# **Supplementary information**

Scalable Synthesis of Hide substance(Hs)-Chitosan(Cs)–Hydroxyapatite(HAp):A novel bio-composite from industrial wastes and its efficiency in dye removal.

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### General characterization of the material:

### Solid state NMR analysis:

The solid-state NMR experiments were performed on a Bruker Avance-III HD 400 WB NMR spectrometer (9.4 T). The 1H, 13C, 31P and 15N resonance frequencies were 400.07, 100.61, 161.95 and 40.54 MHz respectively. Samples were packed in a 4-mm-diameter zirconia rotor with a Kel-F cap. All the NMR spectra of the samples were recorded at room temperature using a double resonance 4 mm MAS probe. 13C cross polarization/total sideband suppression (CP/TOSS) experiment was performed at a spinning speed of 7 kHz to get sideband free spectrum. Typically, 3k scans were acquired with a relaxation delay of 4s. For 15N, CPMAS experiments carried out at a spinning speed of 5 kHz. 12k scans were employed with a relaxation delay of 5s to obtain good signal to noise ratio. For all the CP experiments contact time of 3 ms with a linear ramp on the 1H contact pulse (50% slop) and 1H 90° pulse length of 4 µs were employed. In the case of 31P, single pulse excitation with proton decoupling experiment at a spinning speed of 5 kHz was carried out for 64 scans, recycle delay of 4s and 31P 90° pulse length of 4 µs. SPINAL-64 decoupling sequence was used to decouple protons during the 13C/15N/31P acquisition by employing radiofrequency field strength of 83 kHz. FID was subjected to an appropriate exponential multiplication function prior to Fourier transform. 13C chemical shifts were referenced externally to the carbonyl signal of glycine at 176.03 ppm, 15N was referenced to solid ammonium chloride at 35.9 ppm and 31P was referenced to ADP at 1 ppm.

# FTIR analysis:

Approximately 40 mg powdered experimental sample was blended with 100 mg spectroscopic grade KBr for the preparation of pellets and FTIR spectra of the samples were taken in Shimadzu FTIR 8300 spectrophotometer with 100 scans and resolution 4 cm<sup>-1</sup>.

# **XPS** spectra

XPS analysis of composite (Hs-Cs-HAp) and HAp samples were carried out using monochromatic X-ray sources Al k $\alpha$  (E =1486.6 ev) and Mg k $\alpha$  (E = 1253.6 ev) by Omicron energy analyser. Deconvolution of the superimposed peak were done using XPS peak (software version 4.1).

#### **XRD** analysis:

Powder X-ray diffraction patterns were obtained using Seifert C 3000 instrument with the following operating conditions – 40kV and 30 mA with a Cu/Ni radiation at  $\lambda$ =1.5406. The relative intensity was recorded in a scattering range (20) of 10-100°.

#### **Optical microscopic and SEM analysis:**

The macro structure of the material has been analysed by Novax zoom stereo microscope by Euromax Microscopen B.V. Holland. For the optical microscope study samples were dried under vacuum oven for 1.5 hr. at 105 °C followed by cooling to RT under desiccating condition prior to analysis. SEM images were taken on a FEI Quanta 200 F, using a tungsten filament doped with lanthanum hexaboride (LaB6) as an X-ray source, fitted with an ETD

detector with high vacuum mode, using secondary electrons and an acceleration tension of 10 or 30 kV. Samples were analyzed by spreading them on a carbon tape. Energy dispersive X-ray spectroscopy (EDX) was used in connection with SEM for the elemental analysis. The elemental mapping was also collected with the same spectrophotometer.

## **TEM analysis:**

The samples for TEM analysis were prepared by mounting an ethanol-dispersed sample on a lacey carbon Formvar coated Cu grid and TEM images of the prepared samples were collected using a Jeol JEM 2100 microscope. Energy dispersive X-ray Spectrometer used for analysis for the elemental analysis and characterisation using TEM 2100 model which can be operated under 80KV to 200KV, having LAB 6 and Tungsten as a filament used for producing high intensity electron beam. Oxford detector (EDS, Oxford Energy Max 80) is used here for analysis under EDS Mode and by using Bright Field and Dark Field Detector for STEM images and here we are using AZtech Software of OXFORD instrument for achieving a better results.

### BET surface area and porosity analysis:

Brunauer-Emmett-Teller (BET) surface area and total pore volume of the material were obtained by measuring their nitrogen adsorption isotherm at 77 K in a surface area and porosity analyser (Quantachrome ASIQwin Autosorb Automated Gas Sorption System). Analysis of macroporous in of the material was done using Hg intrusion Quantachrome Poremaster instrument.

#### **TGA analysis:**

TGA of the samples were performed in TFAQ50 V 6.1 Build 181 instrument. Approximately 10 g of the each sample was encapsulated non hermetically in a platinum pan and scanned from 30 °C to 500 °C at a heating rate of 20 °C per minute under nitrogen atmosphere. Each experiment was repeated thrice for better accuracy of the result.



Figure S1. Solid state C<sup>13</sup> NMR spectra of (a) Hs, (b) Cs, (c) Hs-Cs-HAp (composite)



Figure S2. Solid state N<sup>15</sup> NMR spectra of (a) Hs, (b) Cs, (c) Hs-Cs-HAp (composite)



Figure S3. Solid state P<sup>31</sup> NMR spectra of (a) Hs, (b) Cs, (c) Hs-Cs-HAp (composite)



Figure S4. FTIR spectra of the (a) Hs, (b) Cs, (c) HAp, (d) Hs-Cs-HAp.



Figure S5. XPS spectra of Hs-Cs-HAp



Figure S6. XPS spectra of pure HAp



Figure S7. Elemental mapping of Hs-Cs-HAp; A: Carbon, B: Nitrogen, C: Oxygen, D: Phosphorous, E: Sodium, F: Calcium



Figure S8. TEM EDS of rod like crystalline aggregate (Fig. 2c) found in Hs-Cs-HAp.



Figure S9. Effect of pH and temperature over MB and SY adsorption by Hs-Cs-HAp.



Figure S10. Langmuir and Freundlich isotherms plots for MB-Hs-Cs-HAp and SY-Hs-Cs-HAp adsorption system.



Figure S11. Calculated  $R_L$  value at different  $C_0$  for MB and SY from Langmuir isotherm.



Figure S12. Comparison of adsorption of dyes (a) MB, (b) SY by composite and HAp.



Figure S13. Kinetics of MB and SY adsorption by Hs-Cs-HAp



**Figure S14.** Pseudo-1st order and Pseudo-2<sup>nd</sup> order plots of MB-Hs-Cs-HAp and SY-Hs-Cs-HAp kinetic data.



Figure S15. Boyd plot of MB-Hs-Cs-HAp and SY-Hs-Cs-HAp kinetics data.