ISCI, Volume 17

Supplemental Information

Super-Enhancer-Associated LncRNA UCA1

Interacts Directly with AMOT to Activate

YAP Target Genes in Epithelial Ovarian Cancer

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5kb



Figure S1. No evidence of super-enhancer activity at the OR10H1 gene locus, Related to Figure 2

(A) H3K27ac signal at the *OR10H1* gene locus is shown at log scale. (B) Super-enhancer signals positively correlate with *OR10H1* expressions (Spearman's correlation, r=0.47, *P*=0.043). Counts per million (CPM) of *UCA1* from RNA-Seq is shown.



Figure S2. Overexpressed *UCA1* correlates with lower YAP phosphorylation in primary high-grade serous EOCs, Related to Figure 4A

(A) Comparison of RPPA data for YAPpS127 between ovarian cancers with (Altered) or without (Unaltered) *UCA1* amplification. (B) Total YAP is not significantly different between the two groups. Z-score threshold was set at 2. Data are from TCGA (<u>http://www.cbioportal.org</u>). *P*-value was determined using Student's *t*-test, while the *Q*-value was derived from Benjamini-Hochberg procedure.



Figure S3. iRAP identifies the direct interacting proteome of UCA1, Related to Figure 5A

(A) Probe designing for UCA1-iRAP. Two sets of non-overlapping antisense DNA probes for UCA1 were designed and labeled with biotin at their 3' end. An antisense DNA probe complementary to 1-20 nt of U1snRNA was included as a positive control. (B) RT-qPCR analysis of RNA retrieved from iRAP experiments. Data shown are mean \pm SD from 4 independent experiments. (C) Peptides were retrieved from U1 iRAP-MS for direct binding proteins of U1 snRNA. Numbers indicate the starting amino acid position and the ending amino acid position of the peptide retrieved or the full length of target protein. Protein length is not drawn in scale. (D) The same peptides were reproducibly retrieved from UCA1 iRAP-MS experiments. A selection of representative proteins is shown. Numbers indicate the starting amino acid position and the ending amino acid position of the peptide retrieved or the full length of target protein. Protein length is not drawn in scale. (D)



Figure S4. LncRNA UCA1 predominantly localizes in cytoplasm, Related to Figure 6

(A) RT-qPCR measurement of cytoplasmic *18S* rRNA, nuclear *U6* snRNA and *UCA1* in OVCA429 fractionated samples. (B) RT-qPCR measurement of cytoplasmic *18S* rRNA, nuclear *U6* snRNA and *UCA1* in OSEC4C2 fractionated samples. Data shown are mean \pm SD from 3 independent experiments.

LncRNA Name	Importance-score	Raw- score	Chr	Start	End	Width	Width Strand Gene T	
TTTY14	1.06572E+13	1.723	chrY	18872501	19077416	204916	-	lincRNA
PSMB1	99159107045	2.907	chr6	170535117	170553341	18225	-	protein_coding
ZFAS1	48072327683	2.585	chr20	49278178	49295738	17561	+	antisense
TUG1	40492304785	1.923	chr22	30970677	30979395	8719	+	antisense
FGD5-AS1	35063860805	2.147	chr3	14920347	14948424	28078	-	antisense
RP11-554D15.1	29197615843	-1.956	chr6	74069451	74690727	621277	+	lincRNA
CTB-5506.12	23922544460	1.501	chr19	14137179	14171267	34089	+	antisense
GAS5	22959917722	3.715	chr1	173863900	173868882	4983	-	processed_transcript
RP5-1061H20.4	22563332287	2.269	chr1	229258281	229271028	12748	-	lincRNA
RP11-64D24.2	18447025405	2.552	chr11	114210616	114356571	145956	-	antisense
RP11-6N17.9	17963712426	1.799	chr17	47945424	47974438	29015	+	processed_transcript
OIP5-AS1	16976708028	2.315	chr15	41283990	41309737	25748	+	processed_transcript
SNHG15	16315350372	2.006	chr7	44983023	44986961	3939	-	lincRNA
RP11-865I6.2	15818503929	1.567	chr8	68848742	68852763	4022	-	lincRNA
ILF3-AS1	15338973685	1.479	chr19	10651862	10653844	1983	-	lincRNA
SNHG8	11732839093	2.472	chr4	118278709	118279823	1115	+	lincRNA
NOP14-AS1	11574773571	1.796	chr4	2934899	2961738	26840	+	antisense
LINC00662	11461006830	1.478	chr19	27684580	27793940	109361	-	lincRNA
AC005083.1	10597230777	2.135	chr7	20217577	20221700	4124	+	processed_transcript
LINC00648	10263227364	2.269	chr14	47764954	47795092	30139	-	lincRNA
AC004112.4	10158300092	2.129	chr7	112328189	112409623	81435	-	antisense
RP4-758J18.2	9884322048	1.601	chr1	1399522	1402046	2525	+	processed_transcript
RP11-38J22.3	9686546170	3.22	chr1	206117783	206126805	9023	+	antisense
RP11-458F8.4	7896100113	1.608	chr7	66902857	66906297	3441	+	lincRNA
SNHG9	7431812371	1.628	chr16	1964959	1965509	551	+	lincRNA
AJ006995.3	7240196117	-1.844	chr21	27954922	27985295	30374	+	lincRNA
LINC00339	6936706793	2.163	chr1	22024531	22031225	6695	+	lincRNA
LINC00578	6647351610	2.907	chr3	177441921	177752305	310385	+	lincRNA
RP11-47A8.5	6296575822	2.823	chr10	102642792	102644140	1349	-	lincRNA
SNHG17	6267686164	3.08	chr20	38420588	38435353	14766	-	processed_transcript
LINC00910	6258879085	2.145	chr17	43369845	43389199	19355	-	lincRNA
OSER1-AS1	6149104986	2.444	chr20	44210960	44226027	15068	+	lincRNA
C8orf31	6007255795	1.443	chr8	143039209	143059942	20734	+	processed_transcript
RP5-1132H15.1	5980054129	1.991	chr7	66119603	66165011	45409	-	antisense

Table S1. Prognostic lncRNAs in high-grade serous EOC, Related to Figure 2A

-	RP11-430C7.4	5776440431	2.282	chr1	204603035	204616565	13531	+	antisense
	RP11-191L9.4	5456541984	1.633	chr22	47631674	47855600	223927	+	lincRNA
	RP11-115C21.2	5322360392	2.492	chr8	6403551	6407142	3592	-	antisense
	RP11-296I10.3	5160521700	2.558	chr16	70156340	70173448	17109	-	antisense
	UCA1	5062358192	1.923	chr19	15828961	15836320	7360	+	processed_transcript
	RP11-799D4.4	5028502297	1.529	chr17	35231450	35242963	11514	-	antisense
	RP11-526P5.2	4914956869	4.806	chr10	2501783	2567239	65457	+	lincRNA
	LINC00284	4896454541	2.783	chr13	43908669	44030461	121793	+	lincRNA
	FAM157C	-4806404688	2.891	chr16	90102271	90186204	83934	+	lincRNA
	CTC-444N24.11	-5585259828	1.666	chr19	57304305	57308562	4258	+	lincRNA
	Clorf132	-5802882217	1.689	chr1	207801518	207879096	77579	-	lincRNA
	RMRP	-6118555929	1.777	chr9	35657751	35658018	268	-	lincRNA
	RAD51-AS1	-7420839971	1.729	chr15	40686724	40695107	8384	-	processed_transcript
	MIR205HG	-17249780561	1.462	chr1	209428820	209432838	4019	+	processed_transcript
	RASSF8-AS1	-23114357641	2.725	chr12	25939329	25959765	20437	-	antisense
	NEAT1	-29777169121	1.896	chr11	65422774	65445540	22767	+	lincRNA

Table S2. Super-enhancer-associated lncRNAs in high-grade serous EOCs, Related to Figure 2A

LncRNA Name	Chr	Start	End	Width	Strand	Gene Type
ZNF503-AS2	chr10	77160759	77168738	7980	+	processed_transcript
ZEB2-AS1	chr2	145275664	145279058	3395	+	antisense
ZBTB11-AS1	chr3	101395274	101398061	2788	+	antisense
WT1-AS	chr11	32457064	32480315	23252	+	antisense
UNC5B-AS1	chr10	72976981	72977985	1005	-	antisense
UCA1	chr19	15939771	15947130	7360	+	processed_transcript
UBXN8	chr8	30589764	30624522	34759	+	processed_transcript
TMPRSS4-AS1	chr11	117886487	117957508	71022	-	antisense
TMEM72-AS1	chr10	45306472	45455137	1E+05	-	antisense
THAP9-AS1	chr4	83814162	83822113	7952	-	antisense
TFAP2A-AS1	chr6	10409573	10416679	7107	+	antisense
STK4-AS1	chr20	43592435	43595043	2609	-	lincRNA
SSSCA1-AS1	chr11	65337131	65337744	614	-	antisense
SRP14-AS1	chr15	40331512	40359491	27980	+	lincRNA
SPPL2B	chr19	2328614	2355099	26486	+	processed_transcript
SNORA67	chr17	7476144	7485342	9199	+	processed_transcript
SNORA59B	chr17	19460524	19461224	701	+	sense intronic
SNHG9	chr16	2014960	2015510	551	+	lincRNA
SNHG16	chr17	74553848	74561430	7583	+	processed transcript
SNHG15	chr7	45022622	45026560	3939	-	lincRNA
SNHG10	chr14	95998634	96001209	2576	-	antisense
SNHG1	chr11	62619460	62623386	3927	-	processed transcript
SNAI3-AS1	chr16	88729706	88753594	23889	+	antisense
SLC2A1-AS1	chr1	43424720	43449029	24310	+	lincRNA
SENP3-EIF4A1	chr17	7466604	7482033	15430	+	processed transcript
SEMA3B	chr3	50304990	50314977	9988	+	processed transcript
RPS10P7	chr1	201487831	201499602	11772	+	lincRNA
RNF157-AS1	chr17	74136637	74150731	14095	+	antisense
PTOV1-AS1	chr19	50341896	50354933	13038	-	antisense
PSMG3-AS1	chr7	1609709	1629262	19554	+	lincRNA
PROSER2-AS1	chr10	11891612	11936699	45088	-	antisense
PLAC4	chr21	42548249	42558715	10467	-	antisense
PDCD4-AS1	chr10	112629626	112631991	2366	-	antisense
PCOLCE-AS1	chr7	100187025	100201829	14805	-	antisense
PCED1B-AS1	chr12	47599681	47610239	10559	-	processed transcript
PCBP1-AS1	chr2	70189395	70315978	1E+05	-	antisense
NEAT1	chr11	65190245	65213011	22767	+	lincRNA
MYLK-AS1	chr3	123304389	123363415	59027	+	antisense
MRPL23-AS1	chr11	2004467	2011150	6684	-	antisense
MIR29B1	chr7	130561495	130598069	36575	-	lincRNA
MIR22HG	chr17	1614805	1620468	5664	-	lincRNA
MIR210HG	chr11	565660	568457	2798	-	lincRNA
MIR194-2	chr11	64658827	64660921	2095	-	lincRNA
MIR142	chr17	56408245	56409869	1625	-	antisense
MIR10A	chr17	46656992	46659621	2630	-	sense intronic
MEIS1-AS3	chr2	66653867	66660602	6736	-	antisense
MAP3K14-AS1	chr17	43325292	43345997	20706	+	antisense
MAP3K14	chr17	43340488	43394414	53927	-	processed transcript
LMCD1-AS1	chr3	7994492	8653610	7E+05	-	antisense
LINC01011	chr6	2988201	2991407	3207	+	lincRNA
LINC01004	chr7	104590762	104653491	62730	-	antisense
	/					

LINC00974	chr17	39705858	39710747	4890	-	lincRNA
LINC00938	chr12	46119510	46121558	2049	-	lincRNA
LINC00899	chr22	46435787	46440733	4947	-	processed_transcript
LINC00857	chr10	81967466	81979413	11948	+	lincRNA
LINC00847	chr5	180257957	180262726	4770	+	lincRNA
LINC00638	chr14	105287538	105290055	2518	+	lincRNA
LINC00605	chr14	103653558	103655365	1808	-	lincRNA
LINC00518	chr6	10429488	10435107	5620	-	lincRNA
LINC00511	chr17	70319264	70636611	3E+05	-	lincRNA
LINC00336	chr6	33553883	33561115	7233	-	antisense
LINC00324	chr17	8123960	8127361	3402	-	lincRNA
LINC00319	chr21	44866481	44873773	7293	+	lincRNA
LINC00313	chr21	44881974	44899414	17441	-	lincRNA
LINC00176	chr20	62665697	62671315	5619	+	lincRNA
LINC00163	chr21	46409779	46414001	4223	-	lincRNA
LINC00114	chr21	40110945	40119384	8440	-	lincRNA
LINC00111	chr21	43099341	43117496	18156	+	lincRNA
KRTAP5-AS1	chr11	1592583	1620414	27832	+	antisense
ITGB2-AS1	chr21	46340966	46349593	8628	+	antisense
HPN-AS1	chr19	35549963	35597208	47246	-	antisense
HOXB-AS3	chr17	46626992	46683776	56785	+	antisense
HOXB-AS1	chr17	46620913	46628610	7698	+	antisense
HCG11	chr6	26522076	26526807	4732	+	lincRNA
GATA6-AS1	chr18	19746859	19748929	2071	-	lincRNA
FAM13A-AS1	chr4	89630940	89651254	20315	+	antisense
ENO1-AS1	chr1	8938894	8939953	1060	+	antisense
EMX2OS	chr10	119232726	119304579	71854	-	antisense
EGOT	chr3	4790876	4793274	2399	-	lincRNA
DLGAP1-AS1	chr18	3593730	3598350	4621	+	antisense
CYP1B1-AS1	chr2	38302791	38408997	1E+05	+	antisense
CSNK1G2-AS1	chr19	1952530	1954585	2056	-	antisense
CPEB2-AS1	chr4	14911585	15003669	92085	-	lincRNA
COL18A1-AS2	chr21	46827301	46829980	2680	-	antisense
COL18A1-AS1	chr21	46839631	46844985	5355	-	antisense
CDKN2B-AS1	chr9	21994777	22121096	1E+05	+	antisense
CD27-AS1	chr12	6548167	6560733	12567	-	antisense
BAIAP2-AS1	chr17	79002933	79008501	5569	-	lincRNA
ASB16-AS1	chr17	42253341	42264085	10745	-	antisense
ARHGAP26-AS1	chr5	142239169	142248487	9319	-	antisense
AGAP11	chr10	88752163	88769960	17798	+	processed_transcript
ACTN1-AS1	chr14	69446399	69454180	7782	+	antisense
ABO	chr9	136125788	136150617	24830	-	processed_transcript

Protein	OVCA4 29-1	OVCA 429-2	Contr ol-1	Contr ol-2	Contr ol-3	<i>UCA1</i> KO1-1	<i>UCA1</i> KO1-2	<i>UCA1</i> KO1-3	<i>UCA1</i> KO2-1	<i>UCA1</i> КО2-2	Ratio (KO/ WT)	T test
PEA-15-R-V_GBL1116480	1.04	1.03	1.06	1.07	1.09	0.94	0.91	0.97	0.97	0.97	0.90	9.10E-05
UGT1A-M-V_GBL1116415	1.12	1.11	1.05	1.11	1.13	0.93	0.91	0.89	0.97	0.98	0.85	1.66E-04
c-Met_pY1234_Y1235-R- V_GBL1115572	1.23	1.18	1.18	1.23	1.16	0.92	0.85	0.78	0.95	0.89	0.74	1.67E-04
Caveolin-1-R-V_GBL1115521	1.14	1.16	1.18	1.15	1.28	0.78	0.75	0.72	0.85	0.83	0.66	2.07E-04
H2AX_pS139-R-V_GBL1116473	1.02	1.03	1.04	1.02	1.00	0.97	0.98	0.98	0.97	0.98	0.95	9.14E-04
PCNA-M-C_GBL1116373	1.03	1.06	1.07	1.07	1.01	0.92	0.86	0.87	0.90	0.95	0.86	0.01
LDHA-R-C_GBL1115596	1.14	1.18	1.18	1.28	1.42	0.85	0.54	0.64	1.08	0.82	0.63	0.01
Rb_pS807_S811-R- V_GBL1115559	1.25	1.22	1.09	1.20	1.09	0.93	0.86	0.87	1.09	0.93	0.80	0.01
Bcl2A1-R-V_GBL1116330	1.11	1.04	1.11	1.00	1.03	0.98	0.93	0.95	0.95	0.96	0.90	0.01
MSH6-R-C_GBL1115609	1.08	1.07	1.11	1.10	1.14	0.90	0.78	0.82	1.04	0.93	0.81	0.01
Cyclin-B1-R-V_GBL1115524	1.07	1.10	1.05	1.07	1.03	1.01	1.00	0.95	0.88	0.83	0.88	0.01
PYGM-M-C_GBL1116395	1.07	1.09	1.01	1.08	1.10	0.98	0.86	0.86	1.02	0.90	0.86	0.01
Myosin-11-R-V_GBL1115630	1.01	1.18	0.92	1.27	1.28	0.85	0.50	0.54	1.05	0.79	0.66	0.01
CD49b-M-V_GBL1116425	0.97	0.99	0.95	1.12	1.04	0.94	0.84	0.88	1.04	0.96	0.92	0.02
Lck-R-V_GBL1115538	1.46	1.49	1.03	1.43	1.45	0.73	0.67	0.66	1.32	1.02	0.64	0.02
EGFR_pY1173-R-V_GBL1115526	1.10	1.07	1.10	0.98	1.02	0.93	0.96	0.90	0.95	0.93	0.89	0.02
Gys_pS641-R-V_GBL1115603	1.08	1.12	1.05	1.05	1.03	0.95	0.97	0.90	1.03	0.96	0.90	0.02
RBM15-R-V_GBL1115629	1.09	1.03	1.00	1.03	1.05	0.97	0.84	0.81	1.01	0.97	0.89	0.02
PMS2-R-V_GBL1116324	1.17	1.18	1.04	1.07	1.09	0.96	0.87	0.87	1.05	0.95	0.84	0.02
Src_pY416-R-C_GBL1116447	1.00	1.01	1.01	1.03	1.06	0.96	1.00	0.97	0.94	1.01	0.95	0.02
Transglutaminase-M- V_GBL1116383	0.87	0.96	1.11	1.05	1.09	0.51	0.55	0.56	0.95	0.96	0.69	0.02
XPA-M-V_GBL1116419	1.09	1.15	0.99	1.07	1.01	0.92	0.99	0.94	0.96	0.99	0.90	0.02
RPA32-T-C_GBL1116471	1.06	1.03	0.98	1.04	1.10	0.92	0.88	0.85	1.03	1.04	0.91	0.02
Ets-1-R-V_GBL1116361	1.24	1.18	1.36	1.01	1.24	0.95	0.69	0.73	0.99	0.90	0.70	0.03
Axl-R-V_GBL1116474	1.47	1.09	1.14	1.08	1.13	0.83	0.89	0.91	0.88	0.93	0.75	0.03
UQCRC2-M-C_GBL1116412	1.04	1.04	1.01	1.00	1.04	1.02	0.93	0.94	0.96	1.01	0.95	0.03
ERCC5-R-C_GBL1116336	1.00	1.00	1.08	1.07	1.06	0.99	0.96	0.94	0.93	0.98	0.92	0.04
S6_pS240_S244-R-V_GBL1115562	1.19	1.12	0.99	1.12	1.07	0.96	0.79	0.84	1.12	0.94	0.85	0.04
Glutaminase-R-C_GBL1116346	0.98	0.95	1.07	1.05	1.06	0.87	0.81	0.80	1.02	0.98	0.88	0.04
Rad50-M-V_GBL1116385	1.05	1.05	1.08	0.99	1.01	1.01	1.00	0.97	0.94	1.00	0.95	0.04
eEF2-R-C_GBL1115607	1.05	1.08	1.15	1.07	1.23	0.87	0.43	0.46	1.04	0.83	0.65	0.04
ERCC1-M-V_GBL1116435	1.03	1.08	0.96	1.03	1.02	0.93	0.93	0.93	1.00	0.99	0.93	0.04

Table S3. Differentially expressed proteins caused by disruption of UCA1, Related to Figure 4A

Dvl3-R-V_GBL1116454	1.02	1.03	0.91	1.04	1.05	0.99	0.91	0.90	0.99	0.96	0.94	0.04
Myosin-IIa_pS1943-R- V_GBL1115635	1.23	1.43	1.02	1.28	1.50	0.82	0.17	0.26	1.30	0.90	0.54	0.05
FAK-R-C_GBL1115530	1.11	1.08	1.03	1.06	1.09	1.08	0.89	0.96	1.04	0.94	0.91	0.05
Chk1_pS296-R-V_GBL1116335	1.11	1.11	1.03	1.00	0.99	1.04	1.09	1.03	0.95	0.96	0.97	0.05
p53-R-C_GBL1115545	0.88	0.93	0.91	0.89	0.91	1.01	1.03	1.03	0.99	1.04	1.13	1.89E-04
NDRG1_pT346-R-V_GBL1116491	0.84	0.90	0.82	0.84	0.81	1.20	1.22	1.25	1.07	1.18	1.41	4.42E-04
Gab2-R-V_GBL1115594	0.88	0.91	0.82	0.80	0.80	1.06	1.05	1.05	0.93	0.99	1.20	7.12E-04
PDK1_pS241-R-V_GBL1115550	0.93	0.88	0.94	0.87	0.92	1.14	1.08	1.05	1.02	1.04	1.17	1.49E-03
Claudin-7-R-V_GBL1115586	0.85	0.86	0.82	0.99	0.98	1.06	1.32	1.16	1.45	1.32	1.40	1.52E-03
Tuberin_pT1462-R- V_GBL1116448	0.91	0.92	0.80	0.84	0.85	1.01	1.04	1.02	0.96	0.99	1.16	2.48E-03
INPP4b-R-V_GBL1115611	0.90	0.90	0.96	0.96	0.96	1.13	1.03	1.07	1.10	1.09	1.16	2.57E-03
p27_pT198-R-V_GBL1115587	0.97	0.95	1.00	0.94	0.95	1.01	1.03	1.04	0.99	0.98	1.05	2.92E-03
ACVRL1-R-C_GBL1116458	0.98	0.98	0.96	0.95	1.01	1.04	1.02	0.99	0.97	1.04	1.04	3.58E-03
Pdcd4-R-C_GBL1115583	0.85	0.89	0.88	0.93	0.97	1.02	1.07	1.09	1.04	1.04	1.17	4.02E-03
Tuberin-R-V_GBL1115566	0.96	0.95	0.88	0.98	0.97	1.14	1.04	1.02	1.08	1.04	1.12	4.51E-03
Caspase-3-R-C_GBL1115519	0.93	0.91	0.96	0.92	0.91	1.02	1.05	1.04	0.98	0.96	1.09	0.01
Tyro3-R-V_GBL1116457	0.95	0.96	0.93	0.92	0.93	1.04	1.06	1.00	0.99	1.11	1.11	0.01
c-Abl-R-V_GBL1116347	1.00	0.99	1.04	0.99	0.95	1.04	1.02	1.05	1.02	0.99	1.03	0.01
ER-R-V_GBL1116352	0.91	0.89	0.95	0.91	0.91	1.10	1.04	1.06	0.97	1.11	1.15	0.01
HER2-M-V_GBL1116406	0.92	0.92	0.93	0.91	0.95	1.04	1.06	1.03	0.94	1.06	1.11	0.01
A-Raf-R-V_GBL1116314	0.98	0.95	0.92	0.92	0.89	1.02	1.04	1.08	1.01	0.97	1.10	0.01
MMP2-R-V_GBL1115540	0.91	0.93	0.95	0.92	0.91	1.06	1.03	1.03	0.95	1.00	1.10	0.01
D-alpha-Tubulin-R- V_GBL1116339	1.01	0.98	0.94	0.91	0.93	1.14	1.05	1.04	0.96	0.97	1.08	0.01
CXCR4-R-C_GBL1116362	0.94	0.95	0.96	0.94	0.90	1.02	1.03	1.03	1.01	1.09	1.11	0.01
Cyclophilin-F-M-V_GBL1116413	0.92	0.97	0.91	0.93	0.90	1.03	1.02	1.04	0.96	1.01	1.09	0.01
P-Cadherin-R-C_GBL1116446	0.97	1.00	0.91	0.95	0.92	1.05	1.04	0.94	1.03	1.05	1.08	0.02
PDGFR-beta-R-V_GBL1116319	1.00	0.94	0.96	0.94	0.86	1.01	1.06	1.06	1.00	0.95	1.08	0.02
MEK2-R-V_GBL1116323	1.03	1.00	0.98	0.95	0.95	1.13	1.05	1.01	1.07	1.10	1.09	0.02
TFAM-R-V_GBL1116333	0.88	0.95	0.89	0.89	0.91	0.98	1.13	1.10	0.95	0.97	1.13	0.02
IGFBP5-G-C_GBL1116469	0.93	0.91	0.93	0.94	0.91	0.98	1.06	1.01	0.97	0.99	1.08	0.02
Bad_pS112-R-V_GBL1115511	1.00	0.88	1.01	0.89	0.94	1.14	1.11	1.06	1.06	0.99	1.14	0.02
Notch3-R-C_GBL1116354	0.89	0.88	0.99	0.94	0.97	1.02	1.06	1.04	1.00	1.03	1.10	0.02
mTOR_pS2448-R-C_GBL1115542	0.99	0.99	0.91	1.04	0.98	1.08	1.01	1.01	1.06	1.06	1.06	0.02
eIF4E-R-V_GBL1115571	0.88	0.92	1.01	0.95	0.98	0.98	0.98	1.05	0.99	1.00	1.05	0.02
MDM2_pS166-R-V_GBL1116306	0.96	0.94	1.00	1.01	1.01	1.09	1.02	1.01	1.08	1.07	1.07	0.02

Notch1-R-V_GBL1115610 0.99 1.02 0.91 0.89 0.91 1.08 1.03 1.03 1.03 1.11 0.03 ATM-R-V_GBL1116337 0.91 0.89 0.79 0.92 0.87 1.05 1.12 1.11 0.94 1.00 1.19 0.03 Rictor_pT1135-R-V_GBL1115627 0.93 0.92 0.90 0.99 0.91 1.04 1.03 1.03 0.98 0.98 1.09 0.03 Rictor_PT1135-R-V_GBL1115626 0.95 0.94 0.92 0.97 0.94 1.03 1.03 1.04 0.97 0.98 1.07 0.04 Src-M-V_GBL1116378 1.01 0.93 0.93 0.88 0.81 1.04 1.13 0.99 1.02 1.11 1.16 0.04 YAP_pS127-R-E_GBL1115578 0.83 0.89 0.99 0.96 1.00 1.41 1.12 1.11 1.30 1.01 1.18 0.04 Puma-R-C_GBL1116329 0.89 0.92 0.91 <td< th=""><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th><th></th></td<>													
ATM-R-V_GBL11163370.910.890.790.920.871.051.121.110.941.001.190.03Rictor_pT1135-R-V_GBL11156270.930.920.900.990.911.041.031.030.980.980.981.090.03Rictor-R-C_GBL11156260.950.940.920.970.941.031.031.040.970.981.070.04Src-M-V_GBL11163781.010.930.930.880.811.041.130.991.021.111.160.04YAP_pS127-R-E_GBL11155780.830.890.990.961.001.411.121.111.371.081.300.04Puma-R-C_GBL11163290.890.920.910.910.901.011.021.041.301.011.180.04Pdcd-1L1-G-C_GBL11164260.910.960.810.800.970.961.041.021.000.991.130.05CD29-M-V_GBL11164260.890.930.960.950.950.951.061.020.941.021.021.070.05	Notch1-R-V_GBL1115610	0.99	1.02	0.91	0.89	0.91	1.08	1.03	1.03	1.08	1.03	1.11	0.03
Rictor_pT1135-R-V_GBL11156270.930.920.900.990.911.041.031.030.980.981.090.03Rictor-R-C_GBL11156260.950.940.920.970.941.031.031.040.970.981.070.04Src-M-V_GBL11163781.010.930.930.880.811.041.130.991.021.111.160.04YAP_pS127-R-E_GBL11155780.830.890.990.961.001.411.121.111.371.081.300.04Puma-R-C_GBL11163290.890.920.910.910.901.011.021.041.301.011.180.04Pdcd-1L1-G-C_GBL11164680.910.960.810.800.970.961.041.021.000.991.130.05CD29-M-V_GBL11164260.890.930.960.950.950.951.061.020.941.021.070.05	ATM-R-V_GBL1116337	0.91	0.89	0.79	0.92	0.87	1.05	1.12	1.11	0.94	1.00	1.19	0.03
Rictor-R-C_GBL11156260.950.940.920.970.941.031.031.040.970.981.070.04Src-M-V_GBL11163781.010.930.930.880.811.041.130.991.021.111.160.04YAP_pS127-R-E_GBL11155780.830.890.990.961.001.411.121.111.371.081.300.04Puma-R-C_GBL11163290.890.920.910.910.901.011.021.041.301.011.180.04Pdcd-1L1-G-C_GBL11164680.910.960.810.800.970.961.041.021.000.991.130.05CD29-M-V_GBL11164260.890.930.960.950.951.061.020.941.021.070.05	Rictor_pT1135-R-V_GBL1115627	0.93	0.92	0.90	0.99	0.91	1.04	1.03	1.03	0.98	0.98	1.09	0.03
Src-M-V_GBL1116378 1.01 0.93 0.93 0.88 0.81 1.04 1.13 0.99 1.02 1.11 1.16 0.04 YAP_pS127-R-E_GBL1115578 0.83 0.89 0.99 0.96 1.00 1.41 1.12 1.11 1.37 1.08 1.30 0.04 Puma-R-C_GBL1116329 0.89 0.92 0.91 0.91 0.90 1.01 1.02 1.04 1.30 1.01 1.18 0.04 Pdcd-1L1-G-C_GBL1116468 0.91 0.96 0.81 0.80 0.97 0.96 1.04 1.00 0.99 1.13 0.05 CD29-M-V_GBL1116426 0.89 0.93 0.96 0.95 0.95 1.06 1.02 0.94 1.02 1.07 0.05	Rictor-R-C_GBL1115626	0.95	0.94	0.92	0.97	0.94	1.03	1.03	1.04	0.97	0.98	1.07	0.04
YAP_pS127-R-E_GBL1115578 0.83 0.89 0.99 0.96 1.00 1.41 1.12 1.11 1.37 1.08 1.30 0.04 Puma-R-C_GBL1116329 0.89 0.92 0.91 0.91 0.90 1.01 1.02 1.04 1.30 1.01 1.18 0.04 Pdcd-1L1-G-C_GBL1116468 0.91 0.96 0.81 0.80 0.97 0.96 1.04 1.00 0.99 1.13 0.05 CD29-M-V_GBL1116426 0.89 0.93 0.96 0.95 0.95 1.06 1.02 0.94 1.02 1.07 0.05	Src-M-V_GBL1116378	1.01	0.93	0.93	0.88	0.81	1.04	1.13	0.99	1.02	1.11	1.16	0.04
Puma-R-C_GBL1116329 0.89 0.92 0.91 0.91 0.90 1.01 1.02 1.04 1.30 1.01 1.18 0.04 Pdcd-1L1-G-C_GBL1116468 0.91 0.96 0.81 0.80 0.97 0.96 1.04 1.00 0.99 1.13 0.05 CD29-M-V_GBL1116426 0.89 0.93 0.96 0.95 0.95 1.06 1.02 0.94 1.02 1.07 0.05	YAP_pS127-R-E_GBL1115578	0.83	0.89	0.99	0.96	1.00	1.41	1.12	1.11	1.37	1.08	1.30	0.04
Pdcd-1L1-G-C_GBL1116468 0.91 0.96 0.81 0.80 0.97 0.96 1.04 1.02 1.00 0.99 1.13 0.05 CD29-M-V_GBL1116426 0.89 0.93 0.96 0.95 0.95 1.06 1.02 0.94 1.02 1.07 0.05	Puma-R-C_GBL1116329	0.89	0.92	0.91	0.91	0.90	1.01	1.02	1.04	1.30	1.01	1.18	0.04
CD29-M-V_GBL1116426 0.89 0.93 0.96 0.95 0.95 0.95 1.06 1.02 0.94 1.02 1.07 0.05	Pdcd-1L1-G-C_GBL1116468	0.91	0.96	0.81	0.80	0.97	0.96	1.04	1.02	1.00	0.99	1.13	0.05
	CD29-M-V_GBL1116426	0.89	0.93	0.96	0.95	0.95	0.95	1.06	1.02	0.94	1.02	1.07	0.05

The differentially expressed proteins with $P \leq 0.05$ are shown.

Target Sequence		Lough Otort	art End		Gene		
Access #	Probe Sequence	Length	Start	End	Gene ID	Name	Note
NR_015379	GGACTCATTGTCCAGAAGAA	20	6	25	652995	UCA1	Odd
NR_015379	GTTATCTGTTGTCAGCAGAG	20	61	80	652995	UCA1	Odd
NR_015379	GAGATAGGAGAGGGGCAGCT	20	124	143	652995	UCA1	Odd
NR_015379	AAGCATGTCCGTATAGAAGA	20	231	250	652995	UCA1	Odd
NR_015379	ACGGAATGAGGGCCAGGACA	20	305	324	652995	UCA1	Odd
NR_015379	GAGGAGATCCGATTTGAAAT	20	381	400	652995	UCA1	Odd
NR_015379	TGGTCCAAGGGGCTTCTGAG	20	436	455	652995	UCA1	Odd
NR_015379	CAGCTAGGGTGTCTTCATAT	20	505	524	652995	UCA1	Odd
NR_015379	AATGACCGGGTAGGGTCTGG	20	579	598	652995	UCA1	Odd
NR_015379	TGGGCATGGCTTTATTCTGG	20	654	673	652995	UCA1	Odd
NR_015379	GTTTGTCTGATCGGCTCTCG	20	727	746	652995	UCA1	Odd
NR_015379	GCATGCTGGGATGGCCATTT	20	813	832	652995	UCA1	Odd
NR_015379	TTTTATAGGCGGCAGGTCTT	20	875	894	652995	UCA1	Odd
NR_015379	GACTTTTGACCCAGAGATTT	20	949	968	652995	UCA1	Odd
NR_015379	GTTGTCCATTTCATGAGAGT	20	1023	1042	652995	UCA1	Odd
NR_015379	CCCTCTAACAACAACAACA	20	1111	1130	652995	UCA1	Odd
NR_015379	CTTTGTCTCCTGGATTAAGC	20	1161	1180	652995	UCA1	Odd
NR_015379	TCCCATAGGTGTGAGTGGCG	20	1249	1268	652995	UCA1	Odd
NR_015379	AGCCTTTTGTCCCCATTTTC	20	1320	1339	652995	UCA1	Odd
NR_015379	CAGTAATCAGGCATATTAGC	20	1399	1418	652995	UCA1	Odd
NR_015379	GCTAACAAGGTGCCAGTTAG	20	1467	1486	652995	UCA1	Odd
NR_015379	GACTGCGTGGACACCTTAAA	20	1541	1560	652995	UCA1	Odd
NR_015379	AGTTCCTTCTGGGGATTACT	20	1606	1625	652995	UCA1	Odd
NR_015379	GACGGATATGTGCAGGTCAC	20	1724	1743	652995	UCA1	Odd
NR_015379	CTGTTTCACTTCTTTGTGGT	20	1786	1805	652995	UCA1	Odd
NR_015379	CATAATGGTGGAATGTCGTA	20	1837	1856	652995	UCA1	Odd
NR_015379	GATGGGACTCATTGTCCAGG	20	1925	1944	652995	UCA1	Odd
NR_015379	TAGGCGTGGAAAGTTACAGT	20	2000	2019	652995	UCA1	Odd
NR_015379	CTGTTAATTCACTTGGGTGC	20	2089	2108	652995	UCA1	Odd
NR_015379	AGGTTCCTGTCACGCGTGTC	20	2151	2170	652995	UCA1	Odd
NR_015379	AGGTGCATGGTGGAGAGATG	20	29	48	652995	UCA1	Even
NR_015379	ATAGGGCTGGGGTAGGCTGT	20	102	121	652995	UCA1	Even
NR_015379	TGTTAATTCACTTGGGTGCA	20	178	197	652995	UCA1	Even

Table S4. iRAP antisense DNA probe sequences, Related to Figure 5

NR_015379	AGATTTTGGCACCAAGTGTC	20	251	270	652995	UCA1	Even
NR_015379	GGTCGCAGGTGGATCTCTTC	20	325	344	652995	UCA1	Even
NR_015379	CAGTCTTCAGCCACTAAGCC	20	401	420	652995	UCA1	Even
NR_015379	CCGTAAGAGTTACCCGAAGC	20	469	488	652995	UCA1	Even
NR_015379	GTTTGAGGGGTCAGACTTTT	20	542	561	652995	UCA1	Even
NR_015379	AGAGGGTCCTGCAGATGGAC	20	617	636	652995	UCA1	Even
NR_015379	GAAGAGGAGAGATCAAGCTG	20	683	702	652995	UCA1	Even
NR_015379	TGGAAGCATGGCCTTGGCTG	20	778	797	652995	UCA1	Even
NR_015379	AGGAACGGATGAAGCCTGCT	20	838	857	652995	UCA1	Even
NR_015379	GAGGATAGGGTCTCAAGATA	20	902	921	652995	UCA1	Even
NR_015379	ACCATGGGCAAGGATCTGAT	20	991	1010	652995	UCA1	Even
NR_015379	TGCCAAATATGTGGAACTGG	20	1060	1079	652995	UCA1	Even
NR_015379	GGCAAAGAGTGAAATGTCCC	20	1134	1153	652995	UCA1	Even
NR_015379	ATAGTTCTGATTGCAGATCC	20	1210	1229	652995	UCA1	Even
NR_015379	CCCTGTTGCTAAGCCGATGA	20	1282	1301	652995	UCA1	Even
NR_015379	GTAGGTGGCGATGAGTTTTT	20	1373	1392	652995	UCA1	Even
NR_015379	CTTGGAACTGCCCTAATAAA	20	1435	1454	652995	UCA1	Even
NR_015379	TCTAGTAAGTTTCGGGTCTA	20	1504	1523	652995	UCA1	Even
NR_015379	GCTGATATTTCTCCGGACTG	20	1578	1597	652995	UCA1	Even
NR_015379	CTTCAGCTCACAGGCCTGAC	20	1685	1704	652995	UCA1	Even
NR_015379	CTGCTCCAGACTTCTTGGCT	20	1759	1778	652995	UCA1	Even
NR_015379	GTGGGTTAATTAGTTAAGGC	20	1815	1834	652995	UCA1	Even
NR_015379	GATTGATCAGTTGGGGCAGG	20	1886	1905	652995	UCA1	Even
NR_015379	GTCACAAGGTGCATGGTGGA	20	1950	1969	652995	UCA1	Even
NR_015379	GGGCAGCTTTATAGGGCTTG	20	2022	2041	652995	UCA1	Even
NR_015379	CCAATCAGGCTTTGTGTGAG	20	2117	2136	652995	UCA1	Even
NR_015379	AGGAAAGGAAAACCACATGA	20	2244	2263	652995	UCA1	Even
NR_004430.2	CTCCCCTGCCAGGTAAGTAT	20	1	20	26871	U1	

All the antisense DNA probes are labeled with biotin at the 3' end.

Table S5. iRAP peptide sequences, Related to Figure 5

Sequence	Proteins	Start	End	U1	UCA1 -Odd1	UCA1- Even1	UCA1 -Odd2	UCA1- Even2
GGGGGQDNGLEGLGNDSR	sp P08621 RU17_HUMAN	394	411	5	2			
ASMQQQQQLASAR	sp Q9Y3Y2 CHTOP_HUMAN	39	51	4	1			
AVQGGGATPVVGAVQGPVPGMPPMTQAP FFFLAALVIDNGSGMCK	sp/P09012/SNRPA_HUMAN	124	152	4	1		1	
MMLGTEGGEGFVVK	sp P31943 HNRH1_HUMAN	1	14	3			1	
NMGGPYGGGNYGPGGSGGSGGYGGR	sp P22626 ROA2 HUMAN	326	350	3	1			
QEMQEVQSSR	sp P22626 ROA2_HUMAN	191	200	3				
QQEVETELK	sp P08621 RU17_HUMAN	79	87	3	2			
RGGADVNIR	sp P08621 RU17_HUMAN	201	209	3	1			
STVHEILCK	$sp P08021 K017_n0MAN$ $sp P07355 ANXA2_HUMAN\cdot sp A$	2	87 10	3	1		1	
ESTGAQVQVAGDMLPNSTER	sp Q15365 PCBP1_HUMAN;sp Q1	125	144	2	1		1	
SEMTPEELQK	sp P62316 SMD2_HUMAN	9	18	2				
TDYNASVSVPDSSGPER	sp P61978 HNRPK_HUMAN	70	86	2				
DSYVGDEAQSKR	sp P60/09 ACTB_HUMAN;sp P63	51	62	2				
DSYESYGNSR	sp P38159 RBMX_HUMAN;sp Q9	283	292	2				
DGMDNQGGYGSVGR	sp P31942 HNRH3 HUMAN	288	301	2				
NQGGYGGSSSSSSYGSGR	sp P09651 ROA1_HUMAN	353	370	2				1
MWDPHNDPNAQGDAFK	sp P08621 RU17_HUMAN	88	103	2				
VNYDTTESKLR MLACOVL DINL A AEDV	sp P08621 RU17_HUMAN	110	120	2				
NIAGQ V LDINLAAEPK SI KI OA SNVTNK NDPK	sp P0/910 HINKPC_HUMAN	2	89 17	1				
HRSCNTDDCPPGSODFREVOCSEFDSIPFR	sp O9H324 ATS10 HUMAN	593	622	1				
MSATSVDTQR	sp Q9BYJ9 YTHD1_HUMAN	1	10	1	1	1		
VADLTEQYNEQYGAVR	sp Q9BWF3 RBM4_HUMAN	183	198	1				
EVYQQQQYGSGGR	sp Q99729 ROAA_HUMAN	233	245	1				
LIEE A DA DOIN	sp Q96RQ3 MCCA_HUMAN	624 285	035	1				
YLSSVSSOETOGGPLAPMTGTIEK	splQ96RQ3/MCCA_HUMAN	285 636	659	1	1			
TMFENVTR	sp Q96QA5 GSDMA_HUMAN	2	9	1	-			1
AGGGPTLQCPPPSSPEK	sp Q96N66 MBOA7_HUMAN	272	288	1				
KIQNDAGVR	sp Q92945 FUBP2_HUMAN;sp Q9	347	355	1				
DAFADAVQK	sp Q92945 FUBP2_HUMAN	321	80	1				
IGOOPOOPGAPPOODYTK	sp Q92945 FUBP2_HUMAN	629	646	1				
SDYSTGGPPPGPPPAGGGGGGGGGGGGGGGG	sp Q92945 FUBP2_HUMAN	2	40	1				
SVSLTGAPESVQK	sp Q92945 FUBP2_HUMAN	191	203	1				
VQISPDSGGLPER	sp Q92945 FUBP2_HUMAN	178	190	1				
APILIATUVASK	sp Q92841 DDX1/_HUMAN;sp P1	469	480	1				
SSOSSSOOFSGIGR	sp Q92841 DDX17_HUMAN	671	684	1				
VLEEANQAINPK	sp Q92841 DDX17_HUMAN	536	547	1				
EQMEKVEADLTR	sp Q8N8E3 CE112_HUMAN	623	634	1				
QDNTPKR DLACDSEETWITCDNIK	sp Q8N118 CP4X1_HUMAN	275	281	1				1
DLOFONKK	sp Q80VB/ C103A_HUMAN	733	928 740	1	1	1	1	1
VAPDEHPILLTEAPLNPK	sp Q51B30 CE102_HCMAN	97	114	1	1	1	1	1
GGSYSQAASSDSAQGSDVSLTA	sp Q31612 1B73_HUMAN	342	363	1				
VLVDQTTGLSR	sp Q15717 ELAV1_HUMAN	137	147	1				
QKNKSGETVVLK	sp Q15652 JHD2C_HUMAN	2249	2260	1				
MOSNKTENI EK	splQ15505 PCBP1_HUMAN;splQ1	4/	11	1				
SSSVGSSSSYPISPAVSR	sp Q15149 PLEC_HUMAN	4384	4401	1				
AAAAVQGGR	sp Q15005 SPCS2_HUMAN	2	10	1				
IFVGGLSPDTPEEK	sp Q14103 HNRPD_HUMAN	184	197	1				
SSGGSYRDSYDSYATHNE	sp Q14011 CIRBP_HUMAN	155	172	1				
VI VNDAOK	sp Q13263 TIF1B_HUMAN	312	319	1				
AATASAGAGGIDGKPR	sp Q02978 M2OM HUMAN	2	17	1				
GDATVSYEDPPTAK	sp Q01844 EWS_HUMAN	411	424	1				
ATVQQLEGR	sp Q01469 FABP5_HUMAN	2	10	1	1			
SSGPTSLFAVTVAPPGAR	sp Q00839 HNRPU_HUMAN	187	204	1				
MIOVYFPK	splQ00525/MPCP_HUMAN	190	199	1				
AADPPAENSSAPEAEQGGAE	sp P67809 YBOX1 HUMAN	305	324	1				
GAEAANVTGPGGVPVQGSK	sp P67809 YBOX1_HUMAN	119	137	1				
GENLVSMTVEGPPPKDTGIAR	sp P63162 RSMN_HUMAN;sp P14	74	94	1				
TITLEVEPSDTIENVK	sp P62987 RL40_HUMAN;sp P629	12	27	1				
ΑΑŲΘΕΡΥΥΥΓΚ ΕСΤΙΕΑΤΑ	$sp r02020 RAN_HUMAN$ $sp P62633 CNRP_HUMAN$	170^{2}	12	1				
SSNECFKCGR	sp P62633 CNBP_HUMAN	2	11	1				
VAQLEQVYIR	sp P62318 SMD3_HUMAN	55	64	1				
GDNITLLQSVSN	sp P62304 RUXE_HUMAN	81	92	1				
ADIQTER EITALADSTN42	sp P62280 RS11_HUMAN	2	8	1			1	
AGFAGDDAPR	sp P60709 ACTB_HUMAN;sp P63	19	28	1	1		1	1

DDDIAALVVDNGSGMCK	sp P60709 ACTB HUMAN	2	18	1				
HTGPNSPDTANDGFVR	sp P55795 HNRH2 HUMAN;sp P3	99	114	1				
MMLSTEGR	sp P55795 HNRH2_HUMAN	1	8	1				
MLGPEGGEGEVVK	sp P52597 HNRPF_HUMAN	2	14	1				
MGPAMGPALGAGIER	sp P52272 HNRPM_HUMAN	592	606	1				
OGGGGGGGGSVPGIER	sp P52272 HNRPM_HUMAN	380	403	1				
SCEDVCCCVCSCCCCCCC	ap D51001 DOA2_HUMAN	255	276	1				
ALVDCDCTOVD	spir 51991 KOA5_HUMAN	255	370	1				
ALVDGPCTQVR	sp P50914 KL14_HUMAN	30	40	1		1		
ATTATMATSGSAR	sp P38919 IF4A3_HUMAN	2	14	1		1		
DDGYSTKDSYSSRDYPSSR	sp P38159 RBMX_HUMAN;sp Q9	211	229	1				
DVYLSPR	sp P38159 RBMX_HUMAN;sp Q9	204	210	1				
YDDYSSSR	sp P38159 RBMX_HUMAN;sp Q9	310	317	1				
RGPPPPR	sp P38159 RBMX HUMAN	94	101	1				
SDLYSSGR	sp P38159 RBMX HUMAN	332	339	1				
GEATVSFDDPPSAK	splP35637 FUS_HUMAN:splO928	335	348	1				
GLPWSCSADEVOR	sp P31943 HNRH1_HUMAN	17	29	1				
MIGTEGGEGEVVK	sp P31943 HNRH1_HUMAN	2	14	1				
MIGTEGGEGEVVKVR	sp[P31943]HNRH1_HUMAN	2	16	1				
STGEAEVOEASK	ap D21042 HND H2 HIMAN	56	67	1				
DIGEALVQLASK	spjr 51942/minKii5_hUMAN	50	101	1				
DISSSIVVSIQK	sp P31483 11A1_HUMAN	90	101	1				
FGQGGAGPVGGQGPR	sp P23246 SFPQ_HUMAN	667	681	1				
MEKTLETVPLER	sp P22626 ROA2_HUMAN	1	12	1				
TLETVPLER	sp P22626 ROA2_HUMAN	4	12	1				
SISLYYTGEK	sp P19338 NUCL_HUMAN	458	467	1				
AGEAPTENPAPPTQQSSAE	sp P16989 YBOX3_HUMAN	354	372	1				
SEAGEATTTTTTTLPQAPTEAAAAAPQDPA	sp P16989 YBOX3 HUMAN	2	33	1				
DNEETGFGSGTR	sp P15144 AMPN HUMAN	924	935	1				
AEAEAOAEELSFPR	sp P11498 PYC HUMAN	929	942	1				
ALAVSDLNR	sp P11498 PYC_HUMAN	1062	1070	1				
SK SESPK EPEOL R	splP09651/ROA1_HUMAN	2	14	1				
DCGAWGTEOP	app002820 EC1 HUMAN	65	74	1				1
EVERATIALD	approvide the second se	61	74	1				1
EVSSAINALK	spip09012 SNRPA_HUMAN	01	/0	1				
	sp P09012 SNRPA_HUMAN	84	96	1		1		
KAVQGGGATPVVGAVQGPVPGMPPMTQA	sp P09012 SNRPA_HUMAN	123	152	1		1	1	
TDSDIIAK	sp P09012 SNRPA_HUMAN	89	96	1				
AETREERMER	sp P08621 RU17_HUMAN	60	69	1				
DPIPYLPPLEK	sp P08621 RU17_HUMAN	17	27	1				
GGADVNIR	sp P08621 RU17_HUMAN	202	209	1				
GSERGRDEAR	sp P08621 RU17_HUMAN	384	393	1				
IHMVYSK	sp P08621 RU17 HUMAN	132	138	1				
KEELRGGGGDMAEPSEAGDAPPDDGPPGE	sp P08621 RU17 ⁻ HUMAN	306	346	1	1			
VLVDVER	spP08621 RU17_HUMAN	174	180	1	1			
AAVAGEDGR	sp P07910 HNRPC_HUMAN:sp P0	65	73	1				
ASNVTNKTDPR	sp P07910 HNRPC_HUMAN	2	12	1				
VPPPPPIAR	sp[P07910]HNRPC_HUMAN	143	151	1				
AMCEOAVALAD	cplD05165DCCA_HUMAN	215	225	1				
	spir05105 rCCA_HUMAN	313	525	1				
MADALDN I VIK	sp P05105 PCCA_HUMAN	400	4/0	1				
MADEAVCVGPAPISK	sp P05165 PCCA_HUMAN	105	119	1				
V VEEAPSIFLDAEIR	sp P05165 PCCA_HUMAN	299	313	1				
SGAQASSTPLSPTR	sp P02545 LMNA_HUMAN	12	25	1				
MEDSASASLSSAAATGTSTSTPAAPTAR	sp O76021 RL1D1_HUMAN	1	28	1				
TGYTLDVTTGQR	sp O60506 HNRPQ_HUMAN;sp O	131	142	1				
MEKENQKLEASR	sp O60343 TBCD4_HUMAN	856	867	1				
PAASITSKPATLTTTSATSK	sp O43670 ZN207_HUMAN	343	362	1				
NDNQETAAMKPENLKK	sp A2A2Z9 AN18B_HUMAN	266	281	1				
QMEEELLFLK	tr Q8WYG7 Q8WYG7 HUMAN	12	21					1
STILQQQFNR	sp Q9Y4G6 TLN2 HUMAN	432	441		1	1		
OTDAOSASSPKK	sp 09Y3R0 GRIP1 HUMAN	754	765			1		
GVLOOGAGALGSSAOGVK	sp 09H8X9 ZDH11 HUMAN	297	314		1			
GGGEOETOELASK	spl0960R8PURB_HUMAN	25	37		1			
SSGOMAOKESESK	spl096089KI20B_HUMAN	105	117		-		1	
MKOKOFVMEOSP	splQ90Q0911120B_11011111	1	12				1	1
DGAILCOPVITNIGSI SI GVCGPOGP	spl006BF3/TMIG2_HUMAN	62	86					1
STEDKMETMD	splQ2001511WIG2_HUMAN	107	206				1	1
SI EFKIMETWIK SLOVECEWDEAESEWAEV	spiQow IQ4jCF070_HUMAN	197	200		1	1	1	
DOAEOWLD	spiQonynojen TA_numan	257	264		1	1		
DQAEQWLK	SPIQ8N4X5 AFIL2_HUMAN	237	204		1	1		
FYEMYLLINK	sp Q8IZJ1 UNC5B_HUMAN	5//	586		1			
NSLSSIMKNKDK	sp Q81Y51 11GD4_HUMAN	51	62				1	
KNDQALQLTQMDKMHK	splQ86Z20/CC125_HUMAN	337	352				1	
GSGSGQSPSYGR	sp Q86YZ3 HORN_HUMAN;CON	897	908					1
HGSSSGSSSR	sp Q86YZ3 HORN_HUMAN;CON	2149	2158				1	1
QGSSAGSSSSYGQHGSGSR	sp Q86YZ3 HORN_HUMAN;CON	507	525				1	
YGQQGSGSGQSPSR	sp Q86YZ3 HORN_HUMAN;CON	649	662					1
FEVNAKFLGVDMER	sp Q86VI3 IQGA3 HUMAN	1575	1588		1	1		
DVDAAYVSK	sp Q7Z794 K2C1B HUMAN:CON	274	282				1	1
VTVQTDDSNK	sp Q6ZP68 ATPUN HUMAN	105	114					1
SGTNNHVTVAIEN	sp O5W0Z9 ZDH20_HUMAN	353	365				1	1
OESTSKADLNCSKNK	sp O5W0B1 RN219 HUMAN	328	342			1	-	-
IOSSOPMSLK	spl05U4N7lGDF50_HUMAN	2	11			1	1	
IFISSPCCPR	sp O5T749 KPRP_HIMAN	327	336			1	1	1
COEFFWIR	splQ5FWF4/ZRAR3 HUMAN	940	947			1	1	1
	JULVII TELLADJ HUMAN	740	271				1	1

FSNSSSSNEFSK	sp Q5D862 FILA2_HUMAN;CON	404	415				1	
AGSLQLSSMSAGNSSLR	sp Q53SF7 COBL1_HUMAN	380	396		1	1		
IQKEEEEILMANKR	sp Q4VCS5 AMOT_HUMAN	727	740	1	1		1	
DMLMQER	sp Q3ZCV2 CA177_HUMAN	391	397		1	1	1	
MEPIYPFARPQMNTR	sp Q15468 STIL_HUMAN	1	15	1	1			
ERDAALK	sp Q14980 NUMA1_HUMAN	598	604				1	
NEWRMITAMNTIR	sp Q14145 KEAP1_HUMAN	495	507	1				
KPEYDLEEDDQEVLK	sp Q13416 ORC2_HUMAN	52	66	1				
MQLDNPSK	sp Q13085 ACACA_HUMAN	818	825	1				
NVLLNNSEK	sp Q08J23 NSUN2_HUMAN	569	577	1	1			
ENAGEDPGLAR	sp P81605 DCD_HUMAN	43	53	1	1		1	
DSYVGDEAQSK	sp P60709 ACTB_HUMAN;sp P63	51	61			1	1	
MNALDLNMKTK	sp P30519 HMOX2_HUMAN	206	216		1			
GGGGNFGPGPGSNFR	sp P22626 ROA2_HUMAN	214	228	1	1			
NLKNSQMCQK	sp P21439 MDR3_HUMAN	670	679			1		
NMLSQVNYRVPNMR	sp P19174 PLCG1_HUMAN	179	192	1	1			
RQQLNEMLK	sp P11532 DMD_HUMAN	2572	2580			1		
KNSYMNPEKK	sp P08473 NEP_HUMAN	737	746		1			
AQGYSGLSVK	sp P07996 TSP1_HUMAN;CON	1055	1064			1		
GPDPSSPAFR	sp P07996 TSP1_HUMAN;CON	51	60			1		
GTSQNDPNWVVR	sp P07996 TSP1_HUMAN;CON_	969	980			1		
SSPVVIDASTAIDAPSNLR	sp P02751 FINC_HUMAN	1892	1910			1	1	
TSLDEALQWR	sp O15539 RGS5_HUMAN	53	62	1	1			
VAASIGNAQK	sp O00151 PDLI1 HUMAN	247	256	1				

The numbers under each probe are the peptide hits obtained by mass spectrometry for iRAP protein samples.

ANTIBODY	SOURCE	IDENTIFIER	NOTE
β-tubulin	Sigma Aldrich	Cat#T8328, RRID: AB_1844090	1:2,000
U1-70k	EMD Millipore	Cat#05-1588, RRID: AB_10805959	1:1,000
AMOT	Bethyl laboratories	Cat#A303-305A, RRID: AB_10951678	1:500
AXL	R&D Systems	Cat#AF-154, RRID: AB_354852	1:1,000
CYR61	Cell Signaling Technology	Cat#14479S, RRID: N/A	1:1,000
CTGF	Abcam	Cat# ab6992, RRID: AB_305688	1:1,000
YAP	Santa Cruz	Cat#sc-15407, RRID: AB_2273277	1:100-1,000, WB
YAP	Cell Signaling Technology	Cat#4912S, RRID: AB_2218911	IP
YAP (phosphoS127)	Abcam	Cat#ab76252, RRID: AB_1524578	1:1,000
Phospho-LATS1	Cell Signaling Technology	Cat#8654S, RRID: AB_10971635	1:100
LATS1	Cell Signaling Technology	Cat#3477T, RRID: AB_2133513	1:500
GFP	Santa Cruz	Cat#sc-9996, RRID: AB_627695	1:2,000
GAPDH	Fitzgerald Antibody	Cat#10R-G109A, RRID: AB_1285808	1:2,000
β-actin	Santa Cruz	Cat#sc-47778, RRID: AB_626632	1:2,000
Rabbit IgG	Thermo Fisher Scientific	Cat#10500C, RRID: AB_2532981	IP

Table S6. Antibody information used in this study, Related to Figure 4, 5, & 6

Transparent Methods

Cell Culture. FTSEC33 and FTSEC246 cells were grown in DMEM/F12 media supplied with 10% FBS (HyClone, catalogue number: SH3007103) and L-glutamine. OSEC4 and OSEC11 were cultured in NOSE-CM media consisting of MCDB105: Medium 199 (1:1), 15% FBS, 10 ng/mL epidermal growth factor (Sigma Aldrich, catalogue number: E9644-2MG), 0.5 mg/mL hydrocortisone (Sigma Aldrich, catalogue number: H0888), 5 mg/mL insulin (Sigma, catalogue number: I1882-100MG), and 34 mg protein/mL bovine pituitary extract (Thermo Fisher Scientific, catalogue number: 13028014). CaOV3 cells were grown in RPMI1640 supplied with 10% FBS and L-glutamine. OVCA429 cells were cultured in EMEM supplied with 10% FBS, L-glutamine, sodium pyruvate (Lonza, catalogue number: 13-115E), and nonessential amino acids (Lonza, catalogue number: 13-114E). UWB1.289 cells were grown in mammary epithelial growth medium (MEGM): RPMI1640 (1:1) supplied with 3% FBS. HEK293T cells were cultured in DMEM supplied with 10% FBS.

H3K27ac ChIP-Seq and Super-enhancer Identification. H3K27ac ChIP-Seq and data analysis for primary ovarian cancer specimens were performed as described previously (Lawrenson et al., 2018). H3K27ac ChIP-Seq data for FTSEC33, FTSEC246, OSEC4, and OSEC11 are publicly available (Coetzee et al., 2015). The AQUAS pipeline (https://github.com/kundajelab/chipseq_pipeline) was used to processed ChIP-Seq data. Reads were aligned to the reference human genome (hg19), filtered by read quality and duplicate reads removed. MACS2 (https://pypi.python.org/pypi/MACS2) was used for peak calling. For the cell lines, two technical replicates were generated and the final peaks were obtained using a naive overlap approach, where the peaks are included if they overlap more than 50% between the two technical replicates. After alignment, homer (http://homer.ucsd.edu/homer/) was used to identify super-enhancers, using a super slope parameter of 2 and a minimum distance of ten thousand base pairs. For defining a set of high-grade serous ovarian cancer super-enhancers, we selected super-enhancers that were called in at least two high-grade serous ovarian cancer samples. For the FTSEC and OSEC sets of super-enhancers, super-enhancers were called individually in each technical replicate, then all the super-enhancers that overlapped both technical replicates within the same cell

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line were selected to get the union set. To identify super-enhancer-associated lncRNAs, we directly overlapped the expressed lncRNAs with super-enhancers in high-grade serous EOC.

Survival and Super-enhancer-association Analysis. We used the superpc R package to perform the survival analysis using data from profiling of high-grade serous EOCs performed by TCGA (Cancer Genome Atlas Research, 2011). As training set 80% of the samples were randomly selected (377 samples) and the other 20% (77 samples) were used as the test set. Information about vital status was obtained from TCGA. We defined the threshold parameter as 1.4, n.components=3 and prediction.type="continuous". The significant genes were obtained in order of decreasing importance score. Super-enhancer domains in each cell line were identified using published procedures (Lovén et al., 2013; Whyte et al., 2013). To associate super-enhancers to lncRNAs we used the GenomicFeatures package (version 1.28.4) to manipulate the enhancer and lncRNAs genomic annotations. Gene coordinates were obtained using TxDb.Hsapiens.UCSC.hg19.knownGene (version 3.2.2) and org.Hs.eg.db (version 3.4.1) libraries.

RNA-Seq Data Generation and Analysis. Primary ovarian cancer specimens were homogenized and total RNA was extracted using TRIzol LS (Thermo Fisher Scientific, catalogue number: 10296028). Ribosomal RNA (rRNA) was depleted using RiboMinus Transcriptome Isolation Kit (Thermo Fischer Scientific, catalogue number: K155002). Poly (A)+ RNA was then isolated using Dynabeads Oligo (dT)₂₅ (Thermo Fischer Scientific, catalogue number: 61002). Twenty nanograms rRNA-poly (A)+ RNA was used to prepare each RNA-Seq library. External RNA Controls Consortium (ERCC) spike-ins (Thermo Fischer Scientific, catalogue number: 4456740) were added as control for normalization of the samples. Strand-specific RNA-Seq libraries were constructed using the NEBNext Ultra Directional RNA Library Prep Kit (NEB, catalogue number: E7420). The resulting library concentrations were quantified using the Nanodrop. Libraries were sequenced to generate paired-end 75 bp reads on NextSeq 500 platform (Illumina) in high output running mode. Sequencing was performed at the Molecular Genomics Core facility at the University of Southern California. RNA-Seq data were analyzed using Partek Flow and Partek Genomics Suite software after mapping reads to reference lncRNA gene mode (Gencode V16) with Tophat2. **Data Availability.** H3K27ac ChIP-Seq data for normal precursor cells and primary tumors as well as RNA-Seq data for primary tumors have been deposited in the Gene Expression Omnibus database (GEO: GSE121103).

(+)-JQ1 treatment of the ovarian cancer cells. CaOV3 or UWB1.289 cells were treated with 10 nM (+)-JQ1 (Tocris, catalogue number: 4499) or an equal volume of DMSO (Sigma Aldrich, catalogue number: D2650) for 16 h before harvesting for RNA extraction.

Reverse transcription and real time quantitative PCR (RT-qPCR). RNA was extracted using TRIzol LS (Thermo Fisher Scientific, catalogue number: 10296028). M-MLV reverse transcriptase (Promega, catalogue number: M5301) and random hexamers (Promega, catalogue number: C1181) were used for reverse transcription. Gene expression was quantified by RT-qPCR using iQ SYBR Green supermix (Bio-Rad, catalogue number: 170-8886). The relative gene expression was calculated using the $2^{-\Delta\Delta Ct}$ method (Schmittgen and Livak, 2008). Two hundred and fifty nanograms cDNA was used for RT-qPCR analysis on CFX96 Touch Real-Time PCR Detection System (Bio-Rad) using the following primer pairs. UCA1-F: 5'CTTCTGCATAGGATCTGCAATCAG3', UCA1-R: TTTTGTCCCCATTTTCCATCATACG. U1-F: CCAGGGCGAGGCTTATCCATT, U1-R: GCAGTCCCCCACTACCACAAAT. U6-F: GCTTCGGCAGCACATATACTAAAAT, U6-R: CGCTTCACGAATTTGCGTGTCAT. AXL-F: GGTGGCTGTGAAGACGATGA, AXL-R: CTCAGATACTCCATGCCACT. CYR61-F: AAACCCGGATTTGTGAGGT, CYR61-R: GCTGCATTTCTTGCCCTTT. OR10H1-F: CCCAAAAGTCCCCAGTCTCT, OR10H1-R: CTCCTTGTTCCTGAGGCTGA. GAPDH-F: TGCCAAATATGATGACATCAAGAA, GAPDH-R: GGAGTGGGTGTCGCTGTTG, 18S-F: CAGCCACCCGAGATTGAGCA, 18S-R: TAGTAGCGACGGGCGTGTG, β -actin-F: TCACCCACACTGTGCCCATCTACGA, β -actin-R: CAGCGGAACCGCTCATTGCCAATGG.

CRISPR/Cas9 KO of *UCA1***.** Guide RNAs (gRNAs) were designed using the CRISPR design tool (<u>http://crispr.mit.edu/</u>). Two gRNAs were cloned into PX458 (Addgene plasmid number 48138) - *UCA1* promoter: GGTTTCCTTTTAGATGACGG, *UCA1* intron: GGCTAGAATATTTGCAGGCG, following the methods of the Zhang laboratory (Ran et al., 2013). Target cells were co-transfected with the two gRNAs

expressing plasmids or the vector backbone only as a control, using the BioT transfection reagent (Bioland Scientific, catalog number: B01-01). Positive cells were isolated using an Aria II flow cytometer (BD Biosciences) to sort single cells into 96 well plates. For validation of the deletion, PCR was performed using Sequal Prep[™] Long PCR Kit with the following primers - Forward: GACACTGCATTGTGTGCGTT, Reverse: TCCCTGTTGCTAAGCCGATG.

Mouse xenografts. All *in vivo* work was performed with the approval of the USC Institutional Animal Care and Use Committee. Ten million cells were injected with 25% Matrigel subcutaneously into female nu/nu mice (aged 6-8 weeks). Tumor growth was measured by digital caliper measurement and animals euthanized when tumor diameter reached >1.5 cm, or after 8 weeks.

RPPA. Cells were plated into 6 well plates overnight. Cells were washed with PBS and lysed for 20 minutes in buffer containing 1% Triton X-100, 50 mM HEPES, pH 7.4, 150 mM NaCl, 1.5 mM MgCl₂, 1 mM EGTA, 100 mM NaF, 10 mM Na pyrophosphate, 1 mM Na₃VO₄, 10% glycerol, plus PhosSTOP Protease Inhibitor Cocktail (Roche, catalog number: 4906837001) and Protease Inhibitor Cocktail (Roche, catalog number: 11873580001). Insoluble proteins were removed by centrifugation and normalized proteins denatured by boiling in 4× SDS sample buffer (40% Glycerol, 8% SDS, 0.25 M Tris-HCl, pH 6.8 plus 10% 2-mercaptoethanol).

iRAP. Two sets of UCA1 antisense DNA probes with 3' biotin labels were ordered from siTOOLs Biotech GmbH. A U1 antisense probe (Chu et al., 2015) was included as a positive control. OVCA429 were cultured on 150 mm tissue culture plates and used at 90% confluency 96 h after seeding. Cells were washed with 10 mL ice-cold PBS and crosslinked in the presence of 10 mL ice-cold PBS using a Stratalinker (UV Stratalinker 1800, Stratagene) at 254 nm wavelength with 0.8 J/cm2. Cells were collected and centrifuged at 1,000 g for 5 min at 4 °C. The supernatant was removed and cells used directly for iRAP or snap frozen in liquid nitrogen and stored at -80 °C until use. Cells were lysed in Lysis Buffer (10× the mass of pellet, 10 mM Tris-HCl, pH7.0, 50 mM EDTA, 1% SDS) containing freshly added PhosSTOP Protease Inhibitor Cocktail (Roche, catalog number: 4906837001) and Protease Inhibitor Cocktail (Roche, catalog number: 11873580001), PMSF (Sigma Aldrich, catalog number: 93482) and Superase-in (Ambion, catalog number: AM2696). Crosslinked cells were lysed using a Covaris E220 evolution Focused-Ultrasonicator at 4 °C, 20% output, 200 burst cycles, for 15 min for each 20 million cells in 1 mL lysate buffer. Sonicated samples were centrifuged at 16,000 g for 10 min at 4 °C. Cell lysates representing 400 million cells were combined with 2 nM U1 or UCA1-odd/even probe sets in two volumes of hybridization buffer (750 mM NaCl, 1% SDS, 50 mM Tris-HCl pH 7.0, 1 mM EDTA, 15% formamide, with fresh PMSF, phosphatase inhibitor, proteinase inhibitor cocktail, and Superase-in). Hybridization was performed at 37 °C with rotation overnight. RNA-protein complexes were precipitated by adding 2 mL Streptavidin Dynabeads M-270 (Thermo Fisher Scientific, catalog number: 65036) for 30 min at 37 °C. Dynabeads were washed with 2×NaCl and sodium citrate (SSC), 0.5% SDS, and fresh PMSF for 5×5 min.

RIP. Twelve microliter Dynabeads Protein A (Thermo Fisher Scientific, catalog number: 10001D) were washed with 200 μ L HBS (150 mM NaCl, 10 mM HEPES, pH7.5 by KOH and incubated with 2 μ g AMOT antibody, 2 μ g YAP antibody or 2 μ g rabbit IgG isotype control in the presence of 80 μ L HBS buffer at room temperature for 1 h. Eight million CaOV3, OVCA429, or UWB1.289 cells were lysed with 800 μ L cell lysis buffer (10 mM HEPES, pH7.5 by KOH, 150 mM NaCl, 0.1% NP-40, 5 mM EGTA, supplemented with 1× protease inhibitor cocktail and Superase-in) at 4°C for 1 h. Cell debris and insoluble proteins were removed by centrifugation at 4°C, 12,000 g for 10 min, and the supernatants were incubated with antibody-conjugated Dynabeads at 4°C for 1 h. The Dynabeads were then washed 3 times with wash buffer (150 mM NaCl, 10 mM HEPES, pH7.5 by KOH, 0.1% NP-40). Half of the sample was eluted with 12 μ L 1×Laemmli sample buffer (Bio-Rad, catalog number: 1610747) and used for Western blotting, TRIzol LS was added to the other half before proceeding with RNA extraction (Thermo Fisher Scientific, catalog number: 10296028).

Plasmids. Lenti GFP-AMOT p80 and lenti GFP-AMOT p130 were gifts from Dr. Kun-Liang Guan (Addgene plasmid numbers 32830 & 32828). To generate a *UCA1* overexpression plasmid, the full cDNA sequence for *UCA1* was purchased from Genewiz and cloned into the LeGO-iT vector. LeGO-iT was a gift from Dr. Boris Fehse (Addgene plasmid number 27361). Lentiviral particles were produced from co-transfecting HEK293T cells with LeGO-iT+*UCA1*, Lenti GFP-AMOT p80, or Lenti GFP-AMOT p130 together with lentiviral

packaging plasmids pMD2.G and psPAX2 (Gifts from Dr. Didier Trono, Addgene plasmid numbers 12259 & 12260). LeGO-iT has an IRES-tdTomato marker that was used for flow cytometric selection on Aria II cell sorter (BD Biosciences) after transduction.

Transfection. Pooled siRNAs targeting human *UCA1* (Dharmacon, catalog number: R-188002-00-0005), AMOT (Dharmacon, catalog number: L-015417-01-0005), YAP (Dharmacon, catalog number: L-012200-00-0005) or non-targeting scramble controls (Dharmacon, catalog number: D-001810-01-20) were transfected into CaOV3, UWB1.289, or OVCA429 cells using Lipofectamine RNAiMAX transfection reagent (Thermo Fisher Scientific, catalog number: 13778075), DharmaFECT 3 (Dharmacon, catalog number: T-2003-03), or DharmaFECT 1 transfection reagent (Dharmacon, catalog number :T-2001-03), respectively.

Cellular fractionation. Cells were fractionated as previously described (Bahn et al., 2015) with some modifications. Six million OVCA429 cells were treated with the plasma membrane lysis buffer (10 mM Tris-HCl, pH 7.5, 0.15% Nonidet P-40, 150 mM NaCl) on ice for 5 min after homogenization by flicking. For CaOV3 and UWB1.289 cells, six million cells were lysed for 4 min with lysis buffer and for OSEC4C2 cells, eight million cells were treated with lysis buffer for 2 min. Centrifuge at 15,000 g for 10 min at 4°C after loading the lysate onto 24% sucrose cushion (24% RNase-free sucrose in plasma membrane lysis buffer) using large orifice tips. The supernatant (cytoplasmic fraction) was retained after centrifugation, and the pellet (nuclear fraction) was washed with 1×PBS/1 mM EDTA and resuspended in 200 µL of 1×PBS/1 mM EDTA.

Western Blotting. Protein samples were run on 4-20% gradient precast protein gel (Bio-Rad, catalogue number: 456-1096) and transferred onto PVDF membrane (Bio-Rad, catalogue number: 1704157). After 1 h blocking, membranes were incubated with corresponding antibody at 4°C overnight. Membranes were washed three times with Tris-buffered saline containing 0.5% Tween 20 (TBST) before incubating with HRP-conjugated secondary antibody or Clean-Blot IP detection reagent (Thermo Fisher Scientific, catalogue number: 21230) for IP samples at room temperature for 2 h. Then the membranes were incubated briefly with ECL Western Blotting substrate (Thermo Fisher Scientific, catalogue number: 32106) after three times wash with

TBST. The membranes were exposed to HyBlot Autoradiography Film (Denville Scientific, catalogue number:

E3018). Antibody information is listed in Supplementary Table 6.

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