Synthesis and Characterization of Tunable, pH-Responsive Nanoparticle-Microgel Composites for Surface-Enhanced Raman Scattering (SERS) Detection

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Figure S1. TGA curves for (a) $PS_{20}P2VP_{80}$ (M), $PS_{20}P2VP_{80}$ (M)/AuCl₄⁻, $PS_{20}P2VP_{80}$ (M)/Au NPs, (b) $PS_{40}P2VP_{60}$ (M), $PS_{40}P2VP_{60}$ (M)/AuCl₄⁻, $PS_{40}P2VP_{60}$ (M)/Au NPs, (c) $PS_{60}P2VP_{40}$ (M), $PS_{60}P2VP_{40}$ (M)/AuCl₄⁻, $PS_{60}P2VP_{40}$ (M)/Au NPs, (d) $PS_{40}P2VP_{60}$ (M), and a sample of $PS_{40}P2VP_{60}$ (M) treated to the UV photoreduction condition.

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Figure S2. (a) Titration curve relating D_H (by DLS) to pH for $PS_{20}P2VP_{80}$ (M)/ Au NPs. SEM images of (b) $PS_{40}P2VP_{60}$ (M)/Au NPs and (c) $PS_{60}P2VP_{40}$ (M)/Au NPs. Scale bar = 2 μ m.

Movie S1. Video depicting the 3-dimensional reconstruction of a PS_0P2VP_{100} (M)/Au NPs composite that was prepared from photoreduced PS_0P2VP_{100} (M)/AuCl₄⁻ (generated by loading with KAuCl₄ using a pH = 1, H₂SO₄ solution). A series of TEM images were acquired by tilting the TEM grid from -70° to +70° with 2° increments. The Inspect3D software used a contrast thresholding condition to identify Au NPs. These NPs are shown with red pixels in the later portion of the video and Figure S3. The magnification is 320,000x.



Figure S3. (a) TEM image of PS_0P2VP_{100} (M)/Au NP. Inset shows the identification process for Au NPs by a contrast thresholding using Inspect3D software on the TEM instrument. The PS_0P2VP_{100} (M)/Au NP was prepared from photoreduced PS_0P2VP_{100} (M)/AuCl₄⁻, generated from a H_2SO_4 solution (pH = 1) of PS_0P2VP_{100} (M) and loading with KAuCl₄. The software-identified Au NPs are tracked in a tomography video (Movie S1) that was generated from tilting the sample from -70° to +70°. (b) Frames, distinguished by the angle of the tilt, from the tomography video show that some of the Au NPs do not move significantly as the sample is tilted indicating that many are located in the within the PS_0P2VP_{100} (M) particle not only on its surface. See Movie S1 of the Supporting Information.



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Figure S4. TGA-FTIR absorbance maps of (a) PS_0P2VP_{100} (M), (b) $PS_{20}P2VP_{80}$ (M), (c) $PS_{40}P2VP_{60}$ (M), (d) $PS_{60}P2VP_{40}$ (M), (e) $PS_{40}P2VP_{60}$ (M) treated to the photoreduction condition, (f) $PS_{20}P2VP_{80}$ (M) / AuCl₄⁻, (g) $PS_{40}P2VP_{60}$ (M)/AuCl₄⁻, (h) $PS_{60}P2VP_{40}$ (M)/AuCl₄⁻, (i) $PS_{20}P2VP_{80}$ (M)/Au NPs, (j) $PS_{40}P2VP_{60}$ (M)/Au NPs, (k) $PS_{60}P2VP_{40}$ (M)/Au NPs, (l) PS_0P2VP_{100} (M)/Ag NPs, (m) $PS_{20}P2VP_{80}$ (M)/Ag NPs, and (n) $PS_{40}P2VP_{60}$ (M)/Ag NPs. (o) FTIR tracer plots for v_{benz} stretch at 1635 cm⁻¹ from the effluent of $PS_{20}P2VP_{80}$ (M)/Au NPs and $PS_{20}P2VP_{80}$ (M) from TGA sample zone. For (a-n), the absorbance scale is shown as a colored intensity bar beside each map.

Figure S5. (a) Raman spectra of the PS_xP2VP_y (M)/Au NPs showing no signal for vibrational modes in the polymer microgel. (b) Raman spectra of the PS_xP2VP_y (M)/Au NPs exposed to solution-state crystal violet (CV) (concentrations indicated in legends) showing no signal for vibrational modes in the polymer microgel or the CV molecule. In (b), the Raman spectra of solution-state CV is shown at bottom for comparison. (c) S/N of the 1620 cm⁻¹ signal for CV ([CV] = 1.0×10^{-8} M) versus the diameter of free Au NPs (in each case the [Au NPs] = 2.5×10^{-11} M) The Au NPs were synthesized and purified according to a literature procedure.¹

Figure S6. Overlay plot for the Raman spectra of the $PS_{20}P2VP_{80}$ (M)/Ag NPs without (red curve) and with CV in the solution ([CV] = 1.1 x 10⁻⁸ M).

Figure S7. Plot for the S/N at 1620 cm⁻¹ for CV (pH = 7; [CV] = 1.1×10^{-8} M) vs. time using PS₂₀P2VP₈₀ (L)/Ag NPs (squares) and PS₀P2VP₁₀₀ (L)/Ag NPs (triangles). On each day of the study, the pH of solutions of PS_xP2VP_y (L)/Ag NPs and CV was adjusted to 2 and then adjusted back to a pH value of 7, at which point the SERS spectra of CV was reacquired. The first data point (time = 0 days) therefore represents a condition that matches that for n = 1 from Figure 6d.

Figure S8. STEM images of PS_0P2VP_{100}/Ag NPs (L) after 0 pH cycles (a) and after 3 pH cycles (b). STEM image of $PS_{20}P2VP_{80}/Ag$ NPs (L) after 0 pH cycles (c) and after 3 pH cycles (d). A lacey carbon support is present in all images (web-like substrate). Scale bars represent 100 nm.

Figure S9. (a) Tracking of S/N of the 1620 cm⁻¹ signal for CV as the PS_0P2VP_{100}/Ag NPs (original polymer solution : pH 2.5 ultrapure water/HCl = 1:50) are cycled between the latex (integer pH cycles) and the microgel (half integer pH cycles) states by adding 41 uL of 1.71 M HCl and 82 uL of 3.0 M NaOH respectively, roman numerals represent a specific condition for the sample that that was characterized by UV-Vis (b-g).

(a)

Figure S10. (a) Tracking of S/N of the 1620 cm⁻¹ signal for CV as the $PS_{20}P2VP_{80}/Ag$ NPs (original polymer solution : pH 2.5 ultrapure water and HCl 1:50) are cycled between the latex (integer pH cycles) and the microgel (half integer pH cycles) states by adding 41 uL of 1.71 M HCl and 82 uL of 3.0 M NaOH respectively, roman numerals represent a specific condition for the sample that that was characterized by UV-Vis (b-g).

References

(1) Frens, G. Controlled Nucleation for the Regulation of the Particle Size in Monodisperse Gold Suspensions. *Nat. Phys. Sci.* **1973**, *241*, 20-22.