Supplementary Table 1. Primers used in this study

Purpose	Gene	Primer sequence (5' to 3')
quantitative RT-PCR	*FLC*	TCCGGCAAGCTCTACAGCTT GCTGTTTCCCATATCGATCAAGGA
	FT	CTGCTACAACTGGAACAACCTTTG TTTGCCTGCCAAGCTGTCGAA
	SOC1	ATGAATTCGCCAGCTCCAATATGC TGGTGCTGACTCGATCCTTAGTA
	TSF	CACTGGAAATGCCTTTGGCAATGA CTTCCGAGTTGCCGGAACAA
	CO	CTCACTACAACGACAATGGTTCCA CTTGGGTGTGAAGCTGTTGTGA
	SVP	TTCTGTAGCTCCAGCATGAAGGA AGATGGCTGATCAAGCTTCTCCAA
	LHP1	GCTCACAATCCTCATCTCCTGATT CCTACAAGGGCTGTGTACAAACA
	TEM1	GAGCCGGTGATGTGGTTTGT GACCGGACTTTCCAGTGGATATA
	MYB33	GCACGTATGGCTGCACATTTG GCTCGTTGTCGCCTCTTGATA
	ELF6	AGGAGAGACAGATGCTCAGGAAAT GAGGCAGAAAATGCGAGGTCTTA
semi-quantitative RT-PCR	*ANAC075* full	ATGAACAAGAGTAATCCTGCTG TTACCCATGATGATCTTGGTTGTC
	ANAC075 downstream	GGAGGGAAACTCCTCAGACT GGTAGAAGAAGTGCAGCCCATTA
	ACT2	ACATTGTGCTCAGTGGTGGA GAGATCCACATCTGCTGGAAT
Promoter amplification (for fusion with GUS)	*ANAC075*	TAACAAGGCGCGCCGGAAATGGGATAAAACGGAGCTCAAAT ATTCCTGGATCCATCTCAATCTCGAATATCTTTGATCA
CDS amplification without a stop codon (for entry clone construction)	*ANAC075*	GGGGACAAGTTTGTACAAAAAAGCAGGCTCCATGAACAAGAGTAATCCTGCTGGTTCGG GGGGACCACTTTGTACAAGAAAGCTGGGTCCCCATGATGATCTTGGTTGTCAGAAGAGT
	ANAC052	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGGTCGCGAATCTGTGGCTGTTG GGGGACCACTTTGTACAAGAAAGCTGGGTTTTGTCCATTAGCATTGTTCTTCTTG
	JMJ14	GGGGACAAGTTTGTACAAAAAAGCAGGCTTCATGGATCAGCTTGCATCTCTAGCAGAGT GGGGACCACTTTGTACAAGAAAGCTGGGTTAGGACTTATCTCCATCTTATCAACCAAAGT