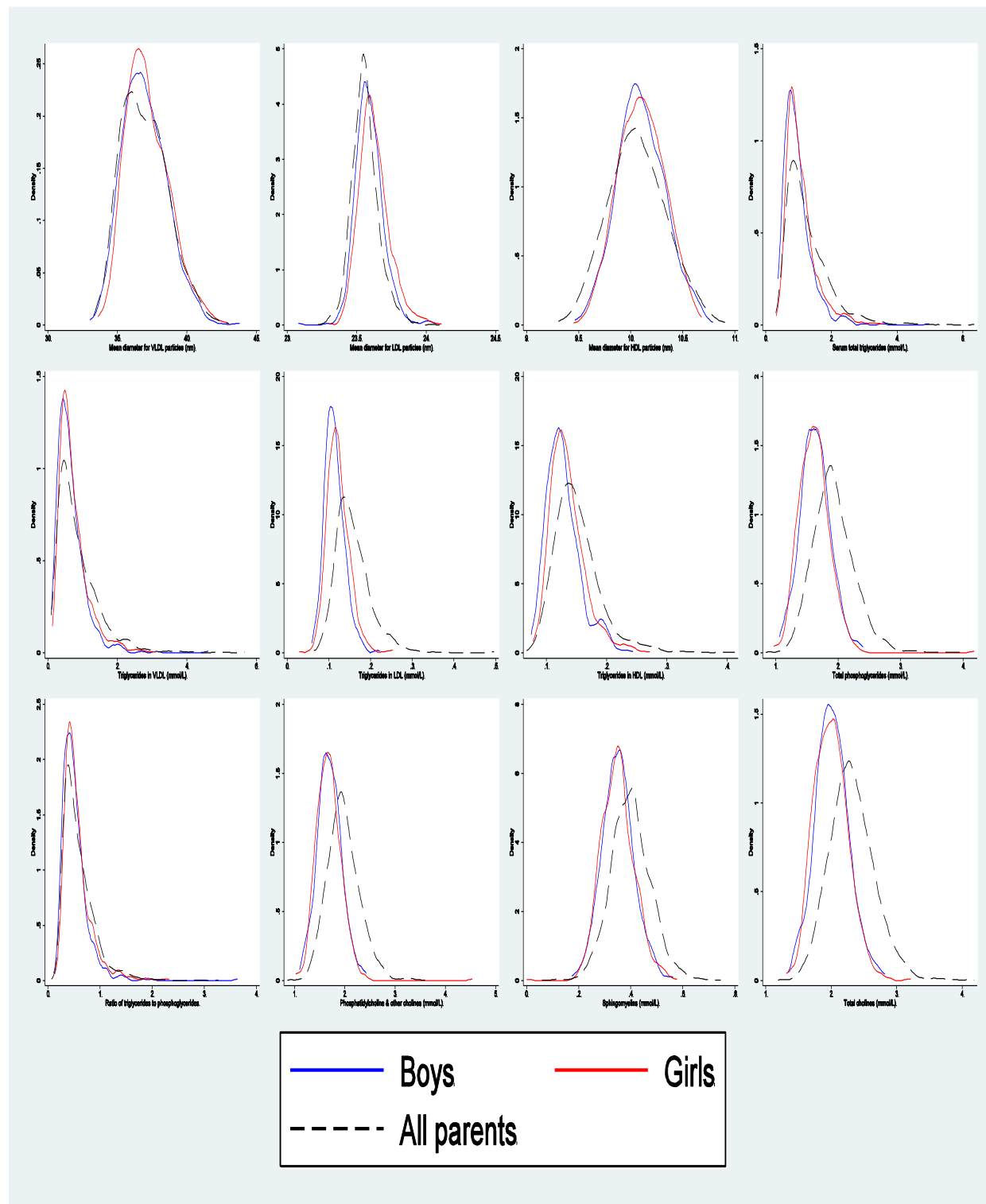
Supplementary Figure 4



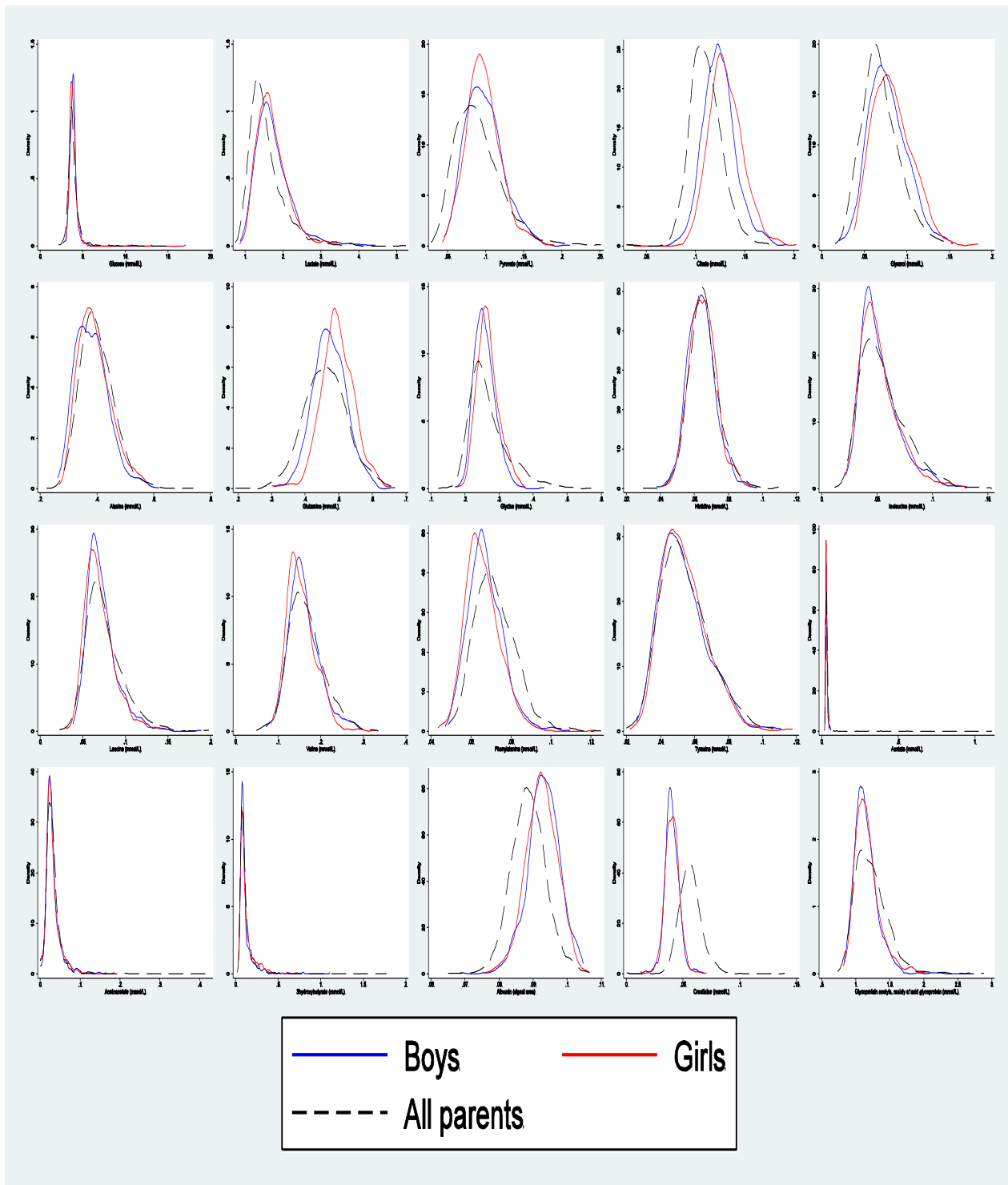
Supplementary Figure 5



Supplementary Figure 6



Supplementary Figure 7



STROBE Statement—checklist of items that should be included in reports of observational studies

Paper title: Metabolomics: Population epidemiology and concordance in 11-12 year old

Australians and their parents

Person completing checklist: Susan Ellul

	Item No	Recommendation	Page number	Line number
Title and abstract	1	(a) Indicate the study's design with a commonly used term in the title or the abstract	2	14-15
		(b) Provide in the abstract an informative and balanced summary of what was done and what was found	2	9-13 25-32
Introduction				
Background/rationale	2	Explain the scientific background and rationale for the investigation being reported	4-5	Pg 4 (3-9 ,21-22, 31-32, 33-35) Pg 5 (5-7)
Objectives	3	State specific objectives, including any prespecified hypotheses	5	8-11
Methods				
Study design	4	Present key elements of study design early in the paper	5	14-22
Setting	5	Describe the setting, locations, and relevant dates, including periods of recruitment, exposure, follow-up, and data collection	5-6	Pg 5 (17-20) Pg 6 (3-32)
Participants	6	(a) <i>Cohort study</i> —Give the eligibility criteria, and the sources and methods of selection of participants. Describe methods of follow-up (b) <i>Case control study</i>—Give the eligibility criteria, and the sources and methods of case ascertainment and control selection. Give the rationale for the choice of cases and controls <i>Cross-sectional study</i> —Give the eligibility criteria, and the sources and methods of selection of participants	5	23-31
		(b) <i>Cohort study</i> —For matched studies, give matching criteria and number of exposed and unexposed <i>Case control study</i>—For matched studies, give matching criteria and the number of controls per case		
Variables	7	Clearly define all outcomes, exposures, predictors, potential confounders, and effect modifiers. Give diagnostic criteria, if applicable	7-9	Pg 7 (4-17) Pg 8 (table 1, 9-16) Pg 9 (1-24)
Data sources/ measurement	8*	For each variable of interest, give sources of data and details of methods of assessment (measurement). Describe comparability of assessment methods if there is more than one group	7-9	Pg 7 (4-17) Pg 8 (table 1, 9-16) Pg 9 (1-24)
Bias	9	Describe any efforts to address potential sources of bias	9 -10	Pg 9 (31-35) Pg 10 (16-18)

Study size	10	Explain how the study size was arrived at	11	5-14, figure 1
Quantitative variables	11	Explain how quantitative variables were handled in the analyses. If applicable, describe which groupings were chosen and why	9-10	Pg 9 (28-35) Pg 10 (3-15)
Statistical methods	12	(a) Describe all statistical methods, including those used to control for confounding	9-10	Pg 9 (28-35) Pg 10 (3-15)
		(b) Describe any methods used to examine subgroups and interactions	9-10	Pg 9 (28-35) Pg 10 (3-15)
		(c) Explain how missing data were addressed	9 -10	Pg 9 (31-35) Pg 10 (16-18)
		(d) <i>Cohort study</i> —If applicable, explain how loss to follow-up was addressed	9	31-35
		(e) <i>Case-control study</i>—If applicable, explain how matching of cases and controls was addressed		
		(e) Describe any sensitivity analyses	10	16-18

Results			Page number	Line number
Participants	13*	(a) Report numbers of individuals at each stage of study—eg numbers potentially eligible, examined for eligibility, confirmed eligible, included in the study, completing follow-up, and analysed	11 and figure 1	5-12
		(b) Give reasons for non-participation at each stage	11 6 figure 1	8-12 7-11
		(c) Consider use of a flow diagram	figure 1	
Descriptive data	14*	(a) Give characteristics of study participants (eg demographic, clinical, social) and information on exposures and potential confounders	12 (table 2)	2
		(b) Indicate number of participants with missing data for each variable of interest	figure 1 table 2 sup Table 1	2
		(c) <i>Cohort study</i> —Summarise follow-up time (eg, average and total amount)	NA	
Outcome data	15*	<i>Cohort study</i> —Report numbers of outcome events or summary measures over time	sup table 1	
		<i>Case-control study</i> —Report numbers in each exposure category, or summary measures of exposure	NA	
		<i>Cross-sectional study</i> —Report numbers of outcome events or summary measures	NA	
Main results	16	(a) Give unadjusted estimates and, if applicable, confounder-adjusted estimates and their precision (eg, 95% confidence interval). Make clear which confounders were adjusted for and why they were included	13 supp tables 2-6	21-24
		(b) Report category boundaries when continuous variables were categorized	NA	

		(c) If relevant, consider translating estimates of relative risk into absolute risk for a meaningful time period	NA	
Other analyses	17	Report other analyses done—eg analyses of subgroups and interactions, and sensitivity analyses	supp tables 2-6	
Discussion				
Key results	18	Summarise key results with reference to study objectives	14	3-10
Limitations	19	Discuss limitations of the study, taking into account sources of potential bias or imprecision. Discuss both direction and magnitude of any potential bias	14	18-28
Interpretation	20	Give a cautious overall interpretation of results considering objectives, limitations, multiplicity of analyses, results from similar studies, and other relevant evidence	14-17	Pg 14 (30-32) Pg 15 (all) Pg 16 (all) Pg 17 (1-2)
Generalisability	21	Discuss the generalisability (external validity) of the study results	14	18-28
Other information				
Funding	22	Give the source of funding and the role of the funders for the present study and, if applicable, for the original study on which the present article is based	18-19	Pg 18 (21-33) Pg 19 (1-2)

*Give information separately for cases and controls in case-control studies and, if applicable, for exposed and unexposed groups in cohort and cross-sectional studies.

Note: An Explanation and Elaboration article discusses each checklist item and gives methodological background and published examples of transparent reporting. The STROBE checklist is best used in conjunction with this article (freely available on the Web sites of PLoS Medicine at <http://www.plosmedicine.org/>, Annals of Internal Medicine at <http://www.annals.org/>, and Epidemiology at <http://www.epidem.com/>). Information on the STROBE Initiative is available at www.strobe-statement.org.