

# Supplemental Material

## “Set-up and Validation of a High Throughput Screening Method for Human Monoacylglycerol Lipase (MAGL) Based on a New Red Fluorescent Probe”.

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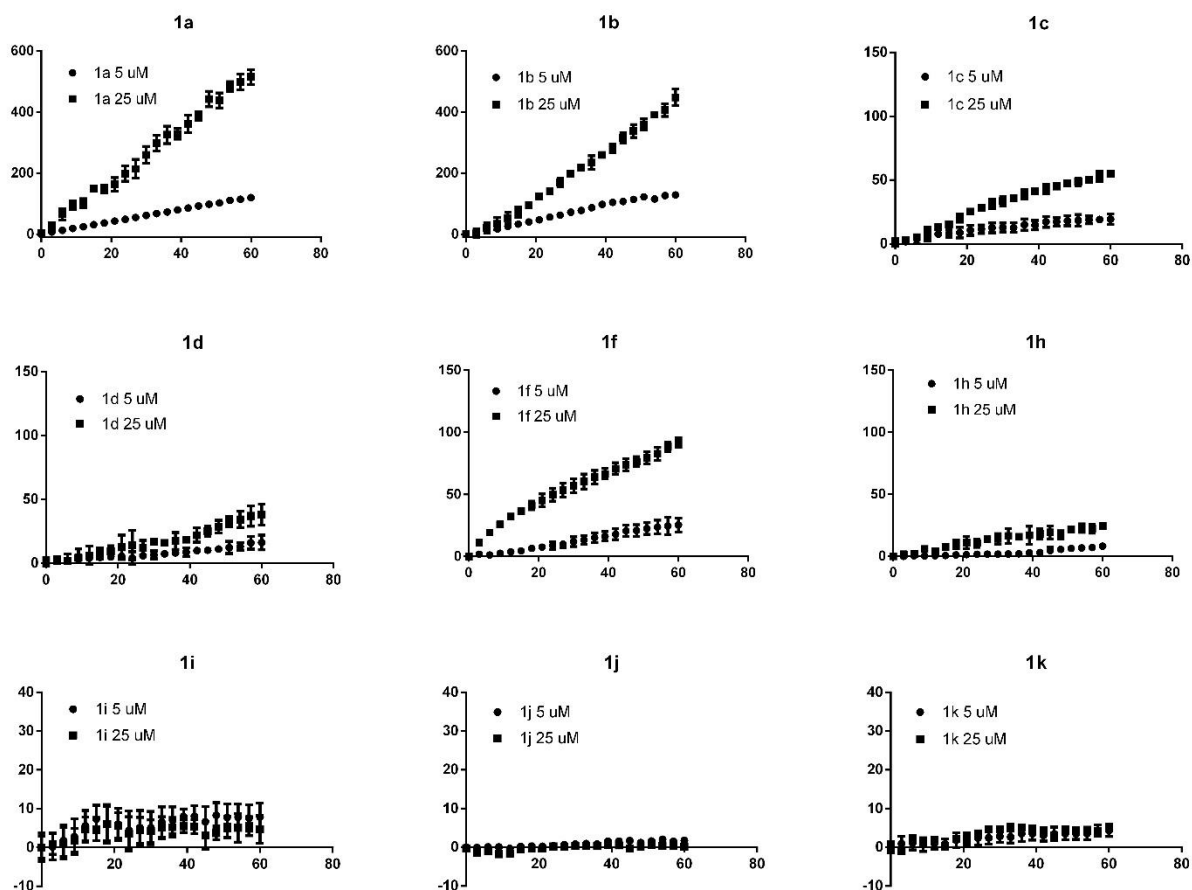
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**Table S1:** Mobile phases used for the separation of synthesis products.

Compound(s)	Yields	Mobile phase	m.p.
<b>1a</b>	65%	CH <sub>2</sub> Cl <sub>2</sub> /acetone, 98:2	220-221 °C <sup>a</sup>
<b>1b</b>	71%	Petroleum ether/AcOEt gradient, from 7:3 to 5:5.	138-140 °C <sup>a</sup>
<b>1c</b>	68%	Petroleum ether/AcOEt 8:2.	111-112 °C
<b>1d</b>	72%	Petroleum ether/AcOEt gradient, from 8:2 to 7:3.	117-119 °C
<b>1e</b>	75%	CH <sub>2</sub> Cl <sub>2</sub> /acetone, 98:2	124-125 °C
<b>1f</b>	66%	Petroleum ether/AcOEt 8:2.	82 °C
<b>1g</b>	62%	CH <sub>2</sub> Cl <sub>2</sub> /acetone, 98:2	Oil <sup>b</sup>
<b>1h</b>	90%	Petroleum ether/AcOEt 75:25.	144-145 °C
<b>1i</b>	82%	CH <sub>2</sub> Cl <sub>2</sub> /acetone, 98:2	124-125 °C
<b>1j</b>	92%	CH <sub>2</sub> Cl <sub>2</sub> /acetone, 98:2	94-96 °C
<b>1k</b>	92%	Petroleum ether/ AcOEt 9:1 to remove side products, then CH <sub>2</sub> Cl <sub>2</sub> /acetone 98:2.	259-261 °C <sup>a</sup>

<sup>a</sup> Guilbault, G.G., and Kramer, D.N. (1965). Resorufin Butyrate and Indoxyl Acetate as Fluorogenic Substrates for Cholinesterase. *Analytical Chemistry*, 37(1), 120-123.

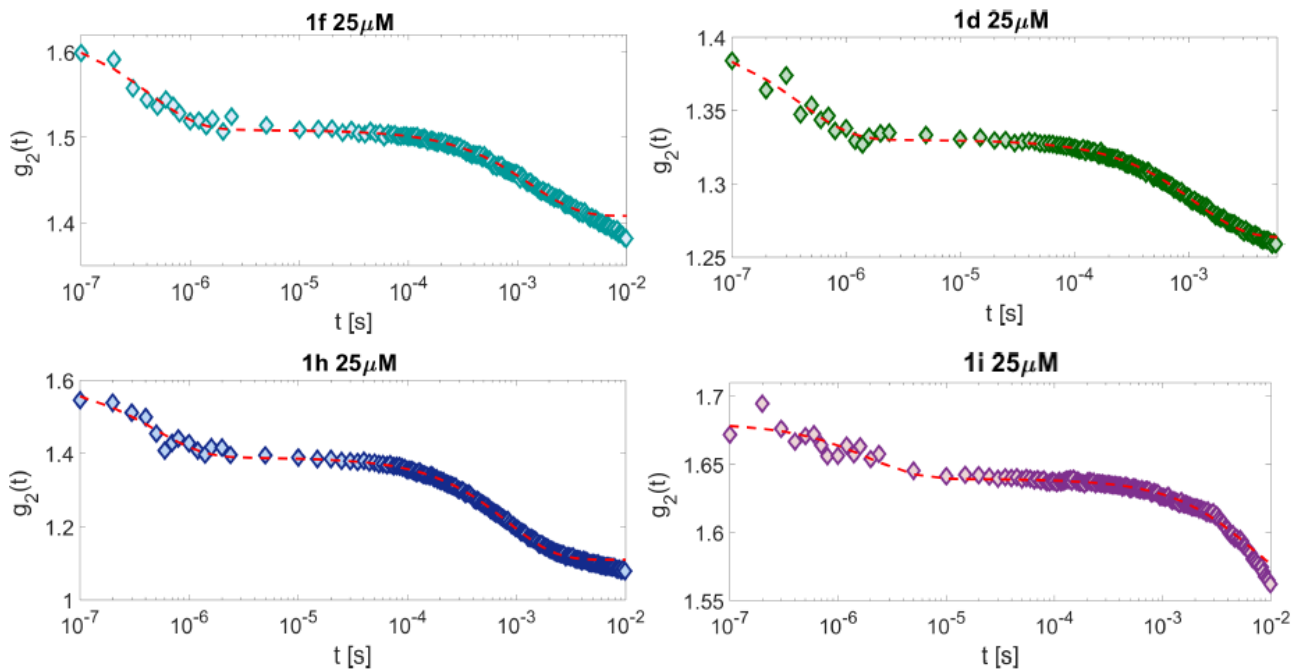
<sup>b</sup> Lauria, S., Casati, S., and Ciuffreda, P. (2015). Synthesis and characterization of a new fluorogenic substrate for monoacylglycerol lipase and application to inhibition studies. *Analytical and Bioanalytical Chemistry*, 407(26), 8163–8167. <https://doi.org/10.1007/s00216-015-8991-9>



**Figure S1:** Comparison between spontaneous hydrolysis of **1a-d**, **1f**, and **1h-k** at lowest (circles) and highest (squares) concentrations assessed (5  $\mu\text{M}$  and 25  $\mu\text{M}$ ). For all compounds, data have been normalized subtracting the  $t_0$  fluorescence value from all the next fluorescence value read. Data are mean  $\pm$  standard error of independent experiments.

**Table S2:** Ligand: MAGL interactions.

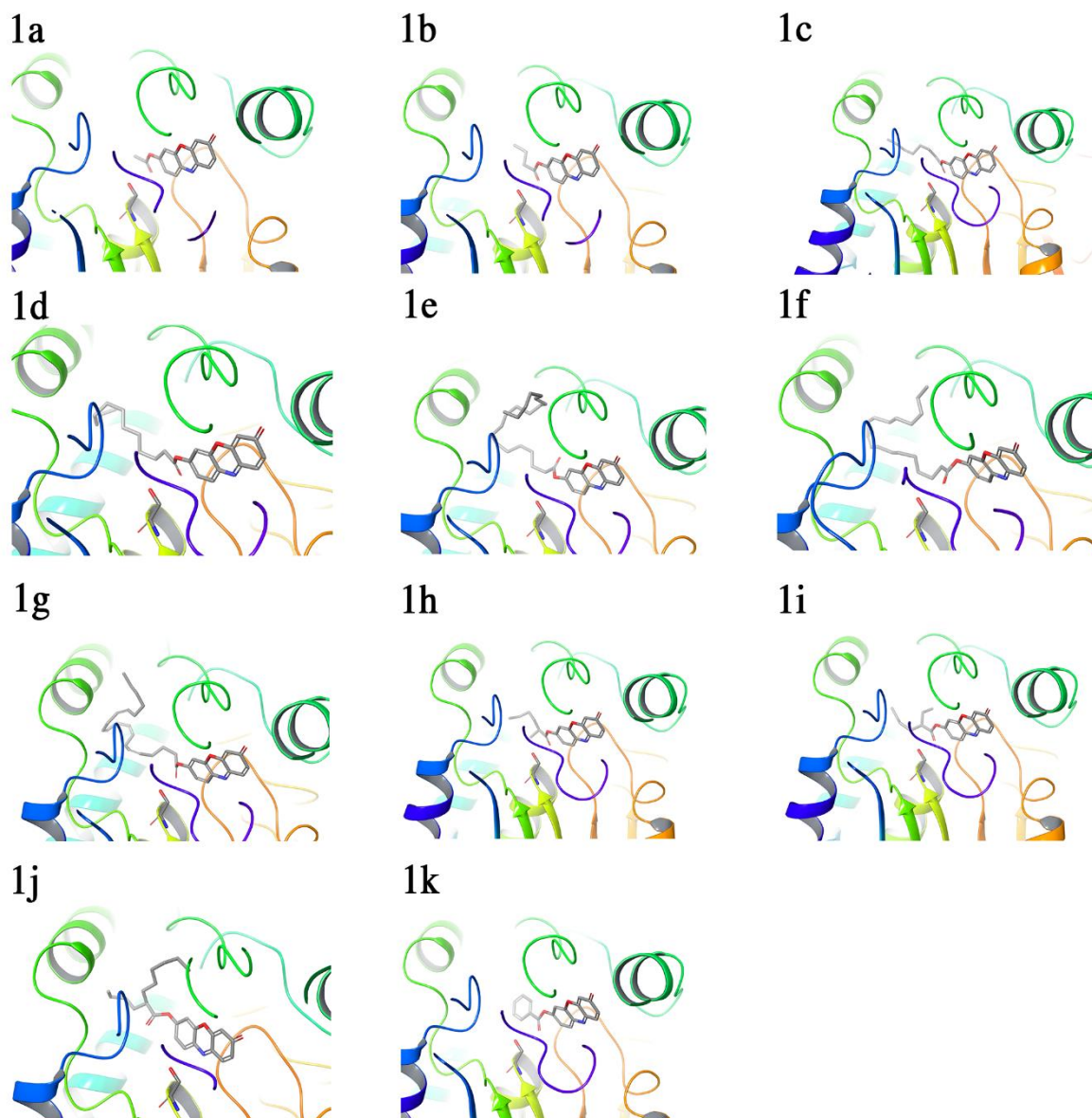
Compound	H-bond	Pi-Pi interaction
<b>1a</b>	Ala51, Met123	His269
<b>1b</b>	Ala51, Met123	His269
<b>1c</b>	Ala51	His269
<b>1d</b>	Ala51	His121, His269
<b>1e</b>	Ser122	His121, His269
<b>1f</b>	Ser122	
<b>1g</b>	Ala51, Ser122, Met123	His121, His269
<b>1h</b>	Ala51, Met123	His121, His269
<b>1i</b>	Ala51, Met123	His121, His269
<b>1j</b>	Ala51	
<b>1k</b>	Ala51	His121



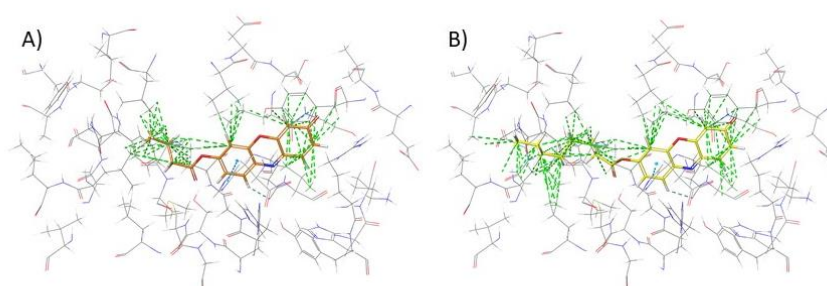
**Figure S2:** Time correlation functions for the scattered intensity (at  $\theta = 90^\circ$ ). Red dashed lines: double exponential fit. The fastest decay time is of the order of fractions of  $\mu$ s which does not correspond to diffusive motion. Whereas the slower decay times reveal nano-sized particles. The corresponding decay times and radius are reported in Tab.3S.

**Table S3:** Decay times and corresponding radius.

SAMPLE	$\tau$ [ms]	R [nm]
<b>1f</b>	$2.7 \pm 0.2$	$287 \pm 25$
<b>1d</b>	$2.3 \pm 0.2$	$246 \pm 16$
<b>1h</b>	$1.7 \pm 0.1$	$182 \pm 13$
<b>1i</b>	$13 \pm 8$	$1390 \pm 860$



**Figure S3:** Binding poses of tested compounds into the MAGL binding site. MAGL is represented as ribbons; tested compounds and Ser122 are represented as sticks without hydrogen atoms.

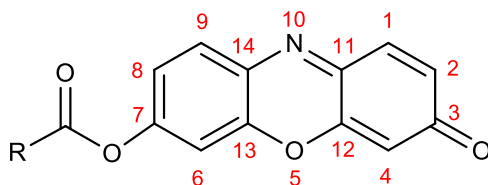


**Figure S4:** MAGL complexes. Compound **1b** (A) and **1c** (B) are shown in orange and yellow sticks, respectively; only MAGL residues participating to the binding site are reported in stick representation; green traces point out residues involved in hydrophobic interactions.

## $^1\text{H}$ and $^{13}\text{C}$ -NMR data

### *Instruments*

$^1\text{H}$ -NMR spectra were recorded in  $\text{CDCl}_3$  (isotopic enrichment 99.95%) solutions at 300 K using a Bruker AVANCE 500 instrument (500.13 MHz for  $^1\text{H}$ , 125.76 MHz for  $^{13}\text{C}$ ) using 5 mm inverse detection broadband probes and deuterium lock. Chemical shifts ( $\delta$ ) are given as parts per million relative to the residual solvent peak (7.26 ppm for  $^1\text{H}$ ) and coupling constants ( $J$ ) are in Hertz. The experimental error in the measured  $^1\text{H}$ - $^1\text{H}$  coupling constants is  $\pm 0.5$  Hz. The splitting pattern abbreviations are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet, and bs, broad peak. For two-dimensional experiments, Bruker microprograms using gradient selection (gs) were applied.



**Figure S5.** IUPAC numbering of 7-hydroxyresorufinyl moiety.

**Table S4.** Assignment of  $^1\text{H}$  and  $^{13}\text{C}$ -NMR Chemical Shift in  $\text{CDCl}_3$ .

	<i>1a</i>		<i>1c</i>		<i>1b</i>		<i>1f</i>		<i>1k</i>		<i>1d</i>	
	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)
CH(1)	7.46 (d) 9.8	134.8	7.47 9.8	134.9	7.46 9.9	134.8	7.46 9.8	134.8	7.49 9.8	134.8	7.47 9.8	134.8
CH(2)	6.89 (dd) 9.8, 2.0	135.2	6.90 9.8, 2.0	135.1	6.89 9.9, 2.1	135.1	6.90 9.8, 2.0	135.1	6.91 9.8, 2.0	135.2	6.90 9.8, 2.0	135.1
C(3)		186.3		186.3		186.3		186.3		186.3		186.3
CH(4)	6.36 (d) 2.0	107.2	6.37 2.0	107.2	6.36 2.1	107.2	6.36 2.0	107.2	6.38 2.0	107.3	6.37 2.0	107.2
CH(6)	7.18 (d) 2.4	109.7	7.18 2.4	109.7	7.18 2.4	109.7	7.17 2.4	109.7	7.32 2.4	109.9	7.18 2.4	109.7
C(7)		153.5		153.7		153.6		153.7		153.8		153.7
CH(8)	7.15 (dd) 8.6, 2.4	119.3	7.15 8.6, 2.4	119.3	7.15 8.6, 2.4	119.3	7.15 8.6, 2.4		7.30 8.6, 2.4	119.4	7.15 8.6, 2.4	119.3
CH(9)	7.82 (d) 8.6	131.1	7.83 8.6	131.1	7.82 8.6	131.1	7.83 8.6		7.89 8.6	134.2	7.83 8.6	131.1
C(11)		148.3		148.2		148.2		148.2		148.3		148.2
C(12)		149.3		149.3		149.3		149.3		149.3		149.3
C(13)		144.3		144.4		144.3		144.4		144.5		144.4
C(14)		131.2		131.2		131.1		131.2		131.2		131.2
COO		168.5		171.4		171.2		171.4		164.4		171.4
OCOCH <sub>2</sub>			2.63 (t) 7.5	34.4	2.62 7.0	36.2	2.63 7.0	34.4			2.63 7.5	34.4
CO CH <sub>2</sub> CH <sub>2</sub>			1.79 (tt) 7.5, 7.5	31.6	1.84 7.0, 7.4	18.3	1.80 7.0, 7.0	22.7			1.79 7.5, 7.5	24.8
CH=CH							5.42-5.34 (2H, m, 9', 10')	129.7 (9'), 130.1 (10')				

CH=CH- CH <sub>2</sub>							2.08-2.02 (4H, m, 8', 11')	27.1 (8'), 27.2 (11')				
CH <sub>2</sub> -FAC <sup>a</sup>			1.27-1.47 (20 H, m, 4'-7')	29.0, 28.9, 24.8, 24.7			1.45-1.26 (20 H, m, 17')	22.7 (17'), 29.0, 29.1, 29.2, 29.4, 29.5, 29.6, 29.7 (4'-7' and 12'- 15'), 31.9 (16')			1.47-1.24 (16 H, m, 4'-11')	22.7, 29.1, 29.2, 29.3, 29.5, 29.6, 31.9 (4'-11')
CH <sub>3</sub>	2.38	21.2	0.93 7.0	14.1	1.09 7.4	13.6		14.1				
<i>o</i> -H <sub>ar</sub>									8.24 (d) 7.7	130.4		
<i>m</i> -H <sub>ar</sub>									7.58 (t) 7.7	128.8		
<i>p</i> -H <sub>ar</sub>									7.72 (t) 7.7	131.2		

<sup>a</sup>fatty acid chain

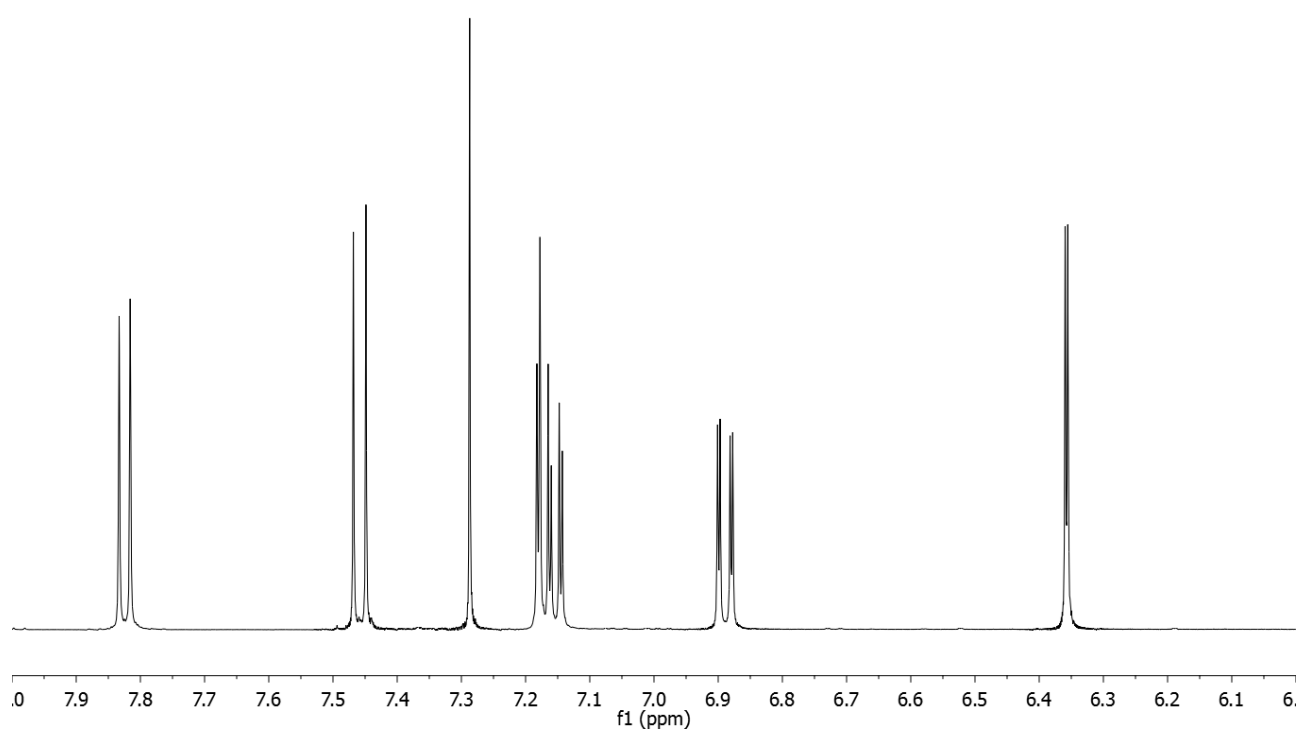
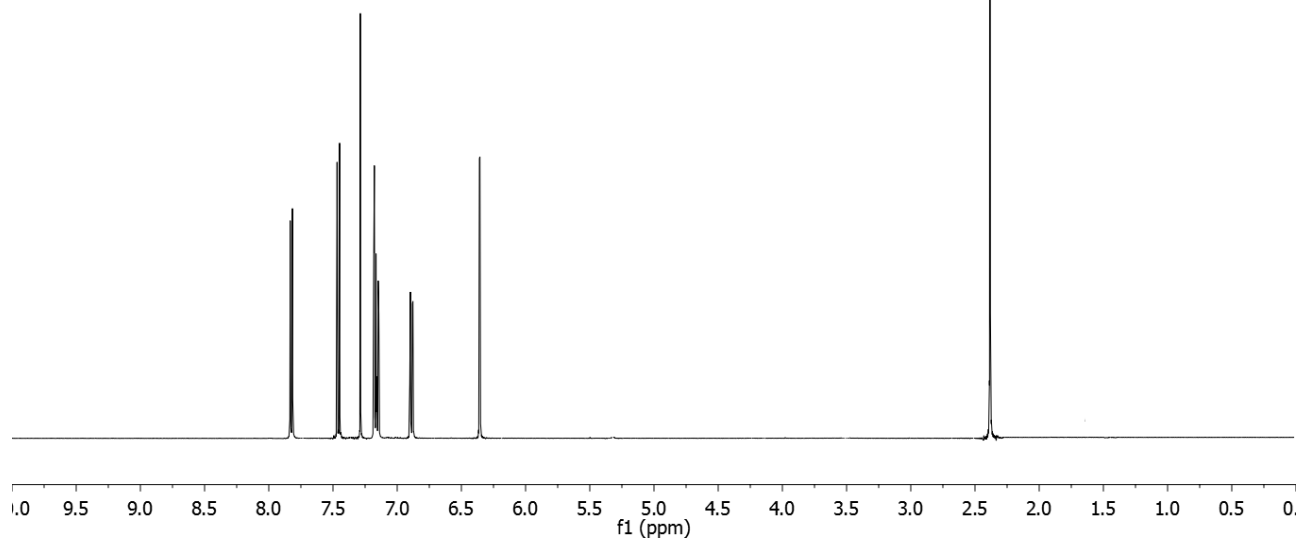
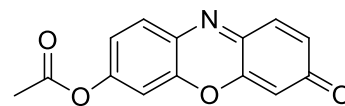


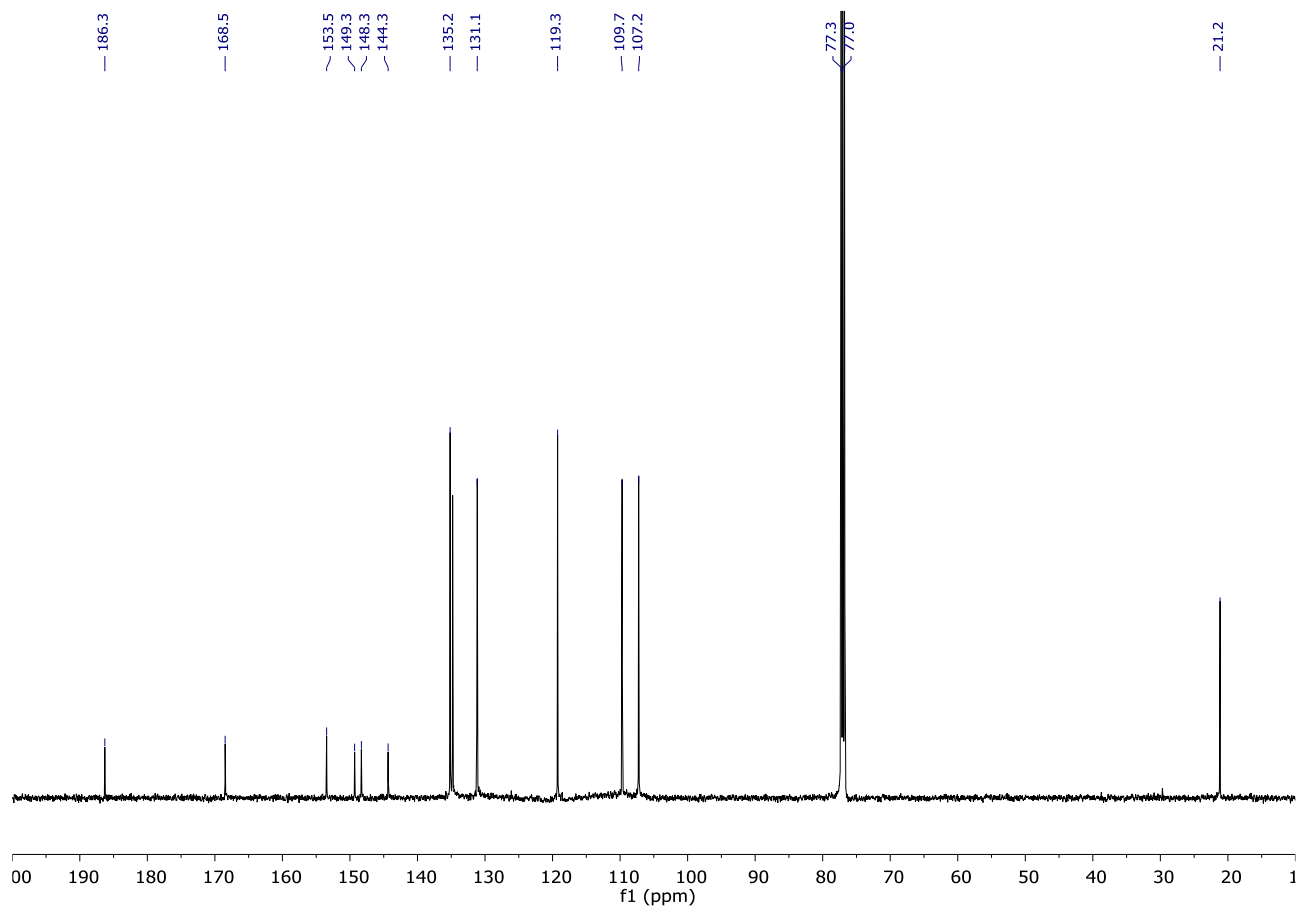
## Continued from Table S4...

	<i>1j</i>		<i>1h</i>		<i>1i</i>		<i>1e</i>		<i>1g</i>	
	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)	$\delta$ (H)	$\delta$ (C)
CH(1)	7.46 (d) 9.8	134.8	7.46 (d) 9.8	134.8	7.47 (d) 9.8	134.8	7.46 9.8	134.8	7.46 9.8	134.8
CH(2)	6.89 (dd) 9.8, 2.0	135.1	6.89 (dd) 9.8, 2.0	135.1	6.90 (dd) 9.8, 2.0	135.1	6.89 9.8, 2.0	135.1	6.89 9.8, 2.0	135.2
C(3)		186.3		186.3		186.3		186.3		186.3
CH(4)	6.36 (d) 2.0	107.2	6.36 (d) 2.0	107.2	6.36 (d) 2.0	107.2	6.36 2.0	107.2	6.36 2.0	107.3
CH(6)	7.15 (d) 2.4	109.7	7.16 (d) 2.4	109.7	7.16 (d) 2.4	109.8	7.18 2.4	109.7	7.17 2.4	109.7
C(7)		153.8		153.8		153.8		153.7		153.6
CH(8)	7.13 (dd) 8.6, 2.4	119.4	7.14 (dd) 8.6, 2.4	119.3	7.13 (dd) 8.6, 2.4	119.4	7.14 8.6, 2.4	119.3	7.14 8.6, 2.4	
CH(9)	7.83 (d) 8.6	131.1	7.83 (d) 8.6	131.1	7.83 (d) 8.6	131.1	7.82 8.6	131.1	7.82 8.6	
C(11)		148.2		148.2		148.2		148.2		148.3
C(12)		149.3		149.4		149.3		149.3		149.4
C(13)		144.3		144.4		144.4		144.4		144.3
C(14)		131.2		131.2		131.1		131.2		131.2
COO		174.2		174.5		174.1		171.4		
OCOCH <sub>n</sub> ( <sub>2,1</sub> )(CH <sub>2</sub> )	2.63 (tt) 7.0, 7.0	45.9	2.75 (tq) 7.0	39.7	2.58 (tt) 7.7, 7.0	47.4	2.63 7.5	34.4	2.65 (t) 7.0	33.7 (2')
OCOCH <sub>n</sub> ( <sub>2,1</sub> )(CH <sub>2</sub> )	1.83-1.74 (m) 1.67-1.59 (m)	32.4 32.1	1.84 (3') 7.0, 7.0	33.3	1.80 (3') 7.4, 7.0	33.3	1.79 7.5, 7.5	24.8	1.88 (3') 7.0, 7.3	24.6 (3')

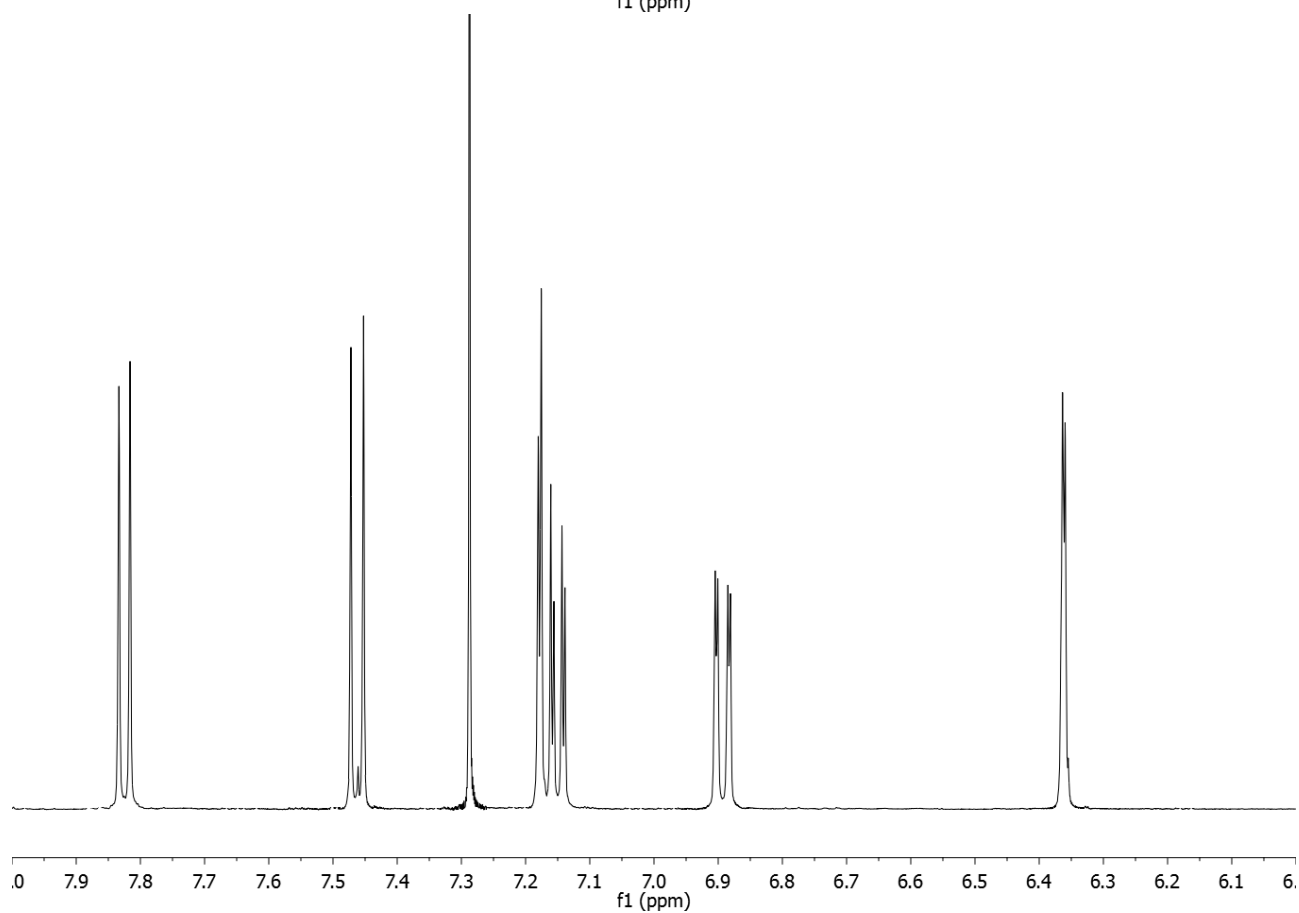
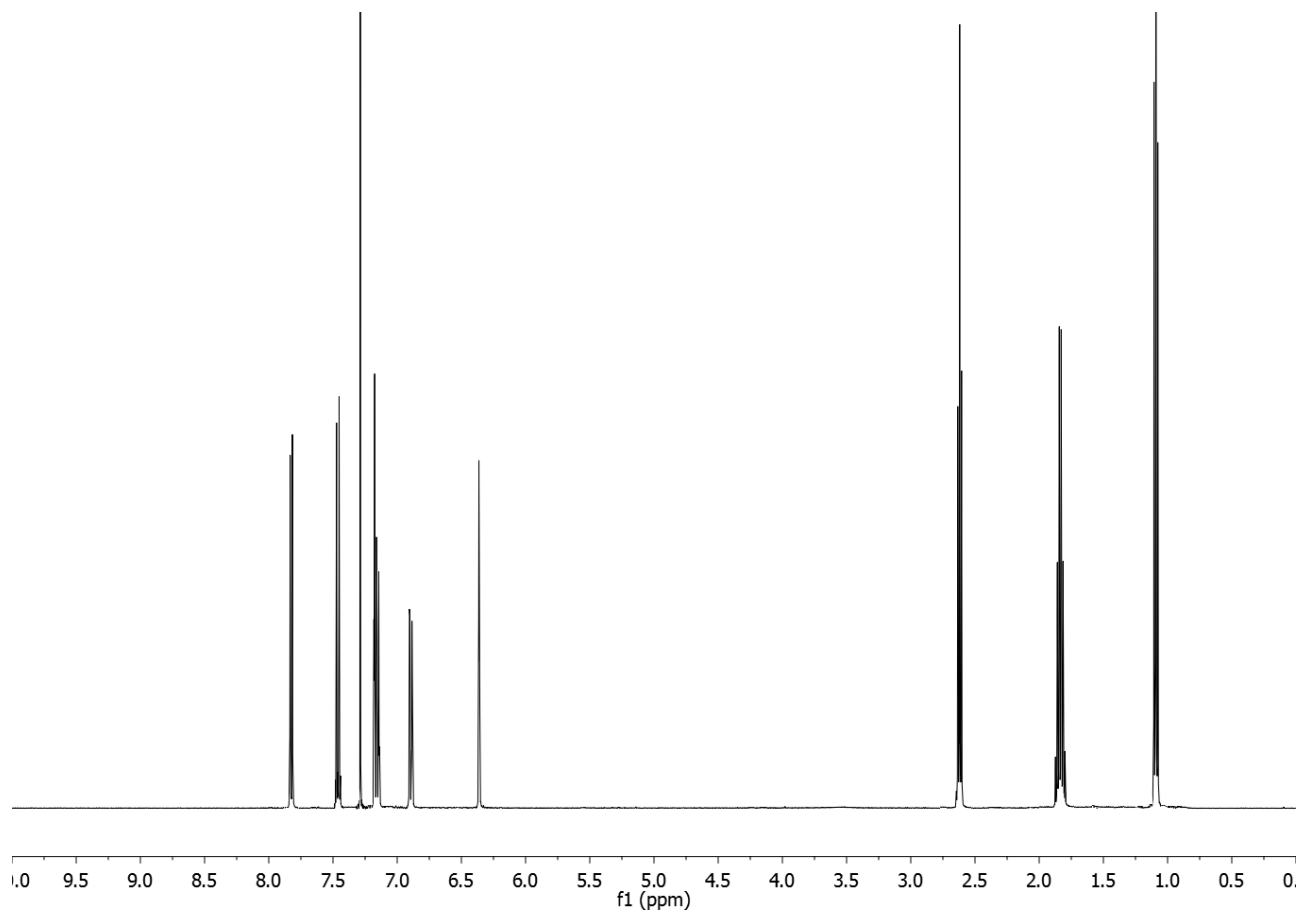
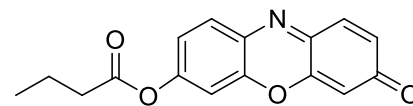
CH=CH				-		-			5.52-5.33 (8H, m, 5',6',8',9', 11',12', 14',15')	130.6,129.5,128.7, 128.5,128.4,128.1,1 27.8,127.5 (5',6',8',9', 11',12', 14',15')
CH=CH-CH <sub>2</sub>				-		-			2.88-2.82 (6H, m, 7', 10', 13')	27.7 (7', 10', 13')
CH <sub>2</sub> -FAC <sup>a</sup>	1.44-1.28 (12 H, m)	31.7, 29.7, 29.2, 27.5, 22.6, 22.6	1.64-1.58 1.45-1.39 (4H)	29.4 22.6	1.74-1.62 1.46-1.36 (6H)	31.6 29.6 25.4	1.46-1.25 (32 H, m, 4'-19')	22.7, 248, 29.1, 29.2, 29.3, 29.5, 29.6, 29.7, 31.9 (4'-19')	2.25 (4', dt) 7.0, 7.3 1.39-1.28 (6 H, m, 17'-19')	22.6 (19'), 29.3 (17'), 31.5 (18'), 26.5 (4'), 27.2 (16')
CH <sub>3</sub>	0.96 (t) 0.92 (t) 7.0	14.1 14.0	1.34 (d) 0.97 (t) 7.0	10.4 16.9	1.06 (t) 7.4 0.97 (t) 7.0	11.9 14.0	0.90 (t) 7.0	14.1	0.91 (t) 7.6	14.1

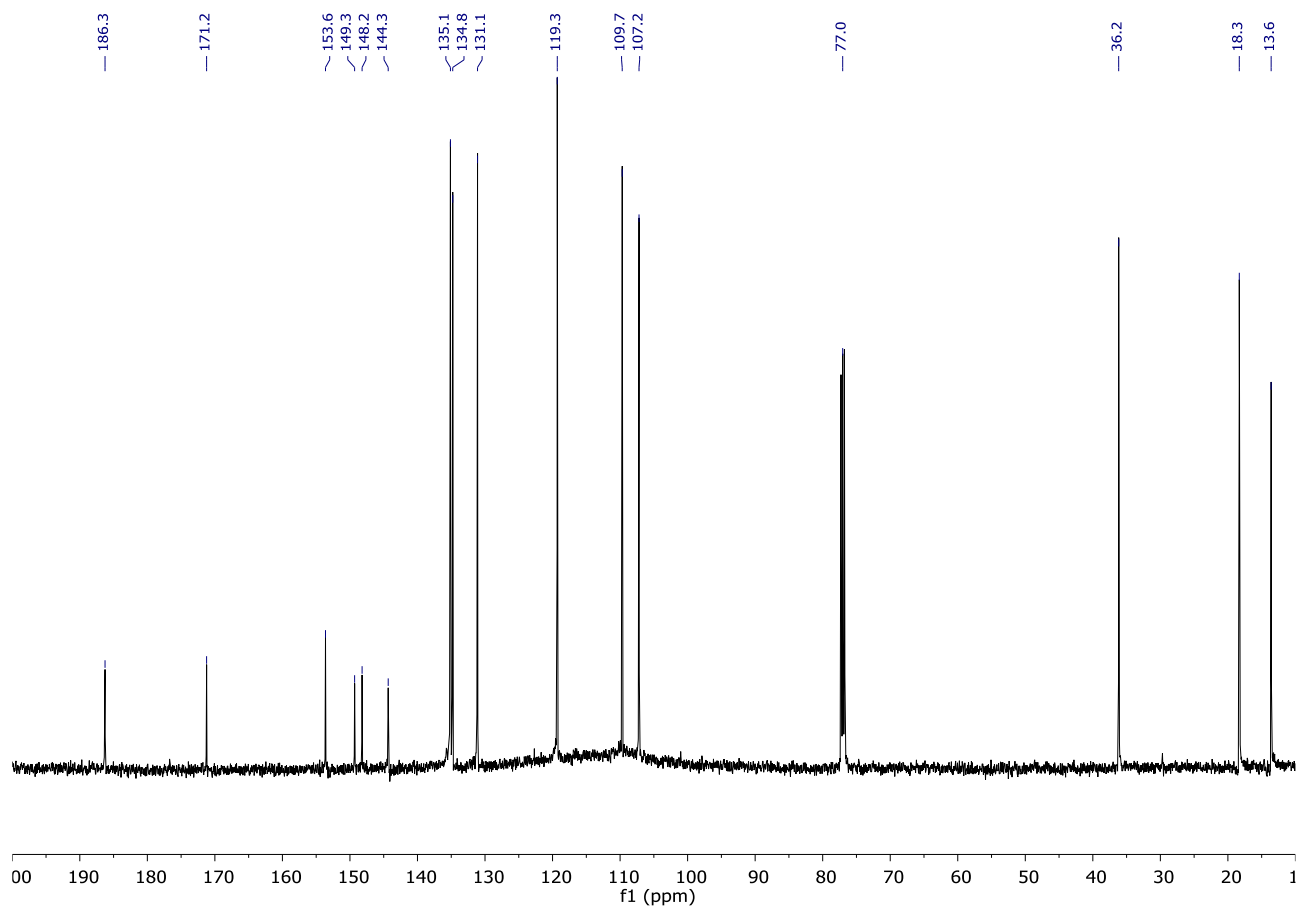
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-acetate (**1a**)



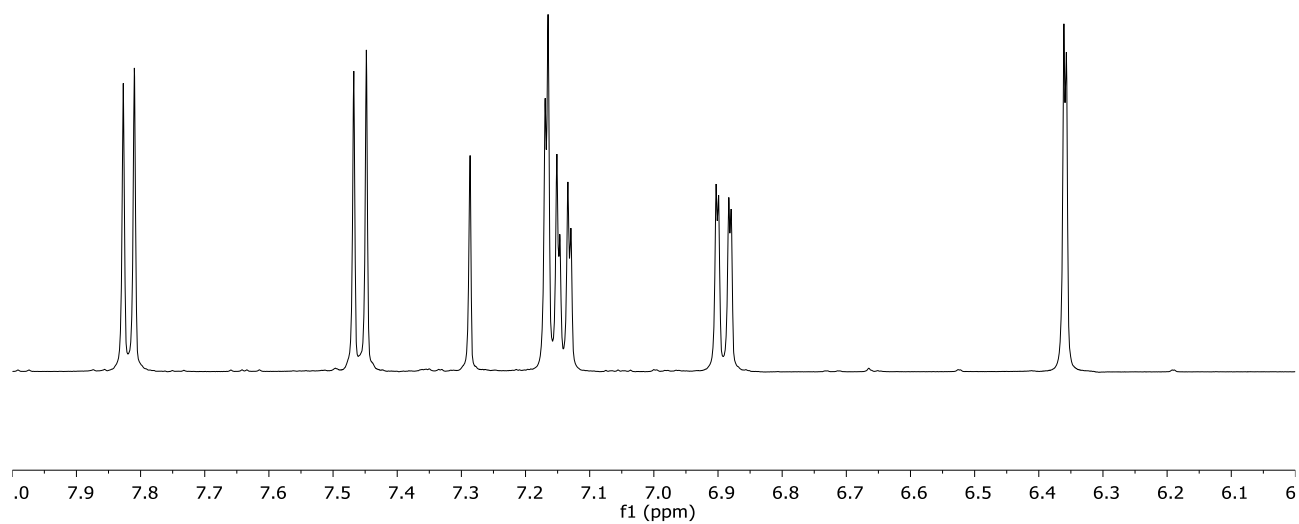
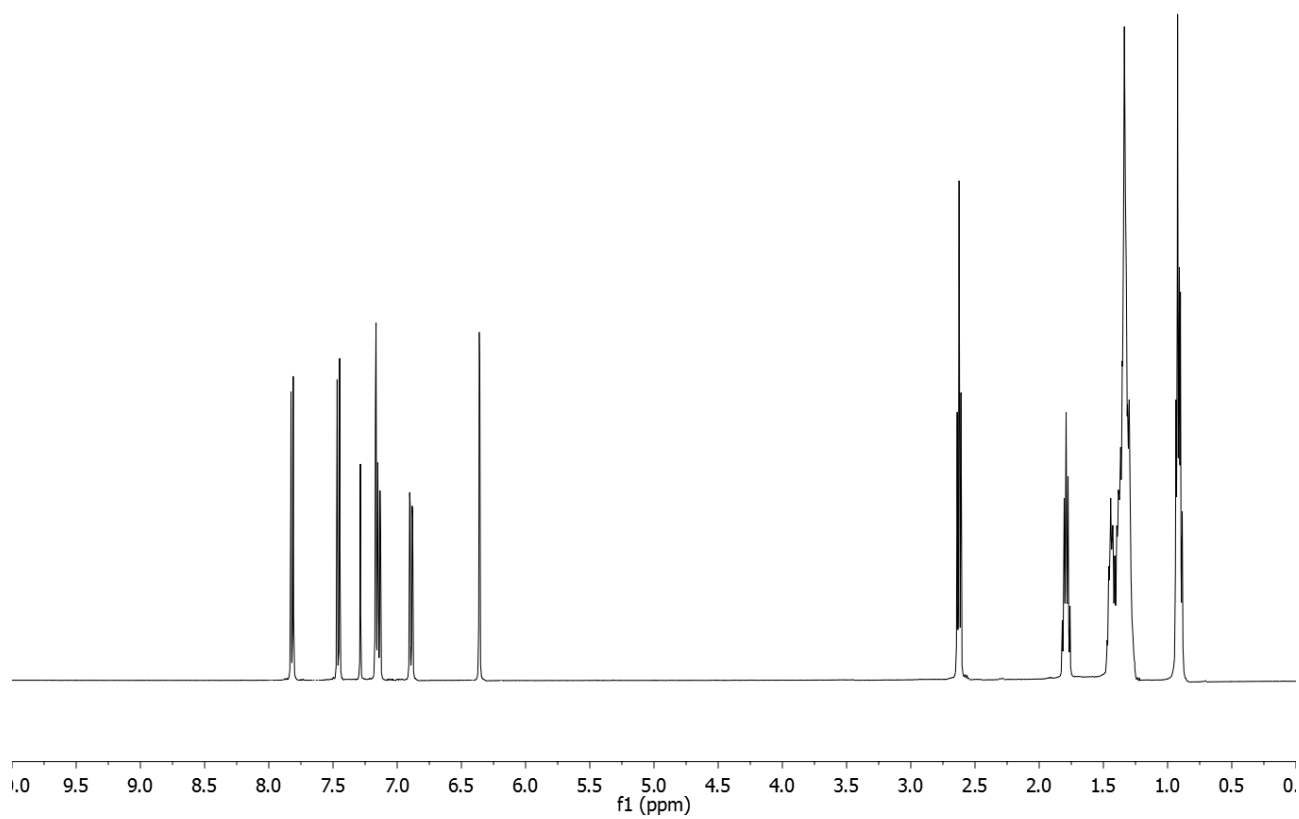
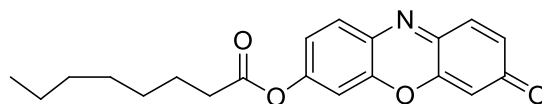


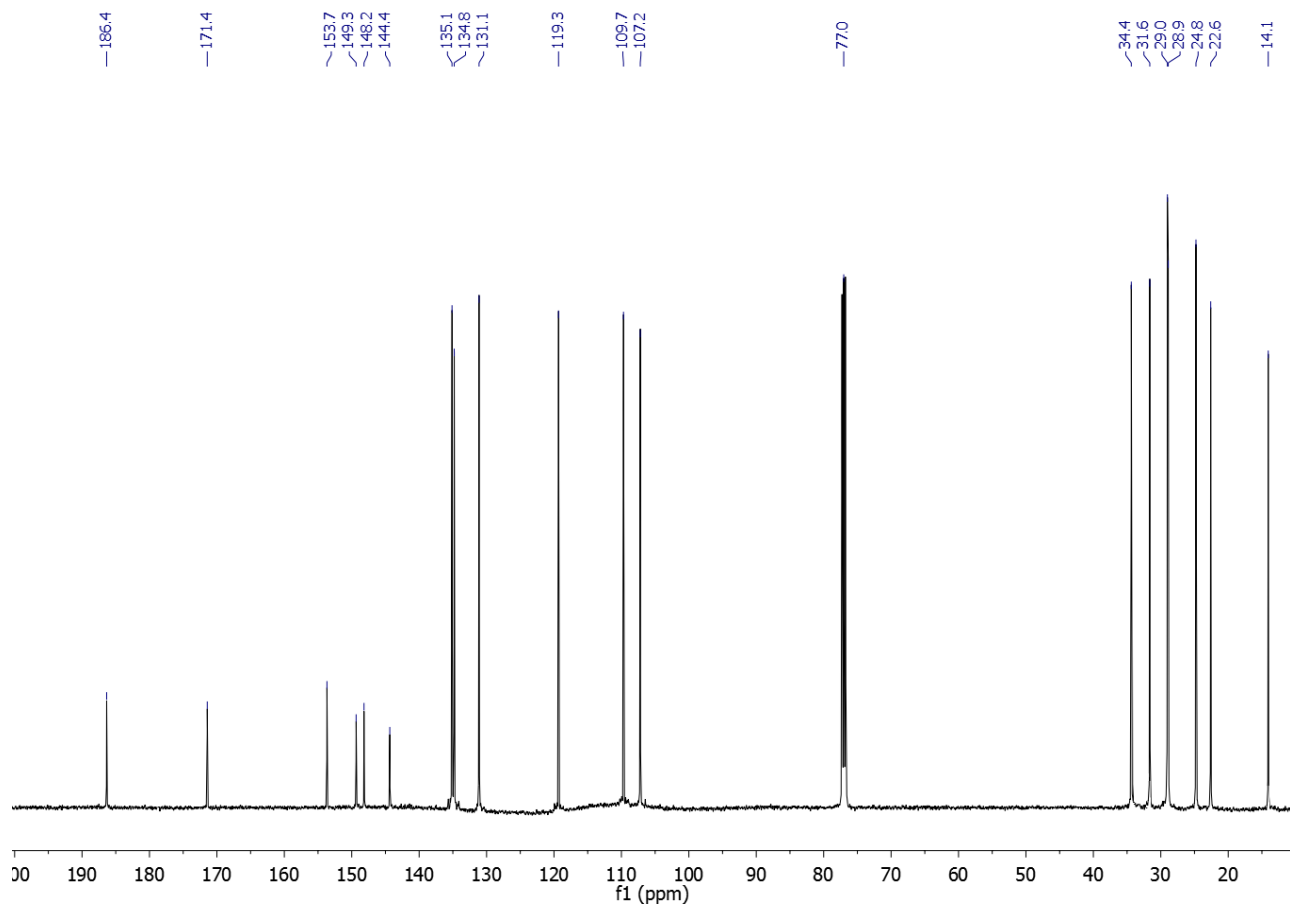
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-butylate (**1b**)





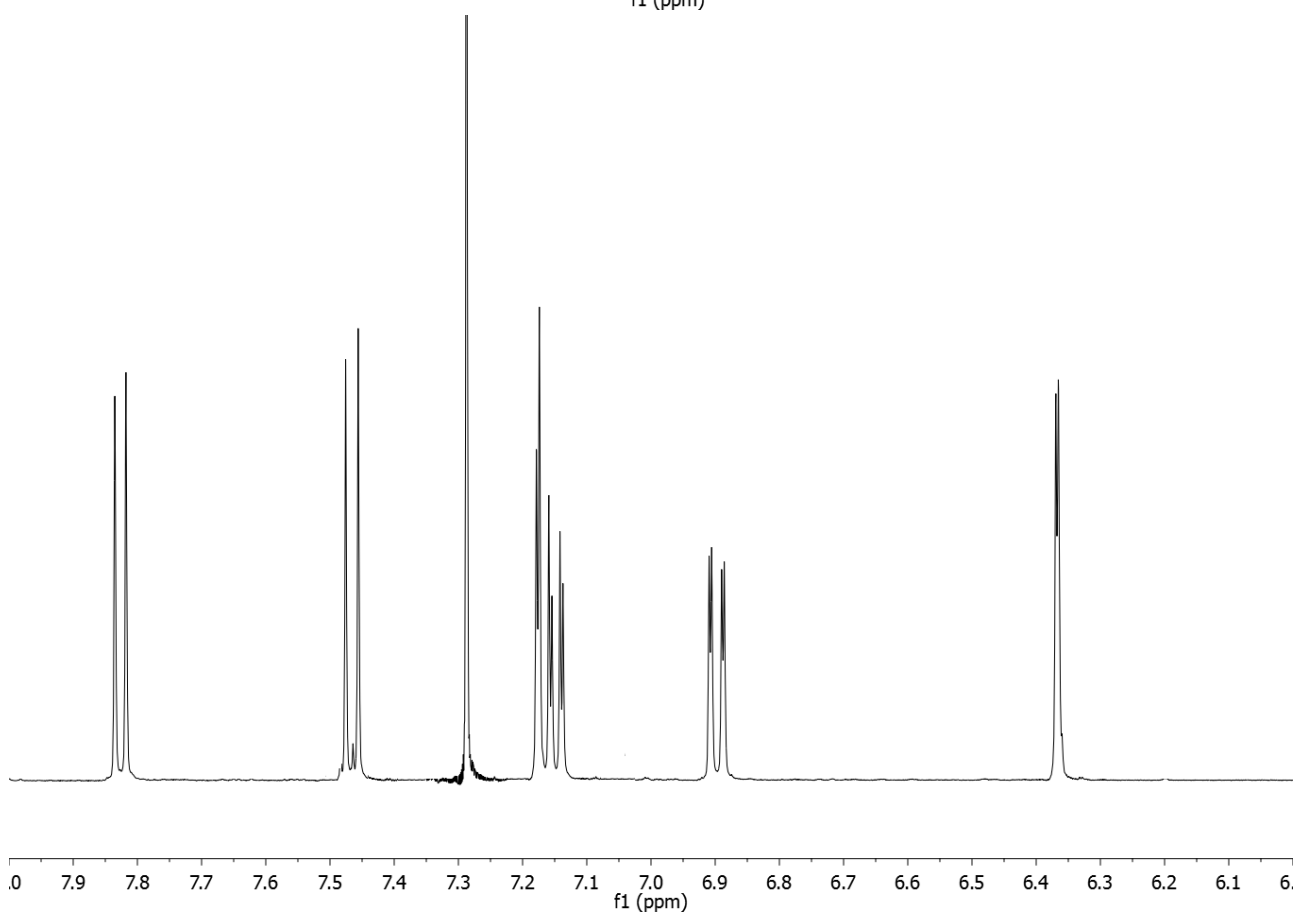
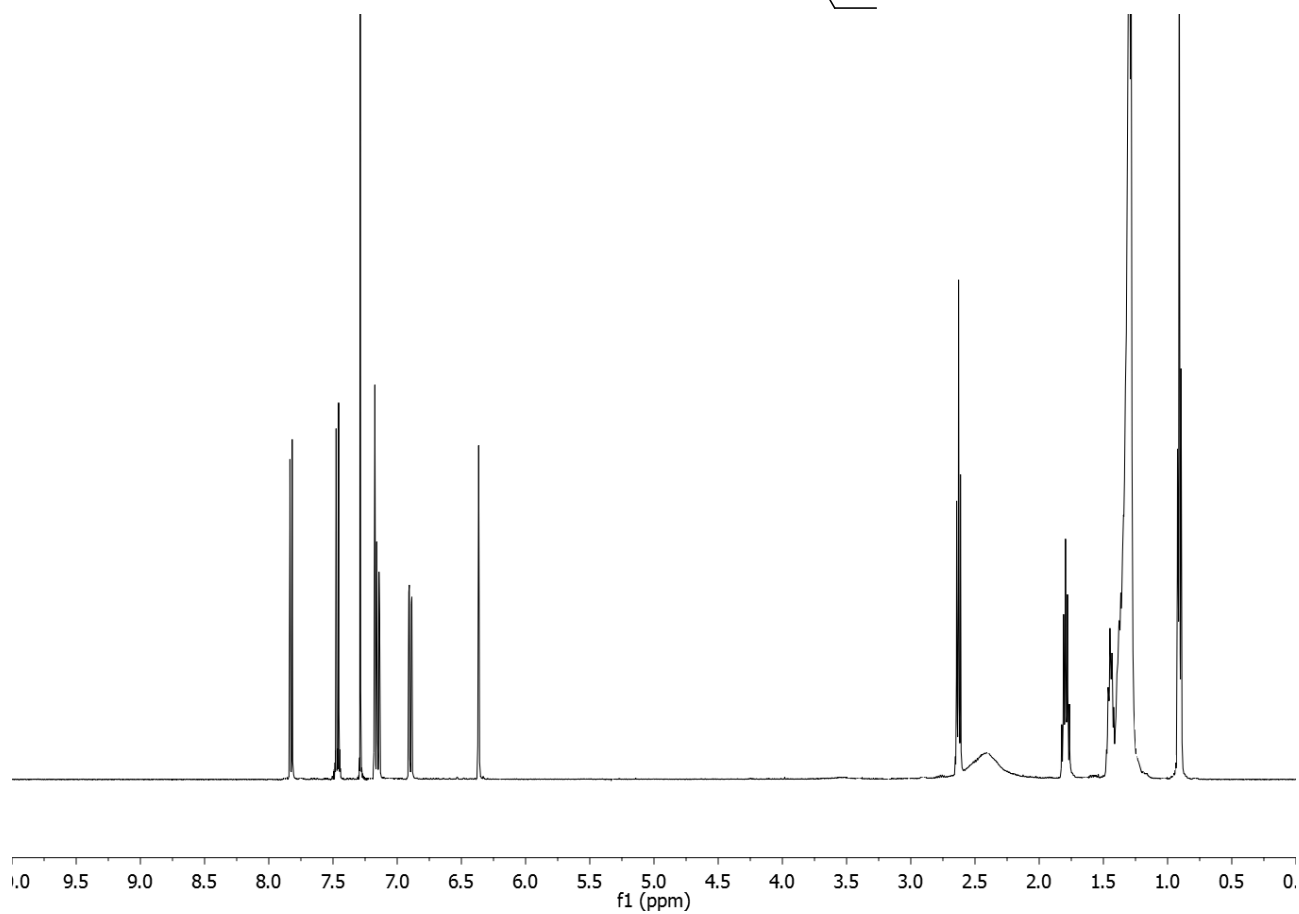
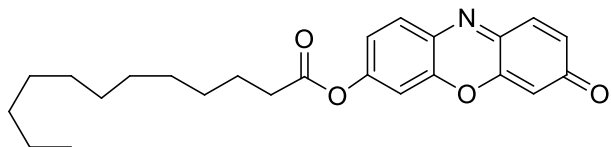
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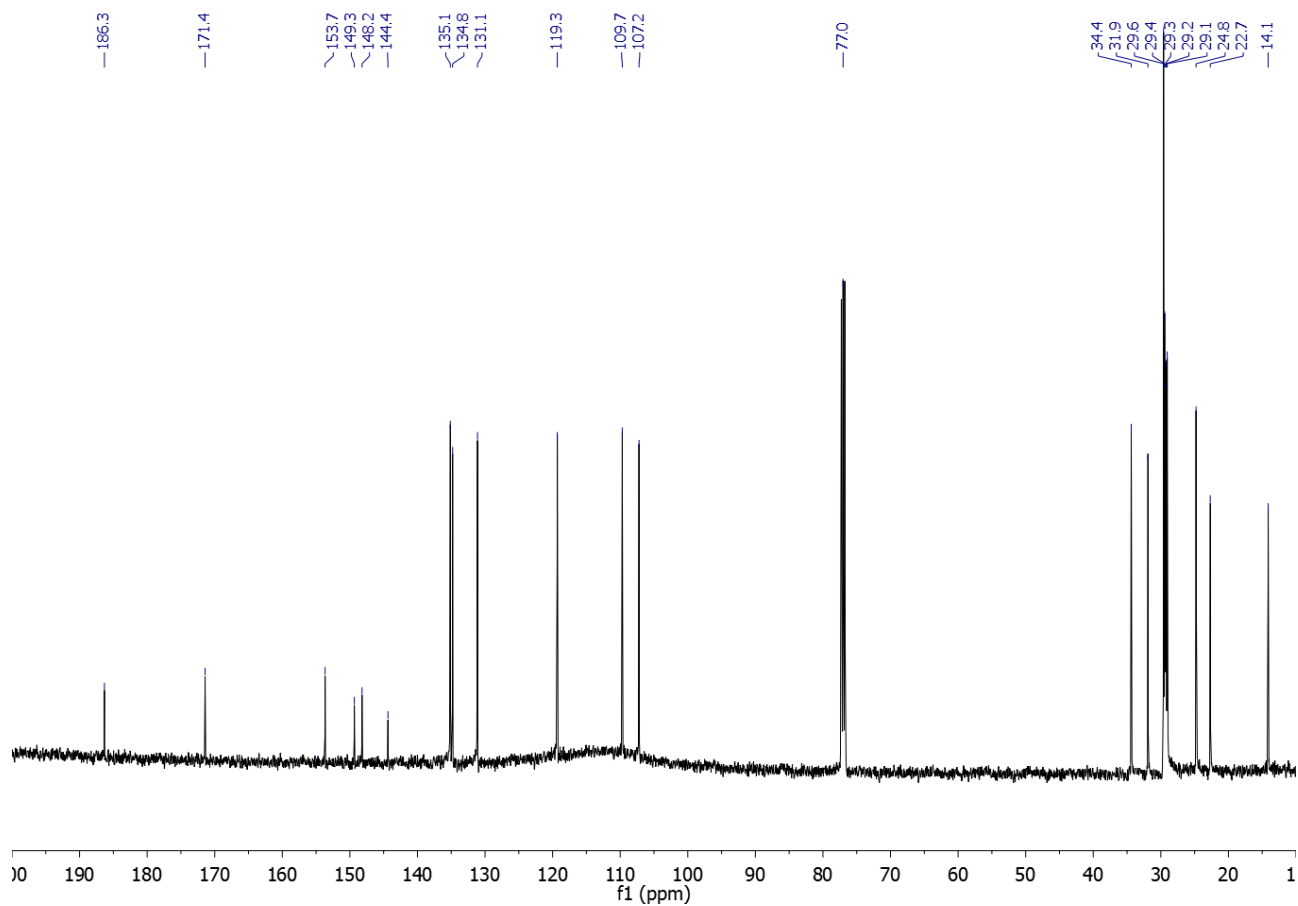




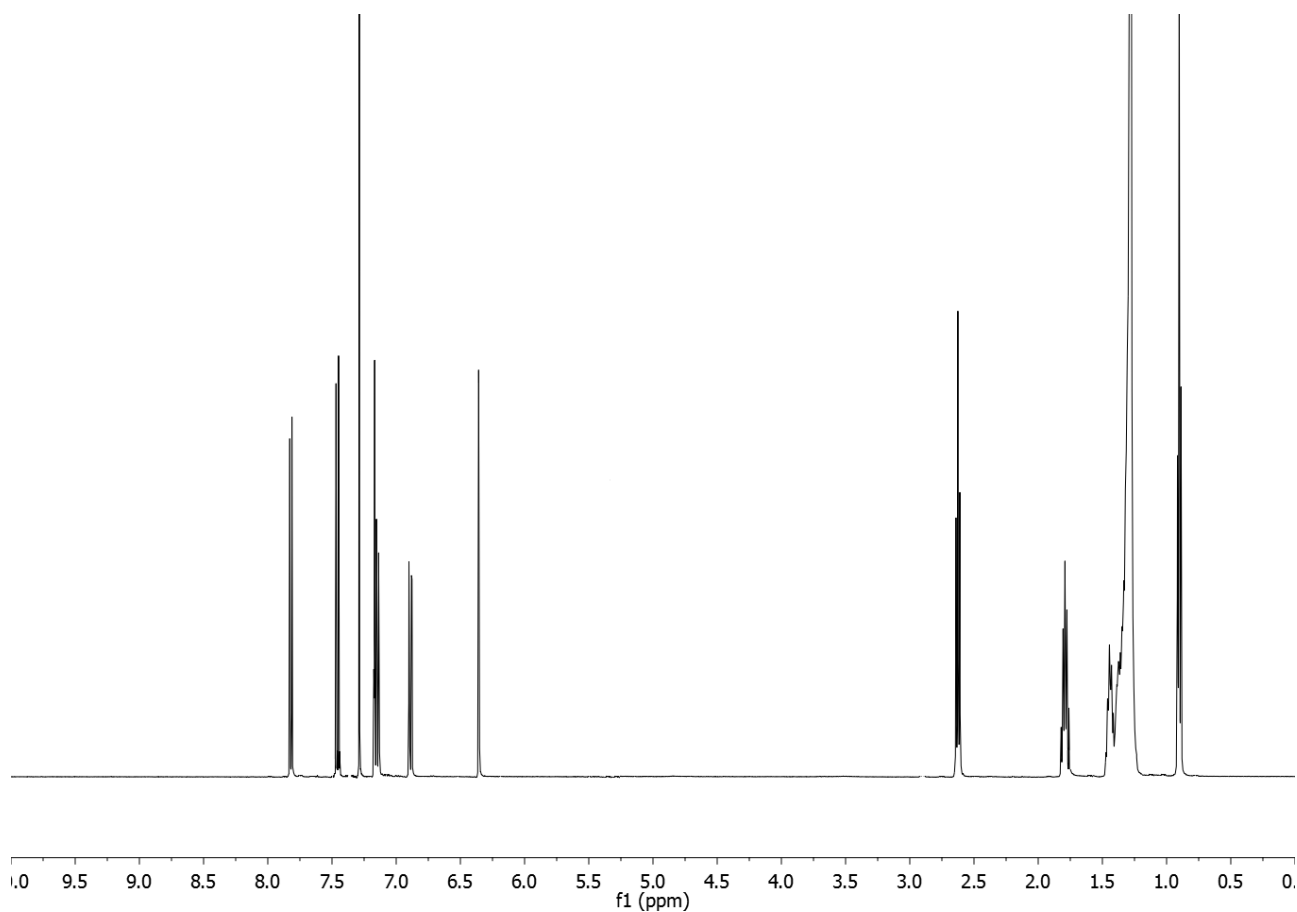
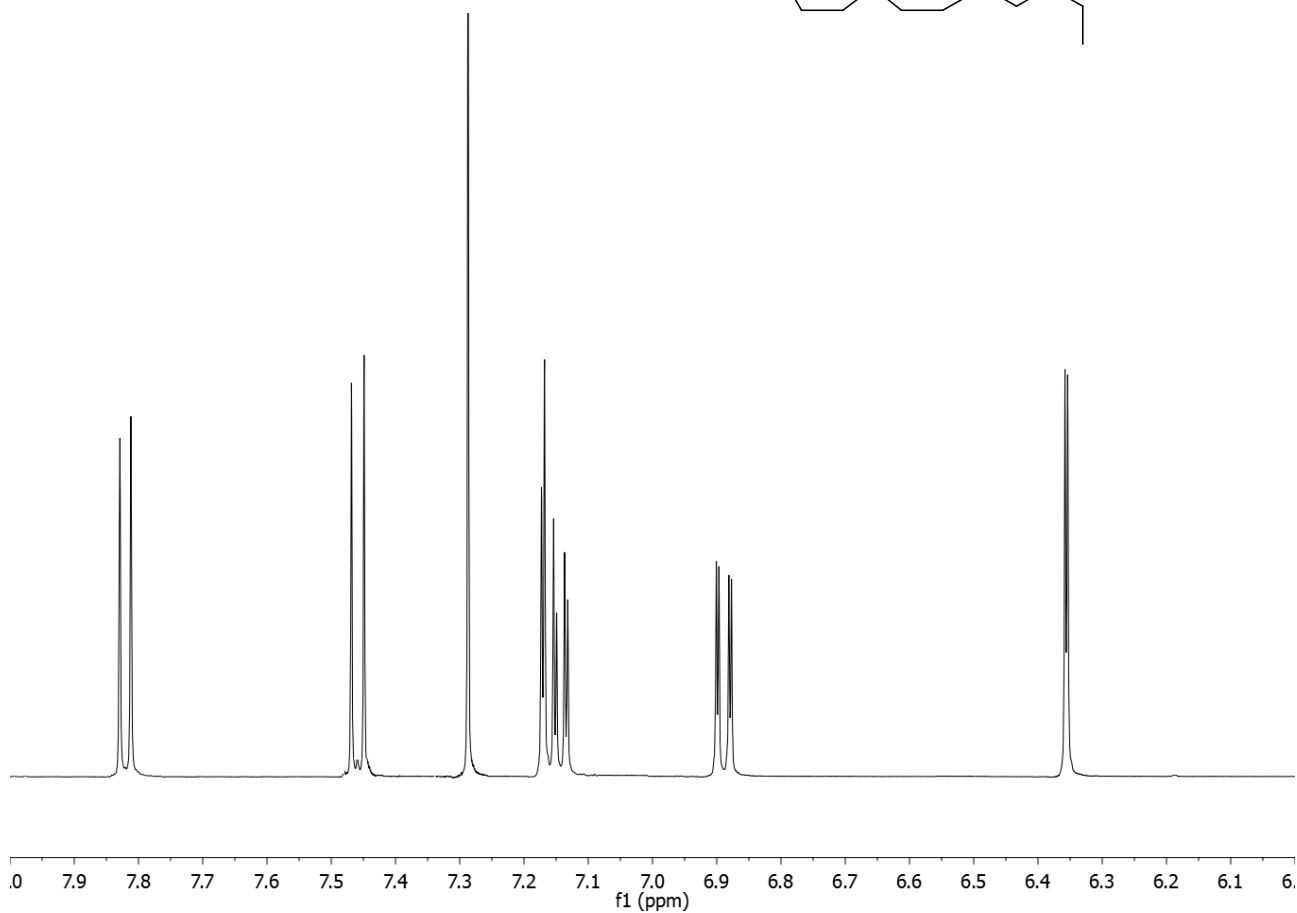
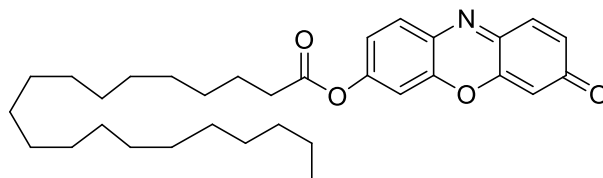


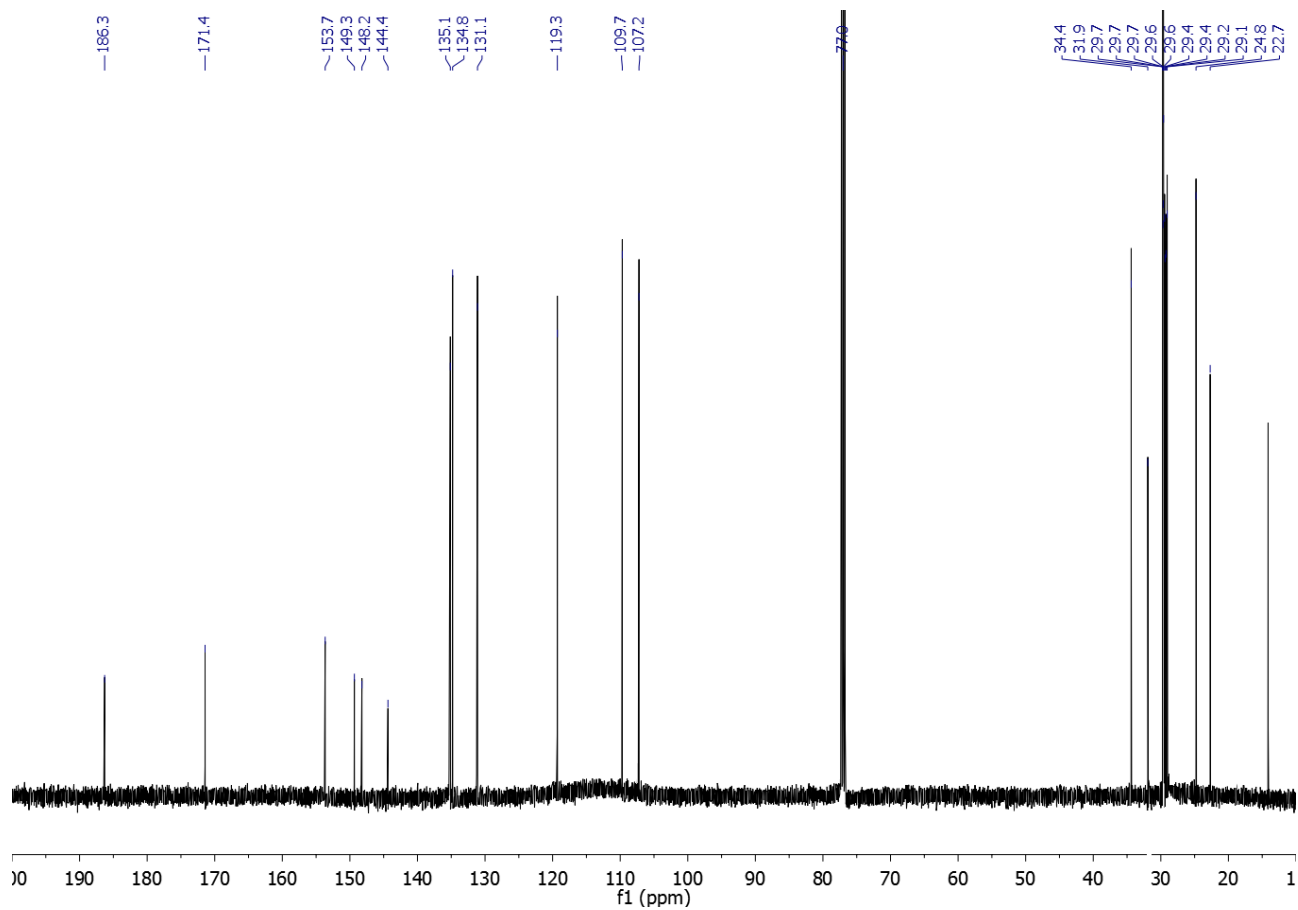
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-dodecanoate (**1d**)



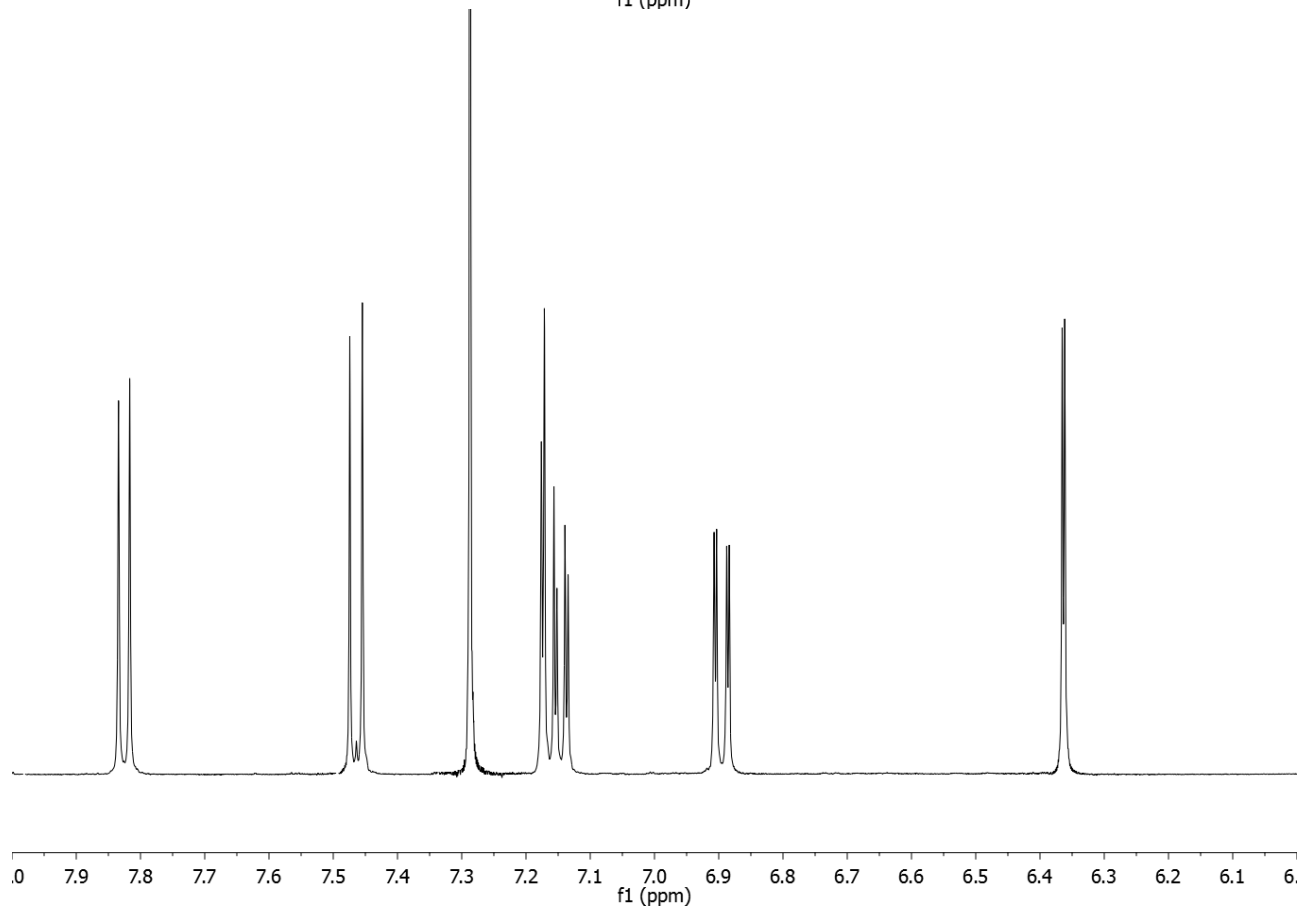
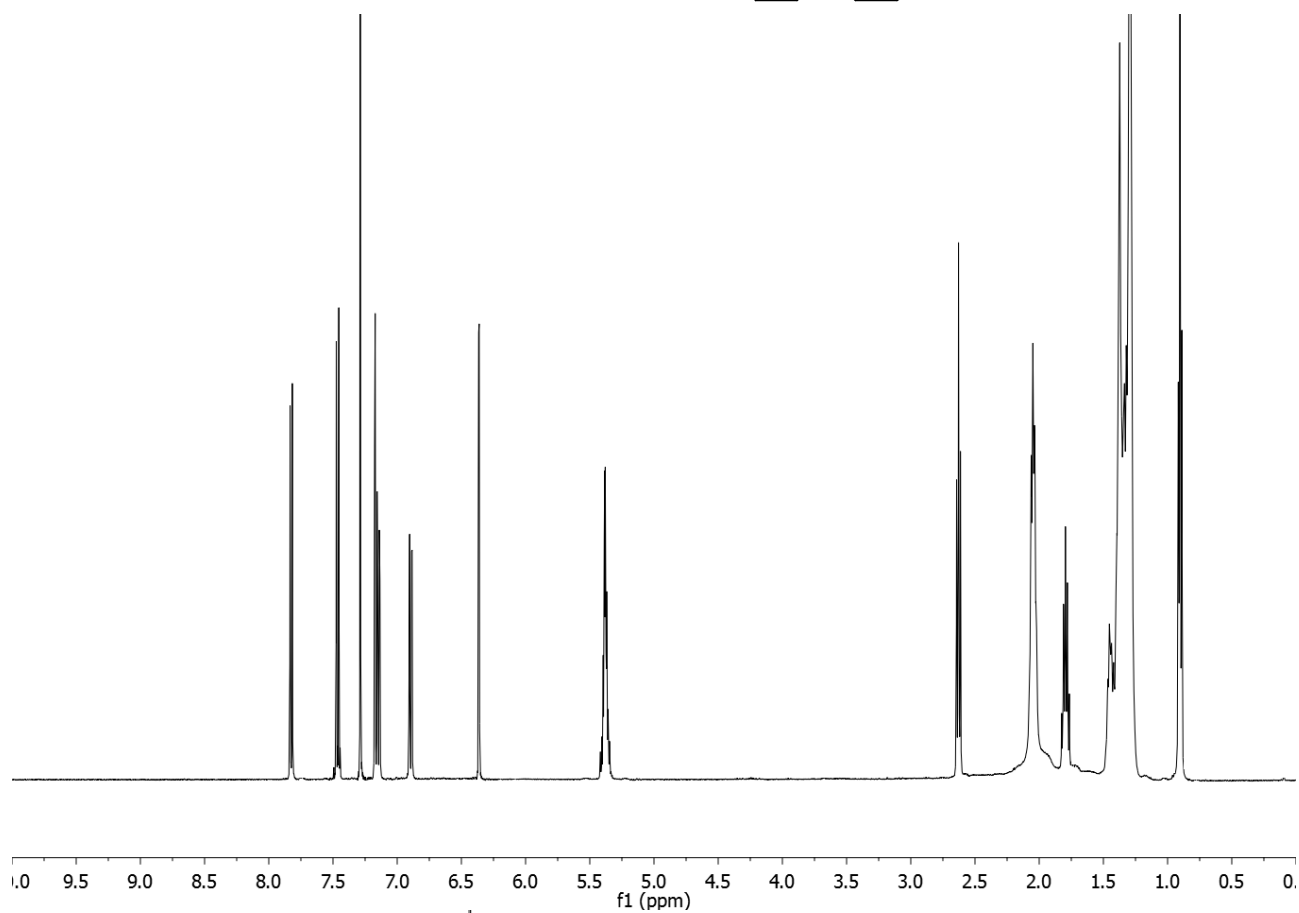
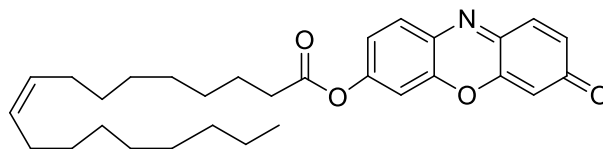


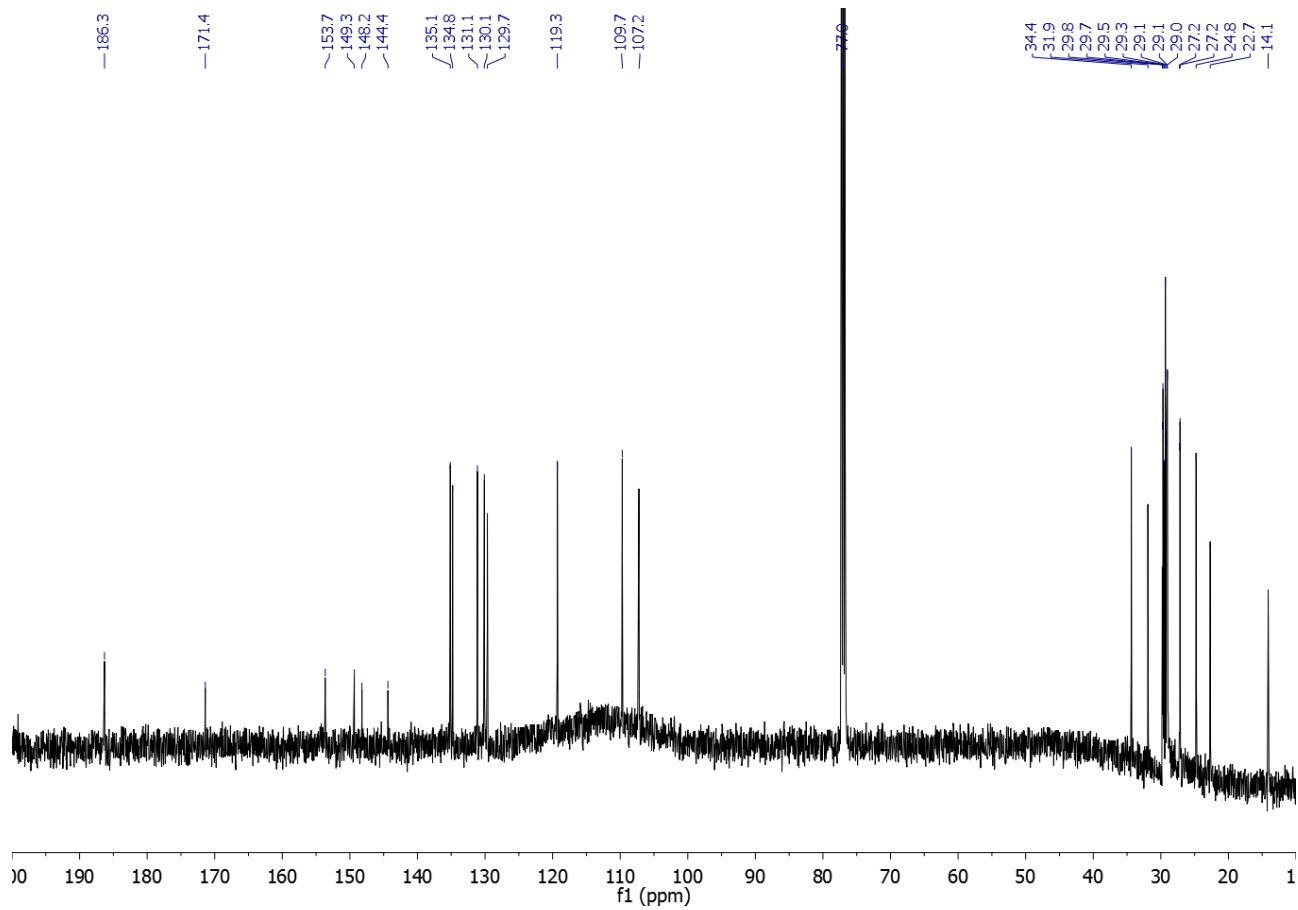
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-icosanoate (1e)



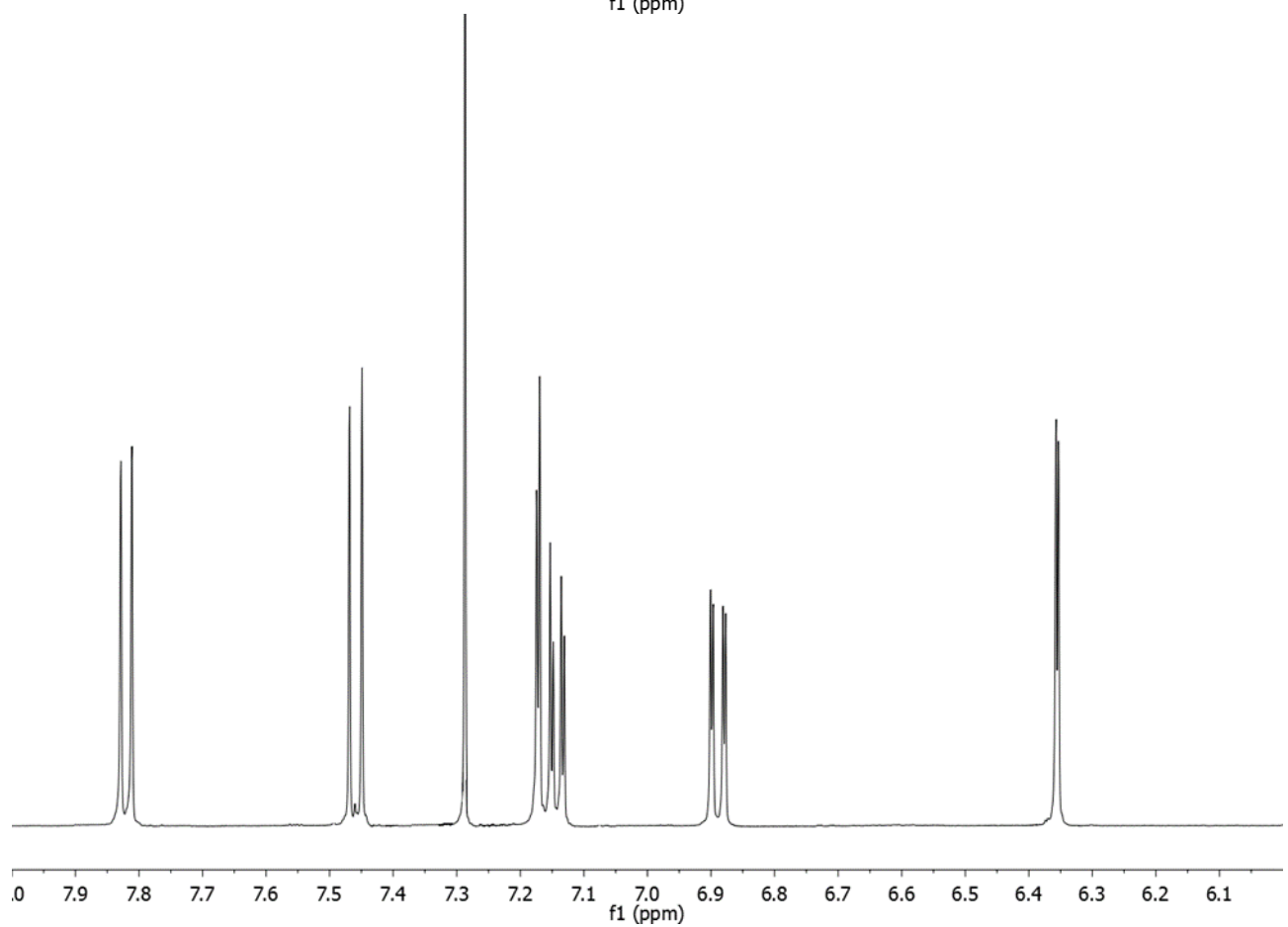
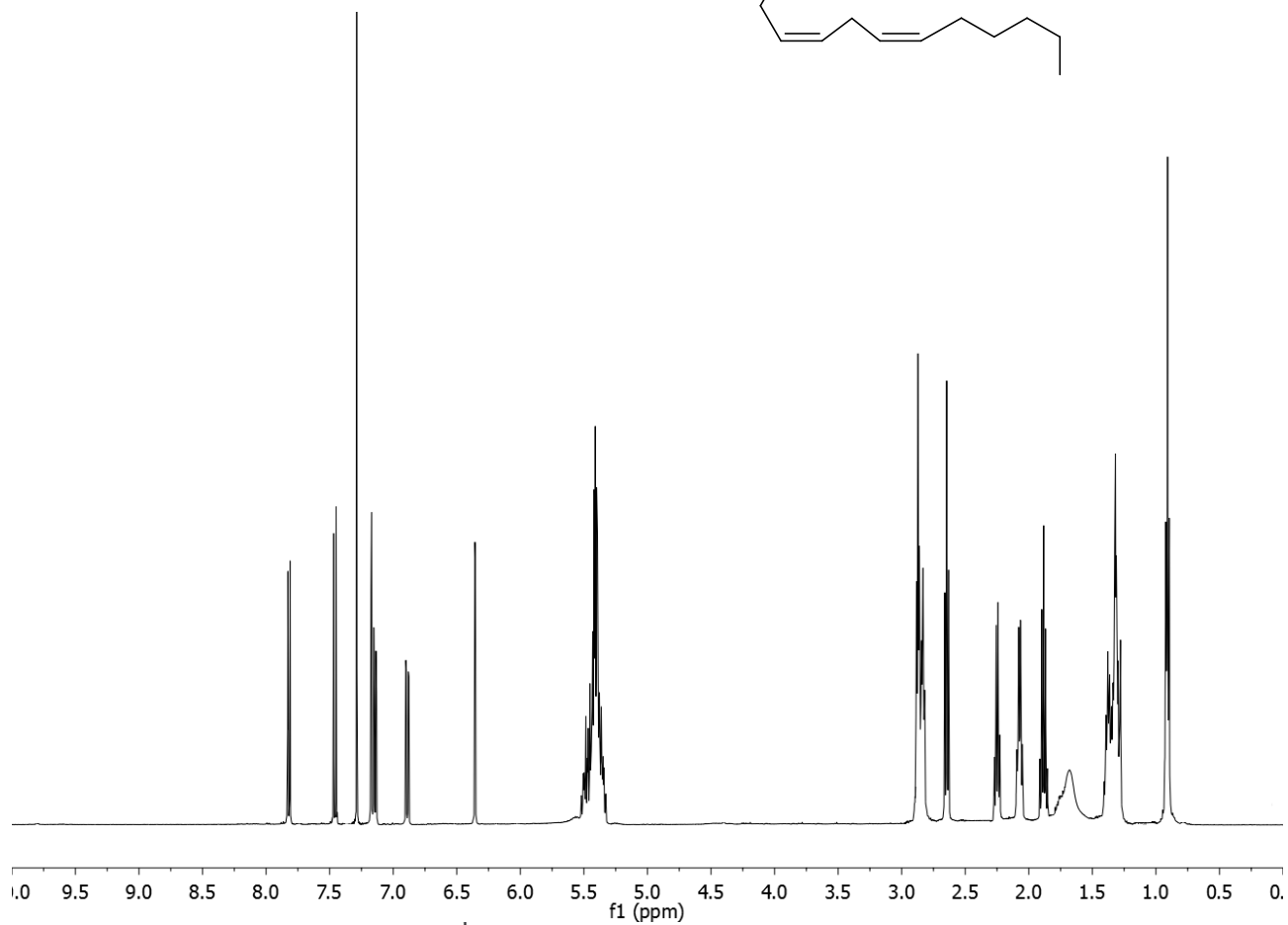
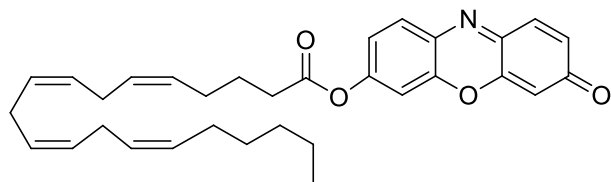


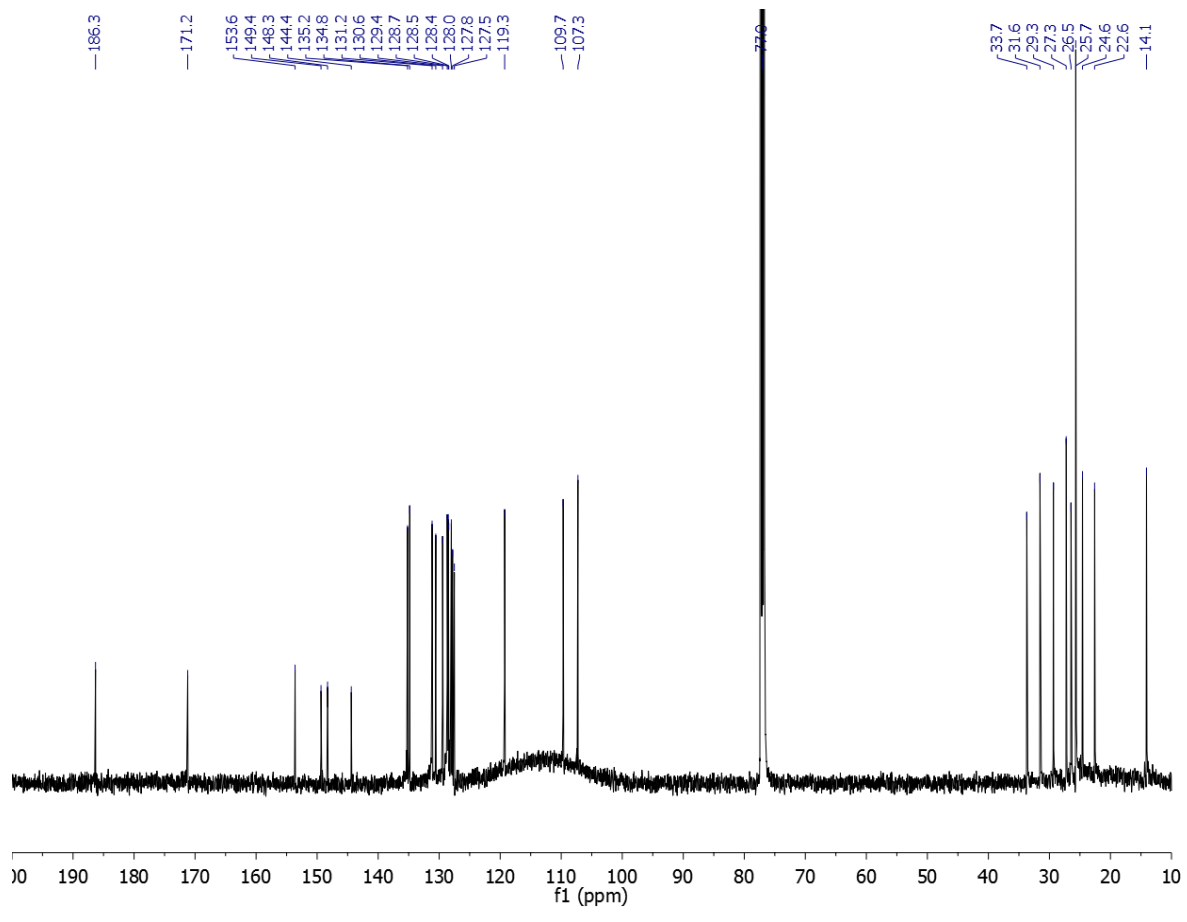
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra  
of 7-hydroxyresorufinyl-oleate (**1f**)





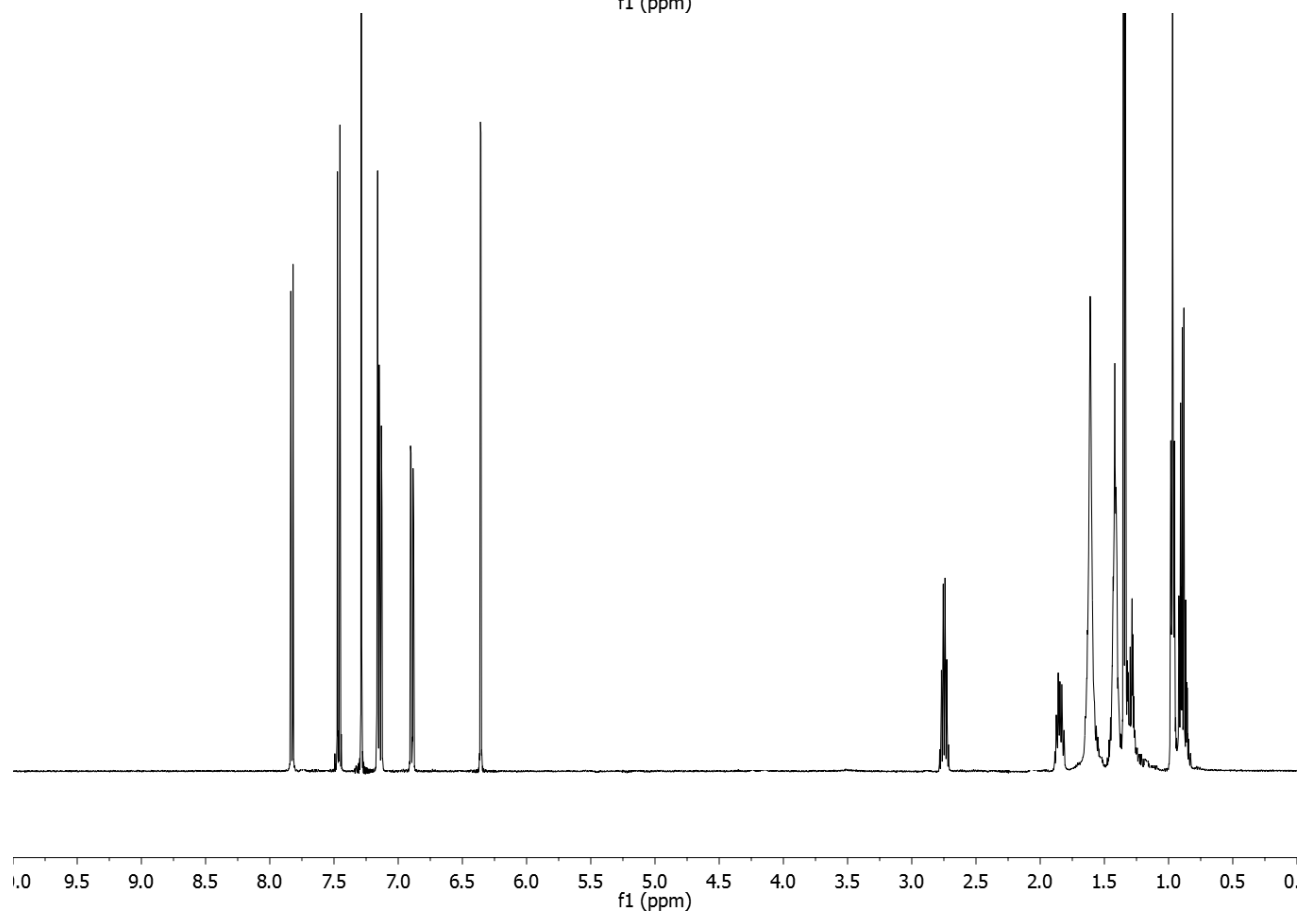
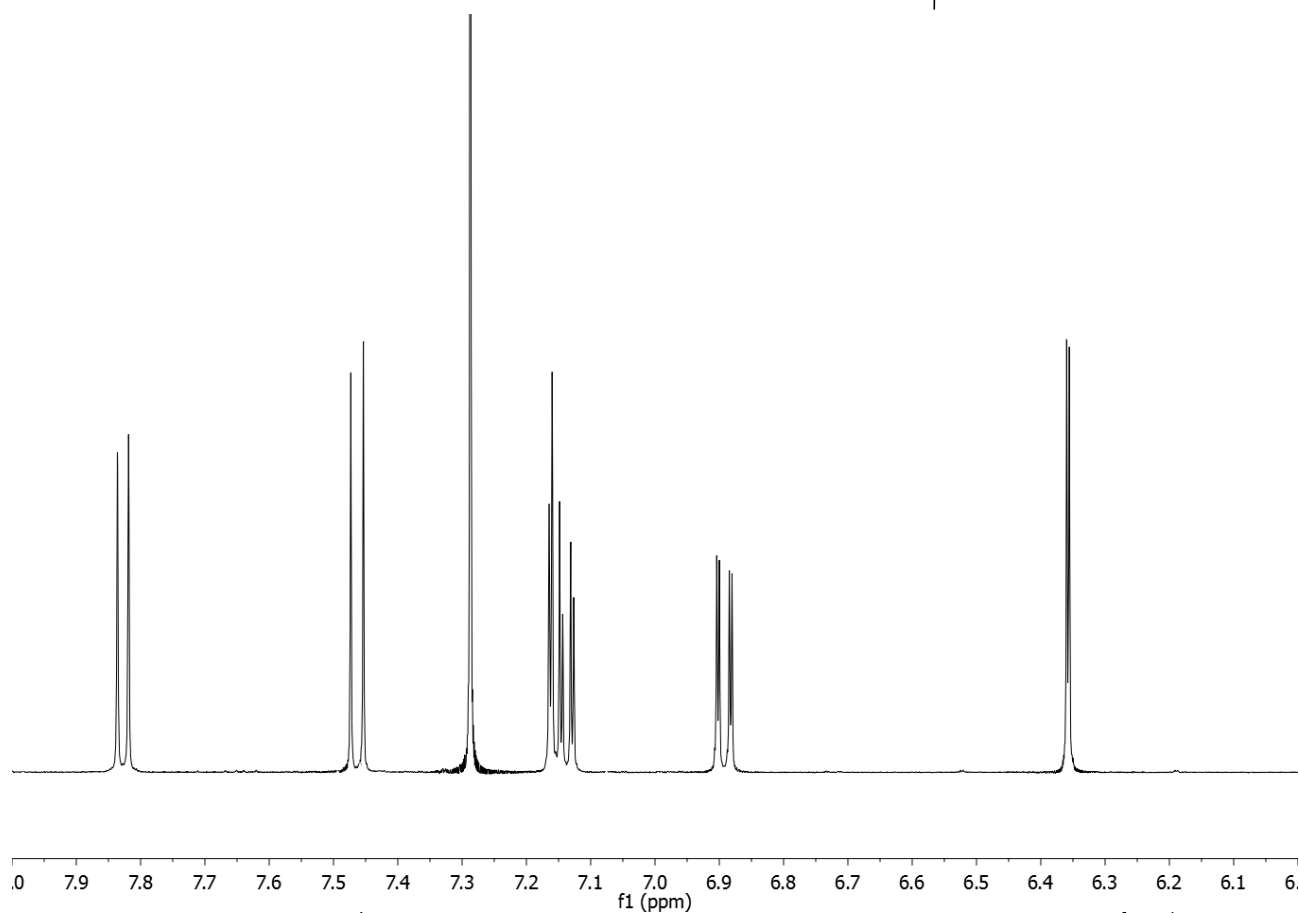
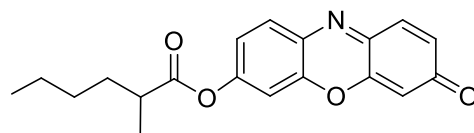
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-arachidonate (**1g**)

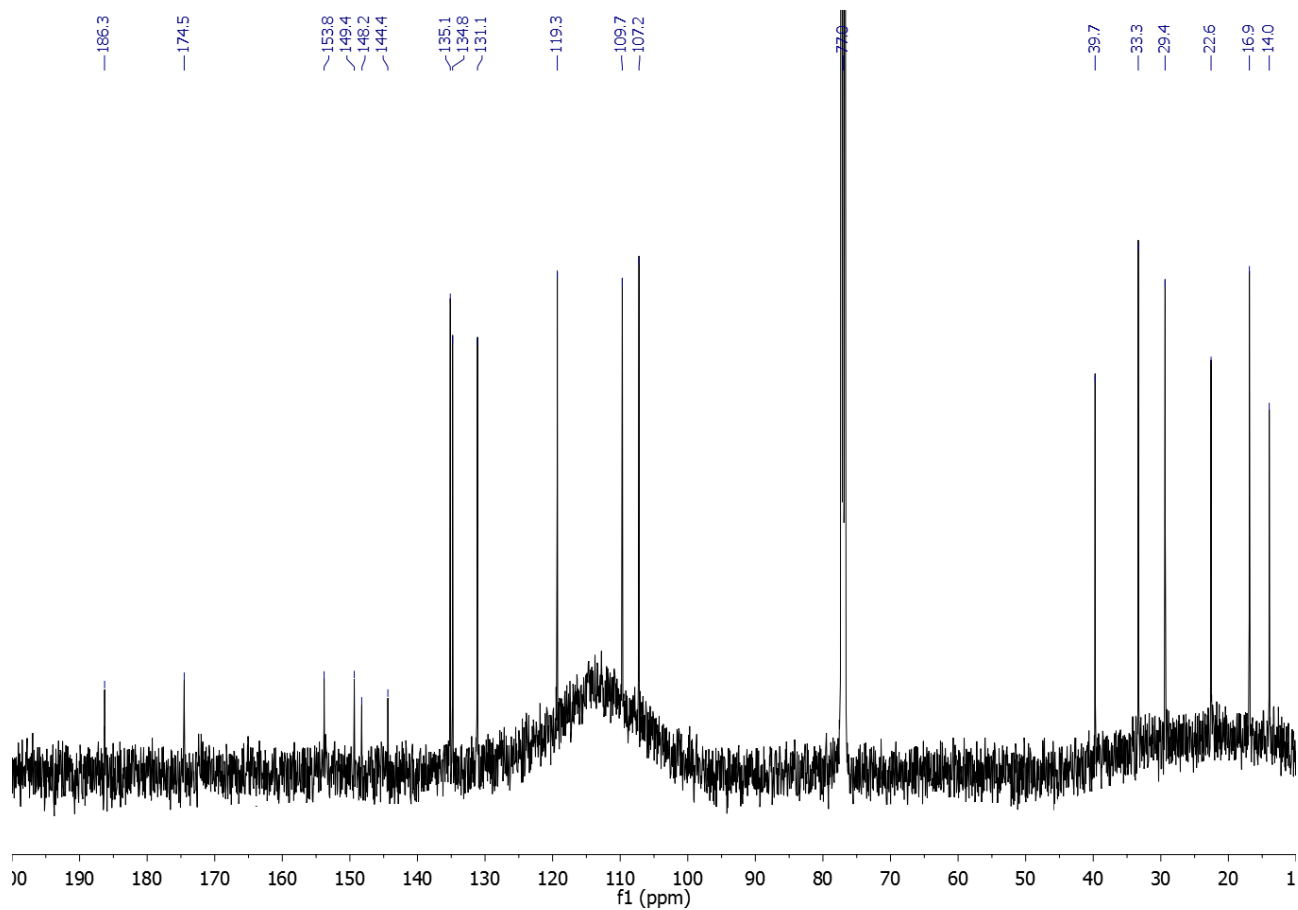




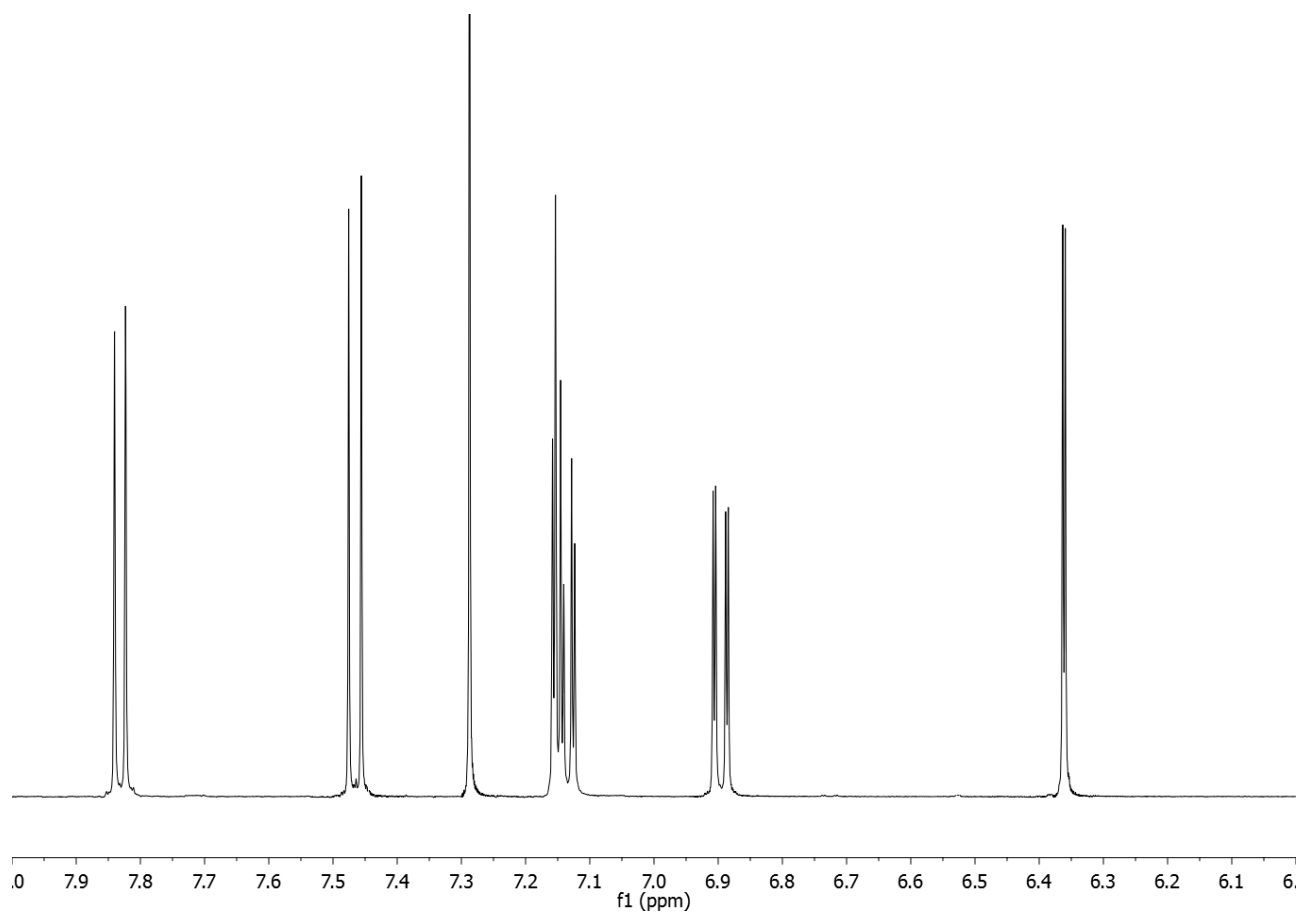
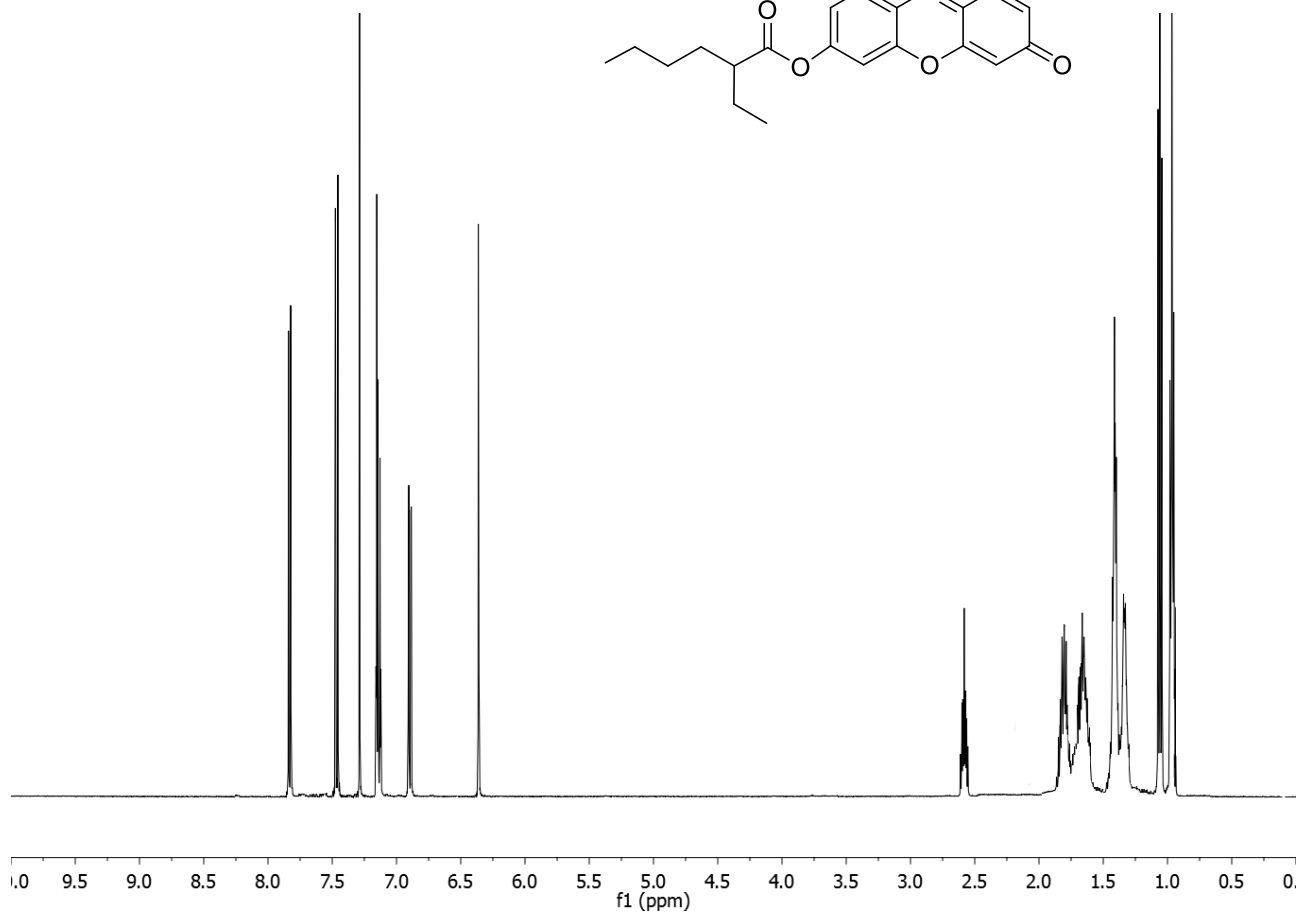
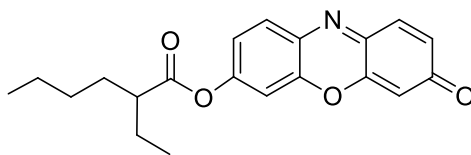


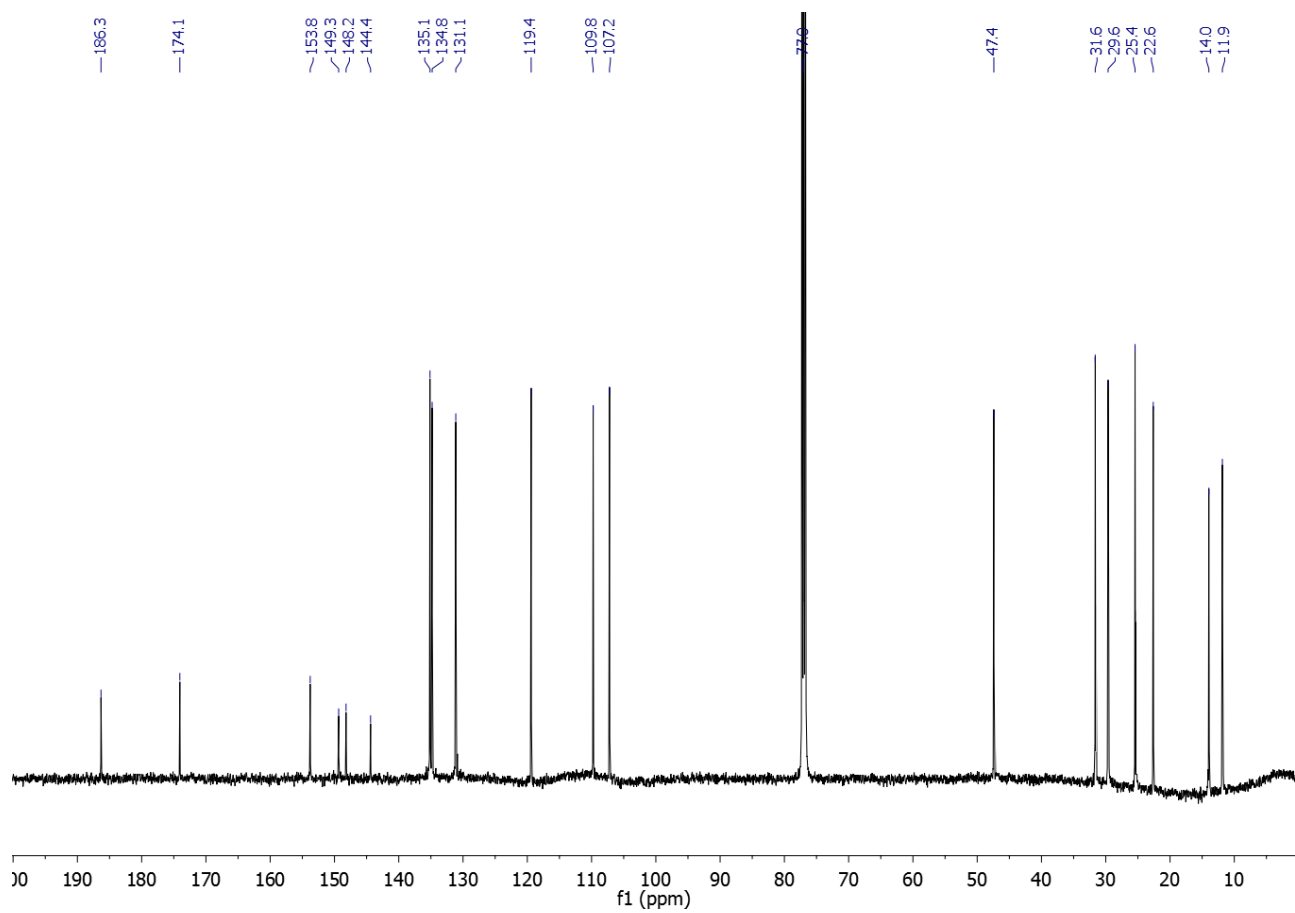
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-2-methylhexanoate (**1h**)



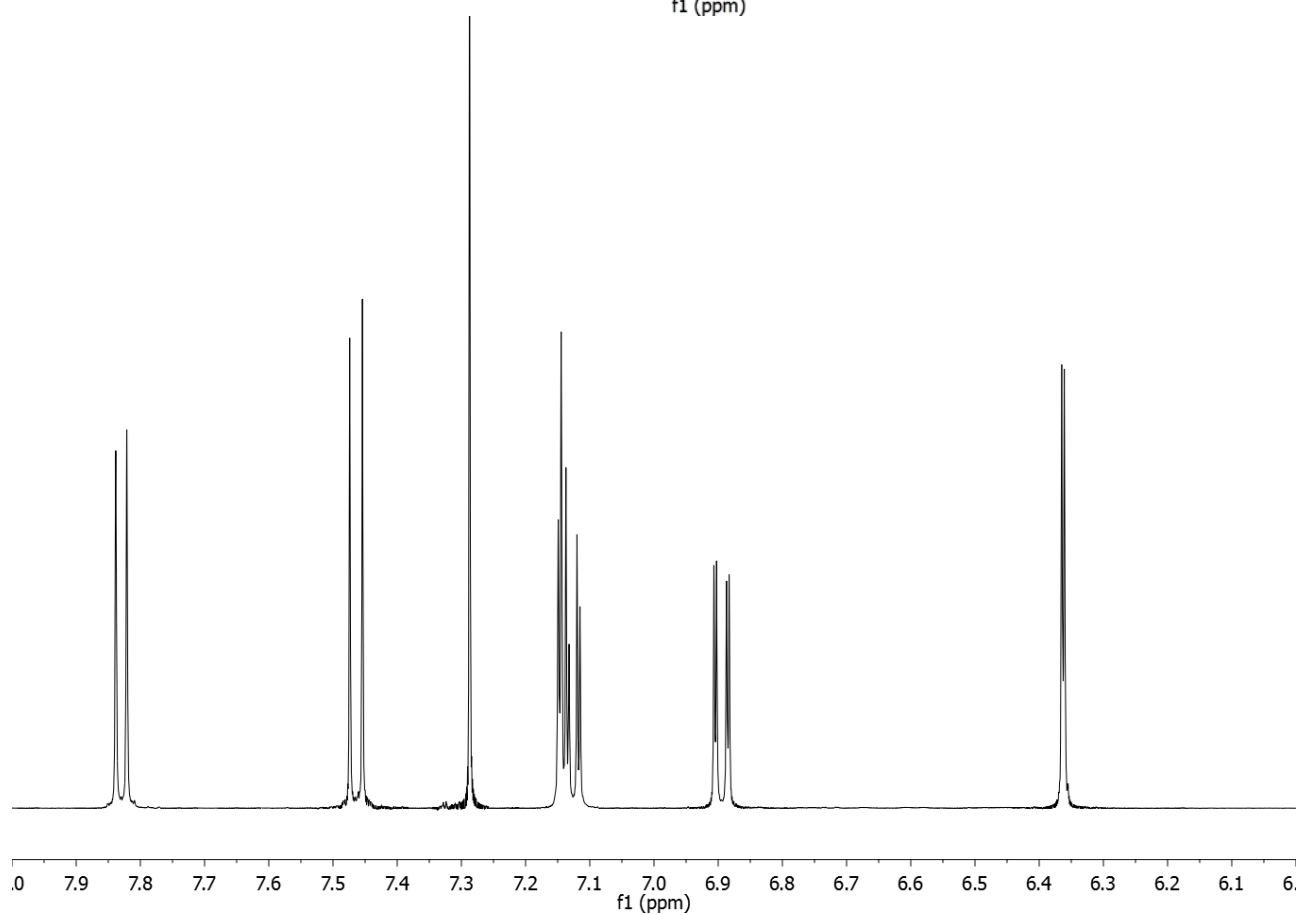
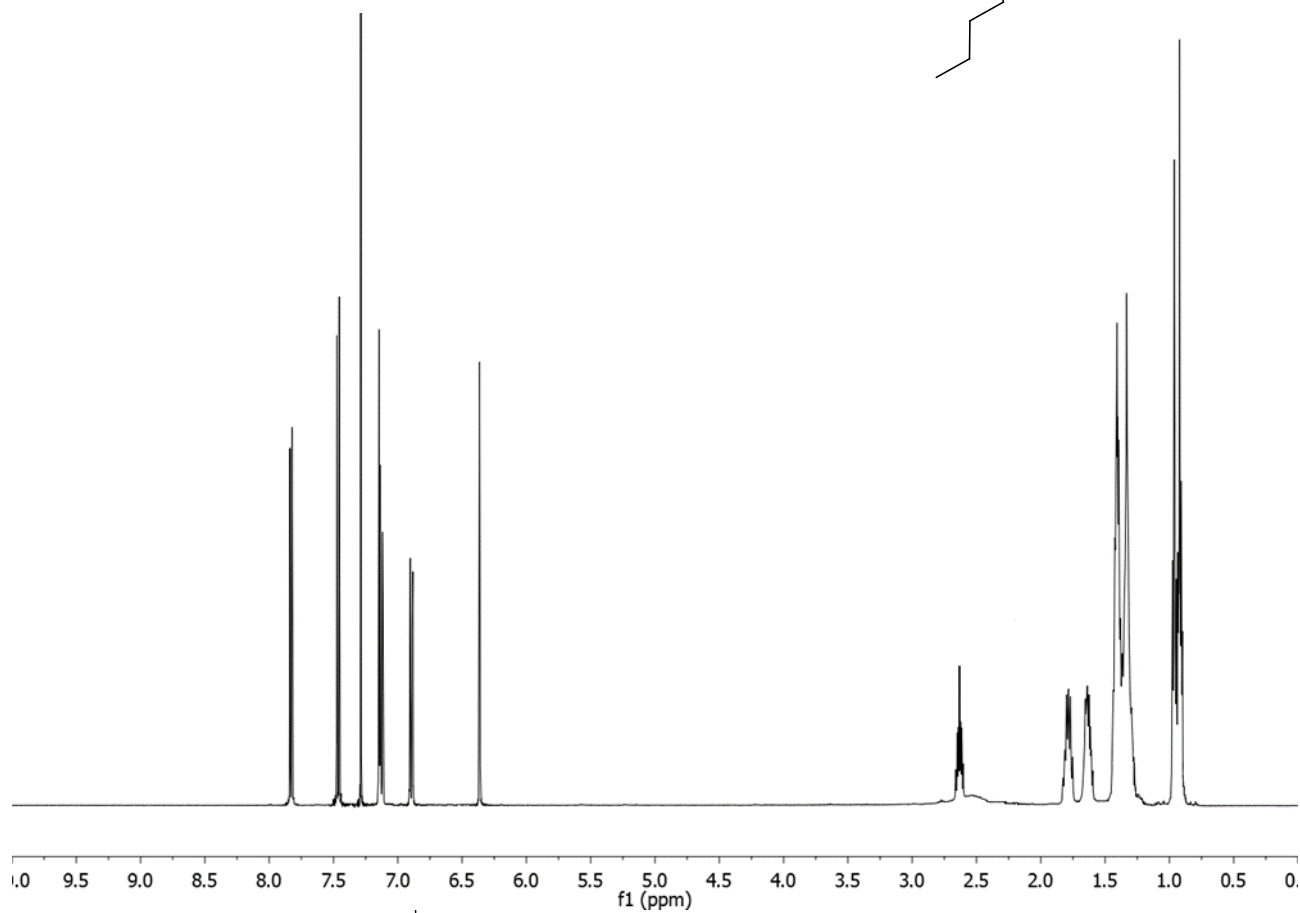
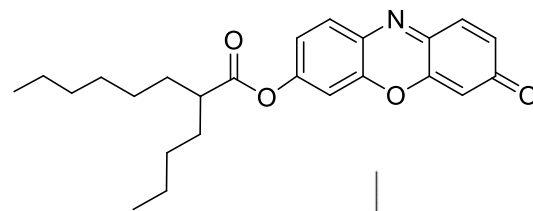


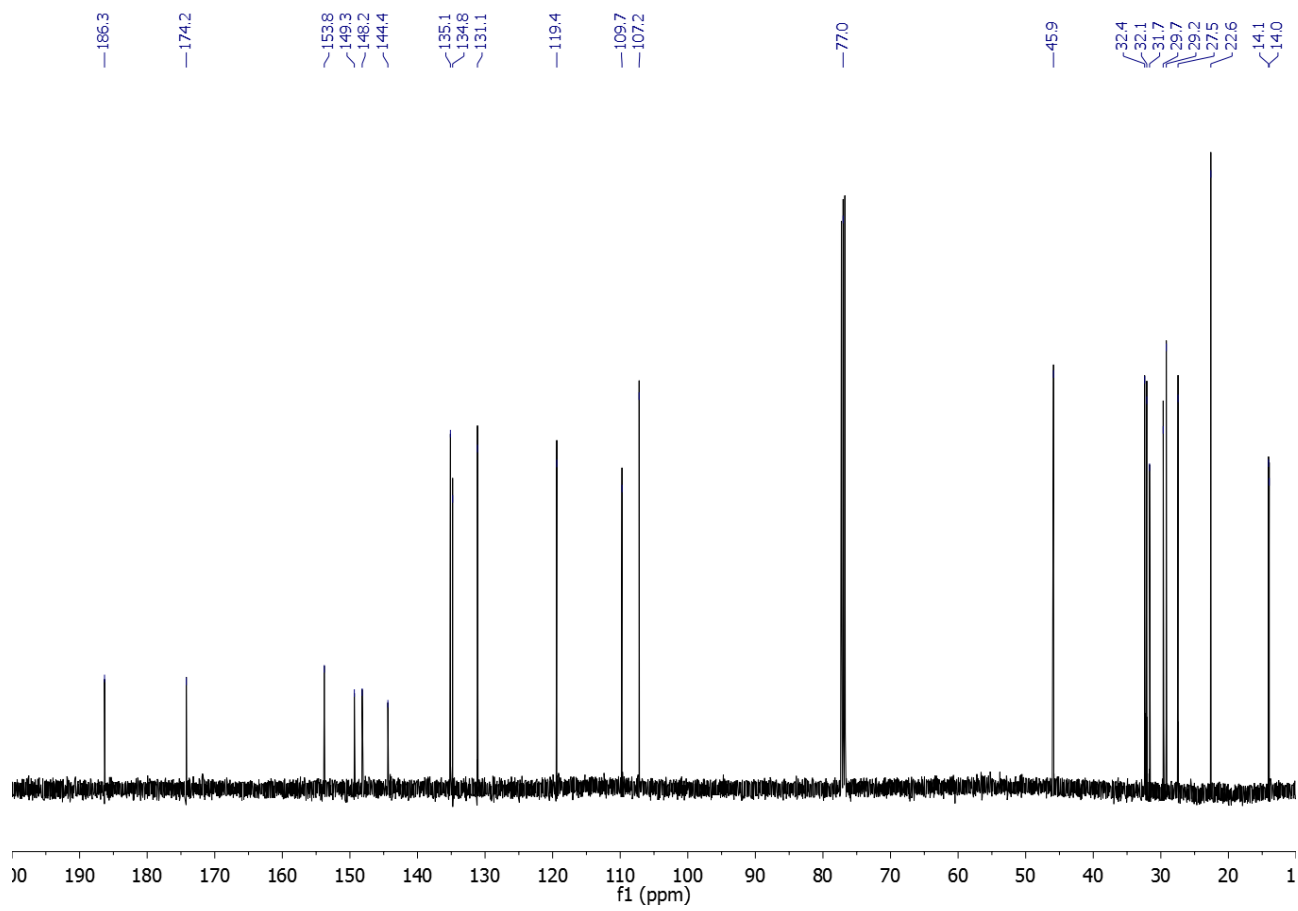
$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-2-ethylhexanoate (**1i**)





$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra  
of 7-hydroxyresorufinyl-2-butyloctanoate (**1j**)





$^1\text{H}$  and  $^{13}\text{C}$  NMR spectra of 7-hydroxyresorufinyl-2-benzoate (**1k**)

