Supplemental Material

Primer	Forward	Reverse						
Human-GAPDH	CGTATTGGGCGCCTGGTCAC	ATGATGACCCTTTTGGCTCC						
Human-cyclin A2	CGCTGGCGGTACTGAAGTC	GAGGAACGGTGACATGCTCA T						
Human-KLF4	TCTCAAGGCACACCTGCGAA	TAGTGCCTGGTCAGTTCATC						
Human-p21	TGTCCGTCAGAACCCATGC	AAAGTCGAAGTTCCATCGCTC						
Human-p27	TAATTGGGGCTCCGGCTAACT	TGCAGGTCGCTTCCTTATTCC						
Human-F2RL1	TTGTGTTTGTGGTGGGTTTGC	ACCAGATGACAGAGAGGAGG T						
Human-PDK2	CTTCAGCAAGTTCTCCCCGTC	TCGGGAAGCAGGTTGATCTC T						
Human-TXNIP	GCCAGCCAACTCAAGAGACAA	TTCGAGCAGAGACAGACACC C						
Human-BRMS1	GCCTCCAAGCAAAGACACAGA	AGCTCTCCTCTTCTGACTCTG T						
Human-PDK1	AGAGGGTTACGGGACAGATGC	GTCTTTGGGTTCTCTGCTGG G						
Human-CD44	ATGGGTTCATAGAAGGGCACG	TGTCATACTGGGAGGTGTTG GA						
Human-RhoA	TTGTTGGTGATGGAGCCTGTG	GCCAACTCTACCTGCTTTCCA						
Human-GAB1	CAGACAGCGAGGAGGAGATG A	GGTGGATGGTGGTGCTGTAT T						
Human-PTEN	CTCAGCCGTTACCTGTGTGTG	AGGTTTCCTCTGGTCCTGGTA						
Human-AKT2	AGGCACGGGCTAAAGTGAC	CTGTGTGAGCGACTTCATCCT						
Human-AKT3	GGAGAGGAAGAGATGGATGC CT	CCACTTGCCTTCTCTCGAACC						
Human-mTOR	CCCTCCATCCACCTCATCAGT	CGCCAAGACACAGTAGCGAA T						
Human-PIK3CG	AACACCGACCTCACAGTTTTT	CTCAAGCCACACATTCCACAG						
Human-cyclin B1	GGTTGTTGCAGGAGACCATGT	AACATGGCAGTGACACCAAC C						
Human-cyclin D1	TTCATTTCCAATCCGCCCTCC	TGTGAGGCGGTAGTAGGACA G						
Human-FGF2	AGAAGAGCGACCCTCACATCA	CGGTTAGCACACACTCCTTTG						
Human-MMP1	AAAATTACACGCCAGATTTGC C	GGTGTGACATTACTCCAGAGT TG						
Human-MMP9	AGACCTGGGCAGATTCCAAAC	CGGCAAGTCTTCCGAGTAGT						
Human-VEGF-A	CCTCCGAAACCATGAACTTT	CCACTTCGTGATGATTCTGC						
Human-VEGF-B	GATGGCCTGGAGTGTGTG	CACACTGGCTGTGTTCTTCC						
Human-VE- cadherin	TGTTCACGCATCGGTTGTTCA	TACATGACAGAGGCGTGGTC T						

Table S1. Primer sequences

Human- VEGF- C	GGCTGGCAACATAACAGAGA	GTGGCATGCATTGAGTCTTT
Human- FAK	GTTATCCCAGTCCGAGGTCCA	TGACCTGGATAGATGCTGCC A
Mouse- Gapdh	AGTGTTTCCTCGTCCCGTAGA	TGAGGTCAATGAAGGGGTCG T
Mouse- F2rl1	AGTTCTCTGCGTCCATCCTCA	GCGGGGTGTTTCTTCTTCGTT
Mouse- Pik3cg	TGCGGAGTTCTACCACCGAT	TAATGCAGGCAGTCTAGGGT C
Mouse-p21	TCTTCCCCATCTTCGGCCTTA	AGTTTGGAGACTGGGAGAGG G

Table S2.	Antibodies,	source	and	dilution
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Antibody	Source	Dilution(application)
AKT1/2/3	Abcam#ab131443	1:10000(WB)
		1:100(IF)
p27	Abcam#ab32034	1:1000 (WB)
		1:100 (IF)
MMP1	Abcam#ab38929	1:1000(IF))
mTOR	Abcam#ab2732	1:2000(WB)
		1:100(IF)
PTEN	Abcam#ab32199	1:10000(IF))
cyclinA2	Abcam#32386	1:10000 (WB)
		1:200(IF)
p21	Abcam#109520	1:2000 (WB)
		1:1000(IF)
F2RL1	Abcam#180953	1:2000 (WB)
		1:100(IF)
cyclinB1	Abcam#181593	1:2000 (WB)
		1:500(IF)
PDK2	Novusbio#NBPI-87307	1:200(WB)
		1:100(IF)
KLF4	Abcam#215036	1:10000(WB)
VEGF-A	Abcam#46154	1:100(WB)
cyclinD1	Abcam#134175	1:10000(WB)
		1:50(IF)
FGF2	Abcam#8880	1:1000(WB)
		1:200(IF)
TXNIP	Abcam#188865	1:1000(WB)
BRMS1	Abcam#57082	1:100(WB)
β-actin	SIGMA#A5441	1:3000(WB)
MMP9	Abcam#38929	1:250(IF)

VE-cadherin	Abcam#33168	1:100(IF)
CD34	Abcam#8158	1:50(IF)
PCNA	Cell Signaling#D3H8P	1:800(IF)
Alexa Fluor 594 donkey	Molecular	1:1000(IF)
anti-rabbit IgG	Probes#A21207	

	Ctrl-T			ТСо			TCo+VO			Ctrl-C			CCo			CCo+VO		
cyclinA2	7.60	7.34	7.37	7.41	7.64	7.63	7.11	7.01	7.06	8.07	7.70	8.68	7.68	7.30	8.16	7.60	7.34	7.37
cyclinB1	5.97	5.74	6.01	6.63	6.42	6.97	6.19	5.72	5.59	6.10	6.29	6.02	5.80	5.84	5.95	5.86	5.98	5.69
cyclinD1	6.01	5.77	4.87	7.74	7.12	6.41	6.14	5.86	5.89	5.97	7.08	6.40	5.00	5.90	5.38	4.92	6.35	5.83
FGF2	9.68	9.02	9.68	10.07	9.20	9.92	10.01	9.01	9.41	10.46	9.34	10.23	9.93	9.08	10.16	9.68	9.02	9.59
MMP1	14.97	14.97	14.97	15.43	15.20	15.89	14.54	14.30	14.49	16.01	16.08	15.83	15.98	15.74	15.38	14.97	15.44	14.97
MMP9	13.60	14.93	14.87	14.92	16.00	15.80	14.30	16.12	16.32	15.99	14.90	16.23	16.57	15.17	16.31	14.93	13.60	14.25
VEGF-A	8.70	10.90	8.35	9.23	11.40	8.95	8.25	10.32	8.23	9.43	8.49	10.49	7.96	5.99	8.20	8.08	6.55	8.35
VEGF-B	0.18	7.19	7.64	0.76	8.05	8.44	-0.05	7.04	7.16	-0.23	7.43	8.52	-0.78	7.05	8.04	0.18	8.03	9.24
VE-cadherin	8.60	9.81	9.46	9.39	10.15	10.45	9.13	10.02	10.28	11.28	10.29	10.64	10.59	10.09	10.16	9.81	9.00	8.60
p21	5.18	5.03	4.96	4.85	5.01	4.27	5.21	6.09	4.85	5.04	4.27	4.62	5.20	4.70	5.07	5.18	5.03	4.96
p27	8.28	8.22	8.24	7.50	7.76	7.74	7.66	7.83	8.04	7.76	7.77	7.62	7.99	7.70	7.78	8.28	8.22	8.24
PIK3CG	10.62	12.21	11.25	11.77	13.18	11.99	11.36	13.04	11.66	10.99	9.87	9.95	11.52	10.61	11.04	11.77	11.17	10.92
PDK1	9.33	11.27	8.87	9.80	12.04	9.05	9.49	11.15	8.43	8.87	9.44	9.78	9.17	9.89	10.27	8.98	9.50	9.83
PDK2	9.59	8.98	10.01	10.05	9.67	10.45	9.78	9.23	9.90	9.59	8.98	9.58	10.82	10.09	10.47	9.97	9.41	10.11
AKT2	19.53	20.88	19.79	20.02	21.09	20.50	18.87	20.08	18.93	18.87	18.28	19.79	19.75	20.30	20.97	19.33	18.89	20.34
AKT3	9.16	9.02	9.50	9.54	9.54	9.68	8.84	8.82	9.54	9.16	8.27	9.76	9.75	8.92	10.13	9.05	8.53	9.65
mTOR	8.77	8.15	8.95	9.84	9.66	9.39	9.01	8.34	8.67	8.95	9.27	8.86	9.32	10.12	9.24	8.79	9.16	8.93

Table S3. Delta CT means of genes in C918 cells with VO

Table S4. Delta CT means of genes in C918 cells

		Ctrl			TCo		CCo					
CD44	-0.75	3.74	4.03	0.32	5.38	5.71	-0.38	4.02	4.47			
RhoA	4.62	4.28	4.45	4.52	3.96	3.97	5.16	4.62	5.16			
GAB1	10.92	11.01	11.38	10.23	10.18	10.92	12.16	11.81	12.49			
PTEN	8.07	0.79	1.47	7.09	-0.10	0.93	7.73	0.43	1.30			

		RPE-Ctr	l		RPE-ESC	2		C918-Ctr	1	C918-ESC				
PAR2	11.32	13.90	5.57	9.94	11.43	3.55	15.06	12.64	9.63	12.70	10.01	7.66		
PIK3CG	13.05	13.05	4.56	12.00	12.35	3.98	6.53	10.94	6.53	7.37	11.52	6.86		
PDK1	10.20	8.63	8.99	8.88	7.09	8.43	7.93	7.93	4.07	8.62	9.04	8.16		
PDK2	8.84	2.55	7.87	8.00	1.92	7.48	9.59	9.59	8.08	9.64	9.77	9.02		
AKT2	19.89	20.73	13.52	19.55	20.16	12.56	14.34	14.54	20.03	14.54	14.70	20.27		
АКТЗ	8.67	8.67	2.27	7.99	8.26	1.81	2.07	9.02	7.50	2.51	9.44	8.75		
mTOR	2.96	2.54	2.54	2.75	2.02	2.33	8.66	8.82	8.93	9.74	9.47	9.33		
PTEN	6.13	5.25	6.95	6.47	6.01	7.23	7.96	7.96	7.75	7.45	7.75	7.65		
cyclin A2	8.01	8.49	9.01	7.84	7.25	7.33	6.94	8.02	6.78	7.31	8.79	8.04		
cyclin B1	6.82	5.97	7.00	6.70	5.98	5.42	4.02	3.79	4.99	4.71	4.97	5.94		
cyclin D1	5.29	5.08	5.45	4.95	3.95	4.18	4.60	3.33	4.84	5.97	4.94	6.38		
p21	4.41	6.00	5.27	5.99	6.61	6.11	5.98	6.43	5.98	5.32	5.44	5.61		
p27	7.58	7.51	6.96	8.05	8.17	7.81	9.68	9.36	9.93	9.32	9.04	9.97		
KLF4	9.01	12.61	10.99	7.22	10.22	8.94	6.97	6.66	9.07	8.87	10.40	9.83		

Table S5. Delta CT means of genes in RPE and C918 cells

	PBS							ESC									ESC-CM				
F2RL1	4.91	4.82	4.15	4.35			3.9	3.76	3.56						4.05	4.84	4.91				
PIK3CG	7.16	7.82	7.44	8.25	7.17	8.02	6.17	7.55	7.04	7.22	6.79				7.49	7.63	8.6	8.15	8.02		
p21	0.68	0.4	0.41	0.35			0.83	1.61	0.97	0.91	1.74	0.4	0.11	0.43	0.79	1.03	0.3	1.48			

Table S6. Delta CT means of genes in skin tissues

Table S7. Delta CT means of genes in tumor tissues

	PBS						ESC-CM											
F2RL1	10	10.56	9.98	10.11	11.06	10.03	9.32	9.8	9.72	9.89		10.05	9.7	9.8	10.4	10.15		
PIK3CG	10.15	10.14	11.21	10.71	11.43		12.57	11.76	11.65	12.11	11.57	11.36	11	11.28	11.02	11.26	11.42	11.21
p21	5.13	5.64	5.22	5.4			4.61	4.88	5.3			5.33	5.59	5.44	5.57			



Figure S1. Characteristics of ESCs. (A) Representative images of morphology by phase microscopy. **(B)** The expression of KLF4, OCT4 and GFP as determined by immunofluorescent staining. **(C)** Teratoma formed by ESCs stained with hematoxylin and eosin for histological analysis. Scale bar, 50 μm. **(A-C)**



Figure S2. Characteristics of MSCs. (A) Representative images of morphology by phase microscopy. Scale bar, 100 µm. **(B)** Flow cytometry analysis of cell surface markers of MSCs. **(C)** Oil red O staining showing

adipogenic differentiation of MSCs. (D) Alizarin red S staining showing osteogenic differentiation of MSCs. Scale bar, 50 μ m. (C-D)

Figure S3



Figure S3. The ESCMe significantly inhibits the growth and metastasis capacity of C918 cells. (A) Scatter diagram of the proportions of apoptotic C918 cells sorted from Ctrl, TCo, TCo+VO, CCo, and CCo+VO groups, as assessed by flow cytometry. **B-F**, Representative images of clone formation (**B**), wound healing at 0 and 9 h (**C**), migration (**D**), invasion (**E**), and VM (**F**) of C918 cells sorted from Ctrl, TCo, TCo+VO, CCo, and CCo+VO groups. (**G**), Immunofluorescence assays of FGF2, MMP1, MMP9, and VE-cadherin in C918 cells sorted from Ctrl, TCo, TCo+VO, CCo, and CCo+VO groups. Scale bars: 100 μm (**D-E**), 400 μm (**F**), and 50 μm (**G**).



Figure S4. C918 cells treated with ESCs have reduced malignancy and tumorigenesis. (**A**) Western blotting of VEGF-A and FGF2 in C918 cells. β-actin served as the internal control. **B-C**, RT-qPCR (**B**) and Western blot analyses (**C**) of TXNIP and BRMS1 in C918 cells sorted from Ctrl, TCo, and CCo groups (n = 3 biological repeats). β-actin served as the internal control. (**D**) TUNEL assay of tumor tissues obtained from mice subcutaneously injected with C918 cells sorted from Ctrl, TCo, or CCo groups. Arrows mark

apoptotic cells. (**E**), PAS-CD34 staining of tumor tissues obtained from mice subcutaneously injected with C918 cells sorted from Ctrl, TCo, or CCo groups. Red arrowheads mark VM; black arrowheads mark normal endothelialdependent vessels. (**F**) H&E staining of tumor tissues obtained from mice subcutaneously injected with C918 cells sorted from Ctrl, TCo, or CCo groups and N:C ratio analysis. . Scale bar, 50 μ m. (**D-F**) Data are means ± SEMs. **P< 0.01; ***P< 0.001; ****P< 0.0001.



Figure S5. The ESCMe influences the expression of cell cycle promoters and senescence markers. Immunofluorescence staining of cell cycle promoters (cyclin A2, cyclin B1, cyclin D1) and senescence markers (p21, and

p27) in RPE and C918 cells sorted from the Ctrl group (C918 cells co-cultured with RPE cells) and ESC group (C918 cells co-cultured with RPE cells and ESCs). Scale bar, 50 μ m.





Figure S6. ESC treatment alters the expression of PI3K pathway factors in RPE cells and C918 cells. Immunofluorescence staining of PDK2, MTOR, and PTEN in RPE and C918 cells sorted from the Ctrl and ESC groups. Scale bar, 50 μm.





Figure S7. The ESCMe suppresses C918 tumor growth in xenograft mouse models. (A) Schematic workflow of the *in vivo* experiment. Mice were subcutaneously injected with C918 cells and then treated with PBS, ESCs, or ESC-CM. (B) H&E staining of tumor tissue and N:C ratio analysis. (C)-(D), Representative images of H&E staining in the livers and spleens of mice treated with PBS, ESCs, or ESC-CM. (E) Sketch map of the TK suicide gene. (F) In vivo imaging of ESCs dyed with DiR. Scale bar, 50 µm. (B-D) Data are means \pm SEMs. *P< 0.05; **P< 0.01; ***P< 0.001; ***P< 0.0001.