

## **Online Supplement**

### **Supplemental Methods**

#### **Reagents and cells**

Purified batroxobin, plasminogen, and factor XIIIa (fXIIIa) were obtained from Enzyme Research Laboratories (ERL). The chemotactic agent C5a was obtained from R&D systems. The RAW264.7 cell line was obtained from the ATCC and was used in low passage numbers (i.e., less than 10 passages). Transwell migration inserts with 5  $\mu$ M pores were from Corning. Murine fibrinogen was purified from citrate-plasma by ammonium sulfate precipitation as described.<sup>1</sup> Aliquots were confirmed to be free of endotoxin contamination prior to use via ToxinSensor Chromogenic LAL assay (GenScript; minimum detection limit, 0.005 EU/mL). Primary bone marrow-derived macrophages (BMDMs) were generated from wildtype mice as described.<sup>2</sup>

#### **Immunohistochemistry**

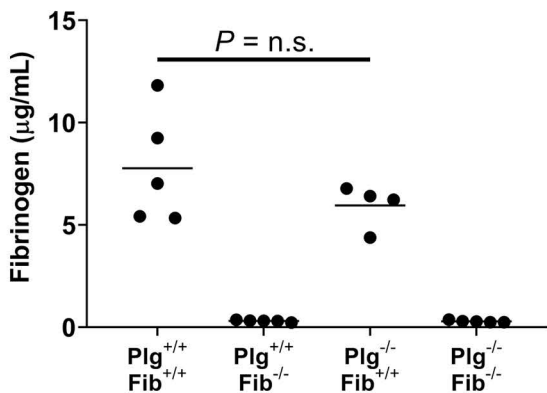
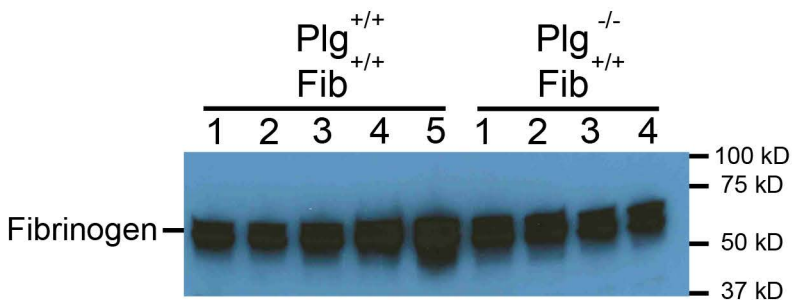
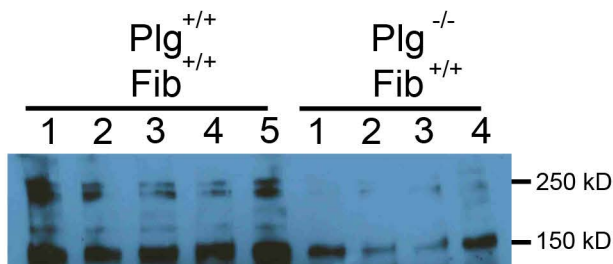
Immunohistochemistry was performed as in fibrin immunofluorescence with slight modifications. Sections were incubated with rat anti-F4/80 antibody (BM8, ThermoFisher Scientific MF48000, 1:500) O/N at 4°C and washed in PBS (3 times). Secondary antibody (Biotinylated goat anti-rat IgG, Vector Laboratories Inc. BA-9400, 1:500) was added for 1 hour at room temperature. Sections were then washed, developed, dehydrated and #1.5 cover slips mounted with ProLong™ Gold Antifade Reagent (Thermo Fisher Scientific). Images were captured on an Olympus BX43 Clinical upright microscope (40x) using Olympus DP74 color camera. A fixed square area was selected around the tissue and the color deconvolution plugin within Image J (NIH) was used to measure DAB/Hematoxylin ratio within the region. Three to five individual fields were measured per tissue and mean values were calculated in Microsoft® Excel for Mac (version 16.15).

1. Prasad JM, Gorkun OV, Raghu H, et al. Mice expressing a mutant form of fibrinogen that cannot support fibrin formation exhibit compromised antimicrobial host defense. *Blood*. 2015;126(17):2047-2058.
2. Ryu JK, Petersen MA, Murray SG, et al. Blood coagulation protein fibrinogen promotes autoimmunity and demyelination via chemokine release and antigen presentation. *Nat Commun*. 2015;6:8164.

## Supplemental Figure Legends

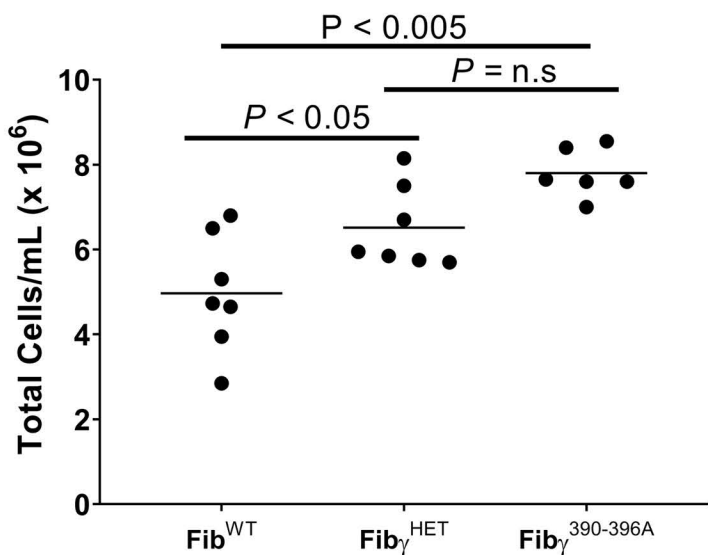
Supplemental Figure 1. **Diminished high-molecular weight fibrin species in Plg<sup>-</sup> animals following thioglycollate-induced peritonitis.** (A) Fibrinogen ELISA reveals similar levels of fibrinogen in the peritoneal lavage fluid of both Plg<sup>+</sup> and Plg<sup>-</sup> animals after thioglycollate-induced peritonitis. (B) Western blot analysis, under reducing conditions, of the peritoneal lavage fluid also confirms similar fibrinogen level between Plg<sup>+</sup> and Plg<sup>-</sup> mice. (C) Interestingly, increased high-molecular weight fibrin species was observed in the lavage fluid from Plg<sup>+</sup> mice compared to Plg<sup>-</sup> animals.

Supplemental Figure 2. **Gene dosage effect on macrophage migration in Fiby<sup>390-396A</sup> animals.** Total cells (A) and macrophages (B) were assessed following thioglycollate-induced peritonitis in Fib<sup>WT</sup>, Fiby<sup>HET</sup>, and Fiby<sup>390-396A</sup> mice. Fiby<sup>HET</sup> animals demonstrated an intermediate phenotype between the Fib<sup>WT</sup> and Fiby<sup>390-396A</sup> mice. Both Fiby<sup>HET</sup> and Fiby<sup>390-396A</sup> mice had a statistically significant increase in macrophage migration compared to Fib<sup>WT</sup> animals.

**A****Fibrinogen  
Peritoneal Fluid****B****C**

# A

## Total Cell Counts Peritoneal Lavage



# B

## Macrophages Peritoneal Lavage

