

Supporting Information

Pd^{II}-Catalyzed Enantioselective C(sp³)-H Activation/Cross-Coupling Reactions of Free Carboxylic Acids

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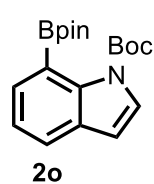
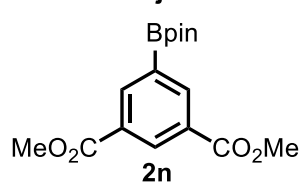
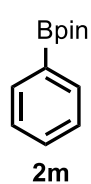
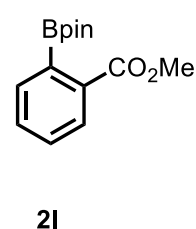
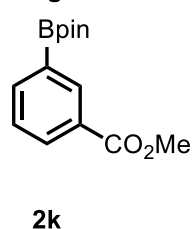
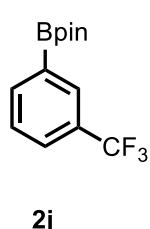
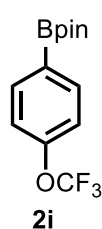
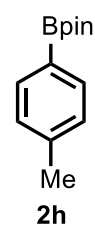
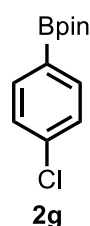
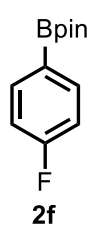
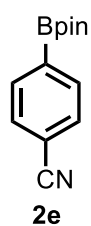
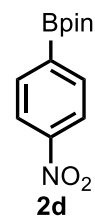
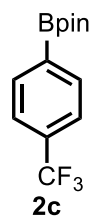
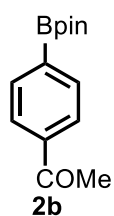
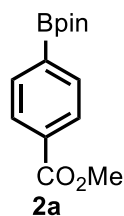
1. General Information

Carboxylic acids were obtained from the commercial sources or synthesized following literature procedures. Alkyl iodides were obtained from the commercial sources. Solvents were obtained from Sigma-Aldrich, Oakwood and Acros and used directly without further purification. Analytical thin layer chromatography was performed on 0.25 mm silica gel 60-F254. Visualization was carried out with UV light and Bromocresol Green Stain. ^1H NMR was recorded on Bruker DRX-600 instrument (600 MHz). Chemical shifts were quoted in parts per million (ppm) referenced to the literature values of tetramethylsilane. The following abbreviations (or combinations thereof) were used to explain multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, m = multiplet, br = broad. Coupling constants, J , were reported in Hertz unit (Hz). ^{13}C NMR spectra were recorded on Bruker DRX-600 instrument (150 MHz), and were fully decoupled by broad band proton decoupling. Chemical shifts were reported in ppm referenced to either the center line of a triplet at 77.0 ppm of chloroform- d or the center line of a multiplet at 29.84 ppm of acetone- d^6 . High-resolution mass spectra (HRMS) were recorded on an Agilent Mass spectrometer using ESI-TOF (electrospray ionization-time of flight). Enantiomeric ratio (er) were determined on a Agilent SFC system or Waters SFC system using commercially available chiral columns.

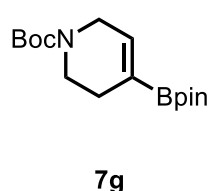
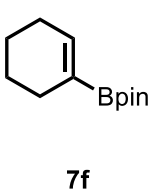
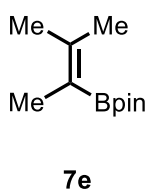
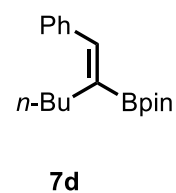
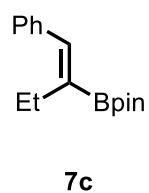
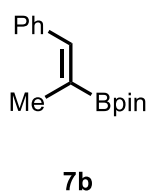
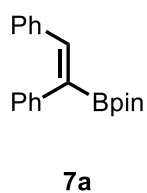
2. Substrate Structures

Substrates **1a**, **1p**, **4a**, **4q**, **4u** were purchased from commercial sources. **1q-1u**, **4p**, **4r-4t** was synthesized from the reported procedure.^{1,5,14}

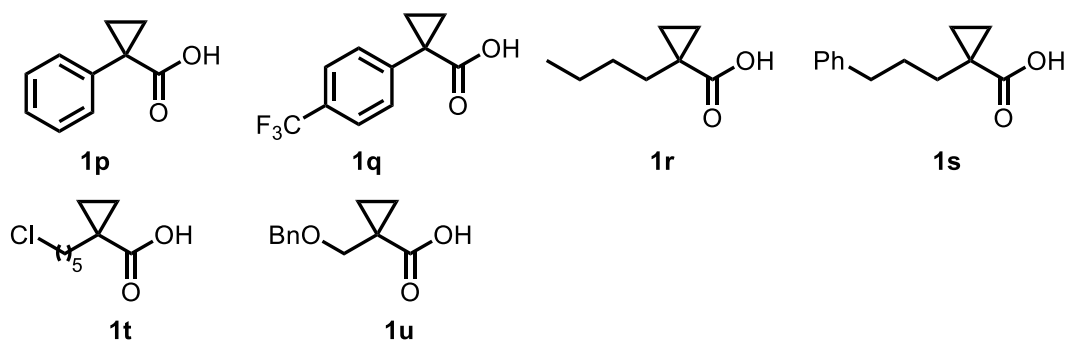
Arylboronic acid pinacol ester



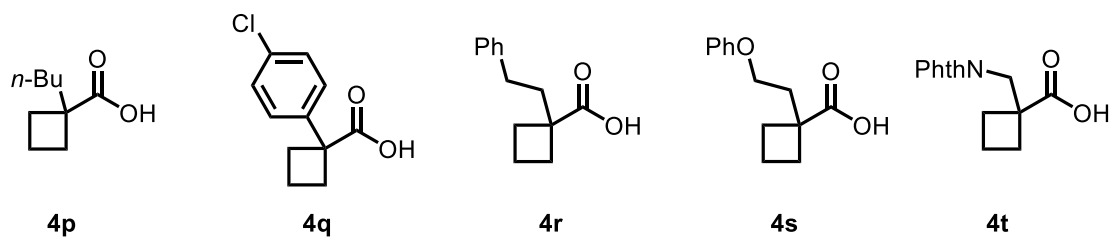
Vinylboronic acid pinacol esters



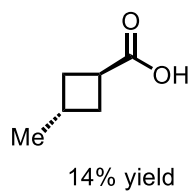
α -Substituted cyclopropanecarboxylic acids



α -Substituted cyclobutanecarboxylic acids

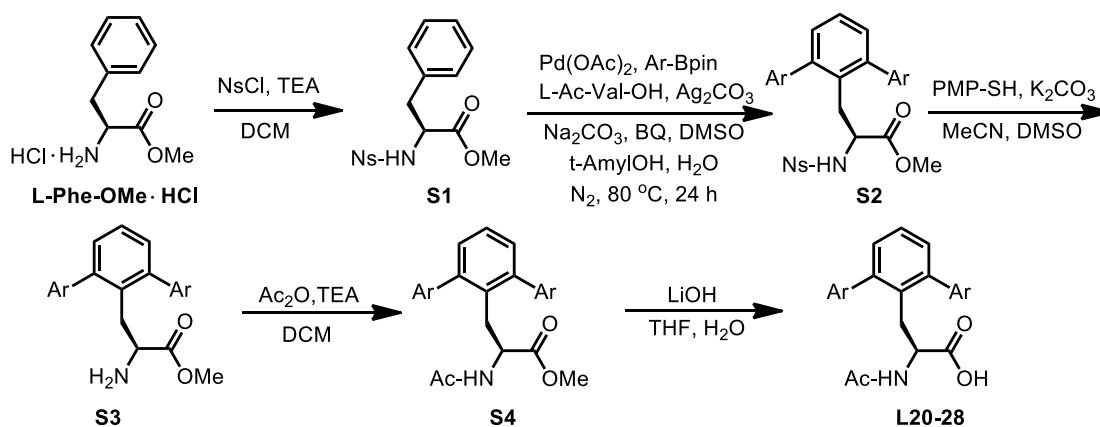


Unsuccessful substrate



3. Experimental Section

3.1.1 General Procedure for the Preparation of Ligands L20-28



4-Nitrobenzenesulfonyl chloride (NsCl, 50 mmol, 1.0 equiv) was added to a cooled (0 °C) solution of L-phenylalanine methyl ester hydrochloride (L-Phe-OMe·HCl, 50 mmol, 1.0 equiv) and triethylamine (TEA, 150 mmol, 3.0 equiv) in DCM (150 mL). After being stirred at room temperature for 12 h, the reaction mixture was poured into H₂O. The organic layer was separated, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum. The residue was purified by trituration with a mixture of 50% DCM/hexane to give **S1**.

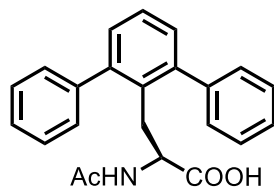
S1 (8.0 mmol, 1.0 equiv), Pd(OAc)₂ (0.075 equiv), Ar-BPin (2.0 equiv), L-Ac-Val-OH (0.2 equiv), Ag₂CO₃ (2.0 equiv), Na₂CO₃ (2.0 equiv), BQ (0.5 equiv), H₂O (5.0 equiv), and DMSO (0.4 equiv) were weighed in air and placed in a Schlenk tube with a magnetic stir bar. *t*-AmylOH (50 mL) was added, and the reaction vessel was evacuated and backfilled with nitrogen (×3). The reaction mixture was heated to 80 °C for 24 h under vigorous stirring. After being cooled to room temperature, the reaction mixture was diluted with EtOAc and filtered through a pad of Celite eluting with EtOAc. The filtrate was concentrated under vacuum and the resulting residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane = 1: 3) to give **S2**.

4-Methoxybenzenethiol (PMP-SH, 20.0 mmol, 4.0 equiv) and potassium carbonate (20.0 mmol, 4.0 equiv) were added to a solution of **S2** (5.0 mmol, 1.0 equiv) in MeCN (40 mL) and DMSO (1.5 mL). After being stirred at room temperature for 12 h, the reaction mixture was diluted with EtOAc, washed with H₂O and brine, dried over anhydrous Na₂SO₄, filtered, and concentrated under vacuum. The residue was purified by column chromatography on silica gel (eluent: EtOAc/hexane = 1: 1) to give **S3**.

Ac₂O (12.0 mmol, 3.0 equiv) was added to a solution of **S3** (4.0 mmol, 1.0 equiv) and triethylamine (12.0 mmol, 3.0 equiv) in DCM (20 mL). After being stirred at room temperature for 3 h, the reaction mixture was quenched with saturated NH₄Cl solution and extracted with DCM. The combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated to give **S4**, which could be used directly in the next step without further purification.

LiOH (8.0 mmol) was added to a suspension of **S4** in THF (8 mL), and H₂O (4 mL) at 0 °C. The reaction mixture was allowed to warm to room temperature and stirred for 8 h. The reaction mixture was quenched with 10% aqueous citric acid solution and extracted EtOAc (3

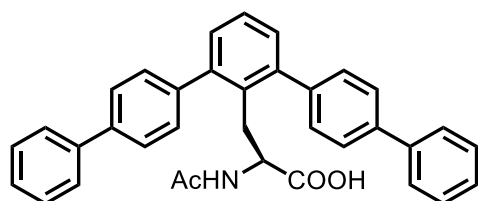
x 20 mL). The combined organic layers were washed with brine, dried over anhydrous Na_2SO_4 , filtered and concentrated to give the desired ligand, further purification could be conducted by recrystallization or reversed phase flash column.



(S)-3-([1,1':3',1''-terphenyl]-2'-yl)-2-acetamidopropanoic acid (L20)

^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 11.92 (s, 1H), 7.47 – 7.40 (m, 4H), 7.39 – 7.33 (m, 7H), 7.30 (t, $J = 7.6$ Hz, 1H), 7.11 (d, $J = 7.6$ Hz, 2H), 3.66 (q, $J = 7.7$ Hz, 1H), 3.13 (dd, $J = 14.1$, 7.5 Hz, 1H), 3.01 (dd, $J = 14.1$, 8.1 Hz, 1H), 1.55 (s, 3H); ^{13}C NMR (150 MHz, $\text{DMSO}-d_6$) δ 172.52, 168.26, 143.23, 141.67, 132.30, 129.32, 129.21, 128.19, 126.91, 126.13, 51.59, 31.04, 22.12.

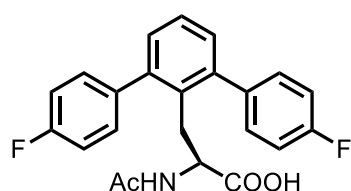
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{23}\text{H}_{22}\text{NO}_3^+$ $[\text{M}+\text{H}]^+$ 360.1600, found 360.1601.



(S)-3-([1,1':4',1'':3'',1''':4''',1''''-quinquephenyl]-2'-yl)-2-acetamidopropanoic acid (L21)

^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 11.99 (s, 1H), 7.83 – 7.66 (m, 8H), 7.53 – 7.45 (m, 9H), 7.41 – 7.37 (m, 2H), 7.35 (t, $J = 7.6$ Hz, 1H), 7.19 (d, $J = 7.6$ Hz, 2H), 3.80 (q, $J = 7.7$ Hz, 1H), 3.26 (dd, $J = 14.1$, 7.4 Hz, 1H), 3.11 (dd, $J = 14.1$, 8.2 Hz, 1H), 1.56 (s, 3H); ^{13}C NMR (150 MHz, $\text{DMSO}-d_6$) δ 172.60, 168.32, 142.90, 140.87, 139.72, 138.49, 132.51, 129.89, 129.41, 128.99, 127.47, 126.57, 126.41, 126.29, 51.64, 31.33, 22.15.

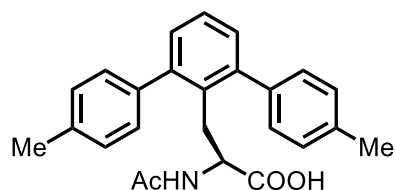
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{35}\text{H}_{30}\text{NO}_3^+$ $[\text{M}+\text{H}]^+$ 512.2226, found 512.2228.



(S)-2-acetamido-3-(4,4''-difluoro-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L22)

^1H NMR (600 MHz, $\text{DMSO}-d_6$) δ 12.02 (s, 1H), 7.50 (d, $J = 8.0$ Hz, 1H), 7.40 – 7.36 (m, 4H), 7.30 (t, $J = 7.6$ Hz, 1H), 7.25 (d, $J = 8.8$ Hz, 4H), 7.12 (d, $J = 7.6$ Hz, 2H), 3.70 (q, $J = 7.8$ Hz, 1H), 3.11 (dd, $J = 14.1$, 7.3 Hz, 1H), 2.96 (dd, $J = 14.2$, 8.2 Hz, 1H), 1.57 (s, 3H); ^{13}C NMR (150 MHz, $\text{DMSO}-d_6$) δ 172.51, 168.32, 161.34 (d, $J = 241.5$ Hz), 42.21, 137.87 (d, $J = 3.0$ Hz), 132.75, 131.20 (d, $J = 9.0$ Hz), 129.52, 126.22, 115.03 (d, $J = 21.0$ Hz), 51.36, 31.21, 22.15; ^{19}F NMR (376 MHz, DMSO) δ -116.14.

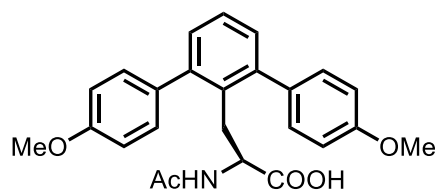
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{23}\text{H}_{20}\text{F}_2\text{NO}_3^+$ $[\text{M}+\text{H}]^+$ 369.1411, found 369.1409.



(S)-2-acetamido-3-(4,4''-dimethyl-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L23)

$^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) δ 11.92 (s, 1H), 7.33 (d, $J = 7.6$ Hz, 1H), 7.23 (s, 9H), 7.06 (d, $J = 7.6$ Hz, 2H), 3.69 (q, $J = 7.8$ Hz, 1H), 3.14 (dd, $J = 14.0, 7.5$ Hz, 1H), 3.04 – 2.99 (m, 1H), 2.35 (s, 6H), 1.56 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, $\text{DMSO-}d_6$) δ 172.57, 168.34, 143.20, 138.87, 135.86, 132.34, 129.25, 129.11, 128.80, 126.06, 51.62, 31.06, 22.14, 20.80.

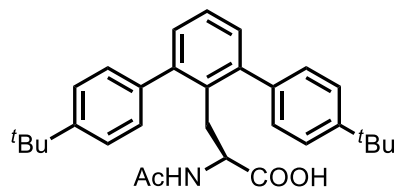
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{25}\text{H}_{26}\text{NO}_3^+$ $[\text{M}+\text{H}]^+$ 388.1913, found 388.1914.



(S)-2-acetamido-3-(4,4''-dimethoxy-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L24)

$^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) δ 11.93 (s, 1H), 7.32 (d, $J = 7.6$ Hz, 1H), 7.29 – 7.26 (m, 4H), 7.24 (d, $J = 7.6$ Hz, 1H), 7.06 (d, $J = 7.5$ Hz, 2H), 7.00 – 6.96 (m, 4H), 3.79 (s, 6H), 3.70 (q, $J = 7.7$ Hz, 1H), 3.15 (dd, $J = 14.0, 7.3$ Hz, 1H), 3.01 (dd, $J = 14.0, 8.4$ Hz, 1H), 1.57 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, $\text{DMSO-}d_6$) δ 172.63, 168.30, 158.12, 142.91, 134.05, 132.71, 130.32, 129.25, 126.01, 113.62, 55.06, 51.57, 31.12, 22.17.

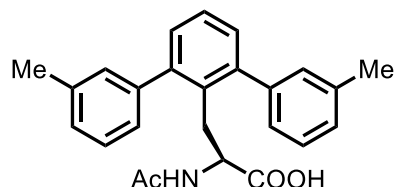
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{25}\text{H}_{26}\text{NO}_5^+$ $[\text{M}+\text{H}]^+$ 420.1811, found 420.1818.



(S)-2-acetamido-3-(4,4''-di-tert-butyl-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L25)

$^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) δ 11.89 (s, 1H), 7.46 – 7.41 (m, 4H), 7.35 (d, $J = 7.4$ Hz, 1H), 7.30 – 7.26 (m, 5H), 7.09 (d, $J = 7.6$ Hz, 2H), 3.65 (q, $J = 7.6$ Hz, 1H), 3.14 (dd, $J = 14.1, 7.6$ Hz, 1H), 3.00 (dd, $J = 14.1, 7.7$ Hz, 1H), 1.54 (s, 3H), 1.32 (s, 18H); $^{13}\text{C NMR}$ (150 MHz, $\text{DMSO-}d_6$) δ 172.75, 168.09, 149.01, 143.09, 138.81, 132.59, 129.26, 128.88, 126.08, 124.92, 51.63, 40.07, 34.25, 31.19, 22.11.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{31}\text{H}_{38}\text{NO}_3^+$ $[\text{M}+\text{H}]^+$ 472.2852, found 472.2847.

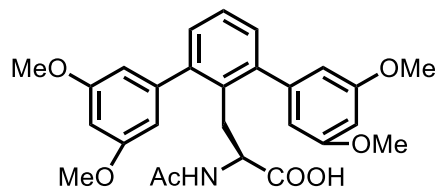


(S)-2-acetamido-3-(3,3''-dimethyl-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L26)

$^1\text{H NMR}$ (600 MHz, $\text{DMSO-}d_6$) δ 11.93 (s, 1H), 7.36 – 7.26 (m, 4H), 7.18 – 7.12 (m, 6H), 7.08 (d, $J = 7.6$ Hz, 2H), 3.70 (q, $J = 7.7$ Hz, 1H), 3.13 (dd, $J = 14.0, 7.3$ Hz, 1H), 2.99 (dd, $J = 14.1, 8.2$ Hz, 1H), 2.35 (s, 6H), 1.56 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, $\text{DMSO-}d_6$) δ 172.59,

168.27, 143.28, 141.66, 137.19, 132.31, 129.86, 129.18, 128.06, 127.53, 126.29, 125.99, 51.72, 31.13, 22.14, 21.13.

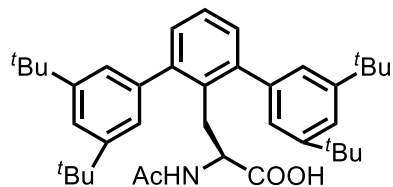
HRMS (ESI-TOF) m/z Calcd for $C_{25}H_{26}NO_3^+$ $[M+H]^+$ 388.1913, found 388.1914.



(S)-2-acetamido-3-(3,3',5,5''-tetramethoxy-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L27)

1H NMR (600 MHz, $DMSO-d_6$) δ 11.99 (s, 1H), 7.40 (d, $J = 7.7$ Hz, 1H), 7.26 (dd, $J = 7.9, 7.2$ Hz, 1H), 7.11 (d, $J = 7.6$ Hz, 2H), 6.50 – 6.48 (m, 6H), 3.89 – 3.84 (m, 1H), 3.77 (s, 12H), 3.20 (dd, $J = 14.1, 7.1$ Hz, 1H), 3.04 (dd, $J = 14.1, 8.6$ Hz, 1H), 1.60 (s, 3H); ^{13}C NMR (150 MHz, $DMSO-d_6$) δ 172.70, 168.39, 160.07, 143.63, 143.08, 132.32, 129.06, 125.91, 107.48, 99.05, 55.21, 51.62, 31.36, 22.15.

HRMS (ESI-TOF) m/z Calcd for $C_{27}H_{30}NO_7^+$ $[M+H]^+$ 480.2022, found 480.2026.

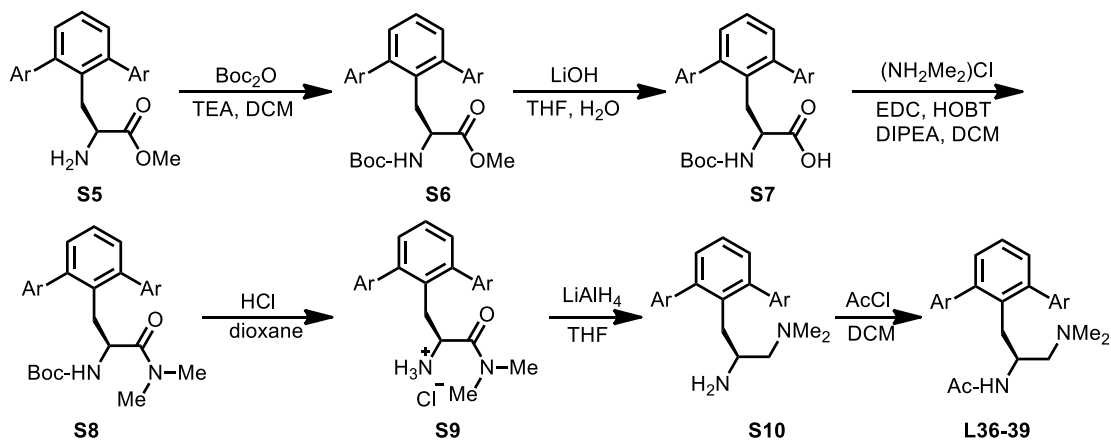


(S)-2-acetamido-3-(3,3',5,5''-tetra-tert-butyl-[1,1':3',1''-terphenyl]-2'-yl)propanoic acid (L28)

1H NMR (600 MHz, $CDCl_3$) δ 7.44 (t, $J = 1.8$ Hz, 2H), 7.35 (dd, $J = 8.1, 7.0$ Hz, 1H), 7.28 – 7.26 (m, 2H), 7.21 (d, $J = 1.9$ Hz, 4H), 5.12 (d, $J = 7.2$ Hz, 1H), 4.07 (ddd, $J = 11.9, 7.1, 3.4$ Hz, 1H), 3.28 (dd, $J = 14.6, 3.5$ Hz, 1H), 3.14 (dd, $J = 14.6, 11.8$ Hz, 1H), 1.75 (s, 3H), 1.36 (s, 36H); ^{13}C NMR (150 MHz, $CDCl_3$) δ 172.81, 170.97, 150.76, 143.93, 140.08, 130.99, 129.35, 126.32, 120.92, 52.75, 34.54, 31.06, 29.33, 22.44.

HRMS (ESI-TOF) m/z Calcd for $C_{39}H_{54}NO_3^+$ $[M+H]^+$ 584.4104, found 584.4114.

3.1.2 General Procedure for the Preparation of Ligands L36-39



Boc_2O (8.0 mmol, 2.0 equiv) was added to a solution of **S5** (4.0 mmol, 1.0 equiv) and triethylamine (8.0 mmol, 2.0 equiv) in DCM (20 mL). After being stirred at room temperature for 3 h, the reaction mixture was quenched with saturated NH_4Cl solution and extracted with

DCM. The combined organic layers were dried over anhydrous Na_2SO_4 , filtered and concentrated to give **S6**, which could be used directly in the next step without further purification.

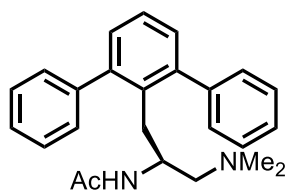
LiOH (8.0 mmol) was added to a suspension of **S6** in THF (8 mL), and H_2O (4 mL) at 0 °C. The reaction mixture was allowed to warm to room temperature and stirred for 8 h. The reaction mixture was quenched with 10% aqueous citric acid solution and extracted EtOAc (3 x 20 mL). The combined organic layers were washed with brine, dried over anhydrous Na_2SO_4 , filtered and concentrated to give **S7**, further purification could be conducted by recrystallization or reversed phase flash column.

The corresponding Boc-protected amino acid (**S7**) (4 mmol), dimethylammonium chloride (8.8 mmol, 2.2 equiv) and benzotriazol-1-ol hydrate (HOBt) (4 mmol, 1.0 equiv) were added to a round bottom flask equipped with a magnetic stir bar. The solid mixture was dissolved in DCM (40 mL), and 1-ethyl-(3-(3-dimethylamino)propyl)-carbodiimide hydrochloride (EDC) (4.8 mmol, 1.2 equiv) was added at 0 °C. The resulting solution was stirred at 0 °C as N-ethyl-N,N-diisopropylamine (DIPEA) (9.6 mmol, 2.4 equiv) was added slowly. The reaction solution was allowed to warm to r.t. and stirred for about 3 h, after which the solution was poured into a separatory funnel, diluted to 150 mL with additional DCM, and washed with approximately 25 mL of 10% w/w aqueous citric acid. The organic layer was separated and subsequently washed with 25 mL each of saturated aqueous NaHCO_3 and brine. The organics were dried over anhydrous Na_2SO_4 , filtered, and concentrated in vacuo to provide corresponding amide (**S8**) which could be directly used in the next step without further purification.

To the Boc-protected amino amide (**S8**) was added 4 N HCl/dioxane solution (4 ml). The resulting solution was stirred at room temperature for 2 h. Then, the volatile components were evaporated in vacuo, and the residue was subsequently used in the following reduction step.

To a solution of **S9** in THF (24 ml) was added a solution of LiAlH_4 in THF (6.0 mmol, 1.5 equiv) dropwise under N_2 at 0 °C. Then, the mixture was heated to reflux for 12 h, before being cooled down and diluted with ether. The mixture was cooled to 0 °C, and 0.28 ml of water was added slowly followed by 15% w/w NaOH aqueous solution (0.28 ml) and water (0.84 ml). The resulting suspension was then warmed to room temperature and stirred for 15 min before MgSO_4 was added. The mixture was stirred for additional 15 min before filtration. The filtrate was collected, and the solvent was removed in vacuo to provide diamine compound (**S10**) which could be used in next step without purification.

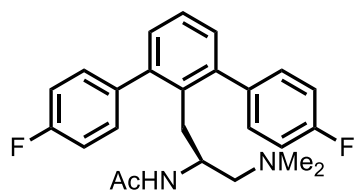
To a solution of the synthesized diamine compound (**S10**) in DCM (8 ml) was added acetyl chloride (8 mmol, 2.0 equiv) at 0 °C. Then the solution was stirred at room temperature for 2 h. The volatile components were evaporated in vacuo, and the residue was dissolved in 8 ml of water. The resulting solution was extracted with ether (10 ml x 3), then the aqueous phase was alkalinized with 15% w/w NaOH aqueous solution until $\text{pH} > 13$. The alkalinized mixture was extracted with ether (10 ml x 3), and the organic layers were concentrated to provide the desired MPAAM Ligand. Further purification could be conducted by recrystallization or reversed phase flash column.



(S)-N-(1-([1,1':3',1''-terphenyl]-2'-yl)-3-(dimethylamino)propan-2-yl)acetamide (L36)

^1H NMR (600 MHz, CDCl_3) δ 7.46 – 7.40 (m, 8H), 7.38 – 7.35 (m, 2H), 7.29 – 7.27 (m, 1H), 7.20 – 7.17 (m, 2H), 4.60 (d, $J = 8.7$ Hz, 1H), 3.74 (dd, $J = 10.0, 5.8$ Hz, 1H), 3.12 (dd, $J = 14.2, 4.2$ Hz, 1H), 2.80 (dd, $J = 14.2, 10.7$ Hz, 1H), 1.85 (dd, $J = 12.0, 7.0$ Hz, 1H), 1.72 (s, 6H), 1.69 (s, 3H), 1.61 (dd, $J = 12.0, 7.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 169.00, 143.33, 142.40, 133.44, 129.76, 129.59, 128.36, 127.02, 126.03, 63.99, 48.06, 45.20, 32.38, 23.41.

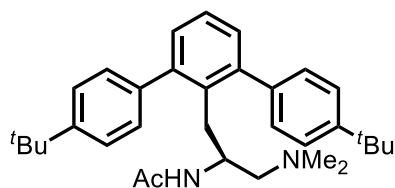
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{25}\text{H}_{29}\text{N}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ 373.2280, found 373.2282.



(S)-N-(1-(4,4''-difluoro-[1,1':3',1''-terphenyl]-2'-yl)-3-(dimethylamino)propan-2-yl)acetamide (L37)

^1H NMR (600 MHz, CDCl_3) δ 7.37 (dd, $J = 8.5, 5.5$ Hz, 4H), 7.29 – 7.27 (m, 1H), 7.17 – 7.12 (m, 5H), 4.74 (d, $J = 8.8$ Hz, 1H), 3.78 – 3.71 (m, 1H), 3.03 (dd, $J = 14.2, 4.2$ Hz, 1H), 2.77 (dd, $J = 14.2, 10.4$ Hz, 1H), 1.92 (dd, $J = 12.1, 7.5$ Hz, 1H), 1.80 (s, 6H), 1.70 (s, 3H), 1.67 – 1.62 (m, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 169.05, 162.04 (d, $J = 246.0$ Hz), 142.36, 138.22 (d, $J = 3.0$ Hz), 133.58, 131.27 (d, $J = 7.5$ Hz), 129.86, 126.19, 115.27 (d, $J = 21.0$ Hz), 63.66, 47.68, 45.05, 32.71, 23.34; ^{19}F NMR (376 MHz, CDCl_3) δ -115.70.

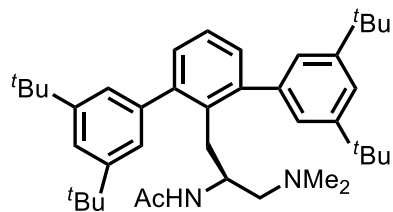
HRMS (ESI-TOF) m/z Calcd for $\text{C}_{25}\text{H}_{26}\text{F}_2\text{N}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ 409.2091, found 409.2096.



(S)-N-(1-(4,4''-di-tert-butyl-[1,1':3',1''-terphenyl]-2'-yl)-3-(dimethylamino)propan-2-yl)acetamide (L38)

^1H NMR (600 MHz, CDCl_3) δ 7.48 – 7.44 (m, 4H), 7.36 – 7.32 (m, 4H), 7.25 – 7.23 (m, 1H), 7.18 – 7.15 (m, 2H), 4.59 (d, $J = 9.0$ Hz, 1H), 3.80 – 3.72 (m, 1H), 3.22 (dd, $J = 14.2, 3.4$ Hz, 1H), 2.70 (dd, $J = 14.3, 11.2$ Hz, 1H), 1.89 (dd, $J = 11.9, 6.3$ Hz, 1H), 1.69 (s, 6H), 1.68 (s, 3H), 1.65 – 1.60 (m, 1H), 1.37 (s, 18H); ^{13}C NMR (150 MHz, CDCl_3) δ 168.50, 149.38, 142.77, 138.97, 133.30, 129.10, 129.00, 125.52, 124.83, 63.89, 47.81, 44.79, 34.12, 31.64, 30.96, 22.97.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{33}\text{H}_{45}\text{N}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ 485.3532, found 485.3529.



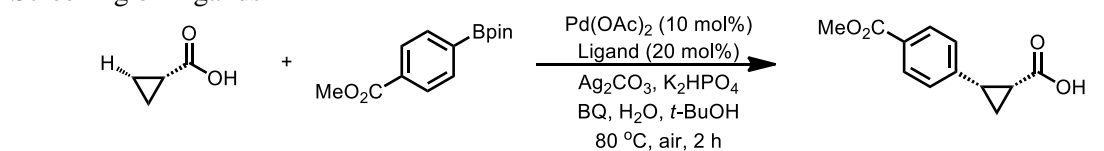
(S)-N-(1-(dimethylamino)-3-(3,3',5,5'-tetra-tert-butyl-[1,1':3',1''-terphenyl]-2'-yl)propa-2-yl)acetamide (L39)

¹H NMR (600 MHz, CDCl₃) δ 7.42 (t, *J* = 1.8 Hz, 2H), 7.30 – 7.26 (m, 5H), 7.20 (d, *J* = 7.5 Hz, 2H), 4.72 (d, *J* = 9.2 Hz, 1H), 3.87 – 3.81 (m, 1H), 3.14 (dd, *J* = 14.2, 3.8 Hz, 1H), 2.80 (dd, *J* = 14.1, 11.1 Hz, 1H), 1.93 (d, *J* = 7.0 Hz, 1H), 1.72 (s, 6H), 1.69 (s, 3H), 1.66 – 1.62 (m, 1H), 1.38 (s, 36H); ¹³C NMR (150 MHz, CDCl₃) δ 168.91, 150.86, 150.71, 144.30, 141.51, 133.34, 129.63, 125.94, 120.89, 64.25, 47.49, 45.04, 34.97, 32.40, 31.58, 23.52.

HRMS (ESI-TOF) *m/z* Calcd for C₄₁H₆₁N₂O⁺ [M+H]⁺ 597.4784, found 597.4791.

3.2 Optimization of Conditions of Arylation for Cyclopropanecarboxylic Acid

Screening of Ligands^{a,b}



No Ligand				
11% yield	38% yield, 96:4 er	11% yield	9% yield	3% yield
13% yield	8% yield	N.R.	N.R.	N.R.
N.R.	23% yield, 96:4 er	17% yield	19% yield	14% yield
7% yield	22% yield, 86:14 er	14% yield	34% yield, 95:5 er	15% yield

^a Conditions: **1a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), Ligand (20 mol%), Ag₂CO₃ (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (10.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

Screening of Ag Salts and Pd source^{a,b}

1a			2a			3a		
Ag Salt	Yield	er	Pd source	Yield	er			
no change	70%	97:3	PdCl ₂	16%	-			
AgOAc	43%	96:4	Pd(TFA) ₂	37%	96:4			
AgOPiv	10%	-	Pd(MeCN) ₄ (BF ₄) ₂	34%	94:6			
AgTFA	NR	-	Pd(MeCN) ₂ Cl ₂	35%	93:7			
AgF	8%	-	Pd(PhCN) ₂ Cl ₂	30%	94:6			
Ag ₂ O	31%	93:7	[Pd(allyl)Cl] ₂	23%	92:8			
			Pd(pph ₃) ₂ Cl ₂	20%	91:9			

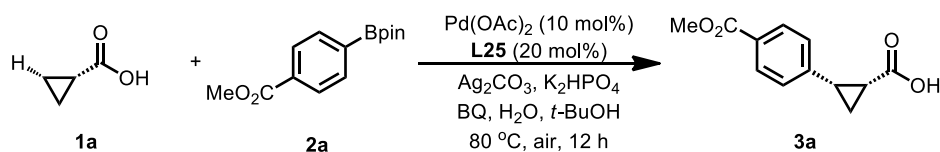
^a Conditions: **1a** (0.1 mmol), **2a** (1.5 equiv), Pd (10 mol%), **L25** (20 mol%), Ag (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (10.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

Screening of Solvent and Temperature^{a,b}

Entry	Solvent	Temperature	Yield	er
1	<i>t</i> -BuOH	80 °C	70%	97:3
2	<i>t</i> -AmylOH	80 °C	56%	94:6
3	THF	80 °C	16%	-
4	Dioxane	80 °C	19%	-
5	MeCN	80 °C	N.R.	-
6	HFIP	80 °C	21%	89:11
7	DMF	80 °C	N.R.	-
8	<i>t</i> -BuOH	70 °C	61%	97:3
9	<i>t</i> -BuOH	90 °C	71%	90:10

^a Conditions: **1a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), **L25** (20 mol%), Ag₂CO₃ (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (10.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

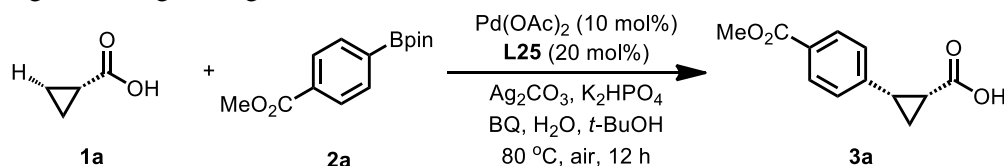
Screening of Bases^{a,b}



Base	Yield	er	Base	Yield	er
None	3%	-	LiF	NR	-
NaOAc	11%	-	KOAc	32%	94:6
NaHCO ₃	9%	-	KHCO ₃	12%	-
Na ₂ CO ₃	23%	94:6	K ₂ CO ₃	29%	94:6
NaH ₂ PO ₄	NR	-	KH ₂ PO ₄	18%	-
Na ₂ HPO ₄	5%	-	K ₂ HPO ₄ ·3H ₂ O	45%	93:7
Na ₃ PO ₄	18%	-	K ₃ PO ₄	47%	96:4
Li ₂ CO ₃	NR	-	KF	11%	-
LiOAc	NR	-	Cs ₂ CO ₃	26%	95:5
Li ₃ PO ₄	NR	-	CsOAc	11%	-

^a Conditions: **1a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), **L25** (20 mol%), Ag₂CO₃ (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (10.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

Screening of loading of reagents^{a,b}

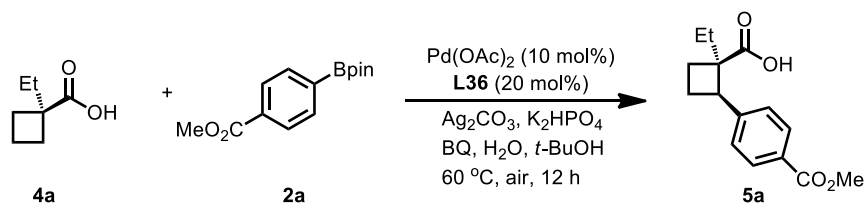


Change	Yield	Base	Yield
No Change	70%	Ag ₂ CO ₃ (1.0 equiv)	57%
K ₂ HPO ₄ (1.0 equiv)	61%	Ag ₂ CO ₃ (2.0 equiv)	68%
K ₂ HPO ₄ (2.0 equiv)	67%	ArBpin (1.0 equiv)	57%
K ₂ HPO ₄ (3.0 equiv)	46%	ArBpin (2.0 equiv)	70%

^a Conditions: **1a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), **L25** (20 mol%), Ag₂CO₃ (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (10.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

3.3 Optimization of Conditions of Arylation for Cyclobutanecarboxylic Acid

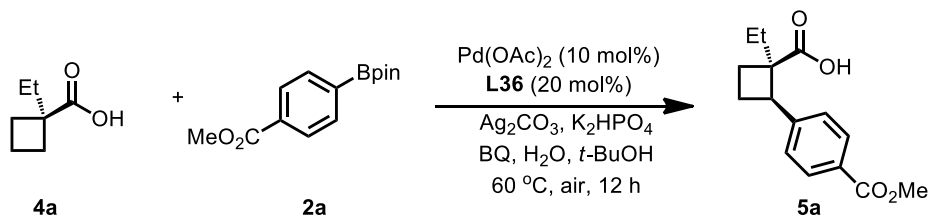
Screening of Solvent and Temperature^{a,b}



Entry	Solvent	Temperature	Yield	er
1	<i>t</i> -BuOH	60 °C	62%	94:6
2	<i>t</i> -AmylOH	60 °C	38%	85:15
3	THF	60 °C	23%	86:14
4	Dioxane	60 °C	12%	-
5	MeCN	60 °C	N.R.	-
6	HFIP	60 °C	32%	64:36
7	DMF	60 °C	N.R.	-
8	<i>t</i> -BuOH	80 °C	60%	90:10
9	<i>t</i> -BuOH	50 °C	46%	94:6

^a Conditions: **4a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), **L36** (20 mol%), Ag₂CO₃ (2.0 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (5.0 equiv), *t*-BuOH (1.0 mL), 60 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

Screening of loading of reagents^{a,b}

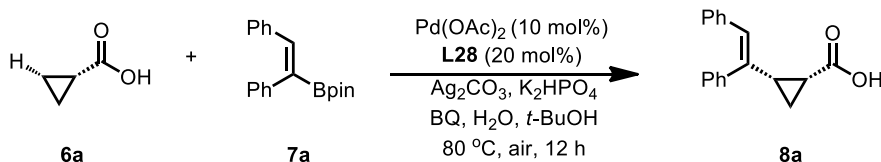


Change	Yield	Base	Yield
No Change	62%	Ag ₂ CO ₃ (1.0 equiv)	53%
K ₂ HPO ₄ (1.0 equiv)	50%	Ag ₂ CO ₃ (2.0 equiv)	62%
K ₂ HPO ₄ (2.0 equiv)	52%	ArBpin (1.0 equiv)	51%
K ₂ HPO ₄ (3.0 equiv)	43%	ArBpin (2.0 equiv)	62%

^a Conditions: **4a** (0.1 mmol), **2a** (1.5 equiv), Pd(OAc)₂ (10 mol%), **L36** (20 mol%), Ag₂CO₃ (2.0 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (5.0 equiv), *t*-BuOH (1.0 mL), 60 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

3.4 Optimization of Conditions of Vinylation for Cyclopropanecarboxylic Acid

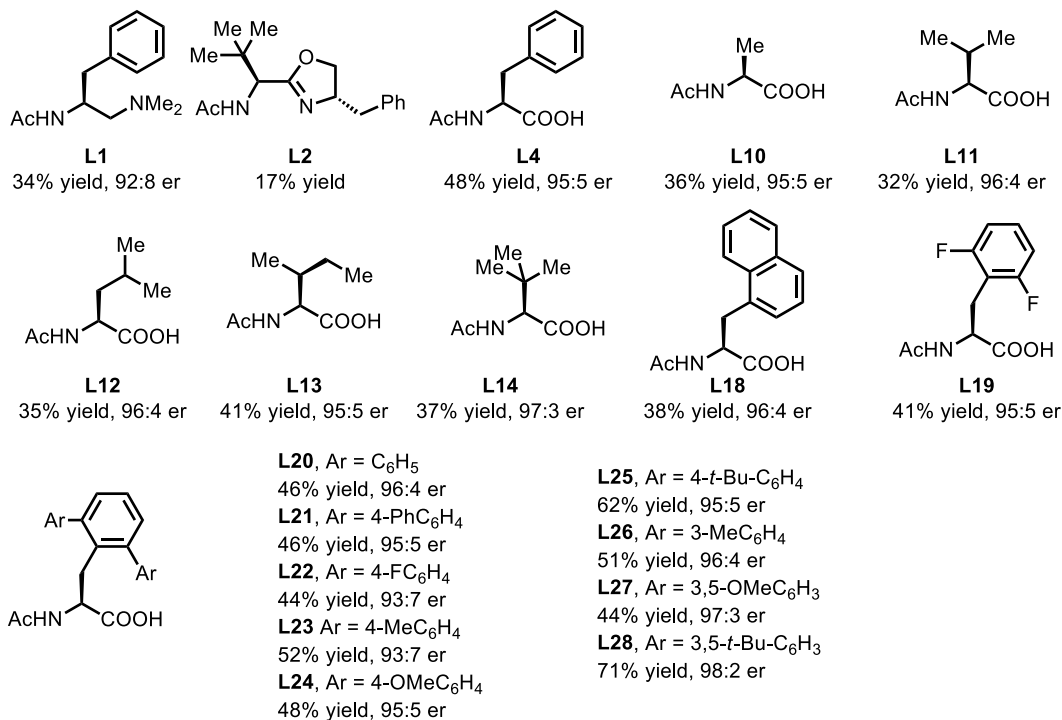
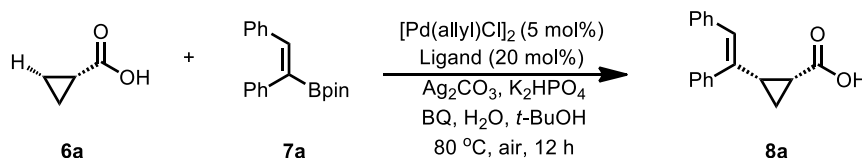
Screening of Ag Salts and Pd source^{a,b}



Ag Salt	Yield	er	Pd source	Yield	er
no change	48%	96:4	PdCl ₂	18%	-
AgOAc	19%	-	Pd(TFA) ₂	52%	95:5
AgOPiv	12%	-	Pd(MeCN) ₄ (BF ₄) ₂	63%	98:2
AgTFA	7%	-	Pd(MeCN) ₂ Cl ₂	53%	97:4
AgF	17%	-	Pd(PhCN) ₂ Cl ₂	52%	96:4
Ag ₂ O	29%	96:4	[Pd(allyl)Cl] ₂	71%	98:2
			Pd(pph ₃) ₂ Cl ₂	54%	96:4

^a Conditions: **6a** (0.1 mmol), **7a** (2.0 equiv), Pd (10 mol%), **L28** (20 mol%), Ag (1.5 equiv), K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (2.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

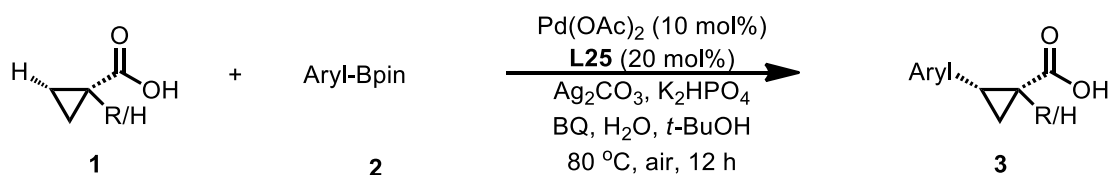
Screening of Ligand^{a,b}



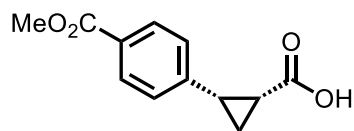
^a Conditions: **6a** (0.1 mmol), **7a** (2.0 equiv), [Pd(allyl)Cl]₂ (5 mol%), Ligand (20 mol%), Ag₂CO₃ (1.5 equiv),

K₂HPO₄ (1.5 equiv), BQ (0.5 equiv), H₂O (2.0 equiv), *t*-BuOH (1.0 mL), 80 °C, air, 12 h. ^b ¹H NMR yields, using CH₂Br₂ as an internal standard.

3.5 Enantioselective Arylation of Cyclopropanecarboxylic acid



General procedure for enantioselective arylation of cyclopropanecarboxylic acid: A 2-dram vial equipped with a magnetic stir bar was charged with Pd(OAc)₂ (2.2 mg, 10 mol%) and L25 (9.4 mg, 20 mol%) in *t*-BuOH (1.0 ml) and then stirred at the rate of 300 rpm at room temperature for 5 min. The appropriate cyclopropanecarboxylic acid substrate (0.10 mmol), Ag₂CO₃ (41.4 mg, 0.15 mmol), K₂HPO₄ (26.0 mg, 0.15 mmol), Aryl-Bpin (0.15 mmol), BQ (5.4 mg, 0.05 mmol), H₂O (18.0 mg, 1.0 mmol) was then added. Subsequently the vial was capped and closed tightly. The reaction mixture was then stirred at the rate of 300 rpm at 80 °C for 12 h. After being allowed to cool to room temperature, the mixture was diluted with ethyl acetate and 0.1 ml of acetic acid was then added. The mixture was passed through a pad of Celite with ethyl acetate as the eluent to remove any insoluble precipitate. The resulting solution was concentrated, and the residual mixture was dissolved with a minimal amount of acetone and loaded onto a preparative TLC plate. The pure product was then isolated using preparative TLC with ethyl acetate and hexane (1/4 to 1/1) as the eluent and 1% v/v of acetic acid as additive.



(1R,2S)-2-(4-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3a)

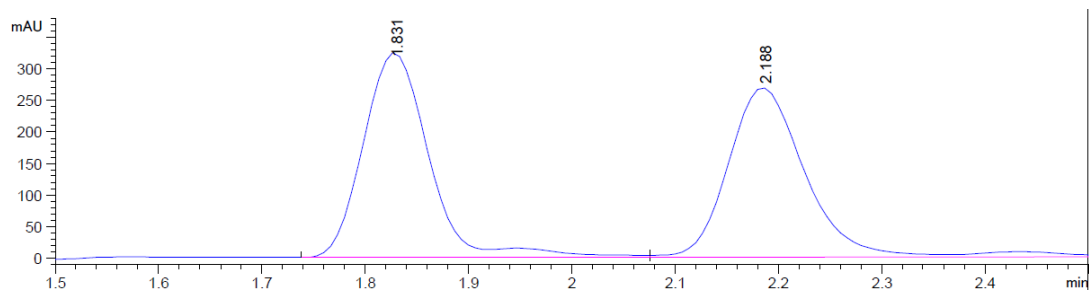
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (70% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 30% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 1.825 min (major) and 2.171 min (minor), 97:3 er); [α]_D²⁰ = -6.7 (c = 1.0, CHCl₃).

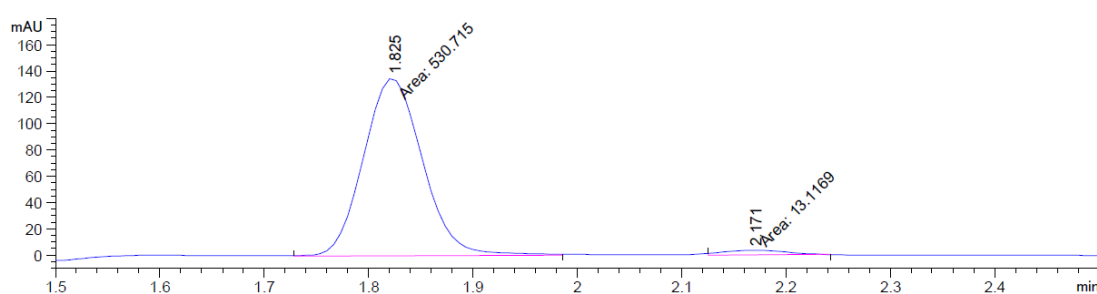
¹H NMR (600 MHz, CDCl₃) δ 7.92 (d, *J* = 8.3 Hz, 2H), 7.29 (d, *J* = 7.8 Hz, 2H), 3.91 (s, 3H), 2.63 (q, *J* = 8.5 Hz, 1H), 2.08 (ddd, *J* = 9.2, 7.8, 5.6 Hz, 1H), 1.69 (dt, *J* = 7.6, 5.4 Hz, 1H), 1.42 (ddd, *J* = 8.5, 7.8, 5.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.11, 167.05, 141.35, 129.30, 129.23, 128.59, 52.02, 26.32, 21.65, 12.24.

HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₃O₄⁺ [M+H]⁺ 221.0808, found 221.0814.

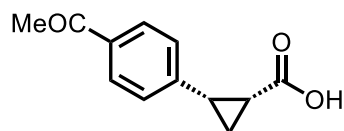
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	1.831	BV R	0.0670	1432.96143	323.97009	50.1015
2	2.188	VV R	0.0791	1427.15442	268.27094	49.8985



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	1.825	MM	0.0653	530.71539	135.44972	97.5881
2	2.171	MM	0.0651	13.11685	3.35660	2.4119



(1R,2S)-2-(4-acetylphenyl)cyclopropane-1-carboxylic acid (**3b**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (62% yield).

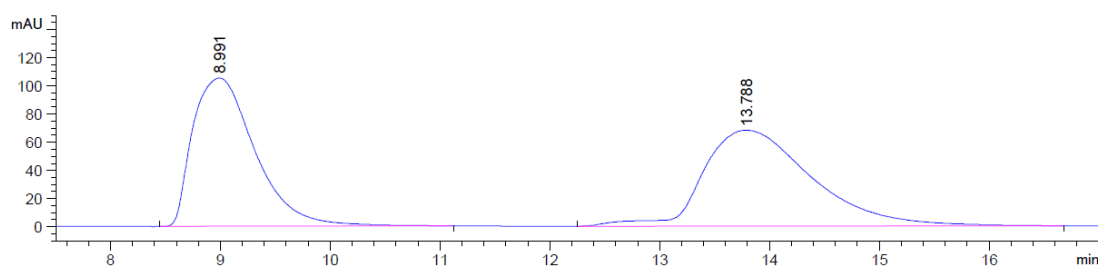
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 9.023 min (major) and 14.154 min (minor), 98:2 er);

¹H NMR (600 MHz, CDCl₃) δ 7.84 (d, *J* = 8.3 Hz, 2H), 7.32 (d, *J* = 8.1 Hz, 2H), 2.64 (q, *J* = 8.5 Hz, 1H), 2.58 (s, 3H), 2.10 (ddd, *J* = 9.3, 7.8, 5.6 Hz, 1H), 1.71 (dt, *J* = 7.8, 5.4 Hz, 1H), 1.44 (td, *J* = 8.2, 5.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 197.95, 175.99, 141.63, 135.66, 129.47, 128.04, 26.58, 26.32, 21.70, 12.29.

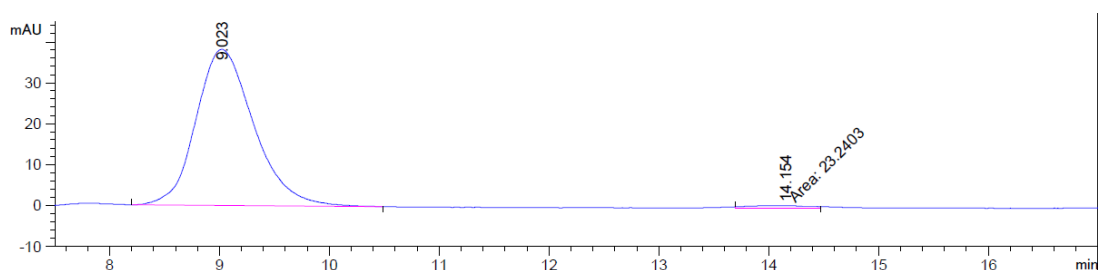
HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₃O₃⁺ [M+H]⁺ 205.0859, found 205.0857.

The absolute stereochemistry was assigned based on comparing the specific rotation of **3m**

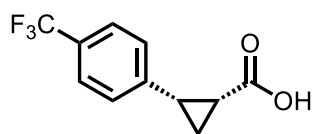
with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.991	BB	0.6176	4144.10840	105.20343	47.0553
2	13.788	BB	1.0413	4662.78320	68.06223	52.9447



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.023	BB	0.5499	1393.16235	38.16215	98.3592
2	14.154	MM	0.6197	23.24029	6.25024e-1	1.6408



(1R,2S)-2-(4-(trifluoromethyl)phenyl)cyclopropane-1-carboxylic acid (3c)

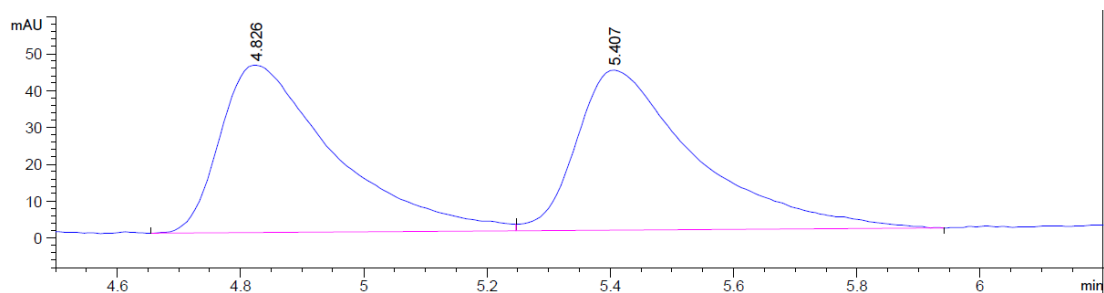
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (69% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 4.897 min (major) and 5.495 min (minor), 97:3 er);

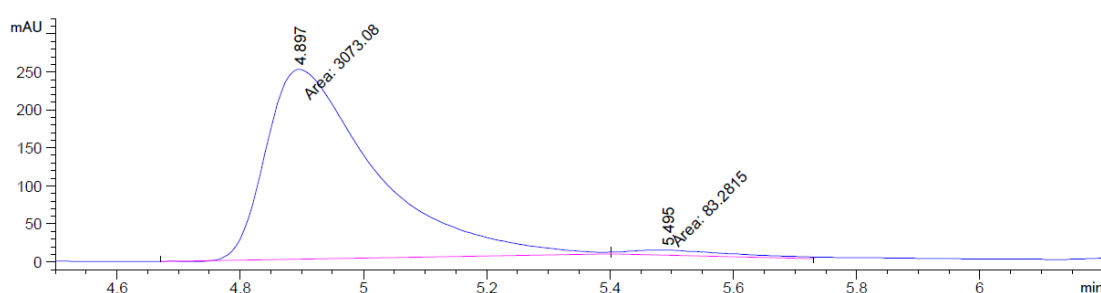
¹H NMR (600 MHz, CDCl₃) δ 7.49 (d, *J* = 8.5 Hz, 2H), 7.33 (d, *J* = 8.4 Hz, 2H), 2.64 (q, *J* = 8.6 Hz, 1H), 2.09 (ddd, *J* = 9.3, 7.7, 5.6 Hz, 1H), 1.69 (dt, *J* = 7.7, 5.4 Hz, 1H), 1.43 (ddd, *J* = 8.7, 7.8, 5.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.43, 140.02 (q, *J* = 1.5 Hz), 129.59, 128.97 (q, *J* = 31.5 Hz), 124.82 (q, *J* = 3 Hz), 124.22 (q, *J* = 270 Hz), 26.09, 21.58, 12.16. ¹⁹F NMR (376 MHz, CDCl₃) δ -62.69.

HRMS (ESI-TOF) m/z Calcd for $C_{11}H_{10}F_3O_2^+ [M+H]^+$ 231.0627, found 231.0631.

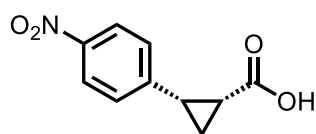
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.826	BV	0.1818	583.19299	45.41054	50.0033
2	5.407	VB	0.1866	583.11597	43.42923	49.9967



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.897	MM	0.2046	3073.07617	250.32617	97.3615
2	5.495	MM	0.2029	83.28146	6.84065	2.6385



(1R,2S)-2-(4-nitrophenyl)cyclopropane-1-carboxylic acid (**3d**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (63% yield).

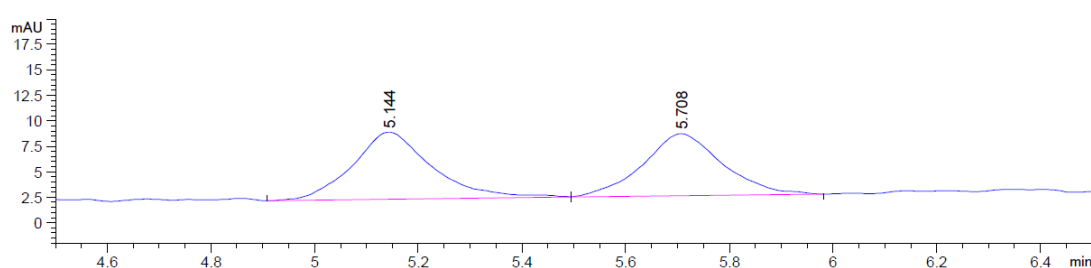
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 5.160 min (major) and 5.715 min (minor), 97:3 er);

¹H NMR (600 MHz, CDCl₃) δ 8.11 (d, J = 8.8 Hz, 2H), 7.39 (d, J = 8.7 Hz, 2H), 2.67 (q, J = 8.5 Hz, 1H), 2.15 (ddd, J = 9.3, 7.8, 5.6 Hz, 1H), 1.72 (dt, J = 7.7, 5.5 Hz, 1H), 1.53 – 1.47 (m,

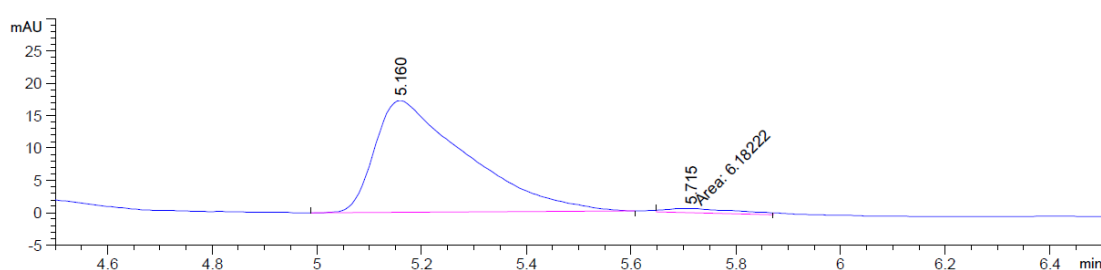
¹H); ¹³C NMR (150 MHz, CDCl₃) δ 175.87, 146.84, 143.71, 130.14, 123.13, 26.05, 21.90, 12.57.

HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₀NO₄⁺ [M+H]⁺ 208.0604, found 208.0607.

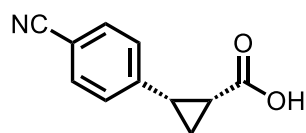
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.144	BV	0.1571	55.05377	4.97626	49.4209
2	5.706	VB	0.1719	56.34407	4.69940	50.5791



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.160	BB	0.1687	211.52269	17.29119	97.1603
2	5.715	MM	0.1533	6.18222	6.72103e-1	2.8397



(1R,2S)-2-(4-cyanophenyl)cyclopropane-1-carboxylic acid (**3e**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (66% yield).

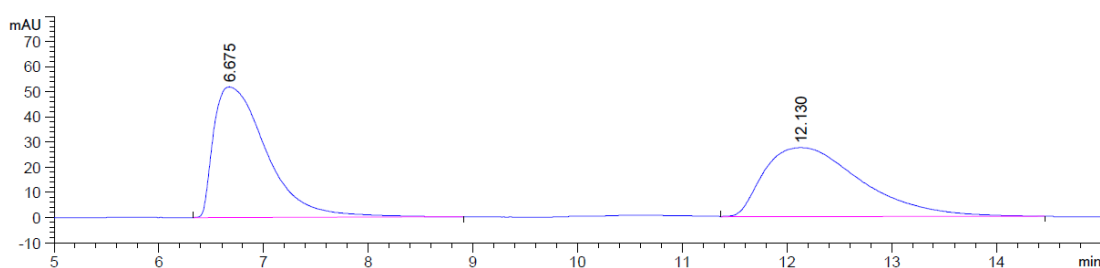
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 5% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 6.706 min (major) and 12.382 min (minor), 97:3 er);

¹H NMR (600 MHz, CDCl₃) δ 7.54 (d, *J* = 8.3 Hz, 2H), 7.33 (d, *J* = 8.2 Hz, 2H), 2.64 (q, *J* = 8.5 Hz, 1H), 2.12 (ddd, *J* = 9.2, 7.8, 5.6 Hz, 1H), 1.69 (dt, *J* = 7.8, 5.5 Hz, 1H), 1.51 – 1.40 (m,

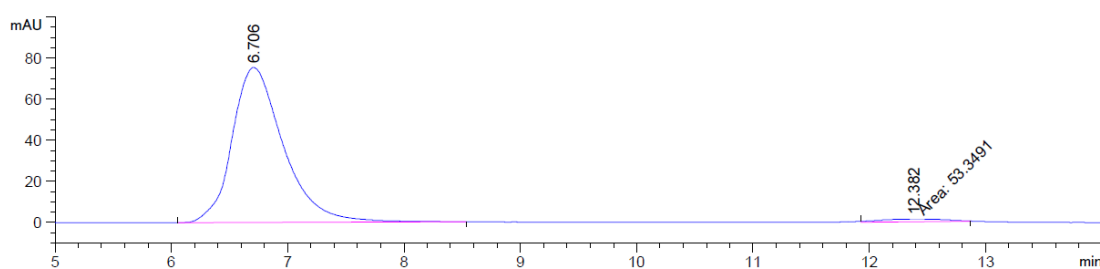
^1H); ^{13}C NMR (150 MHz, CDCl_3) δ 176.23, 141.55, 131.68, 130.06, 118.90, 110.52, 26.28, 21.82, 12.27.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{11}\text{H}_{10}\text{NO}_2^+$ $[\text{M}+\text{H}]^+$ 188.0706, found 188.0704.

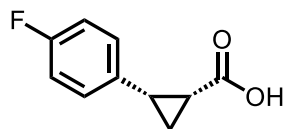
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.675	BB	0.5512	1818.46094	51.84768	50.6245
2	12.130	BB	0.8779	1773.59778	27.37505	49.3755



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.706	BB	0.4634	2305.85181	75.26052	97.7387
2	12.382	MM	0.6413	53.34911	1.38640	2.2613



(1R,2S)-2-(4-fluorophenyl)cyclopropane-1-carboxylic acid (**3f**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (51% yield).

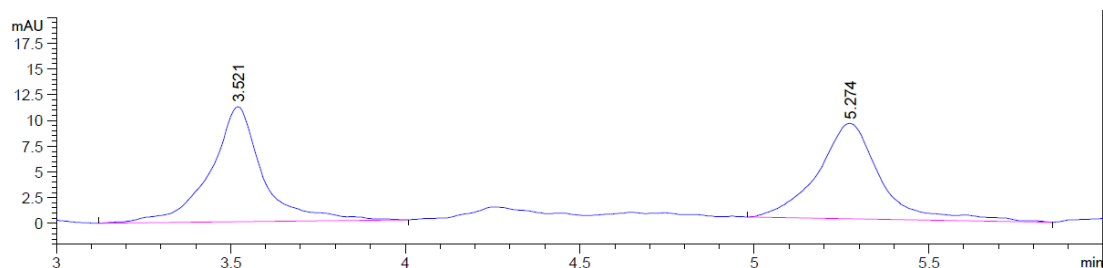
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% i PrOH / CO_2 , flow rate 2 mL/min, retention time 3.394 min (major) and 5.098 min (minor), 97:3 er);

^1H NMR (600 MHz, CDCl_3) δ 7.20 (dd, $J = 8.4, 5.5$ Hz, 2H), 6.93 (t, $J = 8.7$ Hz, 2H), 2.59 (q,

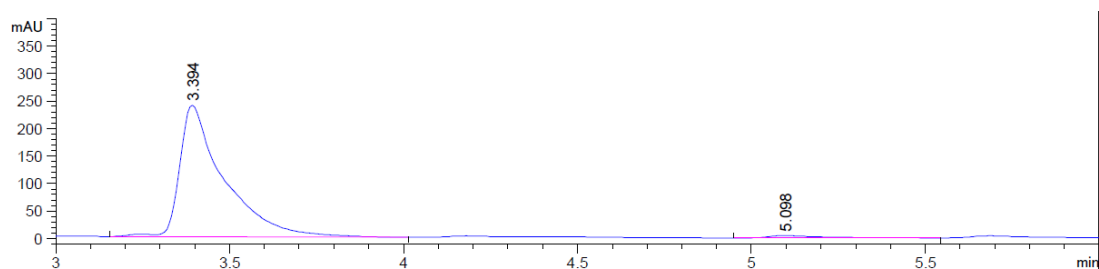
$J = 8.5$ Hz, 1H), 2.05 (ddd, $J = 9.3, 7.9, 5.5$ Hz, 1H), 1.63 (dt, $J = 7.6, 5.4$ Hz, 1H), 1.38 (ddd, $J = 8.7, 7.7, 5.1$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 175.84, 161.74 (d, $J = 243$ Hz), 131.58 (d, $J = 1.5$ Hz), 130.75 (d, $J = 7.5$ Hz), 114.80 (d, $J = 22.5$ Hz), 25.70, 21.22, 12.21; ^{19}F NMR (376 MHz, CDCl_3) δ -116.34.

HRMS (ESI-TOF) m/z . Calcd for $\text{C}_{10}\text{H}_{10}\text{FO}_2^+$ $[\text{M}+\text{H}]^+$ 181.0659, found 181.0658.

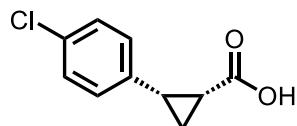
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.521	BB	0.1459	116.74403	11.17264	50.2261
2	5.274	BB	0.1798	115.69290	9.25463	49.7739



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.394	VB R	0.1222	2117.96899	239.30923	97.4252
2	5.098	BV	0.1702	55.97525	4.46813	2.5748



(1R,2S)-2-(4-chlorophenyl)cyclopropane-1-carboxylic acid (**3g**)

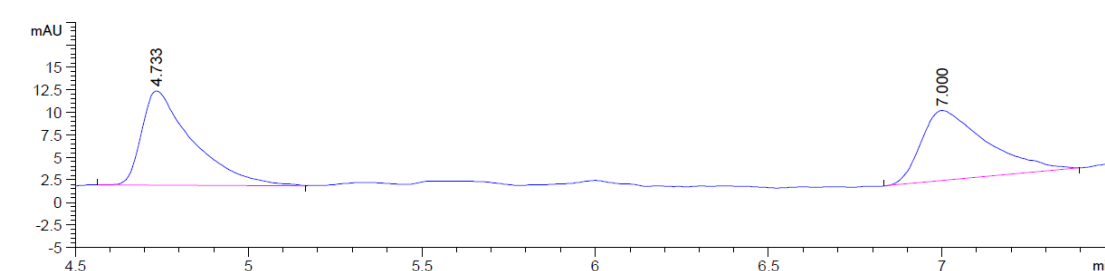
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (70% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 4.726 min (major) and 6.986 min (minor), 97:3 er);

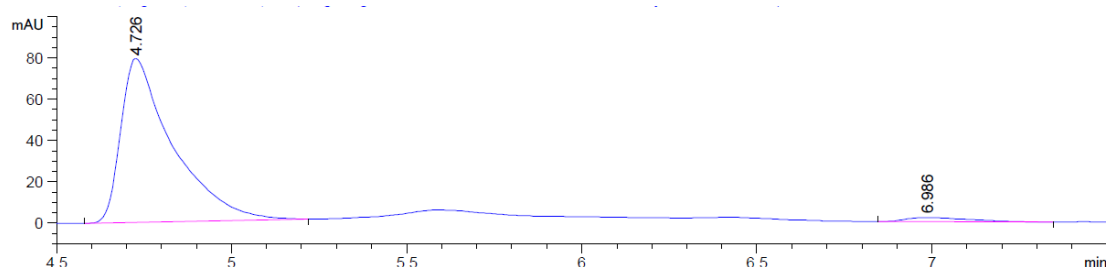
¹H NMR (600 MHz, CDCl₃) δ 7.22 (d, *J* = 8.5 Hz, 2H), 7.17 (d, *J* = 8.1 Hz, 2H), 2.58 (q, *J* = 8.5 Hz, 1H), 2.08 – 2.03 (m, 1H), 1.64 (dt, *J* = 7.7, 5.4 Hz, 1H), 1.39 (ddd, *J* = 8.7, 7.7, 5.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.83, 134.40, 132.56, 130.59, 128.10, 25.82, 21.34, 12.15.

HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₀ClO₂⁺ [M+H]⁺ 197.0364, found 197.0362.

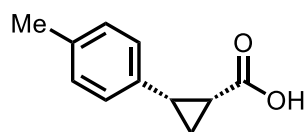
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.733	BB	0.1428	107.01307	10.51189	51.2681
2	7.000	BB	0.1884	101.71914	7.78732	48.7319



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.726	BB	0.1442	818.38550	79.45596	97.1407
2	6.986	BB	0.1774	24.08867	2.01673	2.8593



(1R,2S)-2-(p-tolyl)cyclopropane-1-carboxylic acid (**3h**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (62%

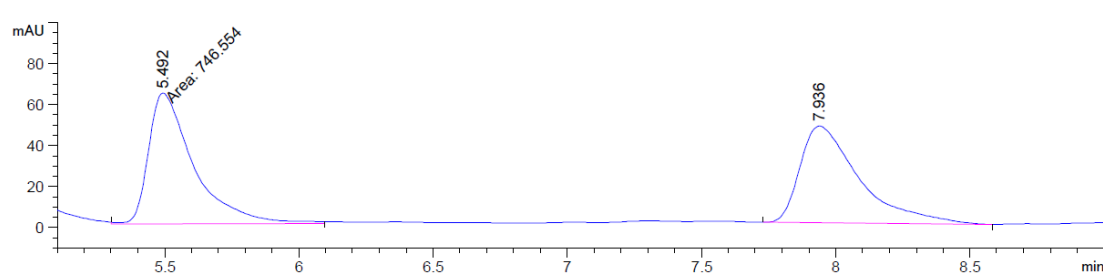
yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% ^tPrOH / CO₂, flow rate 2 mL/min, retention time 5.470min (major) and 7.932 min (minor), 99:1 er);

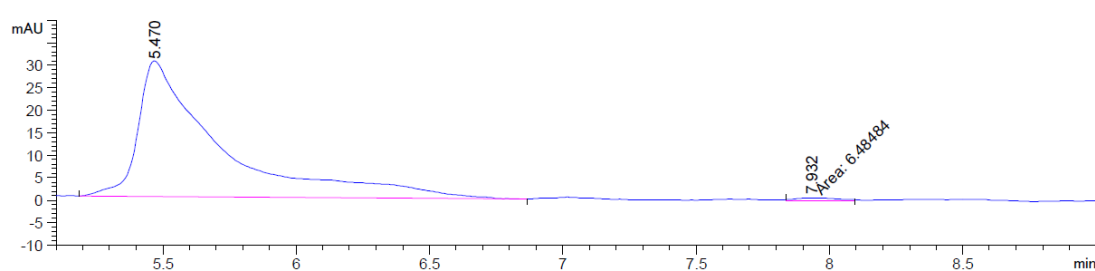
¹H NMR (600 MHz, CDCl₃) δ 7.13 (d, *J* = 7.7 Hz, 2H), 7.05 (d, *J* = 7.7 Hz, 2H), 2.59 (q, *J* = 8.6 Hz, 1H), 2.30 (s, 3H), 2.06 – 2.00 (m, 1H), 1.64 (dt, *J* = 7.8, 5.4 Hz, 1H), 1.35 (td, *J* = 8.2, 4.9 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.58, 136.28, 132.85, 129.09, 128.69, 26.17, 21.23, 21.09, 12.06.

HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₃O₂⁺ [M+H]⁺ 177.0910, found 177.0905.

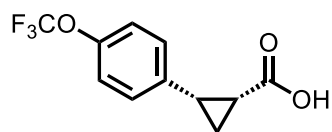
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.492	MM	0.1950	746.55359	63.81255	50.8004
2	7.936	BB	0.2329	723.02826	47.15403	49.1996



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.470	BB	0.2745	642.39697	30.15806	99.0006
2	7.932	MM	0.1920	6.48484	5.62989e-1	0.9994



(1R,2S)-2-(4-(trifluoromethoxy)phenyl)cyclopropane-1-carboxylic acid (**3i**)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl

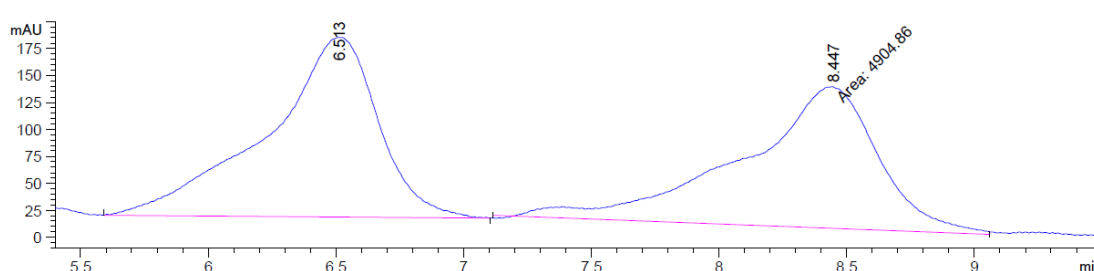
acetate = 3/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (73% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 3% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 6.532 min (major) and 8.508 min (minor), 97:3 er);

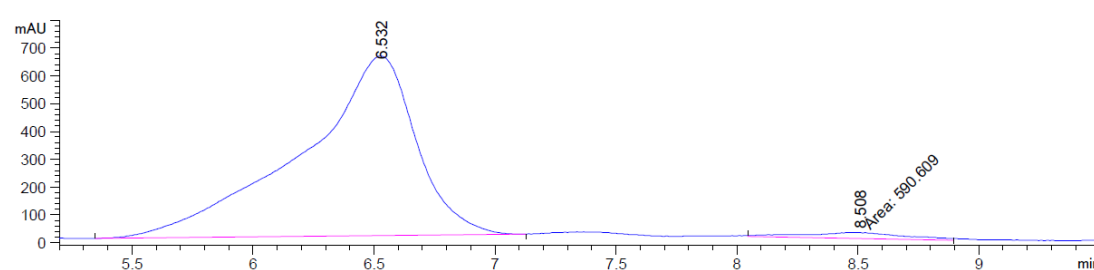
¹H NMR (600 MHz, CDCl₃) δ 7.22 (d, *J* = 8.7 Hz, 2H), 7.08 (d, *J* = 8.3 Hz, 2H), 2.58 (q, *J* = 8.5 Hz, 1H), 2.04 (ddd, *J* = 9.1, 7.7, 5.6 Hz, 1H), 1.62 (dt, *J* = 7.7, 5.4 Hz, 1H), 1.38 (td, *J* = 8.2, 5.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.88, 147.98 (q, *J* = 1.5 Hz), 134.70, 130.58, 120.47 (q, *J* = 255 Hz), 120.37, 25.69, 21.45, 12.12; ¹⁹F NMR (376 MHz, CDCl₃) δ -58.12.

HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₀F₃O₃⁺ [M+H]⁺ 247.0577, found 247.0579.

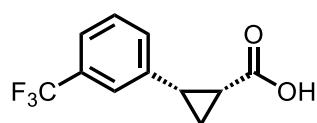
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.513	BV R	0.3599	5056.48486	166.71335	50.7611
2	8.447	MM	0.6216	4904.86084	131.51088	49.2389



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.532	VV R	0.4272	2.12447e4	646.74084	97.2952
2	8.508	MM	0.4780	590.60895	20.59183	2.7048



(1*R*,2*S*)-2-(3-(trifluoromethyl)phenyl)cyclopropane-1-carboxylic acid (**3j**)

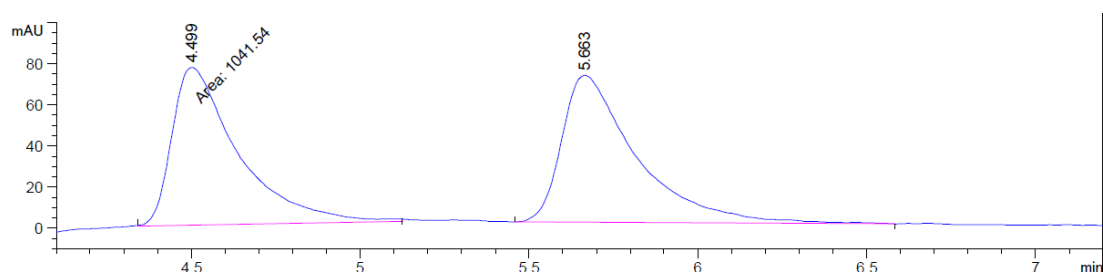
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (77% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 4.529 min (major) and 5.711 min (minor), 92:8 er);

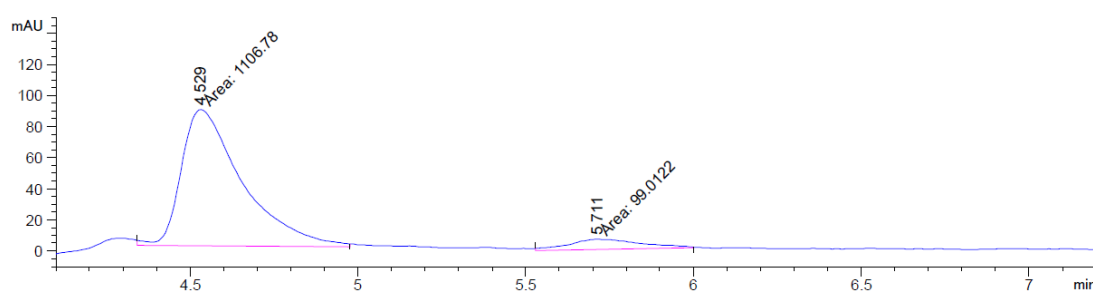
¹H NMR (600 MHz, CDCl₃) δ 7.48 (s, 1H), 7.46 (d, *J* = 7.6 Hz, 1H), 7.39 (d, *J* = 7.8 Hz, 1H), 7.34 (t, *J* = 7.7 Hz, 1H), 2.64 (q, *J* = 8.6 Hz, 1H), 2.08 (ddd, *J* = 9.2, 7.8, 5.6 Hz, 1H), 1.68 (dt, *J* = 7.7, 5.4 Hz, 1H), 1.43 (ddd, *J* = 8.7, 7.8, 5.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.38, 136.92, 132.59, 130.28 (q, *J* = 31.5 Hz), 128.31, 126.11 (q, *J* = 4.5 Hz), 124.11 (q, *J* = 270 Hz), 123.62 (q, *J* = 4.5 Hz), 26.06, 21.40, 12.22; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.90.

HRMS (ESI-TOF) *m/z* Calcd for C₁₁H₁₀F₃O₂⁺ [M+H]⁺ 231.0627, found 231.0628.

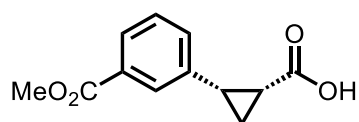
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.499	MM	0.2261	1041.54480	76.76731	48.7093
2	5.663	BV R	0.2211	1096.74170	71.42764	51.2907



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.529	MM	0.2106	1106.78430	87.58733	91.7886
2	5.711	MM	0.2513	99.01224	6.56730	8.2114



(1R,2S)-2-(3-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3k)

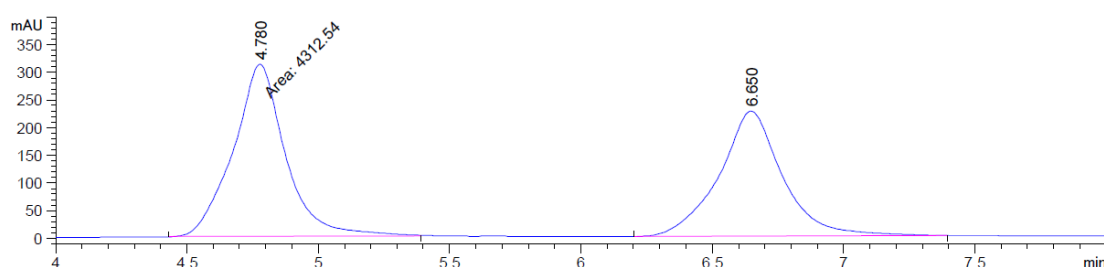
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (67% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 4.789 min (major) and 6.673 min (minor), 95:5 er);

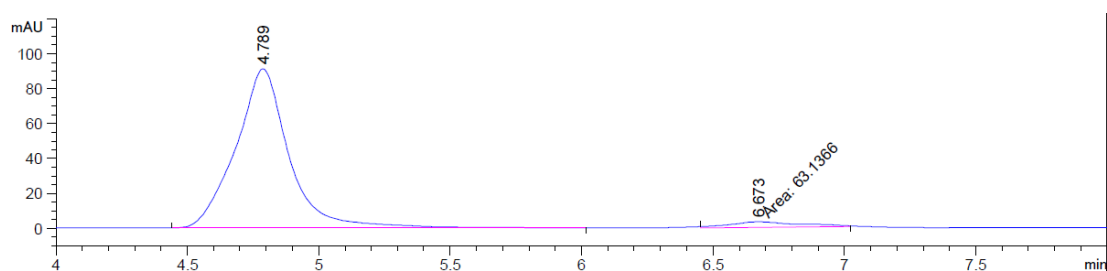
¹H NMR (600 MHz, CDCl₃) δ 7.93 (s, 1H), 7.87 (d, *J* = 7.8 Hz, 1H), 7.40 (d, *J* = 7.6 Hz, 1H), 7.30 (t, *J* = 7.7 Hz, 1H), 3.91 (s, 3H), 2.64 (q, *J* = 8.5 Hz, 1H), 2.10 – 2.03 (m, 1H), 1.70 (dt, *J* = 7.7, 5.4 Hz, 1H), 1.41 (td, *J* = 8.2, 5.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.83, 167.06, 136.33, 133.70, 130.62, 129.87, 128.09, 127.97, 52.08, 26.11, 21.28, 12.15.

HRMS (ESI-TOF) *m/z*. Calcd for C₁₂H₁₃O₄⁺ [M+H]⁺ 221.0808, found 221.0807.

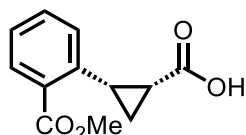
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.781	BB	0.1945	139.56223	10.01887	52.3888
2	6.652	BB	0.2377	126.83482	7.48401	47.6112



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.789	BB	0.1984	1284.97974	91.18695	95.3167
2	6.673	MM	0.3232	63.13662	3.25587	4.6833



(1R,2S)-2-(2-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3l)

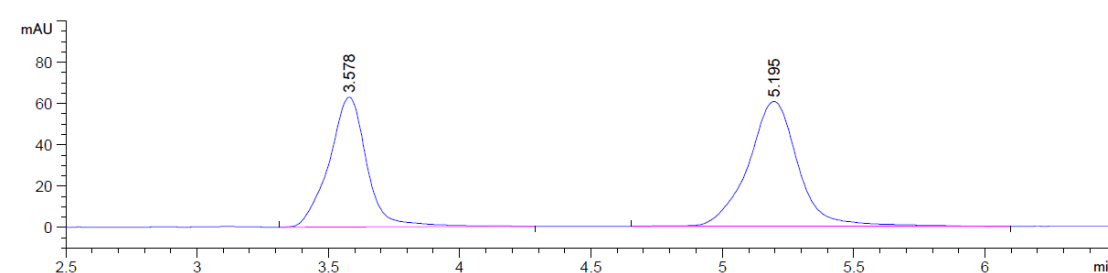
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (54% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.271 min (major) and 4.985 min (minor), 99:1 er);

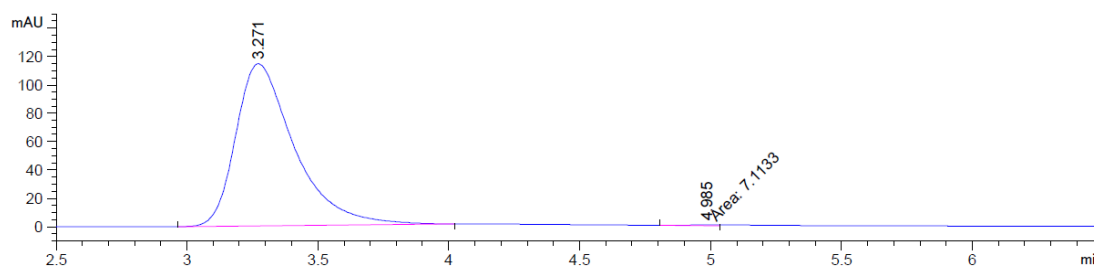
¹H NMR (600 MHz, CDCl₃) δ 7.83 (d, *J* = 7.8 Hz, 1H), 7.41 (t, *J* = 7.6 Hz, 1H), 7.32 (d, *J* = 7.7 Hz, 1H), 7.30 – 7.25 (m, 1H), 3.82 (s, 3H), 2.98 (q, *J* = 8.6 Hz, 1H), 2.12 (ddd, *J* = 9.3, 7.7, 5.4 Hz, 1H), 1.60 (dt, *J* = 8.0, 5.2 Hz, 1H), 1.43 (td, *J* = 8.1, 5.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 177.08, 167.76, 137.61, 131.58, 131.08, 130.88, 130.08, 126.76, 51.95, 26.30, 21.71, 13.43.

HRMS (ESI-TOF) *m/z* Calcd for C₁₂H₁₃O₄⁺ [M+H]⁺ 221.0808, found 221.0807.

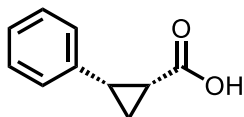
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.578	MM	0.1670	629.59418	62.83921	47.7988
2	5.195	MM	0.1983	687.58069	57.79146	52.2012



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.271	BB	0.2318	1781.41797	114.32291	99.6023
2	4.985	MM	0.1472	7.11330	8.05398e-1	0.3977



(1R,2S)-2-phenylcyclopropane-1-carboxylic acid (3m)

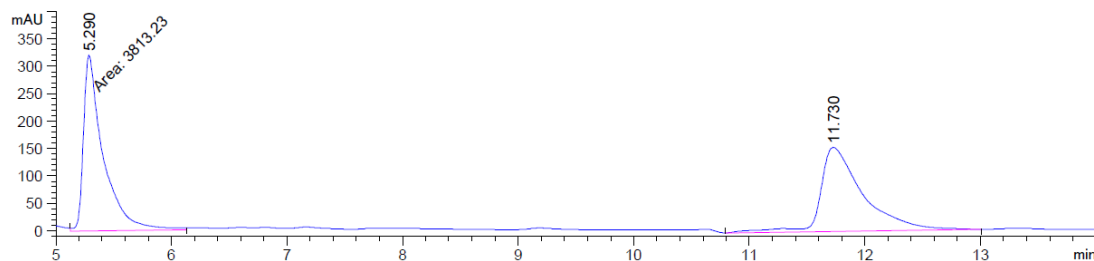
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (68% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 5.287 min (major) and 11.697 min (minor), 98:2 er);

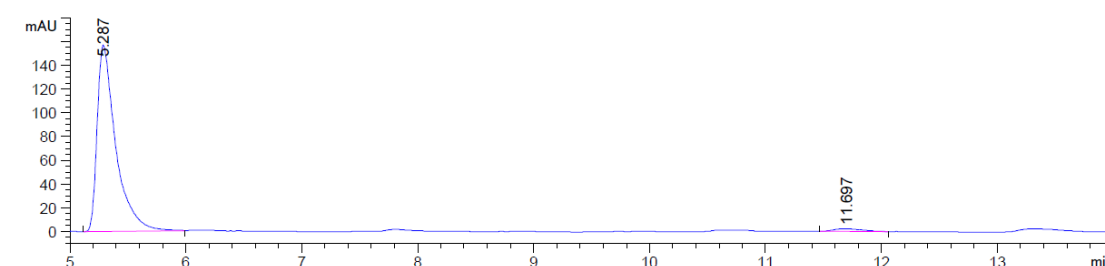
¹H NMR (600 MHz, CDCl₃) δ 7.26 – 7.22 (m, 4H), 7.20 (td, *J* = 5.9, 2.4 Hz, 1H), 2.62 (q, *J* = 8.6 Hz, 1H), 2.04 (ddd, *J* = 9.3, 7.7, 5.6 Hz, 1H), 1.66 (dt, *J* = 7.7, 5.3 Hz, 1H), 1.37 (ddd, *J* = 8.7, 7.7, 5.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.04, 135.91, 129.24, 127.94, 126.76, 26.48, 21.26, 12.00.

HRMS (ESI-TOF) *m/z* Calcd for C₁₀H₁₁O₂⁺ [M+H]⁺ 163.0754, found 163.0752.

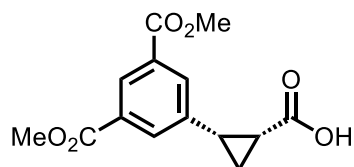
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.290	MM	0.1982	3813.23438	320.63879	49.4393
2	11.730	VB R	0.3614	3899.72827	153.09192	50.5607



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.287	BB	0.1640	1745.86206	156.71941	97.9298
2	11.697	BB	0.2069	36.90720	2.27959	2.0702



(1R,2S)-2-(3,5-bis(methoxycarbonyl)phenyl)cyclopropanecarboxylic acid (3n)

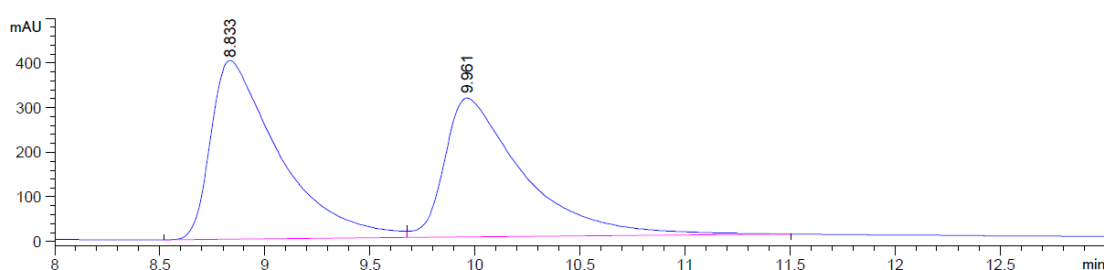
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (66% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 8.903 min (major) and 10.053 min (minor), 97:3 er);

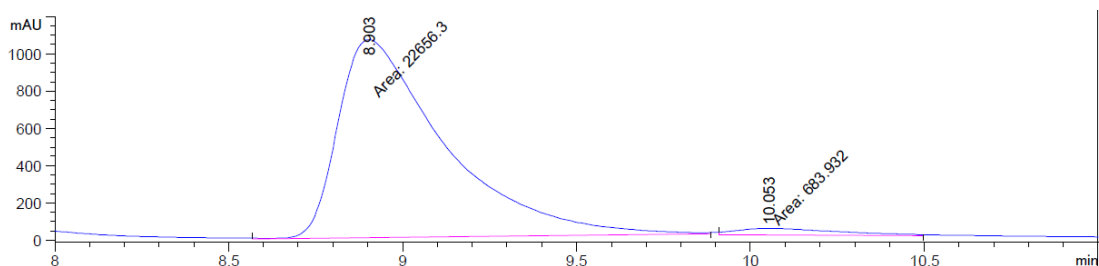
¹H NMR (600 MHz, CDCl₃) δ 8.51 (s, 1H), 8.10 (s, 2H), 3.93 (s, 6H), 2.66 (s, 1H), 2.13 (d, *J* = 7.4 Hz, 1H), 1.75 (s, 1H), 1.46 (s, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 166.28, 161.00, 134.71, 130.27, 129.23, 116.15, 52.36, 24.68, 24.52, 12.32.

HRMS (ESI-TOF) *m/z* Calcd for C₁₄H₁₃O₆⁻ [M-H]⁻ 277.0712, found 277.0719.

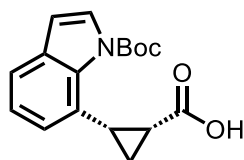
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.833	BV	0.3194	9046.69336	400.76804	52.8028
2	9.961	VV R	0.3767	8086.30322	311.68176	47.1972



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.903	MM	0.3549	2.26563e4	1064.05310	97.0697
2	10.053	MM	0.3360	683.93158	33.92376	2.9303



(1R,2S)-2-(1-(tert-butoxycarbonyl)-1H-indol-7-yl)cyclopropanecarboxylic acid (3o)

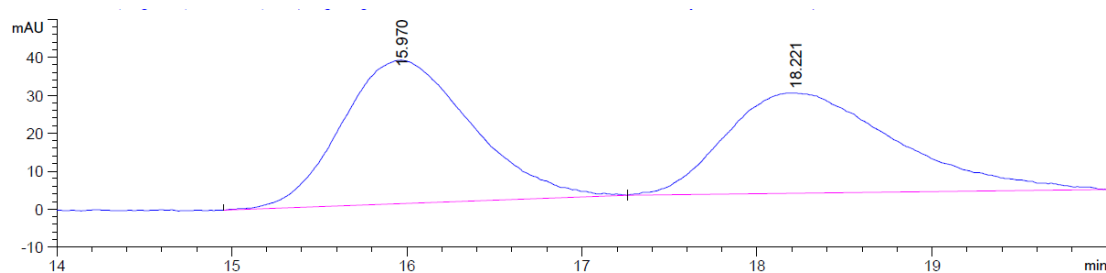
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (47% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 16.895 min (major) and 19.070 min (minor), 99:1 er);

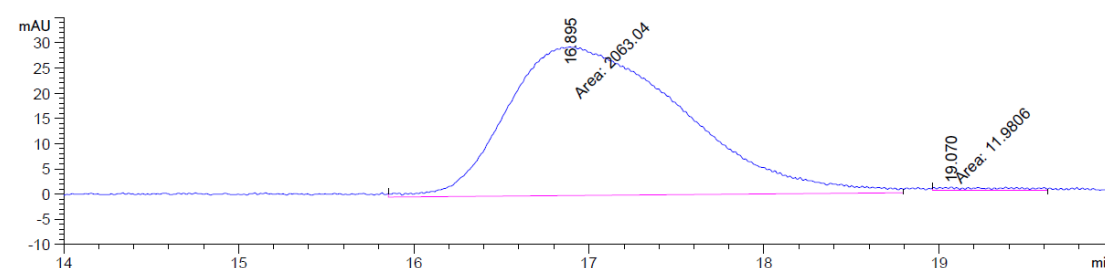
¹H NMR (600 MHz, CDCl₃) δ 8.00 (s, 1H), 7.55 (d, *J* = 3.4 Hz, 1H), 7.42 (dd, *J* = 1.8, 0.9 Hz, 1H), 7.21 (dd, *J* = 8.5, 1.8 Hz, 1H), 6.49 (dd, *J* = 3.6, 0.8 Hz, 1H), 2.72 (q, *J* = 8.6 Hz, 1H), 2.08 – 2.02 (m, 1H), 1.74 – 1.70 (m, 1H), 1.66 (s, 9H), 1.41 – 1.37 (m, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 180.47, 175.83, 149.77, 130.43, 130.20, 126.03, 125.72, 121.42, 114.60, 107.25, 28.19, 26.56, 21.32, 12.26, 9.05.

HRMS (ESI-TOF) *m/z* Calcd for C₁₇H₁₈NO₄ [M-H]⁻ 300.1236, found 300.1228.

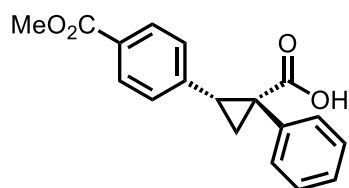
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.972	BV R	0.6088	1993.61938	39.50036	51.2019
2	18.171	MM	1.1199	1900.02429	28.27714	48.7981



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.895	MM	1.1670	2063.04224	29.46432	99.4226
2	19.070	MM	0.3590	11.98057	5.56213e-1	0.5774



(1R,2R)-2-(4-(methoxycarbonyl)phenyl)-1-phenylcyclopropane-1-carboxylic acid (3p)

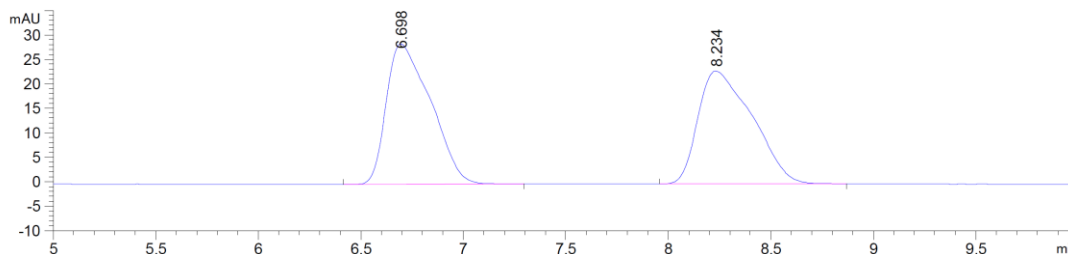
Substrate **1p** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (64% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 20% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 6.300 min (major) and 7.776 min (minor), 98:2 er);

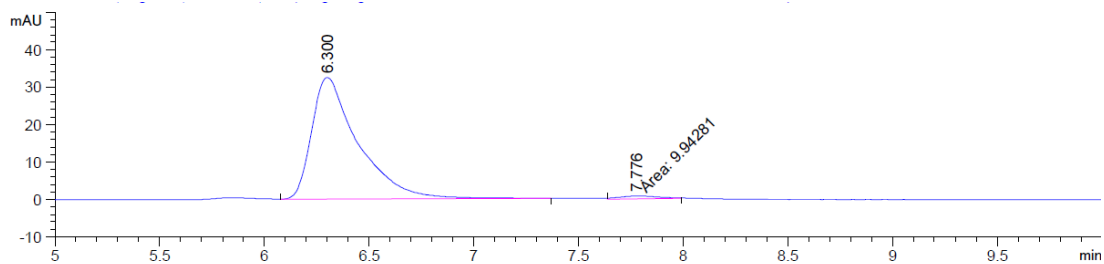
¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, *J* = 8.3 Hz, 2H), 7.48 – 7.41 (m, 2H), 7.35 (td, *J* = 7.9, 5.7 Hz, 4H), 7.31 – 7.27 (m, 1H), 3.93 (s, 3H), 2.89 (t, *J* = 8.4 Hz, 1H), 2.26 (dd, *J* = 7.8, 5.0 Hz, 1H), 1.70 (dd, *J* = 9.0, 5.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.85, 167.05, 141.33, 139.32, 130.19, 129.37, 129.20, 128.66, 128.38, 127.66, 52.06, 37.83, 34.38, 19.25.

HRMS (ESI-TOF) *m/z*. Calcd for C₁₈H₁₆O₄Na⁺ [M+Na]⁺ 319.0941, found 319.0943.

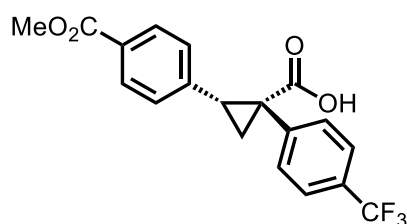
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.698	BB	0.2132	431.89594	28.76545	50.3701
2	8.234	BB	0.2627	425.54874	23.11917	49.6299



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.300	BB	0.2215	500.26663	32.50945	98.0512
2	7.776	MM	0.2147	9.94281	7.71851e-1	1.9488



(1R,2R)-2-(4-(methoxycarbonyl)phenyl)-1-(4-(trifluoromethyl)phenyl)cyclopropane-1-carboxylic acid (3q)

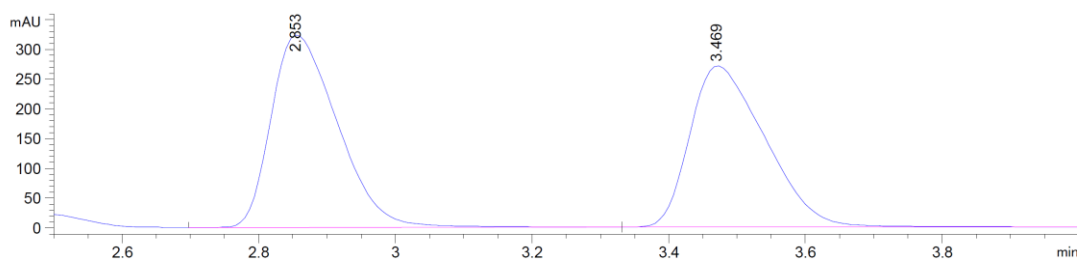
Substrate **1q** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (69% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 20% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 2.741 min (major) and 3.339 min (minor), 97:3 er);

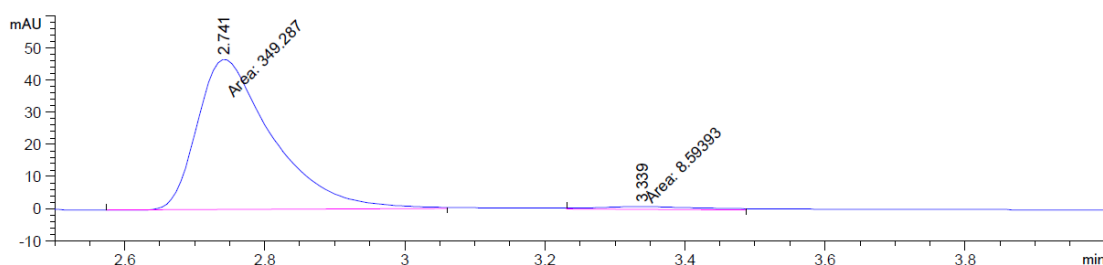
¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, *J* = 8.4 Hz, 2H), 7.60 (d, *J* = 8.1 Hz, 2H), 7.55 (d, *J* = 8.1 Hz, 2H), 7.35 (d, *J* = 8.0 Hz, 2H), 3.93 (s, 3H), 2.88 (t, *J* = 8.5 Hz, 1H), 2.31 (dd, *J* = 7.9, 5.2 Hz, 1H), 1.72 (dd, *J* = 9.0, 5.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.49, 167.02, 143.07, 140.67, 130.63, 129.87 (q, *J* = 33.0 Hz), 129.98, 129.76, 129.54, 129.43, 129.18, 128.87, 125.35 (q, *J* = 3.0 Hz), 123.97 (q, *J* = 270 Hz), 52.12, 37.41, 34.51, 19.20; ¹⁹F NMR (376 MHz, CDCl₃) δ -62.82.

HRMS (ESI-TOF) *m/z* Calcd for C₁₉H₁₆F₃O₄⁺ [M+H]⁺ 365.0995, found 365.0994.

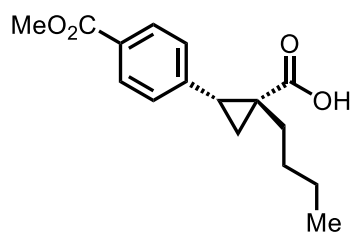
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.853	BB	0.1063	2174.49072	323.80118	51.2016
2	3.469	BB	0.1233	2072.42944	270.79803	48.7984



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.741	MM	0.1243	349.28665	46.85023	97.5987
2	3.339	MM	0.1612	8.59393	8.88326e-1	2.4013



(1R,2R)-1-butyl-2-(4-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3r)

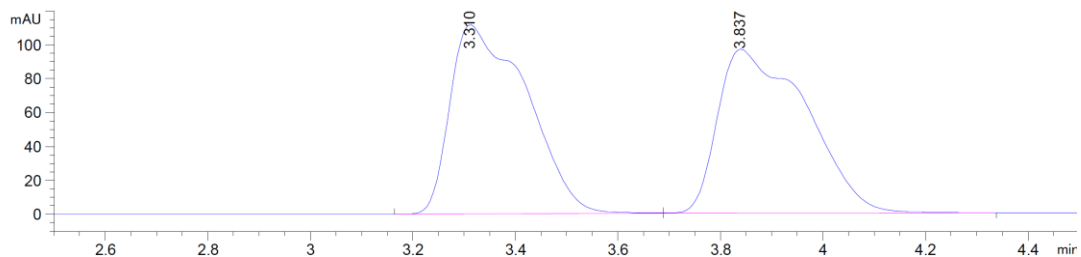
Substrate **1r** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (63% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 20% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.240 min (major) and 3.738 min (minor), 95:5 er);

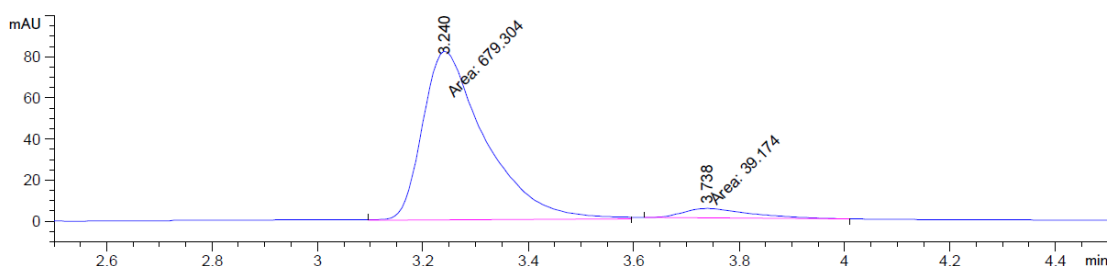
¹H NMR (600 MHz, CDCl₃) δ 7.89 (d, *J* = 8.4 Hz, 2H), 7.21 (d, *J* = 8.2 Hz, 2H), 3.90 (s, 3H), 2.38 (t, *J* = 8.0 Hz, 1H), 2.06 – 1.98 (m, 1H), 1.88 (ddd, *J* = 7.5, 5.0, 1.0 Hz, 1H), 1.52 – 1.39 (m, 2H), 1.35 – 1.26 (m, 3H), 1.18 (dd, *J* = 8.7, 5.1 Hz, 1H), 0.91 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 177.71, 167.10, 142.12, 129.14, 129.02, 128.22, 35.27, 33.44, 33.23, 29.74, 22.72, 18.63, 13.93.

HRMS (ESI-TOF) *m/z* Calcd for C₁₆H₂₀O₄⁺ [M+H]⁺ 277.1434, found 277.1437.

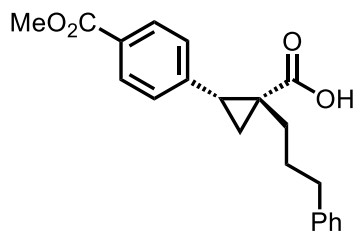
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.310	BV	0.1411	1157.89111	111.58231	49.9134
2	3.837	VB	0.1621	1161.91125	96.72337	50.0866



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.240	MM	0.1378	679.30371	82.13483	94.5476
2	3.738	MM	0.1443	39.17395	4.52614	5.4524



(1R,2R)-2-(4-(methoxycarbonyl)phenyl)-1-(3-phenylpropyl)cyclopropane-1-carboxylic acid (3s)

Substrate **1s** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (44% yield).

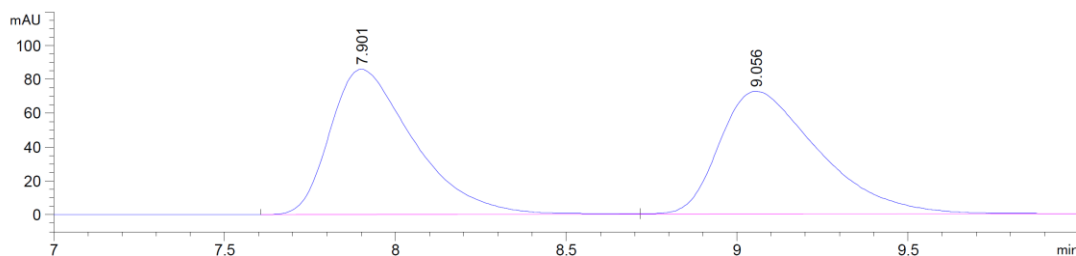
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 20% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 7.675 min (major) and 8.768 min (minor), 96:4 er);

¹H NMR (600 MHz, CDCl₃) δ 7.88 (d, *J* = 8.3 Hz, 2H), 7.28 (t, *J* = 7.6 Hz, 2H), 7.22 – 7.15 (m, 5H), 3.88 (s, 3H), 2.63 (t, *J* = 7.7 Hz, 2H), 2.36 (t, *J* = 8.1 Hz, 1H), 2.05 (ddd, *J* = 13.7, 10.9, 5.1 Hz, 1H), 1.93 – 1.76 (m, 3H), 1.37 (ddd, *J* = 13.8, 11.0, 5.2 Hz, 1H), 1.16 (dd, *J* =

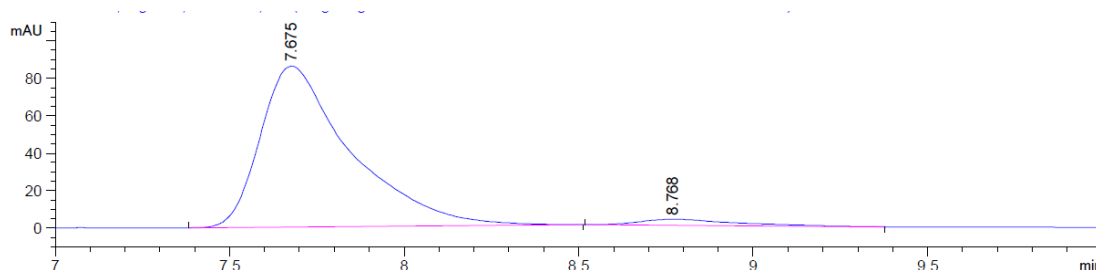
8.7, 5.0 Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 177.54, 167.08, 141.97, 141.91, 129.15, 129.05, 128.31, 128.29, 125.77, 51.97, 35.78, 35.14, 33.55, 32.98, 29.17, 18.78.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{21}\text{H}_{23}\text{O}_4^+$ $[\text{M}+\text{H}]^+$ 339.1591, found 339.1595.

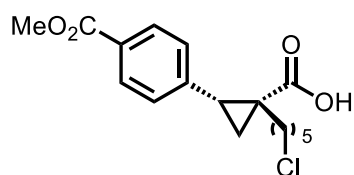
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.901	BV	0.2568	1463.35474	85.87017	50.0170
2	9.056	VBA	0.3032	1462.35803	72.52623	49.9830



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.675	BB	0.2599	1545.83704	85.93114	96.1478
2	8.768	BB	0.2778	61.93466	3.19981	3.8522



(1R,2R)-1-(5-chloropentyl)-2-(4-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3t)

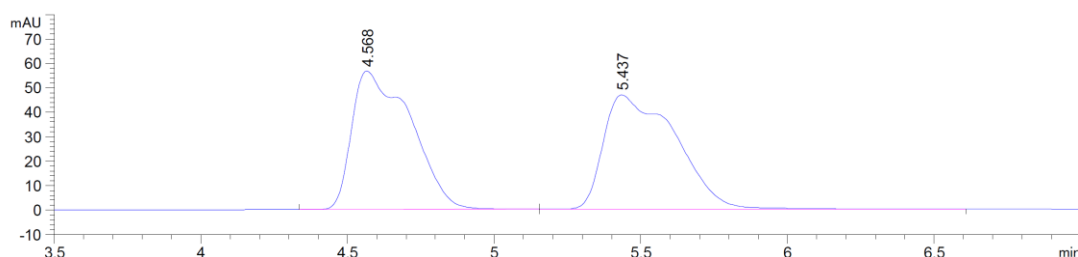
Substrate **1t** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (62% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK[®] AD-3 column, 20% i PrOH / CO_2 , flow rate 2 mL/min, retention time 4.489 min (major) and 5.300 min (minor), 96:4 er);

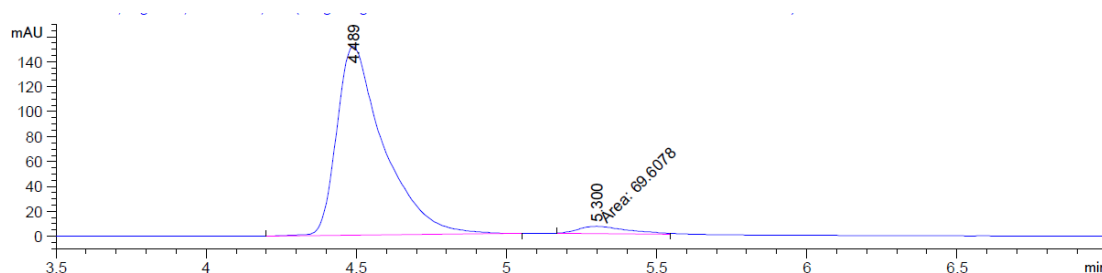
^1H NMR (600 MHz, CDCl_3) δ 7.89 (d, $J = 8.4$ Hz, 2H), 7.22 (d, $J = 8.1$ Hz, 2H), 3.90 (s, 3H), 3.53 (t, $J = 6.7$ Hz, 2H), 2.39 (t, $J = 8.1$ Hz, 1H), 2.04 (ddd, $J = 14.5, 10.6, 4.2$ Hz, 1H), 1.92 – 1.86 (m, 1H), 1.79 (p, $J = 6.8$ Hz, 2H), 1.57 – 1.42 (m, 4H), 1.33 (ddd, $J = 13.8, 10.5, 5.4$ Hz, 1H), 1.20 (dd, $J = 8.7, 5.1$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 177.61, 167.06, 141.89, 129.14, 129.03, 128.28, 51.98, 44.95, 35.36, 33.53, 33.01, 32.34, 26.86, 26.82, 18.72.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{17}\text{H}_{22}\text{ClO}_4^+$ $[\text{M}+\text{H}]^+$ 325.1201, found 325.1205.

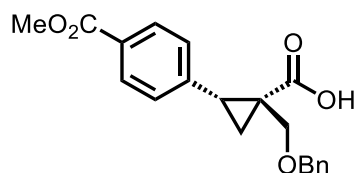
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.568	BB	0.1887	791.70410	56.76482	49.7234
2	5.437	BB	0.2299	800.51172	46.70959	50.2766



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.489	BB	0.1568	1645.09912	151.36237	95.9405
2	5.300	MM	0.1949	69.60776	5.95382	4.0595



(1R,2R)-1-((benzyloxy)methyl)-2-(4-(methoxycarbonyl)phenyl)cyclopropane-1-carboxylic acid (3u**)**

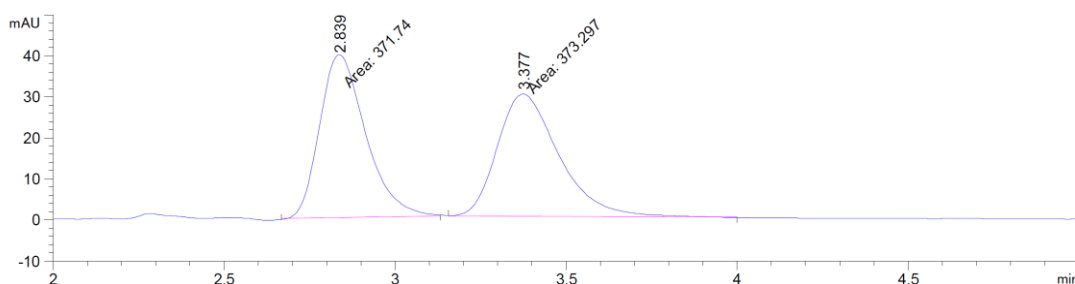
Substrate **1u** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (57% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 20% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 2.835 min (major) and 3.830 min (minor), 98:2 er);

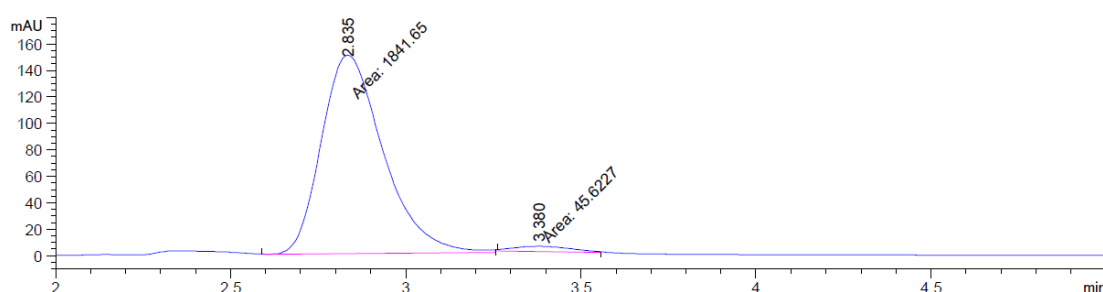
¹H NMR (600 MHz, CDCl₃) δ 7.90 (d, *J* = 8.0 Hz, 2H), 7.37 – 7.27 (m, 5H), 7.24 (d, *J* = 8.0 Hz, 2H), 4.58 (s, 2H), 3.87 (d, *J* = 12.2 Hz, 4H), 3.59 (d, *J* = 10.1 Hz, 1H), 2.58 (t, *J* = 8.3 Hz, 1H), 1.96 (dd, *J* = 7.7, 5.1 Hz, 1H), 1.41 (dd, *J* = 8.9, 5.1 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 175.29, 167.00, 141.19, 137.70, 129.23, 129.13, 128.53, 128.43, 127.78, 127.68, 73.15, 71.85, 51.98, 32.79, 31.00, 16.76.

HRMS (ESI-TOF) *m/z* Calcd for C₂₀H₂₁O₅⁺ [M+H]⁺ 341.1383, found 341.1383.

The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.

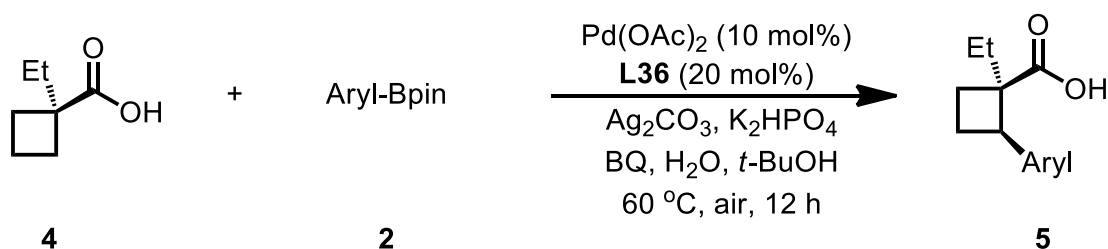


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.839	MM	0.1558	371.74030	39.76003	49.8956
2	3.377	MM	0.2087	373.29666	29.81713	50.1044



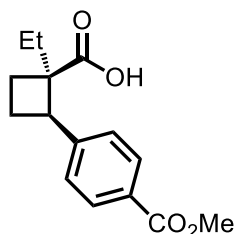
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.835	MM	0.2044	1841.64624	150.13182	97.5826
2	3.380	MM	0.1923	45.62273	3.95362	2.4174

3.6 Enantioselective Arylation of Cyclobutanecarboxylic acid



General procedure for enantioselective arylation of cyclobutanecarboxylic acid:

A 2-dram vial equipped with a magnetic stir bar was charged with Pd(OAc)₂ (2.2 mg, 10 mol%) and **L36** (7.4 mg, 20 mol%) in *t*-BuOH (1.0 ml) and then stirred at the rate of 300 rpm at room temperature for 5 min. The appropriate cyclopropanecarboxylic acid substrate (0.10 mmol), Ag₂CO₃ (55.2 mg, 0.2 mmol), K₂HPO₄ (26.0 mg, 0.15 mmol), Aryl-Bpin (0.15 mmol), BQ (5.4 mg, 0.05 mmol), H₂O (9.0 mg, 0.5 mmol) was then added. Subsequently the vial was capped and closed tightly. The reaction mixture was then stirred at the rate of 300 rpm at 60 °C for 12 h. After being allowed to cool to room temperature, the mixture was diluted with ethyl acetate and 0.1 ml of acetic acid was then added. The mixture was passed through a pad of Celite with ethyl acetate as the eluent to remove any insoluble precipitate. The resulting solution was concentrated, and the residual mixture was dissolved with a minimal amount of acetone and loaded onto a preparative TLC plate. The pure product was then isolated using preparative TLC with ethyl acetate and hexane (1/4 to 1/1) as the eluent and 1% v/v of acetic acid as additive.



(1R,2R)-1-ethyl-2-(4-(methoxycarbonyl)phenyl)cyclobutanecarboxylic acid (**5a**)

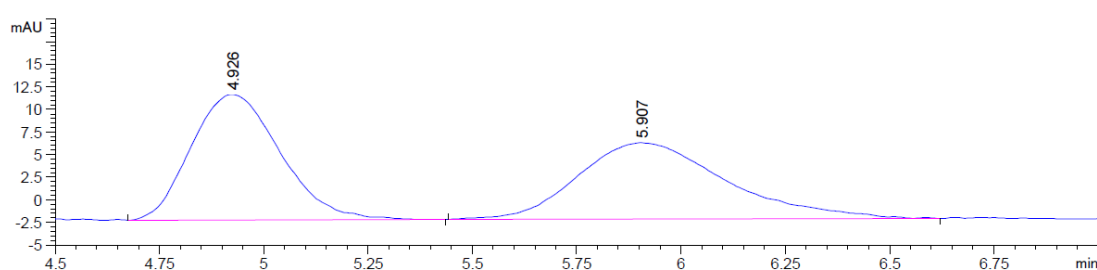
Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (62% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 15% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 4.910 min (minor) and 5.923 min (major), 94:6 er); [α]_D²⁰ = -11.8 (c = 1.0, CHCl₃).

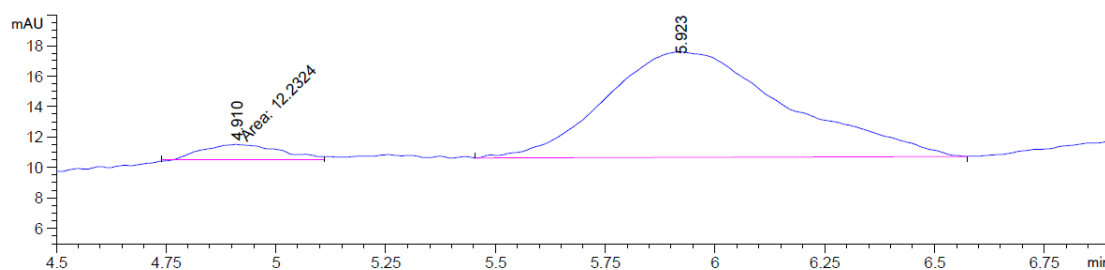
¹H NMR (600 MHz, CDCl₃) δ 7.94 (d, *J* = 8.4 Hz, 2H), 7.24 (d, *J* = 8.2 Hz, 2H), 3.89 (s, 3H), 3.57 (t, *J* = 8.9 Hz, 1H), 2.54 (td, *J* = 10.5, 3.9 Hz, 1H), 2.50 – 2.43 (m, 1H), 2.24 (dtd, *J* = 11.5, 8.9, 3.9 Hz, 1H), 2.17 (dt, *J* = 14.4, 7.2 Hz, 1H), 1.92 – 1.86 (m, 1H), 1.79 – 1.73 (m, 1H), 0.86 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 177.15, 167.05, 145.89, 129.45, 128.45, 127.33, 56.43, 51.98, 50.03, 32.25, 25.37, 21.18, 9.08.

HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₁₇O₄⁻ [M-H] 261.1127, found 261.1132.

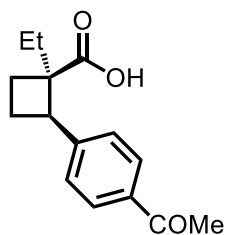
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.926	BB	0.2330	206.27145	13.90880	49.7158
2	5.907	BB	0.3484	208.62941	8.43910	50.2842



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.910	MM	0.2084	12.23242	9.78333e-1	5.8295
2	5.923	BB	0.3590	197.60303	6.92348	94.1705



(1R,2R)-2-(4-acetylphenyl)-1-ethylcyclobutanecarboxylic acid (5b)

Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (65% yield).

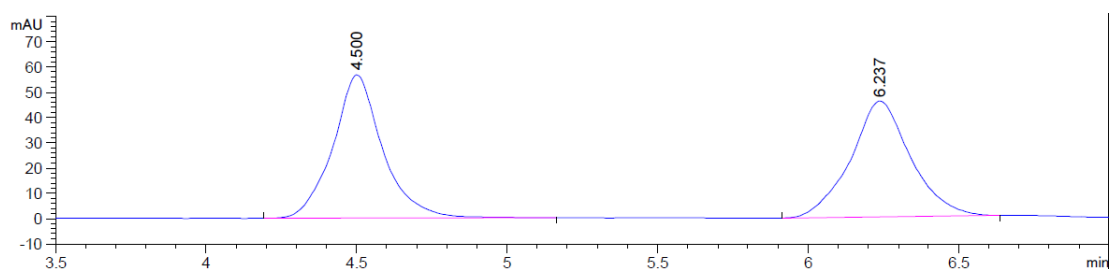
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 4.255 min (minor) and 5.859 min (major), 93:7 er);

¹H NMR (600 MHz, CDCl₃) δ 7.86 (d, *J* = 8.3 Hz, 2H), 7.26 (s, 2H), 3.57 (t, *J* = 8.8 Hz, 1H), 2.57 (s, 3H), 2.54 (dd, *J* = 10.9, 4.1 Hz, 1H), 2.47 (dd, *J* = 11.2, 8.8 Hz, 1H), 2.27 – 2.23 (m, 1H), 2.17 (d, *J* = 6.2 Hz, 1H), 1.92 – 1.87 (m, 1H), 1.77 (dd, *J* = 13.6, 7.2 Hz, 1H), 0.86 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 197.86, 177.29, 146.20, 135.55, 128.26, 127.52,

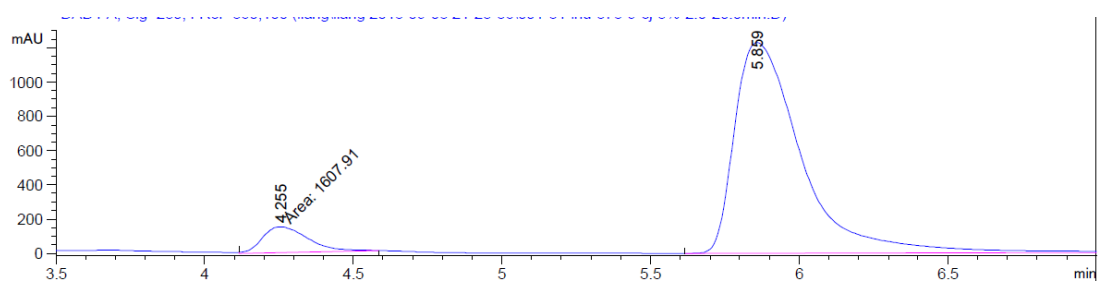
56.43, 50.01, 32.27, 26.54, 25.41, 21.22, 9.10.

HRMS (ESI-TOF) m/z Calcd for $C_{15}H_{17}O_3^-$ $[M-H]^-$ 261.1127, found 261.1132.

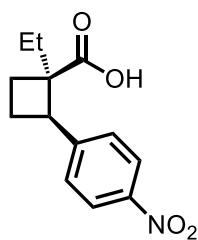
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.500	BB	0.1665	653.67517	56.71861	50.7258
2	6.237	BB	0.1997	634.97046	45.81872	49.2742



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.255	MM	0.1771	1607.90540	151.29349	7.5327
2	5.859	BV R	0.2455	1.97379e4	1229.25208	92.4673



(1R,2R)-1-ethyl-2-(4-nitrophenyl)cyclobutanecarboxylic acid (5c)

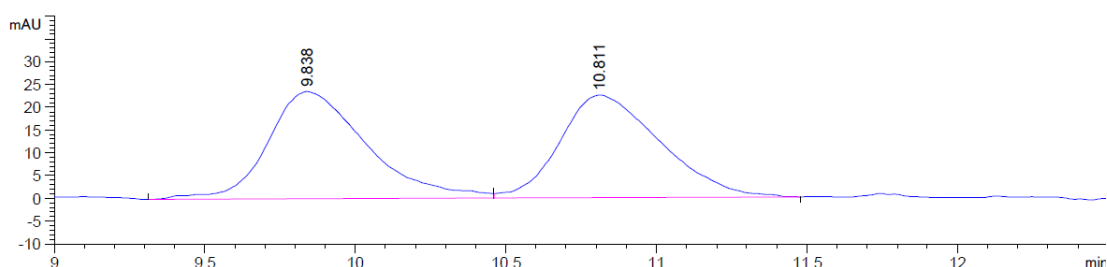
Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (43% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 9.557 min (minor) and 10.402 min (major), 94:6 er);

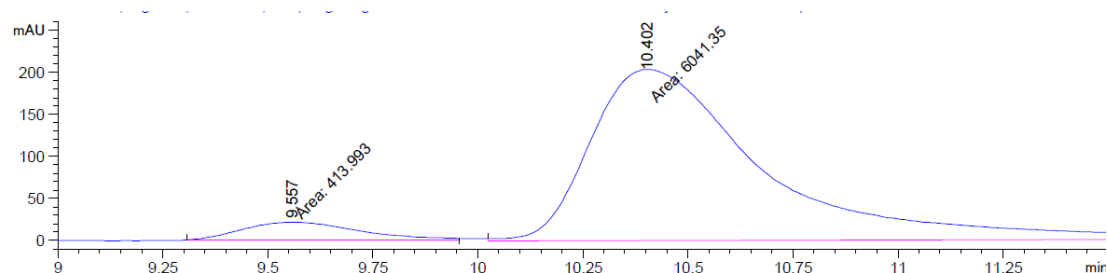
^1H NMR (600 MHz, CDCl_3) δ 8.13 (d, $J = 8.7$ Hz, 2H), 7.32 (d, $J = 8.3$ Hz, 2H), 3.61 (t, $J = 8.8$ Hz, 1H), 2.57 – 2.53 (m, 1H), 2.49 – 2.43 (m, 1H), 2.29 (ddd, $J = 9.1, 7.1, 4.4$ Hz, 1H), 2.19 – 2.15 (m, 1H), 1.95 – 1.91 (m, 1H), 1.80 – 1.75 (m, 1H), 0.86 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 177.10, 148.26, 146.72, 128.12, 123.35, 56.52, 49.63, 32.24, 25.39, 21.23, 9.03.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{13}\text{H}_{14}\text{NO}_4^-$ [M-H] $^-$ 248.0923, found 248.0923.

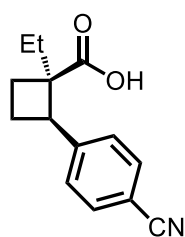
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.838	BV	0.3208	539.44019	23.58702	50.7821
2	10.811	VB	0.3290	522.82501	22.50610	49.2179



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.557	MM	0.3248	413.99335	21.24576	6.4132
2	10.402	MM	0.4934	6041.35449	204.07893	93.5868



(1R,2R)-2-(4-cyanophenyl)-1-ethylcyclobutanecarboxylic acid (5d)

Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (50%

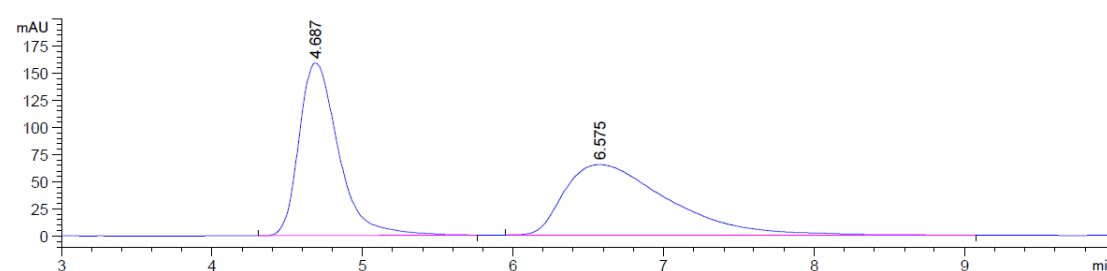
yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 5% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 4.597 min (minor) and 6.464 min (major), 95:5 er);

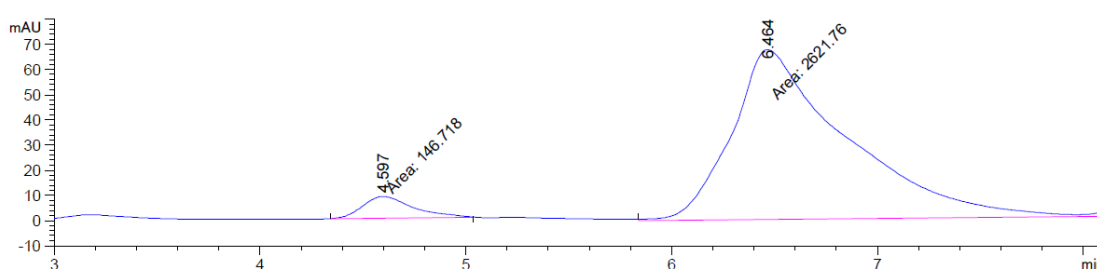
¹H NMR (600 MHz, CDCl₃) δ 7.54 (d, *J* = 8.3 Hz, 2H), 7.25 (d, *J* = 8.2 Hz, 2H), 3.55 (t, *J* = 8.8 Hz, 1H), 2.51 (ddd, *J* = 11.4, 9.6, 4.2 Hz, 1H), 2.42 (dq, *J* = 11.7, 9.0 Hz, 1H), 2.25 (dtd, *J* = 11.5, 8.9, 4.2 Hz, 1H), 2.17 – 2.11 (m, 1H), 1.90 (dt, *J* = 11.5, 8.8 Hz, 1H), 1.75 (dt, *J* = 13.3, 7.4 Hz, 1H), 0.86 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.94, 146.04, 131.86, 128.06, 118.93, 110.31, 56.48, 49.85, 32.17, 25.23, 20.96, 9.06.

HRMS (ESI-TOF) *m/z*. Calcd for C₁₄H₁₄NO₂⁻ [M-H]⁻ 228.1025, found 228.1023.

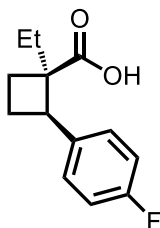
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.687	BB	0.2816	2886.38135	159.12334	48.1444
2	6.575	BB	0.7145	3108.88110	64.94115	51.8556



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.597	MM	0.2843	146.71846	8.60063	5.2996
2	6.464	MM	0.6480	2621.76270	67.42942	94.7004



(1R,2R)-1-ethyl-2-(4-fluorophenyl)cyclobutanecarboxylic acid (5e)

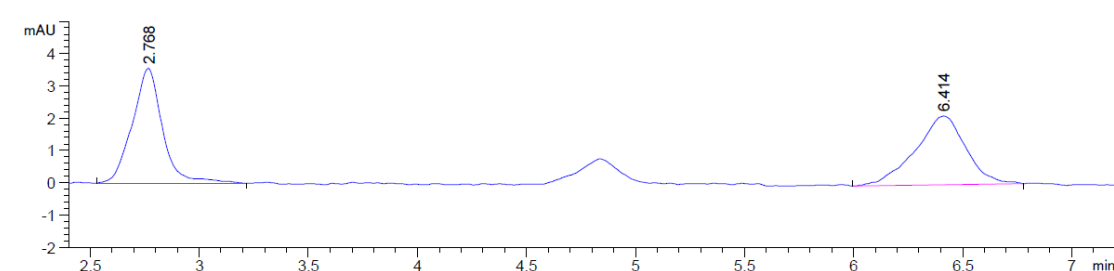
Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (62% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 2.639 min (minor) and 6.111 min (major), 95:5 er);

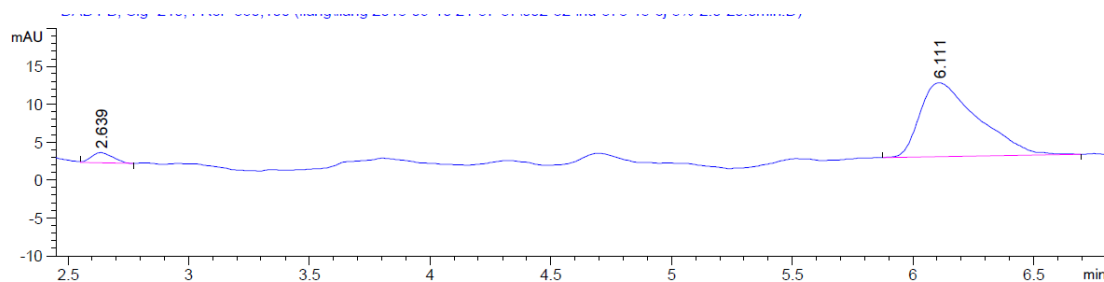
¹H NMR (600 MHz, CDCl₃) δ 7.11 (ddd, *J* = 6.0, 5.4, 2.7 Hz, 2H), 6.93 (t, *J* = 8.7 Hz, 2H), 3.48 (t, *J* = 9.0 Hz, 1H), 2.52 – 2.46 (m, 1H), 2.43 – 2.35 (m, 1H), 2.20 (dtd, *J* = 11.4, 9.0, 4.0 Hz, 1H), 2.16 – 2.10 (m, 1H), 1.86 – 1.80 (m, 1H), 1.70 (dt, *J* = 13.3, 7.4 Hz, 1H), 0.84 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.73, 161.67 (d, *J* = 243 Hz), 136.07 (d, *J* = 3.0 Hz), 128.74 (d, *J* = 7.5 Hz), 114.85 (d, *J* = 21 Hz), 56.42, 49.54, 32.14, 25.14, 21.52, 9.09; ¹⁹F NMR (376 MHz, CDCl₃) δ -116.70.

HRMS (ESI-TOF) *m/z*. Calcd for C₁₃H₁₄FO₂⁻ [M-H]⁻ 221.0978, found 221.0979.

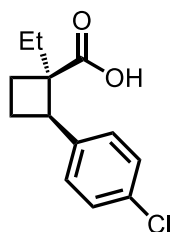
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.768	BB	0.1382	34.19260	3.55512	50.0155
2	6.414	BB	0.2279	34.17139	2.12114	49.9845



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	2.639	VB	0.0920	8.37479	1.35660	4.8913
2	6.111	BB	0.2422	162.84474	9.78151	95.1087



(1R,2R)-2-(4-chlorophenyl)-1-ethylcyclobutanecarboxylic acid (5f)

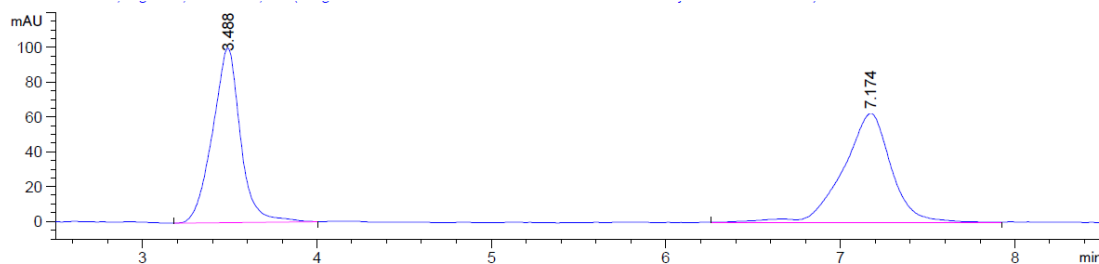
Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a white solid (63% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.581 min (minor) and 7.103 min (major), 92:8 er);

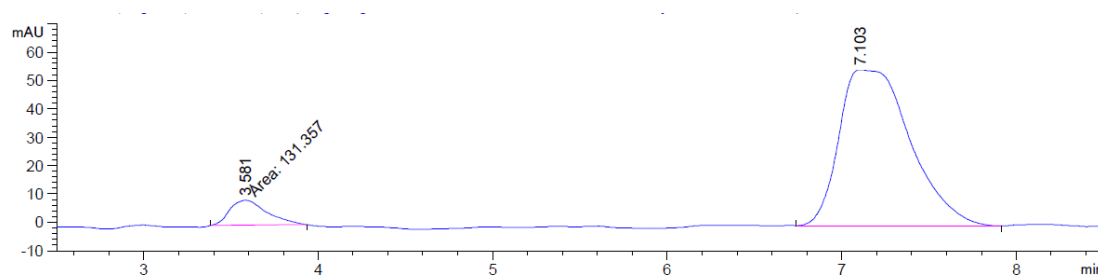
¹H NMR (600 MHz, CDCl₃) δ 7.22 (d, *J* = 8.5 Hz, 2H), 7.09 (d, *J* = 8.1 Hz, 2H), 3.48 (t, *J* = 9.0 Hz, 1H), 2.53 – 2.47 (m, 1H), 2.44 – 2.34 (m, 1H), 2.23 – 2.17 (m, 1H), 2.15 – 2.10 (m, 1H), 1.85 (dt, *J* = 11.4, 8.9 Hz, 1H), 1.75 – 1.67 (m, 1H), 0.85 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.68, 138.92, 132.33, 128.64, 128.20, 56.38, 49.58, 32.19, 25.22, 21.36, 9.10.

HRMS (ESI-TOF) *m/z*. Calcd for C₁₃H₁₄ClO₂ [M-H]⁻ 237.0682, found 237.0687.

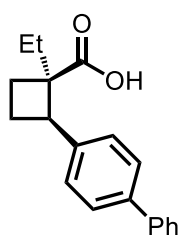
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.487	BB	0.1609	27.71568	2.47056	50.5590
2	7.174	BB	0.2517	27.10285	1.53749	49.4410



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.581	MM	0.2478	131.35725	8.83608	7.7650
2	7.103	BB	0.3747	1560.30310	55.04886	92.2350



(1R,2R)-2-([1,1'-biphenyl]-4-yl)-1-ethylcyclobutanecarboxylic acid (5g)

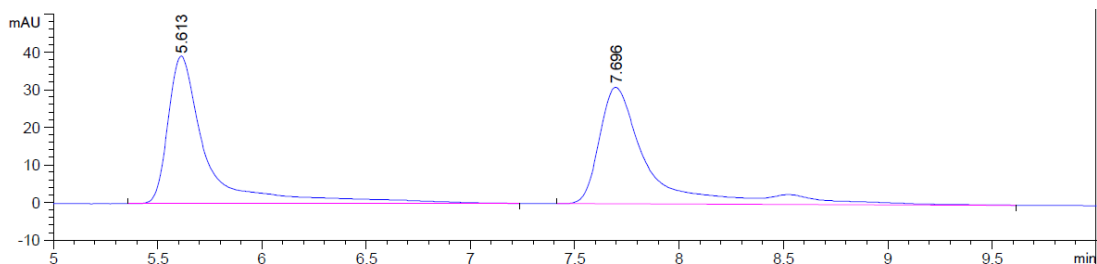
Substrate **4** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a white solid (62% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OD-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 5.720 min (major) and 7.662 min (minor), 95:5 er);

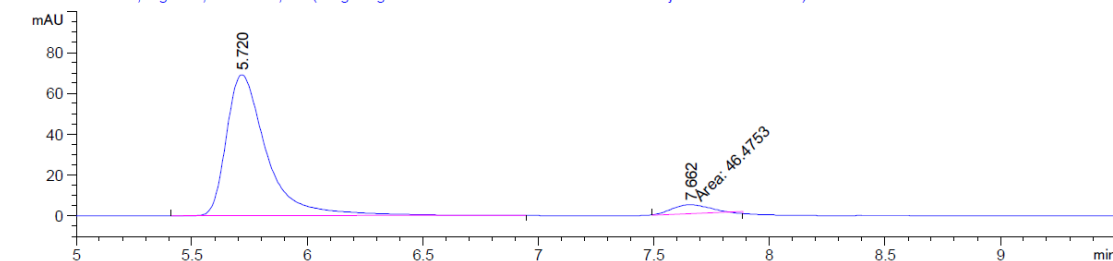
¹H NMR (600 MHz, CDCl₃) δ 7.56 – 7.54 (m, 2H), 7.48 (d, *J* = 8.3 Hz, 2H), 7.42 – 7.39 (m, 2H), 7.33 – 7.30 (m, 1H), 7.25 (d, *J* = 8.1 Hz, 2H), 3.55 (t, *J* = 8.9 Hz, 1H), 2.56 – 2.45 (m, 2H), 2.24 – 2.16 (m, 2H), 1.88 – 1.83 (m, 1H), 1.76 – 1.72 (m, 1H), 0.84 (t, *J* = 7.4 Hz, 3H);
¹³C NMR (150 MHz, CDCl₃) δ 178.01, 140.91, 139.58, 139.38, 128.66, 127.75, 127.05, 126.96, 126.77, 56.46, 50.02, 32.27, 25.33, 21.44, 9.16.

HRMS (ESI-TOF) *m/z* Calcd for C₁₉H₁₉O₂⁻ [M-H]⁻ 279.1385, found 279.1388.

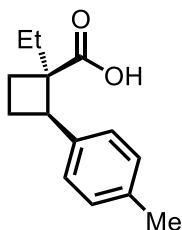
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.613	BB	0.1876	509.75204	39.23637	49.4105
2	7.696	BV R	0.2306	521.91632	30.98957	50.5895



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.720	BB	0.1814	836.39392	69.02428	94.7359
2	7.662	MM	0.1813	46.47529	4.27189	5.2641



(1R,2R)-1-ethyl-2-(p-tolyl)cyclobutanecarboxylic acid (5h)

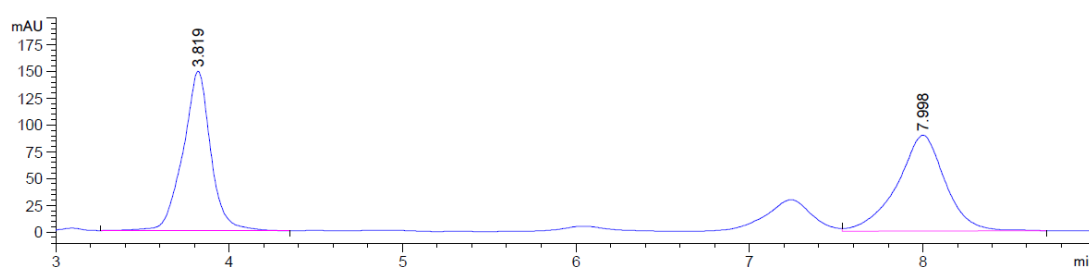
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a white solid (54% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.734 min (minor) and 7.674 min (major), 92:8 er);

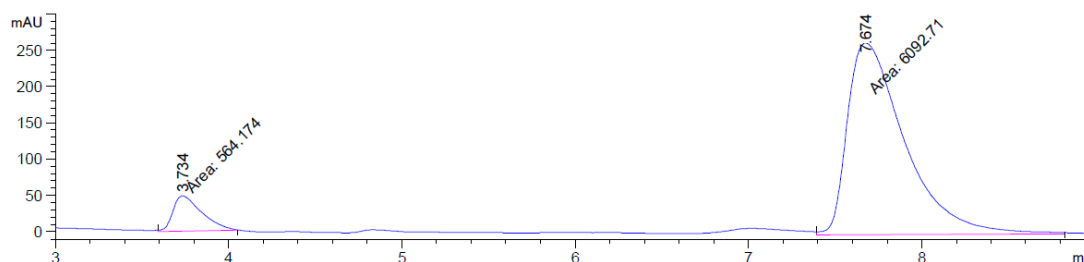
¹H NMR (600 MHz, CDCl₃) δ 7.06 (s, 4H), 3.47 (t, *J* = 9.0 Hz, 1H), 2.53 – 2.48 (m, 1H), 2.45 – 2.39 (m, 1H), 2.29 (s, 3H), 2.21 – 2.13 (m, 2H), 1.85 – 1.79 (m, 1H), 1.73 – 1.68 (m, 1H), 0.84 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.49, 137.38, 136.06, 128.80, 127.19, 56.47, 50.12, 32.22, 25.31, 21.55, 21.03, 9.13.

HRMS (ESI-TOF) m/z Calcd for $C_{14}H_{17}O_2^-$ [M-H]⁻ 217.1229, found 217.1226.

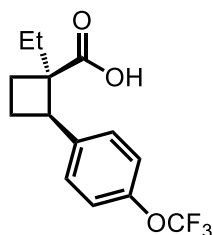
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.819	BB	0.1596	1652.83459	148.80081	49.0380
2	7.998	VB	0.2784	1717.68567	89.28466	50.9620



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.734	MM	0.1929	564.17365	48.75217	8.4750
2	7.674	MM	0.3841	6092.70898	264.39890	91.5250



(1R,2R)-1-ethyl-2-(4-(trifluoromethoxy)phenyl)cyclobutanecarboxylic acid (5i)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (56% yield).

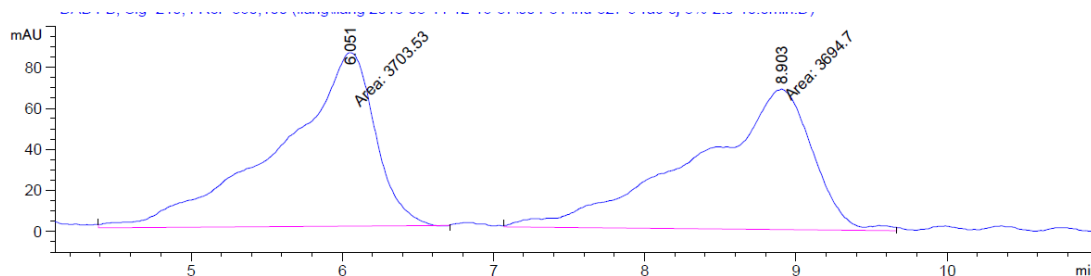
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 4% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 5.888 min (minor) and 8.670 min (major), 93:7 er);

¹H NMR (600 MHz, CDCl₃) δ 7.17 (d, *J* = 8.3 Hz, 2H), 7.09 (d, *J* = 7.8 Hz, 2H), 3.51 (t, *J* =

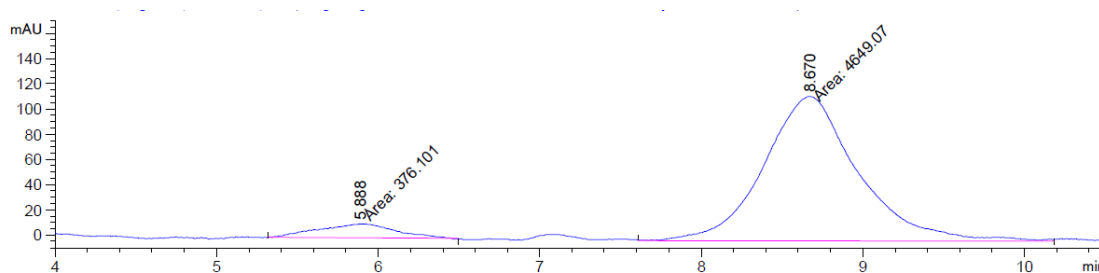
9.0 Hz, 1H), 2.52 – 2.46 (m, 1H), 2.40 (dtd, $J = 11.5, 9.5, 8.5$ Hz, 1H), 2.25 – 2.19 (m, 1H), 2.17 – 2.11 (m, 1H), 1.88 – 1.83 (m, 1H), 1.74 – 1.69 (m, 1H), 0.84 (t, $J = 7.4$ Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 178.86, 147.89, 139.15, 128.58, 120.50, 120.47 (q, $J = 255$ Hz), 56.41, 49.48, 32.15, 25.19, 21.41, 9.01; ^{19}F NMR (376 MHz, CDCl_3) δ -58.14.

HRMS (ESI-TOF) m/z . Calcd for $\text{C}_{14}\text{H}_{14}\text{F}_3\text{O}_3^-$ [M-H] $^-$ 287.0895, found 287.0898.

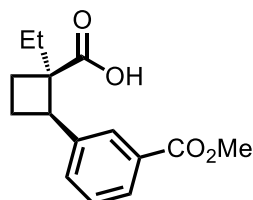
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.051	MM	0.7287	3703.52539	84.70240	50.0597
2	8.903	MM	0.8979	3694.69897	68.58344	49.9403



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.888	MM	0.5542	376.10083	11.30990	7.4843
2	8.670	MM	0.6762	4649.07373	114.59574	92.5157



(1R,2R)-1-ethyl-2-(3-(methoxycarbonyl)phenyl)cyclobutanecarbox (5j)

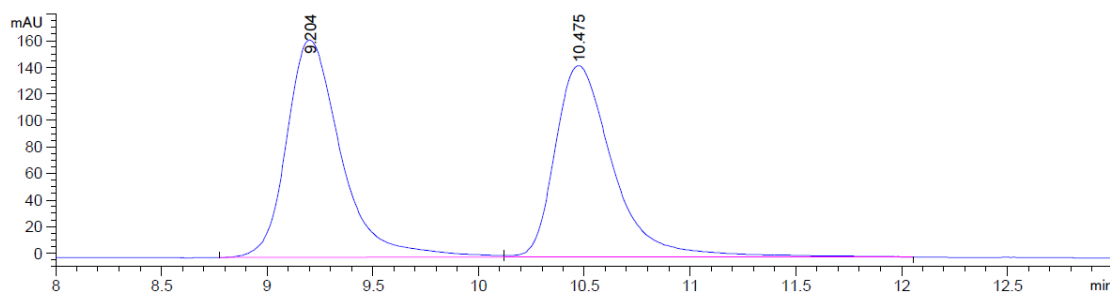
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (64% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 10% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 9.159 min (major) and 10.557 min (minor), 89:11 er);

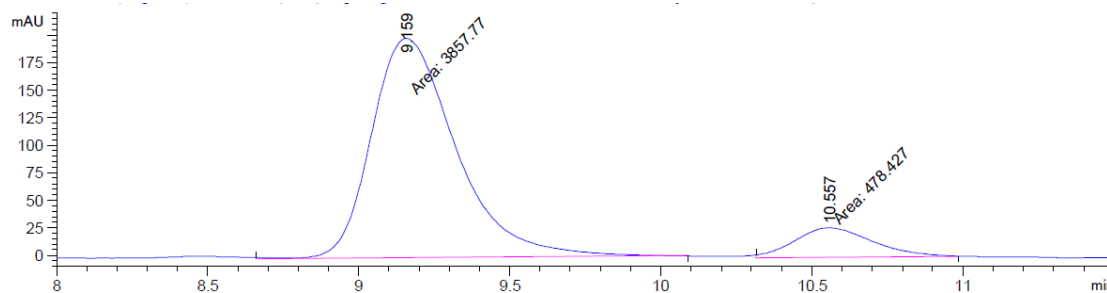
¹H NMR (600 MHz, CDCl₃) δ 7.85 (d, *J* = 7.7 Hz, 1H), 7.82 (s, 1H), 7.35 (d, *J* = 7.7 Hz, 1H), 7.29 (t, *J* = 7.7 Hz, 1H), 3.90 (s, 3H), 3.55 (t, *J* = 8.8 Hz, 1H), 2.53 – 2.43 (m, 2H), 2.25 – 2.19 (m, 1H), 2.15 (dt, *J* = 14.5, 7.2 Hz, 1H), 1.89 – 1.84 (m, 1H), 1.75 – 1.70 (m, 1H), 0.83 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.60, 167.17, 140.78, 131.73, 129.87, 128.59, 128.12, 127.92, 56.39, 52.05, 49.86, 32.20, 25.25, 21.23, 9.09.

HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₁₇O₄ [M-H]⁻ 261.1127, found 261.1130.

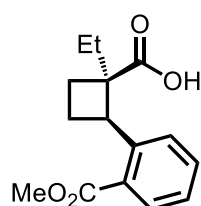
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.204	BV	0.2686	2927.27808	163.69240	51.5808
2	10.475	VB	0.2883	2747.85449	144.24396	48.4192



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.159	MM	0.3229	3857.77026	199.13843	88.9667
2	10.557	MM	0.2999	478.42657	26.58735	11.0333



(1R,2R)-1-ethyl-2-(2-(methoxycarbonyl)phenyl)cyclobutanecarboxylic acid (5k)

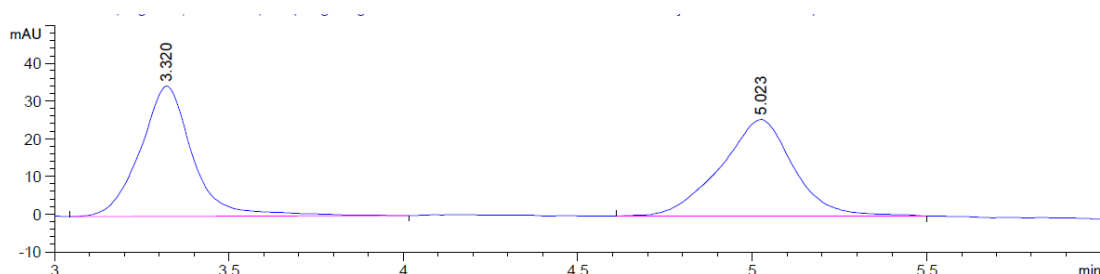
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (60% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.181 min (minor) and 4.768 min (major), 92:8 er);

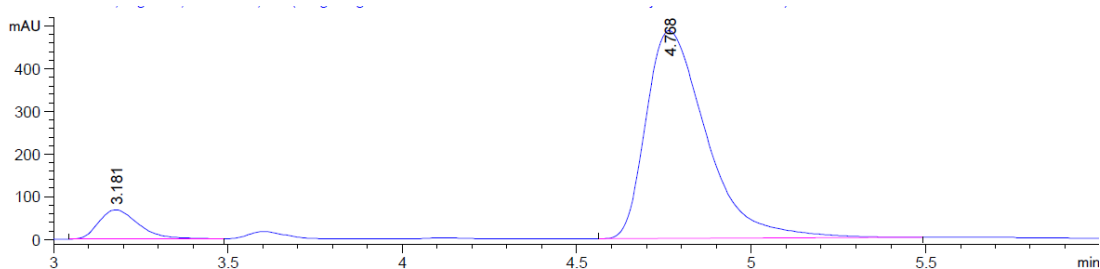
¹H NMR (600 MHz, CDCl₃) δ 7.69 (dd, *J* = 7.8, 1.5 Hz, 1H), 7.62 – 7.59 (m, 1H), 7.54 (td, *J* = 7.6, 1.4 Hz, 1H), 7.28 (td, *J* = 7.6, 1.2 Hz, 1H), 3.96 (s, 3H), 3.91 – 3.87 (m, 1H), 2.80 (d, *J* = 12.1 Hz, 1H), 2.42 – 2.38 (m, 1H), 2.25 – 2.20 (m, 2H), 1.97 – 1.92 (m, 2H), 0.92 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 174.90, 171.37, 141.55, 132.75, 130.48, 129.59, 127.18, 126.72, 55.95, 53.03, 45.81, 31.05, 25.13, 21.05, 9.07.

HRMS (ESI-TOF) *m/z* Calcd for C₁₅H₁₇O₄⁻ [M-H]⁻ 261.1127, found 261.1122.

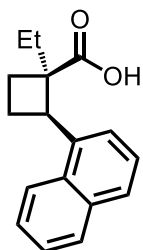
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.320	BB	0.1505	363.30420	34.61173	49.2798
2	5.023	BB	0.2119	373.92389	25.67123	50.7202



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.181	BV	0.1234	549.15814	68.62186	8.5895
2	4.768	BB	0.1839	5844.23730	487.40265	91.4105



(1R,2R)-1-ethyl-2-(naphthalen-1-yl)cyclobutanecarboxylic acid (5l)

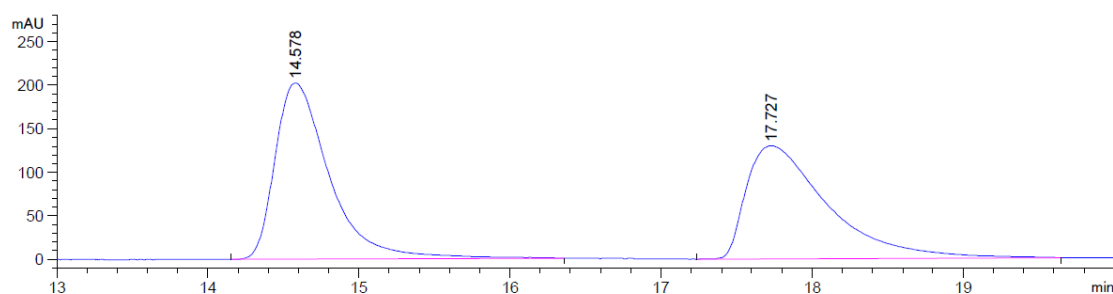
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (56% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AD-3 column, 10% ⁱPrOH / CO₂, flow rate 1 mL/min, retention time 14.396 min (major) and 17.263 min (minor), 92:8 er);

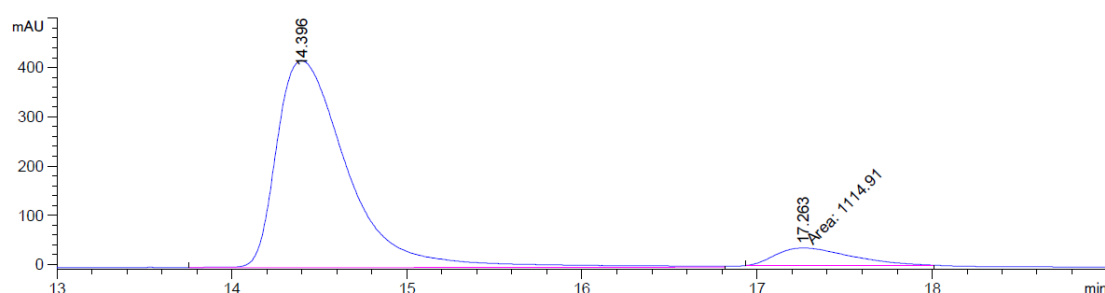
¹H NMR (600 MHz, CDCl₃) δ 8.08 (d, *J* = 8.4 Hz, 1H), 7.81 (d, *J* = 8.0 Hz, 1H), 7.68 (d, *J* = 8.1 Hz, 1H), 7.50 – 7.43 (m, 2H), 7.41 (d, *J* = 7.1 Hz, 1H), 7.36 (t, *J* = 7.6 Hz, 1H), 4.40 (t, *J* = 9.1 Hz, 1H), 2.76 – 2.67 (m, 1H), 2.57 – 2.51 (m, 1H), 2.37 – 2.32 (m, 1H), 2.20 (dtd, *J* = 11.5, 8.6, 3.1 Hz, 1H), 1.96 – 1.88 (m, 2H), 0.85 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.08, 136.09, 133.64, 132.39, 128.64, 127.25, 125.67, 125.36, 125.28, 123.82, 123.66, 57.96, 45.41, 32.49, 24.95, 20.91, 9.38.

HRMS (ESI-TOF) *m/z* Calcd for C₁₇H₁₇O₂ [M-H]⁻ 253.1229, found 253.1225.

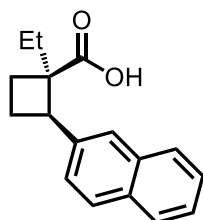
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.578	BB	0.3782	5065.38818	202.24588	51.1350
2	17.727	BB	0.5496	4840.53320	129.64720	48.8650



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.396	BV	0.4351	1.18436e4	419.91861	91.3963
2	17.263	MM	0.5120	1114.90845	36.29034	8.6037



(1R,2R)-1-ethyl-2-(naphthalen-2-yl)cyclobutanecarboxylic acid (5m)

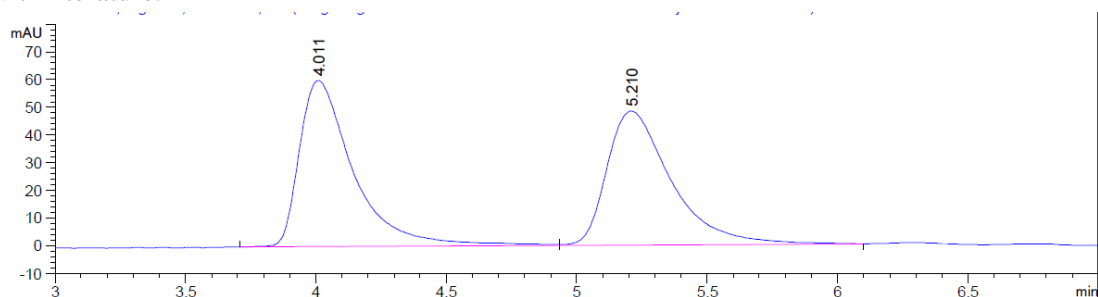
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (58% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IC-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.941 min (major) and 5.166 min (minor), 92:8 er);

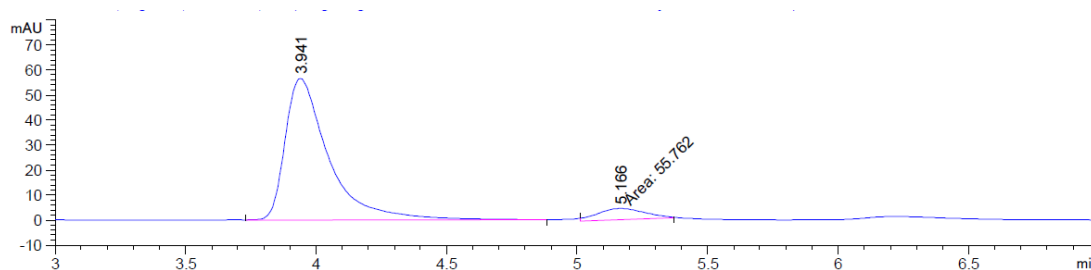
¹H NMR (600 MHz, CDCl₃) δ 7.78 – 7.74 (m, 2H), 7.66 (d, *J* = 8.5 Hz, 1H), 7.58 (s, 1H), 7.45 – 7.38 (m, 2H), 7.27 (dd, *J* = 8.5, 1.8 Hz, 1H), 3.65 (t, *J* = 8.8 Hz, 1H), 2.59 – 2.48 (m, 2H), 2.25 (ddd, *J* = 10.9, 2.9, 1.4 Hz, 1H), 2.20 – 2.15 (m, 1H), 1.89 – 1.83 (m, 1H), 1.75 (dt, *J* = 13.4, 7.4 Hz, 1H), 0.77 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.20, 138.03, 133.29, 132.39, 127.81, 127.54, 127.51, 126.14, 125.79, 125.52, 125.35, 56.50, 50.38, 32.33, 25.36, 21.35, 9.06.

HRMS (ESI-TOF) *m/z* Calcd for C₁₇H₁₇O₂⁻ [M-H]⁻ 253.1229, found 253.1227.

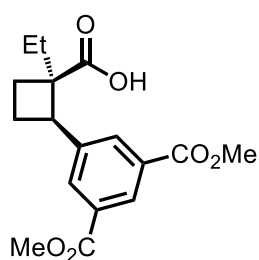
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.011	BV	0.2155	861.07312	59.99102	50.9482
2	5.210	VB	0.2598	829.02124	48.40171	49.0518



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.941	BB	0.1709	653.11957	56.48904	92.1338
2	5.166	MM	0.2074	55.76199	4.48052	7.8662



(1R,2R)-2-(3,5-bis(methoxycarbonyl)phenyl)-1-ethylcyclobutanecarboxylic acid (5n)

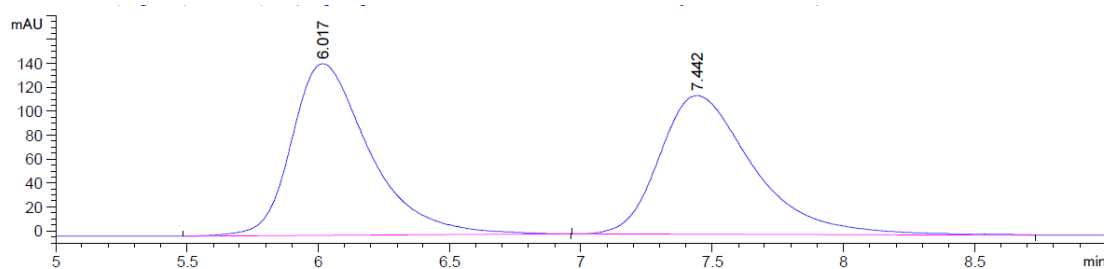
Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (60% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IC-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 5.983 min (major) and 7.467 min (minor), 92:8 er);

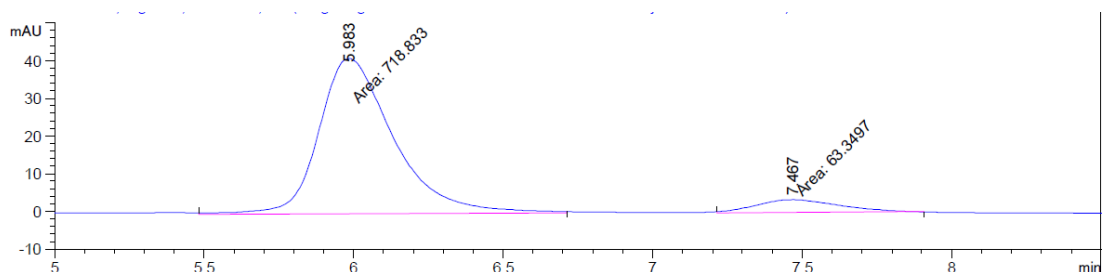
¹H NMR (600 MHz, CDCl₃) δ 8.49 (t, *J* = 1.6 Hz, 1H), 8.02 (dd, *J* = 1.6, 0.6 Hz, 2H), 3.92 (s, 6H), 3.60 (t, *J* = 8.5 Hz, 1H), 2.56 – 2.48 (m, 2H), 2.28 – 2.22 (m, 1H), 2.20 – 2.13 (m, 1H), 1.92 – 1.86 (m, 1H), 1.77 – 1.71 (m, 1H), 0.82 (t, *J* = 7.4 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 178.03, 166.28, 141.46, 132.71, 130.34, 129.09, 56.43, 52.30, 49.48, 32.17, 25.26, 21.18, 9.01.

HRMS (ESI-TOF) *m/z* Calcd for C₁₇H₁₉O₆⁻ [M-H]⁻ 319.1182, found 319.1180.

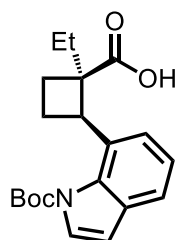
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.017	BB	0.3138	2983.80957	143.97331	50.7996
2	7.442	BB	0.3782	2889.87183	116.17687	49.2004



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.983	MM	0.2907	718.83331	41.21288	91.9009
2	7.467	MM	0.3175	63.34967	3.32499	8.0991



(1R,2R)-2-(1-(tert-butoxycarbonyl)-1H-indol-7-yl)-1-ethylcyclobutanecarboxylic acid (5o)

Substrate **1** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow solid (42% yield).

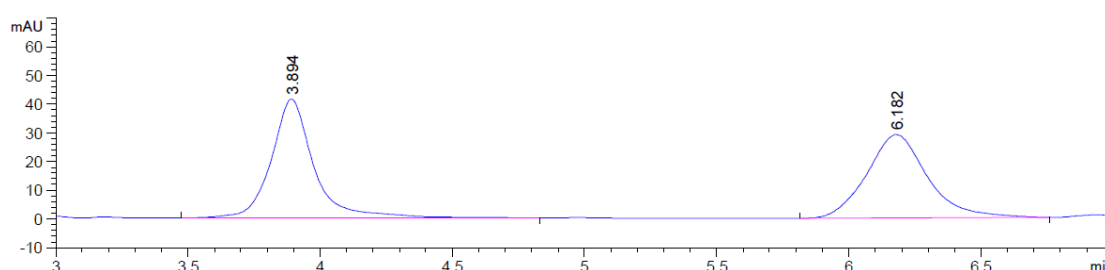
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 3.688 min (minor) and 5.836 min (major), 91:9 er);

¹H NMR (600 MHz, CDCl₃) δ 7.99 (s, 1H), 7.53 (d, *J* = 3.5 Hz, 1H), 7.36 (dd, *J* = 1.7, 0.8 Hz, 1H), 7.13 (d, *J* = 8.6 Hz, 1H), 6.49 (dd, *J* = 3.7, 0.8 Hz, 1H), 3.61 (t, *J* = 8.6 Hz, 1H), 2.53 – 2.51 (m, 1H), 2.25 – 2.20 (m, 1H), 2.20 – 2.16 (m, 1H), 1.88 – 1.79 (m, 2H), 1.74 – 1.71 (m,

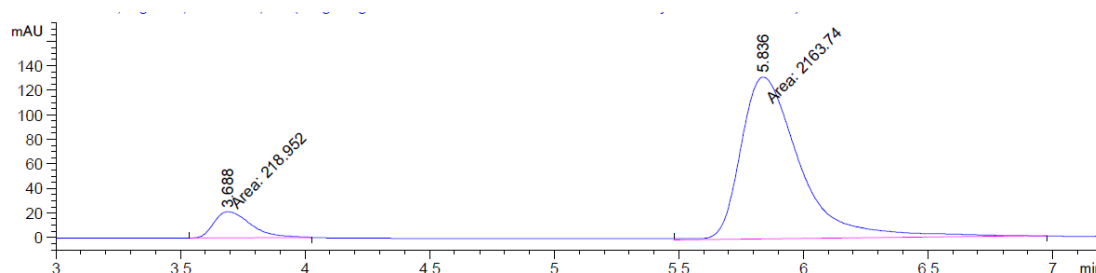
1H), 1.66 (s, 9H), 0.81 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (150 MHz, CDCl_3) δ 182.67, 178.39, 149.78, 134.81, 130.53, 125.89, 124.05, 119.25, 114.65, 107.37, 56.65, 50.40, 32.23, 29.41, 28.18, 25.26, 21.69, 9.11.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{20}\text{H}_{24}\text{NO}_4$ $[\text{M}-\text{H}]^-$ 342.1705, found 342.1700.

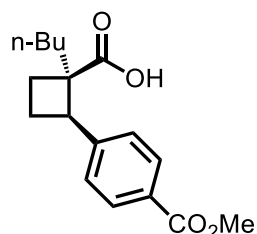
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.894	BB	0.1610	466.13464	41.49893	50.1091
2	6.182	BB	0.2362	464.10437	29.07586	49.8909



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.688	MM	0.1705	218.95163	21.39905	9.1893
2	5.836	MM	0.2735	2163.74097	131.86678	90.8107



(1R,2R)-1-butyl-2-(4-(methoxycarbonyl)phenyl)cyclobutanecarboxylic acid (5p)

Substrate **4p** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (60% yield).

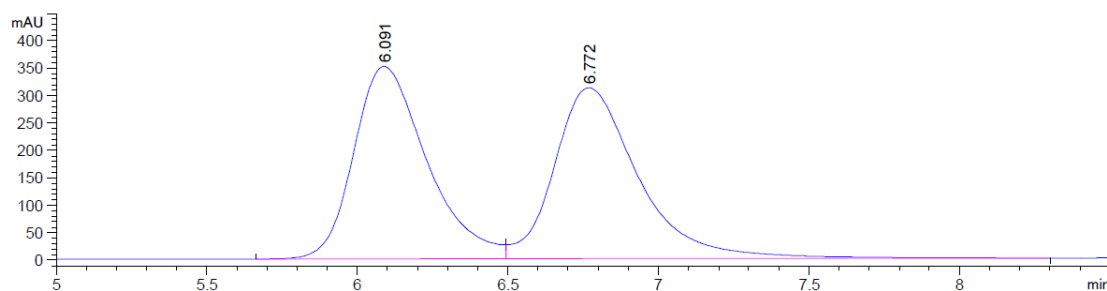
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK®)

IC-3 column, 10% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 5.918 min (major) and 6.620 min (minor), 86:14 er);

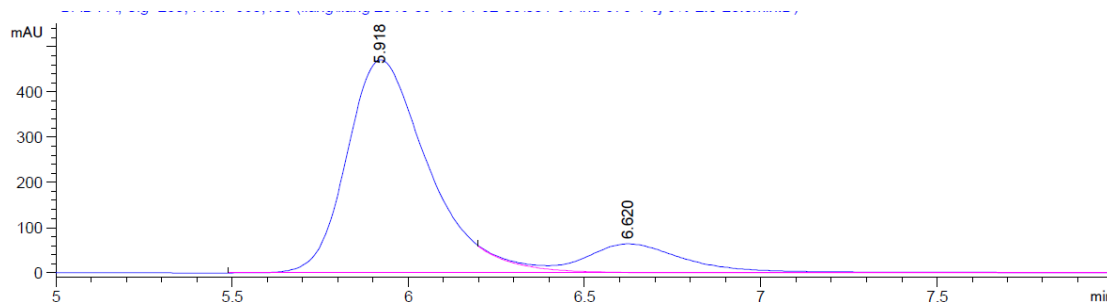
¹H NMR (600 MHz, CDCl₃) δ 7.95 (d, *J* = 8.4 Hz, 2H), 7.24 (s, 2H), 3.90 (s, 3H), 3.78 – 3.71 (m, 2H), 3.15 – 3.09 (m, 1H), 2.71 – 2.67 (m, 1H), 2.16 – 2.11 (m, 2H), 1.61 (p, *J* = 7.7 Hz, 2H), 1.47 (dt, *J* = 14.6, 7.3 Hz, 2H), 1.26 – 1.24 (m, 1H), 1.00 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 173.68, 167.01, 145.95, 129.40, 128.25, 126.91, 61.87, 52.02, 43.98, 38.44, 28.13, 27.02, 23.37, 14.04.

HRMS (ESI-TOF) *m/z* Calcd for C₁₇H₂₁O₄⁻ [M-H]⁻ 289.1440, found 289.1436.

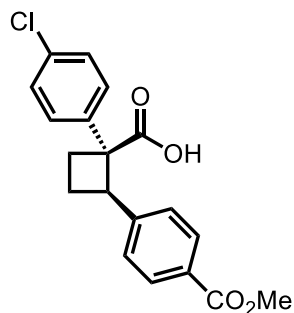
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.091	BV	0.2566	5919.68555	351.25800	49.4354
2	6.772	VB	0.2928	6054.89697	311.50253	50.5646



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.918	BV R	0.2421	7595.69092	471.12335	85.4971
2	6.620	VB E	0.2998	1288.46021	63.22224	14.5029



(1R,2R)-1-(4-chlorophenyl)-2-(4-(methoxycarbonyl)phenyl)cyclobutanecarboxylic acid (5q)

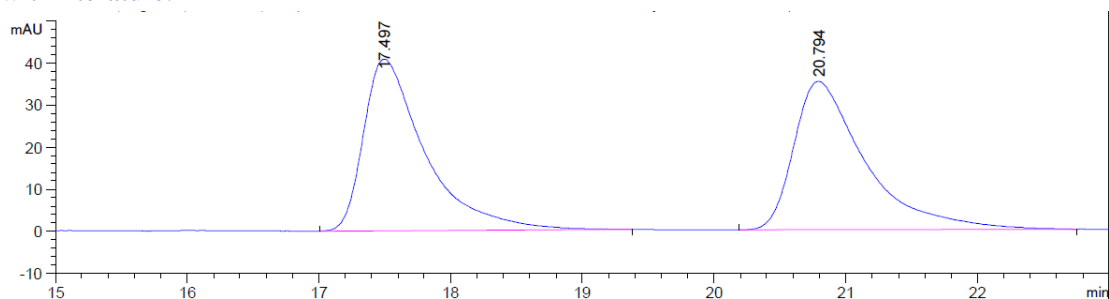
Substrate **4q** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (53% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 16.200 min (minor) and 19.175 min (major), 94:6 er);

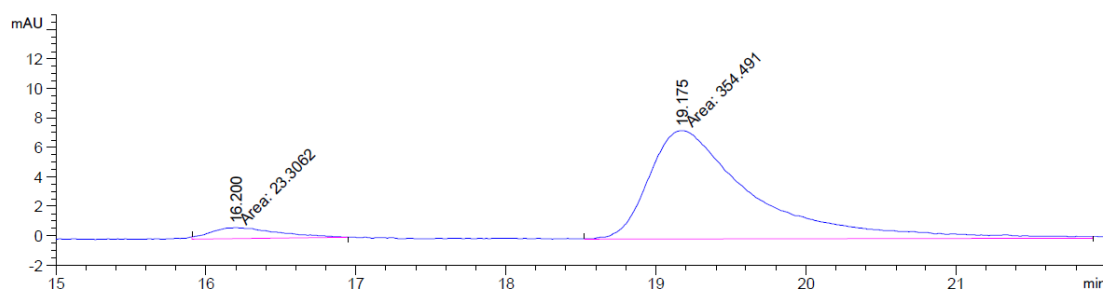
¹H NMR (600 MHz, CDCl₃) δ 8.00 (d, *J* = 8.1 Hz, 2H), 7.41 (d, *J* = 8.2 Hz, 2H), 7.33 (d, *J* = 6.6 Hz, 2H), 7.25 (d, *J* = 8.5 Hz, 2H), 4.17 (t, *J* = 9.4 Hz, 1H), 3.92 (s, 3H), 2.99 – 2.93 (m, 1H), 2.69 – 2.63 (m, 1H), 2.38 – 2.30 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 175.96, 166.91, 145.48, 142.42, 132.91, 129.73, 129.07, 128.52, 127.91, 127.74, 58.68, 52.12, 50.13, 28.68, 23.15.

HRMS (ESI-TOF) *m/z* Calcd for C₁₉H₁₆ClO₄⁻ [M-H]⁻ 343.0737, found 343.0734.

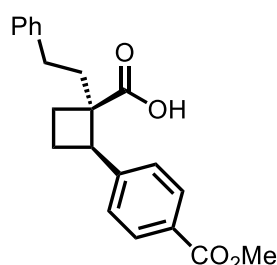
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.497	BB	0.4760	1321.17957	40.77972	49.8186
2	20.794	BB	0.5283	1330.80139	35.26552	50.1814



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.200	MM	0.5180	23.30623	7.49925e-1	6.1690
2	19.175	MM	0.8021	354.49060	7.36629	93.8310



(1S,2R)-2-(4-(methoxycarbonyl)phenyl)-1-phenethylcyclobutanecarboxylic acid (5r)

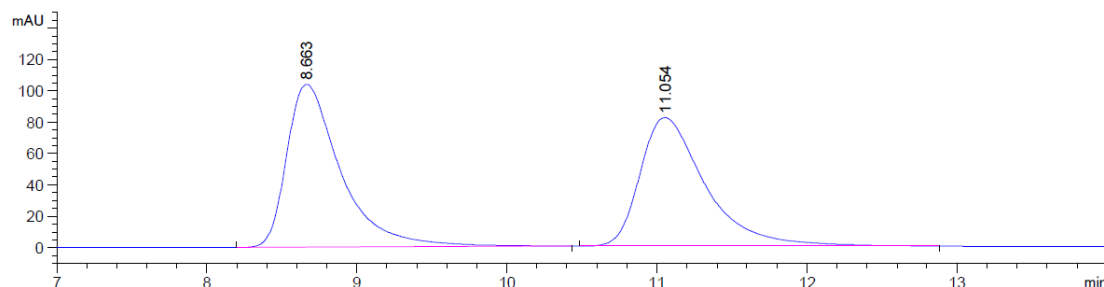
Substrate **4r** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a pale-yellow oil (65% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IC-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 9.028 min (major) and 11.620 min (minor), 93:7 er);

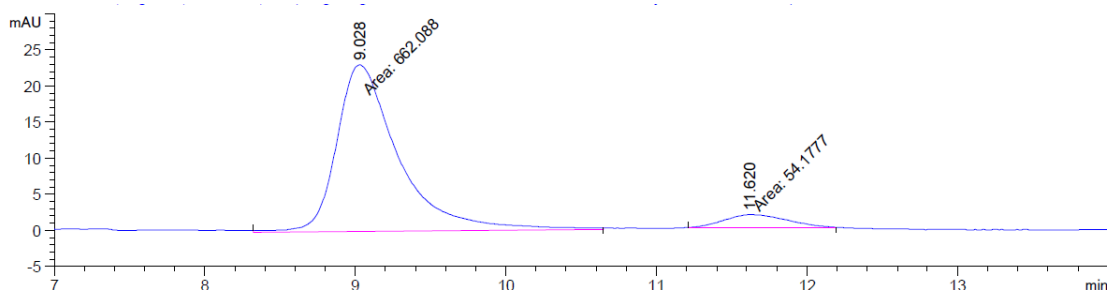
¹H NMR (600 MHz, CDCl₃) δ 7.93 (d, *J* = 8.4 Hz, 2H), 7.29 (t, *J* = 7.6 Hz, 2H), 7.24 – 7.15 (m, 5H), 3.79 (s, 3H), 3.60 (t, *J* = 8.6 Hz, 1H), 2.62 – 2.42 (m, 5H), 2.27 – 2.20 (m, 1H), 2.03 – 1.89 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 178.42, 166.98, 145.32, 141.68, 129.44, 128.56, 128.42, 128.37, 127.32, 125.95, 55.97, 51.92, 50.56, 41.80, 31.41, 26.01, 21.33.

HRMS (ESI-TOF) *m/z* Calcd for C₂₁H₂₁O₄⁻ [M-H]⁻ 337.1440, found 337.1438.

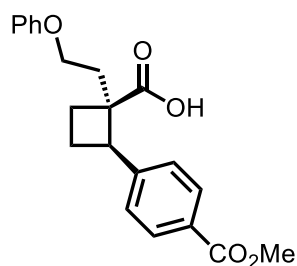
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.663	BB	0.3714	2609.29785	103.73867	51.2837
2	11.054	BB	0.4535	2478.66602	81.79804	48.7163



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.028	MM	0.4777	662.08783	23.09881	92.4361
2	11.620	MM	0.4931	54.17773	1.83108	7.5639



(1S,2R)-2-(4-(methoxycarbonyl)phenyl)-1-(2-phenoxyethyl)cyclobutanecarboxylic acid (5s)

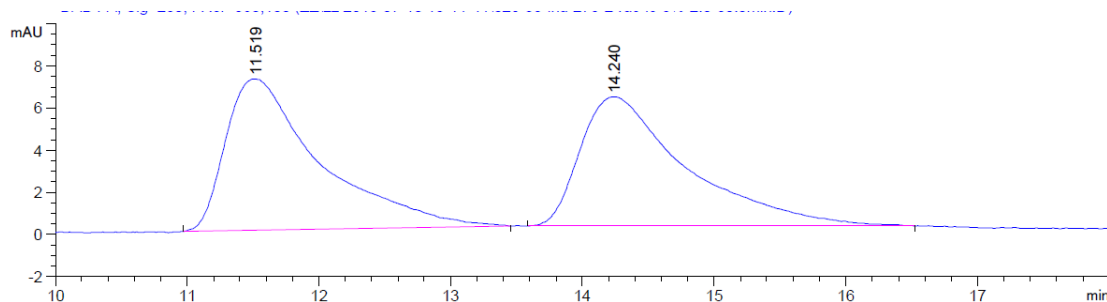
Substrate **4s** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (52% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IC-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 11.010 min (major) and 13.789 min (minor), 84:16 er);

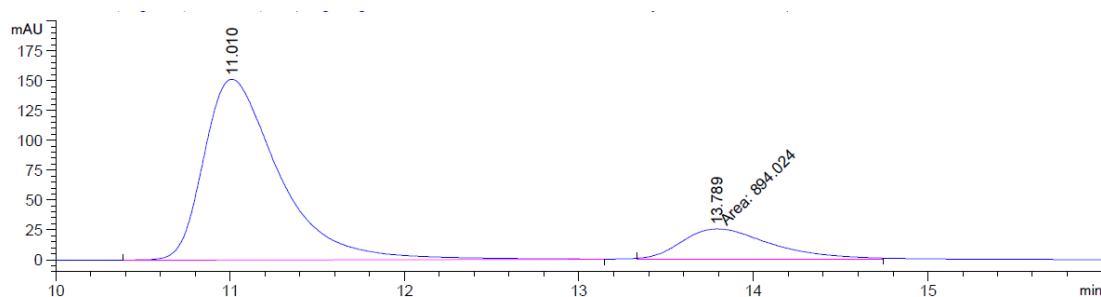
¹H NMR (600 MHz, CDCl₃) δ 7.94 (d, *J* = 8.1 Hz, 2H), 7.26 – 7.22 (m, 4H), 6.95 – 6.90 (m, 1H), 6.84 (d, *J* = 7.8 Hz, 2H), 4.03 – 3.94 (m, 2H), 3.87 (s, 3H), 3.73 (t, *J* = 9.0 Hz, 1H), 2.68 – 2.53 (m, 3H), 2.26 (td, *J* = 7.1, 3.2 Hz, 2H), 2.15 – 2.07 (m, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 177.97, 167.02, 158.53, 145.11, 129.48, 129.44, 128.65, 127.31, 120.76, 114.36, 64.31, 52.01, 50.36, 38.15, 26.23, 24.86, 21.76.

HRMS (ESI-TOF) *m/z* Calcd for C₂₁H₂₂NaO₅⁺ [M+Na]⁺ 377.1365, found 377.1369.

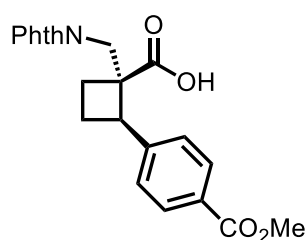
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.519	BB	0.5852	357.24802	7.19614	51.0097
2	14.240	BB	0.6653	343.10483	6.14598	48.9903



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.010	BB	0.4352	4528.30762	151.28094	83.5122
2	13.789	MM	0.5916	894.02399	25.18769	16.4878



(1R,2R)-1-((1,3-dioxoisindolin-2-yl)methyl)-2-(4-(methoxycarbonyl)phenyl)cyclobutane carboxylic acid (5t)

Substrate **4t** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (57% yield).

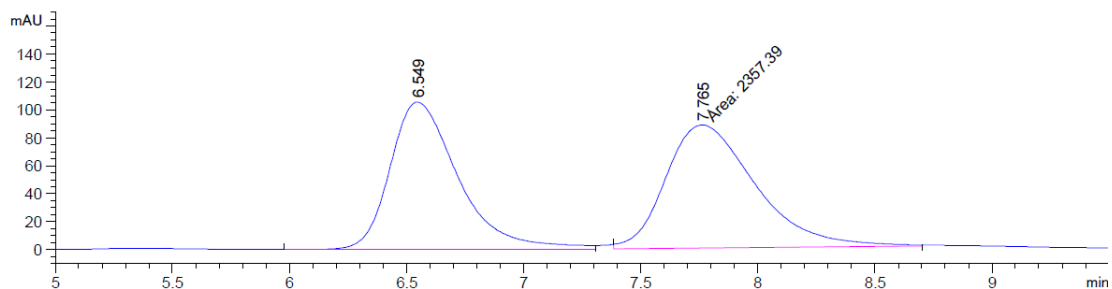
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 6.848 min (minor) and 8.144 min (major), 93:7 er);

¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, *J* = 7.1 Hz, 2H), 7.90 (dd, *J* = 5.4, 3.1 Hz, 2H), 7.77 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.44 (d, *J* = 6.5 Hz, 2H), 4.20 (d, *J* = 15.3 Hz, 1H), 4.01 (d, *J* = 12.8

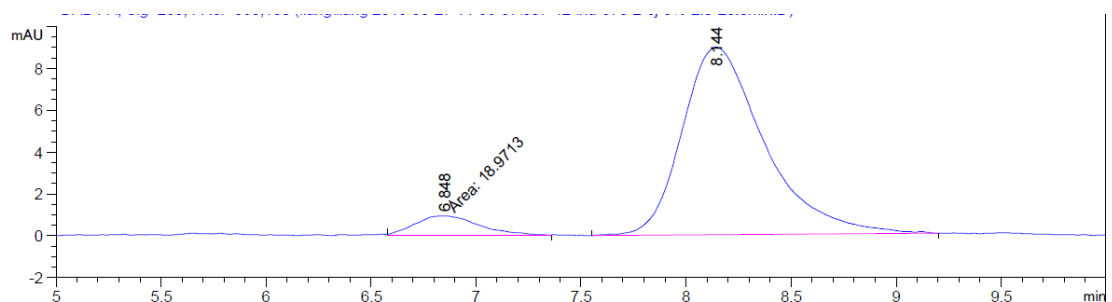
Hz, 1H), 3.89 (s, 3H), 3.81 – 3.76 (m, 1H), 2.61 – 2.55 (m, 1H), 2.41 (t, $J = 10.2$ Hz, 1H), 2.09 (dd, $J = 18.4, 9.5$ Hz, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 185.59, 168.98, 167.10, 144.93, 134.31, 131.88, 129.52, 128.65, 127.47, 123.59, 123.45, 52.01, 45.70, 29.69, 29.21, 23.79, 21.18.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{22}\text{H}_{18}\text{NO}_6$ $[\text{M}-\text{H}]^-$ 392.1134, found 392.1127.

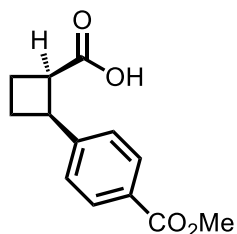
The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.549	BV	0.3108	2141.92163	105.52997	47.6055
2	7.765	MM	0.4455	2357.38965	88.19743	52.3945



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.848	MM	0.3437	18.97134	9.19975e-1	7.1862
2	8.144	BB	0.4041	245.02602	8.98688	92.8138



(1R,2S)-2-(4-(methoxycarbonyl)phenyl)cyclobutanecarboxylic acid (5u**)**

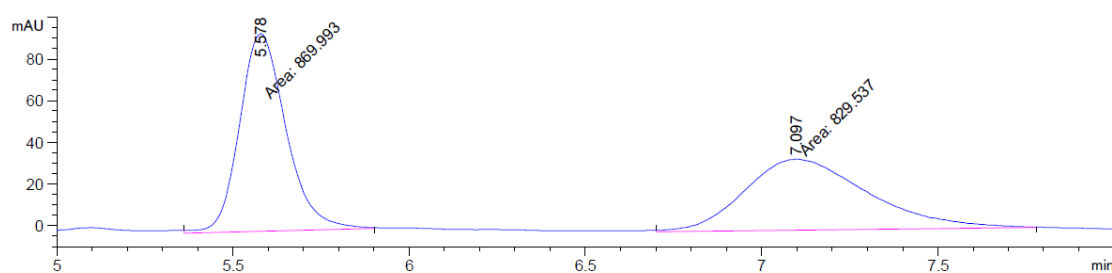
Substrate **4u** was arylated following the general arylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a white solid (61% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 15% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 5.554 min (major) and 7.770 min (minor), 92:8 er);

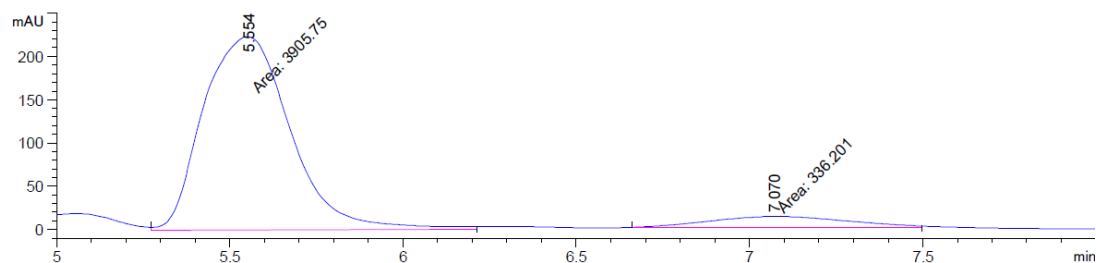
¹H NMR (600 MHz, CDCl₃) δ 7.94 (d, *J* = 8.4 Hz, 2H), 7.25 (d, *J* = 8.2 Hz, 2H), 4.04 – 3.98 (m, 1H), 3.90 (s, 3H), 3.54 (q, *J* = 3.2, 2.4 Hz, 1H), 2.65 – 2.57 (m, 1H), 2.36 – 2.28 (m, 2H), 2.28 – 2.21 (m, 1H); ¹³C NMR (151 MHz, CDCl₃) δ 177.78, 167.09, 146.06, 129.50, 128.36, 127.13, 52.00, 44.56, 42.27, 24.44, 20.53.

HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₃O₄⁻ [M-H]⁻ 233.0814, found 233.0806.

The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.

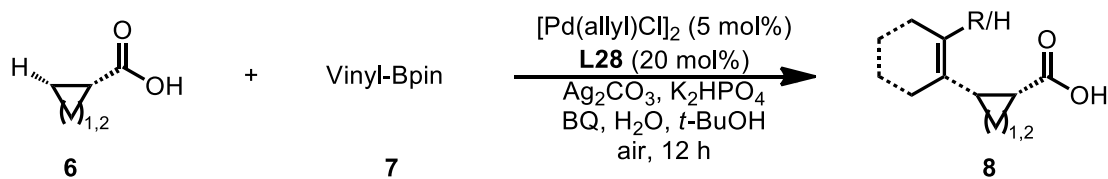


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.578	MM	0.1528	869.99335	94.89941	51.1902
2	7.097	MM	0.4055	829.53741	34.09639	48.8098

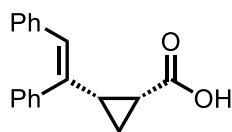


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.554	MM	0.2910	3905.75220	223.69759	92.0744
2	7.070	MM	0.4548	336.20142	12.32098	7.9256

3.7 Enantioselective Vinylation of Cyclopropanecarboxylic acid and Cyclobutanecarboxylic acid



General procedure for enantioselective Vinylation of cyclopropanecarboxylic acid and cyclobutanecarboxylic acid: A 2-dram vial equipped with a magnetic stir bar was charged with $[\text{Pd}(\text{allyl})\text{Cl}]_2$ (1.9mg, 5 mol%) and **L28** (11.7 mg, 20 mol%) in *t*-BuOH (1.0 ml) and then stirred at the rate of 300 rpm at room temperature for 5 min. The appropriate acid substrate (0.10 mmol), Ag_2CO_3 (41.4 mg, 0.15 mmol), K_2HPO_4 (26.0 mg, 0.15 mmol), Vinyl-Bpin (0.2 mmol), BQ (5.4 mg, 0.05 mmol), H_2O (3.6 mg, 0.2 mmol) was then added. Subsequently the vial was capped and closed tightly. The reaction mixture was then stirred at the rate of 300 rpm at 80 °C for 12 h. After being allowed to cool to room temperature, the mixture was diluted with ethyl acetate and 0.1 ml of acetic acid was then added. The mixture was passed through a pad of Celite with ethyl acetate as the eluent to remove any insoluble precipitate. The resulting solution was concentrated, and the residual mixture was dissolved with a minimal amount of acetone and loaded onto a preparative TLC plate. The pure product was then isolated using preparative TLC with ethyl acetate and hexane (1/4 to 1/1) as the eluent and 1% v/v of acetic acid as additive.



(1R,2S)-2-((Z)-1,2-diphenylvinyl)cyclopropanecarboxylic acid (8a)

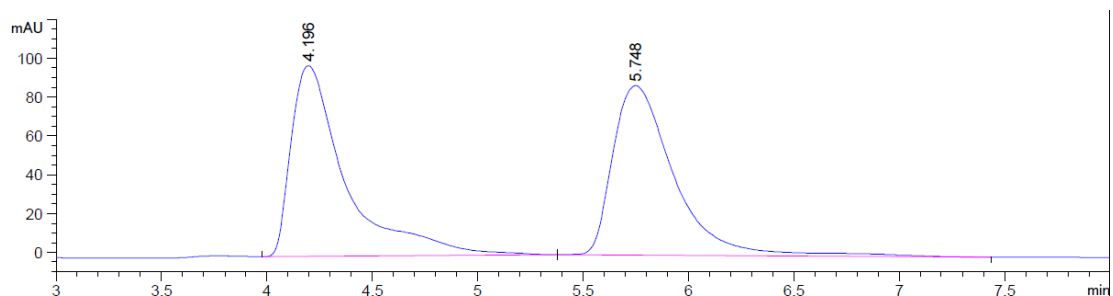
Substrate **6a** was vinylated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (71% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IC-3 column, 10% *i*PrOH / CO_2 , flow rate 2 mL/min, retention time 4.352 min (minor) and 5.836 min (major), 98:2 er);

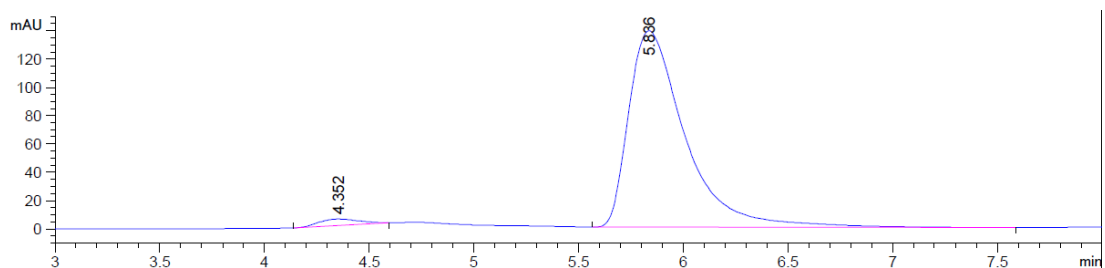
^1H NMR (600 MHz, CDCl_3) δ 7.18 – 7.11 (m, 5H), 7.07 (dt, $J = 4.9, 1.8$ Hz, 3H), 6.96 – 6.90 (m, 2H), 6.55 (s, 1H), 2.38 (q, $J = 8.8, 8.1$ Hz, 1H), 2.05 – 2.00 (m, 1H), 1.70 (dt, $J = 7.9, 5.3$ Hz, 1H), 1.31 (td, $J = 8.0, 5.1$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 176.52, 140.97, 136.93, 135.51, 129.77, 129.28, 128.61, 128.41, 127.83, 127.05, 126.49, 30.91, 21.45, 12.34.

HRMS (ESI-TOF) m/z . Calcd for $\text{C}_{18}\text{H}_{15}\text{O}_2^-$ [M-H] $^-$ 263.1072, found 263.1068.

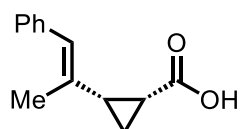
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.196	BB	0.2682	1770.29736	98.25798	49.8954
2	5.748	BB	0.3115	1777.71936	87.32085	50.1046



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	4.352	BB	0.2122	60.82872	4.54320	2.2811
2	5.836	BB	0.2808	2605.83105	138.94025	97.7189



(1R,2S)-2-((E)-1-phenylprop-1-en-2-yl)cyclopropanecarboxylic acid (8b)

Substrate **6b** was vinyated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a white solid (70% yield).

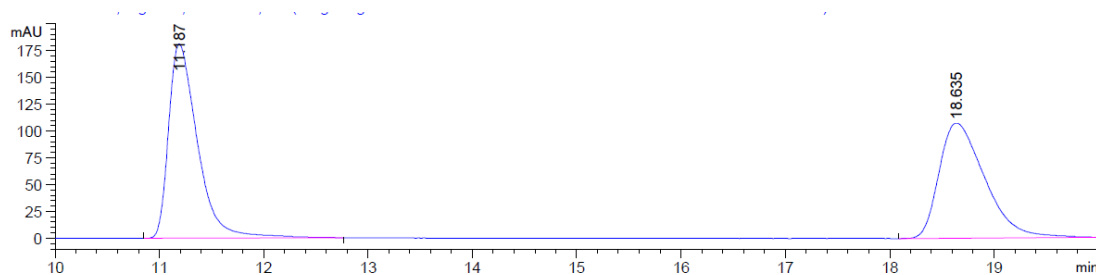
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% ⁱPrOH / CO₂, flow rate 2 mL/min, retention time 11.153 min (major) and 18.721 min (minor), 97:3 er);

¹H NMR (600 MHz, CDCl₃) δ 7.28 (dd, *J* = 8.2, 7.1 Hz, 2H), 7.21 – 7.16 (m, 3H), 6.43 (s, 1H), 2.19 – 2.14 (m, 1H), 1.96 – 1.91 (m, 1H), 1.86 (s, 3H), 1.58 (dt, *J* = 7.7, 5.2 Hz, 1H), 1.23 (td, *J* = 8.0, 5.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 176.64, 137.87, 132.66, 128.85, 128.57, 128.00, 126.20, 30.85, 19.86, 18.65, 12.48.

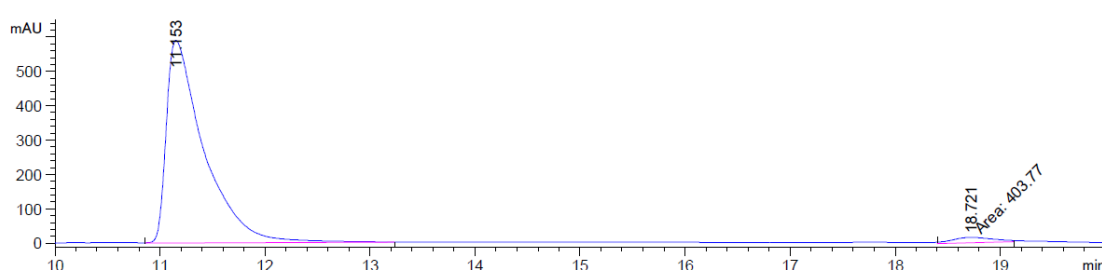
HRMS (ESI-TOF) *m/z* Calcd for C₁₃H₁₃O₂⁻ [M-H]⁻ 201.0916, found 201.0915.

The absolute stereochemistry was assigned based on comparing the specific rotation of **3m**

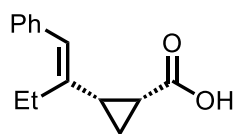
with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.187	BB	0.2919	3079.80225	164.88766	51.0895
2	18.633	BB	0.4418	2948.44263	97.77589	48.9105



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.153	BB	0.3567	1.45963e4	589.75201	97.3082
2	18.721	MM	0.4417	403.76981	15.23415	2.6918



(1R,2S)-2-((E)-1-phenylbut-1-en-2-yl)cyclopropanecarboxylic acid (**8c**)

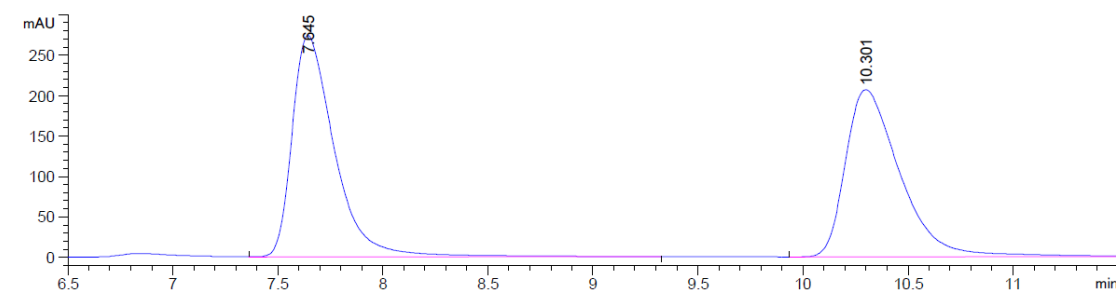
Substrate **6c** was vinyolated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (63% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 7.637 min (major) and 10.295 min (minor), 94:6 er);

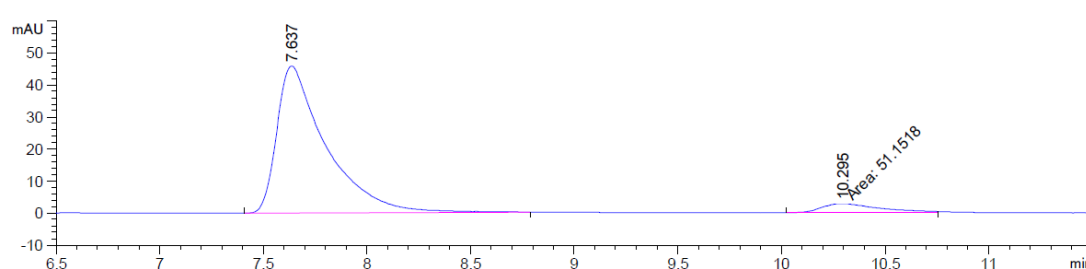
¹H NMR (600 MHz, CDCl₃) δ 7.30 – 7.26 (m, 2H), 7.20 – 7.17 (m, 1H), 7.17 – 7.14 (m, 2H), 6.39 (s, 1H), 2.32 (dq, *J* = 15.0, 7.6 Hz, 1H), 2.21 – 2.15 (m, 2H), 1.97 (ddd, *J* = 9.3, 7.7, 5.4 Hz, 1H), 1.57 (dt, *J* = 7.8, 5.2 Hz, 1H), 1.26 – 1.21 (m, 1H), 1.08 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) δ 177.08, 137.91, 137.59, 128.67, 128.13, 128.02, 126.21, 28.23, 25.18, 20.59, 12.79, 11.73.

HRMS (ESI-TOF) m/z Calcd for $C_{14}H_{15}O_2^-$ [M-H]⁻ 215.1072, found 215.1070.

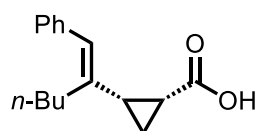
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.645	VB	0.2135	3834.92285	273.60776	50.5871
2	10.301	BB	0.2754	3745.91309	206.68524	49.4129



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.637	BB	0.2301	756.30072	45.92109	93.6650
2	10.295	MM	0.3111	51.15176	2.74055	6.3350



(1R,2S)-2-((E)-1-phenylhex-1-en-2-yl)cyclopropanecarboxylic acid (**8d**)

Substrate **6d** was vinyolated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (61% yield).

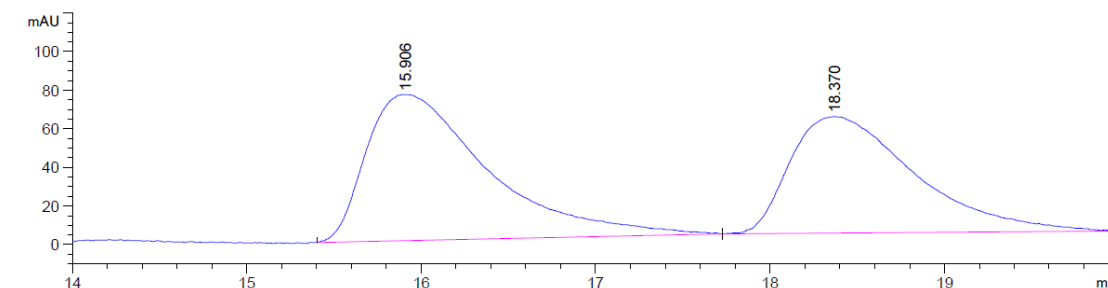
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OD-3 column, 5% *i*PrOH / CO₂, flow rate 1 mL/min, retention time 15.860 min (minor) and 18.022 min (major), 92:8 er);

¹H NMR (600 MHz, CDCl₃) δ 7.29 – 7.26 (m, 2H), 7.21 – 7.12 (m, 3H), 6.41 (s, 1H), 2.32 (ddd, *J* = 13.9, 10.2, 6.4 Hz, 1H), 2.20 – 2.09 (m, 2H), 1.96 (ddd, *J* = 9.3, 7.7, 5.4 Hz, 1H), 1.60 – 1.54 (m, 1H), 1.51 – 1.47 (m, 1H), 1.33 – 1.20 (m, 4H), 0.85 (t, *J* = 7.4 Hz, 3H); ¹³C

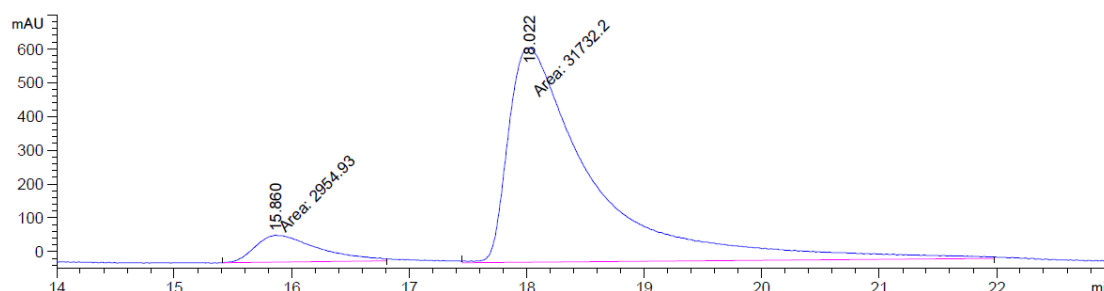
NMR (150 MHz, CDCl₃) δ 176.78, 137.97, 136.60, 128.69, 128.56, 128.00, 126.17, 31.83, 30.28, 28.58, 22.78, 20.49, 13.88, 12.05.

HRMS (ESI-TOF) m/z Calcd for C₁₆H₁₉O₂⁻ [M-H]⁻ 243.1385, found 243.1384.

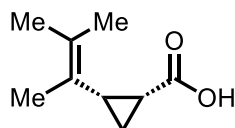
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.875	VV R	0.5591	3928.37988	82.86909	53.1674
2	18.365	MM	0.8272	3460.32300	69.71623	46.8326



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.860	MM	0.6183	2954.92944	79.65041	8.5188
2	18.022	MM	0.8315	3.17322e4	636.05505	91.4812



(1R,2S)-2-(3-methylbut-2-en-2-yl)cyclopropanecarboxylic acid (8e)

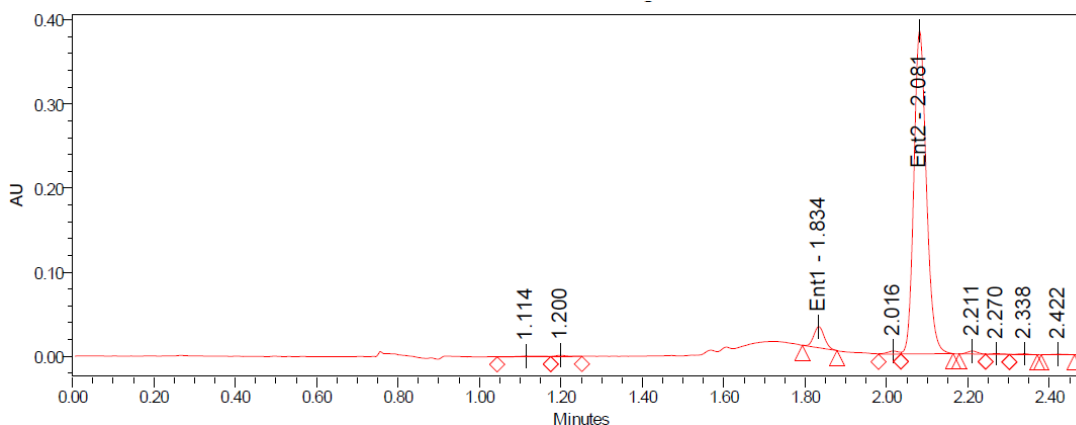
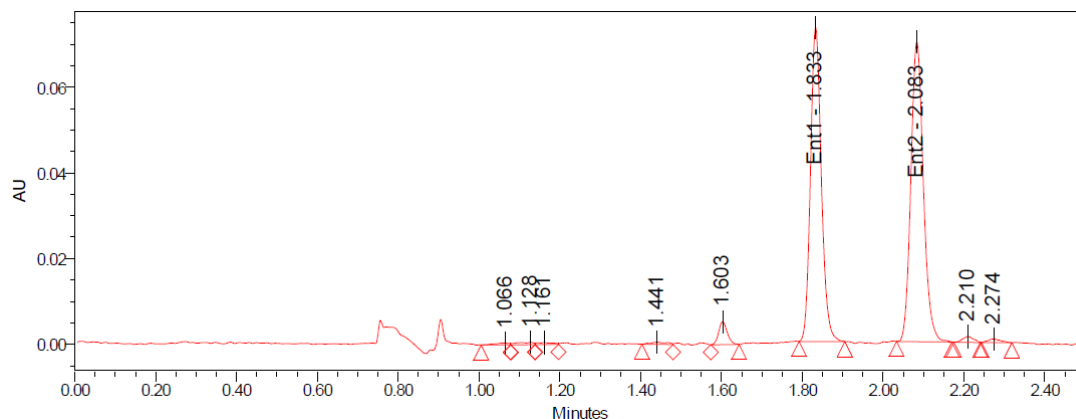
Substrate **6e** was vinyated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (50% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® IG-3 column, 5% MeOH/CO₂, flow rate 4 mL/min, retention time 1.834 min (minor) and 2.081 min (major), 95:5 er);

¹H NMR (600 MHz, CDCl₃) δ 2.05 (t, J = 8.9 Hz, 1H), 1.92 (ddd, J = 8.9, 7.8, 5.3 Hz, 1H),

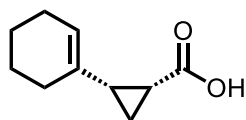
1.74 (s, 3H), 1.66 (s, 3H), 1.64 (s, 3H), 1.36 (dt, $J = 8.0, 5.1$ Hz, 1H), 1.22 – 1.18 (m, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 177.63, 130.57, 122.31, 27.08, 20.92, 20.86, 20.78, 18.25, 14.44. HRMS (ESI-TOF) m/z Calcd for $\text{C}_9\text{H}_{15}\text{O}_2^+$ $[\text{M}+\text{H}]^+$ 155.1072, found 155.1065.

The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Area Summarized by Name

	SampleName	ent1	ent2	ee	Ent1	Ent2
1	HL-5e-rac	47.61	52.39	-4.78	136687	150408
2	HL-5e-chiral	5.22	94.78	-89.55	45185	819942



(1R,2S)-2-(cyclohex-1-en-1-yl)cyclopropanecarboxylic acid (**8f**)

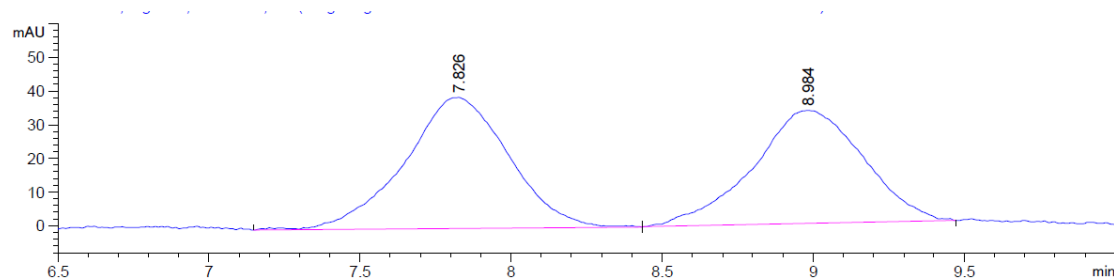
Substrate **6f** was vinylated following the general vinylation procedure (eluent: hexane/ethyl acetate = 5/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (47% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® AS-3 column, 10% *i*PrOH / CO_2 , flow rate 1 mL/min, retention time 7.816 min (major) and 9.113 min (minor), 98:2 er);

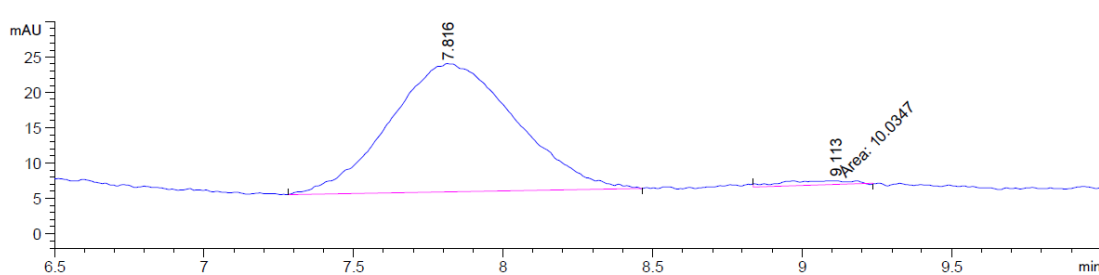
^1H NMR (600 MHz, CDCl_3) δ 5.60 (dq, $J = 4.1, 1.9$ Hz, 1H), 2.07 – 1.96 (m, 3H), 1.91 (t, $J = 8.5$ Hz, 1H), 1.82 (ddd, $J = 9.2, 7.6, 5.5$ Hz, 2H), 1.64 – 1.49 (m, 4H), 1.40 (dt, $J = 7.7, 5.2$ Hz, 1H), 1.10 (td, $J = 8.0, 4.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 176.01, 131.41, 125.03, 28.74, 28.25, 24.74, 22.43, 21.94, 18.80, 10.91.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{10}\text{H}_{15}\text{O}_2^+$ $[\text{M}+\text{H}]^+$ 167.1072, found 167.1065.

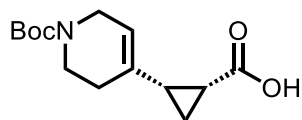
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.826	VV R	0.3030	930.88171	38.93185	52.1549
2	8.984	BV R	0.3058	853.95709	33.68350	47.8451



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.816	BV R	0.3457	532.12018	18.17036	98.1491
2	9.113	MM	0.3155	10.03466	5.30117e-1	1.8509



(1R,2S)-2-(1-(tert-butoxycarbonyl)-1,2,3,6-tetrahydropyridin-4-yl)cyclopropanecarboxylic acid (8g)

Substrate **6g** was vinyllated following the general vinyllation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (51% yield).

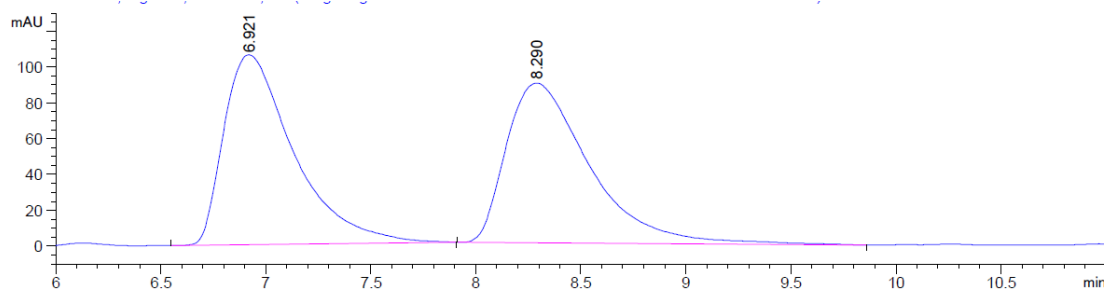
The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK[®] IC-3 column, 10% $^i\text{PrOH}$ / CO_2 , flow rate 2 mL/min, retention time 6.855 min (major) and

8.383 min (minor), 97:3 er);

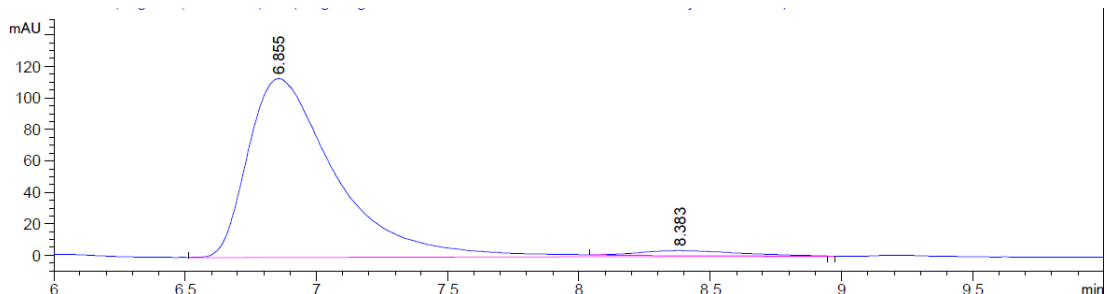
^1H NMR (600 MHz, CDCl_3) δ 5.53 (s, 1H), 3.99 – 3.81 (m, 2H), 3.55 (dt, $J = 12.8, 5.4$ Hz, 1H), 3.31 (s, 1H), 2.21 – 2.13 (m, 1H), 1.96 (q, $J = 8.3$ Hz, 2H), 1.86 (ddd, $J = 9.1, 7.7, 5.5$ Hz, 1H), 1.46 (s, 9H), 1.41 (dt, $J = 7.9, 5.2$ Hz, 1H), 1.16 (td, $J = 8.1, 4.9$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 176.84, 155.02, 131.07, 122.19, 79.55, 31.91, 29.68, 28.46, 27.63, 22.68, 14.11, 11.61.

HRMS (ESI-TOF) m/z Calcd for $\text{C}_{14}\text{H}_{20}\text{NO}_4^-$ [M-H] $^-$ 266.1392, found 266.1392.

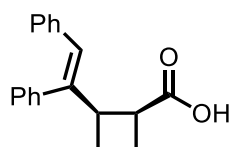
The absolute stereochemistry was assigned based on comparing the specific rotation of **3m** with literature.



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.921	BB	0.3517	2451.06152	106.02688	49.9608
2	8.290	BB	0.4135	2454.91040	89.08801	50.0392



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.855	BV R	0.3346	2656.87109	113.72182	96.8827
2	8.383	VB E	0.3269	85.48772	3.20762	3.1173



(1S,2R)-2-((Z)-1,2-diphenylvinyl)cyclobutanecarboxylic acid (8h)

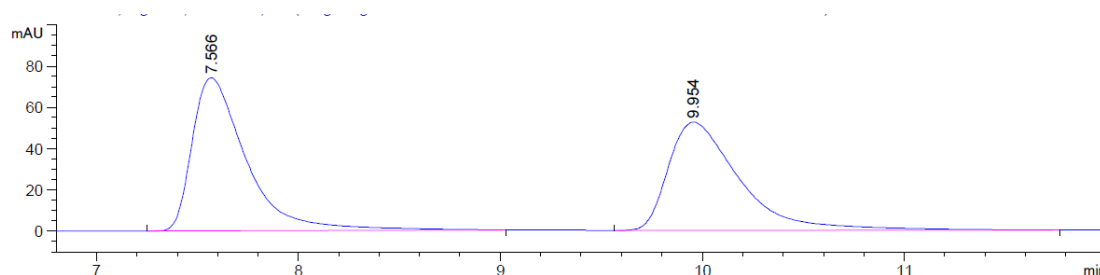
Substrate **6h** was vinylation following the general vinylation procedure (eluent: hexane/ethyl acetate = 2/1 with 1% v/v of acetic acid). The product was obtained as a colorless oil (61% yield).

The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OJ-3 column, 5% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 7.478 min (major) and 9.876 min (minor), 88:12 er);

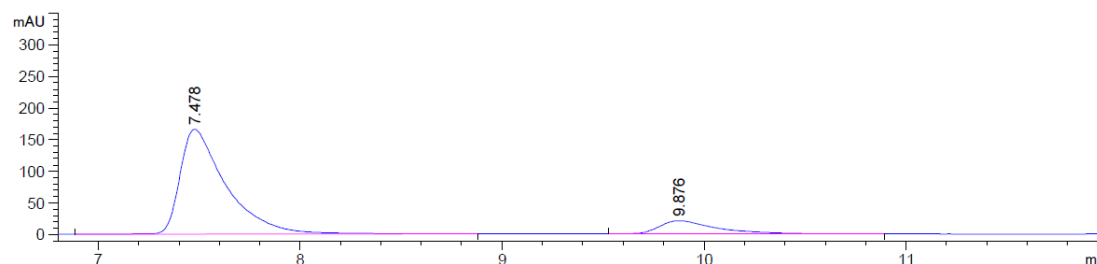
¹H NMR (600 MHz, CDCl₃) δ 7.19 (ddd, *J* = 4.3, 2.3, 1.2 Hz, 3H), 7.14 – 7.09 (m, 2H), 7.05 – 7.03 (m, 3H), 6.95 – 6.89 (m, 2H), 6.47 (s, 1H), 3.81 – 3.74 (m, 1H), 3.28 – 3.20 (m, 1H), 2.78 – 2.70 (m, 1H), 2.14 (dddd, *J* = 9.4, 7.9, 5.9, 3.4 Hz, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 178.51, 141.42, 140.20, 137.07, 129.24, 128.75, 128.36, 127.78, 127.02, 126.91, 126.33, 45.18, 44.40, 24.50, 20.21.

HRMS (ESI-TOF) *m/z* Calcd for C₁₉H₁₇O₂⁻ [M-H]⁻ 277.1229, found 277.1226.

The absolute stereochemistry was assigned based on comparing the specific rotation of **5u** with literature.

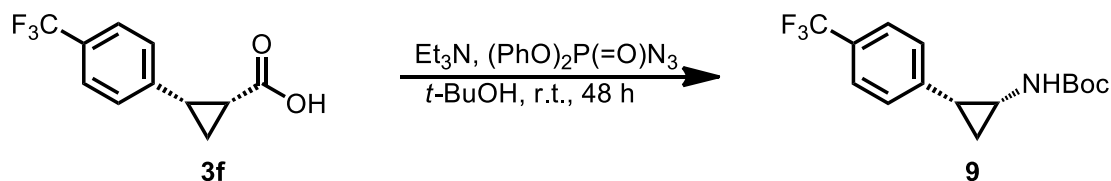


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.566	BB	0.2792	1382.54333	74.26161	51.8243
2	9.954	BB	0.3741	1285.20544	52.41679	48.1757

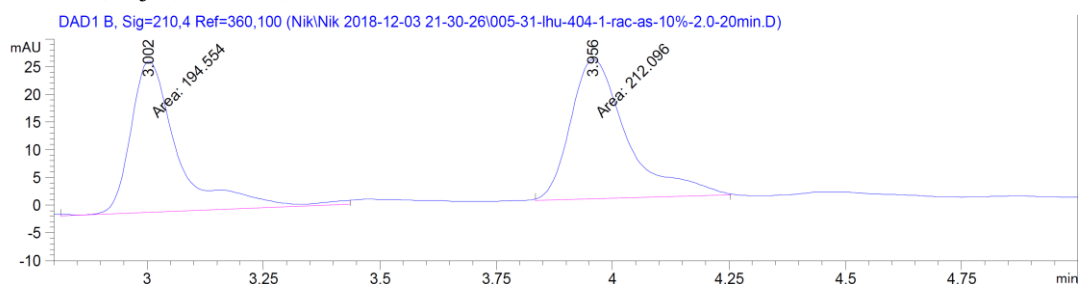


Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.478	BB	0.2308	5079.21924	317.22769	88.1404
2	9.876	MM	0.2967	683.42407	38.38791	11.8596

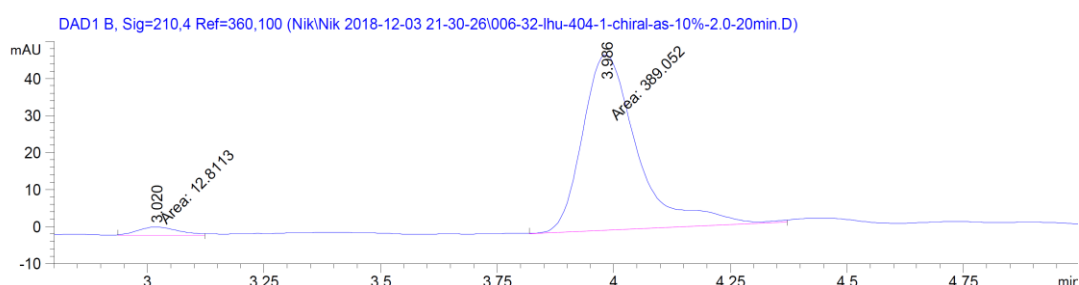
3.8 Synthesis of Cis-Chiral Amine from Carboxylic Acid



To a solution of tert-butyl ((1R,2R)-2-(4-(trifluoromethyl)phenyl)cyclopropyl)carbamate (46 mg, 0.2 mmol, 97:3 er.) and Et₃N (31 uL, 0.22 mmol) in anhydrous *t*-BuOH (1 mL) was added diphenylphosphoryl azide (47 uL, 0.22 mmol) dropwise. The reaction was heated at 80 °C and stirred for 48 h before cooling to r.t. and concentrating in vacuo. Et₂O (20 mL) and water (40 mL) were added. The organic portion was isolated and the aqueous phase was extracted with Et₂O (2 × 30 mL). The organics extracts were combined, washed with sat. aq. NaHCO₃ (20 mL) and brine (20 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography (5 % EtOAc/hexane) to yield the title compound (43.3 mg, 72 %) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.53 (d, *J* = 8.1 Hz, 2H), 7.32 (d, *J* = 7.9 Hz, 2H), 4.35 (s, 1H), 2.97 (s, 1H), 2.28 (d, *J* = 9.0 Hz, 1H), 1.37 (dt, *J* = 8.9, 6.8 Hz, 1H), 1.28 (s, 9H), 1.08 (s, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 156.17, 141.13, 129.01, 128.46 (q, *J* = 36.0 Hz), 124.83 (q, *J* = 4.5 Hz), 124.31 (q, *J* = 270.0 Hz), 79.59, 29.65, 28.07, 22.43, 12.14; ¹⁹F NMR (375 MHz, CDCl₃) δ -62.62. [α]_D²⁰ = 13.2 (c = 1.0, CHCl₃). The enantiomeric purity of the substrate was determined by SFC analysis (CHIRALPAK® OD-3 column, 10% *i*PrOH / CO₂, flow rate 2 mL/min, retention time 3.020 min (minor) and 3.986 min (major), 97:3 er);



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.002	MM	0.1183	194.55423	27.40913	47.8432
2	3.956	MM	0.1399	212.09579	25.26016	52.1568



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	3.020	MM	0.0981	12.81127	2.17736	3.1880
2	3.986	MM	0.1363	389.05157	47.55794	96.8120

4. Reference

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14. Shen, P.-X.; Hu, L.; Shao, Q.; Hong, K.; Yu, J.-Q. *J. Am. Chem. Soc.* **2018**, *140*, 6545.

5. X-Ray Crystallographic Data of Compounds

Figure S1 X-Ray Structure 5p

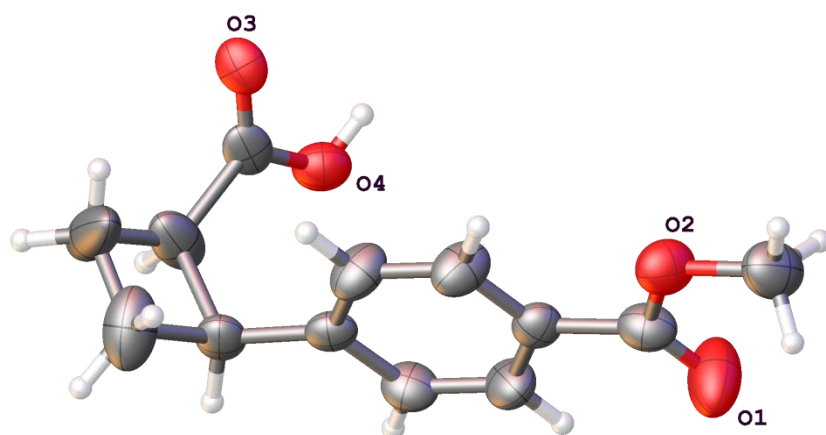


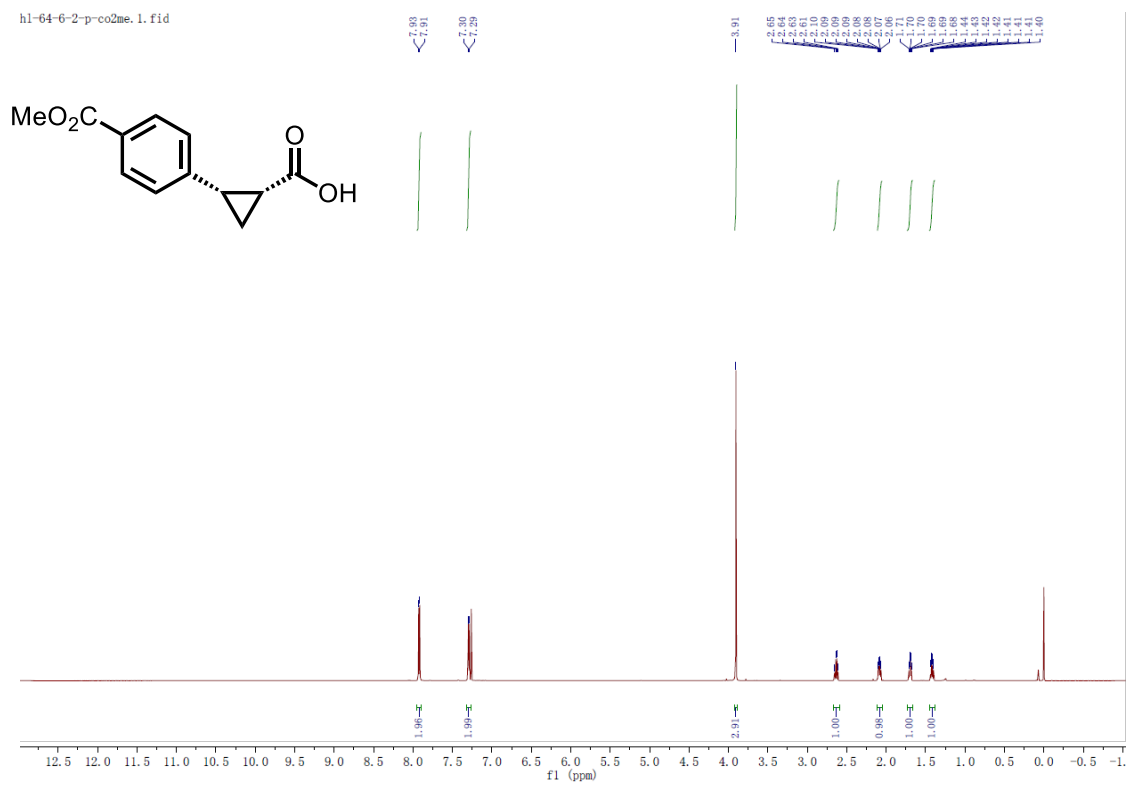
Table 1. Crystal data and structure refinement for 5p

CCDC number	1870087
Empirical formula	C ₁₃ H ₁₄ O ₄
Formula weight	234.24
Temperature	100.0 K
Wavelength	1.54178 Å
Crystal system	Trigonal
Space group	P3 ₁
Unit cell dimensions	a = 15.8015(7) Å = 90°. b = 15.8015(7) Å = 90°. c = 16.3042(10) Å = 120°.
Volume	3525.5(4) Å ³
Z, Z'	12, 4
Density (calculated)	1.324 Mg/m ³
Absorption coefficient	0.815 mm ⁻¹
F(000)	1488
Crystal size	0.20 x 0.17 x 0.11 mm ³
Theta range for data collection	3.229 to 68.408°.
Index ranges	-18<=h<=19, -19<=k<=19, -19<=l<=19
Reflections collected	52265
Independent reflections	8620 [R(int) = 0.0417]
Completeness to theta = 67.679°	100.0 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	0.7531 and 0.6705
Refinement method	Full-matrix least-squares on F ²

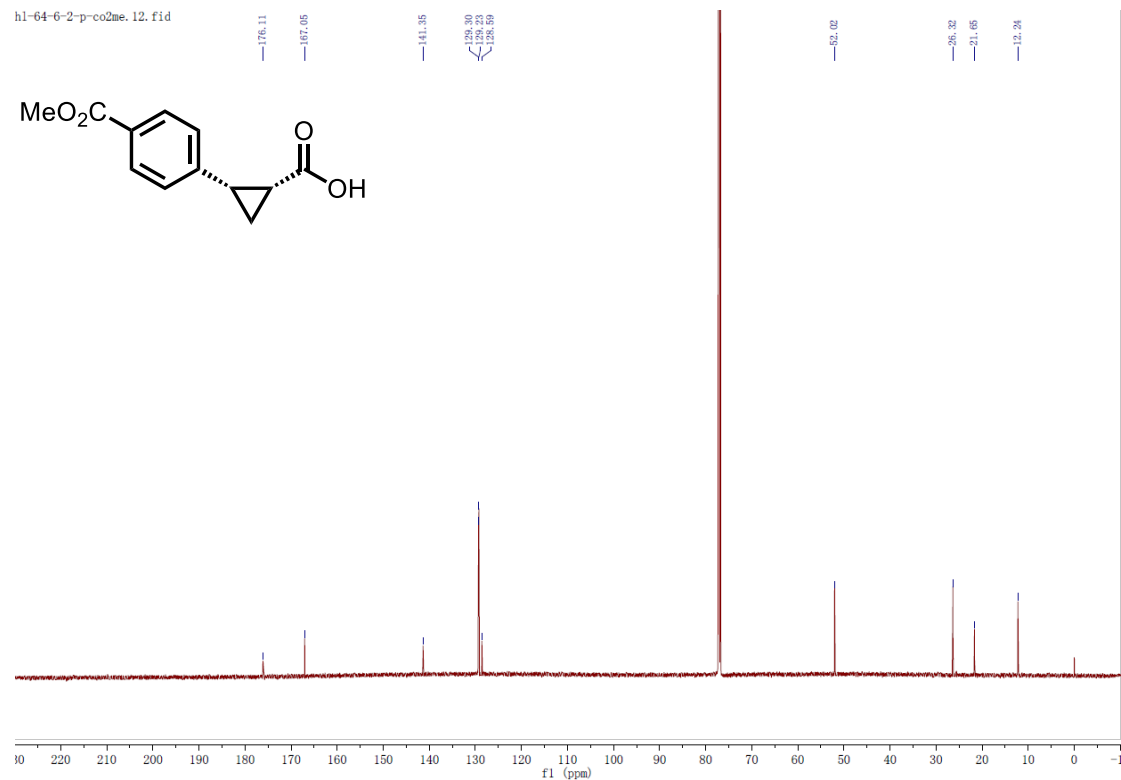
Data / restraints / parameters	8620 / 4 / 634
Goodness-of-fit on F^2	1.039
Final R indices [$I > 2\sigma(I)$]	R1 = 0.0398, wR2 = 0.1082
R indices (all data)	R1 = 0.0413, wR2 = 0.1097
Absolute structure parameter	0.08(5)
Extinction coefficient	n/a
Largest diff. peak and hole	0.328 and -0.171 e. \AA^{-3}

6. NMR Spectra

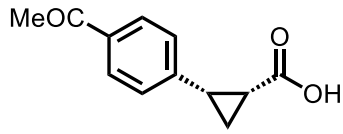
h1-64-6-2-p-co2me.1.fid



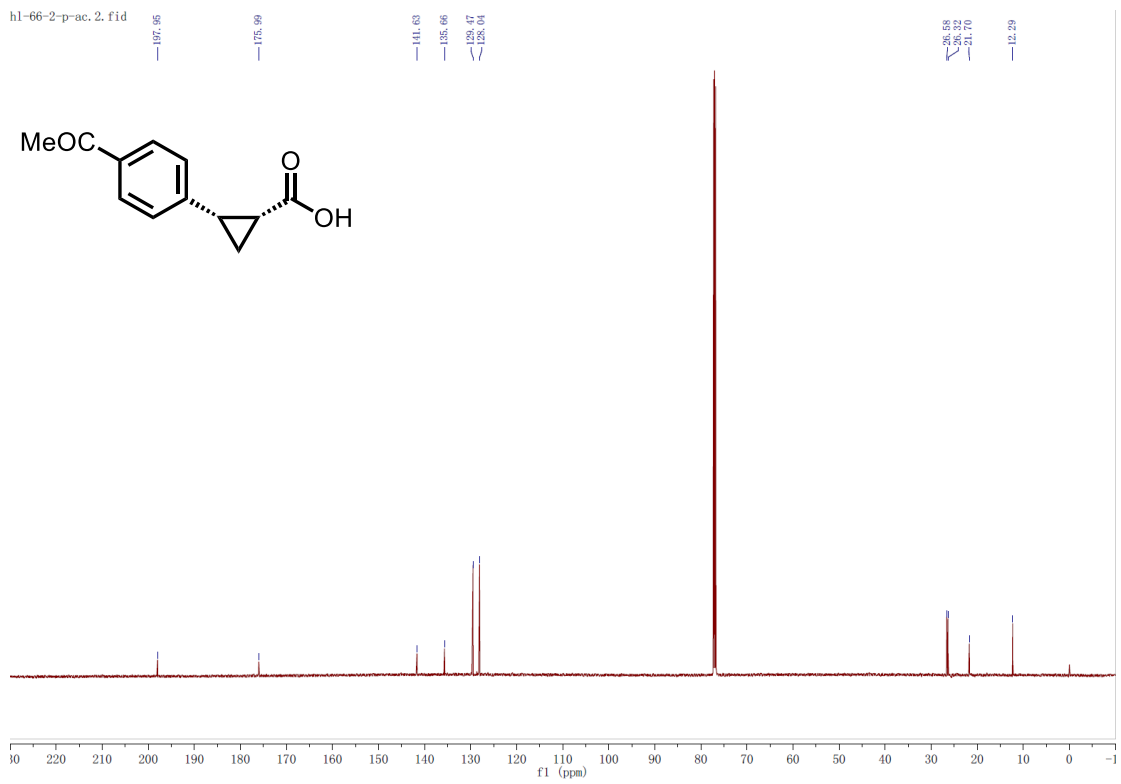
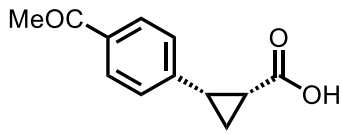
h1-64-6-2-p-co2me.12.fid



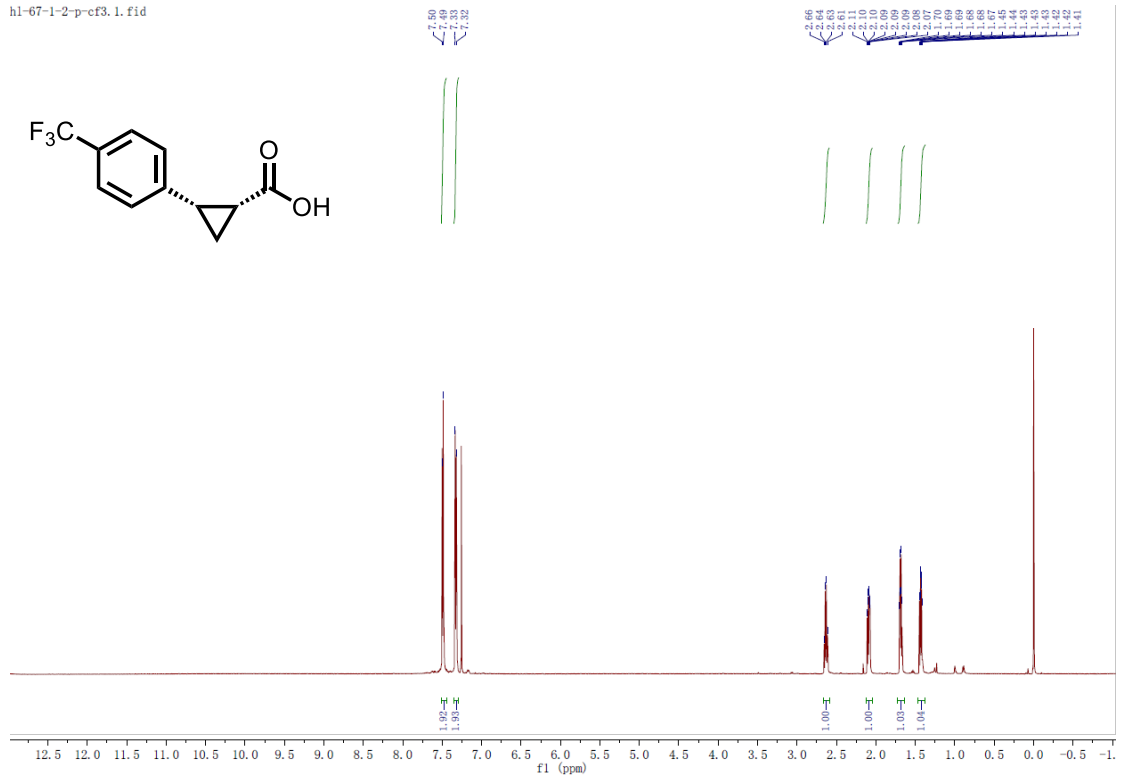
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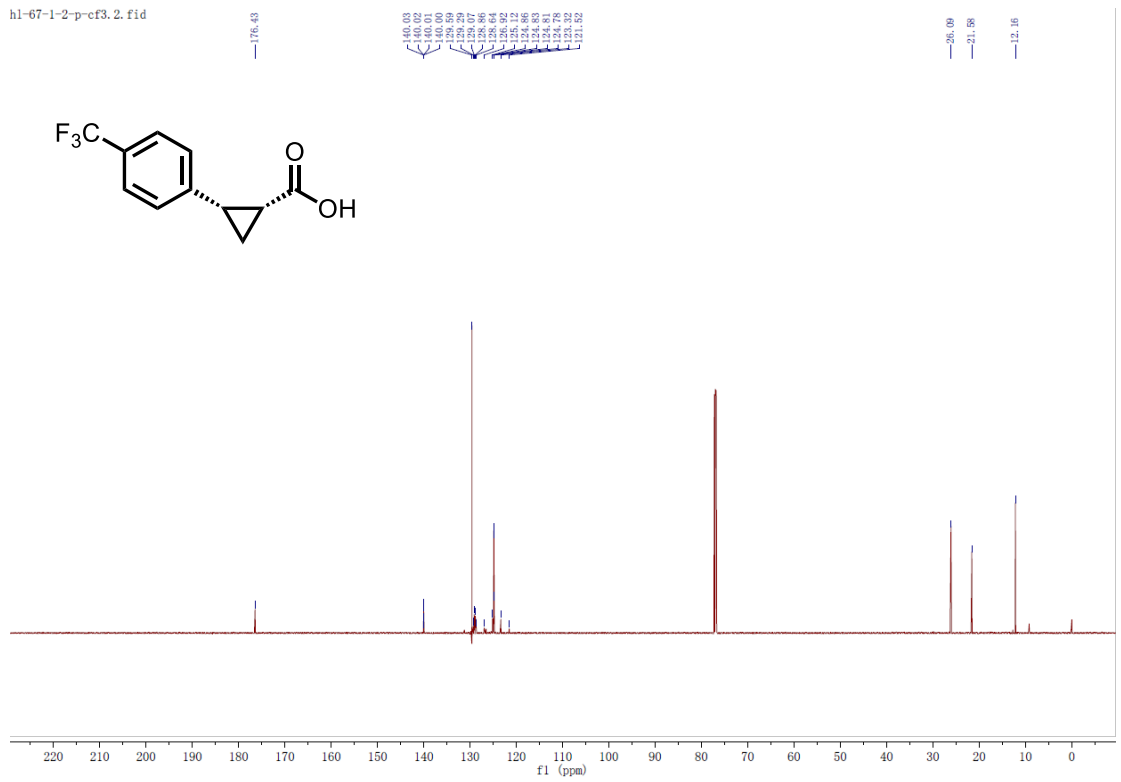
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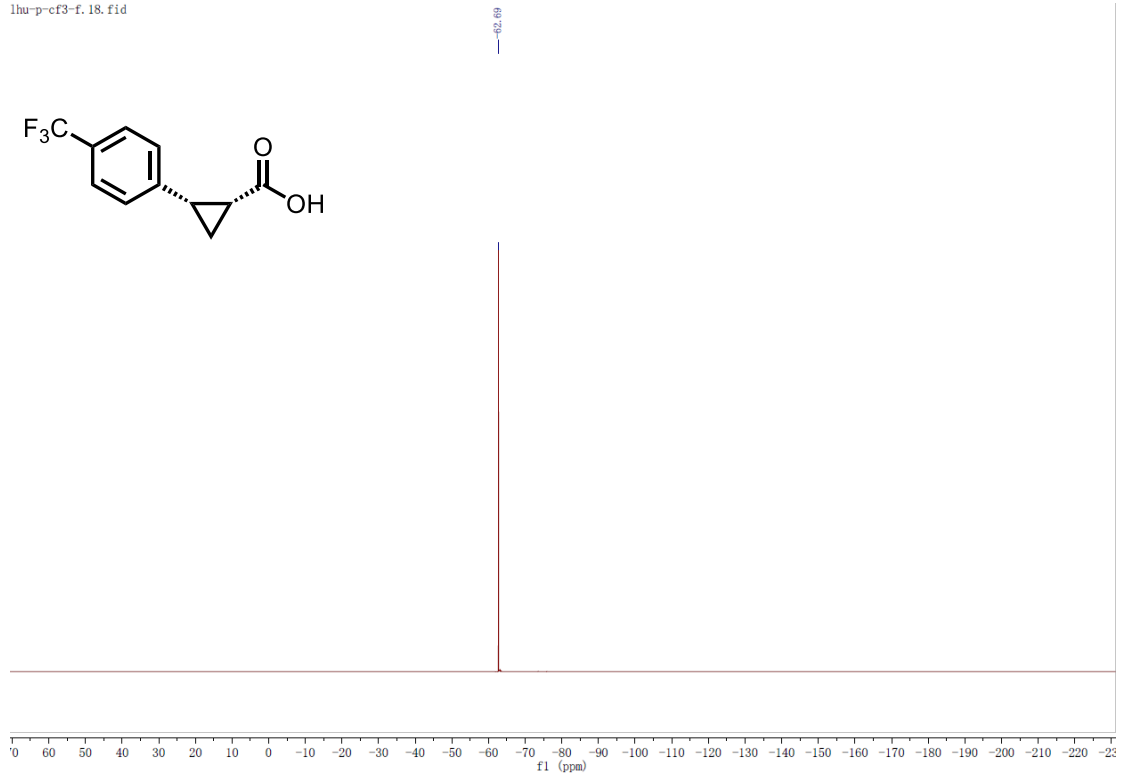
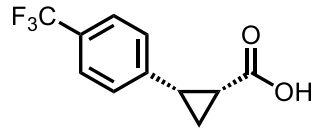
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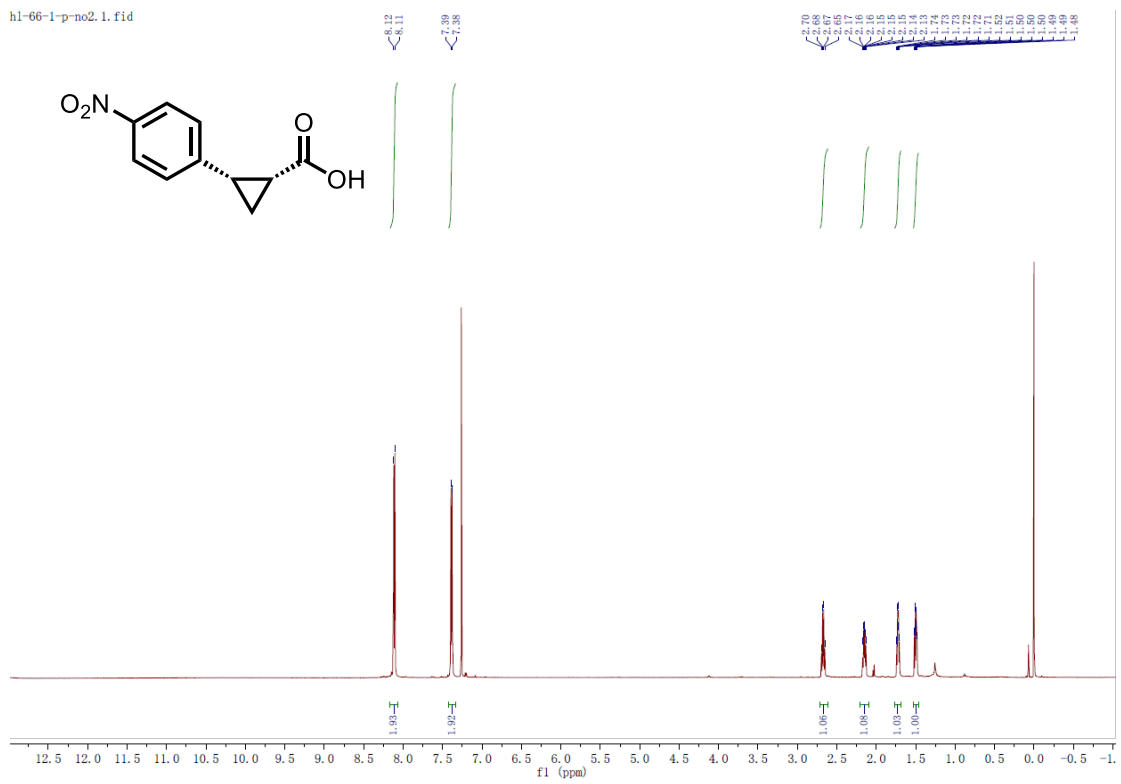
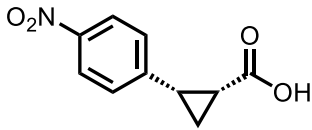
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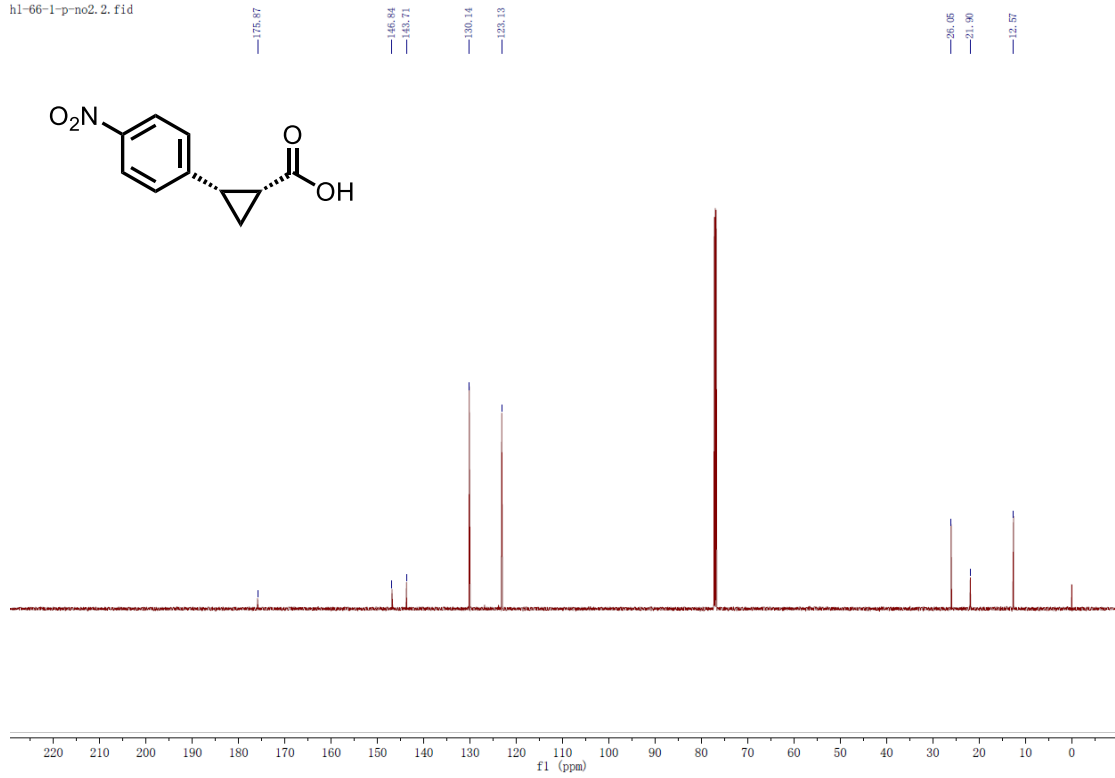
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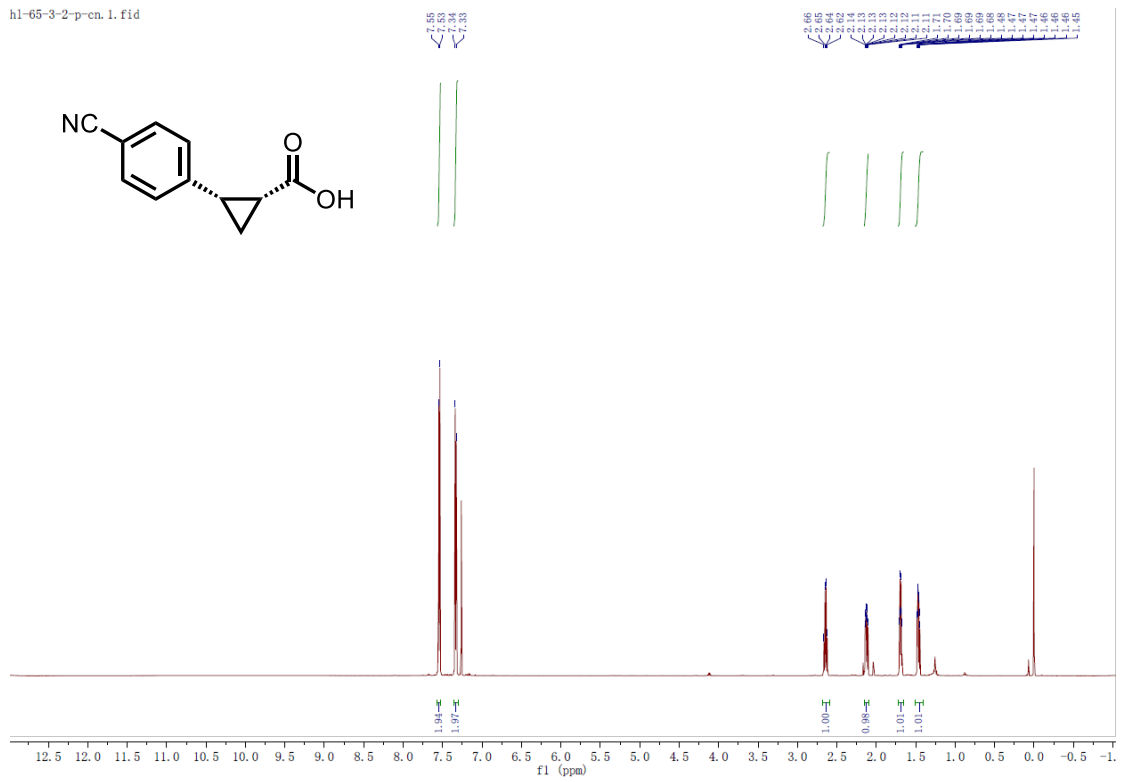
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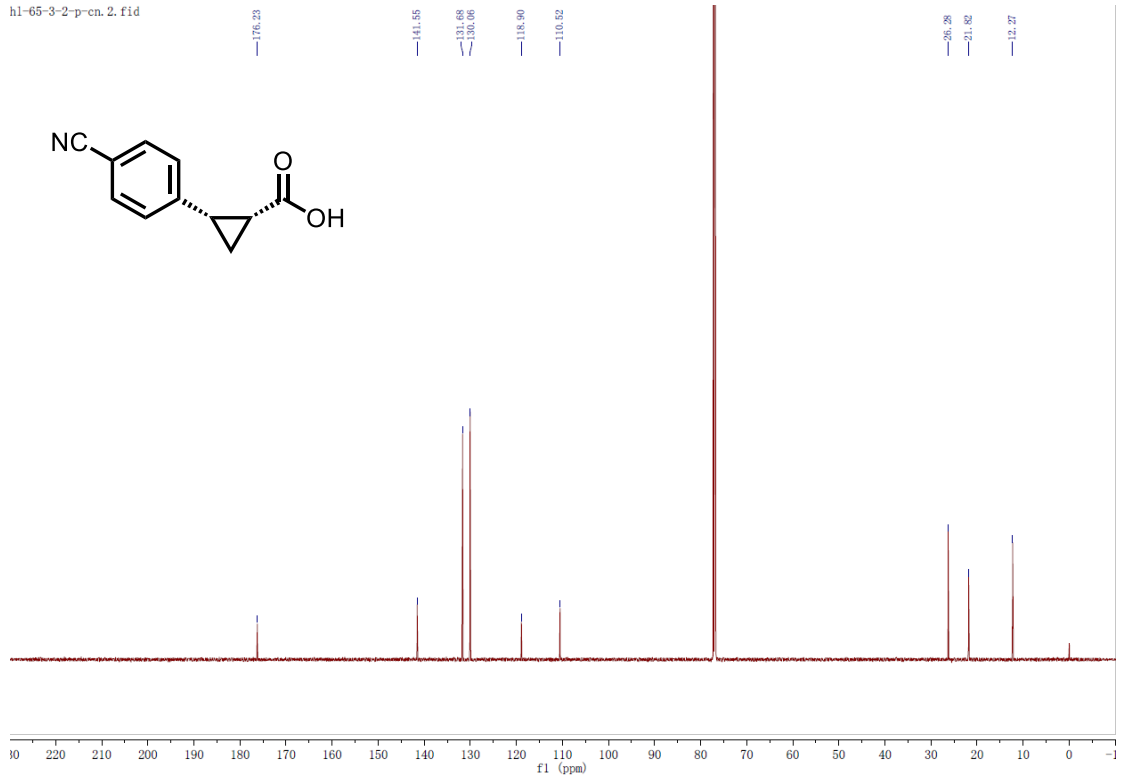
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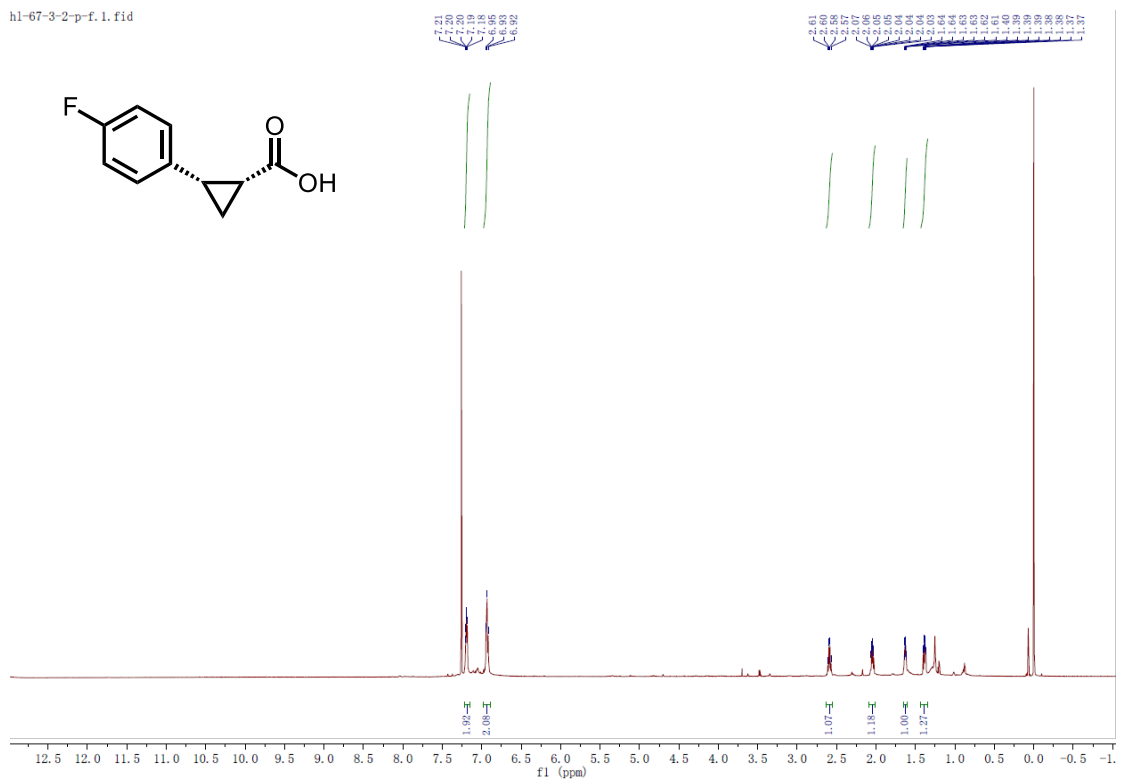
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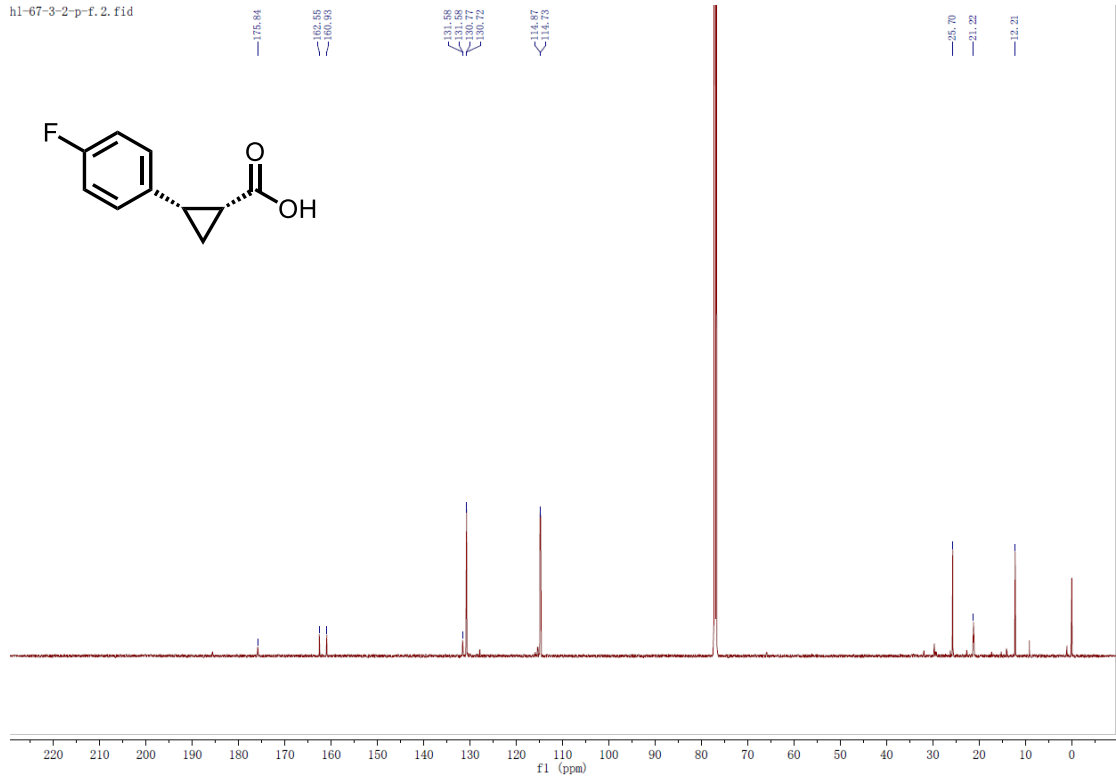
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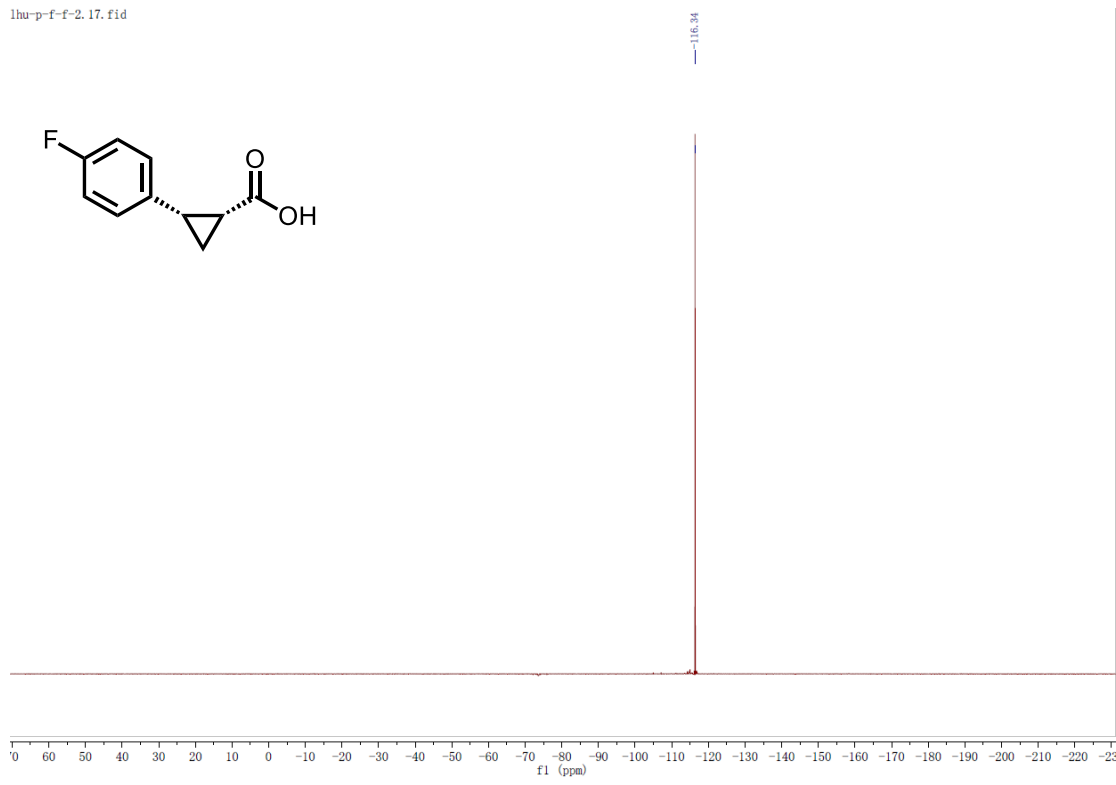
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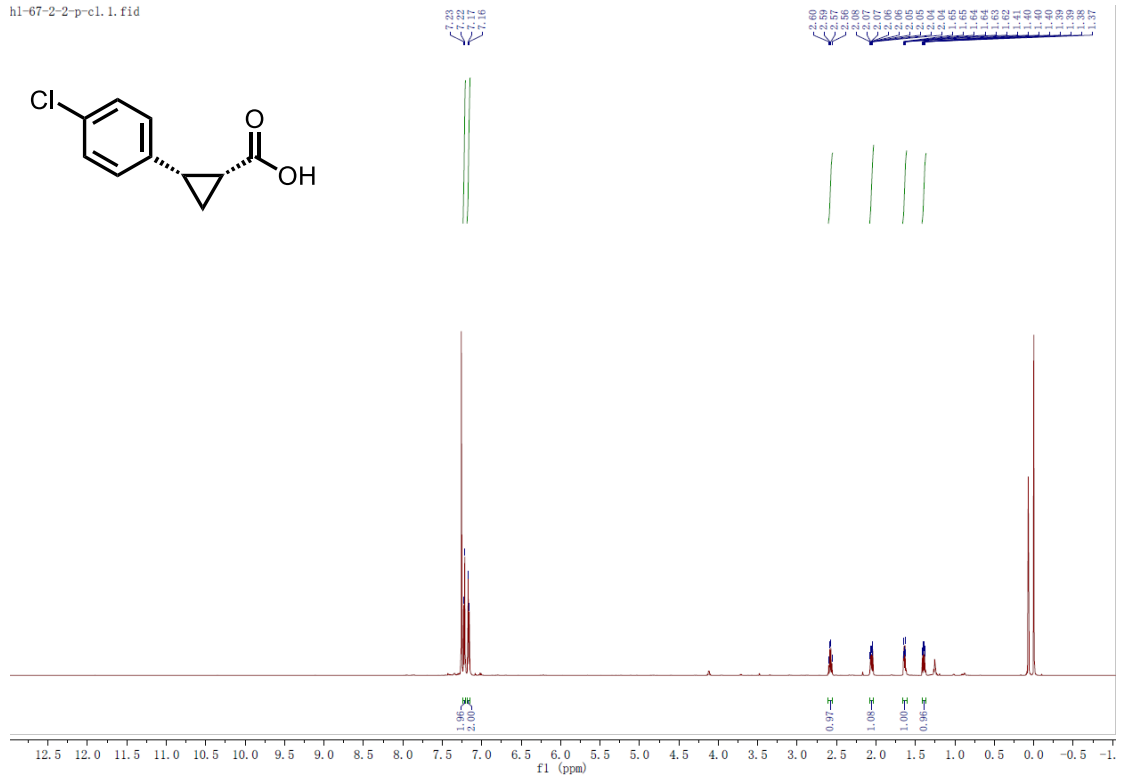
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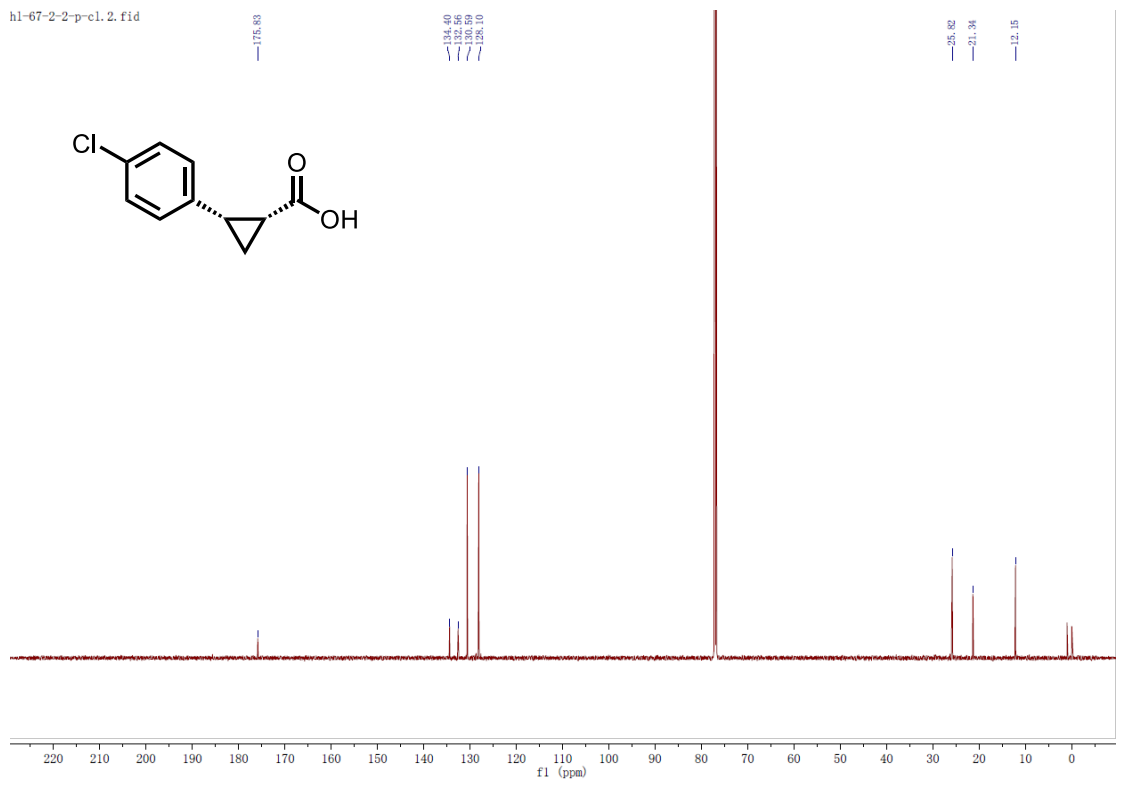
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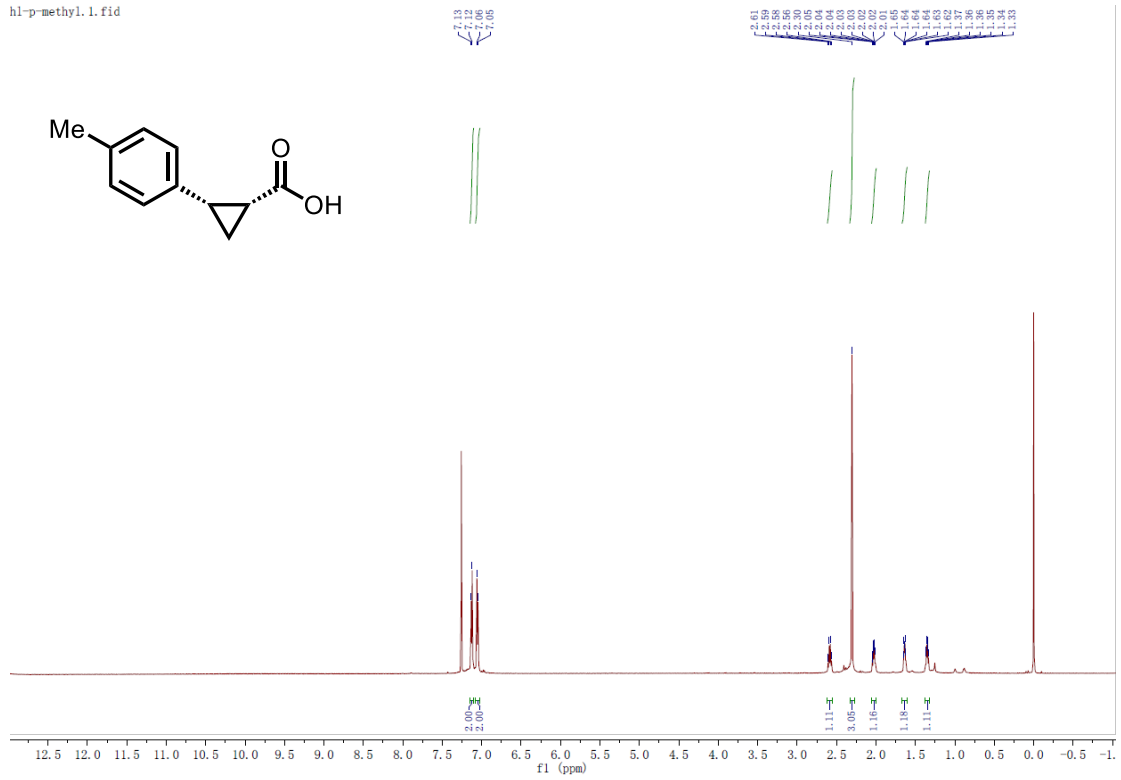
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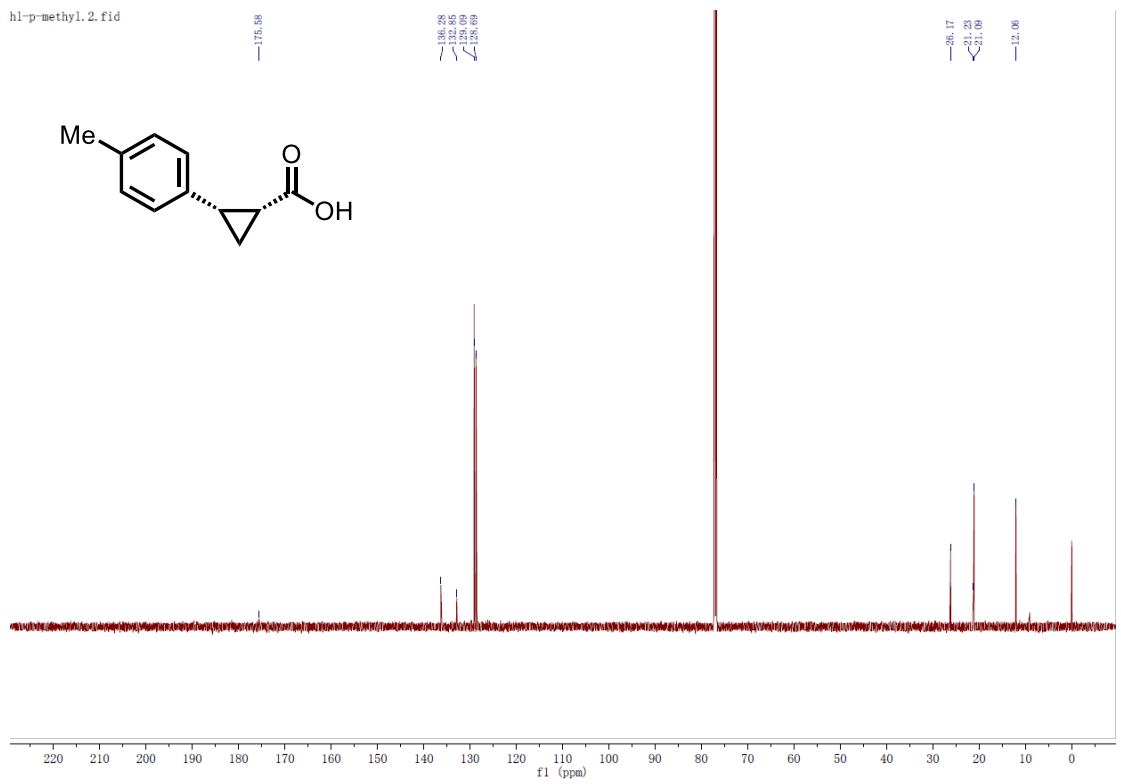
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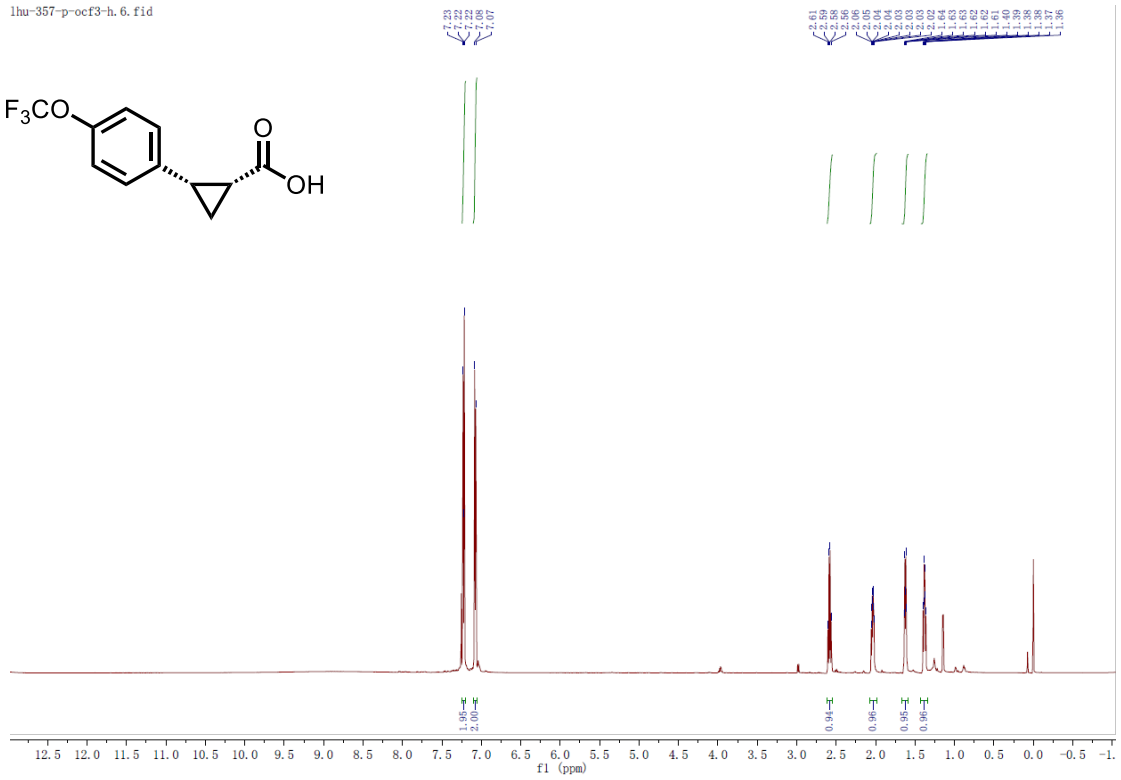
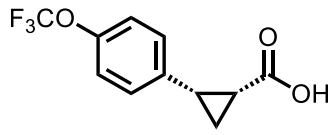
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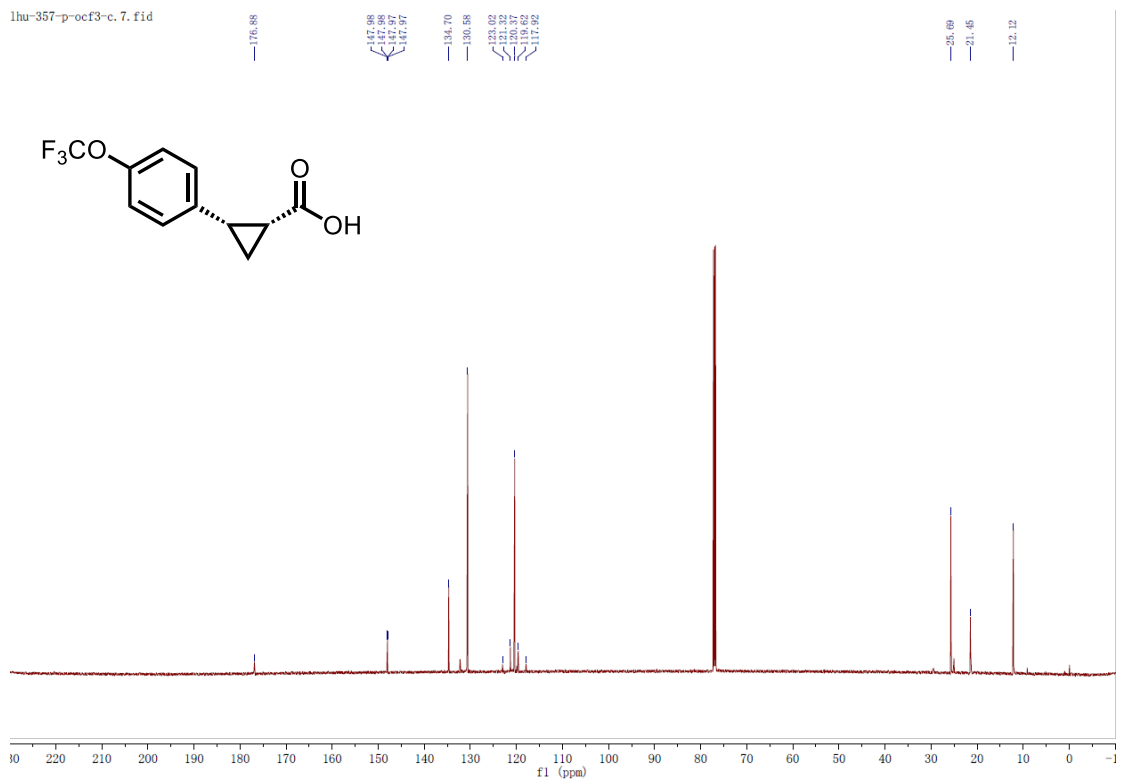
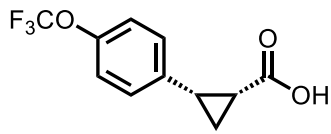
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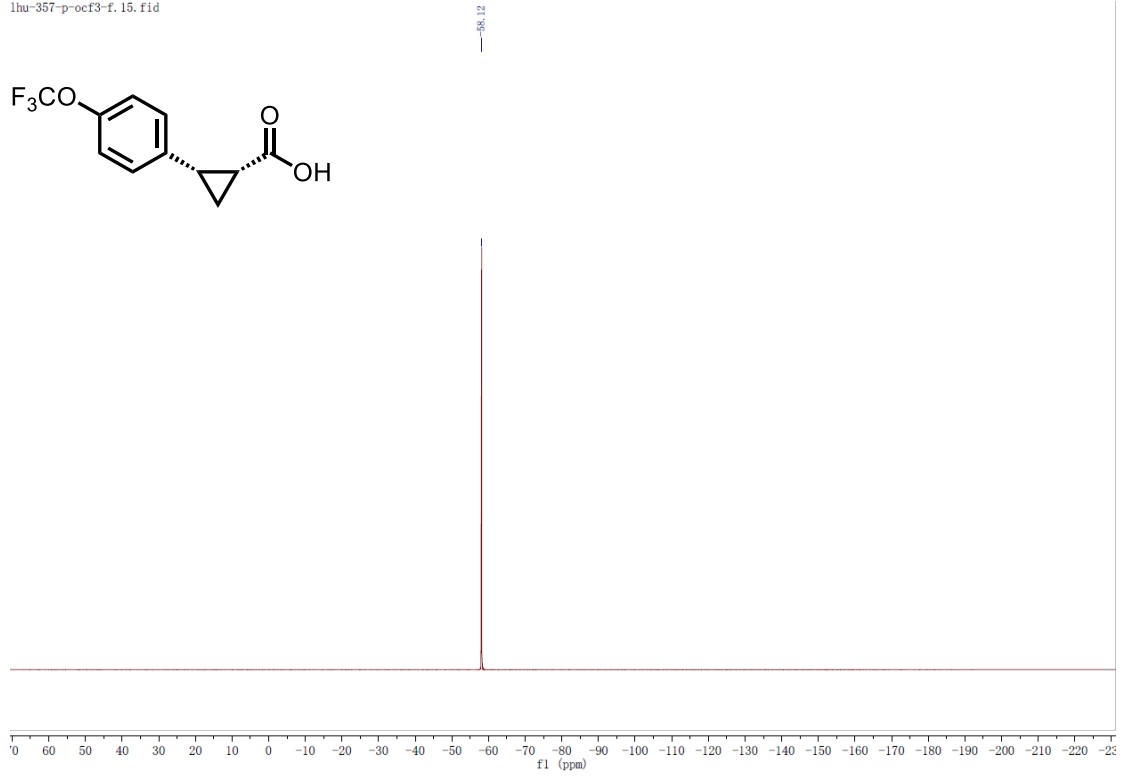
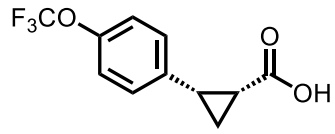
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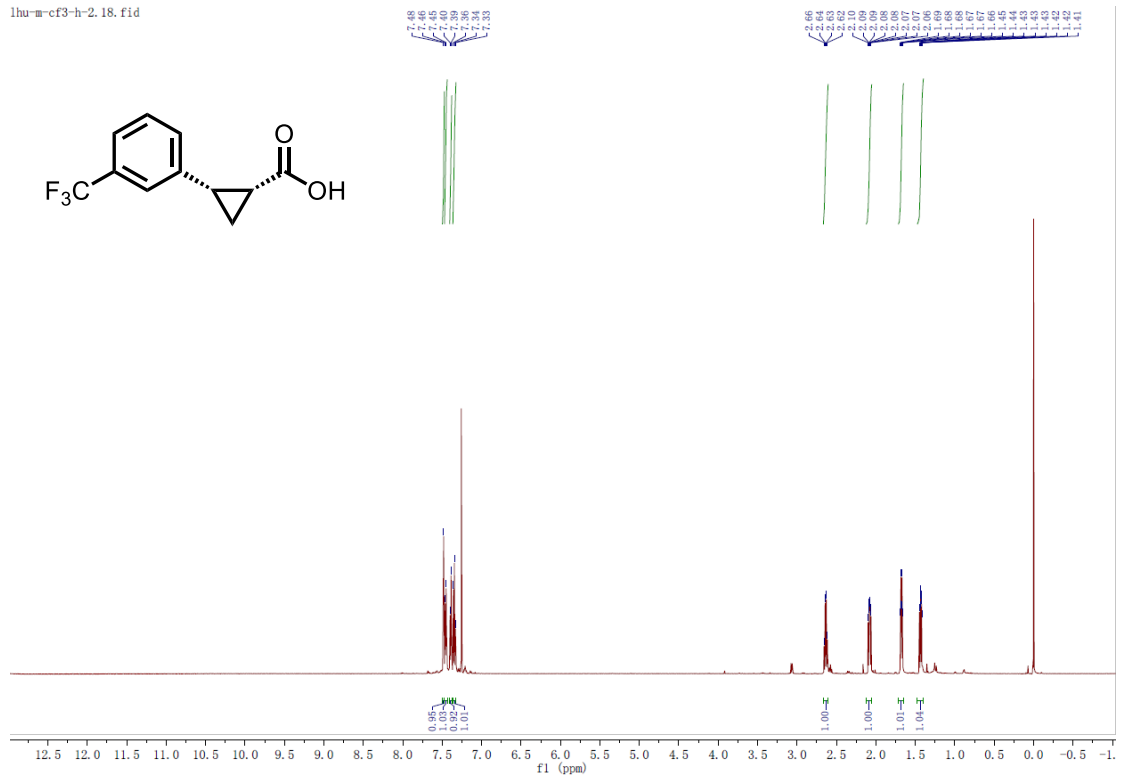
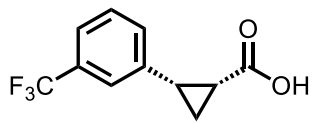
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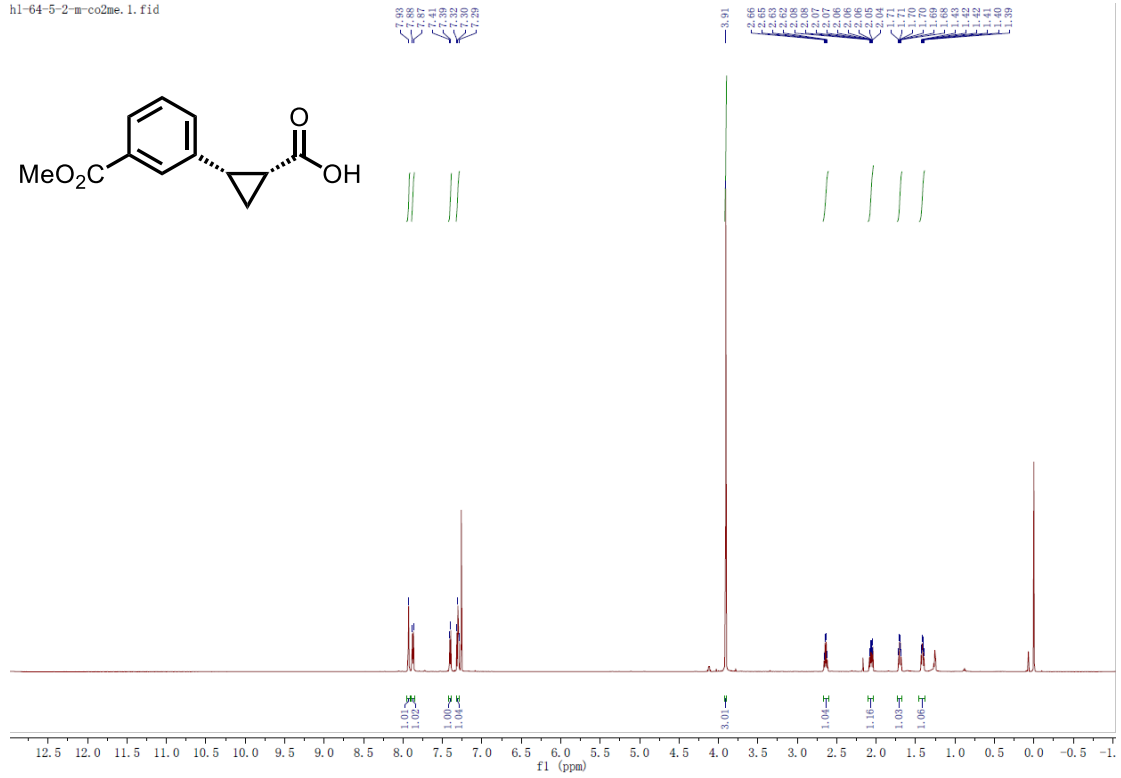
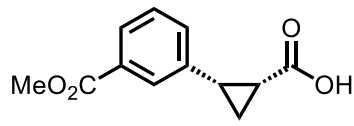
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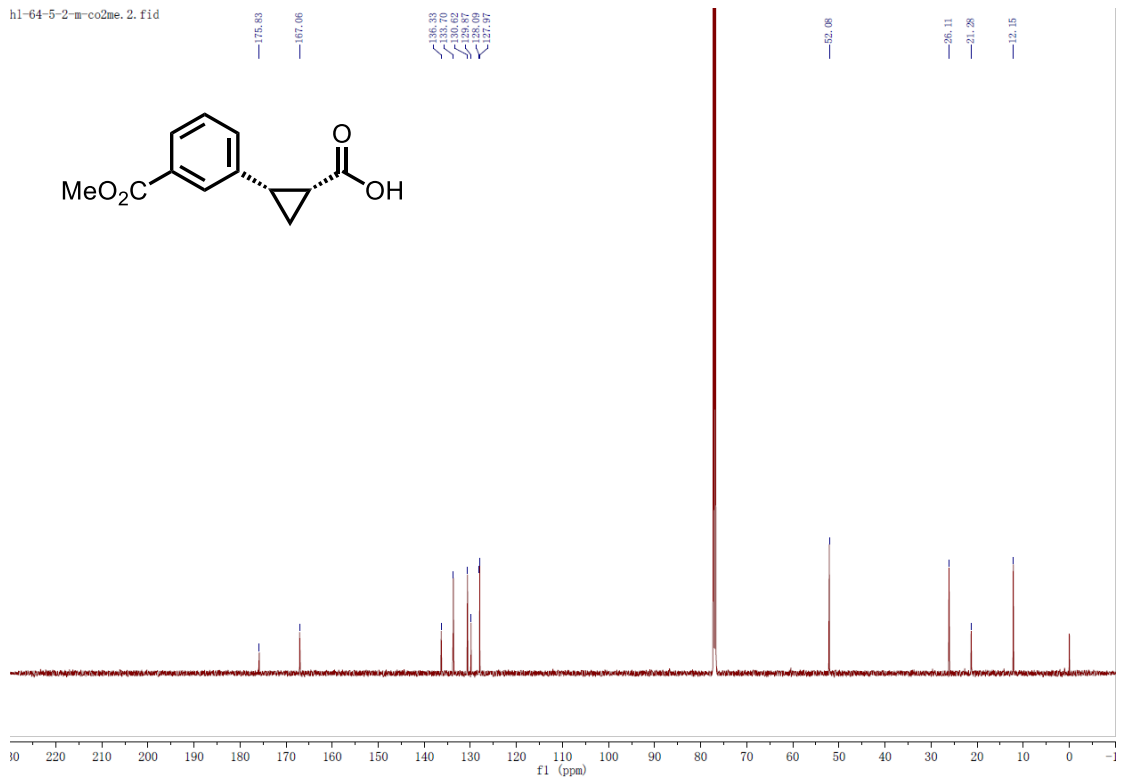
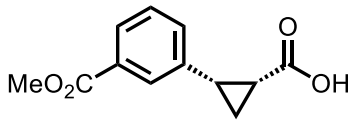
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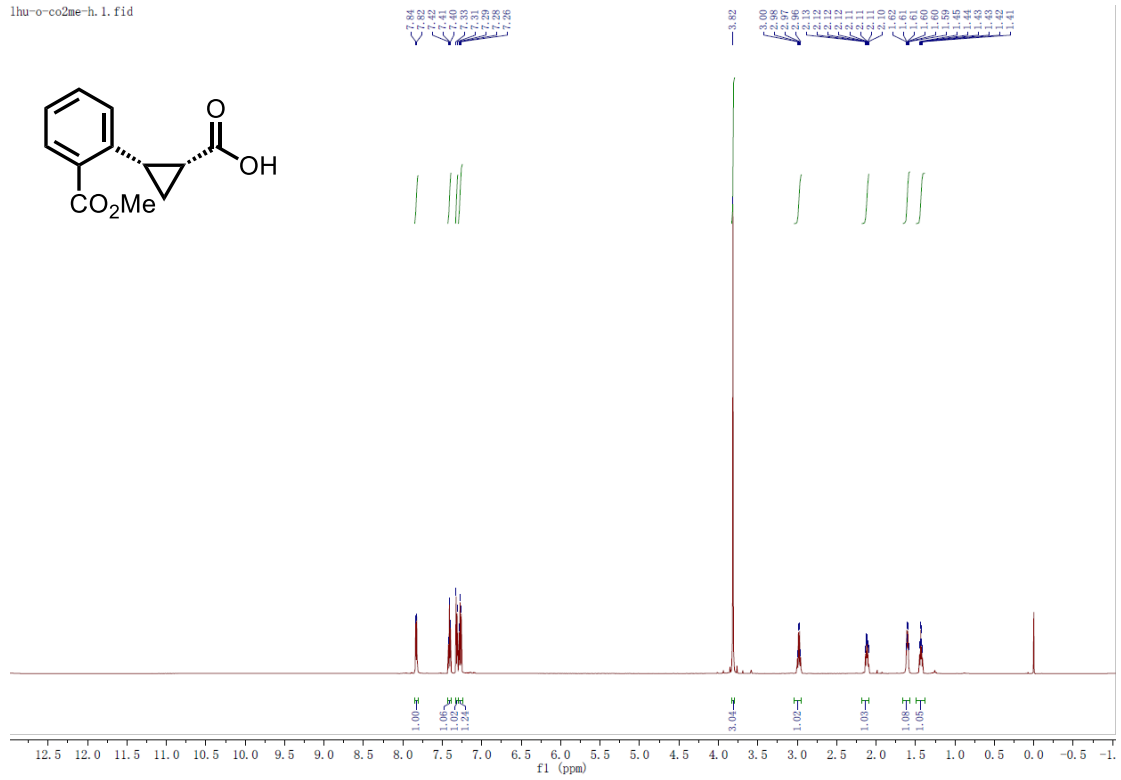
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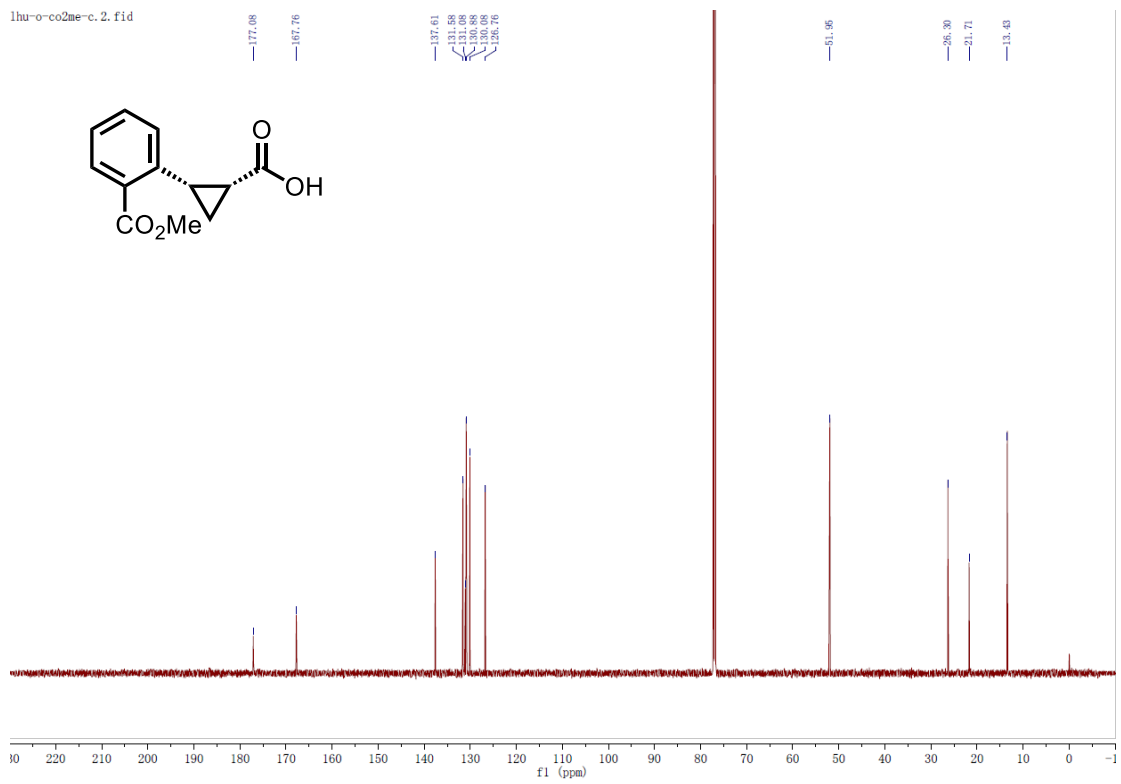
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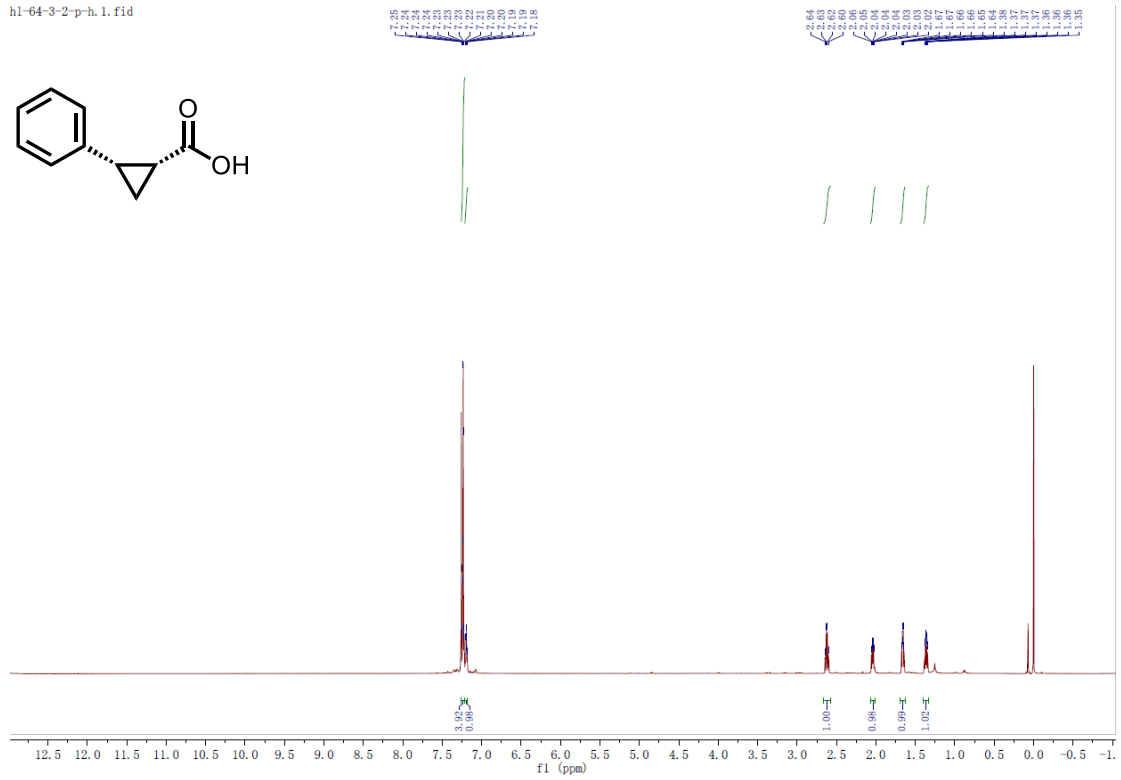
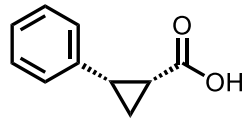
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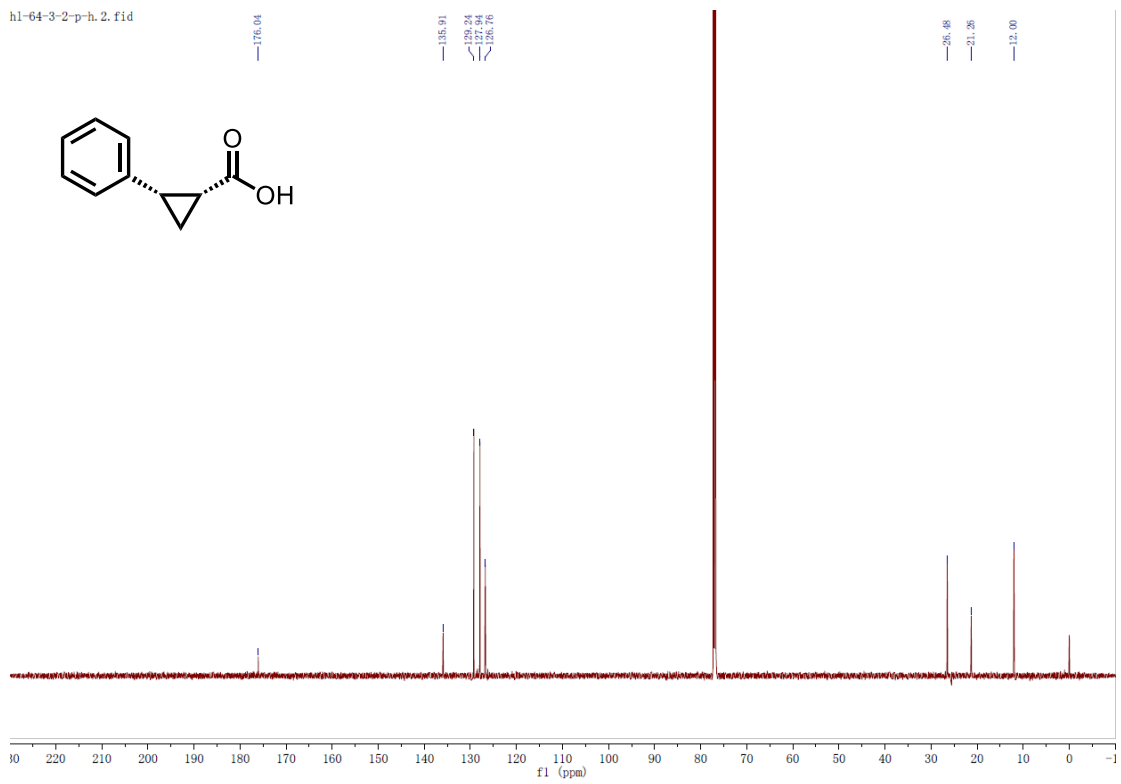
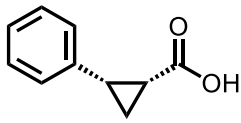
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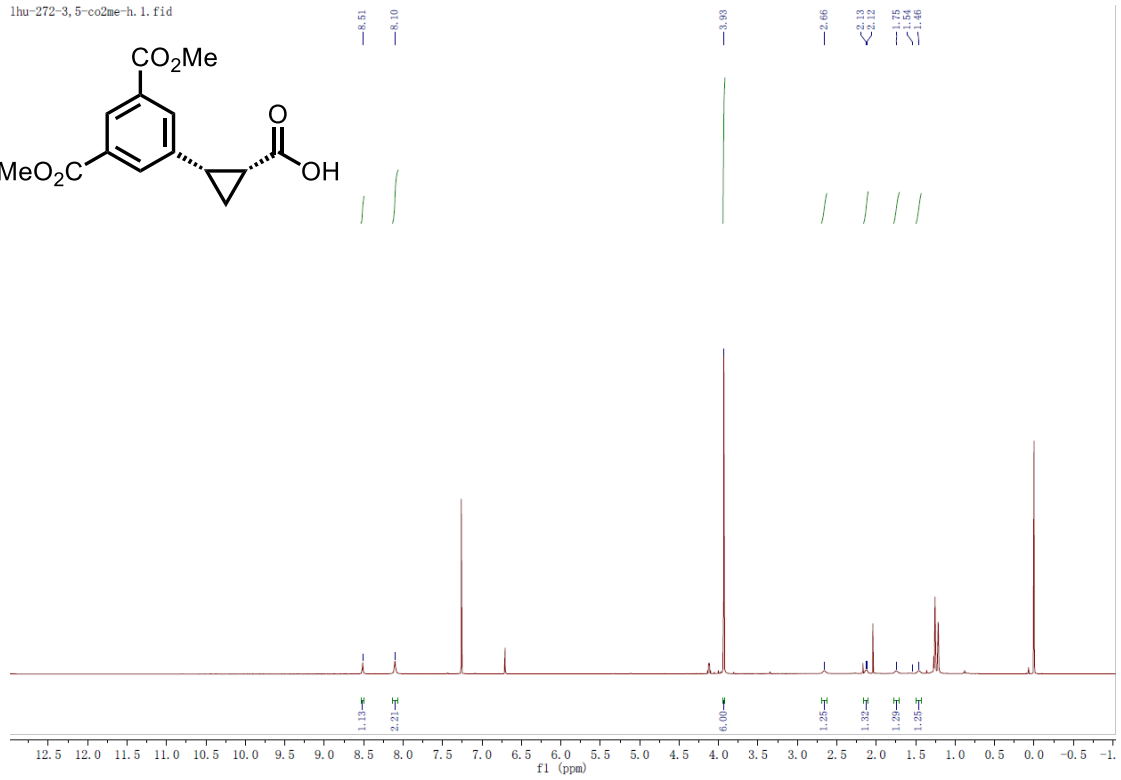
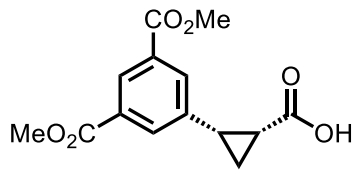
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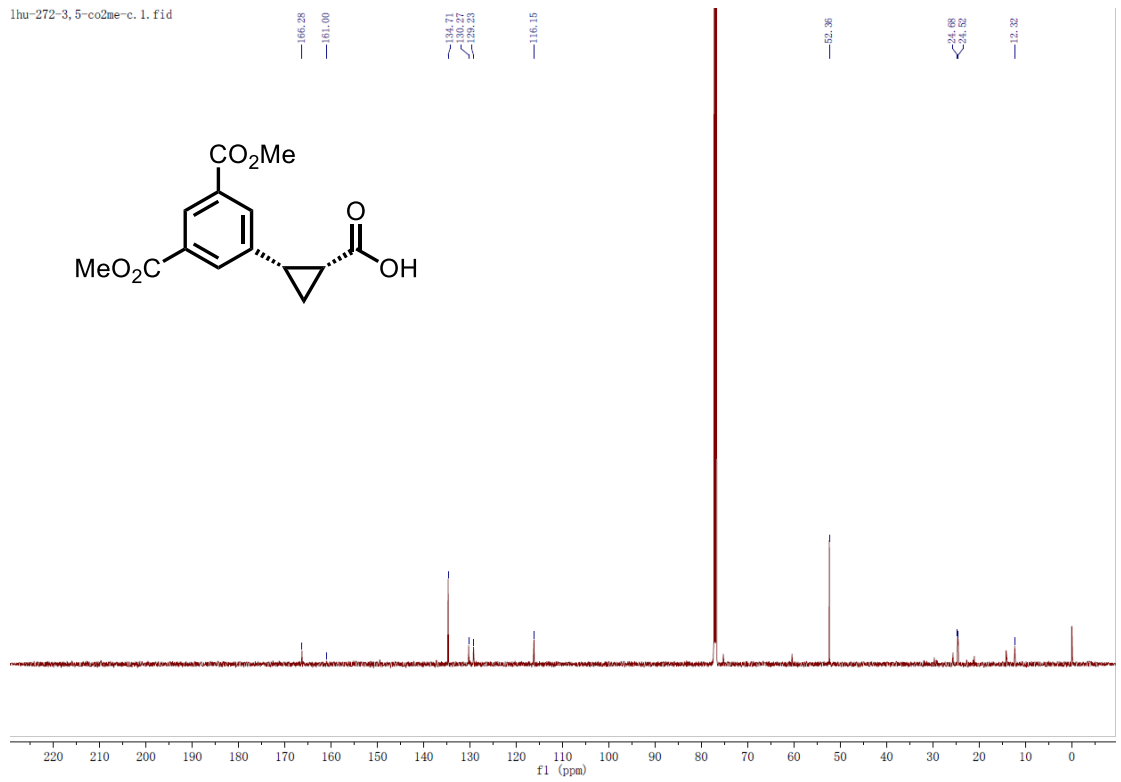
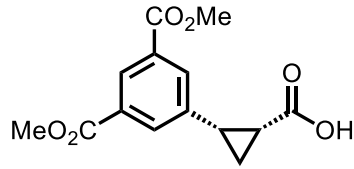
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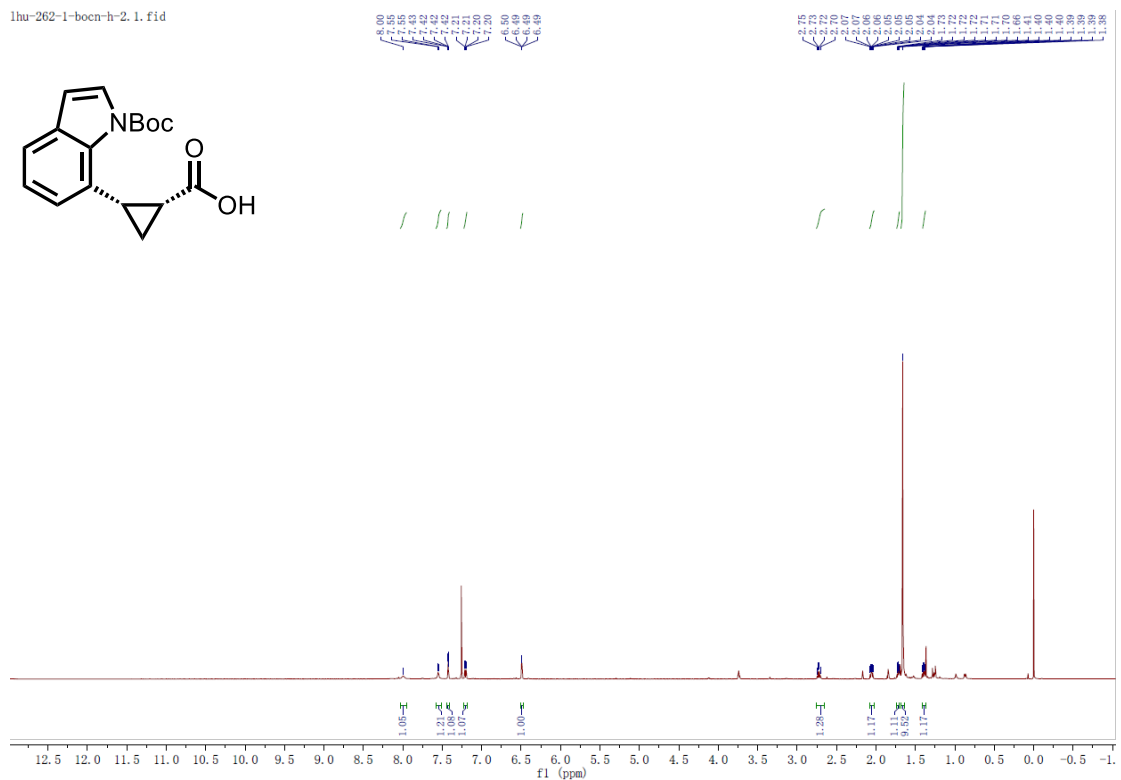
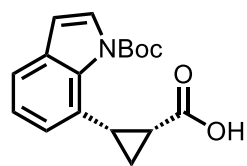
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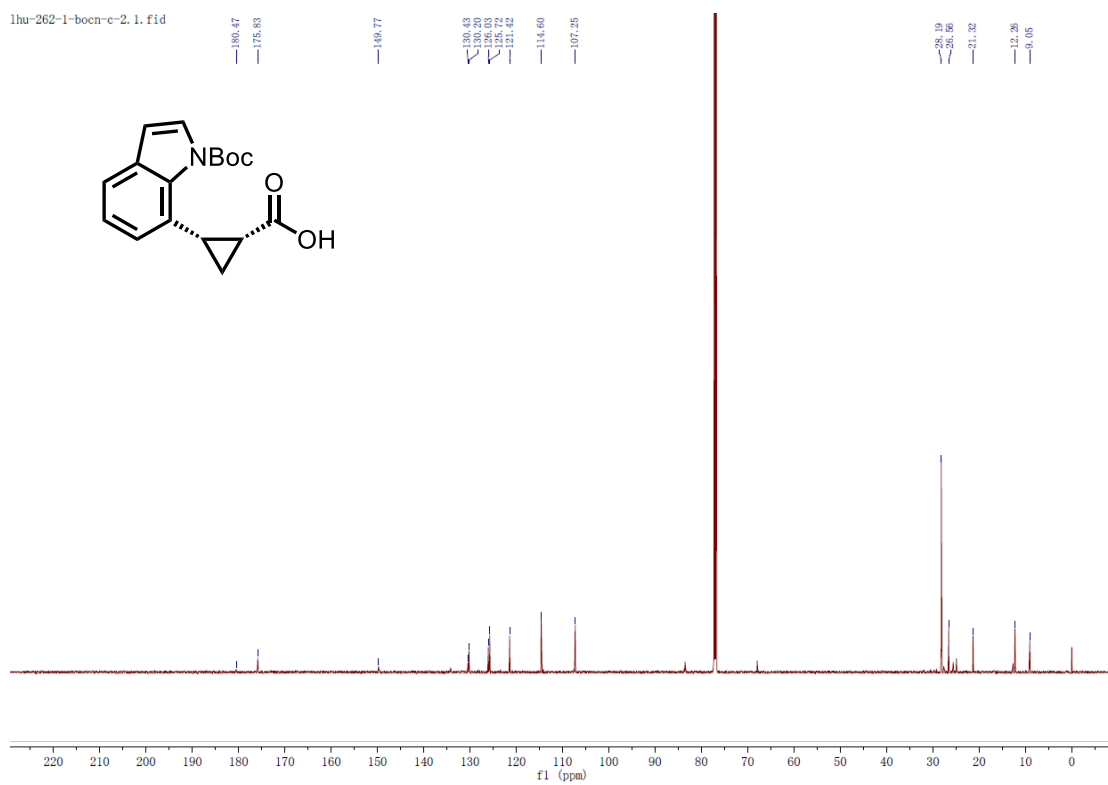
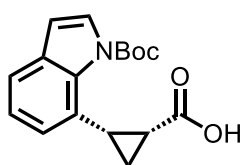
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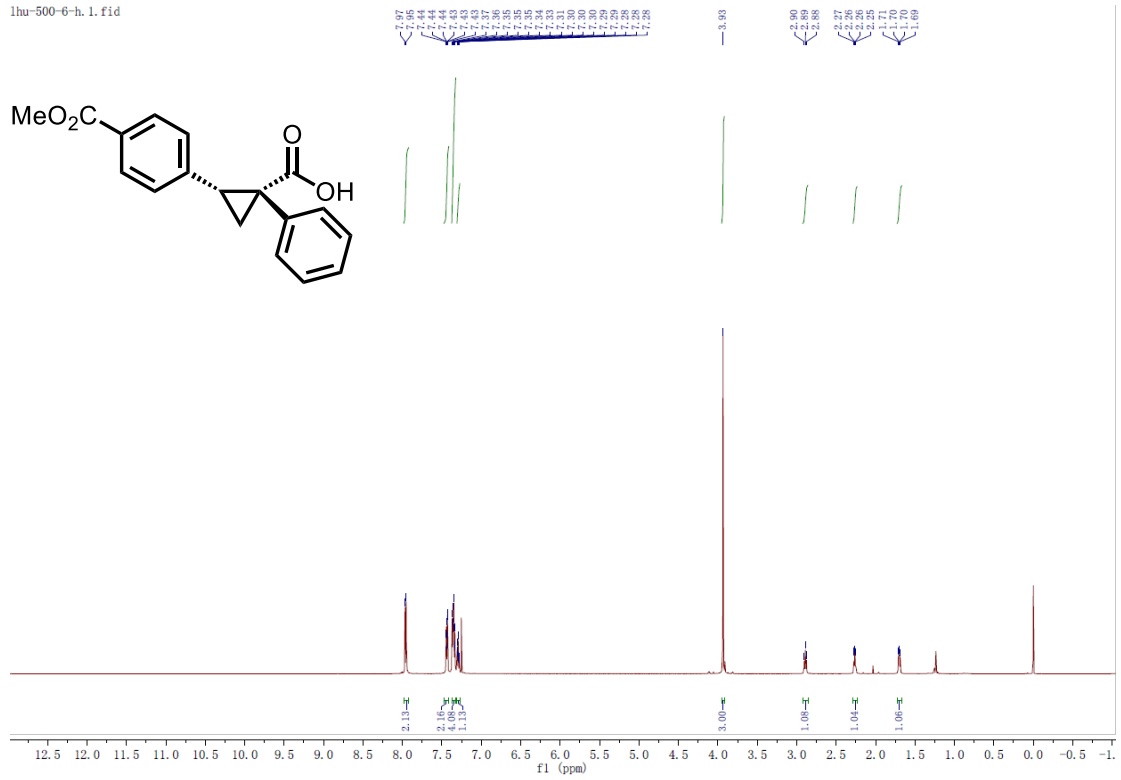
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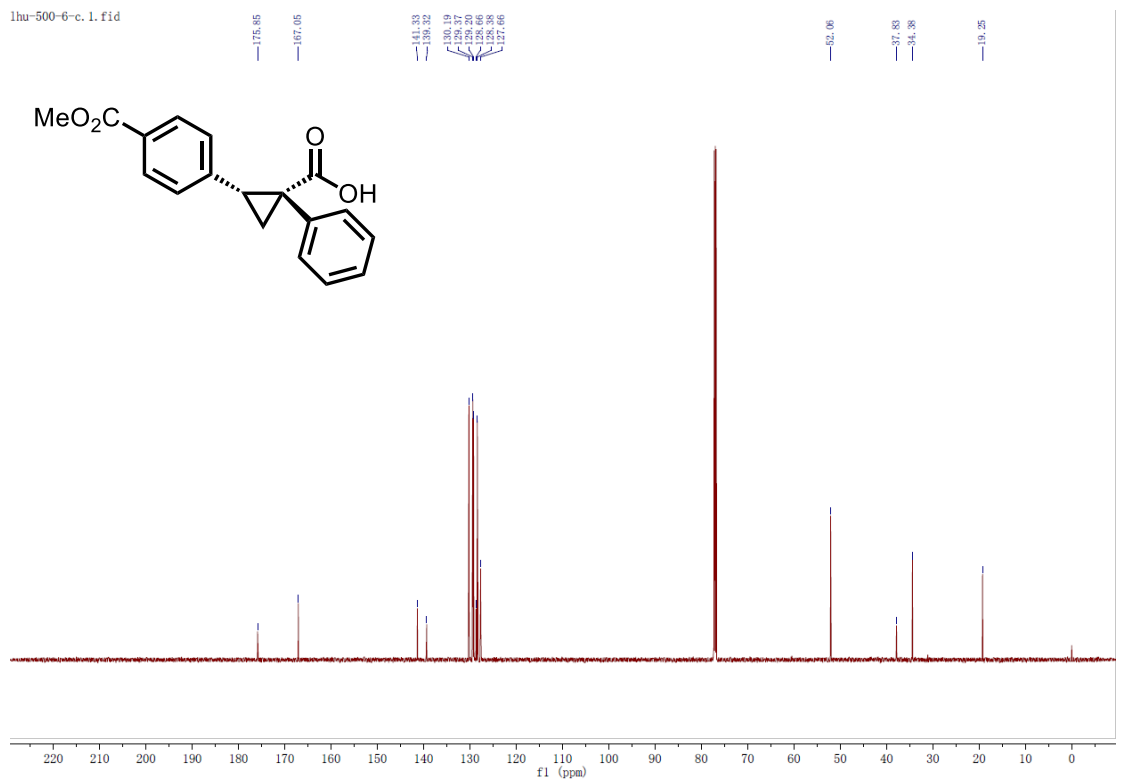
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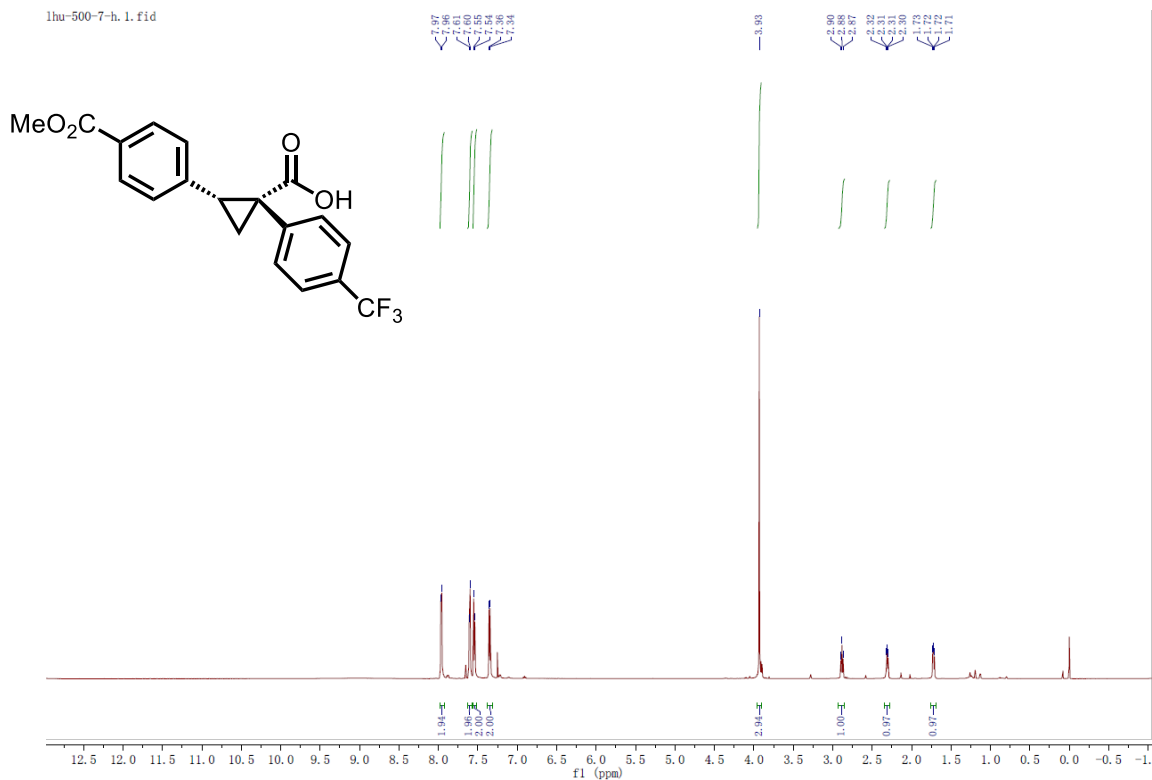
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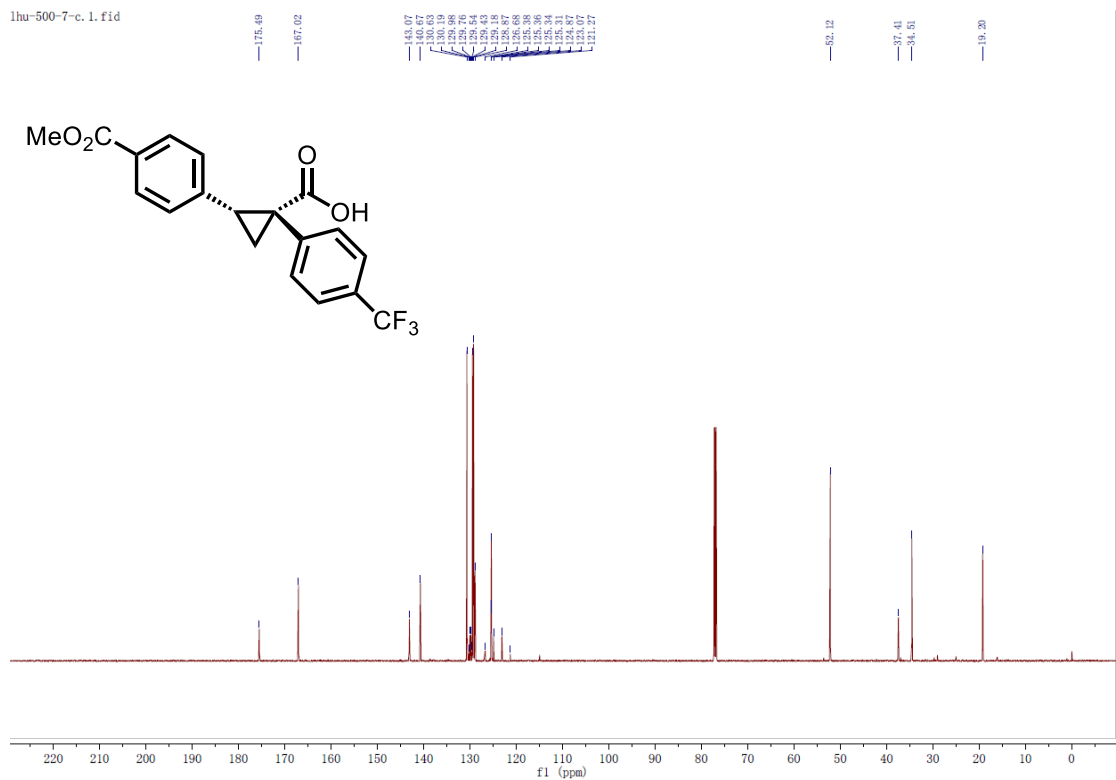
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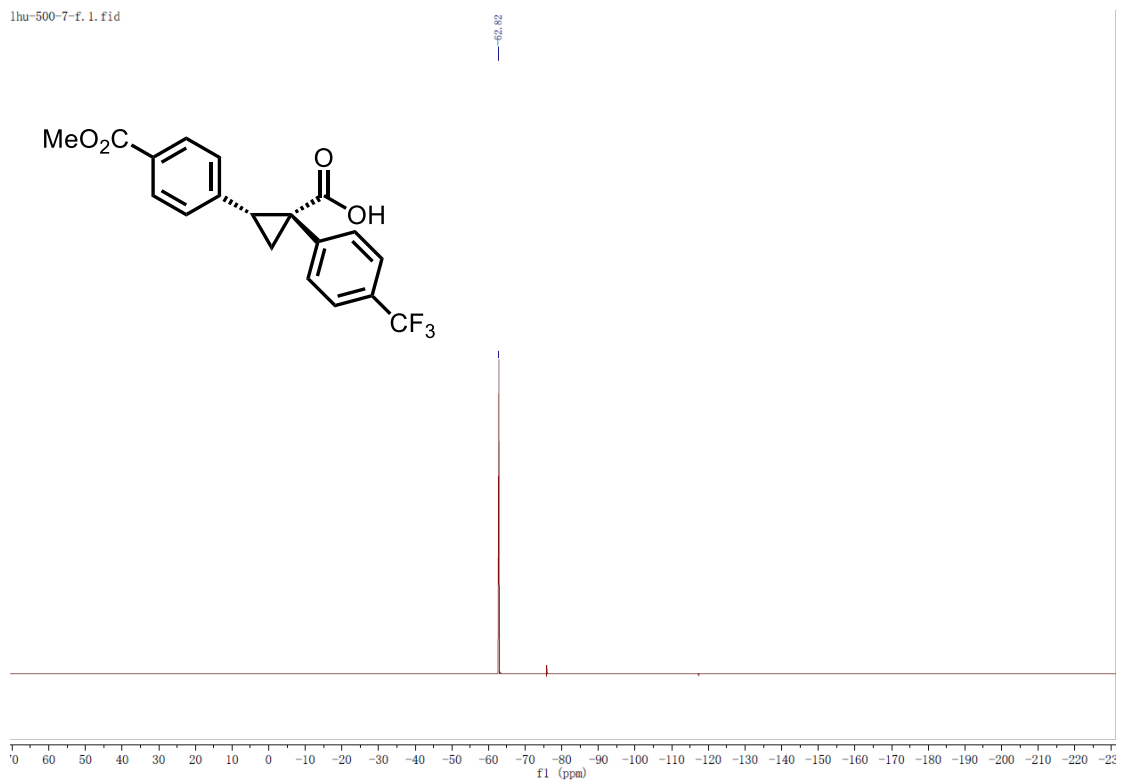
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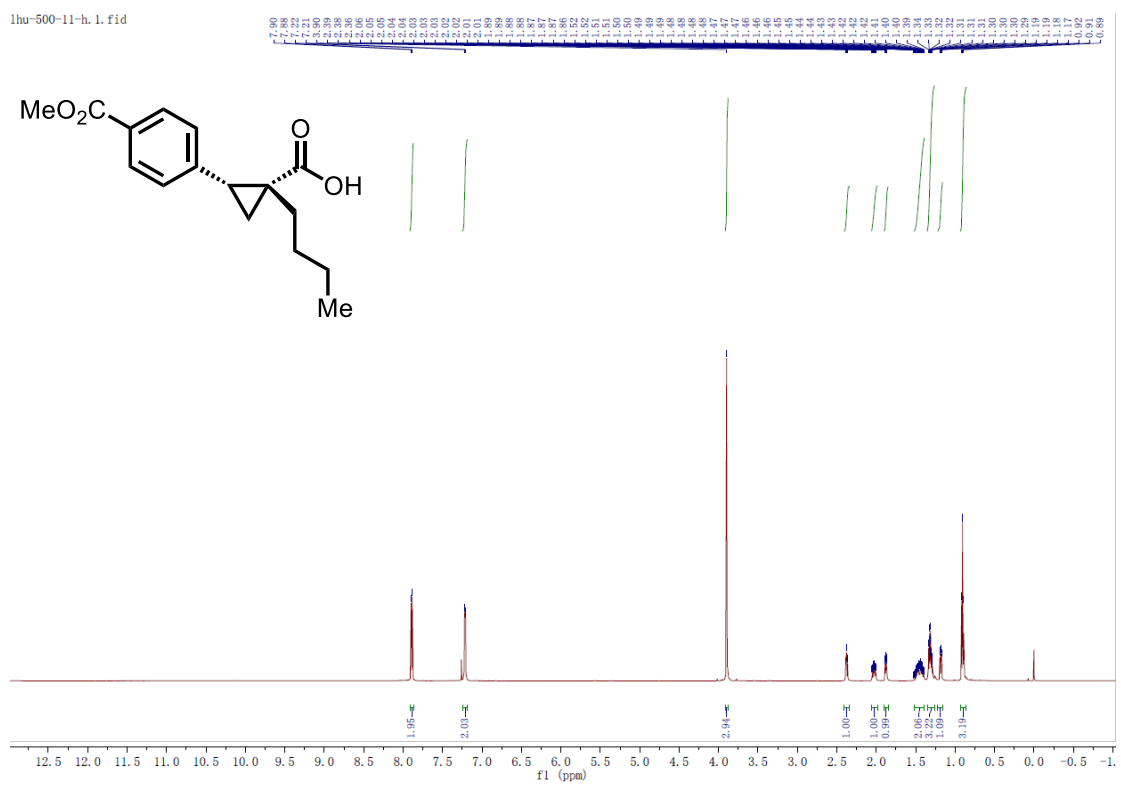
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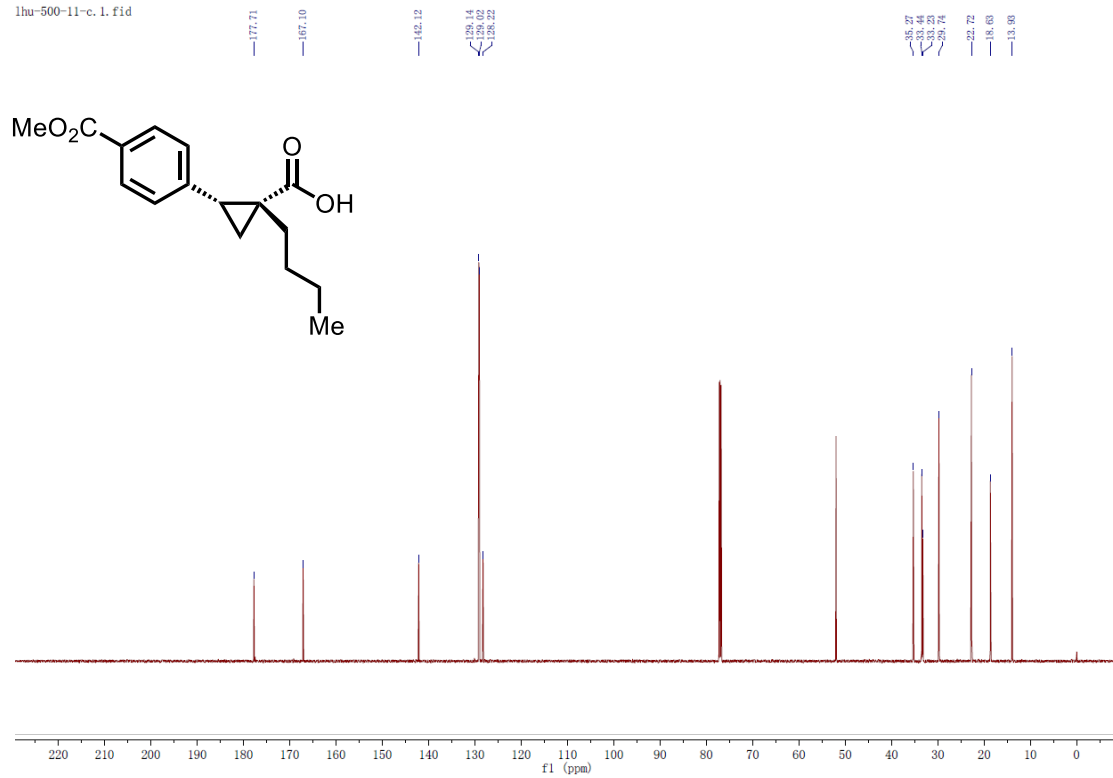
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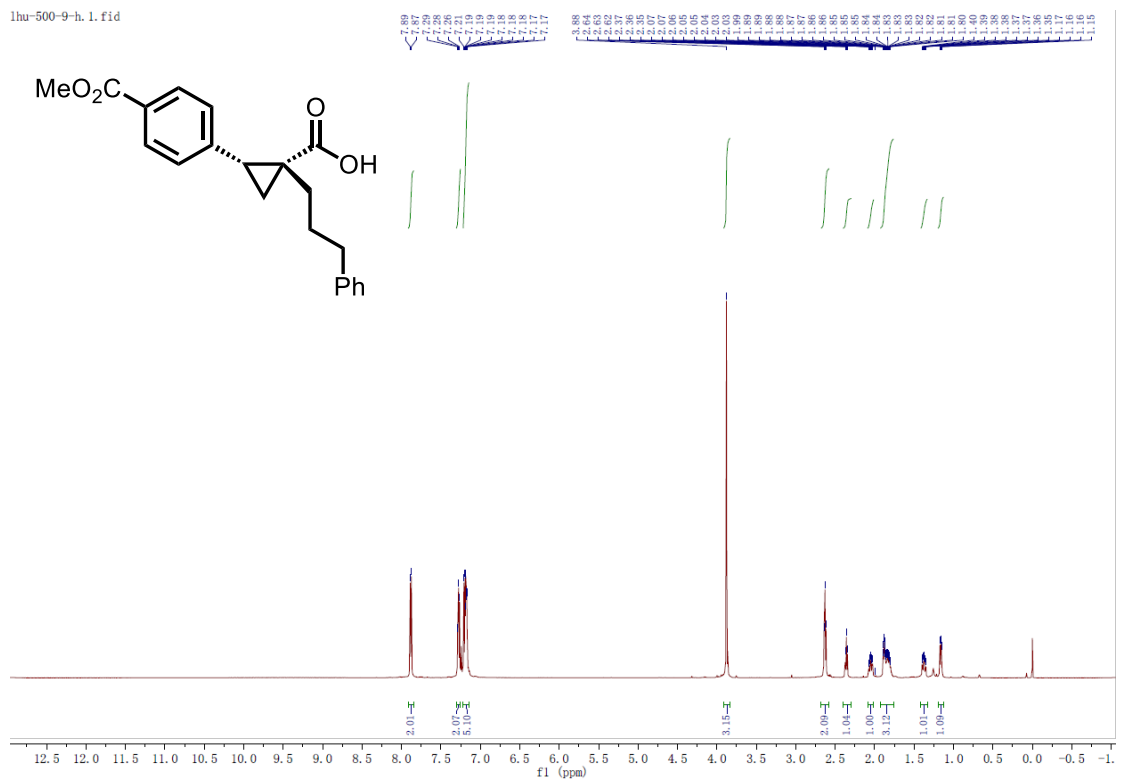
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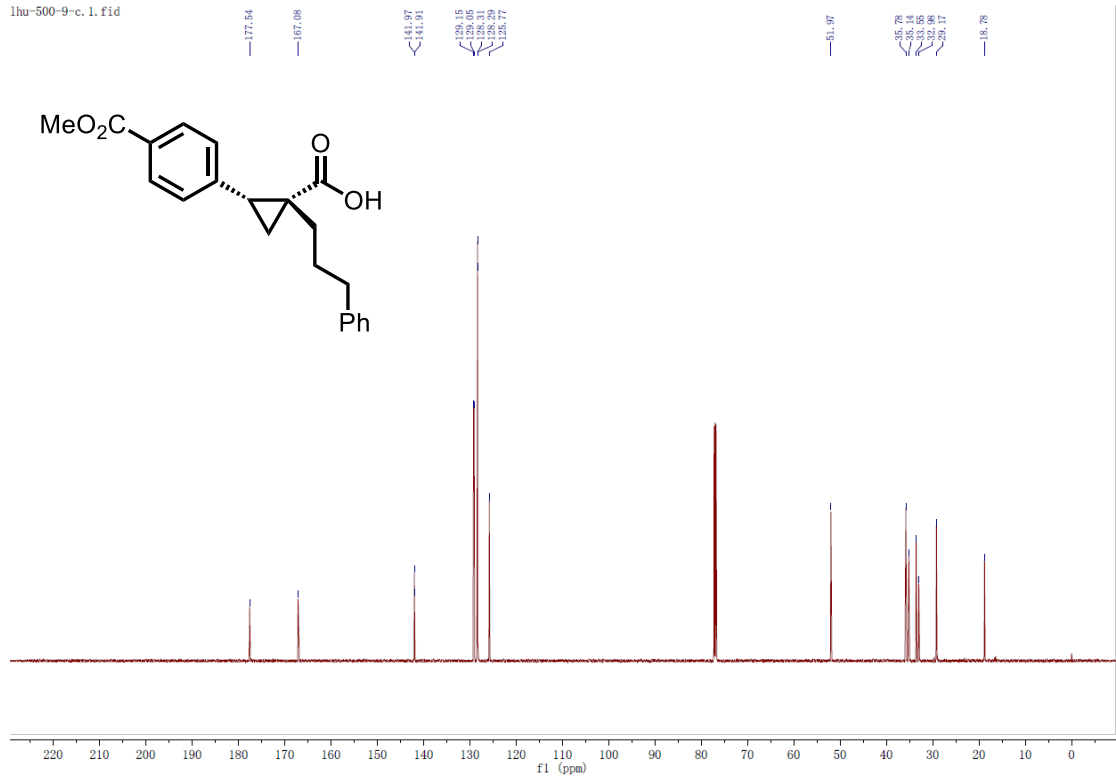
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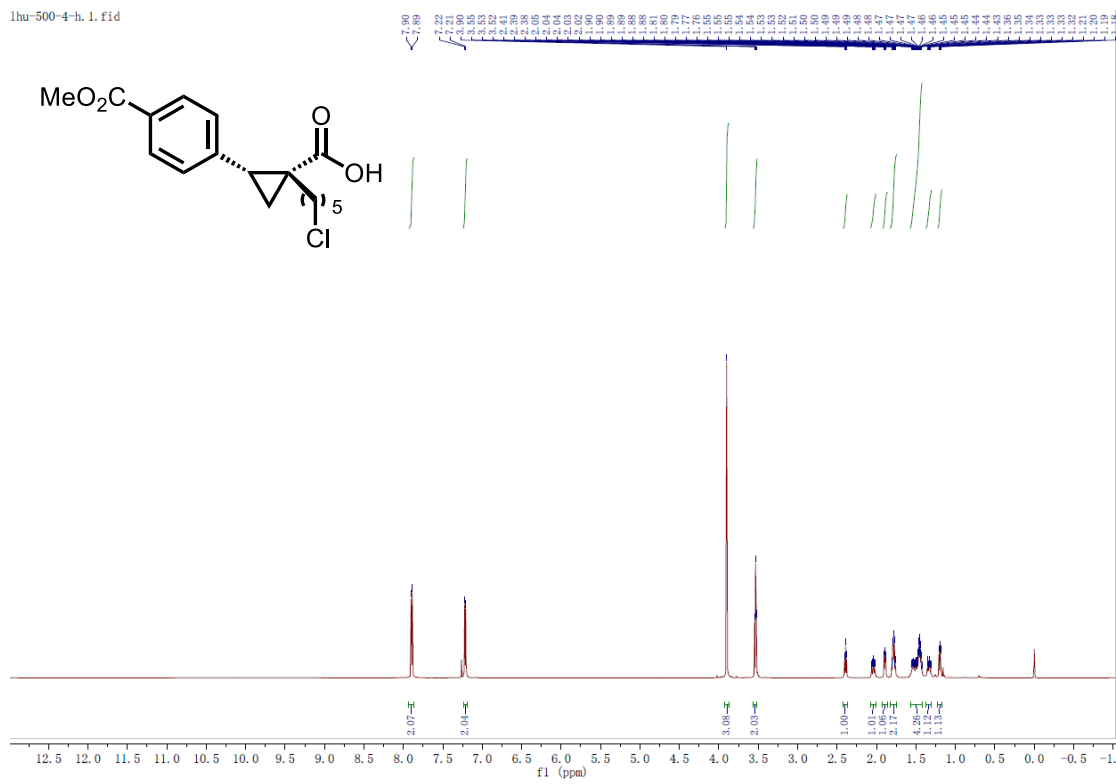
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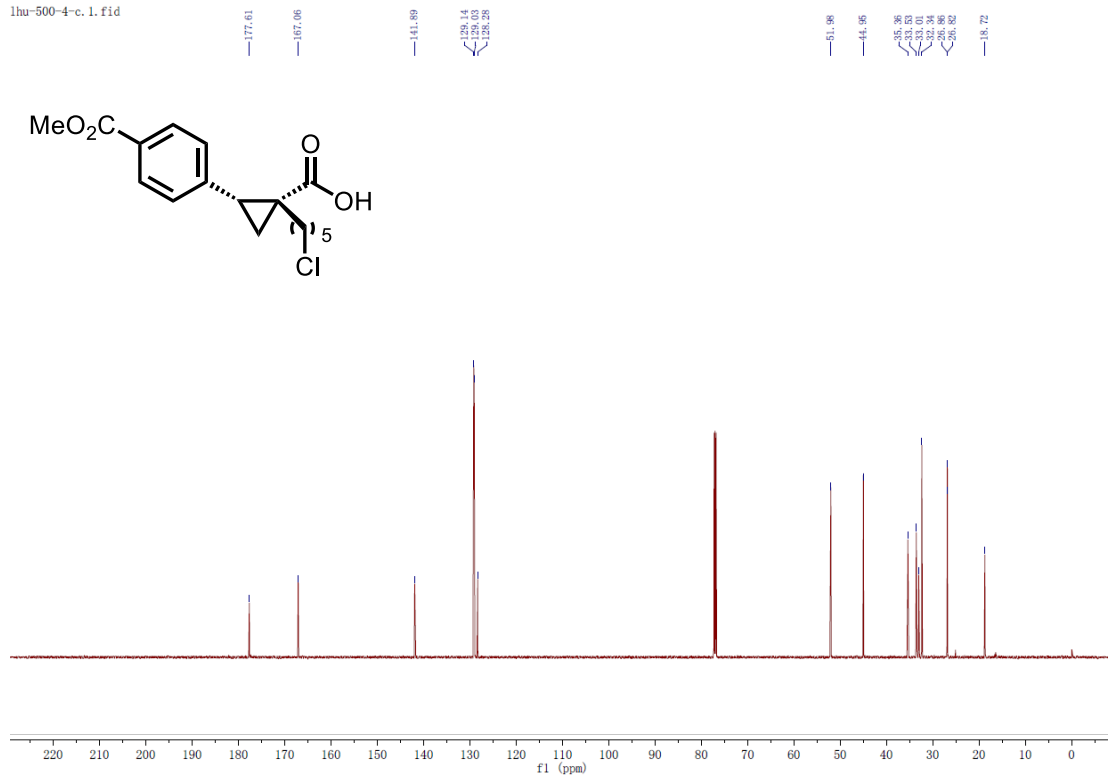
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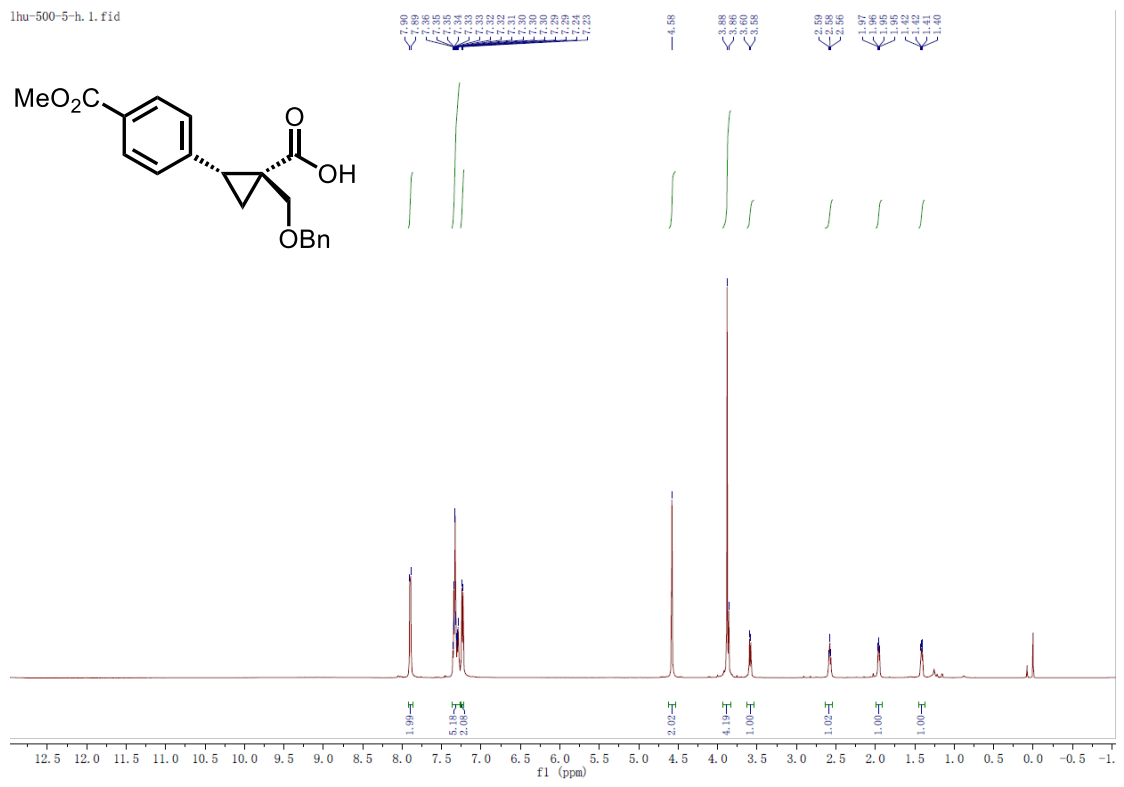
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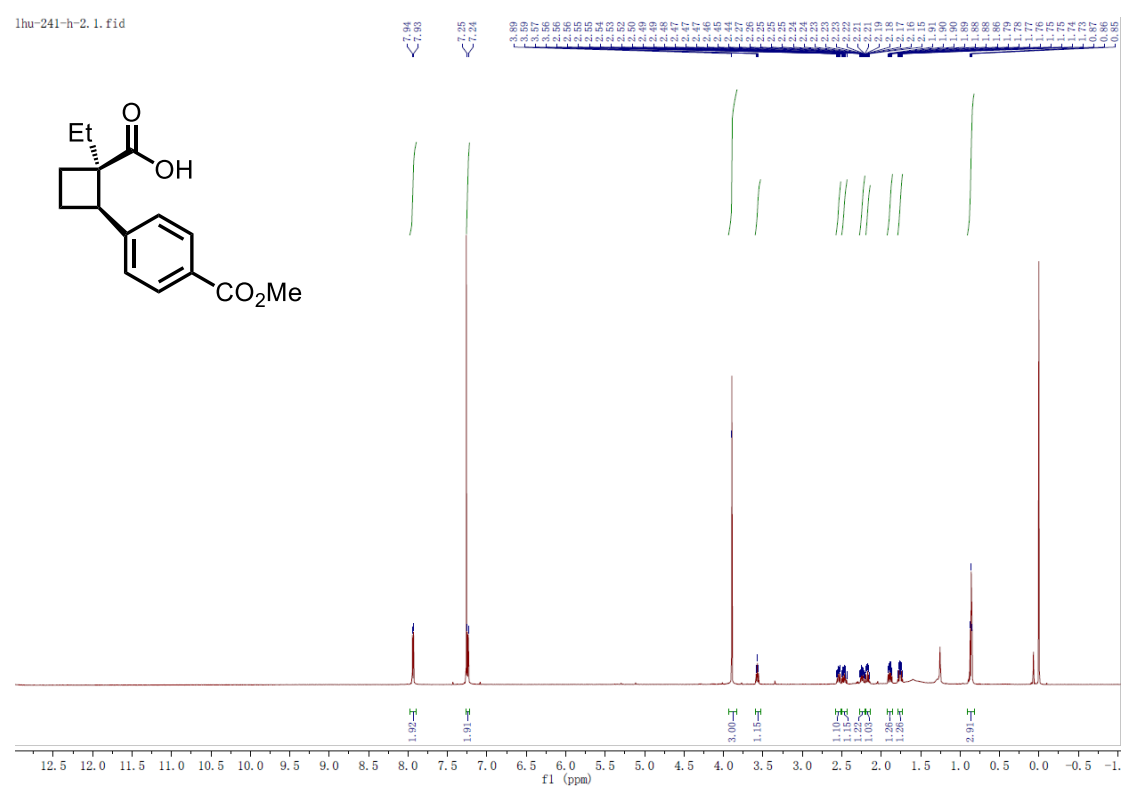
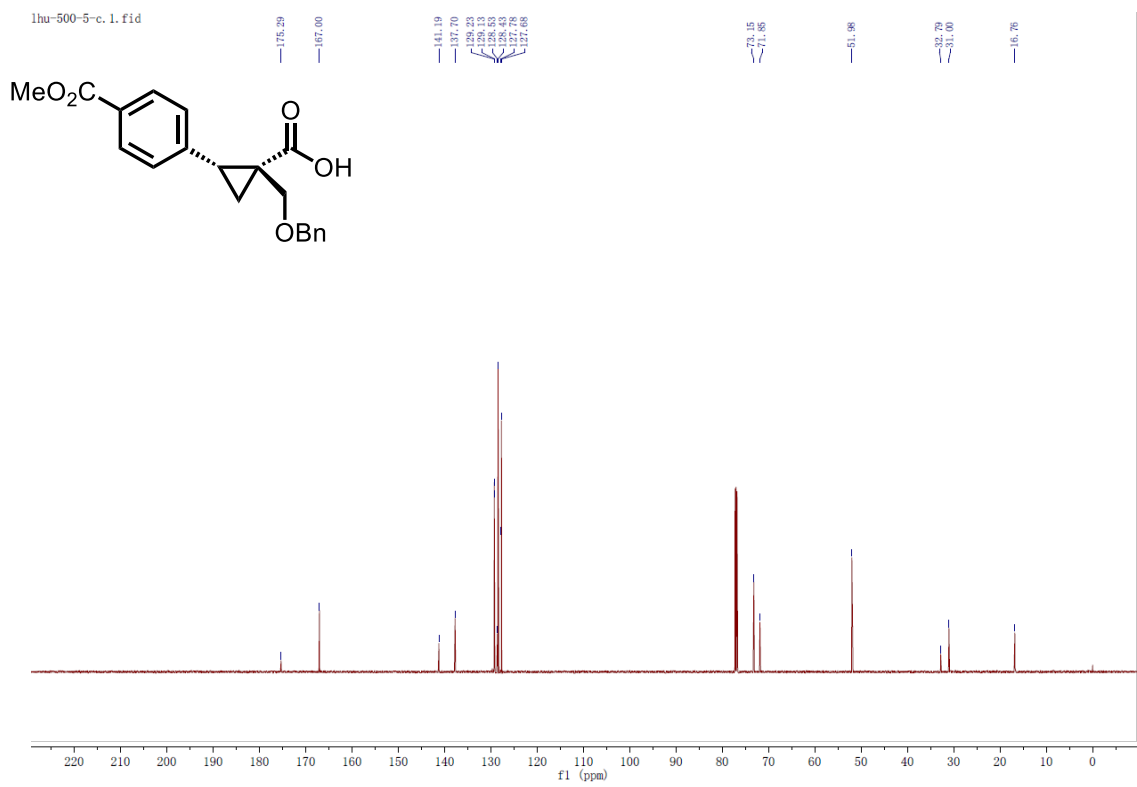


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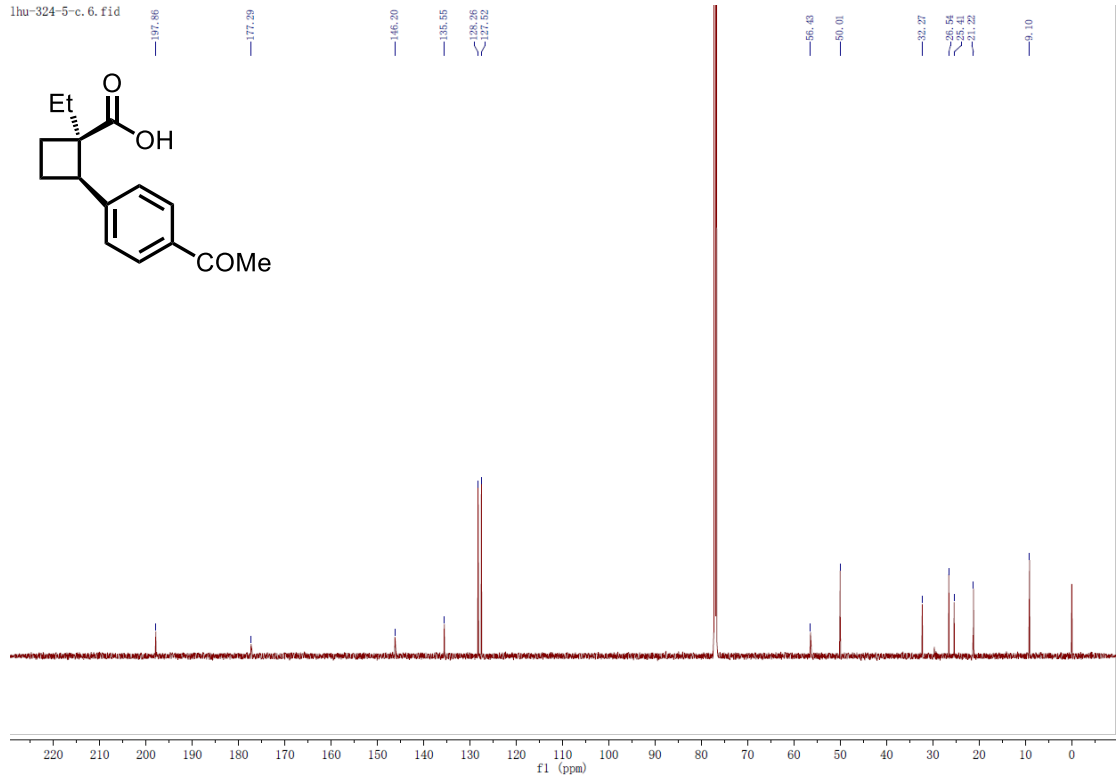


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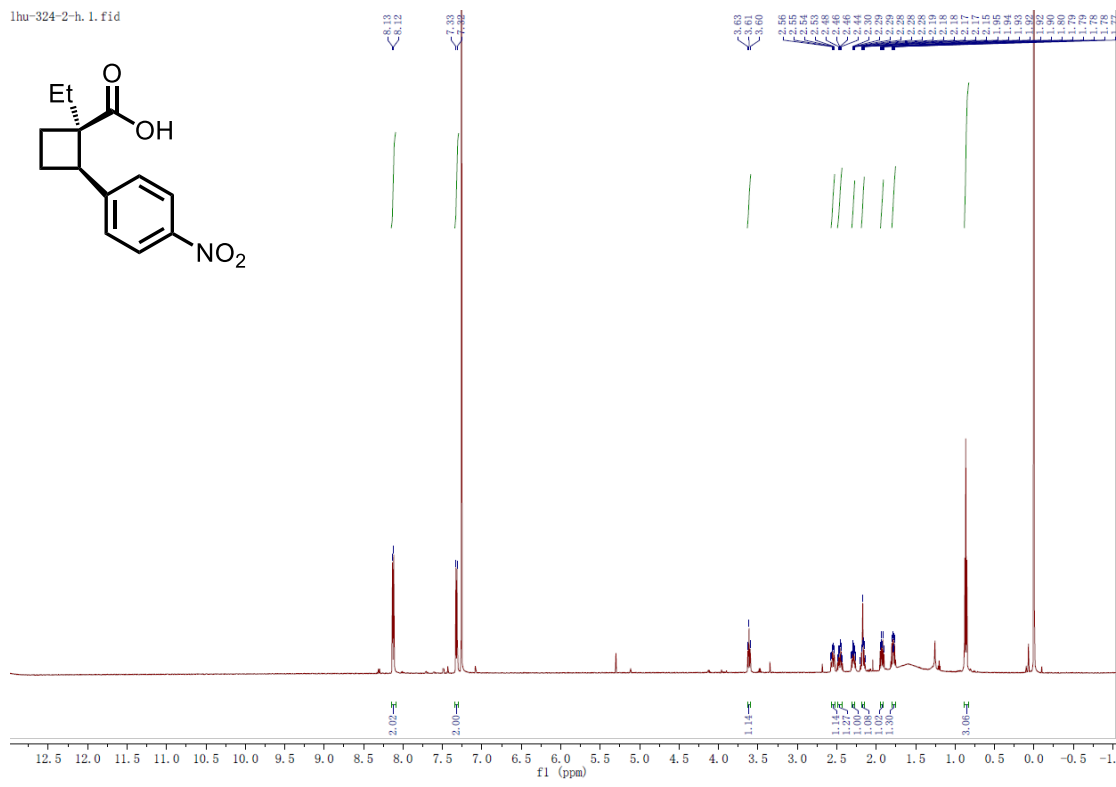




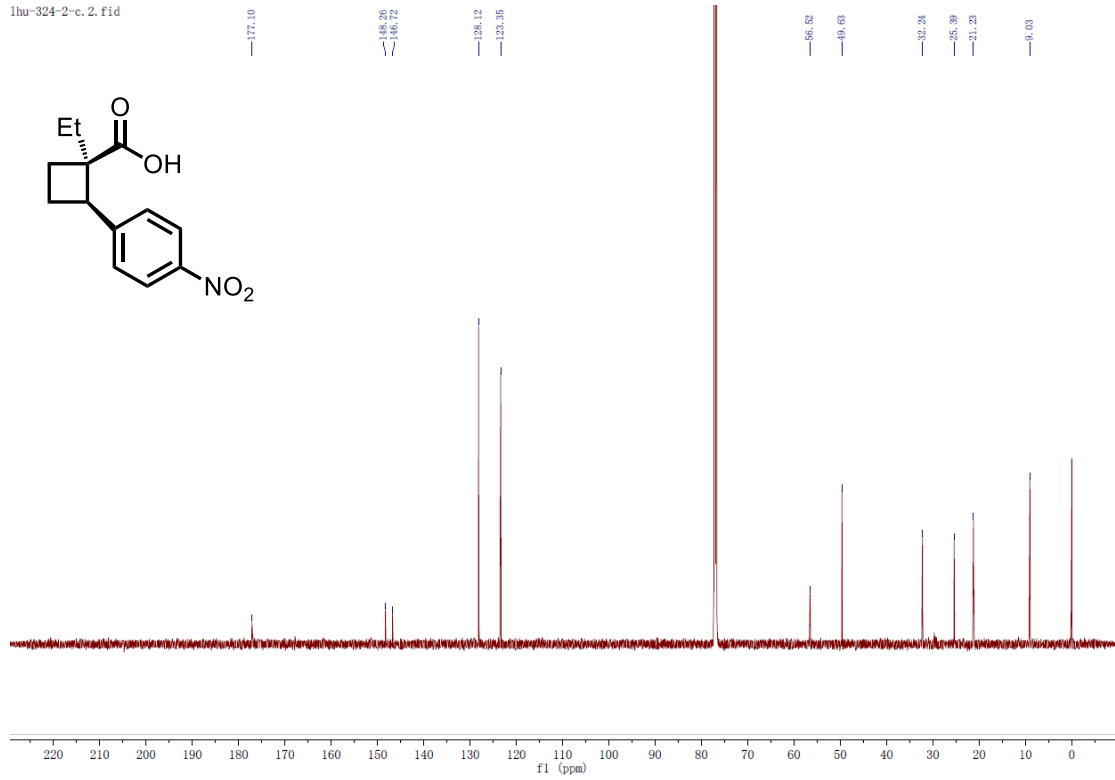
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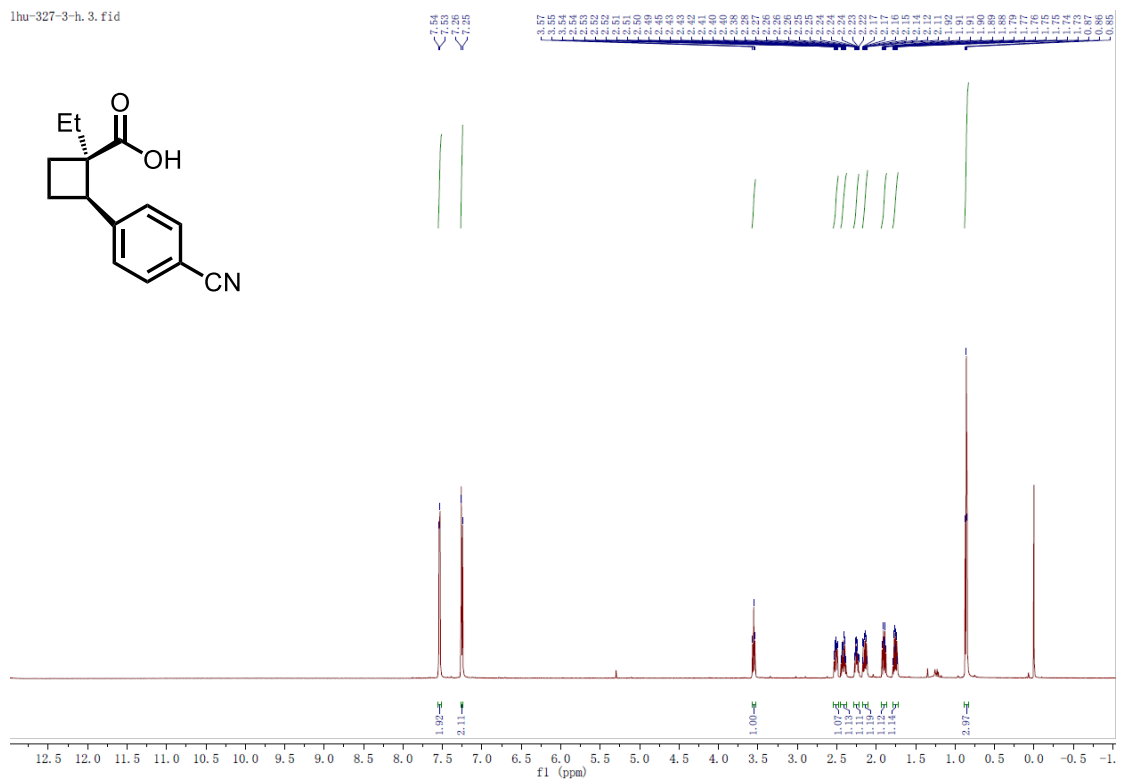
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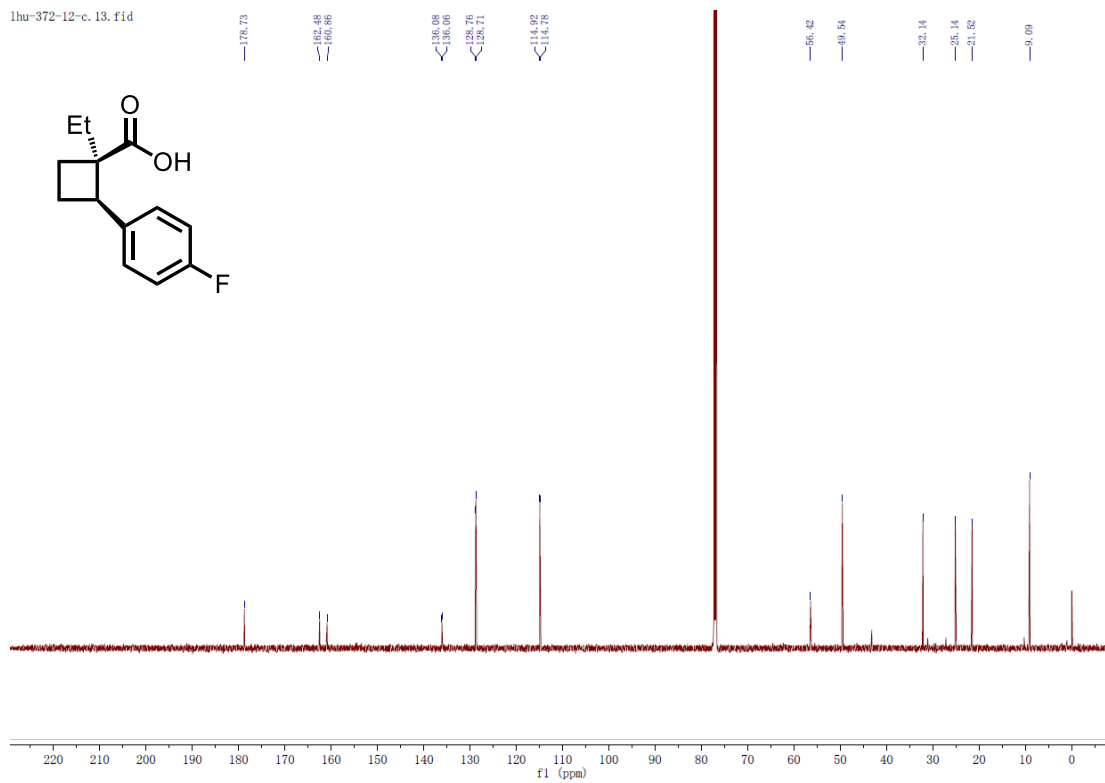
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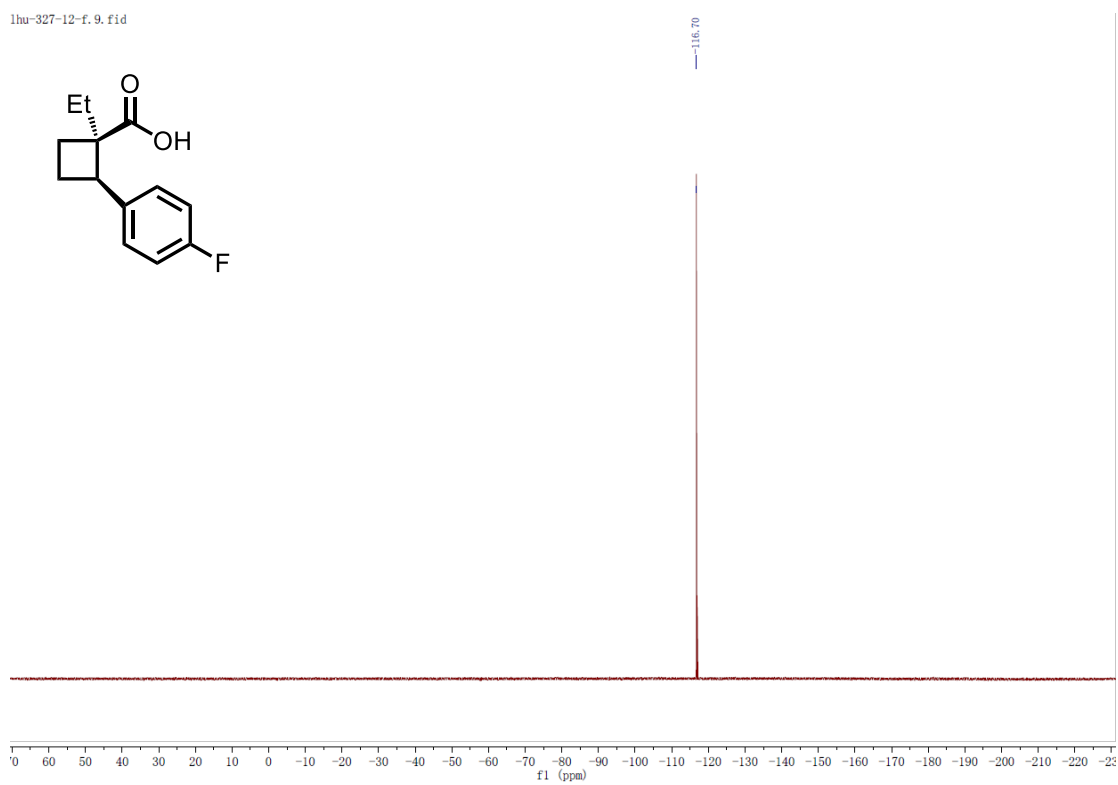
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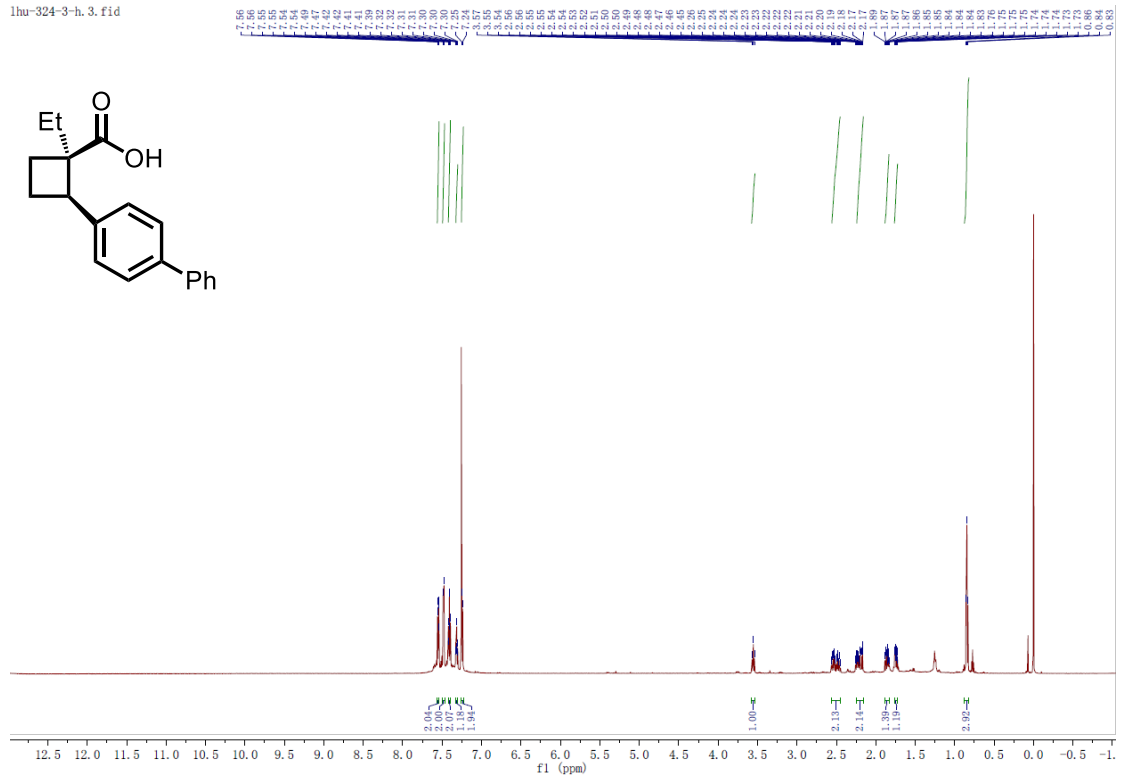
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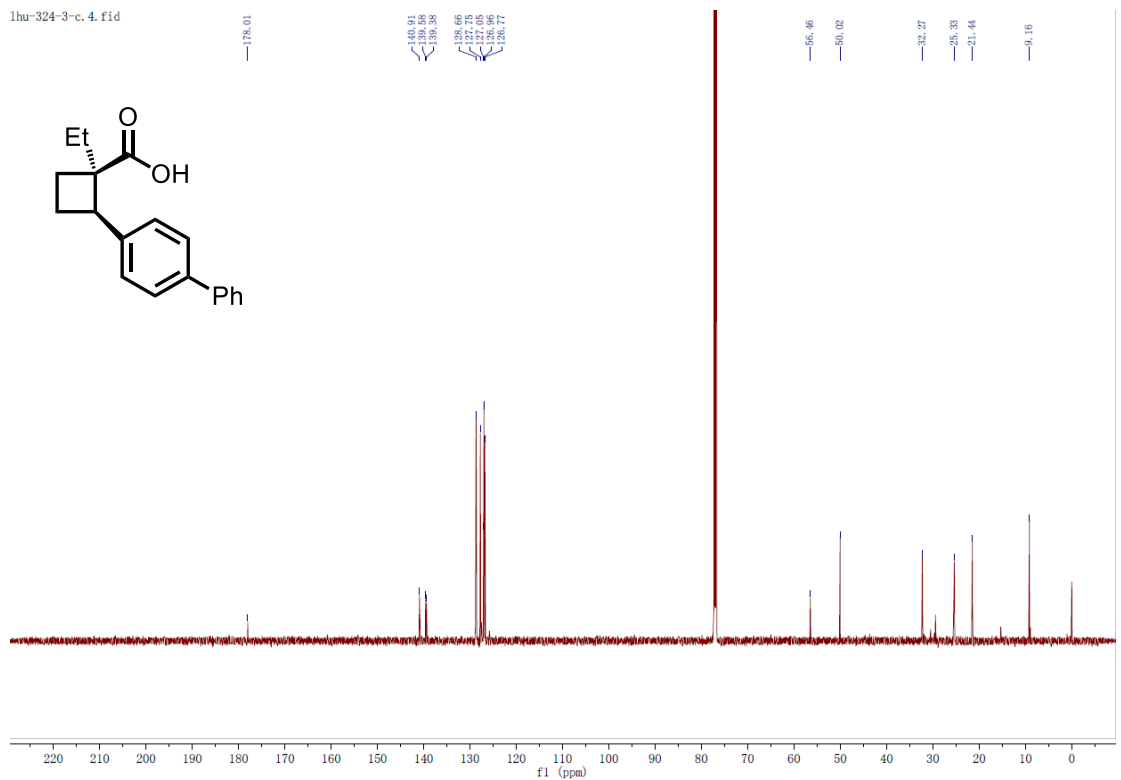
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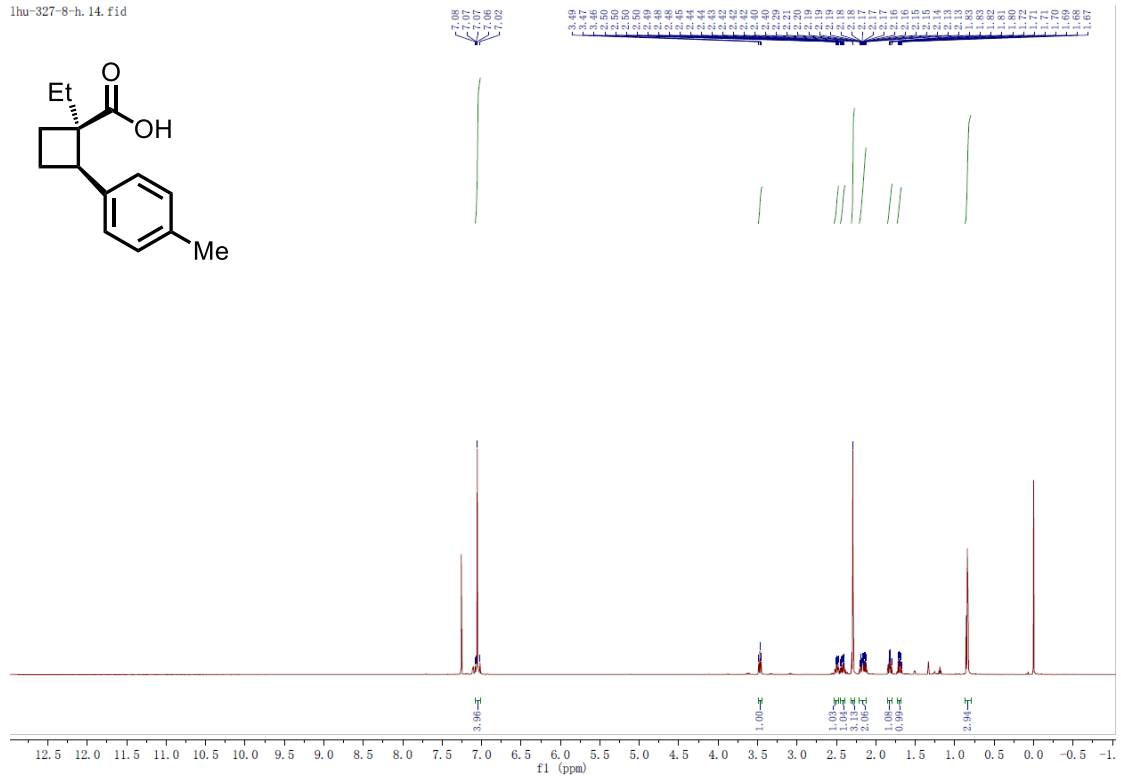
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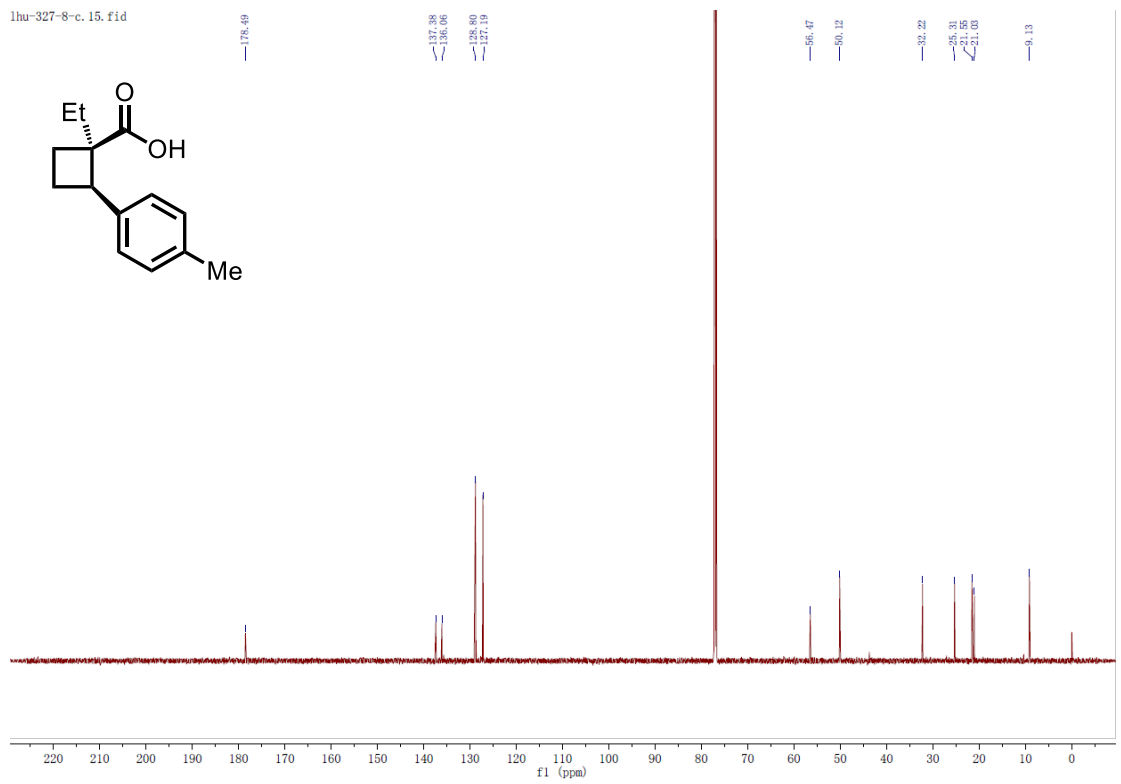
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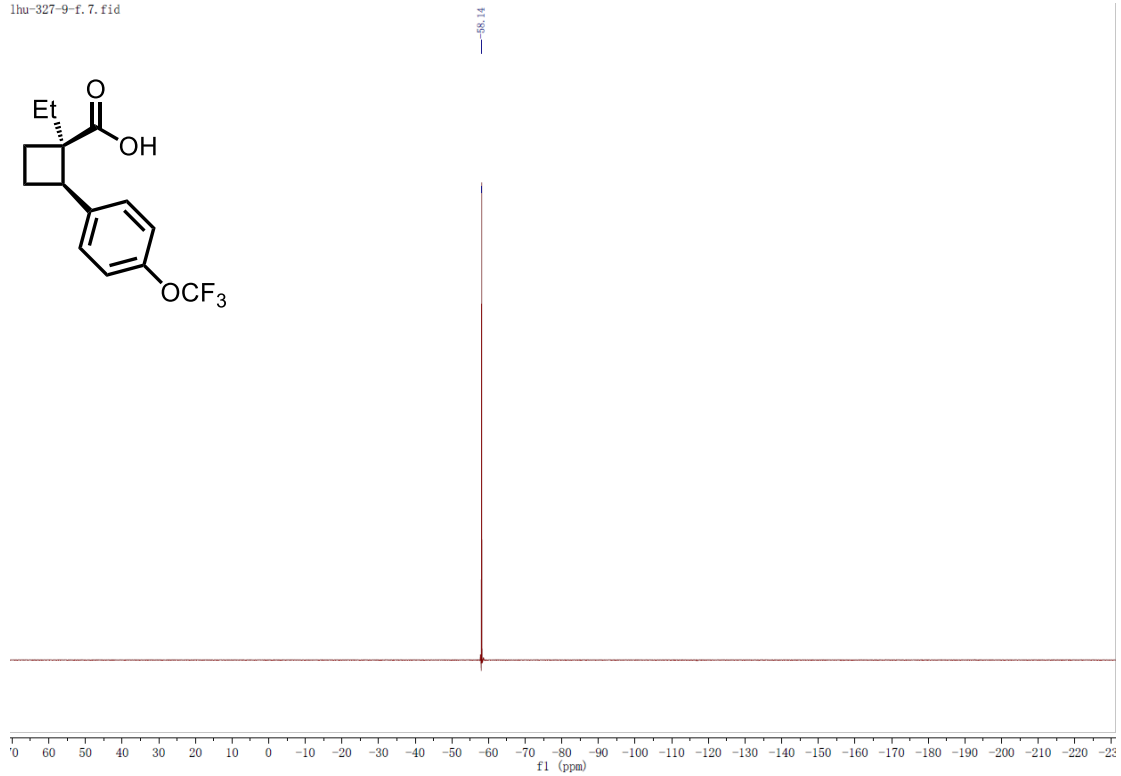
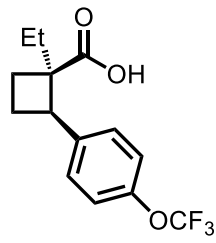
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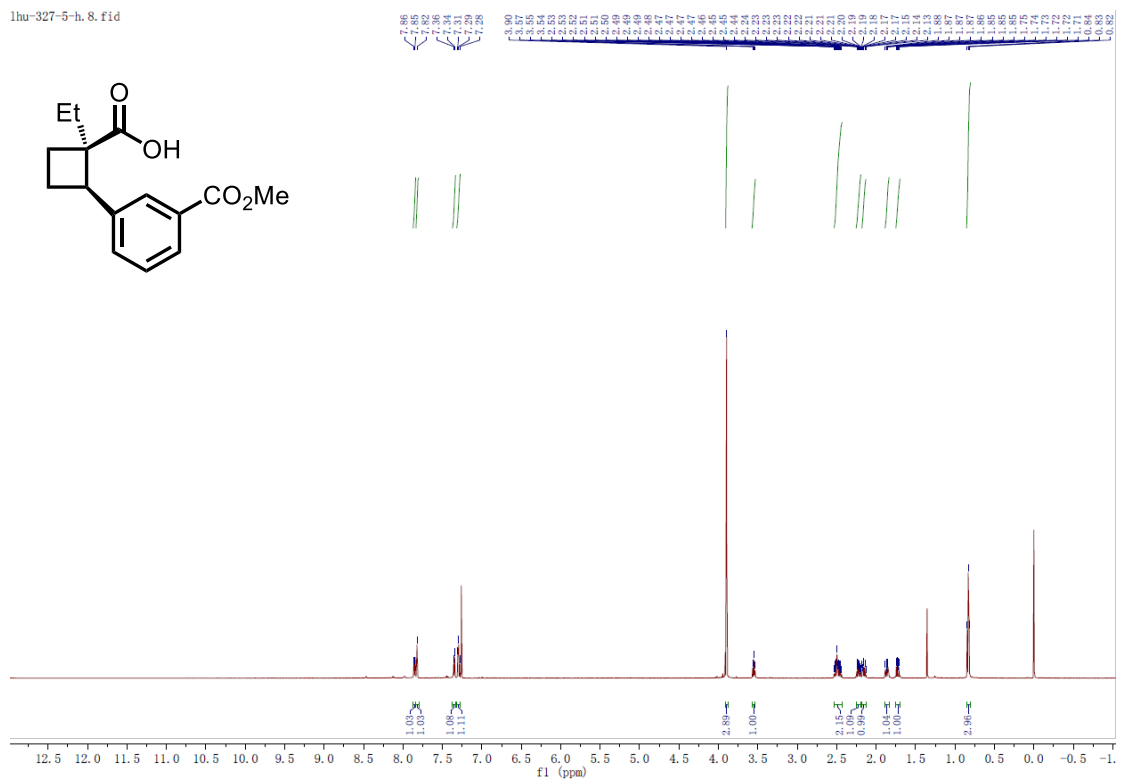
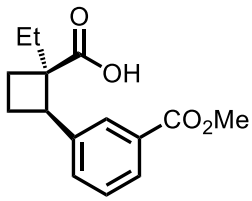
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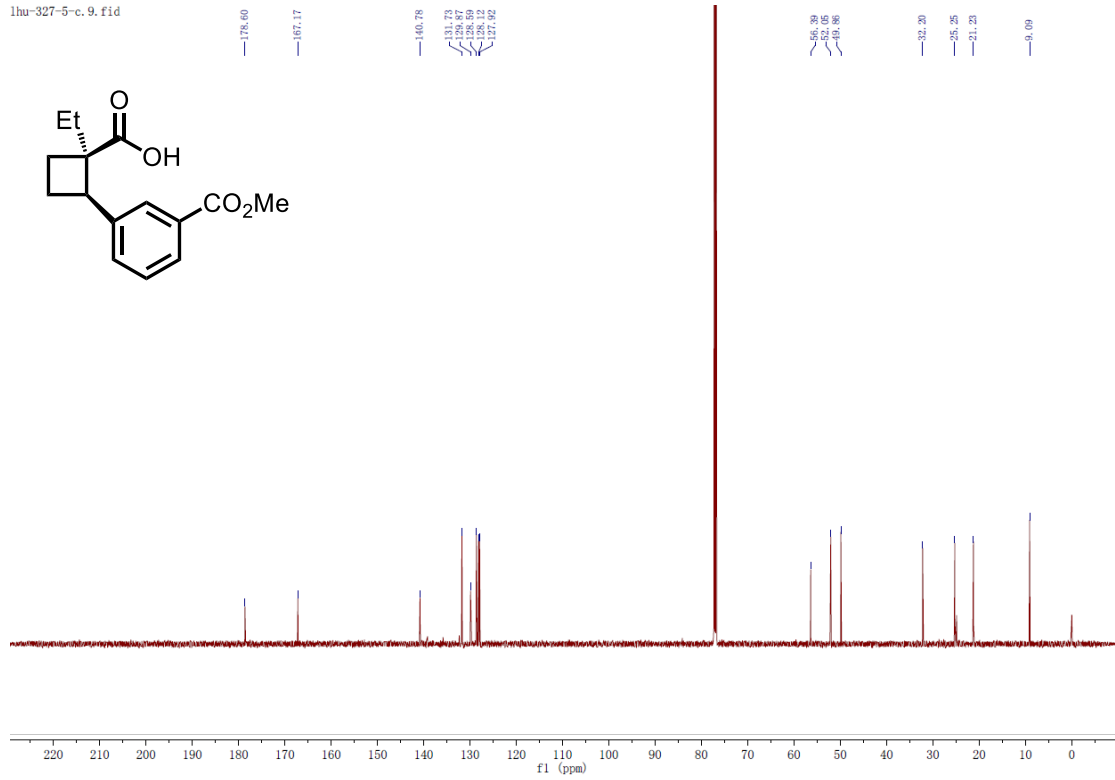
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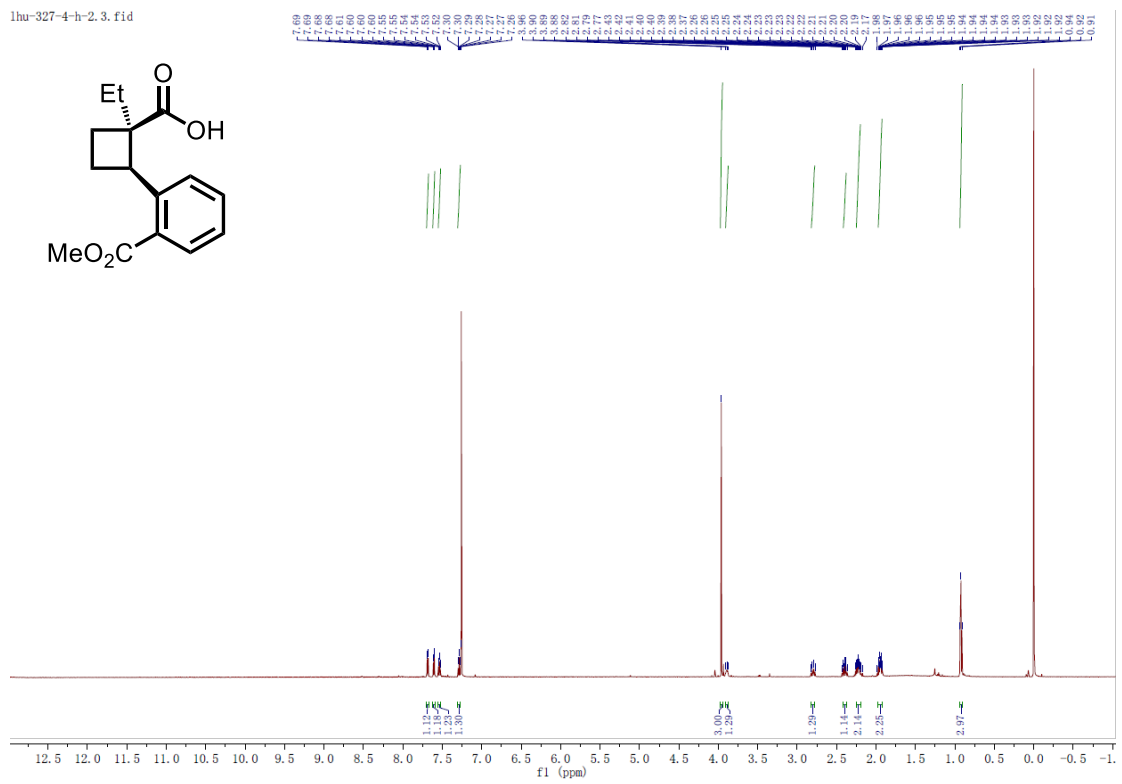
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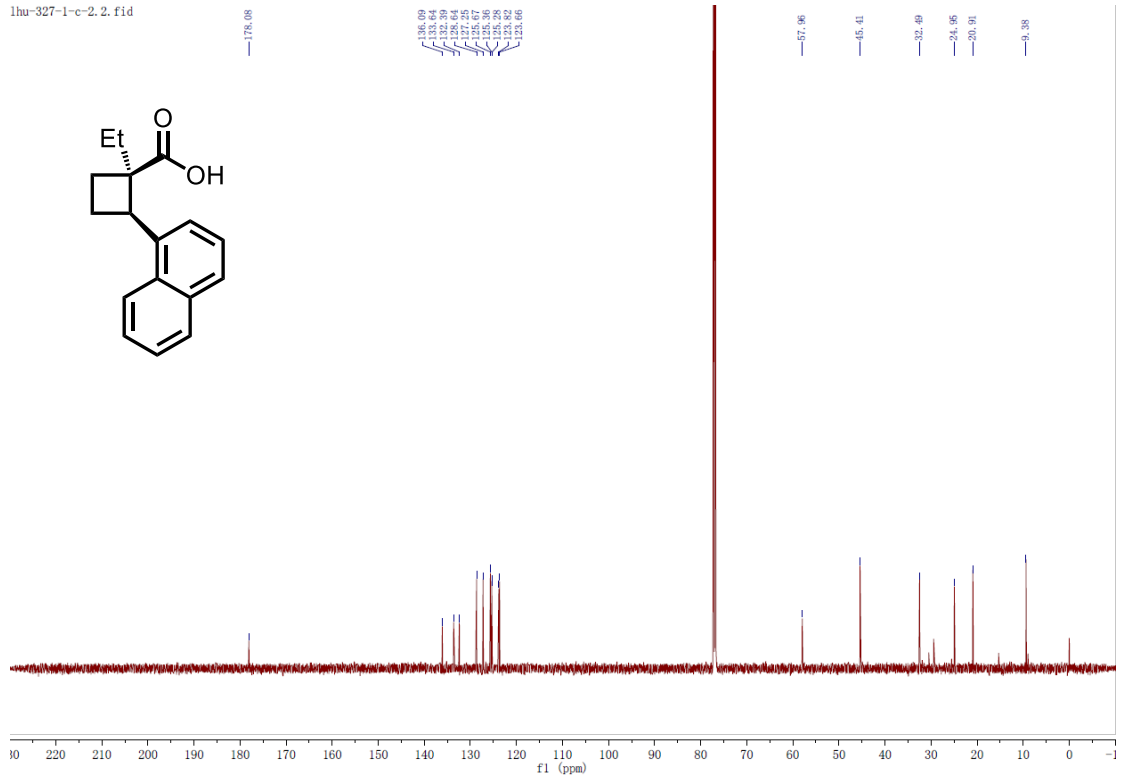
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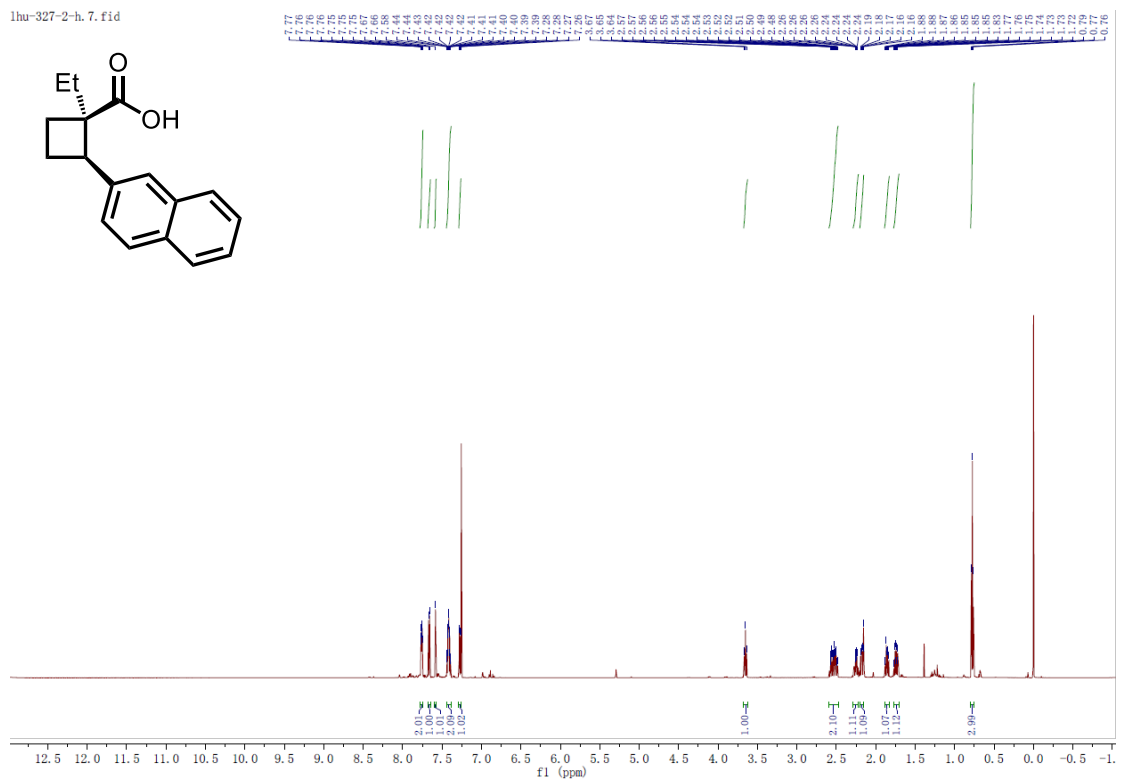
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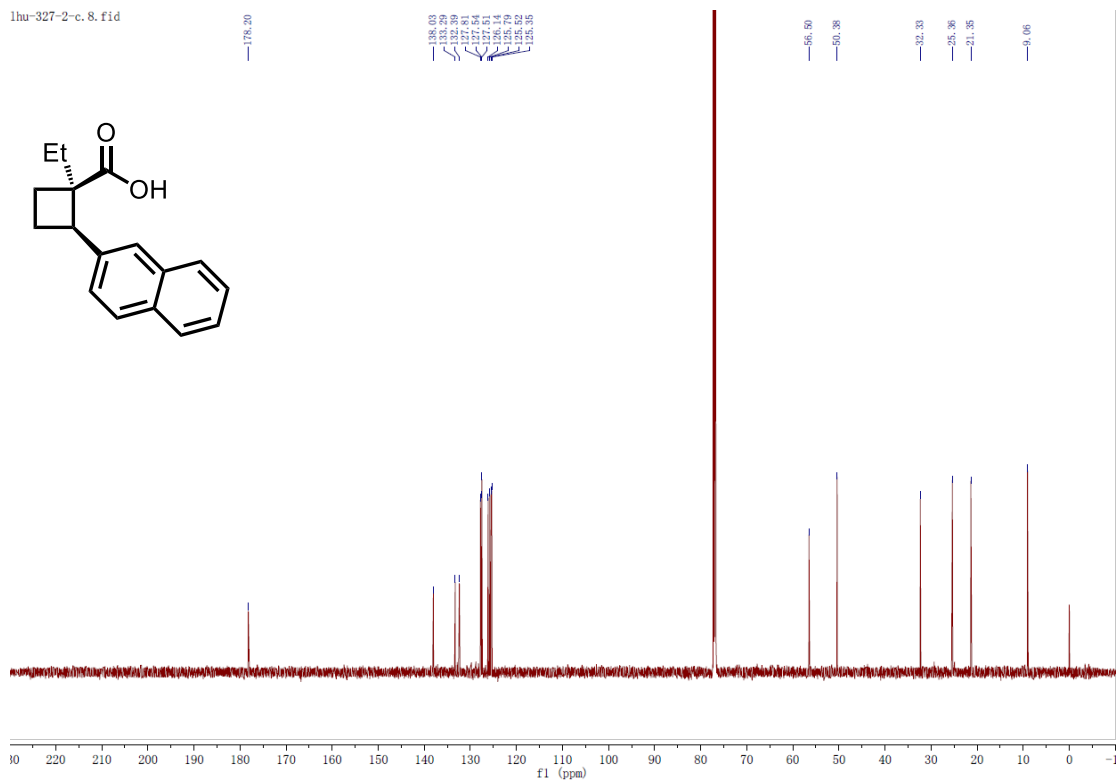
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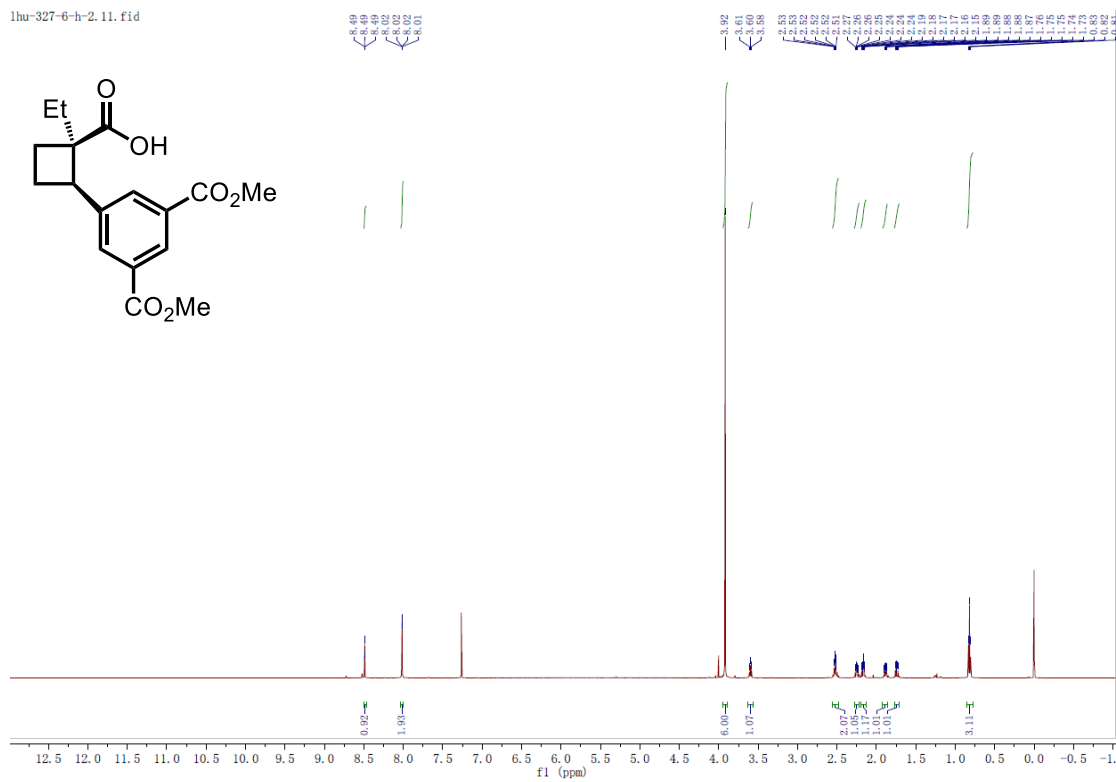
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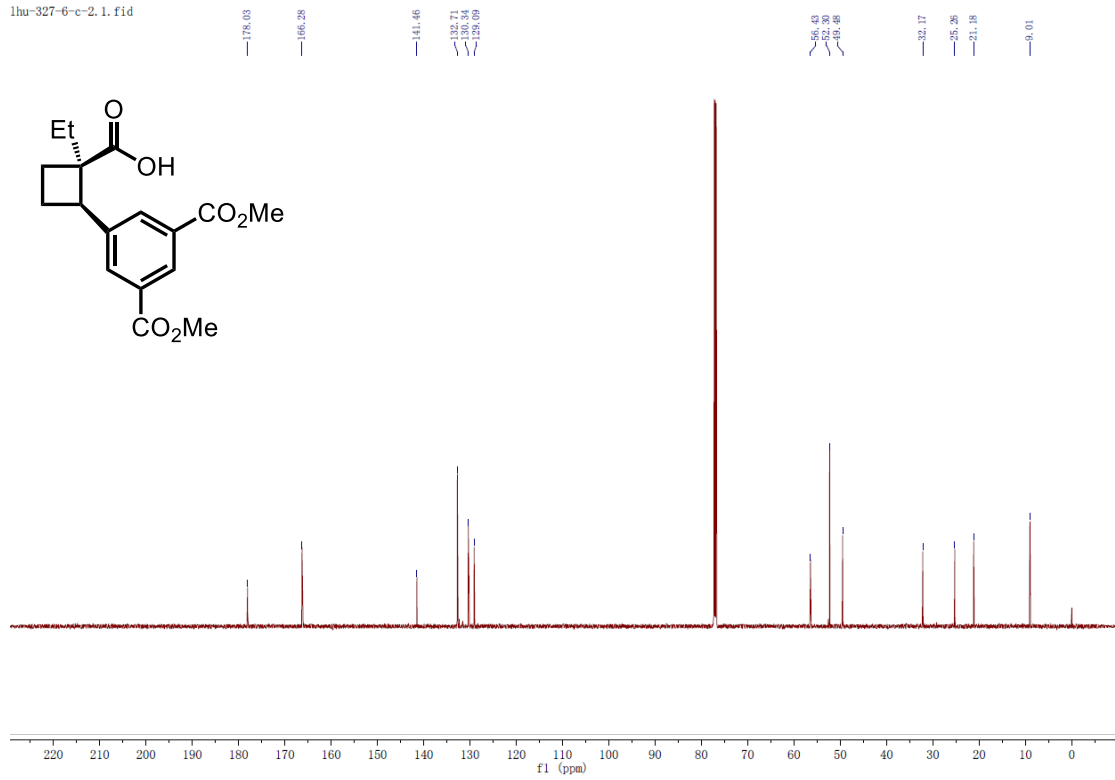
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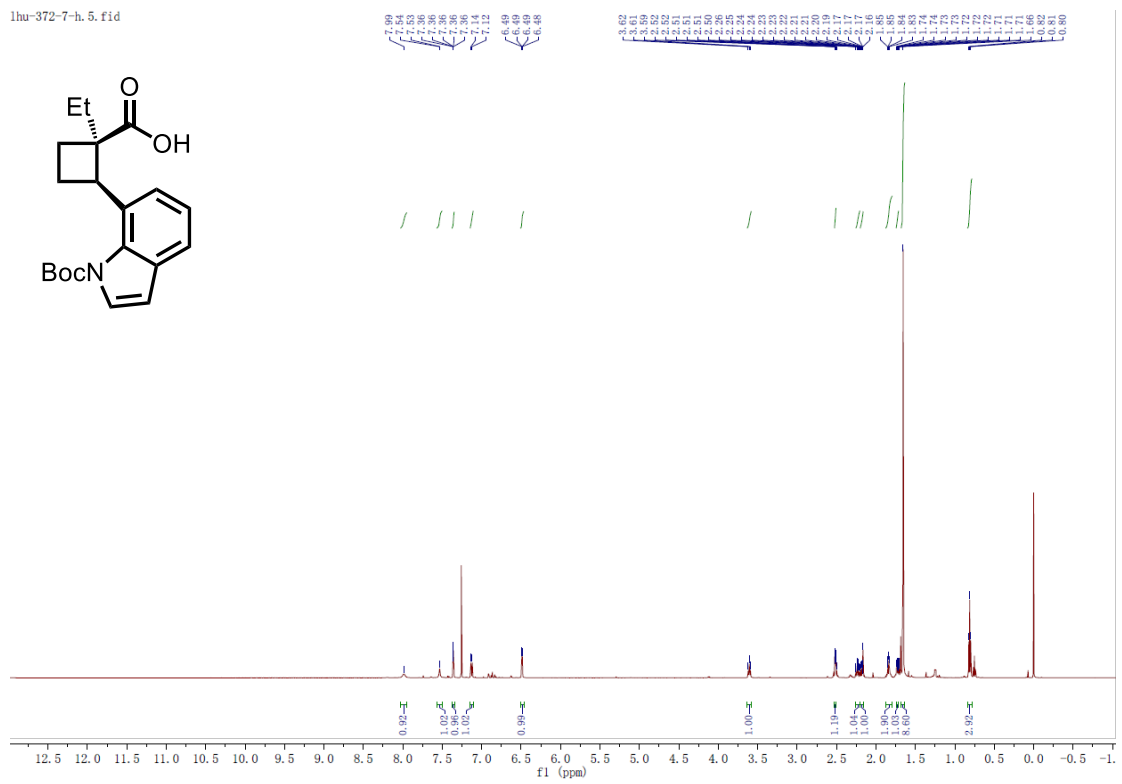
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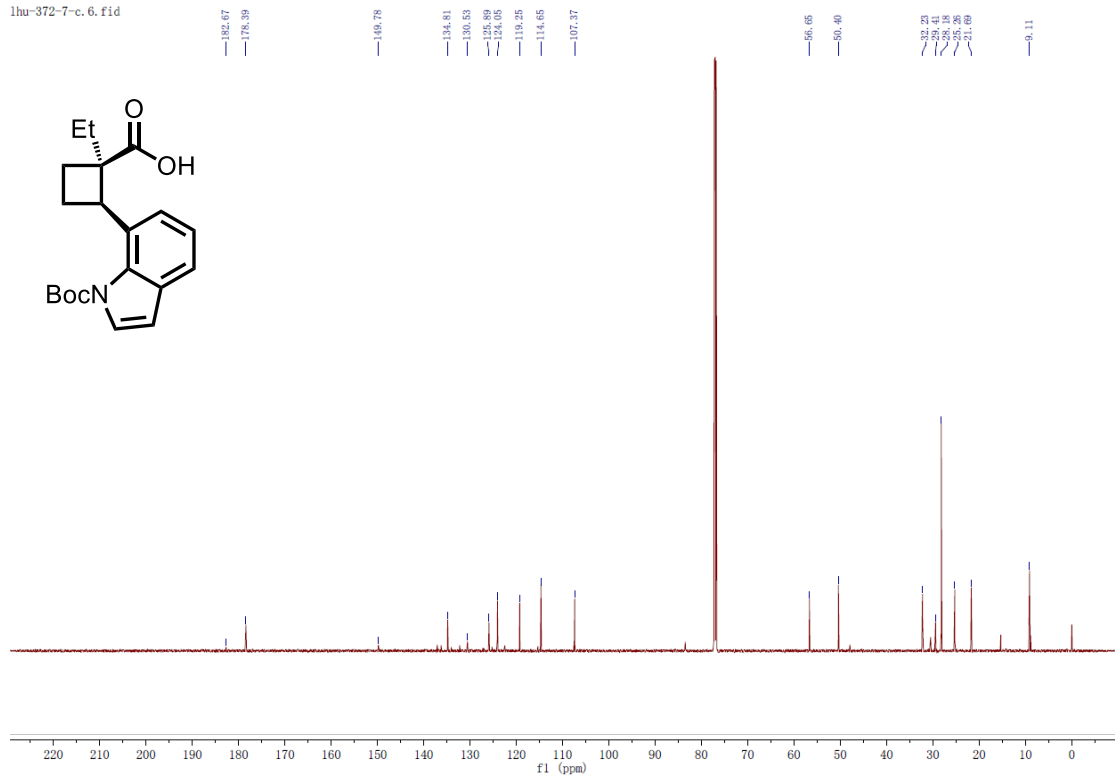
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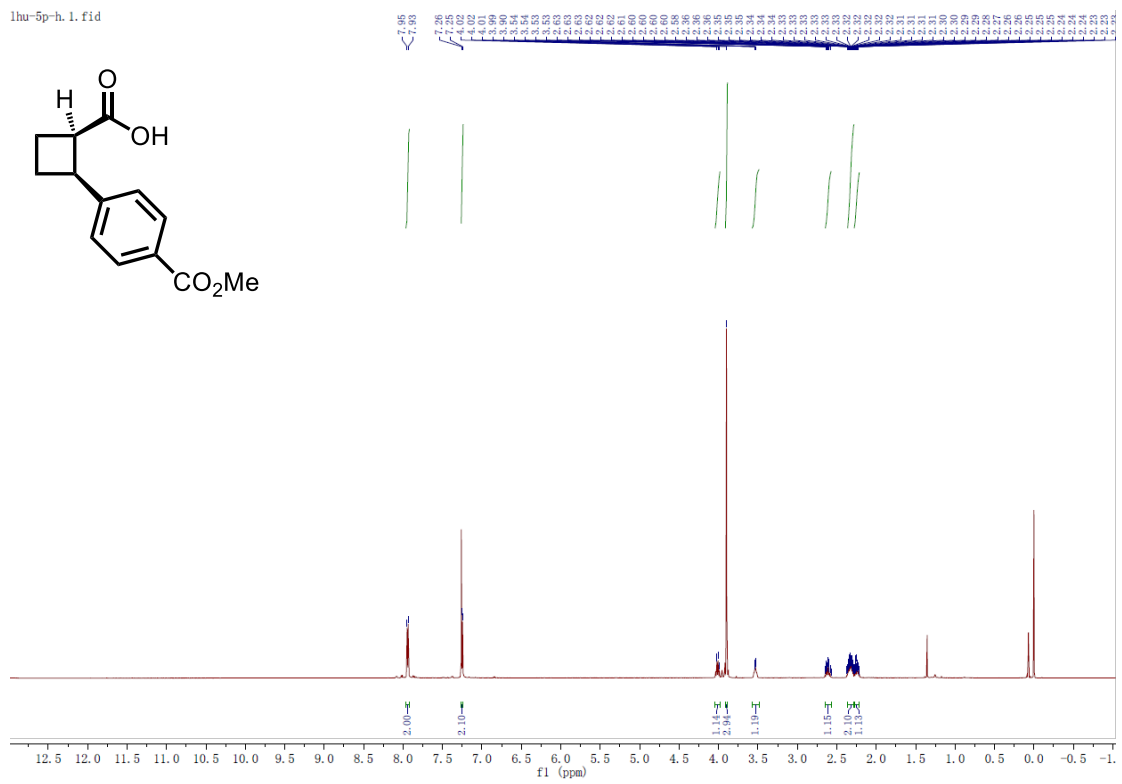
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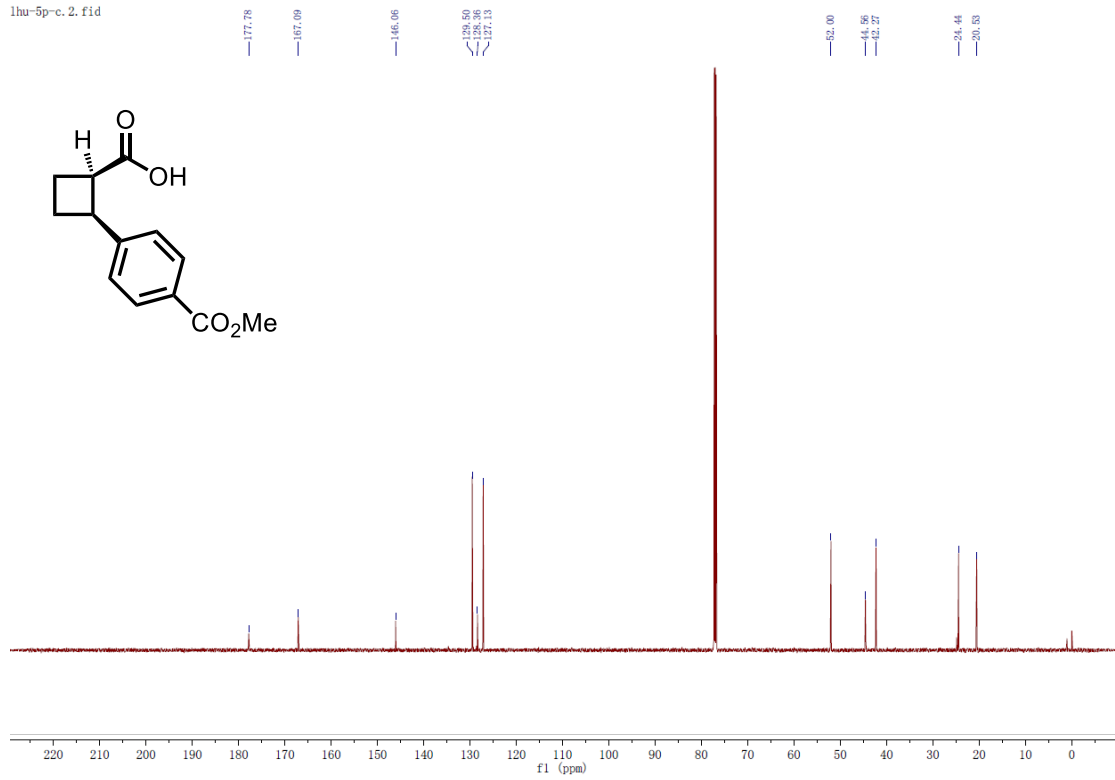
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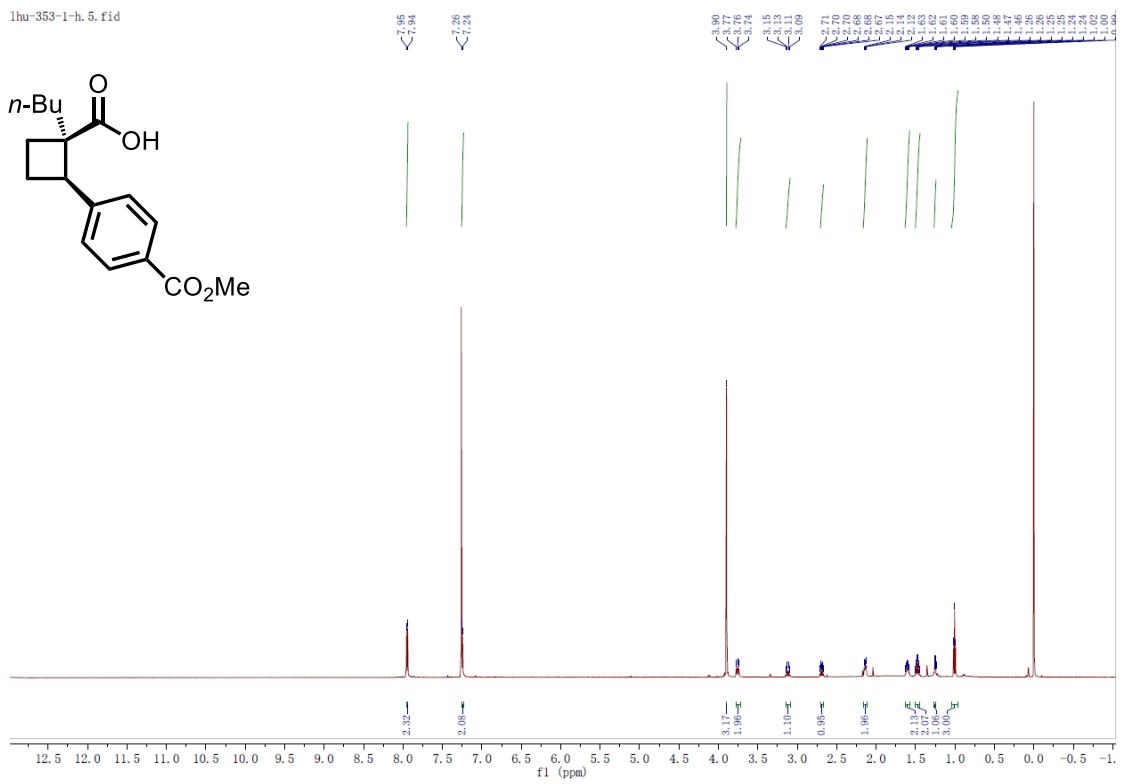
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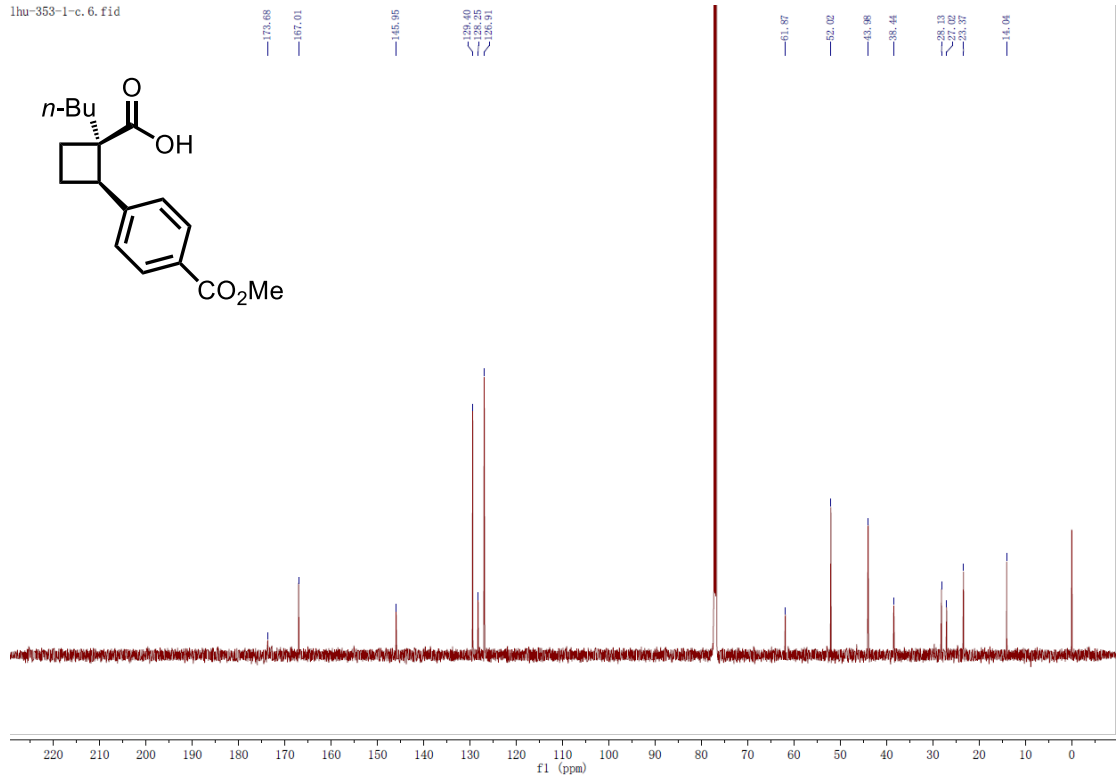
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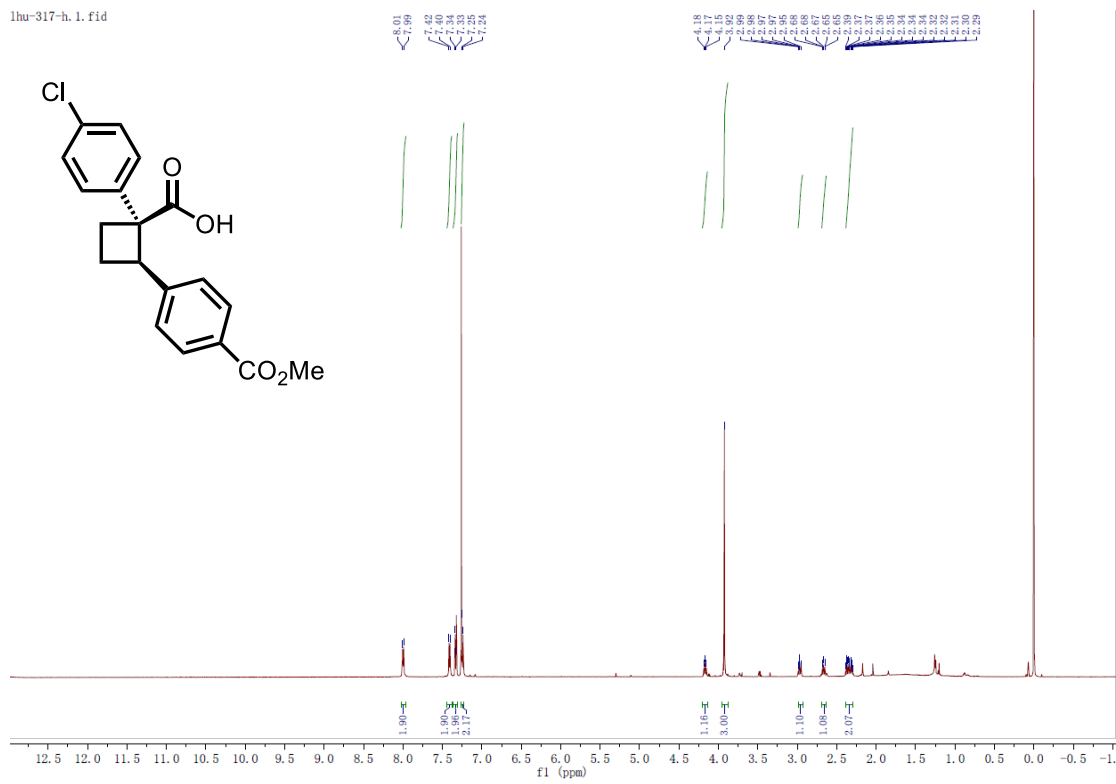
lhu-353-1-h. 5. fid



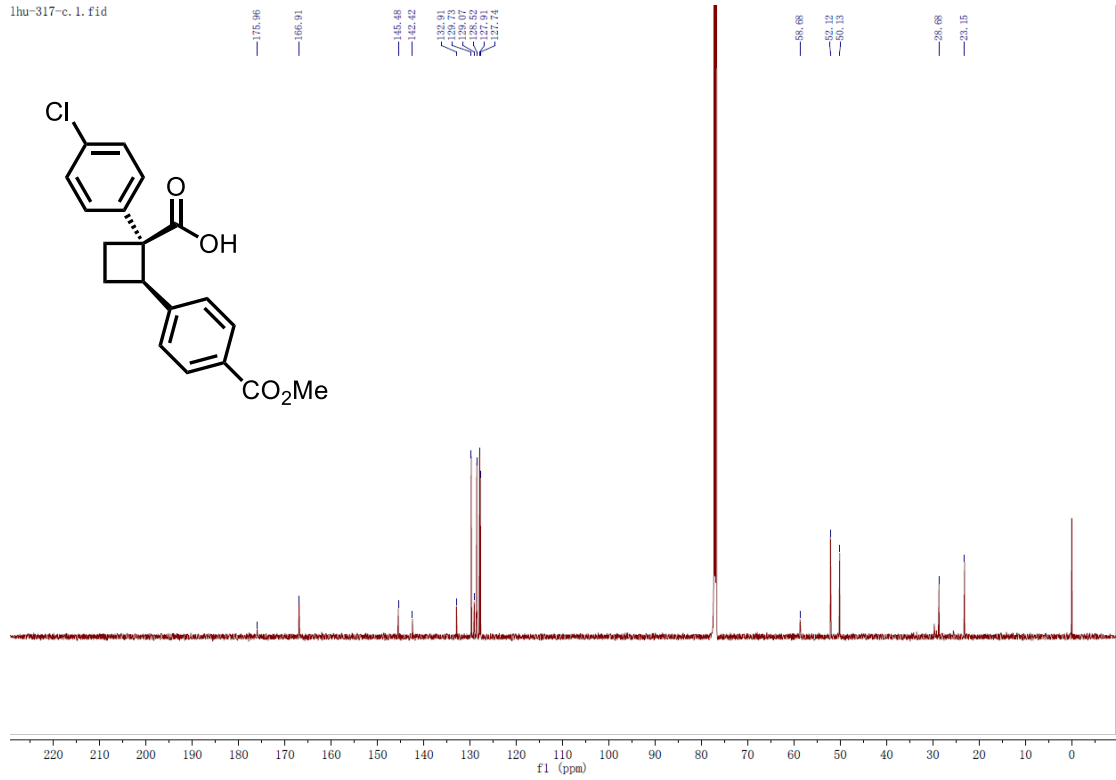
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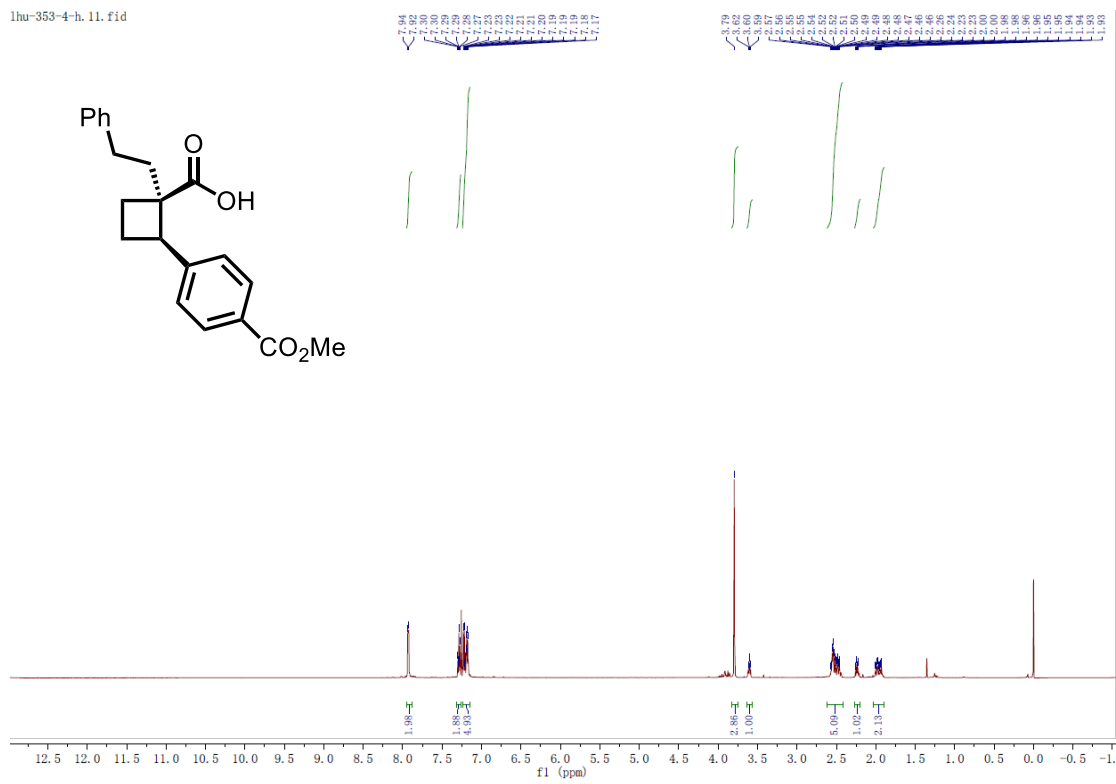
lhu-317-h. 1. fid



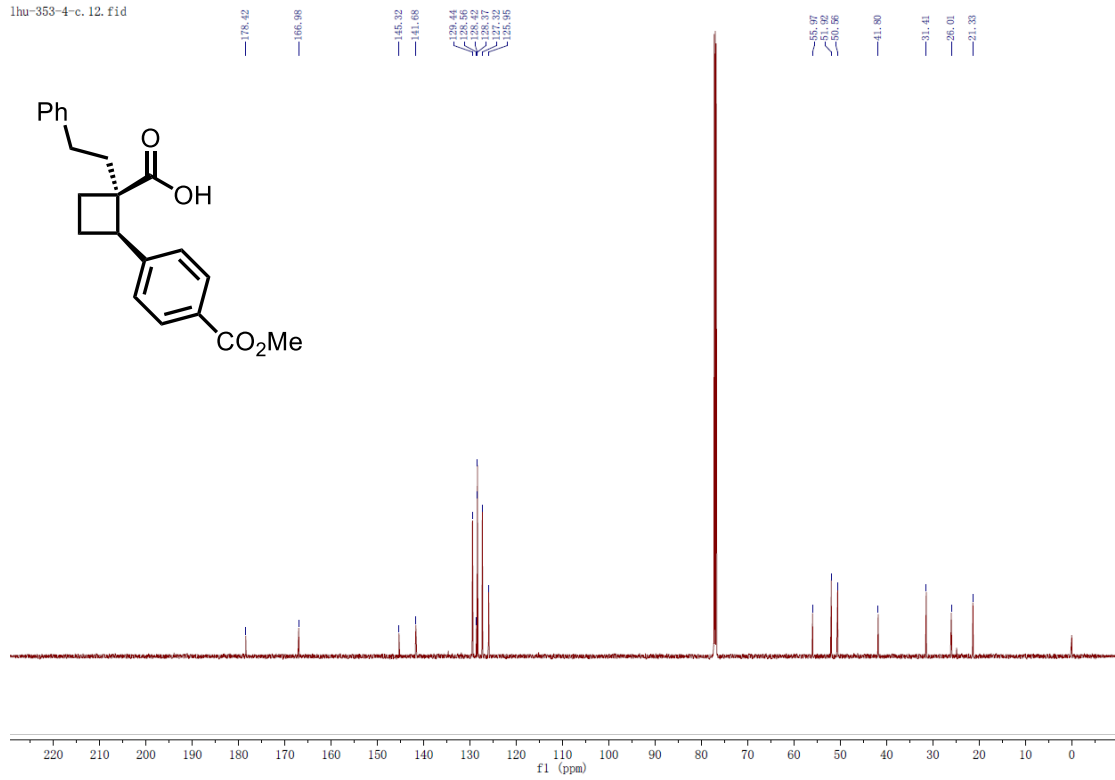
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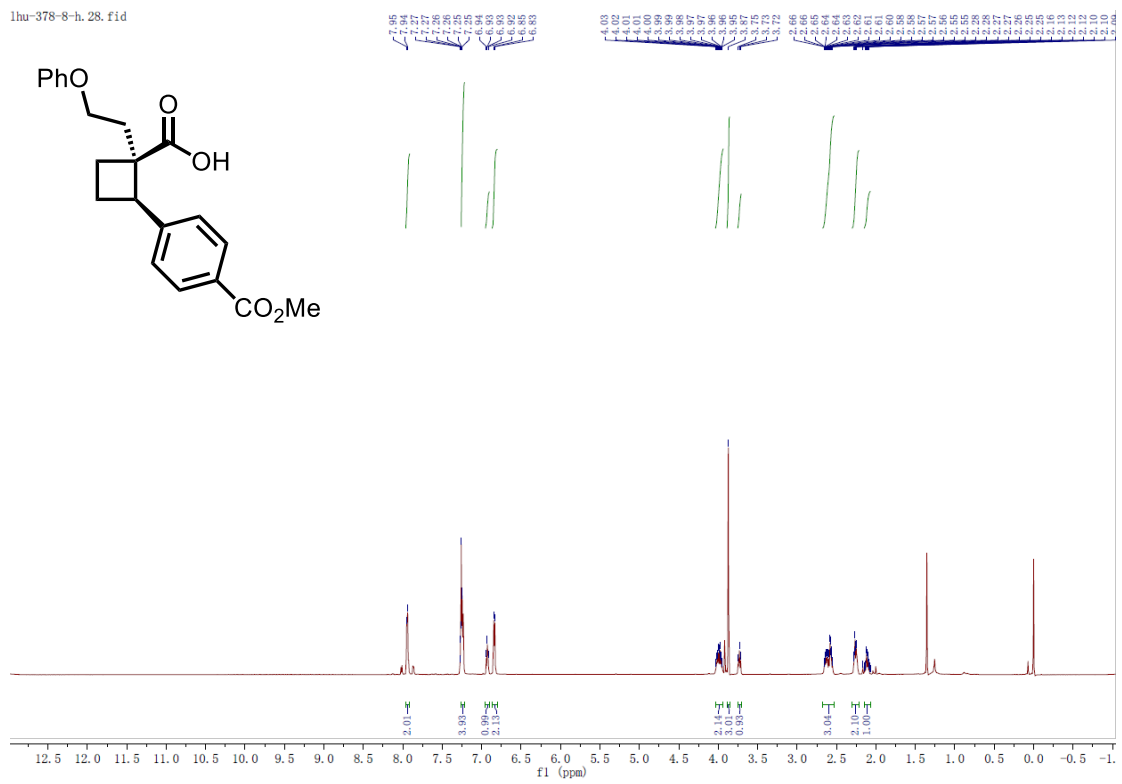
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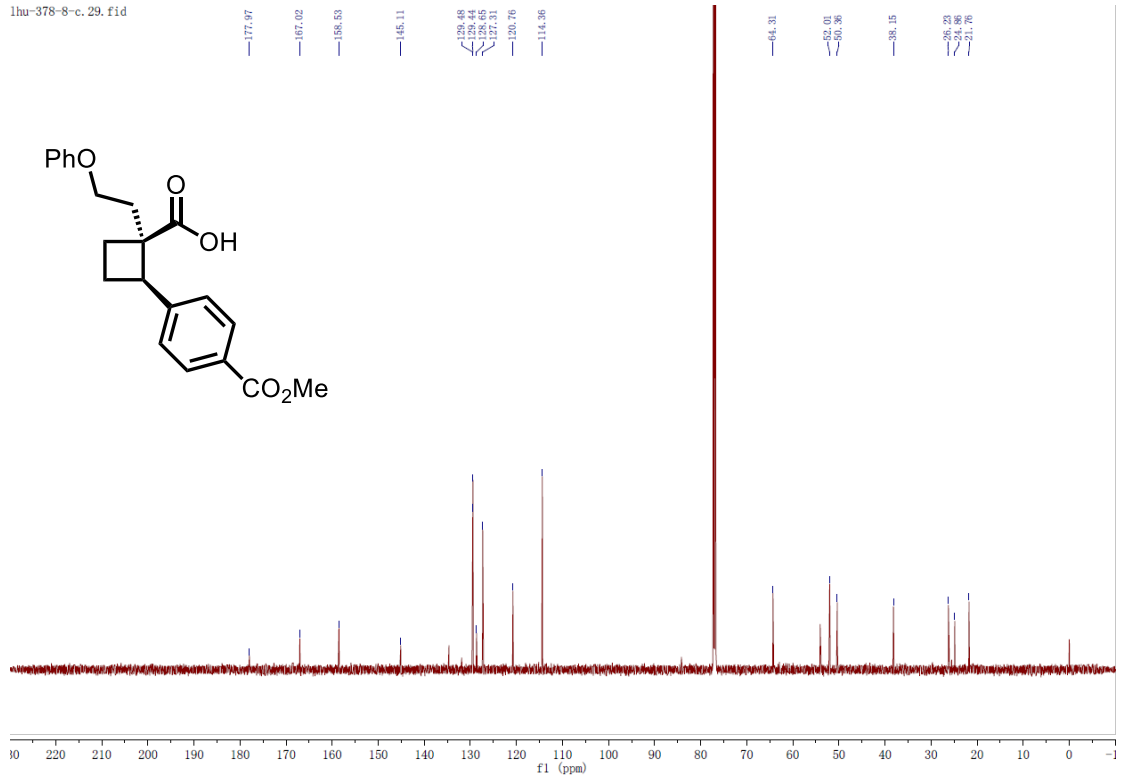
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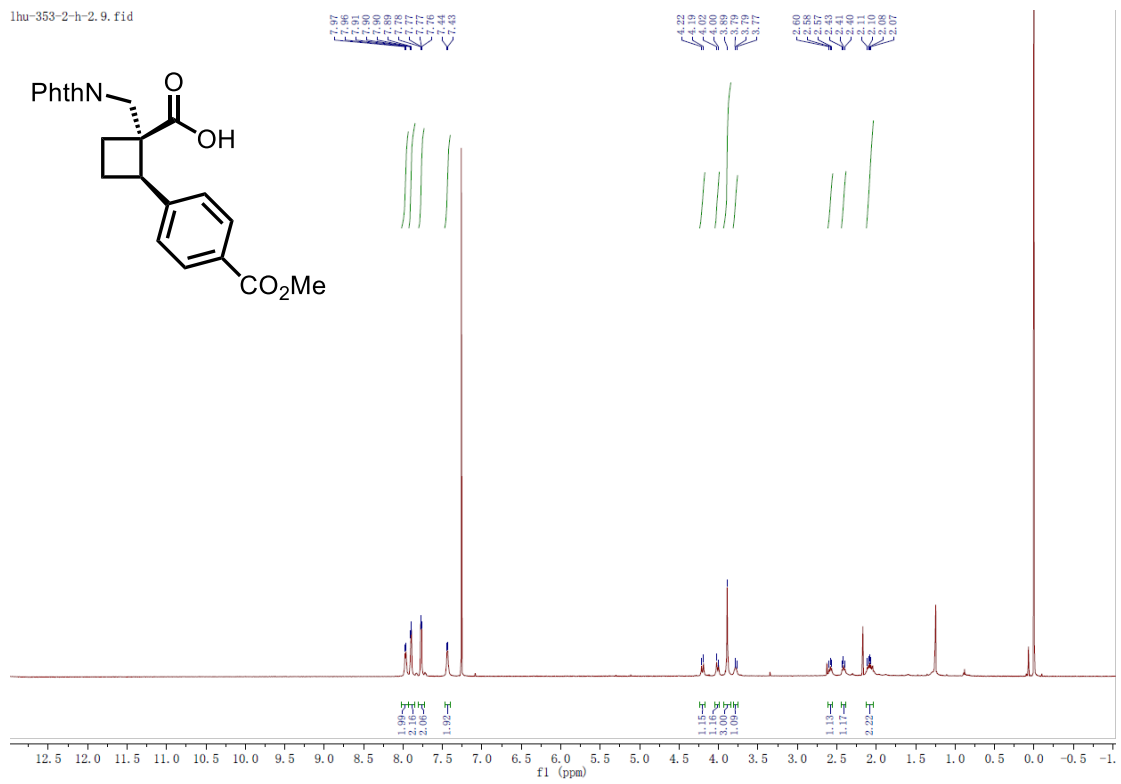
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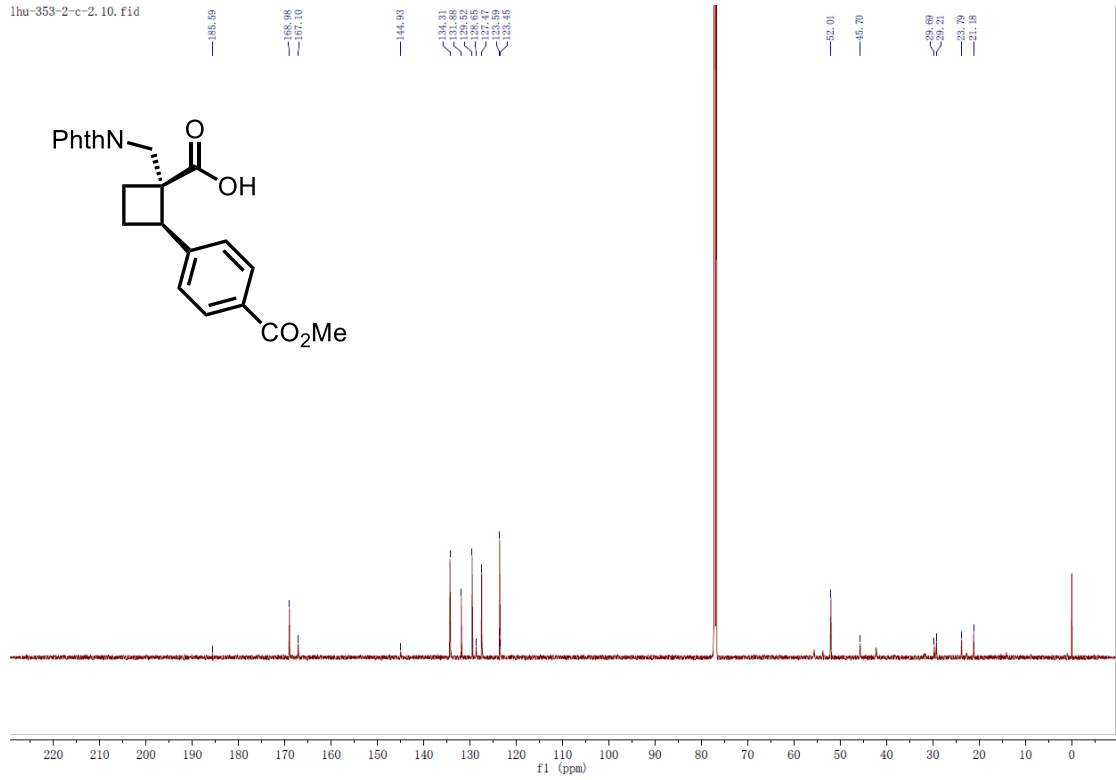
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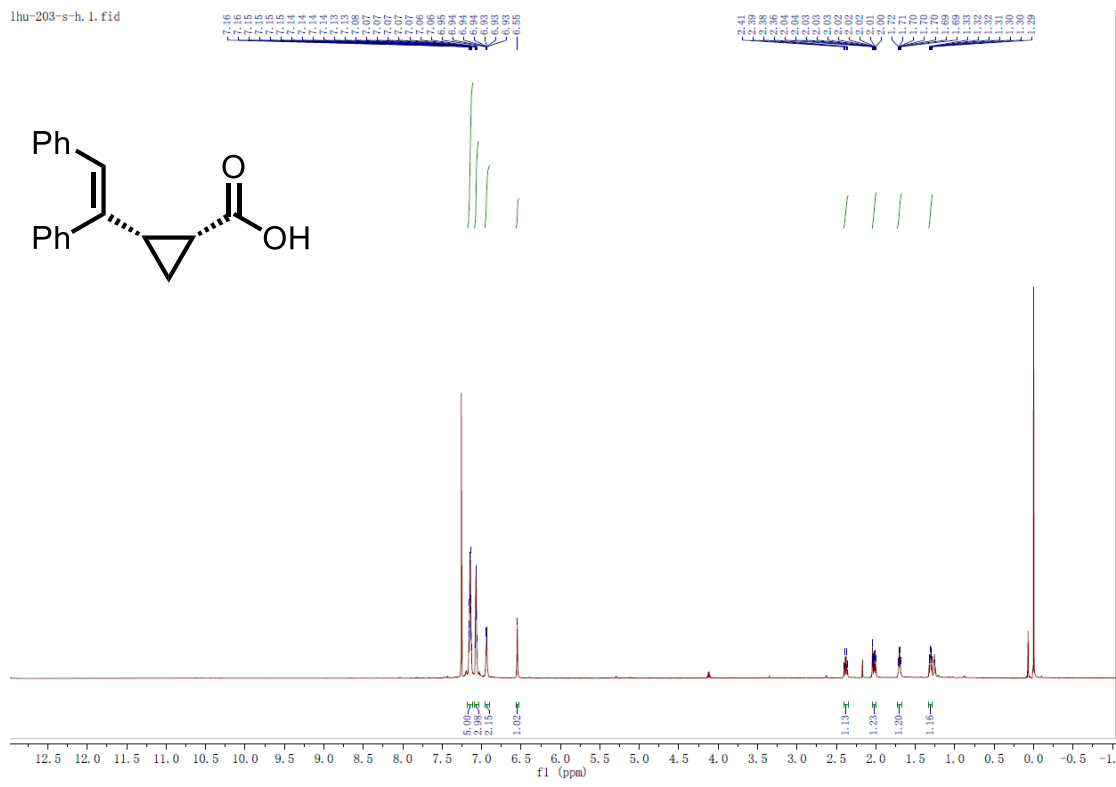
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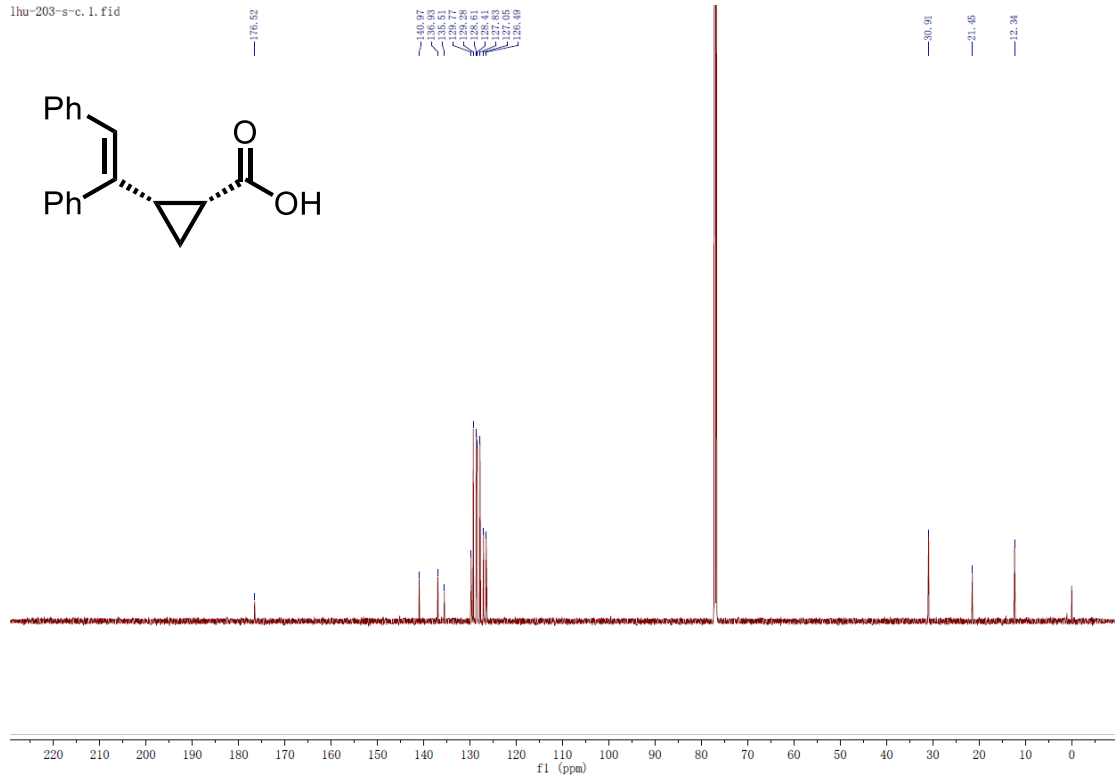
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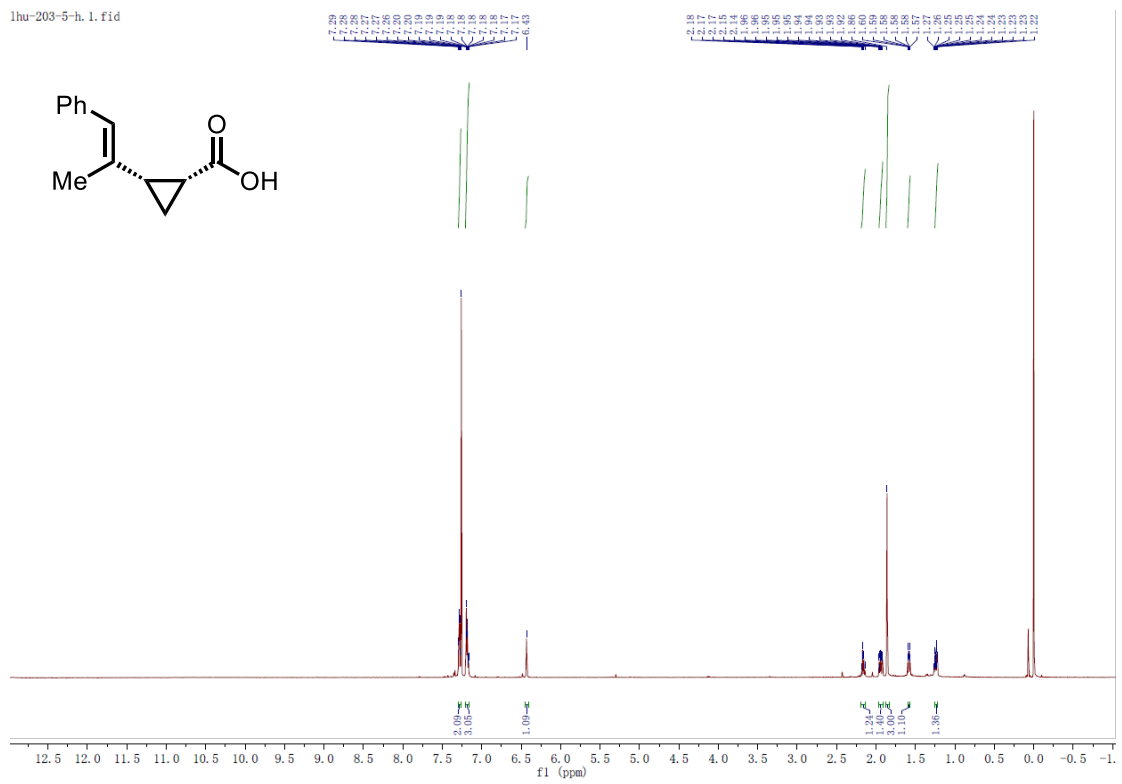
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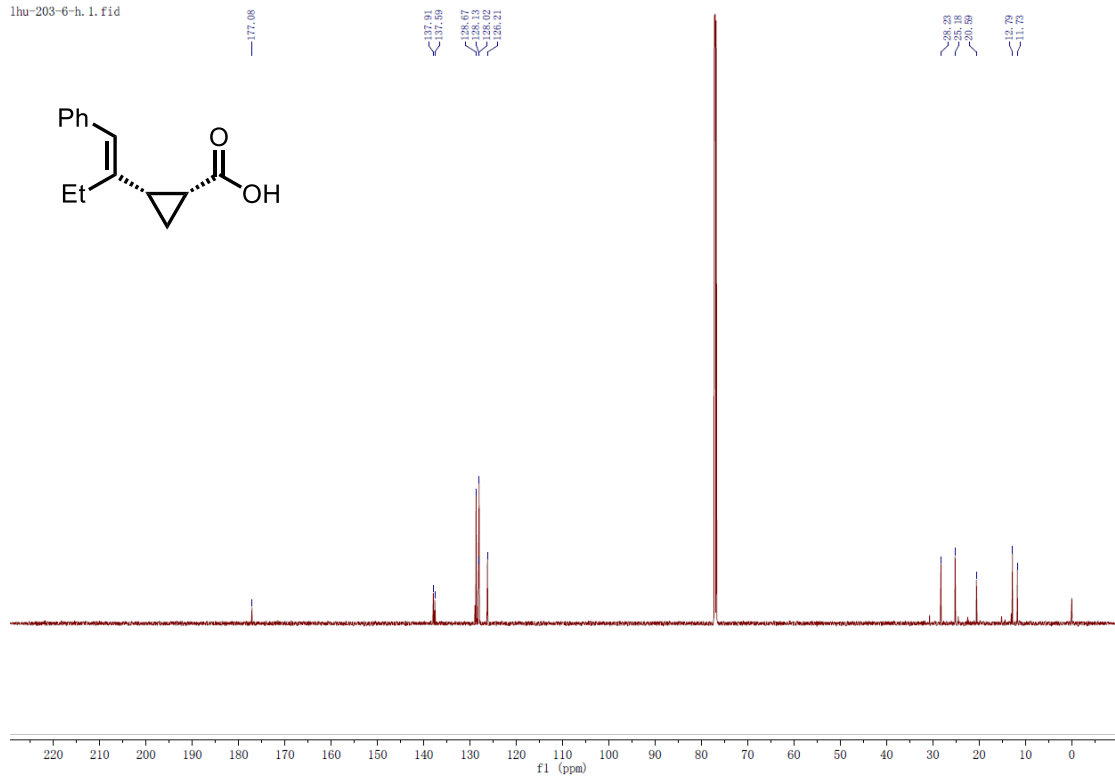
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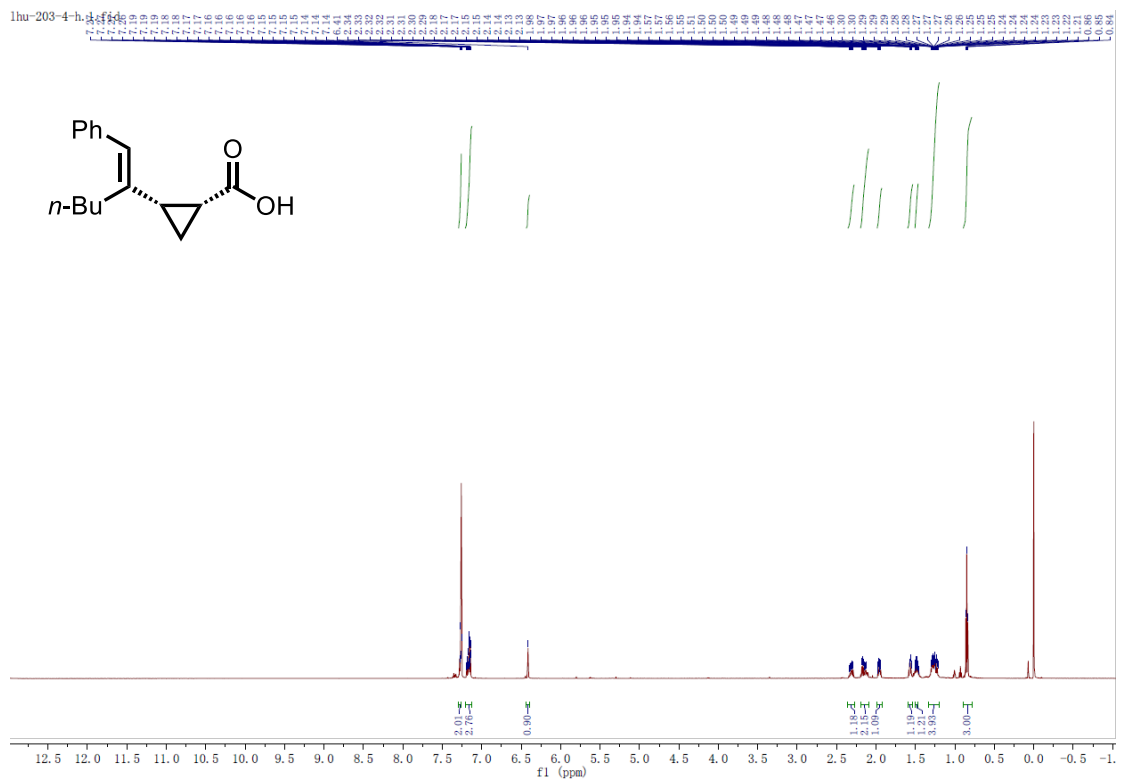
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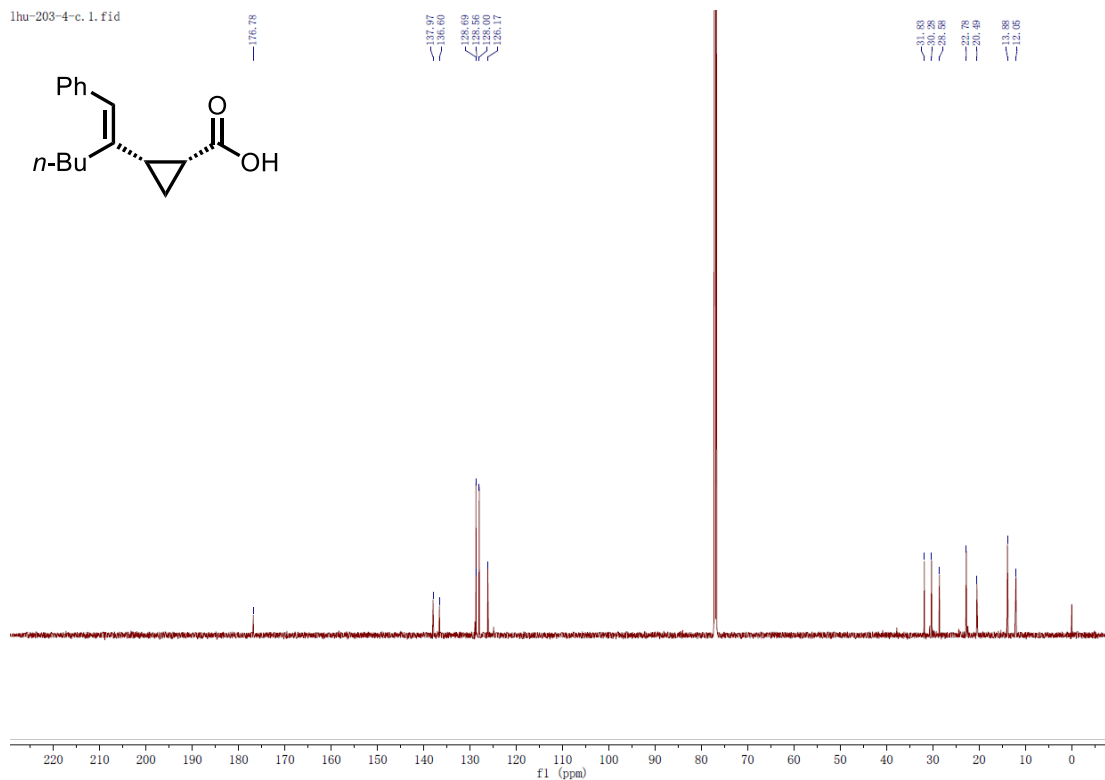
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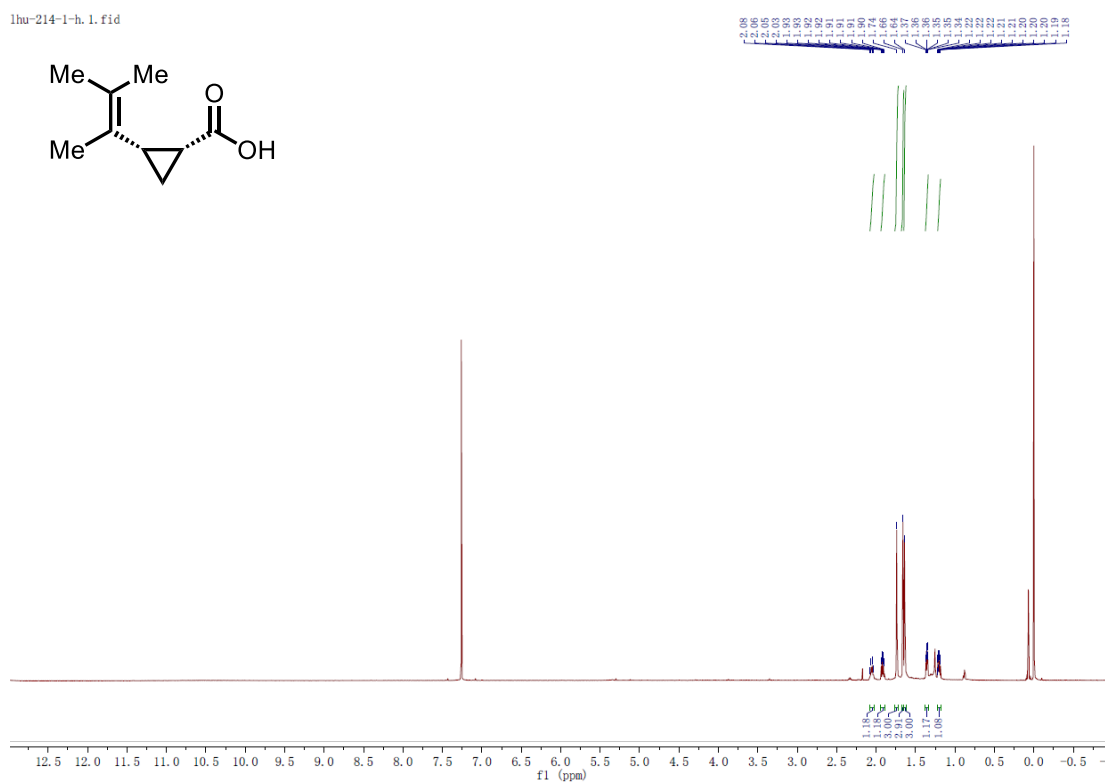
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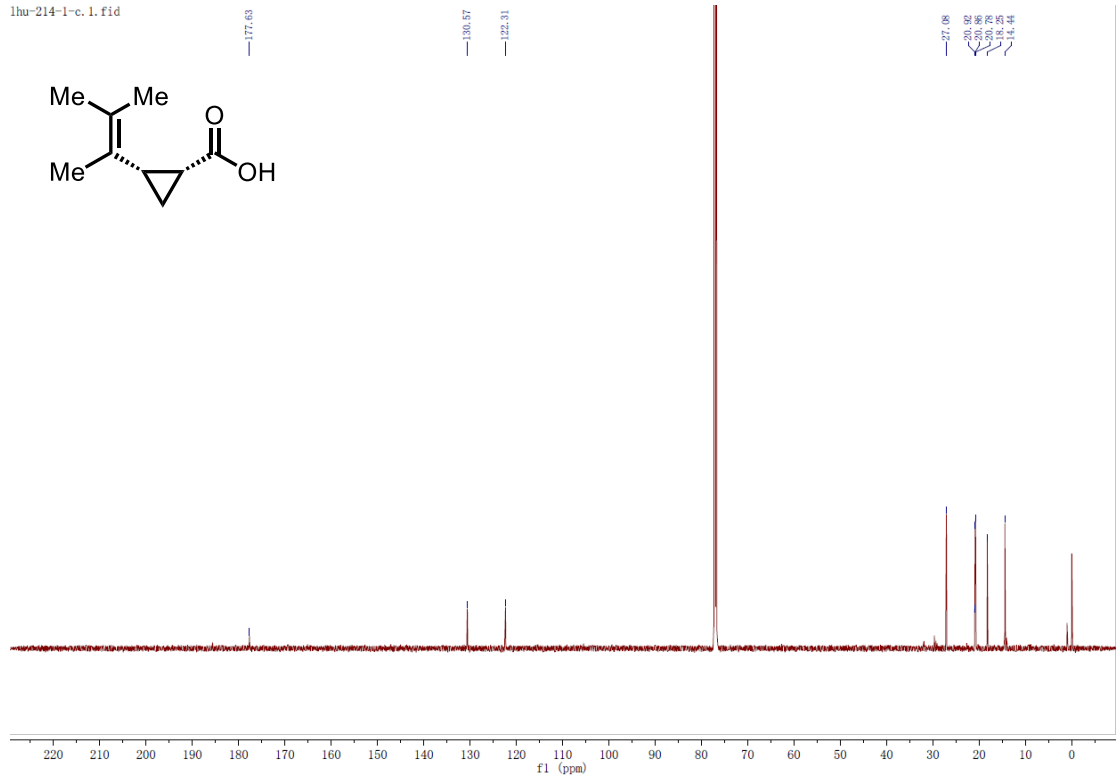
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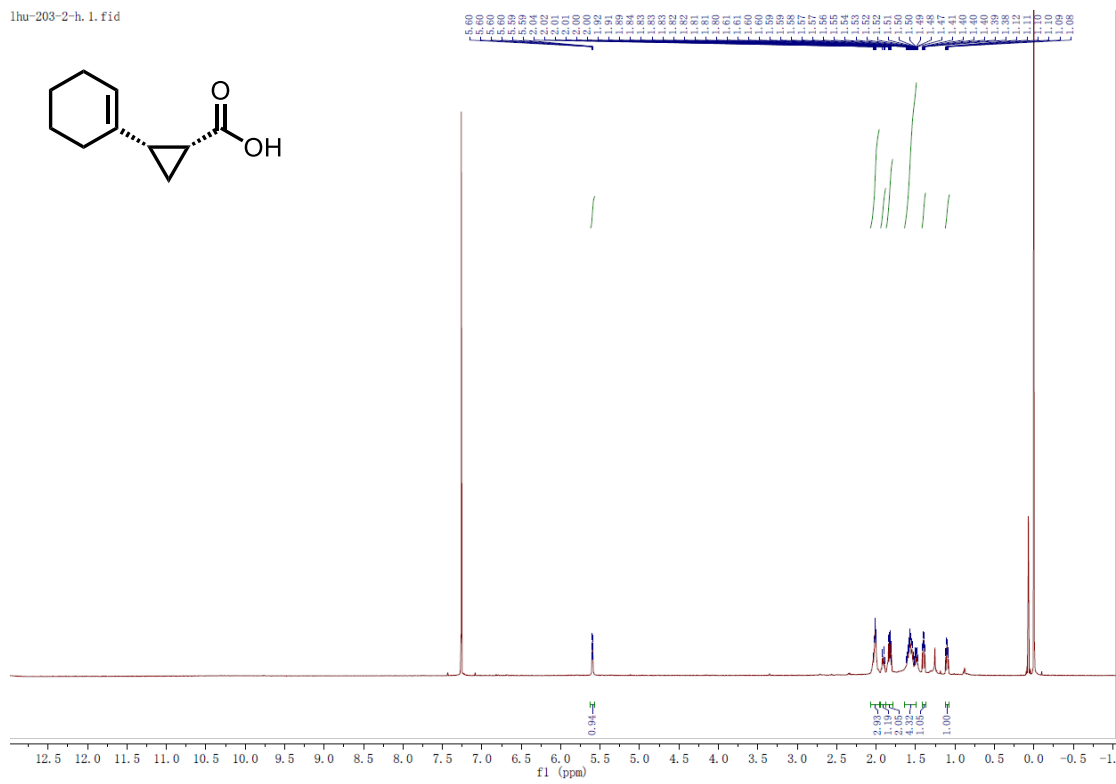
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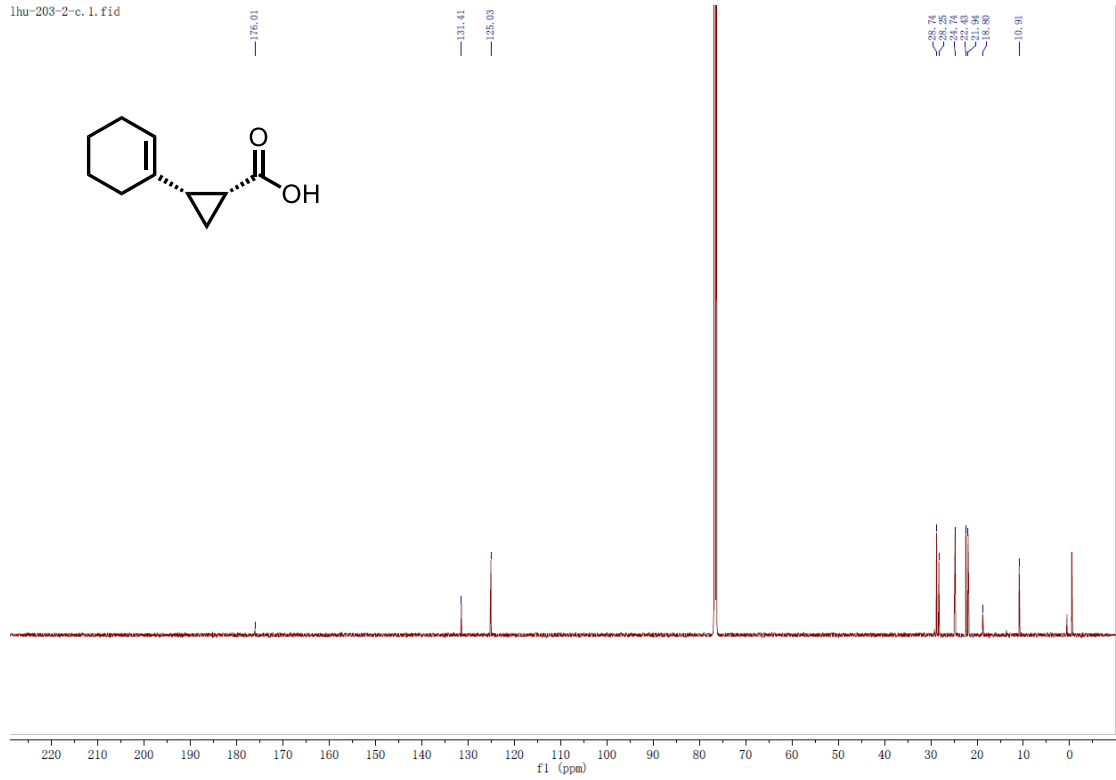
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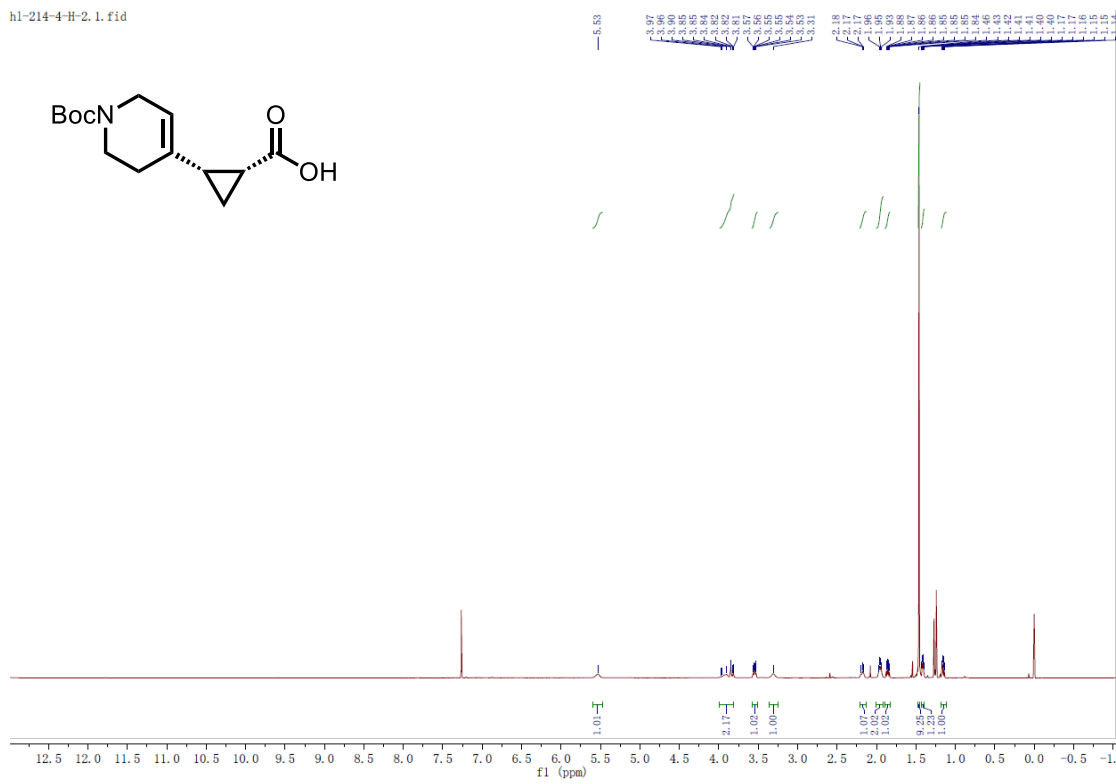
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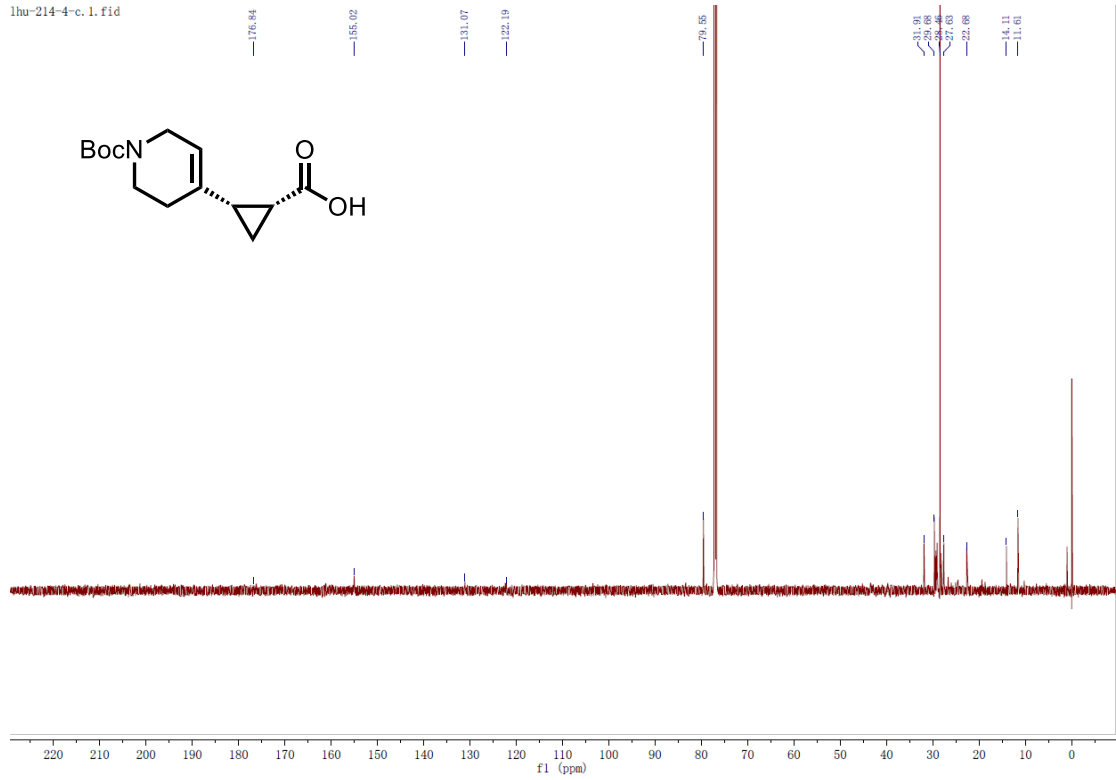
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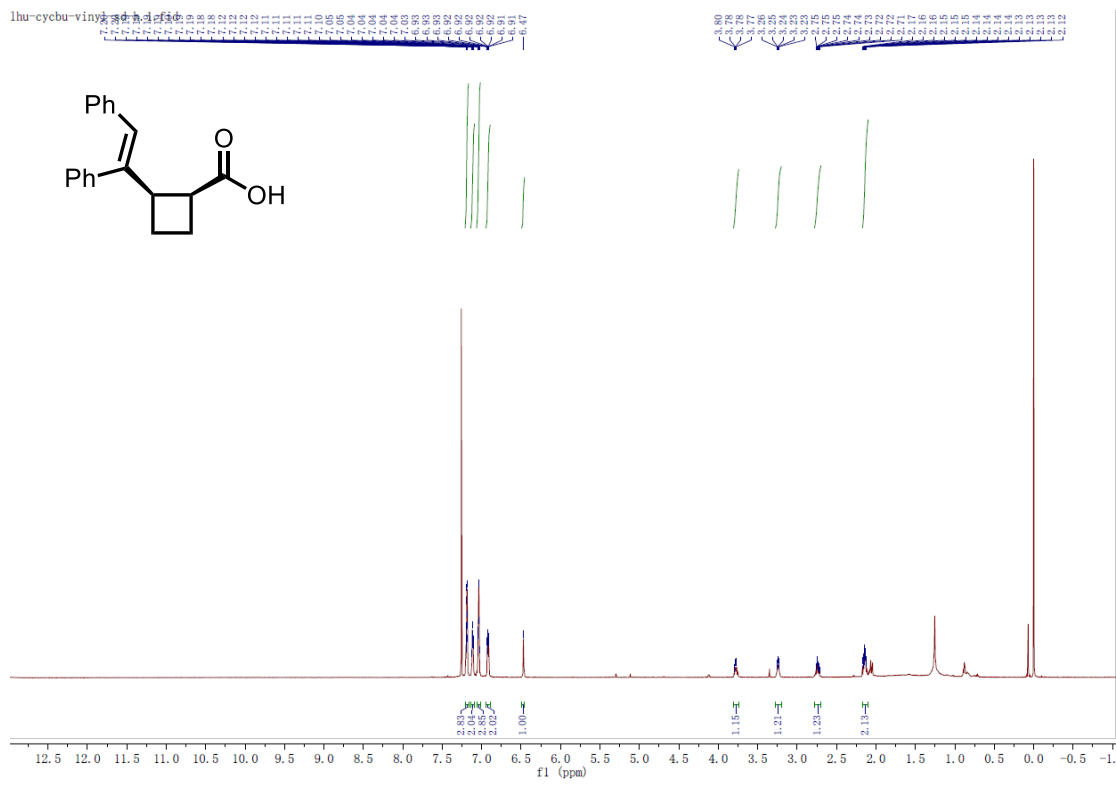
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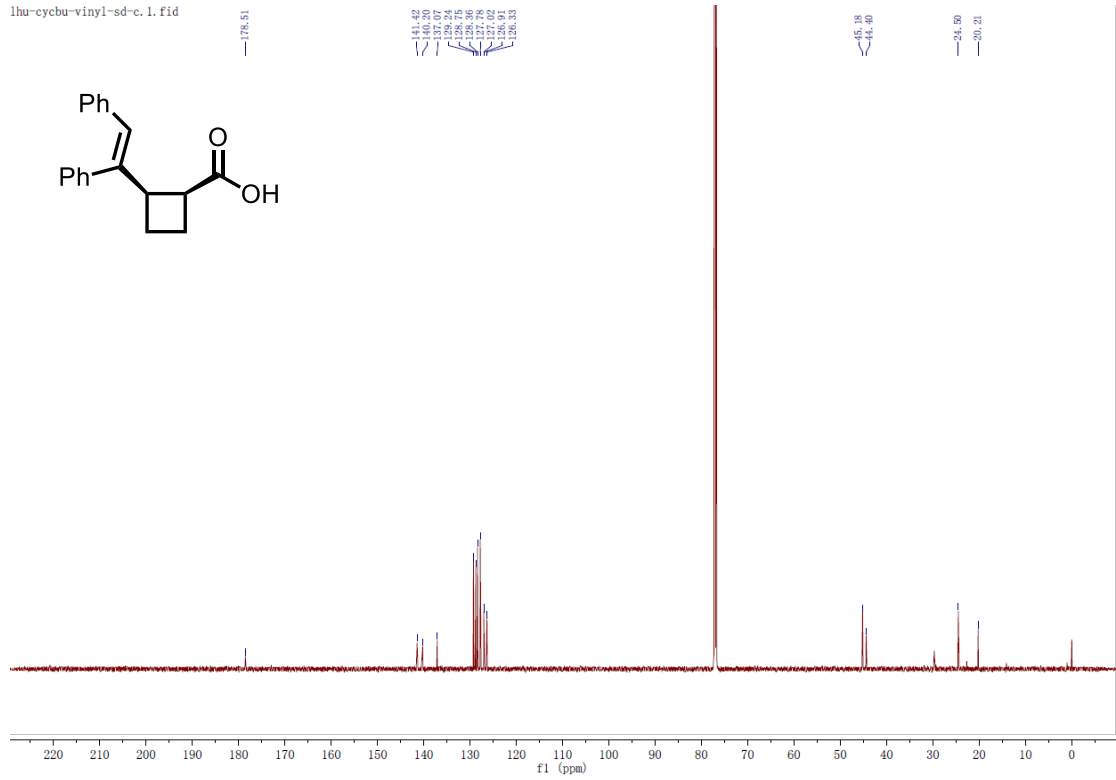
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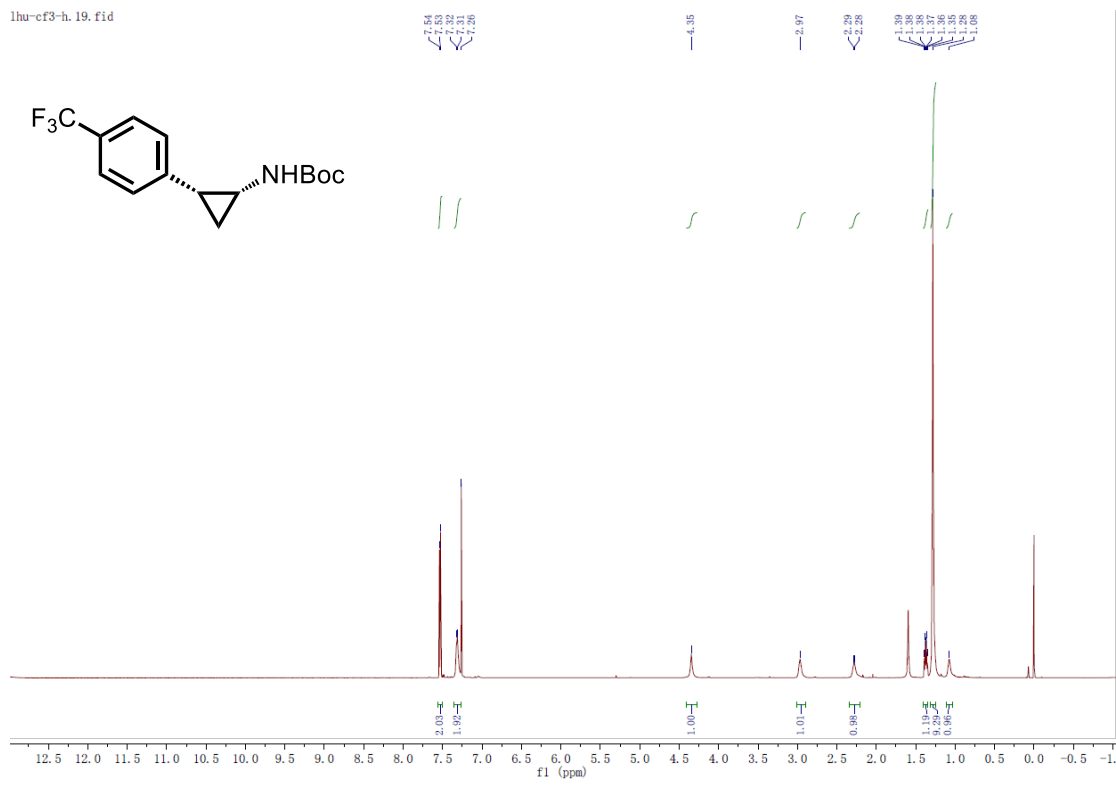
lhu-cycbu-viny...



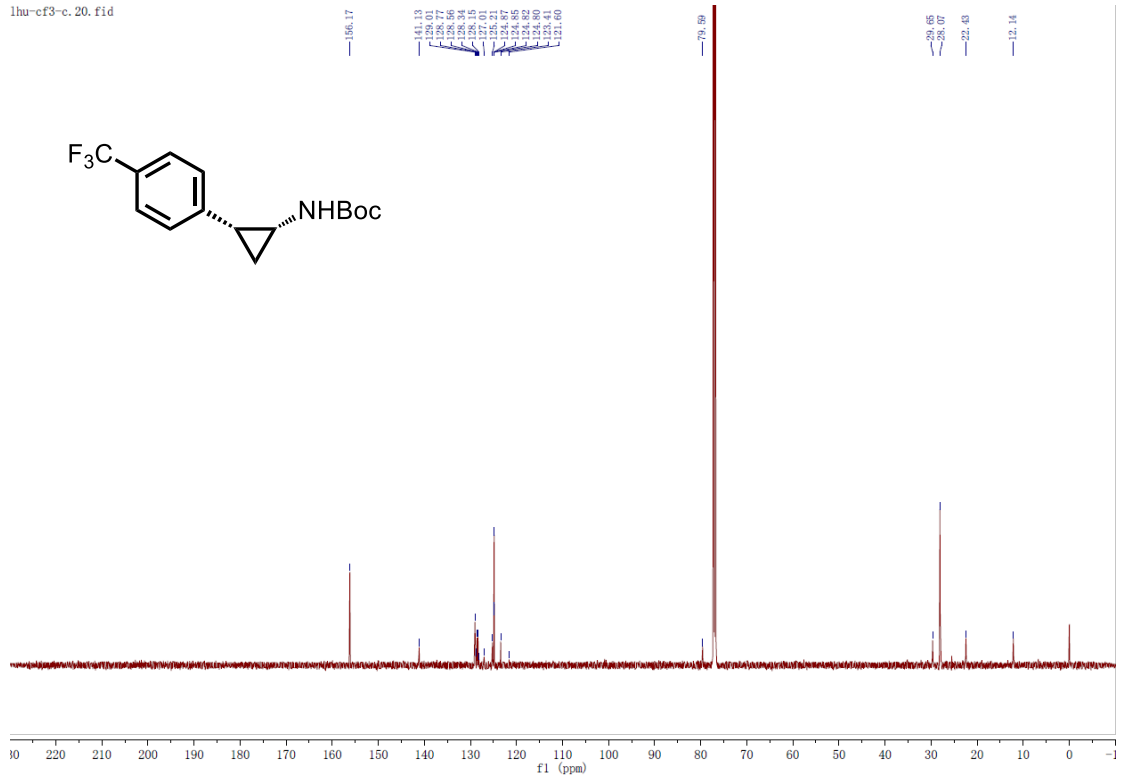
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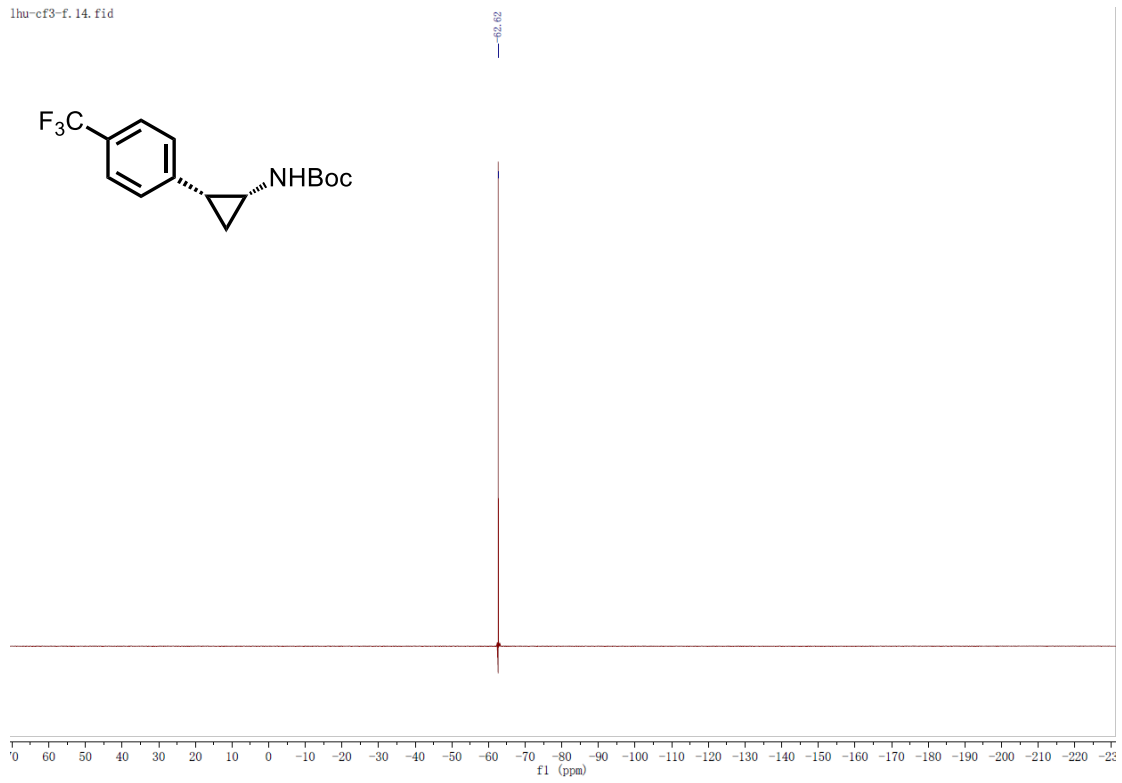
lhu-cf3-h. 19. fid



lhu-cf3-c. 20. fid



lhu-cf3-f. 14. fid



lhu-cf3-h. 3. ser

